Check for updates

OPEN ACCESS

EDITED BY Tiantai Zhang, Chinese Academy of Medical Sciences and Peking Union Medical College, China

REVIEWED BY Wei Peng, Chengdu University of Traditional Chinese Medicine, China Liu Qing-Shan, Minzu University of China, China

*CORRESPONDENCE Rong Wang, I luckchlp369@163.com Wenbin Li, I yfcs2002@163.com

[†]These authors have contributed equally to this work

RECEIVED 14 November 2023 ACCEPTED 10 January 2024 PUBLISHED 01 February 2024

CITATION

Chen L, Jiang L, Shi X, Yang J, Wang R and Li W (2024), Constituents, pharmacological activities, pharmacokinetic studies, clinical applications, and safety profile on the classical prescription Kaixinsan. *Front. Pharmacol.* 15:1338024. doi: 10.3389/fphar.2024.1338024

COPYRIGHT

© 2024 Chen, Jiang, Shi, Yang, Wang and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Constituents, pharmacological activities, pharmacokinetic studies, clinical applications, and safety profile on the classical prescription Kaixinsan

Liping Chen^{1.2†}, Lin Jiang^{1†}, Xiaoyu Shi¹, Jihong Yang¹, Rong Wang^{2*} and Wenbin Li^{2*}

¹School of Pharmacy, Gansu University of Chinese Medicine, Lanzhou, Gansu, China, ²Department of Pharmacy, The 940th Hospital of Joint Logistic Support Force of PLA, Lanzhou, China

Kaixinsan (KXS) is a noteworthy classical prescription, which consists of four Chinese medicinal herbs, namely Polygalae Radix, Ginseng Radix et Rhizoma, Poria, and Acori Tatarinowii Rhizoma. KXS was initially documented in the Chinese ancient book Beiji Qianjin Yaofang written by Sun Simiao of the Tang Dynasty in 652 A.D. As a traditional Chinese medicine (TCM) prescription, it functions to nourish the heart and replenish Qi, calm the heart tranquilize the mind, and excrete dampness. Originally used to treat amnesia, it is now also effective in memory decline and applied to depression. Although there remains an abundance of literature investigating KXS from multiple aspects, few reviews summarize the features and research, which impedes better exploration and exploitation of KXS. This article intends to comprehensively analyze and summarize up-to-date information concerning the chemical constituents, pharmacology, pharmacokinetics, clinical applications, and safety of KXS based on the scientific literature, as well as to examine possible scientific gaps in current research and tackle issues in the next step. The chemical constituents of KXS primarily consist of saponins, xanthones, oligosaccharide esters, triterpenoids, volatile oils, and flavonoids. Of these, saponins are the predominant active ingredients, and increasing evidence has indicated that they exert therapeutic properties against mental disease. Pharmacokinetic research has illustrated that the crucial exposed substances in rat plasma after KXS administration are ginsenoside Re (GRe), ginsenoside Rb1 (GRb1), and polygalaxanthone III (POL). This article provides additional descriptions of the safety. In this review, current issues are highlighted to guide further comprehensive research of KXS and other classical prescriptions.

KEYWORDS

Kaixinsan, constituents, pharmacology, pharmacokinetics, clinical applications

1 Introduction

Due to the distinctive superiority and less adverse effects on the prevention and treatment of sophisticated diseases, Chinese classical prescriptions are attracting more and more attention worldwide (Lu et al., 2021b; Hao et al., 2022). Kaixinsan (KXS) is a noteworthy classical prescription, which was initially described in the Chinese ancient book

Beiji Qianjin Yaofang written by Sun Simiao of the Tang Dynasty in 652 A.D. It was originally applied for the treatment of dementia and morbid forgetfulness and is made up of Polygalae Radix (PR), Ginseng Radix et Rhizoma (GR), Poria, and Acori Tatarinowii Rhizoma (ATR) (Zhang et al., 2019). In light of its long history and efficacy, KXS is listed in a catalog of ancient classic prescriptions (the first batch) in China (Chen et al., 2018; Jiang et al., 2021). According to TCM theories, dysfunctions of the spleen cannot nourish the heart, and thus produce dampness to block the heart, which leads to psychiatric diseases with the pivotal symptoms of dizziness, anhedonia, amnesia, and morbid forgetfulness (Pei et al., 2020). KXS has the function of soothing the nerves, replenishing Qi (vital energy), nourishing the heart, and eliminating dampness, thereby improving the dysfunction of the heart and spleen as well as ameliorating diverse forms of mental diseases in clinical settings (Cao et al., 2020; Hu et al., 2021a).

Furthermore, it is known that KXS follows the rule of drug synergism and compatibility in TCM, in which Poria and ATR play adjuvant roles in facilitating the delivery of the principal herb (PR and GR) to the disease site *in vivo*, thus improving the whole prescription to achieve the best effect (Wang et al., 2010; Guo et al., 2019b).

PR, the dried root of the Polygalaceae family member *Polygala tenuifolia* Willd., *Polygala senega* L. or *Polygala sibirica* L. (https:// mpns.science.kew.org), is designated as the most essential medication in the prescription (Shan et al., 2023a). Its traditional efficacies include soothing the nerves, improving intelligence, dispelling phlegm, and reducing swelling (Chen et al., 2022). With wide-ranging pharmacological properties that include antidepressant, neuroprotective, hypnotic-sedative, antitumor, antioxidant, anti-inflammatory, antiviral, and antiaging effects, it is broadly used in clinics to treat forgetfulness, depression, insomnia, cough, and palpitation (Jiang et al., 2021; Zhang et al., 2023). Moreover, PR is often combined with Poria, and ATR to by eliminating phlegm and dampness and decreasing gastrointestinal toxicity (Sun et al., 2020).

GR is the dried root of *Panax ginseng* C. A. Meyer of the Araliaceae family (https://mpns.science.kew.org). It is acquainted as the king of herbs because it is widely used as medicinal and functional remedies for miscellaneous diseases in China and other East Asian countries (Li et al., 2021a; Lu et al., 2021a). GR can nourish weakness, replenish energy, invigorate the spleen, benefit the lungs, reinforce vital energy, and consolidate robustness (Liu et al., 2020; Potenza et al., 2023). As the minister medicinal in this prescription, GR is helpful to enter the heart and kidney meridian and enhance the tonifying effect of PR.

Poria and ATR are adjuvant drugs in the prescription. Poria is the dried sclerotium of *Poria cocos* (Schw.) Wolf (Pharmacopoeia Commission of the People's Republic of China, 2020). It is traditionally used to promote urination, excrete dampness, invigorate the spleen, and tranquilize the mind. ATR is derived from the dried rhizomes of *Acorus tatarinowii* Schott (Pharmacopoeia Commission of the People's Republic of China, 2020). It can open the orifices (resuscitating), calm the mind, resolve dampness, and harmonize the stomach. Poria cooperates with ATR to strengthen the diuretic effect and sedative activities and thus could promise superior efficacy against amnesia in whole prescription. The Chinese herbal medicine that makes up KXS is illustrated in Figure 1.

Although there are some reports relating to the quality research of KXS, it still failed to form a stable and systematic quality standard system, seriously affecting the therapeutic effects and safety of the prescription (Fu et al., 2019; Bo et al., 2022). Based on the literature to date, the systematic summarization integrating its safety, quality control, and precise mechanisms of action, is still lacking from the point of view of KXS development. Herein, this review discusses current knowledge about the chemical constituents, pharmacological activities, pharmacokinetics, clinical studies, and safety of KXS. It may provide research material and a theoretical foundation for the future study and subsequent development of this classical prescription.

2 Chemical constituents

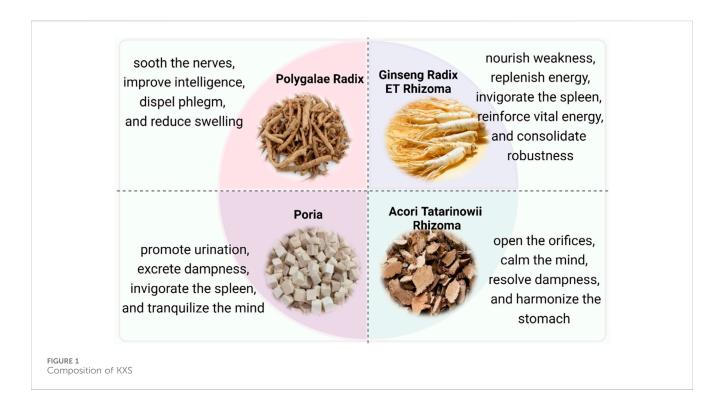
So far, 107 chemical compounds have been isolated from KXS. These components are primarily classified into five types, namely, 65 saponins (1–65), 5 xanthones (66–70), 24 oligosaccharide esters (71–94), 7 triterpenoids (95–101), and 6 other types of compounds (102–107). Of these, saponins occupy the majority, and more than 60 saponins have been reported. In this article, we summarized all chemical constituents isolated in KXS and showed in Table 1, along with their corresponding structures in Figures 2–6.

2.1 Saponins

Saponins are a diverse category of natural compounds that contain one or more sugar chains and a triterpene or steroid aglycone in the basic parent nucleus. This particular category of natural components is a broad-scale distribution in Chinese herbs. Saponins are the predominant active ingredient of KXS, and increasing evidence has indicated that they exert therapeutic properties against memory and cognitive impairment, depression, and neurodegenerative diseases (Lu et al., 2022; Qiu et al., 2022). To date, a total of 65 saponins have been reported in KXS, which mostly consist of polygala saponins and ginsenosides. Structurally, ginsenosides are further classified into four major categories based on the parent ring structure, including the protopanaxadiol type (33–37, 44–46, 50–52, 54–57), the protopanaxatriol type (38–40, 42, 43, 47–49, 53), the oleanane group (41) and pseudoginsenosides of the occillol type (63, 64).

2.2 Xanthones

The name xanthones designates a class of secondary metabolites normally found in a few higher plant families. The polygalaceae family is an important source of natural xanthones which have the basic mother nucleus of 9*H*-xanthen-9-one or xanthone. Currently, 5 xanthones from KXS have been identified, including sibiricaxanthone B (66), sibiricaxanthone F (67), 7-Omethylmangiferin (68), polygalaxanthone III (69), and polygalaxanthone XXVIII (70).



2.3 Oligosaccharide esters

Oligosaccharide esters are a sizable class of natural metabolites with sucrose and trisaccharide esters as a character in their chemical structures and have a variety of physiological activities. At present, a total of 24 oligosaccharide esters have been reported in KXS.

2.4 Triterpenoids

There are currently 7 reported triterpenoids that have been identified from KXS. Poria is rich sources of triterpenoids.

2.5 Other compounds

In addition to the above ingredients, KXS also contains volatile oils (**102–106**) and flavonoids (**107**).

3 Pharmacological activities

Traditionally, KXS has the effect of replenishing Qi, nourishing the heart, and calming the mind. Therefore, it has been used clinically to treat a range of nervous system conditions. Modern studies on KXS have been carried out to demonstrate pharmacological effects. The pharmacological effects are shown in Table 2.

3.1 Anti-Alzheimer's disease effects

Alzheimer's disease (AD) is the most common neurodegenerative disease, manifesting progressive cognitive

impairment, amnesia, and mental behavioral changes (Conti Filho et al., 2023). The curative effects of KXS on amnesia have been verified throughout millennia. It is widely accepted that the gradual enrichment of Amyloid- β (A β) peptide as well as abnormal or hyperphosphorylated Tau are strongly correlated with AD pathogenesis (Khayat et al., 2021; Wegmann et al., 2021; Zhang et al., 2021). As shown in Figure 7, KXS has been proven to possess excellent anti-AD effects by inhibiting neuroinflammation, neurological apoptosis, and oxidative stress, and increasing the supply of neurotransmitters.

AD was induced in male SD rats through an intraperitoneal injection of D-gal (50 mg/kg/d) and a stereotaxic injection of $4\,\mu g\,A\beta_{25\text{-}35}$ (1 $\mu g/\mu L)$ for 6 weeks. The rats were orally administered KXS water extracts (10 g/kg/d) or huperzine A (30 µg/kg) from the third week. The spatial learning and memory were measured with the escape latency, platform crossings, and swimming distance to find the removal platform in the target quadrant of the Morris water maze. Neuronal density hippocampal CA1 regions was performed using in immunohistochemistry staining. According to the results of all the detection indexes, the water extract observably decreased the escape latency and swimming distance, increased platform crossings, increased fluorescence intensity of neuronal nuclei, ameliorated central neurotransmitter loss, reduced Tau hyperphosphorylation via regulating the expression of PI3K/ Akt. Moreover, the extract dramatically decreased the concentrations of inflammatory factors and ROS-induced neuronal apoptosis via the PI3K/Akt/GSK-3ß signaling pathway in the rat hippocampus. In vitro, the extracts potently inhibited the percentage of apoptotic cells and the expressions of Bax/bcl-2 and cleaved-caspase-3, meliorated A_{β25-35}-induced ROS accumulation and LDH release, and reduced the expression of Tau signaling pathway in PC12 cells (at the dose of 1 mg/mL). Based on the

No.	nical constituents isolated fro	Formula	Structure type	Source	References
	Onjisaponin A	C ₈₀ H ₁₂₀ O ₃₉	Saponins	PR	Lv et al. (2016b)
	Onjisaponin B	C ₇₅ H ₁₁₂ O ₃₅	Saponins	PR	Lv et al. (2016b)
	Onjisaponin E	C ₇₁ H ₁₀₆ O ₃₃	Saponins	PR	Lv et al. (2016b)
:	Onjisaponin F	C ₇₅ H ₁₁₂ O ₃₆	Saponins	PR	Lv et al. (2016b)
	Onjisaponin G	C ₇₀ H ₁₀₄ O ₃₂	Saponins	PR	Lv et al. (2016b)
	Onjisaponin J	C ₈₅ H ₁₂₆ O ₄₂	Saponins	PR	Lv et al. (2016b)
	Onjisaponin K	C ₇₆ H ₁₁₂ O ₃₆	Saponins	PR	Lv et al. (2016b)
	Onjisaponin L	C ₈₆ H ₁₂₈ O ₄₃	Saponins	PR	Lv et al. (2016b)
	Onjisaponin O	C ₇₇ H ₁₁₆ O ₃₇	Saponins	PR	Lv et al. (2016b)
0	Onjisaponin R	C ₇₆ H ₁₁₄ O ₃₇	Saponins	PR	Lv et al. (2016b)
1	Onjisaponin T	$C_{83}H_{124}O_{42}$	Saponins	PR	Lv et al. (2016b)
2	Onjisaponin V	$C_{82}H_{122}O_{41}$	Saponins	PR	Lv et al. (2016b)
3	Onjisaponin W	$C_{81}H_{120}O_{40}$	Saponins	PR	Lv et al. (2016b)
4	Onjisaponin X	$C_{87}H_{130}O_{45}$	Saponins	PR	Lv et al. (2016b)
5	Onjisaponin Y	C ₆₉ H ₁₀₂ O ₃₀	Saponins	PR	Lv et al. (2016b)
6	Onjisaponin Z	$C_{71}H_{106}O_{32}$	Saponins	PR	Lv et al. (2016b)
7	Onjisaponin Ng	C ₈₀ H ₁₁₈ O ₃₈	Saponins	PR	Lv et al. (2016b)
8	Onjisaponin Sg	C ₈₇ H ₁₃₀ O ₄₄	Saponins	PR	Liu and Ye (2012)
9	Polygalasaponin XIV	C ₅₄ H ₈₆ O ₂₄	Saponins	PR	Reeju et al., (2017)
	XXIV				
0	polygalasaponin XVIII	C ₅₉ H ₉₄ O ₂₈	Saponins	PR	Reeju et al., (2017)
1	Polygalasaponin XIX	$C_{64}H_{102}O_{32}$	Saponins	PR	Reeju et al., (2017)
2	Polygalasaponin XXI	C ₅₁ H ₆₀ O ₂₇	Saponins	PR	Reeju et al., (2017)
	XXI				
3	Polygalasaponin XXII	C ₅₈ H ₉₂ O ₂₈	Saponins	PR	Reeju et al., (2017)
4	Polygalasaponin XXXII	C ₇₉ H ₁₁₈ O ₃₈	Saponins	PR	Liu and Ye (2012)
5	Polygalasaponin XXXV	C ₆₃ H ₉₈ O ₃₁	Saponins	PR	Reeju et al., (2017)
6	Arilloside E	$C_{66}H_{104}O_{34}$	Saponins	PR	Reeju et al., (2017)
7	Polygalasaponin XXXXII	C ₆₇ H ₁₀₆ O ₃₅	Saponins	PR	Reeju et al., (2017)
8	Senegasaponin B	C ₆₉ H ₁₀₂ O ₃₁	Saponins	PR	Reeju et al., (2017)
9	Senegin II	C ₇₀ H ₁₀₄ O ₃₂	Saponins	PR	Reeju et al., (2017)
0	Senegin III	C ₇₅ H ₁₁₂ O ₃₅	Saponins	PR	Lv et al. (2016b)
1	Senegin IV	C ₈₁ H ₁₂₂ O ₃₉	Saponins	PR	Liu and Ye (2012)
2	Arillatanoside C	C ₆₄ H ₁₀₂ O ₃₃	Saponins	PR	Lv et al. (2016b)
3	Ginsenoside Rb1	C ₅₄ H ₉₂ O ₂₃	Saponins	GR	Lv et al. (2016b)
4	Ginsenoside Rb2	C ₅₃ H ₉₀ O ₂₂	Saponins	GR	Lv et al. (2016b)
5	Ginsenoside Rc	C ₅₃ H ₉₀ O ₂₂	Saponins	GR	Lv et al. (2016b)

TABLE 1 Chemical constituents isolated from KXS.

(Continued on following page)

TABLE 1 (Continued) Chemical constituents isolated from KXS.

No.	Compounds	Formula	Structure type	Source	References
37	Ginsenoside Rd	C48H82O18	Saponins	GR	Lv et al. (2016b)
38	Ginsenoside Re	C48H82O18	Saponins	GR	Lv et al. (2016b)
39	Ginsenoside F1	C ₃₆ H ₆₂ O ₉	Saponins	GR	Lv et al. (2016b)
40	Ginsenoside F4	C ₄₂ H ₇₀ O ₁₂	Saponins	GR	Lv et al. (2016b)
41	Ginsenoside Ro	C ₄₈ H ₇₆ O ₁₉	Saponins	GR	Lv et al. (2016b)
42	Ginsenoside Rg1	C ₄₂ H ₇₂ O ₁₄	Saponins	GR	Lv et al. (2016b)
43	Ginsenoside Rg2	C42H72O13	Saponins	GR	Lv et al. (2016b)
44	20SGinsenoside Rg3	C ₄₂ H ₇₂ O ₁₃	Saponins	GR	Lv et al. (2016b)
45	20RGinsenoside Rg3	C ₄₂ H ₇₂ O ₁₃	Saponins	GR	Lv et al. (2016b)
46	Ginsenoside Rg5	C ₄₂ H ₇₀ O ₁₂	Saponins	GR	Lv et al. (2016b)
47	Ginsenoside Rg6	C42H70O12	Saponins	GR	Lv et al. (2016b)
48	Ginsenoside Rh1	C ₃₆ H ₆₂ O ₉	Saponins	GR	Lv et al. (2016b)
49	Ginsenoside Rf	$C_{42}H_{72}O_{14}$	Saponins	GR	Lv et al. (2016b)
50	Ginsenoside Rh3	C36H60O7	Saponins	GR	Lv et al. (2016b)
51	Ginsenoside Rk1	$C_{42}H_{70}O_{12}$	Saponins	GR	Lv et al. (2016b)
52	Ginsenoside Rk2	C ₃₆ H ₆₀ O ₇	Saponins	GR	Lv et al. (2016b)
53	Ginsenoside Rk3	C36H60O8	Saponins	GR	Lv et al. (2016b)
54	Ginsenoside Rs3	C ₄₄ H ₇₄ O ₁₄	Saponins	GR	Lv et al. (2016b)
55	Ginsenoside Rs4	C444H72O13	Saponins	GR	Lv et al. (2016b)
56	Ginsenoside Rh2	C ₃₆ H ₆₂ O ₈	Saponins	GR	Lv et al. (2016b)
57	Ginsenoside F4/Rg4	C42H70O12	Saponins	GR	Lv et al. (2016b)
58	Notoginsenoside R ₂	C ₄₁ H ₇₀ O ₁₃	Saponins	GR	Liu et al. (2005)
59	Notoginsenoside R ₄	C ₅₉ H ₁₀₀ O ₂₇	Saponins	GR	Liu et al. (2005)
60	Zingibroside R ₁	$C_{42}H_{66}O_{14}$	Saponins	GR	Lv et al. (2016b)
61	Malonyl-ginsenoside Rb1	C ₅₇ H ₉₄ O ₂₆	Saponins	GR	Lv et al. (2016b)
62	Malonyl-ginsenoside Rd	C ₅₁ H ₈₄ O ₂₁	Saponins	GR	Lv et al. (2016b)
63	Pseudoginsenoside F11	C ₄₂ H ₇₂ O ₁₄	Saponins	GR	Lv et al. (2016b)
64	Pseudoginsenoside RT1	C47H74O18	Saponins	GR	Lv et al. (2016b)
65	Tenuifolin	C ₃₆ H ₅₆ O ₁₂	Saponins	PR	Lv et al. (2016b)
66	Sibiricaxanthone B	C ₂₄ H ₂₆ O ₁₄	Xanthones	PR	Lv et al. (2016b)
67	Sibiricaxanthone F	C ₂₉ H ₃₄ O ₁₆	Xanthones	PR	Lv et al. (2016b)
68	7-O-Methylmangiferin	C ₂₀ H ₂₀ O ₁₁	Xanthones	PR	Lv et al. (2016b)
69	Polygalaxanthone III	C ₂₅ H ₂₈ O ₁₅	Xanthones	PR	Wang et al. (2021b)
70	Polygalaxanthone XXVIII	C ₅₃ H ₈₄ O ₂₄	Xanthones	PR	Lv et al. (2016b)
71	3,6′-disinapoyl sucrose	C ₃₄ H ₄₂ O ₁₉	Oligosaccharide esters	PR	Lv et al. (2016a); Lin et al. (2021)
72	Sibiricose A1	C ₂₃ H ₃₂ O ₁₅	Oligosaccharide esters	PR	Lv et al. (2016b)
73	Sibiricose A3	C ₁₉ H ₂₆ O ₁₃	Oligosaccharide esters	PR	Wang et al. (2017b)
74	Sibiricose A5	C ₂₂ H ₃₀ O ₁₄	Oligosaccharide esters	PR	Lv et al. (2016a)
				1	

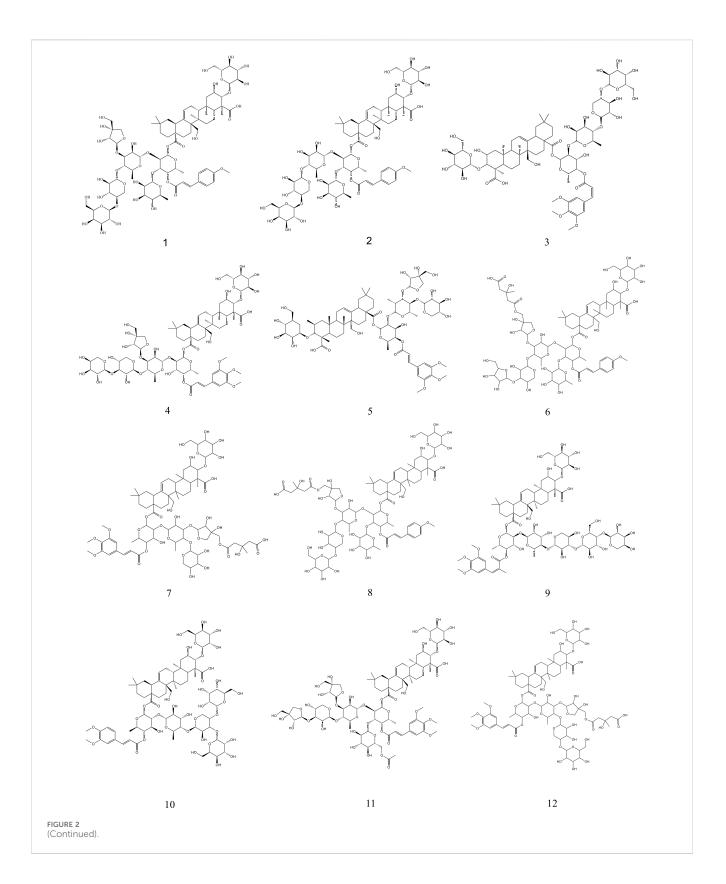
(Continued on following page)

TABLE 1 (Continued) Chemical constituents isolated from KXS.

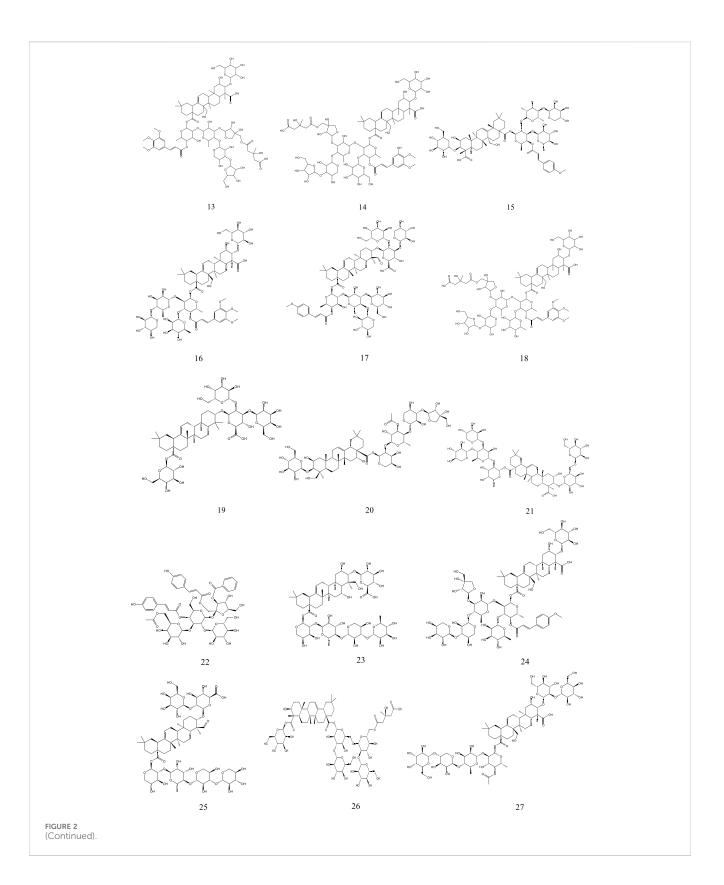
No.	Compounds	Formula	Structure type	Source	References
75	Sibiricose A6	$C_{23}H_{32}O_{15}$	Oligosaccharide esters	PR	Lv et al. (2016a)
'6	Tenuifoliside A	C ₃₁ H ₃₈ O ₁₇	Oligosaccharide esters	PR	Lv et al. (2016b); Lin et al. (2021)
7	Tenuifoliside B	C ₃₀ H ₃₆ O ₁₇	Oligosaccharide esters	PR	Lv et al. (2016b)
'8	Tenuifoliside C	C35H44O19	Oligosaccharide esters	PR	Lv et al. (2016b)
79	Tenuifoliose A	C ₆₂ H ₇₆ O ₃₅	Oligosaccharide esters	PR	Lv et al. (2016b)
60	Tenuifoliose B/D	C ₆₀ H ₇₄ O ₃₄	Oligosaccharide esters	PR	Lv et al. (2016b)
1	Tenuifoliose E/C	C ₅₈ H ₇₂ O ₃₃	Oligosaccharide esters	PR	Lv et al. (2016b)
32	Tenuifoliose H	C ₆₁ H ₇₄ O ₃₄	Oligosaccharide esters	PR	Lv et al. (2016b)
3	Tenuifoliose I/J	C ₅₉ H ₇₂ O ₃₃	Oligosaccharide esters	PR	Lv et al. (2016b)
34	Tenuifoliose K	C ₅₇ H ₇₀ O ₃₂	Oligosaccharide esters	PR	Lv et al. (2016b)
5	Tenuifoliose N	C ₆₃ H ₇₈ O ₃₆	Oligosaccharide esters	PR	Wang et al. (2017a)
6	Tenuifoliose F	C ₆₈ H ₈₆ O ₃₉	Oligosaccharide esters	PR	Lv et al. (2016b)
37	Tenuifoliose G	C ₆₆ H ₈₄ O ₃₈	Oligosaccharide esters	PR	Lv et al. (2016b)
38	Tenuifoliose L	C ₆₇ H ₈₄ O ₃₈	Oligosaccharide esters	PR	Lv et al. (2016b)
9	Tenuifoliose M	C ₆₅ H ₈₂ O ₃₇	Oligosaccharide esters	PR	Lv et al. (2016b)
0	Tenuifoliose O	C ₆₁ H ₇₆ O ₃₅	Oligosaccharide esters	PR	Lv et al. (2016b)
91	Telephiose C	C32H40O18	Oligosaccharide esters	PR	Lv et al. (2016b)
2	Arillanin A	C33H40O18	Oligosaccharide esters	PR	Wang et al. (2017b)
13	Arillanin C	C22H30O14	Oligosaccharide esters	PR	Wang et al. (2017a)
4	Reiniose J	C ₆₀ H ₇₄ O ₃₄	Oligosaccharide esters	PR	Wang et al. (2017b)
5	Dehydropachymic acid	C33H50O5	Triterpenoids	Poria	Lv et al. (2016b)
6	Dehydrotumulosic acid	C31H48O4	Triterpenoids	Poria	Wang et al. (2021b)
7	Dehydrotrametenolic acid	C ₃₀ H ₄₆ O ₃	Triterpenoids	Poria	Wang et al. (2021b)
8	Tumulosic acid	C31H2004	Triterpenoids	Poria	Lv et al. (2016a)
19	Pachymic acid	C ₃₃ H ₅₂ O ₅	Triterpenoids	Poria	Wang et al. (2021b)
.00	Polyporenic acid C	C ₃₁ H ₄₆ O ₄	Triterpenoids	Poria	Lin et al. (2021)
01	Poricoic acid A	C ₃₁ H ₄₆ O ₅	Triterpenoids	Poria	Lin et al. (2021)
02	α-asarone	C ₁₂ H ₁₆ O ₃	Other compounds	ATR	Wang et al. (2021a)
03	β-asarone	C ₁₂ H ₁₆ O ₃	Other compounds	ATR	Wang et al. (2021a)
04	Methyleugenol	C ₁₁ H ₁₄ O ₂	Other compounds	ATR	Lin et al. (2021)
05	Acorenone B	C ₁₅ H ₂₄ O	Other compounds	ATR	Lin et al. (2021)
06	Shyobunol	C ₁₅ H ₂₆ O	Other compounds	ATR	Lin et al. (2021)
.07	Spinosin	C ₂₈ H ₃₂ O ₁₅	Other compounds	ATR	Lin et al. (2021)

results of the *in vivo* and *in vitro* experiments, KXS extracts indicated its potentially beneficial role in ameliorating AD progression (Guo et al., 2019c). An *in vivo* study on AD rats after treatment with KXS indicated that 23 constituents were found in AD rats' plasma, including 1 from PR, 8 from GR, 4 from Poria, 6 from ATR, and 4 that were only discovered in KXS. meanwhile, 10 components were detected in AD rats' brains, including 4 from GR and 6 that were only existed in the whole prescription.

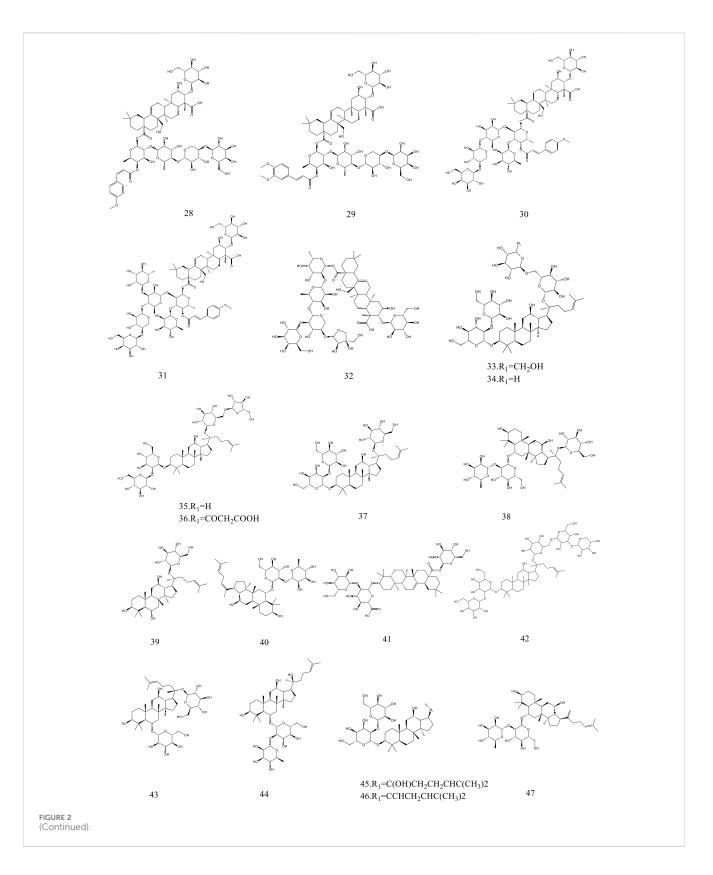
Furthermore, it was found that the peak areas of some constituents in the KXS group were bigger than those in the single herbs group *via* semiquantitative analysis. KXS might mediate oxidative stress, neuroinflammation, and apoptosis both *in vivo* and *in vitro*, especially when the four single herbs were adopted in combination. These results demonstrated the synergistic function and the whole prescription compatibility rationality of KXS to a certain degree (Guo et al., 2019a).



Meta-analysis of animal experiments is believed to provide a high level of research evidence (Chou et al., 2020). Network pharmacology is helpful in elucidating the mechanisms of the TCM formula in the treatment of complex diseases in a comprehensive and systematic manner (Zhou et al., 2022a). To further elucidate the active ingredients and the mechanisms of KXS, Yi et al. (2020) employed meta-analysis, network pharmacology analyses, and molecular docking to investigate the efficacy and potential underlying action mechanisms of KXS against AD from a holistic prospect. The result indicated that aposcopolamine,

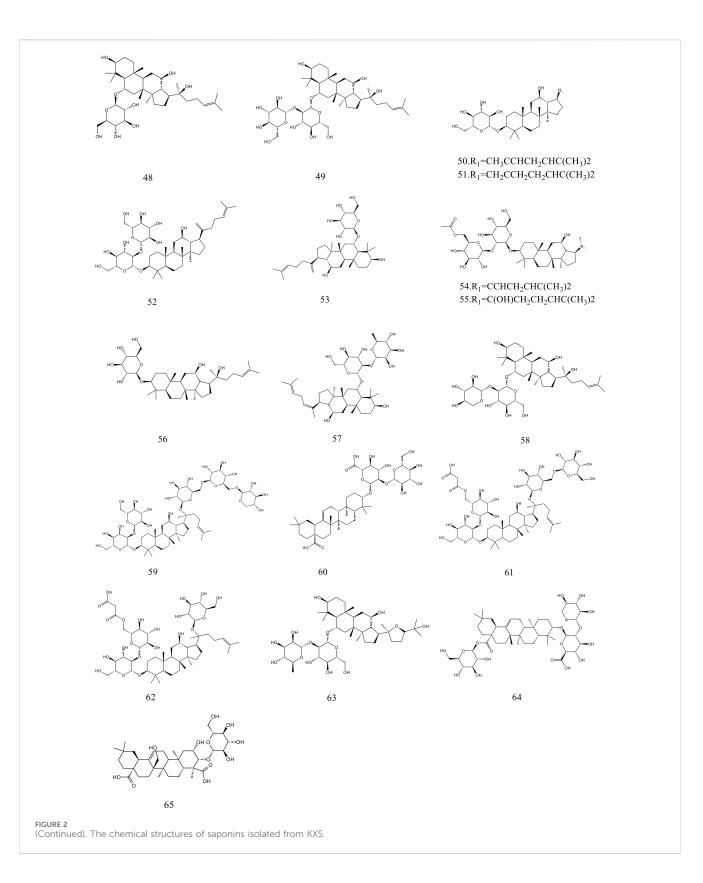


stigmasterol, and inermin exhibited good affinity for several key genes associated with memory and cognition, such as muscarinic acetylcholine receptors, adrenergic receptors, and acetylcholinesterase (AChE). Scopolamine-induced amnesia is an excellent model of behavioural-cum-cognitive deficits to study dementing-related illnesses such as AD (Tang, 2019). In scopolamine (3 mg/kg/d i. p.)-induced cognitive dysfunction mice, Luo et al. (2020) validated the anti-inflammatory effects of the ethanol extracts of KXS (at doses of 1.4 and 2.8 g/kg) by decreasing proinflammatory cytokine levels such as tumor

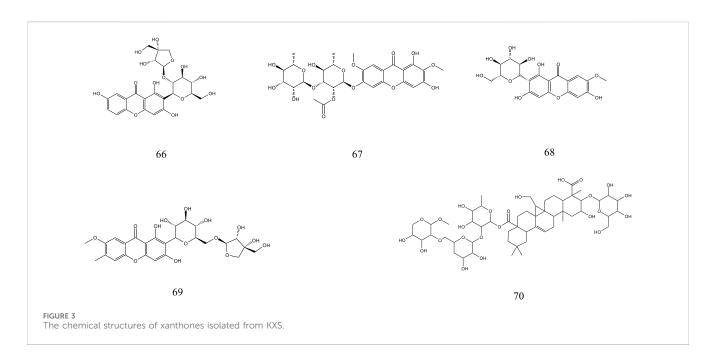


necrosis factor (TNF)- α , interleukin (IL)-6 and IL-1 β both in the hippocampus and cortex, which were consistent with the network pharmacology analyses. More importantly, it was found that cholinergic system dysfunction improvement of KXS was significantly correlated with regulation of the nicotinic

acetylcholine receptor CHRNB2. Senescence accelerated mouse prone 8 (SAMP8) was reported to develop typical AD pathologies, such as an abnormal expression of anti-aging factors, inflammation, oxidative stress, tau hyperphosphorylation, amyloid-(A) deposits, abnormal autophagy activity, and endoplasmic



reticulum stress (Andrea et al., 2022). Therefore, SAMP8 is widely used and considered as an ideal model for AD research, displaying age-related cognitive decline and memory loss (Xie et al., 2020). In a subsequent study, the potential molecular mechanisms of KXS on AD which was predicted by the network pharmacology approach were experimentally validated in SAMP8. The result indicated that the water extract (at doses of 5.4 g/kg) significantly upregulated AKT phosphorylation, inhibited the activation of GSK3 β and CDK5, and suppressed the TLR4/MyD88/NF-KB pathway to suppress neuronal apoptosis and attenuate cognitive impairment, thus alleviating the



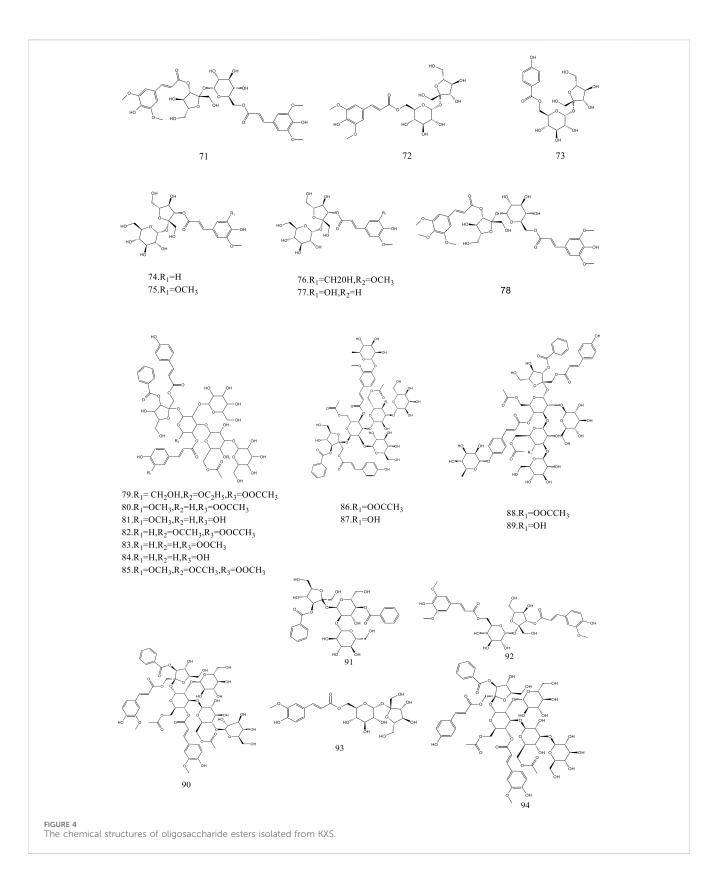
progression of AD (Jiao et al., 2022). Although dozens of *in vivo* and *in vitro* experiments of KXS and its components have supported the anti-AD potentials, explicit pharmacokinetic clinical trials should be executed to further explore the principles of formulating prescriptions and elucidate the correlation between bioactive ingredients and pharmacological function.

3.2 Anti-depression effects

Depression, a common but devastating psychiatric illness characterized by anhedonia, despair, anxiety, cognitive changes, and sleep disturbance, has become a pressing public health problem that leads to a heavy socioeconomic burden (Cladder-Micus et al., 2018). More than 264 million people worldwide are suffering from this debilitating illness (Monroe and Harkness, 2022). Unfortunately, depression is an extremely complex disease caused by multiple factors, and the actual etiology of this disease is still unclarified (McCarron et al., 2021). Nowadays, substantial basic research is ongoing to shed light on depression. Many pathological hypotheses including brain-derived neurotrophic factor (BDNF), neurotransmitter imbalance, over-stimulation of inflammatory, hypothalamic-pituitary-adrenal (HPA) axis disturbances, and endogenous metabolites, have been put forward (Fox and Lobo, 2019).

In terms of TCM theory, depression is believed to be caused by the stagnant of Qi (vital energy), moisture, phlegm, or heat, which ultimately leads to brain dysfunction (Li et al., 2023). KXS has the function of tonifying Qi, nourishing the heart, eliminating dampness, and resolving phlegm, thereby for the treatment of depression function. An increasing number of studies have suggested that KXS was used to effectively treat depression by regulating autophagy, neurotrophic factors, neurotransmitters, synaptogenesis, and inflammation *in vivo* and *in vitro* experiments (Figure 8) (Zhou et al., 2012; Zhu et al., 2012; Zhu et al., 2013; Dong et al., 2017; Fu et al., 2019). Emerging experimental investigations have suggested that chronic stress which causes many impairments in mood, memory, and cognition, has been considered to play a causal role in depression onset (Cruz-Pereira et al., 2020). The chronic unpredictable mild stress (CUMS) model of depressive disorder is currently the most commonly employed, effective, and reliable experimental tool to assess human psychopathology (Antoniuk et al., 2019). In a preclinical study where fluoxetine (10 mg/kg/d) was used as a positive control, intra-gastric administration of KXS water extract at the dosage of 3, 5, or 10 g/kg/day for 2 weeks remarkably ameliorated depressive-like behaviors of eight CUMS rats in sucrose preference test, forced swimming test and open field test, thereby relieving the symptoms of anhedonia, despair, and anxiety. To be more specific, the sucrose preference was markedly increased in the minimum effective dosage (5 g/kg/day) of the treated group compared with the control group (p < 0.05). The improvements in depressive-like behaviors seen with KXS-H (10 g/kg/d) were similar to the results seen with fluoxetine, demonstrating the efficacy of KXS on depression treatment. In parallel, this study confirmed that KXS inhibited inflammationstimulated depression via resisting NLRP3 inflammasome activation and inducing autophagy in vivo and in vitro, as evidenced by the decrease in tumor necrosis factor (TNF-a), interleukin (IL)-6, IL-1β mRNA expression and IL-1β protein expression, as well as the accumulation in ROS content (Yu et al., 2021).

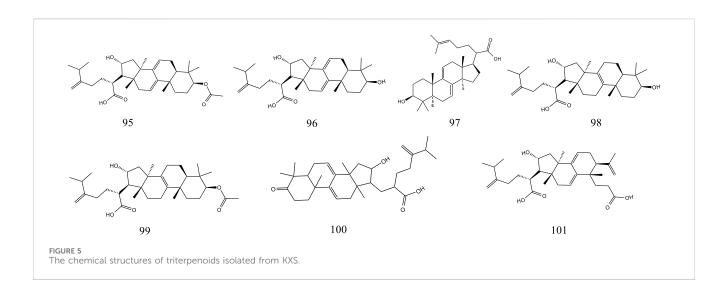
KXS was beneficial for modulating the metabolite pathway of neurotrophins (Zhu et al., 2017), inducing synaptic protein expressions (Zhu et al., 2016), and regulating pro-inflammatory cytokines in microglia (Qu et al., 2021), which might account for the concrete regulation mechanism. According to a network pharmacology investigation, 4 compounds including Asarone, Pachymic acid, Ginsenoside Rg1, and Polygalaxanthone III were identified as the key active components of KXS for treating depression (Du et al., 2022). These compounds might target 6 target genes that were closely related to four pathways, such as

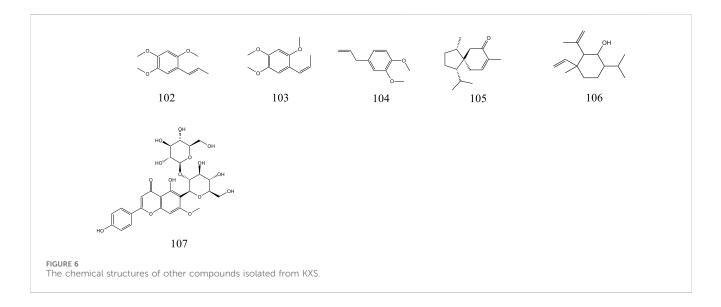


the serotonergic synapse pathway, neuroactive ligand-receptor interaction pathway, MAPK signaling pathway, and PI3K-Akt signaling pathway (Du et al., 2022).

A considerable number of studies have demonstrated that depressive symptoms are the highly prevalent risk factor for

cardiovascular morbidity and mortality (Carney and Freedland, 2016; Bucciarelli et al., 2020; Li et al., 2020). It has been reported that comorbid depression often occurs in patients with coronary heart disease (CHD) (Khandaker et al., 2019), and the comorbidity complicates the depression treatment and worsens the





cardiovascular prognosis (Carney and Freedland, 2016). TCM theory specifies that "Heart Governs the Spirit Light". This means that a dysfunctional heart correlates strongly with mental diseases, such as insomnia, depression, and insanity (Ong et al., 2018). According to one study, KXS water extract (370 mg/kg) provided obvious protection against postmyocardial infarction depression (Hu et al., 2020). Compared to the model group, KXS treatment dramatically improved the levels of ejection fraction and fractional shortening, reduced the levels of lactate dehydrogenase (LDH) and creation kinase MB (CK-MB), increased the activities of catalase (CAT), superoxide dismutase (SOD) and glutathione peroxidase (GSH-PX), and decreased malondialdehyde (MDA) content. In addition, KXS diminished the levels of IL-6, TNF-a, and C-reactive protein (CRP) in myocardial tissue (Hu et al., 2020). Nevertheless, these experiments have obvious limitations in that only one or two doses of KXS were applied, and thus the information on the dose-dependent effect is restricted. As a classical prescription for the treatment of depression, KXS regulates prominent

components of gut and brain functions (Cao et al., 2020). The anti-depression molecular mechanisms of KXS remain to be elucidated. The regulatory effects of KXS by which the microbiota-gut-brain axis affects depression are worth studying.

3.3 Effect on multi-infarct dementia

Multi-infarct dementia (MID), which is characterized by neuronal degeneration and multiple lesions in the cerebral graywhite matter, is the most common type of vascular dementia (VD), accompanied by obvious brain tissue pathological damage and cognitive dysfunction (Jellinger, 2013). Accumulated excitatory amino acid (EAA) and flawed energy metabolism in the brain are usually considered two major pathological hallmarks in the progression of MID (Popa-Wagner et al., 2015). Compared with the model group, pretreated with oral KXS water extract (2.12, 1.06 g/kg, 0.7 mg/kg hydergine was used as positive control) for 45 days

TABLE 2 Pharmacological effects of KXS.

Pharmacological activity	Model	Effective dose	Route	Positive control	Effects	Mechanisms	Application	References
Anti-AD effects	Male SD rats AD model; PC12 cells	10 g/kg	p.o.	Huperzine A	Regulate the levels of central	Ach ↑	In vivo and in vitro	Guo et al. (2019c)
				30 ug/kg	neurotransmitters, reducing the neuroinflammation and ROS-induced	GABA ↑		
					neuronal apoptosis via PI3K/Akt/GSK-3β pathways.	DA ↑		
						5-HT ↑	-	
						5-HIAA ↑		
						р-РІЗК ↑		
						p-Akt ↑		
						p-GSK-3β↑		
						p-Tau↓	-	
						ROS ↓		
						IL-1β ↓		
						TNF-α↓		
						AA ↓		
						Bax ↓		
						Bcl-2 ↑		
						Caspase-3 ↓		
	Kun-Ming mice AD model	1.4, 2.8 g/kg	p.o.	Donepezil 3 mg/kg	Inhibit the inflammation of microglia, alleviate cholinergic system dysfunction.	IL-1 $\beta \downarrow$	In vivo	Luo et al. (2020)
						TNF-α ↓		
						IL-6 ↓		
						CHRNB2 ↑		
	Male SAMP8 mice	5.4 g/kg	p.o.		Attenuate Tau hyperphosphorylation and neuroinflammation by inhibiting the TLR4/	Tau phosphorylation \downarrow	In vivo	Jiao et al. (2022)
					MyD88/NF-KB signaling pathway, therefore suppressing neuronal apoptosis and	p-GSK3β (Tyr216)/ GSK3β↓		
					improving the cognitive function.	p-GSK3β (Ser9)/ GSK3β ↑		
						p-AKT (\$473)/AKT ↑		
						TLR4 ↓		
						MyD88 ↓		

Pharmacological activity	Model	Effective dose	Route	Positive control	Effects	Mechanisms	Application	References
						NF-κB ↓		
						Bax ↓	-	
						Bcl-2 ↑	-	
						Caspase 1 ↓	-	
						Caspase 3 ↓	-	
						MMP ↑		
						ATP ↑		
						EC ↑		
						Glutamate ↓		
						p-NMDAR1 ↓		
						Ptch1 ↑		
						Smo ↑	_	
						Gli1 ↑		
The effects on memory and cognitive functions	Male Kunming mice model of scopolamine-induced cognitive dysfunction	0.7, 1.4, 2.8 g/kg I	p.o.	Donepezil 3 mg/kg	Improve cognitive function through reducing apoptosis and oxidative stress, and regulating	Bcl-2 ↑	In vivo	Xu et al. (2019)
					synapse-associated protein and cholinergic neurotransmitters.	Bax ↓		
					iterioralisinteris.	PSD 95 ↑		
						SYN ↑		
						BDNF ↑		
						ChAT ↑		
						ACh ↑	_	
						AChE ↓	_	
						SOD ↑	-	
						GSH-Px ↑		
						ROS ↓		
						MDA ↓		
	Male SD rat model of PSD-induced cognitive deficit	125, 250, 500 mg/kg	p.o.	Modafinil 30 mg/kg	Improve cognitive function by modulation of neurotransmitter levels and the expression of	GLU ↓	In vivo	Hu et al. (2013)
				00	some genes in the brain that contribute to memory functions.	GABA↓		

(Continued on following page)

Pharmacological activity	Model	Effective dose	Route	Positive control	Effects	Mechanisms	Application	References
						BDNF ↑		
						CREB ↑		
						p-CREB ↑		
	Male SD rat model of simulated	0.3, 0.6 g/kg	p.o.	Huperzine A,	Improve memory deficiency by inhibiting	ROS ↓	In vivo	Qiong et al.
	weightless-ness			0.1 mg/kg	oxidative stress damage	8-OHdG ↓		(2016)
						3-NT ↓		
	Female BALB/c mice model of	1 g/kg	p.o.	Doxorubicin,	Regulate inflammatory responses and reduce	IL-6 ↓	In vivo	Lyu et al. (2021)
	chemotherapy-induced cognitive impairment			5 mg/kg	oxidative stress and neural degeneration.	IL-12p70 ↓	-	
						IL-1β ↓		
						TFN-α ↓		
						IL-4 ↑		
						IL-10 ↑		
						MDA ↓		
						SOD ↑		
						GPx ↑		
						CAT ↑		
						GSH ↑		
Anti-depression effects	Male Wistar rats model of CUMS- induced depression; LPS- induced rat	3, 5, 10 g/kg	p.o.	Fluoxetine, 10	Ameliorate depressive behaviors and inhibited the NLRP3 inflammasome-mediated	IL-6 ↓	In vivo and in vitro	Yu et al. (2021)
	astrocytes			mg/kg	inflammation in vivo and in vitro by inducing	IL-1β↓		
					autophagy.	TNF-α ↓		
						NLRP3 ↓		
						ASC ↓		
						Caspase-1 ↓		
						MDA ↓		
						SOD ↑		
						GSH ↑		
						LC3-II ↑		

(Continued on following page)

10.3389/fphar.2024.1338024

Frontiers in Pharmacology

Pharmacological activity	Model	Effective dose	Route	Positive control	Effects	Mechanisms	Application	References
						Р62↓		
						Beclin1 ↑		
						ROS ↓		
	Male ICR mice model of CUMS-induced	3, 10 g/kg	p.o.	Fluoxetine	Modulate gut micro-environment	Allobaculum ↑	In vivo	Cao et al. (2020)
	depression			4 mg/kg	modification, suppress neuronal inflammation in the brain and inhibition of	Bifidobacterium ↑		
					HPA axis activation.	Turicibacter ↑		
						Coprococcus ↓		
						Mucispirillum ↓		
						Odoribacter ↓		
						Oscillospira ↓		
						IL-6 ↓		
						IL-1 $\beta \downarrow$		
						TNF-α ↓		
						ZO-1 ↑		
						Occludin ↑		
						Claudin-5 ↑		
						ACTH \downarrow		
						Corticosterone \downarrow		
	Male rat model of depression with MI	370 mg/kg	p.o.		Possesse both antidepressive effects and cardioprotective functions	CK-MB ↓	In vivo	Hu et al. (2020)
					cardioprotective functions	LDH ↓		
						SOD ↑	_	
						GSH-PX ↑		
						CAT ↑		
						MDA ↓		
	Male KM mice model of depression	175, 350, 700, 1,400 mg/kg	p.o.	Fluoxetine 28 mg/kg	Regulate the central monoaminergic neurotransmitter system	5-HT ↑	In vivo	Zhou et al. (2012)
		1,700 IIIg/Kg		20 mg/kg	neurorransmitter system	DA ↑		

(Continued on following page)

17

Pharmacological activity	Model	Effective dose	Route	Positive control	Effects	Mechanisms	Application	References
						NE ↑		
	Male ICR mice model of CUMS-induced	1.5 g/kg	p.o.	Fluoxetine	Regulate neurotrophin-Trk signaling pathway	NGF ↑	In vivo	Zhu et al. (2017)
	depression			4 mg/kg		BDNF ↑		
						Trk receptors ↑		
	Male wistar rat model of CUMS-induced	1.5, 5 g/kg	p.o.	Fluoxetine	Beneficial for synaptogenesis by inducing	Synapsin II ↑	In vivo and in vitro	Zhu et al. (2016)
	depression; primary cultures of rat hippocampal neuron			3.6 mg/kg	synaptic protein expressions via cAMP- dependent signaling pathway.	Synaptotagmin ↑		
						PSD95 ↑	-	
						Bt2-cAMP ↑		
	Male ICR mice model of CUMS-induced depression; Mice microglial cell	3, 10 g/kg	p.o.	Fluoxetine 7.2 mg/kg	Decrease pro-inflammatory cytokine expression in microglia via inhibiting TLR4/ IKK/NF-KB pathways.	IL-1 $\beta \downarrow$	In vivo and in vitro	Qu et al. (2021)
	lines (BV2)					IL−2 ↓	_	
						TNF-α ↓		
						TLR4 \downarrow		
						p-NF-KB/NF-KB↓		
						p-IKK/IKK ↓		
	Mouse astrocytes	1–10 ug/mL			Increase expressions of synthesis kinase tPA in neurotrophic factor metabolic pathway	NGF ↑	in vitro	Cao et al. (2018)
					neurorrophic factor inclabolic pathway	BDNF ↑		
						Bt2-cAMP ↑		
						p-CREB ↑		
Anti-fatigue effects	Male SD rats model of fatigue induced by forced wheel running	125, 250, 500 mg/kg	p.o.	Modafinil, 13 mg/kg	Increase exhaustive running time in the treadmill running test	Hepatic/muscle glycogen ↑	In vivo	Cao et al. (2012)
						Testosterone ↑		
						ldh ↓		
						SUN ↓	-	
						BLA ↓		
						β -endorphin \downarrow		
						SOD ↑		
						MDA ↓		

significantly improved cognitive impairment and hippocampal neuron damage of MID rats, rescued mitochondrial functions by upregulating brain energy, ameliorating mitochondrial swelling as well as improving mitochondrial membrane potential under chronic hypoperfusion conditions, suppressed glutamate neurotoxicity through decreasing the concentration of glutamate and the level of p-NMDAR1, and noticeably activated Shh/Ptch1 signaling pathway in the brain tissue of MID rats. Intriguingly, KXS protected PC12 cells against glutamate-induced neurotoxicity via Shh/Ptch1 signaling pathway (Li et al., 2021b). Nevertheless, further in-depth clinical trials should be conducted to investigate the effectiveness of KXS as a treatment for MID.

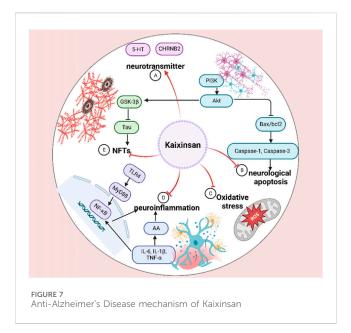
3.4 The effects on cognitive functions

Cognitive dysfunction (CD) is characterized by abnormalities in reduced memory, learning, calculation, attention, comprehension, executive functioning, and psychomotor speed (Knight and Baune, 2018; Xie et al., 2020), along with abnormal mental behaviors, which interfere with patients' daily performance and social abilities (Moussa-Pacha et al., 2020). It is well known that the Morris water maze (MWM) and Y-maze tasks are behavioral experiments designed to determine the spatial memory and learning functions (Wang et al., 2021d).

Appropriate animal models could serve as useful tools for experimental studies of drugs (Zhou et al., 2022b). Scopolamine is a pharmacologic agent that causes serious damage to cognition and functional imaging in various kinds of animals, including rats, mice, dogs, and cats (Klinkenberg and Blokland, 2010), similar to what is seen in cognitive dysfunction in humans. A substantial number of preclinical studies have employed scopolamine as a standard drug for inducing cognitive impairment models in healthy animals (Deng et al., 2019; Amirahmadi et al., 2022). Sleep deficiency or loss leads to the decline of emotion, immunity, memory, and learning (Krone et al., 2021). The rodent models of sleep deprivation show anxiety-like behaviors and cognitive impairment similar to humans (Yin et al., 2017). It is well known that cancer-related disruptions in cognitive function and memory are a common adverse effect of adjuvant chemotherapy (Speidell et al., 2018). Clinical and preclinical studies have indicated that KXS exerts a beneficial effect on CD in different animal models. In twelve scopolamine-induced CD mice, oral administration of 0.7, 1.4, and 2.8 g/kg KXS water extract (3 mg/kg/d of donepezil was used as positive control) for 2 weeks ameliorated learning and memory impairments (shortened the escape latency and increased residence time in target quadrant and the number of target crossings in MWM test, increased the percentage of alternations between the labyrinth arms in the Y-maze), decreased neuronal apoptosis and reversed neuronal degeneration (inhibited the formation of apoptotic bodies and expression of proapoptotic protein Bax, and upregulated the expression of antiapoptotic protein Bcl-2), regulated cholinergic neurotransmitters (increased acetylcholine levels and the activity of choline acetyltransferase), and alleviated oxidative stress damage in the cortex and hippocampus (increased the level and activity of superoxide dismutase and glutathione peroxidase, reduced the level and activity of reactive oxygen species and malondialdehyde) (Xu et al., 2019). Treatment with oral KXS water extract (at daily doses of 500, 250, and 125 mg/kg body weight, 30 mg/kg modafinil was used as positive control) for 14 days improved learning and memory function of paradoxical sleep deprivation (PSD)-induced CD rats by decreasing the glutamic acid (GLU) and y-aminobutyric acid (GABA) levels and modulating of the expression of some genes in the brain tissue, such as BDNF, cyclic AMP response element binding protein (CREB), and phosphorylated-CREB (p-CREB). The minimal active dose was 250 mg/kg, while the memory enhancement effect of 500 mg/kg KXS was equally effective as that of 30 mg/kg modafinil (Hu et al., 2013). Another study (0.1 mg/kg of huperzine A was used as positive control) showed that intragastric administration of 0.3 or 0.6 g/kg of KXS water extract for 2 weeks improved memory deficit induced by simulated weightlessness in ten rats mainly by inhibiting oxidative stress injury (Qiong et al., 2016). Chemotherapy-related cognitive impairment has been observed in 15% and 50% of oncology patients (Rummel et al., 2021). The therapeutic effect of KXS (at the dosage of 1 g/kg daily by gavage over 3 weeks) was studied on doxorubicin (DOX)-related cognitive impairment in 4T1 breast cancer mice. This study suggested that KXS attenuated DOX-induced cognitive impairment by regulating inflammatory responses, decreasing oxidative stress, and reversing neural degeneration. Additionally, it is worthy that KXS is beneficial to enhance quality of life and prolong survival time in DOX-treated tumor-bearing mice (Lyu et al., 2021). In summary, KXS is a promising prescription for the treatment of memory decline. These results gave inspiration for the future development of commercial agents. Nonetheless, studies investigating the clinical pharmacokinetic parameters and safety evaluation of KXS are scarce, which limits further understanding of the medicinal substance of KXS.

3.5 Anti-fatigue effects

Chronic fatigue syndrome (CFS) is a complex disease characterized by extreme fatigue with associated symptoms of anxiety, depression, pain, and various somatic complaints (Zhang et al., 2022b). The forced wheel running model is a popular model that has been used to simulate the chronic fatigue syndrome (Toval et al., 2021). Cao et al. evaluated the effects of KXS on CFS mice induced by forced wheel running. Briefly, 72 male Kunming mice were randomly separated into six groups (n = 12), including a control group, CFS group, Modafinil-treated CFS group (13 mg/kg/ day), and three KXS-treated groups (175, 350 and 700 mg/kg/day). The mice in the control group were not forced to run. Correspondingly, the animals in the other five groups were induced by forced wheel running for 4 weeks. All the mice were anesthetized by intraperitoneal injections of ketamine (80 mg/kg) and xylazine (4 mg/kg). Blood was taken using a disposable syringe from the heart. The liver and muscle of mice had been weighed precisely as instructions. Levels of serum urea nitrogen (SUN), serum lactate dehydrogenase (LDH), liver glycogen (LG), muscle lactic acid (MLA), muscle glycogen (MG), and the expressions of IL-2, and IL-4 secreted by splenocytes were detected. The results showed that electric shock time was prominently reduced in the three KXS-treated groups when compared with the CFS group without drug treatment. Interestingly, the reduction in MLA,



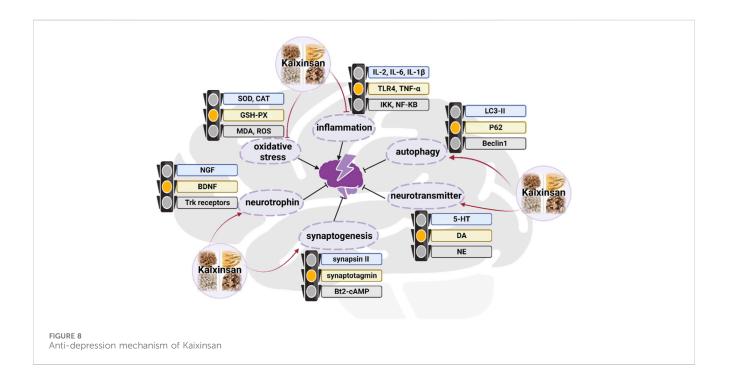
SUN, LDH and IL-4 levels and an increase in LG, MG, MLA, and IL-2 levels were demonstrated in these three KXS-treated groups than those in the CFS group without drug treatment (Cao et al., 2012). This elucidates that KXS exhibits comprehensive anti-fatigue effects. However, the specific mechanisms remain unclear, and further investigation is required to perform at animal, cellular, and molecular levels.

4 Pharmacokinetic studies

Pharmacokinetics (PK) investigation mainly focuses on the changes in the process of medicine absorption, distribution,

metabolism, and excretion (ADME) processes in vivo after oral administration, and provides a helpful approach to confirm the effective ingredients and guide the clinical usage of herbal medicine (Tang et al., 2022). There are few investigations on the PK of whole KXS extracts. Currently, efficient technologies employed in the PK study of KXS are LC-MS/MS, Q-TOF/MS, HPLC-MS/MS, and UPLC-MS/MS, which are used for the determination of rat samples. The parameters are summarized as follows. In preclinical experiments, rats were given oral KXS (10 g/kg), and there were 26 prototype components (ginsenosides, polygala saponins, oligosaccharide esters, terpenoids, sucrose ester, xanthone, and other compounds) in the plasma of rats were assigned for identification using UHPLC-Q-TOF-MS. These components were derived from PR, GR, and Poria (Zhang et al., 2015). Wang et al. (2017a) obtained systematic PK data concerning the activity of KXS in the context of AD by Ultra-fast liquid chromatography-MS/MS. In normal control group, the Cmax of POL, SB, Te, A5, A6, tenuifoliside A (TenA), GRe and GRb1 were 51.57 ± 19.10, 75.30 ± 9.10, 133.9 ± 25.3, 155.4 ± 30.8, 129.2 ± 22.9, 72.94 ± 35.17, 75.47 ± 17.2, and 269.3 ± 12.3 ng/ mL, respectively. The T_{max} were 0.36 \pm 0.12, 0.28 \pm 0.17, 0.39 \pm 0.09, 0.21 ± 0.15 , 0.28 ± 0.17 , 0.15 ± 0.10 , 0.40 ± 0.20 , 0.50 ± 0.28 h, respectively. And the area under the curve (AUC 0- ∞) were 198.2 ± $52.7, 159.2 \pm 58.1, 692.2 \pm 322.7, 270.4 \pm 69.1, 350.6 \pm 103.3, 111.4 \pm$ 44.1, 297.5 ± 42.2, and 4172 ± 902 ng h/mL, respectively. Compared with the normal group, the PK behaviors of SB, Te, and GRe in the AD model significantly illustrated a trend of continuous changes, such as higher bioavailability, better assimilation effect, slower elimination rate, delays in reaching the Cmax and longer mean dwell time.

The basic feature of TCM is that it consists in the form of preparation and formulation by the guidance of compatibility rules, different herbs are usually orchestrated to form a multi-herbs combination. A classical prescription generally contains primary



20

(called "Jun" or "Emperor") and adjuvant herbs (called "Chen, Zuo, and Shi") according to the therapeutic characteristics of TCM, and the latter function as "Assistant", "Vassal" and "Delivery servant" which are generally worked as assistants to increase efficiency, enhance the synergistic effect or to facilitate the delivery of the emperor medicine (Wang et al., 2021a). In the TCM system, the incorporation of a "Delivery servant" into classical prescription is an important compatibility mechanism to support the transportation from the effective constituents to the target organ in treating diseases (Wei et al., 2018). ATR is assumed to act as a "delivering servant" in a multi-herb classical prescription for treatment of the central nervous disease, capable of facilitating the delivery of bioactive ingredients to the brain. KXS is a classical prescription with various PK interactions among its multi-constituents. In the case of KXS, a combination of PR, Poria, and ATR has synergistic effects on enhancement to the nootropic effects of GR. In 2010, Wang et al. investigated the PK profiles and brain distribution of ginsenosides Re, Rb1, and Rg1 after oral administration of KXS with or without ATR to rats. The result indicated that ATR in KXS increased the brain concentrations and bioavailabilities of ginsenosides Re and Rg1, but unlikely influenced the brain-to-plasma AUC ratios. ATR, the presence of the delivering servant, appeared to promote the initial absorption rate of ginsenoside in the gastrointestinal tract (Wang et al., 2010). Similar phenomena were found in the study of investigating the differences of uptaken of ginsenoside and triterpenes after oral administration of single herb and drug pairs in KXS by PK studies in vitro Caco-2 cell monolayers model (Zheng et al., 2018).

5 Clinical applications

According to TCM, depression is known as "Yu Zheng" which is manifested as a mental illness induced by insufficiency of xin-qi, stagnation of vital energy, exuberance of dampness, and phlegm in the body (Zhang and Cheng, 2019). KXS is one of the classical prescriptions for the treatment of depression. The prescription nourishes the heart, tonifies xin-qi, eliminates dampness, and tranquilizes the spirit (Yu et al., 2021). In the clinic, KXS intervention (3.2 g/d) was significantly successful in improving depressive symptoms and cognitive dysfunction in a randomized, double-blind, placebo-controlled trial involving 134 patients diagnosed with depression (Hu et al., 2021a). In addition, Hu et al. (2021b) conducted a clinical observation of 156 patients to investigate the therapeutic impact of KXS (3.2 g/d) combined with fluoxetine (20 mg/d) in the treatment of depression. Combining KXS drastically ameliorated the Hamilton Depression 17 (HAM-D17) rating scores and self-rating depression scale scores, and the serum lipoprotein B, apolipoprotein C3 and albumin levels and lowdensity lipoprotein cholesterol/high-density lipoprotein cholesterol ratio (Hu et al., 2021b).

6 Safety evaluation

Notably, rare investigations have reported the safety evaluation of KXS in preclinical and clinical practice. One report in 2011 indicated that at doses consumed in traditional medicine, KXS may be relatively safe (Mu et al., 2011). To evaluate the preclinical safety of KXS, mice received oral administration of single doses (19.67, 24.59, 30.74, 38.42, 48.03 and 60.04 g/kg, 0.8 mL/mice/time, twice/day) of the water extracts for 7 days in a study on acute toxicity, rats were given oral extracts (1, 3 and 9 g/kg, respectively) for 91 days in a study on sub-chronic toxicity. The study found that the oral median lethal dose (LD50) was 32.59 g/kg, and the mortality rate and adverse effects were clearly dose-dependent in female and male mice. In addition, the no-observed-adverse-effect level (NOAEL) of the extract was 19.67 g/kg for both sexes. No treatment-related clinical signs, mortalities, body weights, food consumption, blood chemistry, and organ parameters (brain, kidney, spleen, adrenal glands, testis, and epididymis) were observed during the sub-chronic toxicity study (Mu et al., 2011).

However, the assessment of the safety of whole classical prescription also depends on the presence of herbs with known toxicity. One of the herbs in KXS that requires monitoring is PR. PR can cause mild nausea and is regarded as a gastrointestinal side effect in routine clinical usage (Pharmacopoeia Commission of the People's Republic of China, 2020). Long-term and excessive doses of raw PR can severely suppress gastrointestinal movement, damage the stomach and small intestine, cause intestinal wall thinning, and damage the gastric wall cell structure (Wang et al., 2018). One study showed that the use of a high dose of onjisaponin B (200 mg/kg via oral administration) which is derived from PR, can induce gastrointestinal congestion and swelling in mice. And the gastrointestinal irritation was decreased when PR was fried with honey (Wen et al., 2015). It is worth noting that the existing studies have indicated that ATR-derived volatile oil could noticeably depress the pulse frequency of cardiac myocytes, and slow down the heart rate at the therapeutic dose (Wu et al., 2009). In addition, α -asarone and β -asarone derived from ATR may have hepatotoxicity, genotoxicity, and carcinogenicity (Chellian et al., 2017). Although this does not indicate that KXS might lead to this toxicity, further preclinical and clinical evidence should be required to confirm the safety through long-term monitoring of the undesired effects of KXS.

The remaining herbs including GR and Poria which have been used both as food and medicine, are generally acknowledged as safe for nutrient supplements and treatment of diseases (Lu et al., 2021a; Li et al., 2022). Nevertheless, the clinical efficacy and safety of herbal medicine are also susceptible to being influenced by germplasm resources and growth conditions (Ren et al., 2020). Pesticide residues in herbal medicine are one of the most important issues affecting safety (Yue et al., 2022). More attention should be paid to the pesticide residues that may accumulate with long-term consumption of KSX.

7 Discussion and conclusion

Classical prescriptions are characterized by significant advantages of their high efficiency, low toxicity, minor side effects, and economic benefits (Liu et al., 2023). KXS, as an outstanding classical prescription applied for the treatment of dementia and morbid forgetfulness, has attracted substantial research interest in China and other countries in East Asia (Shan et al., 2023b). In this review, we systematically summarized the recent advances of KXS on chemical constituents, pharmacological effects, pharmacokinetics, clinical studies, and safety. Previous studies have shown that KXS is a multi-constituent, multi-target and multi-pathway classical prescription with neuroprotective, anti-AD, anti-depressant and anti-fatigue effects. It is commonly used clinically for the treatment of depression and memory decline (Qiong et al., 2016). The PK-based determination of these exposure constituents, together with metabolites after KXS administration will facilitate uncovering bioactive ingredients responsible for the therapeutic action of classical prescription (Zhang et al., 2015). All these investigations have constructed a strong scientific foundation for further research on KXS. However, the following issues still exist and remain to be further studied.

First, although many studies have demonstrated its broad therapeutic potential, the medicinal substances in the classical prescription are incompletely characterized, and the study is limited to the research on some specific and representative chemical ingredients as well as their structure-activity relationships with these bioactive ingredients. For example, the saponins from PR, the ginsenosides from GR, the phenolic acids from Poria, and the volatile oils from ATR (Li et al., 2018b). However, for some components with low contents and the bioactive compositions with high molecular weight, there are fewer studies on KXS, such as the sterols from Poria, the phenylpropanoids from ATR, as well as the polysaccharides from Poria and GR.

Second, for pharmacokinetic studies. In addition to the general analysis of ADME, numerous gut microbial play an important role in herb-taking effects (Gong et al., 2020). Yet the interaction between gut microbiota in the host and bioactive ingredients of KXS remains unclear.

Third, For quality control it is known that classical prescription is notable for its comprehensive medical effects with various complex chemical components (Li et al., 2018). Therefore, discovering and confirming the quality control markers that could reflect the overall efficacy of KXS is another important problem requiring further study.

Fourth, For pharmacological studies. Due to its unique multicomponent and multi-target effect of classical prescription, the detailed molecular mechanism is absent or incomplete in most of the pharmacological studies as well as the following validation experiments. And thus in the future, the application of new technologies and research strategies should be carried out. In addition, the current pharmacology studies on quality controls including animal grouping, administration dosage, and positive drug are not strictly conducted.

In light of the question listed above, some solutions are proposed to solve these problems in the study of this classical prescription.

- (I) Comprehensive chemical ingredients analysis for KXS should be further studied. More efficient and highquality analytical techniques such as high-resolution mass spectrometry techniques, multi-dimensional chromatography, and fingerprint analysis should be applied to clarify the structure-activity relationships.
- (II) KXS can replenish Qi and nourish the heart. Interestingly, the functions of the brain and gut are respectively attributed

to the functions of the heart and spleen in TCM (Zhang et al., 2022a). Thus, the interaction of gut microbial and bioactive ingredients of this prescription is a promising and worthy focus.

- (III) It is necessary to analyze the mechanisms of KXS and enhance the quality control markers based on the bioactive ingredients from this classical prescription. Therefore, novel methods and ideas, such as highthroughput screening in conjunction with clinical outcomes, experimental validations after the network pharmacology definition, and metabolomics analysis should be adapted to further research. It helps to analyze the complex mechanism of action of KXS in various diseases and expand its indications.
- (IV) It is worth noting that the kinds of major active components and their contents vary based on the habitat of medicinal plants. The quality of medicinal plants plays an essential role in ensuring the clinical efficacy and safety of classical prescription. Thus, it is necessary to apply genuine Chinese medicine and/or the medicinal plant cultivated in the main regions to control the quality of KXS. Moreover, the development of robust quality control strategies should be applied to the overall evaluation of KXS quality.

Taken together, the present review has concluded with a summary and insight into the basic information about KXS, which might provide a scientific basis for the related research of KXS. Additionally, this paper contains a brief introduction about the potential value and some new research directions on KXS, thereby providing new research ideas for better exploration and exploitation in the future.

Author contributions

LC: Conceptualization, Funding acquisition, Writing-original draft, Writing-review and editing. LJ: Resources, Software, Writing-review and editing. XS: Data curation, Validation, Writing-original draft. JY: Investigation, Resources, Writing-review and editing. RW: Funding acquisition, Project administration, Supervision, Writing-original draft. WL: Project administration, Supervision, Writing-review and editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Gansu Provincial Science and Technology Project (23ZDFA013-5), China; Innovation Team Project of Lanzhou Science and Technology Bureau (No. 2023-2-62), China; Fund Project of the 940th Hospital (No. 2021yxky043, No. 2023YXKY016), China.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Amirahmadi, S., Farimani, F. D., Akbarian, M., Mirzavi, F., Eshaghi Ghalibaf, M. H., Rajabian, A., et al. (2022). Minocycline attenuates cholinergic dysfunction and neuroinflammation-mediated cognitive impairment in scopolamine-induced Alzheimer's rat model. *Inflammopharmacology* 30 (6), 2385–2397. doi:10.1007/s10787-022-01071-2

Andrea, P., Martina, H., Lucie, H., Veronika, S., Barbora, N., Helena, P., et al. (2022). Age-related metabolic and neurodegenerative changes in SAMP8 mice. *Aging* (Albany NY). 14 (18), 7300–7327. doi:10.18632/aging.204284

Antoniuk, S., Bijata, M., Ponimaskin, E., and Wlodarczyk, J. (2019). Chronic unpredictable mild stress for modeling depression in rodents: meta-analysis of model reliability. *Neurosci. Biobehav Rev.* 99, 101–116. doi:10.1016/j.neubiorev.2018.12.002

Bo, M., Zhang, H., Xu, J., Zhao, H., Jia, X., Wang, G., et al. (2022). Systematic review of Kaixinsan in treating depression: efficacy and pharmacological mechanisms. *Front. Behav. Neurosci.* 16, 1061877. doi:10.3389/fnbeh.2022.1061877

Bucciarelli, V., Caterino, A. L., Bianco, F., Caputi, C. G., Salerni, S., Sciomer, S., et al. (2020). Depression and cardiovascular disease: the deep blue sea of women's heart. *Trends Cardiovasc. Med.* 30 (3), 170–176. doi:10.1016/j.tcm.2019.05.001

Cao, C., Liu, M., Qu, S., Huang, R., Qi, M., Zhu, Z., et al. (2020). Chinese medicine formula Kai-Xin-San ameliorates depression-like behaviours in chronic unpredictable mild stressed mice by regulating gut microbiota-inflammation-stress system. *J. Ethnopharmacol.* 261, 113055. doi:10.1016/j.jep.2020.113055

Cao, C., Xiao, J., Liu, M., Ge, Z., Huang, R., Qi, M., et al. (2018). Active components, derived from Kai-xin-san, a herbal formula, increase the expressions of neurotrophic factor NGF and BDNF on mouse astrocyte primary cultures via cAMP-dependent signaling pathway. *J. Ethnopharmacol.* 224, 554–562. doi:10.1016/j.jep.2018.06.007

Cao, Y., Hu, Y., Liu, P., Zhao, H.-X., Zhou, X.-J., and Wei, Y.-M. (2012). Effects of a Chinese traditional formula Kai Xin San (KXS) on chronic fatigue syndrome mice induced by forced wheel running. *J. Ethnopharmacol.* 139 (1), 19–25. doi:10.1016/j.jep. 2011.08.030

Carney, R. M., and Freedland, K. E. (2016). Depression and coronary heart disease. Nat. Rev. Cardiol. 14 (3), 145-155. doi:10.1038/nrcardio.2016.181

Chellian, R., Pandy, V., and Mohamed, Z. (2017). Pharmacology and toxicology of α and β -Asarone: a review of preclinical evidence. *Phytomedicine* 32, 41–58. doi:10.1016/j. phymed.2017.04.003

Chen, S., Liang, A., and Zhang, B. (2018). Letter to the editor: comment on the regulatios of the simplified registration and approval management for compound recipe of classical prescription of traditional Chinese medicine. *Acta Pharm. Sin. B* 8 (1), 127–129. doi:10.1016/j.apsb.2017.12.003

Chen, Z., Yang, Y., Han, Y., and Wang, X. (2022). Neuroprotective effects and mechanisms of senegenin, an effective compound originated from the roots of polygala tenuifolia. *Front. Pharmacol.* 13, 937333. doi:10.3389/fphar.2022.937333

Chou, Y. H., Ton That, V., and Sundman, M. (2020). A systematic review and metaanalysis of rTMS effects on cognitive enhancement in mild cognitive impairment and Alzheimer's disease. *Neurobiol. Aging* 86, 1–10. doi:10.1016/j.neurobiolaging.2019. 08.020

Cladder-Micus, M. B., Speckens, A. E. M., Vrijsen, J. N., Ar, T. D., Becker, E. S., and Spijker, J. (2018). Mindfulness-based cognitive therapy for patients with chronic, treatment-resistant depression: a pragmatic randomized controlled trial. *Depress Anxiety* 35 (10), 914–924. doi:10.1002/da.22788

Conti Filho, C. E., Loss, L. B., Marcolongo-Pereira, C., Rossoni Junior, J. V., Barcelos, R. M., Chiarelli-Neto, O., et al. (2023). Advances in Alzheimer's disease's pharmacological treatment. *Front. Pharmacol.* 14, 1101452. doi:10.3389/fphar.2023.1101452

Cruz-Pereira, J. S., Rea, K., Nolan, Y. M., O'Leary, O. F., Dinan, T. G., and Cryan, J. F. (2020). Depression's unholy trinity: dysregulated stress, immunity, and the microbiome. *Annu. Rev. Psychol.* 71 (1), 49–78. doi:10.1146/annurev-psych-122216-011613

Deng, G., Wu, C., Rong, X., Li, S., Ju, Z., Wang, Y., et al. (2019). Ameliorative effect of deoxyvasicine on scopolamine-induced cognitive dysfunction by restoration of cholinergic function in mice. *Phytomedicine* 63, 153007. doi:10.1016/j.phymed.2019. 153007

Dong, X. Z., Wang, D. X., Lu, Y. P., Yuan, S., Liu, P., and Hu, Y. (2017). Antidepressant effects of Kai-Xin-San in fluoxetine-resistant depression rats. *Braz J. Med. Biol. Res.* 50 (10), e6161. doi:10.1590/1414-431X20176161

Du, L. J., Zhang, X. N., Li, S. S., Sun, Y. F., Jin, H. Z., Yan, S. K., et al. (2022). Network pharmacological investigation into the mechanism of Kaixinsan powder for the treatment of depression. *Metab. Brain Dis.* 37 (8), 2903–2914. doi:10.1007/s11011-022-01067-5

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Fox, M. E., and Lobo, M. K. (2019). The molecular and cellular mechanisms of depression: a focus on reward circuitry. *Mol. Psychiatry* 24 (12), 1798–1815. doi:10. 1038/s41380-019-0415-3

Fu, H., Xu, Z., Zhang, X. L., and Zheng, G. Q. (2019). Kaixinsan, a well-known Chinese herbal prescription, for alzheimer's disease and depression: a preclinical systematic review. *Front. Neurosci.* 13, 1421. doi:10.3389/fnins.2019.01421

Gong, X., Li, X., Bo, A., Shi, R.-Y., Li, Q.-Y., Lei, L.-J., et al. (2020). The interactions between gut microbiota and bioactive ingredients of traditional Chinese medicines: a review. *Pharmacol. Res.* 157, 104824. doi:10.1016/j.phrs.2020.104824

Guo, S., Wang, J., Wang, Y., Zhang, Y., Bi, K., Zhang, Z., et al. (2019a). Study on the multitarget synergistic effects of kai-xin-san against alzheimer's disease based on systems biology. *Oxid. Med. Cell Longev.* 2019, 1707218. doi:10.1155/2019/1707218

Guo, S., Wang, J., Wang, Y., Zhang, Y., Bi, K., Zhang, Z., et al. (2019b). Study on the multitarget synergistic effects of kai-xin-san against alzheimer's disease based on systems biology. Oxidative Med. Cell. Longev. 2019, 1707218–1707315. doi:10.1155/2019/1707218

Guo, S., Wang, J., Xu, H., Rong, W., Gao, C., Yuan, Z., et al. (2019c). Classic prescription, kai-xin-san, ameliorates alzheimer's disease as an effective multitarget treatment: from neurotransmitter to protein signaling pathway. *Oxid. Med. Cell Longev.* 2019, 9096409. doi:10.1155/2019/9096409

Hao, W., Chen, Z., Wang, L., Yuan, Q., Gao, C., Ma, M., et al. (2022). Classical prescription Huanglian Decoction relieves ulcerative colitis via maintaining intestinal barrier integrity and modulating gut microbiota. *Phytomedicine* 107, 154468. doi:10. 1016/j.phymed.2022.154468

Hu, Y., Chen, C., Wang, Y., Yang, W., Wang, Y., Zhu, W., et al. (2021a). The effects of KaiXinSan on depression and its association with lipid profiles: a randomized, doubleblinded, placebo-controlled trial. *Phytomedicine* 83, 153467. doi:10.1016/j.phymed. 2021.153467

Hu, Y., Liu, M., Liu, P., Yan, J. J., Liu, M. Y., Zhang, G. Q., et al. (2013). Effect of kai xin san on learning and memory in a rat model of paradoxical sleep deprivation. *J. Med. Food* 16 (4), 280–287. doi:10.1089/jmf.2012.2486

Hu, Y., Liu, X., Zhang, T., Chen, C., Dong, X., Can, Y., et al. (2020). Behavioral and biochemical effects of KXS on postmyocardial infarction depression. *Front. Pharmacol.* 11, 561817. doi:10.3389/fphar.2020.561817

Hu, Y., Wang, Y., Chen, C., Yang, W., Zhu, W., Wang, Y., et al. (2021b). A randomized, placebo-controlled, double-blind study on the effects of SZL on patients with mild to moderate depressive disorder with comparison to fluoxetine. *J. Ethnopharmacol.* 281, 114549. doi:10.1016/j.jep.2021.114549

Jellinger, K. A. (2013). Pathology and pathogenesis of vascular cognitive impairmenta critical update. *Front. Aging Neurosci.* 5, 17. doi:10.3389/fnagi.2013.00017

Jiang, N., Wei, S., Zhang, Y., He, W., Pei, H., Huang, H., et al. (2021). Protective effects and mechanism of radix polygalae against neurological diseases as well as effective substance. *Front. Psychiatry* 12, 688703. doi:10.3389/fpsyt.2021.688703

Jiao, Y. N., Zhang, J. S., Qiao, W. J., Tian, S. Y., Wang, Y. B., Wang, C. Y., et al. (2022). Kai-xin-San inhibits tau pathology and neuronal apoptosis in aged SAMP8 mice. *Mol. Neurobiol.* 59 (5), 3294–3309. doi:10.1007/s12035-021-02626-0

Khandaker, G. M., Zuber, V., Rees, J. M. B., Carvalho, L., Mason, A. M., Foley, C. N., et al. (2019). Shared mechanisms between coronary heart disease and depression: findings from a large UK general population-based cohort. *Mol. Psychiatry* 25 (7), 1477–1486. doi:10.1038/s41380-019-0395-3

Khayat, E., Lockhart, C., Delfing, B. M., Smith, A. K., and Klimov, D. K. (2021). Met35 oxidation hinders a β 25-35 peptide aggregation within the dimyristoylphosphatidylcholine bilayer. *ACS Chem. Neurosci.* 12 (17), 3225–3236. doi:10.1021/acschemneuro.1c00407

Klinkenberg, I., and Blokland, A. (2010). The validity of scopolamine as a pharmacological model for cognitive impairment: a review of animal behavioral studies. *Neurosci. Biobehav. Rev.* 34 (8), 1307–1350. doi:10.1016/j.neubiorev.2010.04.001

Knight, M. J., and Baune, B. T. (2018). Cognitive dysfunction in major depressive disorder. Curr. Opin. Psychiatry 31 (1), 26–31. doi:10.1097/YCO.00000000000378

Krone, L. B., Yamagata, T., Blanco-Duque, C., Guillaumin, M. C. C., Kahn, M. C., van der Vinne, V., et al. (2021). A role for the cortex in sleep–wake regulation. *Nat. Neurosci.* 24 (9), 1210–1215. doi:10.1038/s41593-021-00894-6

Li, B., Xu, M., Wang, Y., Feng, L., Xing, H., and Zhang, K. (2023). Gut microbiota: a new target for traditional Chinese medicine in the treatment of depression. *J. Ethnopharmacol.* 303, 116038. doi:10.1016/j.jep.2022.116038

Li, J., Huang, Q., Chen, J., Qi, H., Liu, J., Chen, Z., et al. (2021a). Neuroprotective potentials of Panax ginseng against alzheimer's disease: a review of preclinical and clinical evidences. *Front. Pharmacol.* 12, 688490. doi:10.3389/fphar.2021.688490

Li, L., Zuo, Z.-T., and Wang, Y.-Z. (2022). The traditional usages, chemical components and pharmacological activities of wolfiporia cocos: a review. *Am. J. Chin. Med.* 50 (02), 389-440. doi:10.1142/s0192415x22500161

Li, Q., Sun, Y., Guo, H., Sang, F., Ma, H., Peng, H., et al. (2018a). Quality control of the traditional Chinese medicine Ruyi jinhuang powder based on high-throughput sequencing and real-time PCR. *Sci. Rep.* 8 (1), 8261. doi:10.1038/s41598-018-26520-3

Li, X., He, Y., Zeng, P., Liu, Y., Zhang, M., Hao, C., et al. (2018b). Molecular basis for Poria cocos mushroom polysaccharide used as an antitumour drug in China. *J. Cell. Mol. Med.* 23 (1), 4–20. doi:10.1111/jcmm.13564

Li, X., Wen, W., Li, P., Fu, Y., Chen, H., Wang, F., et al. (2021b). Mitochondrial protection and against glutamate neurotoxicity via shh/ptch1 signaling pathway to ameliorate cognitive dysfunction by kaixin san in multi-infarct dementia rats. *Oxid. Med. Cell Longev.* 2021, 5590745. doi:10.1155/2021/5590745

Li, X., Yang, S., Wang, Y., Yang, B., and Zhang, J. (2020). Effects of a transtheoretical model - based intervention and motivational interviewing on the management of depression in hospitalized patients with coronary heart disease: a randomized controlled trial. *BMC Public Health* 20 (1), 420. doi:10.1186/s12889-020-08568-x

Lin, R., Yin, J., Wu, M., Ding, H., Han, L., Yang, W., et al. (2021). Global identification and determination of the major constituents in Kai-Xin-San by ultra-performance liquid chromatography-quadrupole-Orbitrap mass spectrometry and gas chromatography-mass spectrometry. *J. Pharm. Biomed. Analysis* 206, 114385. doi:10. 1016/j.jpba.2021.114385

Liu, C., and Ye, M. (2012). Characterization of chemical constituents and *in vivo* metabolites of Kai-Xin-San prescription by HPLC/DAD/ESI-MSn. *J. Chin. Pharm. Sci.* 21 (6). doi:10.5246/jcps.2012.06.072

Liu, H., Lu, X., Hu, Y., and Fan, X. (2020). Chemical constituents of Panax ginseng and Panax notoginseng explain why they differ in therapeutic efficacy. *Pharmacol. Res.* 161, 105263. doi:10.1016/j.phrs.2020.105263

Liu, J.-Y., Xiao, S.-Y., Shang, W.-F., Xu, L.-Z., and Yang, S.-L. (2005). A new minor triterpene saponin from kaixin-san prescription. *J. Asian Nat. Prod. Res.* 7 (4), 643–648. doi:10.1080/1028602032000169550

Liu, X., Wu, X., Wang, S., and Qin, X. (2023). Gut microbiome and tissue metabolomics reveal the compatibility effects of Xiaoyaosan on depression based on "gut-liver-kidney" axis. *Phytomedicine* 111, 154628. doi:10.1016/j.phymed.2022. 154628

Lu, G., Liu, Z., Wang, X., and Wang, C. (2021a). Recent advances in Panax ginseng C.A. Meyer as a herb for anti-fatigue: an effects and mechanisms review. *Foods* 10 (5), 1030. doi:10.3390/foods10051030

Lu, J., Wang, X., Wu, A., Cao, Y., Dai, X., Liang, Y., et al. (2022). Ginsenosides in central nervous system diseases: pharmacological actions, mechanisms, and therapeutics. *Phytotherapy Res.* 36 (4), 1523–1544. doi:10.1002/ptr.7395

Lu, J.-Z., Ye, D., and Ma, B.-L. (2021b). Constituents, pharmacokinetics, and pharmacology of gegen-qinlian decoction. *Front. Pharmacol.* 12, 668418. doi:10. 3389/fphar.2021.668418

Luo, Y., Li, D., Liao, Y., Cai, C., Wu, Q., Ke, H., et al. (2020). Systems pharmacology approach to investigate the mechanism of kai-xin-san in alzheimer's disease. *Front. Pharmacol.* 11, 381. doi:10.3389/fphar.2020.00381

Lv, C., He, B., Sui, Z., Li, Q., and Bi, K. (2016a). Identification and determination of the major constituents in Kai-Xin-San by UPLC-Q/TOF MS and UFLC-MS/MS method. *J. Mass Spectrom.* 51 (7), 479–490. doi:10.1002/jms.3773

Lv, C., He, B., Sui, Z., Li, Q., and Bi, K. (2016b). Identification and determination of the major constituents in Kai-Xin-San by UPLC-Q/TOF MS and UFLC-MS/MS method. *J. Mass Spectrom.* 51 (7), 479–490. doi:10.1002/jms.3773

Lyu, W., Ouyang, M., Ma, X., Han, T., Pi, D., and Qiu, S. (2021). Kai-xin-San attenuates doxorubicin-induced cognitive impairment by reducing inflammation, oxidative stress, and neural degeneration in 4T1 breast cancer mice. *Evid. Based Complement. Altern. Med.* 2021, 5521739. doi:10.1155/2021/5521739

McCarron, R. M., Shapiro, B., Rawles, J., and Luo, J. (2021). Depression. Ann. Intern. Med. 174 (5), ITC65–ITC80. doi:10.7326/aitc202105180

Monroe, S. M., and Harkness, K. L. (2022). Major depression and its recurrences: life course matters. *Annu. Rev. Clin. Psychol.* 18, 329–357. doi:10.1146/annurev-clinpsy-072220-021440

Moussa-Pacha, N. M., Abdin, S. M., Omar, H. A., Alniss, H., and Al-Tel, T. H. (2020). BACE1 inhibitors: current status and future directions in treating Alzheimer's disease. *Med. Res. Rev.* 40 (1), 339–384. doi:10.1002/med.21622

Mu, L.-H., Huang, Z.-X., Liu, P., Hu, Y., and Gao, Y. (2011). Acute and subchronic oral toxicity assessment of the herbal formula Kai-Xin-San. *J. Ethnopharmacol.* 138 (2), 351–357. doi:10.1016/j.jep.2011.08.033

Ong, W.-Y., Wu, Y.-J., Farooqui, T., and Farooqui, A. A. (2018). Qi Fu yin-a ming dynasty prescription for the treatment of dementia. *Mol. Neurobiol.* 55 (9), 7389–7400. doi:10.1007/s12035-018-0908-0

Pei, H., Ma, L., Cao, Y., Wang, F., Li, Z., Liu, N., et al. (2020). Traditional Chinese medicine for alzheimer's disease and other cognitive impairment: a review. *Am. J. Chin. Med.* 48 (03), 487–511. doi:10.1142/s0192415x20500251

Popa-Wagner, A., Buga, A. M., Popescu, B., and Muresanu, D. (2015). Vascular cognitive impairment, dementia, aging and energy demand. A vicious cycle. J. Neural Transm. (Vienna) 122 (Suppl. 1), S47–S54. doi:10.1007/s00702-013-1129-3

Potenza, M. A., Montagnani, M., Santacroce, L., Charitos, I. A., and Bottalico, L. (2023). Ancient herbal therapy: a brief history of Panax ginseng. J. Ginseng Res. 47 (3), 359–365. doi:10.1016/j.jgr.2022.03.004

Qiong, W., Yong-Liang, Z., Ying-Hui, L., Shan-Guang, C., Jiang-Hui, G., Yi-Xi, C., et al. (2016). The memory enhancement effect of Kai Xin San on cognitive deficit induced by simulated weightlessness in rats. *J. Ethnopharmacol.* 187, 9–16. doi:10.1016/j.jep.2016.03.070

Qiu, W.-Q., Ai, W., Zhu, F.-D., Zhang, Y., Guo, M.-S., Law, B.Y.-K., et al. (2022). Polygala saponins inhibit NLRP3 inflammasome-mediated neuroinflammation via SHP-2-Mediated mitophagy. *Free Radic. Biol. Med.* 179, 76–94. doi:10.1016/j.freeradbiomed. 2021.12.263

Qu, S., Liu, M., Cao, C., Wei, C., Meng, X. E., Lou, Q., et al. (2021). Chinese medicine formula kai-xin-san ameliorates neuronal inflammation of CUMS-induced depressionlike mice and reduces the expressions of inflammatory factors via inhibiting TLR4/IKK/ NF-kB pathways on BV2 cells. *Front. Pharmacol.* 12, 626949. doi:10.3389/fphar.2021. 626949

Reeju, M., Chiranjivi, T., Wang, N., Ram, C., Xue, D., and Jia, Y. M. (2017). Identification of Polygala Saponins and their metabolites *in vivo* from ethanol extract of Kaixinsan. *Chinese Journal of Experimental Traditional Medical Formulae* 23 (19), 118–123. doi:10.13422/j.cnki.syfjx.2017190118

Ren, J.-I., Zhang, A.-H., Kong, L., Han, Y., Yan, G.-L., Sun, H., et al. (2020). Analytical strategies for the discovery and validation of quality-markers of traditional Chinese medicine. *Phytomedicine* 67, 153165. doi:10.1016/j.phymed.2019.153165

Rummel, N. G., Chaiswing, L., Bondada, S., St Clair, D. K., and Butterfield, D. A. (2021). Chemotherapy-induced cognitive impairment: focus on the intersection of oxidative stress and TNFa. *Cell Mol. Life Sci.* 78 (19-20), 6533–6540. doi:10.1007/s00018-021-03925-4

Shan, X., Lv, S., Huang, P., Zhang, W., Jin, C., Liu, Y., et al. (2023a). Classic famous prescription kai-xin-san ameliorates alzheimer's disease via the wnt/ β -catenin signaling pathway. *Mol. Neurobiol.* doi:10.1007/s12035-023-03707-y

Shan, X. X., Zhou, L. L., Li, D. W., Yin, H. H., Tao, W. W., Li, J. Y., et al. (2023b). Research progress on mechanism of classic famous prescription Kaixin Powder in treatment of Alzheimer's disease. *Chin. Traditional Herb. Drugs* 54 (11), 3685–3695. doi:10.7501/j.issn.0253-2670.2023.11.030

Speidell, A. P., Demby, T., Lee, Y., Rodriguez, O., Albanese, C., Mandelblatt, J., et al. (2018). Development of a human APOE knock-in mouse model for study of cognitive function after cancer chemotherapy. *Neurotox. Res.* 35 (2), 291–303. doi:10.1007/ s12640-018-9954-7

Sun, Y., Feng, G., Zheng, Y., Liu, S., Zhang, Y., Pi, Z., et al. (2020). Putative multiple reaction monitoring strategy for the comparative pharmacokinetics of postoral administration Renshen–Yuanzhi compatibility through liquid chromatography–tandem mass spectrometry. *J. Ginseng Res.* 44 (1), 105–114. doi:10.1016/j.jgr.2018.09.007

Tang, C., Fang, Z., Chu, D., Zhang, L., Tang, Y., Zhou, J., et al. (2022). Pharmacodynamics, network pharmacology, and pharmacokinetics of Chinese medicine formula 9002A in the treatment of alzheimer's disease. *Front. Pharmacol.* 13, 849994. doi:10.3389/fphar.2022.849994

Tang, K. S. (2019). The cellular and molecular processes associated with scopolamineinduced memory deficit: a model of Alzheimer's biomarkers. *Life Sci.* 233, 116695. doi:10.1016/j.lfs.2019.116695

Toval, A., Garrigos, D., Kutsenko, Y., Popović, M., Do-Couto, B. R., Morales-Delgado, N., et al. (2021). Dopaminergic modulation of forced running performance in adolescent rats: role of striatal D1 and extra-striatal D2 dopamine receptors. *Mol. Neurobiol.* 58 (4), 1782–1791. doi:10.1007/s12035-020-02252-2

Wang, B., Feng, X., Liu, S., Qiu, F., Lu, X., and Li, Z. (2021a). Comprehensive quality assessment of kaixin powder by HPLC-DAD quantification and HPLC-QTOF-MS/MS confirmation. *ACS Omega* 6 (17), 11319–11326. doi:10.1021/acsomega.1c00289

Wang, N., Du, N., Peng, Y., Yang, K., Shu, Z., Chang, K., et al. (2021b). Network patterns of herbal combinations in traditional Chinese clinical prescriptions. *Front. Pharmacol.* 11, 590824. doi:10.3389/fphar.2020.590824

Wang, N., Hassan, A., Jia, Y. M., Xue, D., Ram, C., Huang, S. M., et al. (2017a). Identification of polygala oligosaccharide esters and their metabolites in rat plasma after oral administration of ethanol extract of Kai Xin San by UHPLC-MS. *Acta Pharm. Sin.* 52 (10), 1592–1598. doi:10.16438/j.0513-4870.2017-0373

Wang, W., Liao, Q.-P., Quan, L.-H., Liu, C.-Y., Chang, Q., Liu, X.-M., et al. (2010). The effect of Acorus gramineus on the bioavailabilities and brain concentrations of ginsenosides Rg1, Re and Rb1 after oral administration of Kai-Xin-San preparations in rats. *J. Ethnopharmacol.* 131 (2), 313–320. doi:10.1016/j.jep.2010.06.034

Wang, X., Wang, Z., Cao, J., Dong, Y., and Chen, Y. (2021d). Melatonin alleviates acute sleep deprivation-induced memory loss in mice by suppressing hippocampal ferroptosis. *Front. Pharmacol.* 12, 708645. doi:10.3389/fphar.2021.708645 Wang, R., Wu, T., Liu, Y., Yu, M. X., Tao, X. J., and Wang, H. B. (2018). The effect of Radix Polygala and honey stir-baking Radix Polygala on acute gastrointestinal toxicity of mice. *Chin. Med. Mod. Dis. Educ.* 16, 88–90. doi:10.3969/j.issn.1672-2779.2018. 08.037

Wang, X., Zhang, Y., Niu, H., Geng, Y., Wang, B., Yang, X., et al. (2017b). Ultra-fast liquid chromatography with tandem mass spectrometry determination of eight bioactive components of Kai-Xin-San in rat plasma and its application to a comparative pharmacokinetic study in normal and Alzheimer's disease rats. *J. Sep. Sci.* 40 (10), 2131–2140. doi:10.1002/jssc.201601343

Wegmann, S., Biernat, J., and Mandelkow, E. (2021). A current view on Tau protein phosphorylation in Alzheimer's disease. *Curr. Opin. Neurobiol.* 69, 131–138. doi:10. 1016/j.conb.2021.03.003

Wei, P.-l., Gu, H., Liu, J., and Wang, Z. (2018). Development of fangiomics for systems elucidation of synergistic mechanism underlying combination therapy. *Comput. Struct. Biotechnol. J.* 16, 565–572. doi:10.1016/j.csbj.2018.10.015

Wen, L., Xia, N., Tang, P., Hong, Y., Wang, Z., Liu, Y., et al. (2015). The gastrointestinal irritation of polygala saponins and its potential mechanism *in vitro* and *in vivo*. *BioMed Res. Int.* 2015, 918048–8. doi:10.1155/2015/918048

Wu, Q., Yuan, D., Wang, Q., and Wu, X. (2009). Effects of volatile oil of Rhizoma Acori Tatarinowii on morphology and cell viability in cultured cardiac myocytes. *J. Chin. Med. Mater.* 32, 242–245. doi:10.13863/j.issn1001-4454.2009.02.037

Xie, Z., Lu, H., Yang, S., Zeng, Y., Li, W., Wang, L., et al. (2020). Salidroside attenuates cognitive dysfunction in senescence-accelerated mouse prone 8 (SAMP8) mice and modulates inflammation of the gut-brain Axis. *Front. Pharmacol.* 11, 568423. doi:10. 3389/fphar.2020.568423

Xu, Y. M., Wang, X. C., Xu, T. T., Li, H. Y., Hei, S. Y., Luo, N. C., et al. (2019). Kai Xin San ameliorates scopolamine-induced cognitive dysfunction. *Neural Regen. Res.* 14 (5), 794–804. doi:10.4103/1673-5374.249227

Yi, P., Zhang, Z., Huang, S., Huang, J., Peng, W., and Yang, J. (2020). Integrated metaanalysis, network pharmacology, and molecular docking to investigate the efficacy and potential pharmacological mechanism of Kai-Xin-San on Alzheimer's disease. *Pharm. Biol.* 58 (1), 932–943. doi:10.1080/13880209.2020.1817103

Yin, M., Chen, Y., Zheng, H., Pu, T., Marshall, C., Wu, T., et al. (2017). Assessment of mouse cognitive and anxiety-like behaviors and hippocampal inflammation following a repeated and intermittent paradoxical sleep deprivation procedure. *Behav. Brain Res.* 321, 69–78. doi:10.1016/j.bbr.2016.12.034

Yu, S., Liu, S., Wang, N., Yu, D., Qin, M., Wu, J., et al. (2021). Novel insights into antidepressant mechanism of Kai Xin San formula: inhibiting NLRP3 inflammasome activation by promoting autophagy. *Phytomedicine* 93, 153792. doi:10.1016/j.phymed. 2021.153792

Yue, K. X., Li, H. L., Pi, Z. F., Song, F. R., Wang, Y. P., and Liu, Z. Y. (2022). Multiresidue screening of pesticides in Panax Ginseng C. A. Meyer by ultra-highperformance liquid chromatography with quadrupole time-of-flight mass spectrometry. J. Sep. Sci. 45 (10), 1702–1710. doi:10.1002/jssc.202100933

Zhang, B., Li, Y., Liu, J.-w., Liu, X.-w., Wen, W., Cui, Y., et al. (2019). Postsynaptic GluR2 involved in amelioration of aβ-induced memory dysfunction by KAIXIN-san through rescuing hippocampal LTP in mice. *Rejuvenation Res.* 22 (2), 131–137. doi:10. 1089/rei.2018.2080

Zhang, C., Xue, P., Zhang, H., Tan, C., Zhao, S., Li, X., et al. (2022a). Gut brain interaction theory reveals gut microbiota mediated neurogenesis and traditional

Chinese medicine research strategies. Front. Cell. Infect. Microbiol. 12, 1072341. doi:10.3389/fcimb.2022.1072341

Zhang, L., Yong, Y.-Y., Deng, L., Wang, J., Law, B.Y.-K., Hu, M.-L., et al. (2023). Therapeutic potential of Polygala saponins in neurological diseases. *Phytomedicine* 108, 154483. doi:10.1016/j.phymed.2022.154483

Zhang, N., Zhao, L., Su, Y., Liu, X., Zhang, F., and Gao, Y. (2021). Syringin prevents a $\beta_{25\cdot35}$ -induced neurotoxicity in SK-N-sh and SK-N-be cells by modulating miR-124-3p/BID pathway. *Neurochem. Res.* 46 (3), 675–685. doi:10.1007/s11064-021-03240-1

Zhang, X., Li, Q., Lv, C., Xu, H., Liu, X., Sui, Z., et al. (2015). Characterization of multiple constituents in Kai-Xin-San prescription and rat plasma after oral administration by liquid chromatography with quadrupole time-of-flight tandem mass spectrometry. *J. Sep. Sci.* 38 (12), 2068–2075. doi:10.1002/jssc.201500123

Zhang, Y., Jin, F., Wei, X., Jin, Q., Xie, J., Pan, Y., et al. (2022b). Chinese herbal medicine for the treatment of chronic fatigue syndrome: a systematic review and metaanalysis. *Front. Pharmacol.* 13, 958005. doi:10.3389/fphar.2022.958005

Zhang, Y.-W., and Cheng, Y.-C. (2019). Challenge and prospect of traditional Chinese medicine in depression treatment. *Front. Neurosci.* 13, 190. doi:10.3389/fnins.2019. 00190

Zheng, Y., Feng, G., Sun, Y., Liu, S., Pi, Z., Song, F., et al. (2018). Study on the compatibility interactions of formula Ding-Zhi-Xiao-Wan based on their main components transport characteristics across Caco-2 monolayers model. *J. Pharm. Biomed. Analysis* 159, 179–185. doi:10.1016/j.jpba.2018.06.067

Zhou, W., Zhang, H., Wang, X., Kang, J., Guo, W., Zhou, L., et al. (2022a). Network pharmacology to unveil the mechanism of Moluodan in the treatment of chronic atrophic gastritis. *Phytomedicine* 95, 153837. doi:10.1016/j.phymed.2021.153837

Zhou, X. J., Liu, M., Yan, J. J., Cao, Y., and Liu, P. (2012). Antidepressant-like effect of the extracted of Kai Xin San, a traditional Chinese herbal prescription, is explained by modulation of the central monoaminergic neurotransmitter system in mouse. *J. Ethnopharmacol.* 139 (2), 422–428. doi:10.1016/j.jep.2011.11.027

Zhou, Z., Ma, Y., Xu, T., Wu, S., Yang, G.-Y., Ding, J., et al. (2022b). Deeper cerebral hypoperfusion leads to spatial cognitive impairment in mice. *Stroke Vasc. Neurology* 7 (6), 527–533. doi:10.1136/svn-2022-001594

Zhu, K. Y., Mao, Q. Q., Ip, S. P., Choi, R. C., Dong, T. T., Lau, D. T., et al. (2012). A standardized Chinese herbal decoction, kai-xin-san, restores decreased levels of neurotransmitters and neurotrophic factors in the brain of chronic stress-induced depressive rats. *Evid. Based Complement. Altern. Med.* 2012, 149256. : doi:10.1155/2012/149256

Zhu, K. Y., Xu, S. L., Choi, R. C., Yan, A. L., Dong, T. T., and Tsim, K. W. (2013). Kaixin-san, a Chinese herbal decoction containing ginseng radix et rhizoma, polygalae radix, acori tatarinowii rhizoma, and poria, stimulates the expression and secretion of neurotrophic factors in cultured astrocytes. *Evid. Based Complement. Altern. Med.* 2013, 731385. doi:10.1155/2013/731385

Zhu, Y., Chao, C., Duan, X., Cheng, X., Liu, P., Su, S., et al. (2017). Kai-Xin-San series formulae alleviate depressive-like behaviors on chronic mild stressed mice via regulating neurotrophic factor system on hippocampus. *Sci. Rep.* 7 (1), 1467. doi:10.1038/s41598-017-01561-2

Zhu, Y., Duan, X., Cheng, X., Cheng, X., Li, X., Zhang, L., et al. (2016). Kai-Xin-San, a standardized traditional Chinese medicine formula, up-regulates the expressions of synaptic proteins on hippocampus of chronic mild stress induced depressive rats and primary cultured rat hippocampal neuron. *J. Ethnopharmacol.* 193, 423–432. doi:10. 1016/j.jep.2016.09.037

Glossa	ry		
		MG	Muscle glycogen
KXS	Kaixinsan	РК	Pharmacokinetics
GRe	Ginsenoside Re	ADME	Absorption, distribution, metabolism and excretion
GRb1	Ginsenoside Rb1	Ach	Acetylcholine
POL	Polygalaxanthone III	DA	Dopamine
ТСМ	Traditional Chinese medicine	5-HT	5-hydroxytryptamine
LC	Liquid chromatography	5-HIAA	5-hydroxyindoleacetic acid
MS	Mass spectrometry	PI3K	Phosphoinositide 3-kinase
QTOF-MS/	Quadrupole time-of-flight tandem mass spectrometry	Akt	Protein kinase B
MS		ROS	Reactive Oxygen Species
GC	Gas chromatography	TNF-α	Tumor necrosis factor-alpha
UPLC	Ultra-performance liquid chromatography	AA	Arachidonic acid
POL	Polygalaxanthone III	Bax	BCL2-associated X protein
Abeta	Amyloid beta peptide	Bcl-2	B cell lymphoma 2
AChE	Acetylcholinesterase	BHT	Butylated hydroxytoluene
IL	Interleukin	АМРК	AMP-activated protein kinase
BDNF	Brain derived neurotrophic factor	SAMP8	Senescence-accelerated mouse prone 8
HPA	Hypothalamic-pituitary-adrenal	NLRP3	Nucleotide-binding oligomerization domain-like receptor family,
CUMS	Chronic unpredictable mild stress		pyrin domain containing 3
CHD	Coronary heart disease	MMP	Mitochondrial membrane potential
MI	Myocardial ischemia	EC	Energy charge
LDH	Lactate dehydrogenase	NMDAR1	N-methyl-D-aspartate receptor 1
СК	Creation kinase	ChAT	Choline acetyltransferase
CAT	Catalase	SYN	Synaptophysin
SOD	Superoxide dismutase	p-CREB	Phosphorylated-CREB
GSH-PX	Glutathione peroxidase	8-OHdG	8-hydroxy-2'-deoxyguanosine
MDA	Malondialdehyde	3-NT	3-nitrotyrosine
AD	Alzheimer's disease	CUMS	Chronic unpredictable mild stress
CRP	C-reactive protein	m-TOR	Mammalian target of rapamycin
MID	Multi-infarct dementia	BLA	Lactic acid
VD	Vascular dementia	CK-MB	Creation kinase MB
CD	Cognitive dysfunction	NE	Epinephrine
MWM	Morris water maze	NGF	Nerve growth factor
PSD	Paradoxical sleep deprivation		
GLU	Glutamic acid		
GABA	γ-aminobutyric acid		
CREB	Cyclic AMP response element binding protein		
	, <u>1</u> OI		

Doxorubicin

Chronic fatigue syndrome

Serum urea nitrogen

Liver glycogen Muscle lactic acid

DOX

CFS

SUN

LG

MLA