



Anthocyanins in the Management of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review

Rozita Naseri¹, Fatemeh Farzaei², Pouya Haratipour^{3,4}, Seyed Fazel Nabavi⁵, Solomon Habtemariam⁶, Mohammad Hosein Farzaei^{2*}, Reza Khodarahmi^{7*}, Devesh Tewari⁸ and Saeideh Momtaz^{9,10}

¹ Internal Medicine Department, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran, ² Pharmaceutical Sciences Research Center, Kermanshah University of Medical Sciences, Kermanshah, Iran, ³ Department of Chemistry, Sharif University of Technology, Tehran, Iran, ⁴ Phyto Pharmacology Interest Group, Universal Scientific Education and Research Network, Los Angeles, CA, United States, ⁵ Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran, ⁶ Pharmacognosy Research Laboratories, Medway School of Science, University of Greenwich, Kent, United Kingdom, ⁷ Medical Biology Research Center, Kermanshah University of Medical Sciences, Kermanshah, Iran, ⁸ Department of Pharmaceutical Sciences, Faculty of Technology, Kumaun University, Nainital, India, ⁹ Medicinal Plants Research Center, Institute of Medicinal Plants, ACECR, Karaj, Iran, ¹⁰ Toxicology and Diseases Group, The Institute of Pharmaceutical Sciences, Tehran University of Medical Sciences, Tehran, Iran

OPEN ACCESS

Edited by:

Anna Rita Bilia,
Università degli Studi di Firenze, Italy

Reviewed by:

Michael Erich Netzel,
The University of Queensland,
Australia
Gokhan Zengin,
Selçuk University, Turkey

*Correspondence:

Mohammad Hosein Farzaei
mh.farzaei@gmail.com
Reza Khodarahmi
rkhodarahmi@mbrc.ac.ir;
rkhodarahmi@kums.ac.ir

Specialty section:

This article was submitted to
Ethnopharmacology,
a section of the journal
Frontiers in Pharmacology

Received: 14 March 2018

Accepted: 26 October 2018

Published: 04 December 2018

Citation:

Naseri R, Farzaei F, Haratipour P, Nabavi SF, Habtemariam S, Farzaei MH, Khodarahmi R, Tewari D and Momtaz S (2018) Anthocyanins in the Management of Metabolic Syndrome: A Pharmacological and Biopharmaceutical Review. *Front. Pharmacol.* 9:1310. doi: 10.3389/fphar.2018.01310

The term “metabolic syndrome” (MetS) refers to a combination of diabetes, high blood pressure, and obesity. The origin of MetS includes a combination of multiple factors, such as sedentary lifestyle, unhealthy diet choice, and genetic factors. MetS is highly prevalent and adversely affects the general population by elevating risk of cardiovascular complications, organ failure, and much other pathology associated with late-stage diabetes. Anthocyanins (ANTs) are health-promoting bioactive compounds belonging to the flavonoids subclass of polyphenols. Numerous studies have reported the potential therapeutic benefits on MetS syndrome and diabetes from fruits rich in ANTs. This review summarizes the role of several dietary ANTs on preventing and managing MetS as well as the pharmacological mechanisms and biopharmaceutical features of their action. We also discuss potential nanoformulation and encapsulation approaches that may enhance the bioefficacy of ANTs in MetS. Experiments have demonstrated that ANTs may attenuate the symptoms of MetS via improving insulin resistance, impaired glucose tolerance, dyslipidaemia, cholesterol levels, hypertension, blood glucose, protecting β cells, and preventing free radical production. In brief, the intake of ANT-rich supplements should be considered due to their plausible ability for prevention and management of MetS. Additionally, randomized double-blind clinical trials are obligatory for evaluating the bioefficacy and pharmacological mechanisms of ANTs and their pharmaceutical formulations in patients with MetS.

Keywords: anthocyanins, natural pigments, phytochemicals, metabolic syndrome, diabetes mellitus, insulin resistance

INTRODUCTION

Metabolic syndrome (MetS), also known by other names such as “insulin resistance syndrome” or “syndrome X,” was first defined by Kylin in the 1920s as a combination of hyperglycemia, hypertension, and gout. To date, its diagnostic criteria include atherogenic dyslipidemia [low HDL cholesterol and high triglycerides (TGs)], hyperglycemia, insulin resistance (landmark sign of the disease), glucose intolerance, hypertension, and central obesity (Spiegelman and Flier, 2001; Ford et al., 2002; Grundy et al., 2004; Alberti et al., 2005; Russell and Proctor, 2006; Romeo et al., 2012). Hence, MetS is common medical terminology for a combination of diabetes, high blood pressure, and obesity. Furthermore, central obesity or hypertriglyceridaemic waist phenotype contributes to the development of hyperinsulinemia, lipid abnormalities, hyperglycemia, and the activation of inflammatory and prothrombotic mediators with an amplified risk of the prevalence of type 2-diabetes mellitus (DM), cardiovascular diseases (CVD), and many cancers (Berlin et al., 2000; Festa et al., 2001; Lakka et al., 2002; Carr et al., 2004; Gluckman and Hanson, 2004; Hansel et al., 2004; Guilder et al., 2006; Vlachopoulos et al., 2010). DM is a chronic disease diagnosed by hyperglycemia, owing to insufficient insulin production or inadequate cellular sensitivity to insulin, and progressive decline in B-cell function (Kudva and Butler, 1997; American Diabetes Association, 2009). DM is a rising global problem and expected to affect around 380 million by 2025 (Kaul et al., 2013). Several studies suggested that plant derivatives such as polyphenols possess numerous biological activities with anti-inflammatory, antioxidative, and insulin-sensitizing effects (Hämäläinen et al., 2007; Shamim, 2009; Sodagari et al., 2015). Natural supplements and various herbal products, especially ANT-rich food, are claimed to be beneficial in controlling MetS. Thus, in the present review, we reviewed ANT-rich food as potential alternative therapeutic as well as their possible mechanisms of action for managing MetS.

EPIDEMIOLOGY OF METABOLIC SYNDROME (METS)

The worldwide prevalence of MetS varies between 10 and 84% for urban populations based on the region, composition (age, sex, race) of the population, and the definition of MetS. The International Diabetes Federation (IDF) estimated that approximately one-quarter of the global adult population has MetS, of which 28% were men and 34% were women belonging to the atherosclerosis risk in communities (ARIC) study population (Desroches and Lamarche, 2007; Kolovou et al., 2007). In a survey conducted on 8,814 people in the USA, the prevalence of MetS was more than 40% in population between 60 and 69 years (Ford et al., 2002; Day, 2007). According to Amirkalali et al. the prevalence of MetS in Iranian individuals was 36.9%, depending on the adult treatment panel III (ATP III) criteria, 34.6% according to the IDF, and 41.5% based on the Joint Interim Societies (JIS) criteria (Amirkalali et al., 2015). The high prevalence of MetS is responsible for substantial public

health consequences owing to augmented risk of type 2 DM and CVD (Carr et al., 2004). Nowadays, diabetes is becoming a global pandemic with increasing prevalence in India and Asia, whom will be the 7th leading reason of death by the year 2030 according to the World Health Organization (WHO) estimates (World Health Organization, 2012; Maiese, 2015; Munasinghe and Katare, 2016).

CURRENT THERAPEUTIC PROTOCOLS FOR METS

The primary cause of MetS is diet, obesity, physical inactivity, age, and genetic profiles, such as a defect in a single gene, in lamin A/C, O-acyltransferase, 1-acylglycerol-3-phosphate, seipin, the adrenergic receptor, and adiponectin (Steppan et al., 2001; Lakka and Laaksonen, 2007; Schröder, 2007; Abete et al., 2011; Kastorini et al., 2011; Amiot et al., 2016; Martinez-Abundis et al., 2016; Merone and McDermott, 2017). Evidence indicates that combination of lifestyle modifications with effective weight loss and drug therapy may serve as treatment for MetS (Marvasti and Adeli, 2010). First-line recommendations include lifestyle modification as well as introduction of the Mediterranean diet, which includes more fruit and vegetable consumption along with higher monounsaturated fat intake (Esposito et al., 2004). Such an approach may suppress the postprandial glycaemia, serum TG levels, and raise HDL-cholesterol; thus, delaying the transition from impaired glucose tolerance to incidence of type 2 DM, and reducing risk of developing MetS (Tsuda, 2012).

Since insulin resistance plays significant role in regulating diabetes, pharmacological interventions, such as thiazolidinediones and metformin, seem to have supplementary effects in ameliorating diabetes and/or MetS evolution by stimulating muscle glucose uptake and suppressing hepatic glucose production along with AMP-activated protein kinase (AMPK) activation. AMPK is a major cytological regulator of glucose and lipid metabolism, thereby is considered as a potential target for therapeutic management of type 2 DM (Zhou et al., 2001; Hawley et al., 2002; Grewal et al., 2016; Maskimov et al., 2016).

Lipid-lowering agents and low-density lipoprotein (LDL) lowering standard drugs, such as statins and ezetimibe, modify atherogenic dyslipidemia, and CVD in patients with MetS. Other drugs that reduce MetS progression include thiazolidinediones, glucagon-like peptide-1 (GLP-1) agonists, and inhibitors of dipeptidyl peptidase-4 (DPP-4). Once statin therapy and lifestyle modifications are not successful, niacin may be helpful to reduce TG (Marvasti and Adeli, 2010).

To manage diabetes, there are several ongoing drug therapy approaches including sulphonylureas, metformin, and α -glucosidase inhibitors, which suppress and interfere with gut glucose production and absorption, but may become refractory to the treatment over time. It is now clear that the aggressive control of hyperglycaemia by synthetic drugs in patients with MetS may be involved in the progression of various chronic complications, such as retinopathy and nephropathy. Since the utilization of oral antihyperglycemic drugs have limited efficacy

and numerous side effects, complementary and alternative medicines such as acupuncture, herbal medicines, Ayurveda, traditional medicine, and other medicinal approaches may be helpful in the management of MetS.

MOLECULAR PATHOPHYSIOLOGY OF METS

Diverse pathophysiologic factors that may drive the progression of MetS, for instance, insulin resistance with circulating fatty acids accumulation and adiposity are the main factors (Montague and O'rahilly, 2000; Taniguchi et al., 2006; Barazzoni et al., 2018) (Figure 1). Insulin resistance is a physical condition, which is demarcated as a state that needs additional insulin to produce biological effects with decreasing glucose uptake in muscle and adipose tissue. Insulin affects antilipolysis and stimulates lipoprotein lipase via inhibition of lipolysis in adipose tissue. Therefore, when insulin resistance occurs, increasing amounts of fatty acids are produced by high amounts of stored triacylglycerol

molecules, inciting additional lipolysis in adipose tissue. In the liver, insulin resistance leads to flaws in insulin receptor substrate-1 and substrate-2 tyrosine phosphorylation, leading to the activation of protein kinase C. Excessive fatty acids may also impair activation of protein kinase C as well as acyl-coenzyme A (CoA) generation in muscles.

Obesity is linked with accumulation of higher macrophages in adipose tissues and augmented pro-inflammatory cytokines (Di Gregorio et al., 2005). Fat accumulation in adipose tissue, liver, skeletal muscle, heart, and pancreas may increase systemic oxidative stress independent of hyperglycemia (Unger, 2003) as well as adipocytokines or adipokines such as plasminogen activator inhibitor-1 (PAI-1), tumor necrosis factor (TNF)- α , resistin, and leptin (Friedman and Halaas, 1998; Leyva et al., 1998; Matsuzawa et al., 1999; Niemann et al., 2017; Reho and Rahmouni, 2017; Louwen et al., 2018). In a research conducted by Guzmán-Gerónimo et al. a high-fat diet caused increased arterial blood pressure, high levels of TG in plasma, and reduction of HDL-C due to elevation of fatty acid reesterification (Guzmán-Gerónimo et al., 2017). Likewise, the reduction of HDL-C has

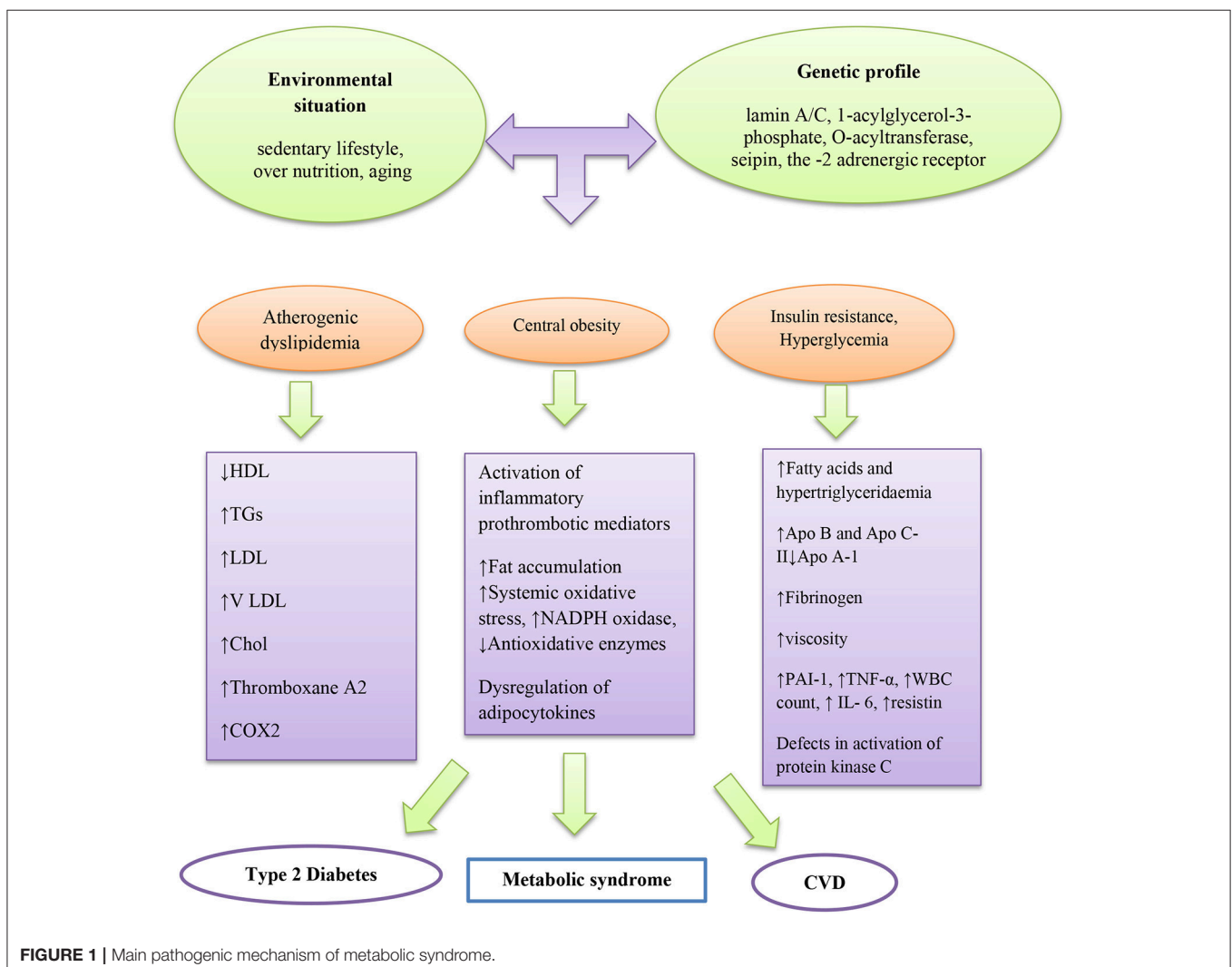


FIGURE 1 | Main pathogenic mechanism of metabolic syndrome.

been reported in humans with MetS (Guzmán-Gerónimo et al., 2017).

ROLE OF OXIDATIVE STRESS IN PATHOGENESIS OF METS

Oxidative stress, a shift of the redox balance, is a deleterious condition that occurs when cellular components including proteins, lipids, and DNA are damaged. Selective radical overgeneration in adipose tissue may possess a prominent role in insulin resistance development, diabetes, and CVD through impairment of muscle glucose uptake and secretion of insulin from β cells (Maddux et al., 2001). Numerous studies revealed that an increased level of reactive oxygen species (ROS) in peripheral blood from accumulated fat is involved in initiation of insulin resistance in different adipose tissue, skeletal muscle, and other diabetic complications. Insulin resistance leads to disruption of many prime oxidative reactions, resulting in undue ROS generation at cellular and mitochondrial levels. Studies have shown that in type 2 diabetic patients, lipid peroxidation increased, while the levels of plasma glutathione (GSH) and GSH-metabolizing enzymes are reduced (Sundaram et al., 1996). Folmer et al. reported that hyperglycemia induces free radical and oxidative stress production in mice (Folmer et al., 2002). An application of about 10–20 mM glucose into the posterior root ganglion neurons resulted in production of O_2^- , H_2O_2 , lipid oxidation and neuronal death (Schmeichel et al., 2003).

Adiponectin, an anti-inflammatory cytokine produced by adipocytes, improves insulin sensitivity and inhibits many inflammatory processes. In cultured adipocytes, oxidative stress was increased when the level of fatty acid was enhanced, this was attributed to the activation of NADPH oxidase and generation of adipocytokines (fat-derived hormones) at a deregulated manner (Hotamisligil et al., 1993; Shimomura et al., 1996; Lara-Castro et al., 2006). NADPH oxidase is a key source of ROS production in adipocytes, which increases in obesity. Treatment with NADPH oxidase inhibitor may decrease ROS production in adipose tissue, attenuate adipocytokines dysregulation, and ameliorate hyperlipidemia and diabetes in obese mice, also may reduce pathogenesis of several vascular diseases like hypertension and atherosclerosis (Iwaki et al., 2003; Farzaei et al., 2017, 2018; Furukawa et al., 2017).

ANTHOCYANINS AS ANTIOXIDANT

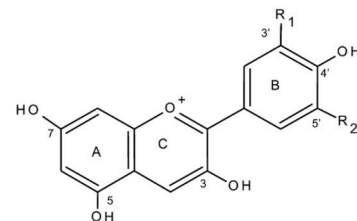
The term “Anthocyanin” is derived from two Greek words, i.e., antos for flower and kyanos means blue. ANTs are one of the most important health-promoting natural plant pigments, which belong to the flavonoids group and polyphenol class of phytochemicals (Dreiseitel et al., 2008; Pojer et al., 2013). Variations in ANTs are a result of the number and degree of methylation, hydroxyl group position, and the number of rings (aliphatic/aromatic) that are attached to the sugar moieties, also are dependent

on the location and type of sugar attachment to the molecule on the basic anthocyanidin skeleton (Deng et al., 2013). Flavylium cation (2-phenylbenzopyrylium) is the fundamental structure that links with either one or more sugar moiety and hydroxyl (-OH) and/or methoxyl (-OCH₃) groups. Cyanidin-3-glucoside, cyanidin-3-(xylose-glucose-galactoside), cyanidin-3-(xylose-feruloyl-glucose-galactoside), cyanidin-3-(xylose-sinapoyl-glucose-galactoside), cyanidin-3-(xylose-galactoside), and cyanidin-3-(xylose-coumaroyl-glucose-galactoside) are some of the most important ANTs (Table 1, Figure 2).

ANTs are water-soluble bioactive compounds widely found in various vegetables and fruits, including berries like cranberries, strawberries, blueberries, blackberries, elderberries, grapes, currants, plums, cherries, red cabbage, red onions, and sweet potatoes. They are usually distributed in fruits and flowers;

TABLE 1 | Major anthocyanidins in plants (Pojer et al., 2013; Fang, 2015).

Selected plant source	Anthocyanidins
Apple, elderberry, blackberry, pear, peach, fig, cherry, red onion, red cabbage, rhubarb, gooseberry	Cyanidin
Banana, red radish, strawberry, potato	Pelargonidin
Pomegranate, black currant, gooseberry, purple carrot, blood orange, egg plant, green bean	Cyanidin and delphinidin
Pomegranate, passion fruit, eggplant, green bean	Delphinidin
Plum, sweet cherry, purple sweet potato	Cyanidin and peonidin
Mango	Peonidin
Bilberry, red grape	Petunidin and malvidin



Anthocyanidins	R1 Substitute	R1 Substitute	MW
Pelargonidin	H	H	271
Cyanidin	OH	H	287
Delphinidin	OH	OH	303
Peonidin	OCH ₃	H	301
Petunidin	OCH ₃	OH	317
Malvidin	OCH ₃	OCH ₃	331

FIGURE 2 | Structures of common anthocyanidins in fruits and vegetables.

however, stems, leaves, and roots of some plants also contain different types of ANTs (Wu and Prior, 2005).

ANTs are natural antioxidants with high reactivity toward ROS, mainly due to their ability to transfer electrons or to donate the hydrogen atoms from various hydroxyl groups to free radicals, to their basic structural compounds and ring orientation, and to the unpaired electron supporting ability of ANTs (Wang et al., 1999; Anderson and Jordheim, 2008; Markakis, 2012). ANTs also have prominent therapeutic effects like anti-inflammatory, anti-viral, anti-carcinogenic, anti-mutagenic, anti-allergic, anti-microbial, improvement of arterial stiffness and antioxidants effects, and are strong lipid peroxidation inhibitors (Kim et al., 2006; Tsang et al., 2018). The antioxidant capacity of ANTs has been proven by several methods, such as oxygen radical absorbance capacity (ORAC), DPPH (2,2-diphenyl-1-picrylhydrazyl) assay, ABTS [2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid)] assay, and etc. (Burns et al., 2000; Wang and Lin, 2000; Prior et al., 2001; Zheng and Wang, 2003; Steed and Truong, 2008; Sodagari et al., 2015; Yue et al., 2018).

BIOPHARMACEUTICAL FEATURES OF ANTHOCYANINS IN METABOLIC SYNDROME

Although ANTs have low absorption and high metabolism, the regular intake of ANTs may have critical and/or beneficial effects on human health. ANTs are absorbed intact as glycosides and their absorption rates are influenced by their chemical structures. ANTs are poorly absorbed after oral administration (about 10–50 nM) in the stomach and small intestine, and the maximal plasma concentration time is about 1.5 h. Individual ANTs absorption efficiency is between 0.12 and 0.25% for non-acylated ANTs, and 0.0079–0.019% for acylated ANTs. The acylated types showed lower affinity for the transporter bilitranslocase (Kay, 2006; Czank et al., 2013; Fernandes et al., 2014; Kay et al., 2017).

Elimination from plasma differs based upon ANTs structures. Non-acylated compounds are eliminated slower compared to the acylated forms. Additionally, variations in ANTs interactions with the transporters (at tissue level) may be responsible for the differences in the plasma kinetics. Several studies have revealed that ANTs are absorbed and excreted intact. Pharmacokinetics data analysis in animal and human subjects suggested that the intestine is the major site of ANTs absorption (Ferrars et al., 2014; Kamiloglu et al., 2015). ANTs are metabolized to glucurono-, sulfo-, or methyl-derivatives in the proximal gastrointestinal (GI) tract like other flavonoids. ANTs clearance from the circulation is suitably rapid (Bub et al., 2001).

The majority of ingested ANTs appears to reach the lower intestine, and are subjected to the microbial catabolism. In intact functioning colon volunteers, a portion of ANTs pass into the large intestine, where it is deglycosylated and the subsequent aglycones are broken down via C-ring fission with fragments of A- and B-ring (González-Barrío et al., 2011).

The investigation of the biopharmaceutical profile and comprehensive researches on different aspects of ANTs bioavailability such as absorption, distribution, metabolism, and excretion (ADME) are gaining tremendous interests recently. Diverse research groups are focusing on various ANTs to enhance their bioavailability against different diseases including cancer (Mueller et al., 2018; Thibado et al., 2018). Animal studies showed that ANTs are mainly absorbed in the intact glycosidic form and reach systemic circulation within 0.25–2 h. ANTs from *Vaccinium myrtillus* (400 mg/kg) reached peak plasma concentrations level (2–3 $\mu\text{g/mL}$) after 15 min and then, declined rapidly within 2 h upon a single oral administration in rats (Huang et al., 2014). After oral administration of cyanidin-3-glucoside (C3G) at 400 mg/kg, the intact form was rapidly observed in the plasma [C_{max} : 0.31 $\mu\text{mol/L}$ (0.14 $\mu\text{g/mL}$)] at 30 min (Feshani et al., 2011; Kalt et al., 2017; Tymchuk et al., 2017).

Most ANTs, especially from berries, are believed to have low bioavailability (Kay et al., 2017). The peak plasma concentration of ANTs from berries ranges between 1 and 120 nmol/L with <1% urinary recovery confirmed by different studies (Kay, 2006) and around 0.005% level at excretion (Stalmach et al., 2012). Metabolism of ANTs occurs by the formation of sulfo-, glucurono-, or methyl-derivatives in the proximal GI tract. Unmetabolized compounds have also been observed in small quantities in the urine and systemic circulation, even though the exact mechanism for absorption is still highly theoretical (Kamiloglu et al., 2015). Several studies reported that the possible transport mechanism is through intestinal glucose transporters, stomach transporters, and tight junction permeability (Passamonti et al., 2003).

In a study conducted on 9 volunteers, 300 g raspberry with 292 μmol ANTs constituted of cyanidin-based components were ingested, and the results showed that only cyanidin-*O*-glucuronide and cyanidin-3-*O*-glucoside were traced with sub-nmol/L peak concentrations (C_{max}) in the plasma, with a T_{max} (Time of Peak Concentration) after 4 and 1 h, respectively. After 0–24 h, only 20 nmol of cyanidin-3-*O*-glucoside was detected in the urine and no other parent ANTs were observed (González-Barrío et al., 2010; Ludwig et al., 2015).

The findings of Felgines and colleagues demonstrated that the excretion of blackberry ANTs in urine occurs as intact and methylated forms with no conjugated or aglycones compounds. Moreover, low amounts of aglycones and ANTs were detected in cecal contents, which suggested microflora adaptation to ANTs degradation. Additionally, ANTs and their metabolites were detected in bile rapidly after oral intake, demonstrating the rapid absorption and metabolism (Felgines et al., 2002). Methylated ANTs were also recorded in rat plasma (Ichihyanagi et al., 2004). In a recent study, consumption of ANT-enriched beverages in milk and water was examined in order to investigate the role of milk on the oral bioavailability of ANTs. The authors recorded the significant effect of milk compared to water on decreasing the C_{max} , the area under curve (AUC) of two individual pelargonidin ANTs (pelargonidin-3-glucuronide and pelargonidin-3-*O*-rutinoside), and C_{max} of pelargonidin-3-*O*-glucoside. The oral bioavailability of these

ANTs decreased in the subjects that consumed beverages in milk by about 50% (Xiao et al., 2017). Oral administration of 100 mg delphinidin-3-glucoside/kg attained the C_{max} in 15 min while the methylated form of delphinidin-3-glucoside showed C_{max} after 1 h, and the presence of ANTs glucuronides in rat plasma suggested that metabolites are produced in the liver, rather than by intestinal flora (Ichiyanagi et al., 2005).

ROLE OF NANOFORMULATION AND ENCAPSULATION METHODS IN ANTHOCYANINS BIO-EFFICACY FOR MANAGEMENT OF METABOLIC SYNDROME

Various ANTs are used in the food industries as an active ingredient, but their degradation is possible after exposure to different factors such as oxygen or light, thus, stability is of prime importance when ANTs are used as a colorant in the food industry. To overcome this problem, microencapsulation is a potential technique (Favaro-Trindade et al., 2010; Nayak and Rastogi, 2010). ANTs present in pomegranate juice were relatively degraded faster in the fresh pomegranate juice than in microencapsulated powder, representing the importance of encapsulation techniques in the preservation of the bioactive compounds (Robert et al., 2010).

A number of polyphenols do not absorbed in GI track of human (Cerdá et al., 2004; Seeram et al., 2006). Therefore, nanoencapsulation may help to conquer the susceptibility of these compounds toward GI hydrolysis, low systemic bioavailability, poor absorption, and short half-life (Shirode et al., 2015). One such example is nano-pelargonidin, which enhanced protection at ~10-fold decreased dose, and is postulated to be used in the formulation of protective drugs for mitochondrial dysfunction management that is often tested in alloxan-induced hyperglycemic L6 cells (Samadder et al., 2017). Nanoformulations can improve drug delivery and bioavailability to the target cells due to their physicochemical properties, making them viable in successfully curing deadly diseases. The same research group evaluated the preventive effects of nanoencapsulated pelargonidin against alloxan-induced DNA damaged cells (L6) by *in vitro* methods, and reported around ~10-fold enhancement in efficacy of nanoencapsulated pelargonidin than pelargonidin (Samadder et al., 2016). Extracts of Chinese herbal medicine named “Shanzhuyu” containing ANTs were also used for the preparation of metal nanoparticles, and unveiled promising anticancer activity against human liver cancer (HepG2) and human prostate cancer (PC-3) cell lines (He et al., 2006, 2017). Apart from the nanoformulations and encapsulation, nano packing is also considered an emerging technique for the preservation of quality in ANT-rich fruits like strawberries (Yang et al., 2010). Although, biocompatible and safe nanoformulation is potentially important and an emerging field to enhance the bioavailability of the ANTs, only limited studies have been conducted in

this regard. Therefore, nanoencapsulation and preparation of different nanoformulations targeting metabolic syndrome is in need.

PHARMACOLOGICAL MECHANISMS OF ANTHOCYANINS IN METABOLIC SYNDROME

Anthocyanins Enriched Extracts

All of ANT-enriched extracts may contain a significant amount of other non-ANT phenolics (flavonoids and/or phenolic acids) and other non-polyphenolic compounds, which may possess favorable impact/effect on the pathogenesis of the MetS. However, below is the pharmacological mechanisms of plant extracts, in which ANTs are considered as the main bioactive and major constituents.

Berry Anthocyanins

Blueberry Anthocyanins

Blueberries (*Vaccinium myrtillus*) from Ericaceae family are particularly high in anthocyanidins, chlorogenic acid, flavonoids, α -linolenic acid, pterostilbene and resveratrol. Myrtocyan is a highly purified extract of *Vaccinium myrtillus*, which contains 36% anthocyanosides including 3-arabinoside, delphinidin 3-galactoside, delphinidin, petunidin 3-arabinoside, petunidin 3-galactoside, cyanidin 3-galactoside, cyanidin 3-glucoside, cyanidin 3-arabinoside, malvidin 3-galactoside, malvidin 3-glucoside, peonidin 3-glucoside, peonidin, 3-galactoside, peonidin 3-arabinoside, and peonidin 3-glucoside (Routray and Orsat, 2011).

Malvidin-3-glucoside possesses anti-inflammatory activity in endothelial cells through inhibition of production of monocyte chemotactic protein-1 (MCP-1), intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1) both in protein and mRNA levels (Huang et al., 2014).

Numerous investigations indicated that blueberries have several beneficial therapeutic properties, such as attenuating age-induced oxidative stress and inflammatory responses (Lau et al., 2007), protecting the kidney (Elks et al., 2015), preventing diabetes (Martineau et al., 2006), protecting against cardiovascular disorders, preventing hyperlipidemia and hypertension (Kalea et al., 2009), and reducing obesity *in vitro* and *in vivo* (Seeram et al., 2002; Kumar et al., 2012).

Blueberries exhibited anti-inflammatory activity *in vitro* via attenuation of the balances of pro-inflammatory cytokines in lipopolysaccharide (LPS)-induced RAW264.7 macrophages (Table 2). In addition, wild blueberry-enriched diet has protective effects on the pro-inflammatory status related to the MetS in the obese Zucker rat by suppressing liver expression of NF- κ B and increasing adiponectin expression (Table 3) (Seeram et al., 2002; Vendrame et al., 2013). Moreover, cellular and animal models of oxidative stress have also been utilized to prove the antioxidant potential of blueberries (Sellappan et al., 2002).

An *in vivo* study showed that blueberry reduced TGs, body weight gain, liver weight, abdominal fat mass, total fat mass, and

TABLE 2 | *In vitro* studies evaluating the protective and therapeutic effects of anthocyanins in metabolic diseases.

Anthocyanins	Cell culture model	Results	References
Cyanidin-3-O- β -glucoside chloride or Cyaniding chloride	HK-2 cells	\uparrow Cholesterol efflux & ABCA1 expression \uparrow PPAR α & LXR α expressions \downarrow ICAM1, MCP1, TGF β 1 & NF κ B	Du et al., 2015
Malvidin-3-Glucoside and Malvidin-3-Galactoside	HUVEC cells	\downarrow ICAM1, MCP1 & VCAM1 \downarrow I κ B α degradation Blocking block the nuclear translocation of p65	Huang et al., 2014
Delphinidin 3-sambubioside-5-glucoside (D3S5G)	H4IIE hepatoma cells	\downarrow I κ B α degradation \downarrow Gluconeogenic enzyme, glucose-6-phosphatase	Rojo et al., 2012
Pelargonidin	L6 skeletal muscle cell	\uparrow Intracellular glucose uptake \downarrow GLUT4, IRS1, IRS2, PI3, GK & PK	Samadder et al., 2017
Pelargonidin	L6 cells	\downarrow Oxidative damage Activation of DNA repaired cascades	Samadder et al., 2016
Cyanidin-3-glucoside	MIN6N pancreatic β -cells	\downarrow Overproduction of reactive oxygen species \downarrow Apoptosis of cell (under high glucose condition) \uparrow Insulin secretion	Lee et al., 2015
Cyanidin-3-O-b-glucoside	THP-1 cells	\downarrow TNF α & IL-6 expression and secretion Blockage of phosphorylation of I κ B α and NF- κ B nuclear translocation	Zhang et al., 2010
Blueberries and Concord grapes (containing malvidin, petunidin, or peonidin)	Mouse embryonic fibroblast cell line 3T3-L1	\uparrow Basal oxygen consumption rate \downarrow Lipid accumulation \uparrow Mitochondrial respiration	Skates et al., 2017
Bilberry extracts	3T3-L1 cells	Inhibition of 3T3-L1 cells differentiation \downarrow PPAR γ \downarrow Sterol regulatory element-binding protein 1c (Srebp1c) \downarrow Phosphorylation of tyrosine residues of IRS1	Suzuki et al., 2011

HUVECs, Human umbilical vein endothelial cells; PPAR α , Peroxisome proliferator-activated receptor alpha; LXR α , Liver X receptor alpha; ICAM1, Intercellular adhesion molecule-1; MCP1, Monocyte chemoattractant protein-1; TGF β 1, Transforming growth factor- β 1; MCP-1, Monocyte chemoattractant protein-1; ICAM-1, Intercellular adhesion molecule-1; VCAM-1, Vascular cell adhesion molecule-1; GLUT4, Glucose transporter 4; GK, Glucokinase; PK, Pyruvate kinase.

improved adipose and skeletal muscle peroxisome proliferator-activated receptors (PPARs) activities that are involved in glucose uptake/oxidation and fat oxidation in obese rats (Table 3) (Seymour et al., 2008). Furthermore, purified blueberry and blueberry juice ANTs attenuated obesity development, elevated serum leptin, and diabetes in mice fed with obesogenic diet (Wallace et al., 2001; Prior et al., 2010).

Blueberries improve hyperglycemia, regulate skeletal muscle glucose uptake, and decrease liver glucose production *in vivo* (Defuria et al., 2009). DeFuria et al. also reported that blueberries consumption attenuated insulin resistance and insulin sensitivity by reducing adipocyte death in weight gain caused by high-fat diet intake in Male C57BL/6 mice (Defuria et al., 2009). Prior et al., indicated that ANTs fraction of blueberries significantly suppressed body fat accumulation and body weight gain in mice (Prior et al., 2010). Highbush Blueberry (*Vaccinium corombosum*) inhibited α -amylase and α -glucosidase activities *in vitro* and can be considered as an anti-diabetic drug (Johnson et al., 2011). In the study by Flores et al., acetonc extract of whole blueberry mitigated postprandial hyperglycemia via α -glucosidases inhibition (Flores et al., 2013).

ANT-enriched extracts of blueberries attenuated cardiac injury induced by cyclophosphamide *in vivo* condition via reduction of arterial blood pressure, and increases in enzyme

activities and heart rate (Liu et al., 2015). It was shown that consumption of fresh blueberries for 75 days in a high cholesterol diet decreased the accumulation of cholesterol and oxidative stress in the guinea pig's aorta and liver. Consumption of blueberry was found to protect against oxidative stress and free radicals in red blood cells *in vivo* (Coban et al., 2013).

Strawberry (*Fragaria x ananassa*) Anthocyanins

Strawberry is a member of the Rosaceae family with abundant amounts of phenolic compounds, particularly ANTs and ellagic acid (Andersen et al., 2004). Strawberries contain different types of ANTs, such as ascyanidin 3-glucoside, pelargonidin 3-glucoside, pelargonidin3-rutinoside, pelargonidin 3-acetylglucoside, and cyanidin 3-rutinoside. Furthermore, 5 carboxypyranopelargonidin 3-glucoside and four purple ANT flavanol complexes consisting of pelargonidin 3-glucoside were detected in strawberries.

Some of the known cardioprotective agents in strawberries including vitamin C, folic acid, potassium, fiber and phytosterols contribute to the antioxidant, anti-inflammatory, and hypocholesterolemic effects of these fruits (Wang and Lin, 2000).

TABLE 3 | *In vivo* studies on animal models evaluating the protective and therapeutic effects of anthocyanins and anthocyanin-rich extracts in metabolic diseases.

Anthocyanin	Animal model	Results	References
Black chokeberry extract	STZ-induced diabetes in rats and mice	Antidiabetic & hypoglycemic effect by ↑insulin secretion, maintaining the round shape of the pancreas, protecting pancreatic β cells, ↓sucrase & maltase activity, ↓LDL-cholesterol and TG	Jurgonski et al., 2008; Kim et al., 2013; Jeon et al., 2018
Black chokeberry anthocyanins	STZ -induced oxidative stress in male wistar rats	↓Body weight gain, ↓lipase, ↓pancreatic amylase & ↓carbohydrates absorption in the digestive system	Qin and Anderson, 2012
Blueberries anthocyanin	Obesogenic diet mice	↓Abdominal fat mass, ↓total fat mass, ↓body weight gain, ↓TGs, ↓liver weight, ↑PPAR	Seymour et al., 2008; Prior et al., 2010
Blueberries anthocyanin	High-fat diet induced weight gain in Male C57BL/6 mice	↓Insulin resistance, hyperglycemia ↓adipocyte death, ↓body fat accumulation & ↓body weight gain	Defuria et al., 2009; Basu et al., 2010
Blueberries anthocyanin	Cyclophosphamide-induced cardiac injury in rats	Attenuates cardiac injury by ↑heart rate & activities of heart enzymes, ↓IL-1β & TNF-α expression, ↑IL-10	Liu et al., 2015
C3G	High-fat diet induced body fat accumulation C57BL/6J mice	↓Hyperglycemia, ↓blood glucose level & modulates insulin ↓Body fat accumulation via ↓lipid synthesis in the liver and white adipose tissue	Tsuda et al., 2003
<i>Hibiscus sabdariffa</i>	High-fat diet induced obesity and liver damage in hamsters	↓Body weight, fat content & liver fat bodies ↓LDL-C & ↓TGs ↓ALT & ↓AST	Huang et al., 2015
Maqui Berry	Diet-induced obese hyperglycaemic C57BL/6J mice	↓Fasting blood glucose levels & glucose tolerance	Schreckinger et al., 2010; Rojo et al., 2012
Mulberry	High-fat diet in db/db mice	↓Fasting blood glucose, serum insulin, ↓leptin, ↓TGs & cholesterol levels ↓and LDL values ↑adiponectin levels	Yan et al., 2016
Pelargonidin	STZ-induced oxidative stress in diabetic neuropathic rat	↑SOD, malondialdehyde, fructosamine & catalase, ↑thiobarbituric acid reactive substances formation, ↓elevated blood glucose levels	Roy et al., 2008; Mirshekar et al., 2010
Pomegranate seed oil	High-cholesterol diet fed male rats	↓Weight raises & ↓body fat mass	Vroegrijk et al., 2011
Purple sweet potato anthocyanin	STZ-induced insulin deficiency in yellow db/db mice	Induced hypoglycemic activity via ↓oral glucose insulin sensitivity	Ludvik et al., 2004
Purple sweet potato anthocyanin and diacylated ANTs	STZ-induced insulin deficiency in obese Zucker fatty rats	Improved glucose tolerance & diabetes signs via ↓hyperinsulinemia & ↓hyperlipidemia as well as ↓TGs & ↓FFA, ↓maltase and ↓maximal blood glucose level & serum insulin secretion	Kusano et al., 2001
Red onion extract	Diet-induced obese hyperglycaemic C57BL/6J mice	↑Insulin sensitivity via upregulation of energy expenditure & biogenesis of mitochondrial skeletal muscle	Morrison et al., 2015
Red onions	High-fat diet in C57BL/6J mice	Attenuated hyperglycemia & ↑insulin sensitivity via ↑energy expenditure and biogenesis of mitochondrial skeletal muscle, ↑glucose tolerance, protecting DNA from oxidative stress o	Eldin et al., 2010; Jung et al., 2011
Sweet potato	STZ-induced insulin deficiency	↓Hyperlipidemia, ↓TGs & FFA ↓maximal blood glucose level & serum insulin, ↓oral glucose insulin sensitivity ↓maltase inhibitory activity	Kusano and Abe, 2000; Matsui et al., 2002

STZ, Streptozotocin; LDL, Low-density lipoprotein; IL, Interleukin; FFAs, Free fatty acids; PPAR, Peroxisome proliferator-activated receptors; TGs, Triglycerides; TNF, Tumor necrosis factor; SOD, Superoxide dismutase; ALT, Alanine transaminase; AST, Aspartate transaminase.

ANTs in strawberries also reduced obesity in mice, inhibited esophageal cancer, suppressed ox-LDL-induced proliferation, reversed behavioral aging in rats and possessed anticarcinogenic and antithrombotic effects (Wang and Lin, 2000; Qin et al., 2009).

Ox-LDL, a marker of oxidative stress, is elevated in subjects with established coronary heart disease (CHD) and is a prognostic marker for the progression of subclinical atherosclerosis (Toshima et al., 2000).

The anti-hyperglycemic effects of Brazilian strawberries have been reported in *in vitro* model (da Silva Pinto et al., 2008). In mice models, freeze-dried strawberry powder was shown to reduce obesity and improved glycemic control in those fed a high-fat diet while ANT-fed mice demonstrated an upregulation of anti-inflammatory adiponectin gene. Serum cholesterol level was lowered following 4 weeks consumption of freeze-dried strawberries due to the presence of phytosterol, fiber, or other phytochemicals. Suppression of LDL-cholesterol as well as lipid peroxidation was also noted. The antioxidant rich phytochemicals in strawberries have been shown to reduce the central nervous system deficits caused by aging in rat models (Andersen et al., 2004). The cardiovascular health benefits of strawberries were also associated to the reduction of thiobarbituric acid-reactive substances in LDL and decrease in lipids oxidative damage in hyperlipidemic subjects. In addition, strawberries ANTs, such as pelargonidin-3-O-glucoside, reduced postprandial inflammation and increased insulin sensitivity in overweight adults (Wang and Lin, 2000).

Maqui Berry (*Aristotelia chilensis*) Anthocyanins

The fruit from *Aristotelia chilensis* (Molina) Stuntz, commonly known as Maqui Berry, Chilean blackberry or “maqui” in Chile and Argentina, is a common wildberry that belongs to the Elaeocarpaceae (Toshima et al., 2000). Maqui berry has recently been reported as one of the healthiest exotic berries due to its particularly high concentration of bioactive polyphenols (Schreckinger et al., 2010). Studies on the phytochemical composition of Maqui berry have confirmed the presence of phenolic acids, proanthocyanidins, and ANTs such as delphinidin-3-sambubioside-5-glucoside. The leaves and fruits of Maqui berry have been used in folk medicine to treat a variety of ailments including sore throat, kidney pains, ulcers, fever, inflammation, and diarrhea.

In vitro studies have demonstrated that Maqui berry significantly inhibits nitrite oxide production, which is comparable to the effect exerted by quercetin, a potent anti-inflammatory agent via inhibition of prostaglandin E2 and the COX-2 on LPS-stimulated RAW 264.7 macrophages (Morazzoni and Bombardelli, 1996; Schreckinger et al., 2010). In another *in vitro* study, the extract of Maqui berry suppressed the production of glucose and attenuated the downregulation of gluconeogenic enzyme and glucose-6-phosphatase. Moreover, oral administration of delphinidin 3-sambubioside-5-glucoside decreased the fasting blood glucose in obese C57BL/6J mice (Pergola et al., 2006; Rojo et al., 2012) and can be a therapeutic agent for managing MetS and diabetes.

The Maqui berry also showed cardioprotective effect against ischaemia–reperfusion heart damage in mice. Maqui berry possessed antioxidant activity and the highest oxygen radical absorbance capacity (ORAC) by inhibiting LDL oxidation and adipogenesis, also played a protecting role against intracellular oxidative stress in human endothelial cells (Schreckinger et al., 2010).

Black Chokeberry Anthocyanins (*Aronia melanocarpa*)

Aronia melanocarpa is one of the richest plant sources of polyphenolic substances, especially ANT glycosides with the highest antioxidant capacity among the polyphenol-rich beverages (Kulling and Rawel, 2008). Black chokeberry decreases weight gain, attenuates insulin resistance, reduces adipogenesis, and plasma concentrations of total cholesterol, LDL-cholesterol, and TGs. *In vitro* experiments showed that the phenolic constituents of black chokeberry exhibited anti-platelet effects as well as vasoactive and vasoprotective properties in porcine coronary arteries (D’alessandro et al., 2012). *In vivo* studies have shown that black chokeberry extract significantly exhibited hypoglycemic and antidiabetic effect by inducing the glucose uptake and glycogen synthesis, increasing insulin secretion, and protecting pancreatic β cells in streptozotocin (STZ)-induced oxidative stress in male wistar rats (Renaud and De Lorgeril, 1992; Valcheva-Kuzmanova and Belcheva, 2006; Jurgonski et al., 2008).

Qin and Anderson reported that diet supplemented with chokeberry extract reduced body weight gain significantly after 4 weeks via lipase and pancreatic amylase inhibition along with reducing carbohydrates absorption in the digestive system (Qin and Anderson, 2012).

Frejnage and Zduńczyk suggested that diets supplemented with 0.4, 0.8, and 1.2% of chokeberry extract suppressed the prooxidative activity *in vivo* by reducing blood malonyldialdehyde content in rats (Frejnage and Zduńczyk, 2008). Olas et al. reported that ANTs of *Aronia* attenuated lipid peroxidation and possessed antioxidative activity in peroxynitrite induced stress *in vitro*. It may be helpful in managing the reduction-oxidation (redox) homeostasis disturbance by inhibiting nuclear factor (NF)- κ B and increasing glutathione peroxidase activity, which confirms the beneficial effect of *Aronia melanocarpa* in patients with MetS and diabetes (Olas et al., 2008). Kim et al. reported that *Aronia* modulated hepatic lipid metabolism and improved antioxidant function in mice (Simeonov et al., 2002; Kim et al., 2013).

It has been found that black chokeberry extract significantly exhibited hypoglycemic and antidiabetic effect *in vivo* by induction of glucose uptake and glycogen synthesis and by elevating insulin secretion. It also helped to maintain the round shape of the pancreas and protected the pancreatic β cells in STZ-induced oxidative stress along with defeating sucrase and maltase activities in male wistar rats (Jeon et al., 2018).

Mulberry Anthocyanins

Mulberry contains water soluble ANTs, such as cyanidin-3-glucoside (47%), cyanidin-3-rutinoside (27%), and

pelargonidin-3-glucoside (1.4%), which has been traditionally used in Chinese medicines. Mulberry was shown to have great antioxidant, anti-inflammatory, and anti-cancer activities in both cultured cells and animal models (Hassimotto et al., 2008; Huang et al., 2013). The dietary supplements with mulberry ANTs mitigate adverse effects of high-glucose against diet-induced obesity in C57BL/6 mice. Yan et al. demonstrated that mulberry ANTs reduced fasting blood glucose, serum insulin and leptin, as well-modulated TGs, cholesterol, and LDL values in high-fat diet in db/db mice (Yan et al., 2016).

Purple Sweet Potato (*Ipomoea batatas*) Anthocyanin

The sweet potato (*Ipomoea batatas*), is a dicotyledonous plant that belongs to Convolvulaceae family; it is large, starchy, and sweet-tasting, making it consumed as a food additive for the prevention and care of type 2 diabetes, anemia, and hypertension. Sweet potato contains a variety of ANTs. An *in vivo* study of sweet potato demonstrated that oral administration improved diabetes, glucose tolerance, hyperinsulinemia, and hyperlipidemia, also lowered TGs and free fatty acid in Zucker fatty rats (Kusano and Abe, 2000). It also exhibited hypoglycemic activity in STZ-induced insulin deficiency in yellow db/db mice. Ludvik et al., observed a reduction in oral glucose insulin sensitivity following Caiapo treatment (Ludvik et al., 2008). Matsui et al. showed that *in vivo* oral administration of the diacylated ANTs derived from *I. batatas* in rats exhibited a potent maltase inhibitory activity, and significantly decreased maximal blood glucose level and serum insulin secretion compared to vehicle treatment (Matsui et al., 2002). It has also been shown that oral administration of Caiapo 4 g/day for 6 weeks lowered total and LDL cholesterol levels as well as blood glucose by increasing insulin sensitivity without affecting insulin secretion. Administration of *I. batatas* could significantly increase the level of adiponectin, which is produced by adipocytes and acts as a modulator of insulin sensitivity (Ludvik et al., 2008).

Pomegranate Seed Anthocyanins

Pomegranate (*Punica granatum*, Punicaceae) is an edible fruit comprising of 80% juice and 20% seed, and cultivated in Mediterranean countries, China, Japan, Russia and the United States. ANTs detected in pomegranate include pelargonidin 3-glucoside, cyanidin 3-glucoside, delphinidin 3-glucoside, pelargonidin 3,5-diglucoside, cyanidin 3,5-diglucoside, and delphinidin 3,5-diglucoside. The proanthocyanidins and ANTs of this plant were found to show antiangiogenic, antioxidant, anti-carcinogenic, and antimicrobial activities, besides, these compounds were shown to inhibit the activities of cyclooxygenase (COX), nitric oxide, and epidermal growth factor receptor (Bagchi et al., 2004; Vasconcelos et al., 2006).

Studies have reported that pomegranate fruit extract demonstrated anti-inflammatory activity by modulating the production of prostaglandin and leukotriene along with inhibition of COX and lipoxygenase. α -Tocopherol from seeds of this plant inhibited sphingolipid synthesis and COX-2 activity. Recent studies have shown that pomegranate wine can inhibit

NF- κ B in vascular endothelial cells. Dietary utilization of pomegranate juice significantly diminished the atherosclerotic lesions formation and decreased LDL oxidation in atherosclerotic mice (Schubert et al., 1999; Aviram et al., 2000; Gil et al., 2000; Aviram and Dornfeld, 2001; Kaplan et al., 2001; Chidambara Murthy et al., 2002). It has shown that pomegranate seed oil can reduce weight gains and food consumption in male rats fed a high-cholesterol diet. Vroegrijk et al. also observed a significant reduction in body fat mass in male C57Bl/J6 mice fed with a high-fat diet (Vroegrijk et al., 2011).

Red Onions (*Allium cepa*) Anthocyanins

Red onions (*Allium cepa*), a widely consumed vegetable with purplish-red skin which comes from anthocyanidins such as cyanidin, belongs to Liliaceae family native of Southwest Asia, and is widely cultivated throughout the world. Red onions are an abundant source of flavonols, including quercetin derivatives, such as quercetin glycosides, and ANTs (Kaplan et al., 2001). Onion significantly decreased the levels of total cholesterol and LDL, and attenuated hypertension and blood cholesterol in diabetic animal models (Kumari and Augusti, 2002). Red onion supplementation attenuated high-fat diet-induced insulin resistance in C57BL/6J mice by limiting adiposity and increasing energy expenditure (Eldin et al., 2010; Jung et al., 2011).

Several *in vivo* studies showed that onion ingestion improves hyperglycemia in diabetic patients via increasing insulin sensitivity, improving glucose tolerance, and protecting DNA from oxidative stress in mice (Mathew and Augusti, 1975; Corzo-Martínez et al., 2007). Morrison et al. explained that the reduction of obesity and improvement of insulin sensitivity might be related to the upregulation of energy expenditure and biogenesis of mitochondrial skeletal muscle in C57BL/6J mice upon red onion extract supplementation (Morrison et al., 2015). These reports confirmed the therapeutic effects of *A. cepa* in patients with MetS and diabetes.

Purified Anthocyanins Cyanidin 3-Glucoside (C3G)

The most common anthocyanidin, cyanidin, is present in 90% of fruits. It is absorbed into blood circulation in an intact form and metabolized to methoxy derivatives in the liver and kidney, and its metabolites may modulate metabolic effects. Studies have shown that the antioxidant activity of cyanidin was more than that of vitamin E and Trolox, and was comparable to that of synthetic antioxidants, such as tert-butylhydroquinone (TBHQ), butylated hydroxytoluene (BHT), and butylated hydroxyanisole (BHA) likely because of free hydroxyl groups on the 3' and 4' positions of cyanidin (Amorini et al., 2001).

It has been proven that C3G has antioxidative and anti-inflammatory properties based on *in vitro* and *in vivo* studies. C3G significantly suppressed the development of high-fat diet induced obesity C57BL/6 mice and modulated the gene expression of adipocytokines in human adipocytes, and reduced inflammation and adipocyte death, but not adipocyte size in high-fat diet mice *in vivo* (Tsuda et al., 2003). C3G also diminished inflammation in isolated vascular endothelial cells and monocytes *in vitro* and possessed an insulin-like effect

in human omental adipocytes and 3T3-L1 cells. Attenuation of gene expression of adipocytokines is also seen in human adipocytes. Other studies have reported that C3G or its aglycone induced upregulation of adiponectin, which enhanced insulin sensitivity in isolated rat and human adipocytes, but these events were not observed *in vivo*. C3G efficiently inhibited free fatty acids (FFAs) and glycerol release from the adipocytes during hyperglycemia in high glucose-induced lipolysis in cultured 3T3-L1 adipocytes. It also increased the activity of AMPK and decreased the activity of glutamine, fructose 6-phosphate, and aminotransferase. C3G reduced hyperglycemia-promoted O-glycosylation of transcription factor Foxo1, resulting in decreased expression of adipose triglyceride lipase. Purple corn is a source of C3G, and has been shown to decrease body fat and hyperglycemia in high-fat diet mice. In another study, C3G increased adipocyte glucose uptake and GLUT4 membrane translocation significantly. Nuclear PPAR γ activity was increased as well as adiponectin (Scazzocchio et al., 2011). C3G isolated from mulberry fruits possesses an antidiabetic effect via decreasing oxidative stress, and increasing antioxidant defense system and cytoprotective activity during glucose-induced apoptosis in MIN6N pancreatic β -cells by depleting generation of ROS, DNA fragmentation, and the rate of apoptosis (Lee et al., 2015). Pure C3G also increases in cholesterol efflux, ABCA1 expression, PPAR α , LXRA, and decreases in proinflammatory molecules, such as ICAM1, MCP1, TGF β 1, and NF- κ B in HK-2 cells (Du et al., 2015). Zhang et al. suggested that I κ -B α and NF- κ B nuclear translocation have a significant role in therapeutic effects of C3G (Zhang et al., 2010). C3G significantly suppressed body fat accumulation induced by a high-fat diet, which is attributed to a reduction in lipid synthesis in the liver and white adipose tissue in C57BL/6J mice. It has also been demonstrated that C3G significantly ameliorated hyperglycemia and insulin sensitivity *in vivo* (Tsuda et al., 2003). A recent *in vivo* study reported that C3G inhibited release of purified platelet granule and protected against CVD and thrombosis (Zhou et al., 2017). Furthermore, it also diminishes blood glucose level and modulates insulin sensitivity in type 2 diabetic mice. It has also been proven that C3G had synergistic effect with acarbose, an inhibitor of α -glucosidase used in the treatment of diabetes. Furthermore, C3G treatment resulted in increased insulin secretion compared with the control diabetic group, and is a potential phytotherapeutic agent for the prevention of diabetes (Zhou et al., 2017).

Cyanidin-3-glucoside and peonidin-3-glucoside in black rice may decrease antioxidant and anti-inflammation activity by protecting against oxidative damage and suppressing nitric oxide synthase in mouse macrophage cell linings (Hu et al., 2003).

Pelargonidin

Lamy et al., showed that delphinidin inhibited phosphorylation of vascular endothelial growth factor (VEGF) receptor-2 in human umbilical vascular endothelial cells (Tonelli et al., 2009). *In vitro* studies showed that a pelargonidin derivative enhanced insulin secretion by β -cells and could be a good anti-diabetic agent via suppression of fasting blood glucose level almost to half of the pretreatment levels. Furthermore, urine sugar decreased

to (non-significant/minor) traces and appeared healthy (Cherian et al., 1992) NF- κ B.

In vivo studies showed that pelargonidin significantly ameliorated the alteration in hyperalgesia through attenuation of oxidative stress in STZ-diabetic neuropathic rat. This compound also diminished diabetes-induced thiobarbituric acid reactive substances formation and reduce antioxidant defensive enzyme superoxide dismutase (Mirshekar et al., 2010). Roy et al., reported that pelargonidin normalized elevated blood glucose levels, improved serum insulin levels, decreased catalase and SOD, and enhanced fructosamine and malondialdehyde levels in diabetic rats (Roy et al., 2008).

Peonidin and Malvidin

Consumption of berries containing (57% malvidin and 33% petunidin or peonidin) was effective to reduce high-fat diet induced metabolic damage through individual significant effects on energy expenditure and increased activity by decreasing mitochondrial respiration and dissipation of the mitochondrial proton gradient (proton leak) in adipose tissue in C57BL/6 mouse model of polygenic obesity (Skates et al., 2017). Bogнар et al. demonstrated that malvidin attenuated LPS-induced NF- κ B, activation of mitogen activated protein kinase (MAPK), poly ADP-ribose polymerase, production of ROS, and depolarization of mitochondria (Bogнар et al., 2013).

CLINICAL STUDIES CONFIRMING THE BENEFICIAL EFFECTS OF ANTS IN METS

ANTS are generally considered as safe remedy without considerable adverse effects and a wide range of pharmacological activities. Studies showed that ANTs have explicit useful effects on MetS features. However, studies on the effects of ANTs in prevention and treatment of MetS in human subjects are limited. The following studies demonstrated that regular consumption of ANTs diet may show protective effects in management and prevention of MetS.

In a systematic review and meta-analysis of 32 clinical studies, it was revealed that ANT-rich food can exert promising preventive and protective effects against cardiometabolic disorders. ANTs substantially decreased glycemic control markers, enhanced fasting and 2-h postprandial glucose level, and possessed favorable effects on controlling the LDL level (Yang et al., 2017).

Two-month *Aronia* extract therapy resulted in considerable decline in systolic and diastolic blood pressure, besides suppressed the LDL, TGs, and Endothelin-1 in 47 subjects (32 women, 15 men). Moreover, *Aronia* fruit juice reduced the elevated cholesterol, LDL, and plasma lipids concentration (Broncel et al., 2007).

A randomized controlled study on 48 participants with MetS, who have been treated with freeze-dried and fresh blueberries for 8 weeks daily exposed that blueberry beverage significantly decreased the plasma oxidized LDL, diastolic, and systolic blood pressure, also ameliorated the serum malondialdehyde and hydroxynonenal levels (Basu et al., 2010).

Regular chokeberry juice drinking (250 mL per day) was recorded to decrease LDL and TGs, increased HDL cholesterol level and led to significant reduction in glucose serum, homocysteine and fibrinogen in men with mild hypercholesterolaemia (Skoczynska et al., 2007). Black chokeberry also decreased glucose concentration and fasting blood glucose in human studies. In addition, it may also diminish oxidative and/or nitrate stress that occurred in platelets from breast cancer patients (Olas et al., 2008). Maqui Berry ANTs consumption in subjects with hyperlipidemia and dyslipidemia depressed LDL and VLDL and increased HDL cholesterol (Alvarado et al., 2016). Delphinol is a proprietary Maqui berry extract with a standardized content of 25% w/w delphinidin glycosides and 35% total ANTs that can significantly inhibit postprandial blood glucose (Hidalgo et al., 2014).

In a factorial randomized design study, 1-month therapy with *Hibiscus sabdariffa* extract powder significantly enhanced HDL-c levels, amended the ratio of TAG/HDL-c, reduced glucose and total cholesterol levels as well as triglycerides in MetS patients (Gurrola-Díaz et al., 2010).

In another randomized controlled trial on 27 subjects with MetS, 4 cups of freeze-dried strawberry beverage daily for 8 weeks, caused hypocholesterolemic effects in study subjects through decreasing the total and LDL-cholesterol levels along with suppression of VCAM-1 circulating levels (Basu et al., 2010). Another study on eight elderly women exhibited that consumption of strawberries, red wine, spinach, or vitamin C can increase human serum antioxidant capacity (Cao et al., 1998).

The simultaneous consumption of blackcurrant ANTs and apple polyphenols was the subject of a clinical study on five postmenopausal women and 20 men, investigating the effect of this mixture completed with a meal containing starch and sucrose on the initial postprandial glycemic response. The mixture was found to be effective in inhibition of the early responses (0–30 min) of plasma glucose and insulin, and reduction of postprandial glycemia. Insulin and incretin excretion were reduced as the secondary results. The promising inhibitory role of ANT and proanthocyanidin-riched diets on the negative effects of high-carbohydrate meals was highlighted by this study (Castro-Acosta et al., 2017). Modulation of lipid and glucose-metabolism, antioxidative, and anti-inflammatory activities were the other outcomes of the inclusion of ANTs in human diets. These findings have been corroborated in an investigation carried out by Kim et al., who chose Açai berries as a rich source of ANTs to be consumed by 37 subjects (12 weeks) suffering from MetS (Kim et al., 2018). In result, the plasma levels of interferon gamma (IFN- γ) and urinary level of 8-isoprostane were decreased. However, all parameters related to the glucose and lipid metabolisms were found to be unchanged after intake of the beverage (Kim et al., 2018). Although their study verified the health-promoting effect of ANT-rich diets in metabolically challenged humans, further clinical investigations are required to warranty these specific results.

A search in www.clinicaltrials.gov has shown that many completed and ongoing clinical trials are evaluating the therapeutic potential of ANTs for the treatment of fatty liver

TABLE 4 | Completed and ongoing clinical trials.

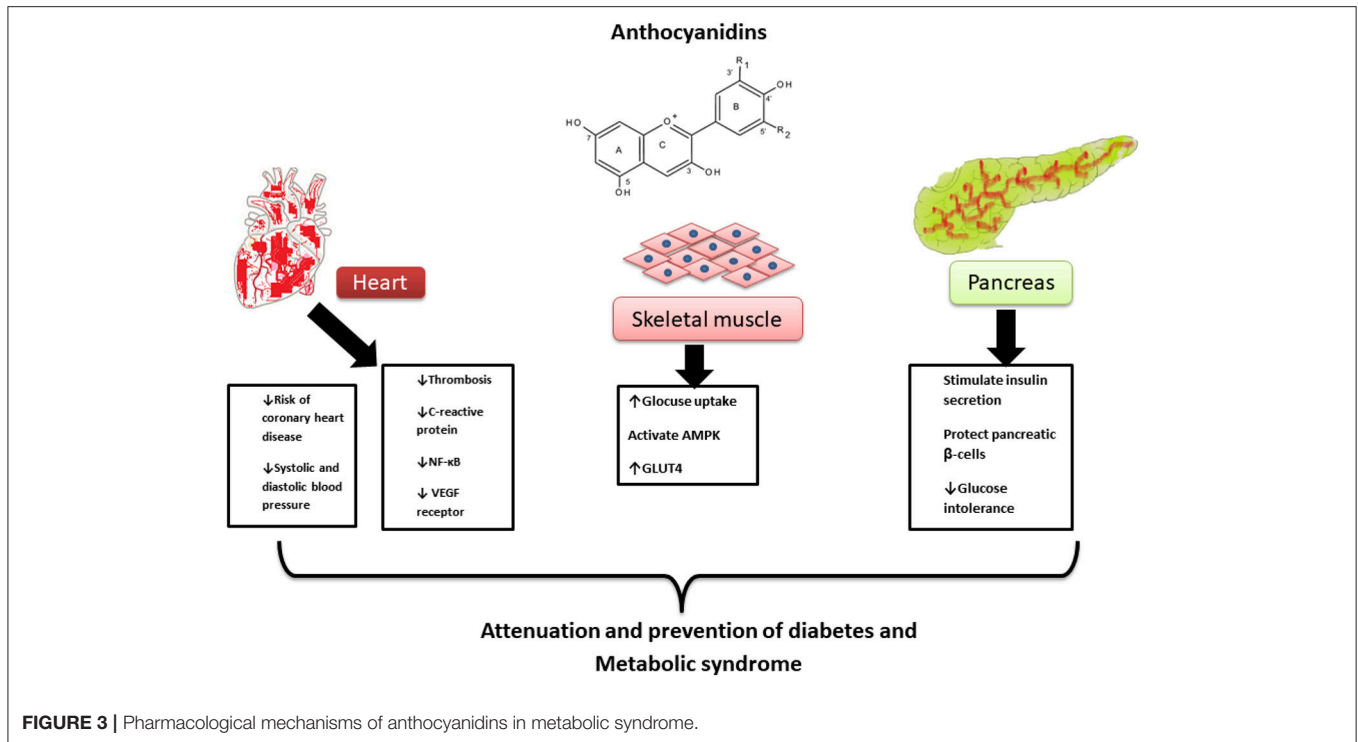
Number	Title
NCT02407522	The improvements of dietary supplement of black rice on MetS (IDSBRMS)
NCT02999256	Effect of cherry juice on fat oxidation and cardio-metabolic markers
NCT01399138	The effect of blueberry powder supplementation on cardiovascular risk factors in subjects with the MetS
NCT00992641	The effect of nordic recommended diet on the features of MetS
NCT01562392	Effects of berries and vegetables on cardiometabolic risk markers and cognitive function
NCT01414647	The health effect of diet rich in nordic berries (berry)
NCT01224743	Effect of fruit and vegetable concentrates on endothelial function in persons with MetS
NCT01154478	Effects of dietary polyphenols and ω -3 fatty acids on cardiovascular risk factors in high risk subjects (Etherpaths)
NCT02035592	The health effects of blueberry ANTs in MetS (ongoing)
NCT01766570	Beneficial effects of a polyphenol enriched beverage on type 2 diabetes prevention and on cardiovascular risk profile of men and women with insulin resistance.
NCT01245270	A single supplement of a standardized bilberry extract modifies glycaemic response
NCT01180712	Study of oral ants on insulin resistance
NCT01860547	Effects of berries and berry fractions on metabolic diseases
NCT02689765	Effect of ants on metabolic profiles in subjects with pre-diabetes
NCT02779985	Goji berries and energy expenditure
NCT02017132	Effect of pomegranate extract intake on body composition and blood pressure.
NCT01568983	The effects of polyphenol-rich berry juice on blood pressure in hypertensive subjects
NCT02459756	Ant-rich blackcurrant and vascular function

disease, CVD, MetS, coronary artery disease, and type 2 diabetes. Retrieved clinical trials registered in www.clinicaltrials.gov are summarized in **Table 4**.

CONCLUSION AND FUTURE PROSPECTS

MetS is closely related to obesity and has a major role in initiating CVD and many other pathological complications of type 2 diabetes.

Diet has an important role in disease management and prevention. ANTs have antioxidant properties and possess a protective role for pancreatic β -cells from glucose-induced oxidative stress. They also act as anti-inflammatory and hypotensive agents (Lietti et al., 1976). Additionally, it has been reported that ANTs cause a reduction in concentrations of TC, LDL-C, and TGs, as well suppress the expression of enzymes responsible for fatty acid synthesis. ANTs protect against CVD, cancer and diabetes; also attenuate the symptoms of MetS such as dyslipidaemia, insulin resistance, impaired glucose tolerance, hypertension, hyperglycemia, and glucosuria. ATNs also inhibit the activity of α -glucosidase against maltase and sucrose, and increase the excretion of insulin in primary



cell culture. ANT-rich extracts showed a lowering effect on plasma lipid profiles in rodent models of hyperlipidaemia. ANTs in chokeberry and purple maize reduced visceral adiposity, systolic blood pressure, and total body fat, moreover, reduced the glucose tolerance, liver, and cardiovascular structure and function. **Figure 3** shows the main pharmacological mechanisms of ANTs in MetS.

This review focused on a group of dietary ANTs, and their positive effect in human health. Our presentation demonstrated that more investigations into the efficacy and intracellular mechanisms of dietary ANTs are necessary to recognize the metabolism, bioefficacy, and main mechanisms of action in MetS. Current evidence establishes that dietary ANTs and its pharmaceutical supplements can be considered as a functional nutritional supplement for the prevention and management of metabolic syndrome and its complications.

REFERENCES

- Abete, I., Goyenechea, E., Zulet, M., and Martinez, J. (2011). Obesity and metabolic syndrome: potential benefit from specific nutritional components. *Nutr. Metab. Cardiovasc. Dis.* 21, B1–B15. doi: 10.1016/j.numecd.2011.05.001
- Alberti, K. G. M., Zimmet, P., and Shaw, J. (2005). The metabolic syndrome a new worldwide definition. *Lancet* 366, 1059–1062. doi: 10.1016/S0140-6736(05)67402-8
- Alvarado, J., Schoenlau, F., Leschot, A., Salgad, A., and Portales, P. V. (2016). Delphinol (R) standardized Maqui berry extract significantly lowers blood glucose and improves blood lipid profile in prediabetic individuals in three-month clinical trial. *Panminerva Med.* 58, 1–6.
- American Diabetes Association (2009). *Introduction*. American Diabetes Association.
- Amiot, M., Riva, C., and Vinet, A. (2016). Effects of dietary polyphenols on metabolic syndrome features in humans: a systematic review. *Obes. Rev.* 17, 573–586. doi: 10.1111/obr.12409
- Amirkalali, B., Fakhrzadeh, H., Sharif, F., Kelishadi, R., Zamani, F., Asayesh, H., et al. (2015). Prevalence of metabolic syndrome and its components in the Iranian adult population: a systematic review and meta-analysis. *Iran. Red Crescent Med. J.* 17:e24723. doi: 10.5812/ircmj.24723
- Amorini, A. M., Fazzina, G., Lazzarino, G., Tavazzi, B., Di Pierro, D., Santucci, R., et al. (2001). Activity and mechanism of the antioxidant properties of cyanidin-3-O-β-glucopyranoside. *Free Radic. Res.* 35, 953–966. doi: 10.1080/10715760100301451

AUTHOR CONTRIBUTIONS

RN and FF did a literature review and prepared the first draft of the manuscript. PH, SN, SH, DT, and SM edited the manuscript and proposed/included some vital modifications. MF and RK design throughout the work and did the final edition of the manuscript.

FUNDING

This work will be supported, financially, by Kermanshah University of Medical Sciences.

ACKNOWLEDGMENTS

Authors thank Kelsi Yu (University of Southern California) for English language revision.

- Andersen, M., Fossen, T., Torskangerpoll, K., Fossen, A., and Hauge, U. (2004). Anthocyanin from strawberry (*Fragaria ananassa*) with the novel aglycone, 5-carboxypyranopelargonidin. *Phytochemistry* 65, 405–410. doi: 10.1016/j.phytochem.2003.10.014
- Anderson, and Jordheim, M. (2008). "Anthocyanins: food applications," in *Proceedings of the 5th Int. Congr. Pigments Foods: For Quality and Health* (Helsinki), 14–16.
- Aviram, M., and Dornfeld, L. (2001). Pomegranate juice consumption inhibits serum angiotensin converting enzyme activity and reduces systolic blood pressure. *Atherosclerosis* 158, 195–198. doi: 10.1016/S0021-9150(01)00412-9
- Aviram, M., Dornfeld, L., Rosenblat, M., Volkova, N., Kaplan, M., Coleman, R., et al. (2000). Pomegranate juice consumption reduces oxidative stress, atherogenic modifications to LDL, and platelet aggregation: studies in humans and in atherosclerotic apolipoprotein E-deficient mice. *Am. J. Clin. Nutr.* 71, 1062–1076. doi: 10.1093/ajcn/71.5.1062
- Bagchi, D., Sen, C., Bagchi, M., and Atalay, M. (2004). Anti-angiogenic, antioxidant, and anti-carcinogenic properties of a novel anthocyanin-rich berry extract formula. *Biochemistry* 69, 75–80. doi: 10.1023/B:BIRY.0000016355.19999.93
- Barazzoni, R., Cappellari, G. G., Ragni, M., and Nisoli, E. (2018). Insulin resistance in obesity: an overview of fundamental alterations. *Eat. Weight Disord.* 23, 149–157. doi: 10.1007/s40519-018-0481-6
- Basu, A., Fu, D. X., Wilkinson, M., Simmons, B., Wu, M., Betts, N. M., et al. (2010). Strawberries decrease atherosclerotic markers in subjects with metabolic syndrome. *Nutr. Res.* 30, 462–469. doi: 10.1016/j.nutres.2010.06.016
- Berlin, E., Nguyen, P., Guenounou, M., Durlach, V., Potron, G., and Leutenegger, M. (2000). Plasma levels of tumor necrosis factor- α (TNF- α) are essentially dependent on visceral fat amount in type 2 diabetic patients. *Diabetes Metab.* 26, 178–183.
- Bognar, E., Sarszegi, Z., Szabo, A., Debreceni, B., Kalman, N., Tucsek, Z., et al. (2013). Antioxidant and anti-inflammatory effects in RAW264. 7 macrophages of malvidin, a major red wine polyphenol. *PLoS ONE* 8:e65355. doi: 10.1371/journal.pone.0065355
- Broncel, M., Kozirog-Kolasiska, M., Andryshowski, G., Duchnowicz, P., Koter-Michalak, M., Owczarczyk, A., et al. (2007). Effect of anthocyanins from *Aronia melanocarpa* on blood pressure, concentration of endothelin-1 and lipids in patients with metabolic syndrome. *Pol. Merk. Lekarski* 23, 116–119.
- Bub, A., Watzl, B., Heeb, D., Reckemmer, G., and Briviba, K. (2001). Malvidin-3-glucoside bioavailability in humans after ingestion of red wine, dealcoholized red wine and red grape juice. *Eur. J. Nutr.* 40, 113–120. doi: 10.1007/s003940170011
- Burns, J., Gardner, P. T., O'neil, J., Crawford, S., Morecroft, I., Mcphail, D. B., L et al. (2000). Relationship among antioxidant activity, vasodilation capacity, and phenolic content of red wines. *J. Agric. Food Chem.* 48, 220–230. doi: 10.1021/jf9909757
- Cao, G., Russell, R. M., Lischner, N., and Prior, R. L. (1998). Serum antioxidant capacity is increased by consumption of strawberries, spinach, red wine or vitamin C in elderly women. *J. Nutr.* 128, 2383–2390. doi: 10.1093/jn/128.12.2383
- Carr, D. B., Utschneider, K. M., Hull, R. L., Kodama, K., Retzlaff, B. M., et al. (2004). Intra-abdominal fat is a major determinant of the National Cholesterol Education Program Adult Treatment Panel III criteria for the metabolic syndrome. *Diabetes* 53, 2087–2094. doi: 10.2337/diabetes.53.8.2087
- Castro-Acosta, M. L., Stone, S. G., Mok, J. E., Mhajan, R. K., Fu, C. I., and Lenihan-Geels, G. N., et al. (2017). Apple and blackcurrant polyphenol-rich drinks decrease postprandial glucose, insulin and incretin response to a high-carbohydrate meal in healthy men and women. *J. Nutr. Biochem.* 49, 53–62. doi: 10.1016/j.jnutbio.2017.07.013
- Cerdá, B., Espín, J. C., Parra, S., Martínez, P., and Tomás-Barberán, F. A. (2004). The potent *in vitro* antioxidant ellagitannins from pomegranate juice are metabolized into bioavailable but poor antioxidant hydroxy-6H-dibenzopyran-6-one derivatives by the colonic microflora of healthy humans. *Eur. J. Nutr.* 43, 205–220. doi: 10.1007/s00394-004-0461-7
- Cherian, S., Kumar, R. V., Augusti, K., and Kidwai, J. (1992). Antidiabetic effect of a glycoside of pelargonidin isolated from the bark of *Ficus bengalensis* Linn. *Indian J. Biochem. Biophys.* 29, 380–382.
- Chidambara Murthy, K. N., Jayaprakasha, G. K., and Singh, R. P. (2002). Studies on antioxidant activity of pomegranate (*Punica granatum*) peel extract using *in vivo* models. *J. Agric. Food Chem.* 50, 4791–4795. doi: 10.1021/jf0255735
- Coban, J., Evran, B., Özkan, F., Çevik, A., Dogru-Abbasoglu, S., and Uysal, M. (2013). Effect of blueberry feeding on lipids and oxidative stress in the serum, liver and aorta of guinea pigs fed on a high-cholesterol diet. *Biosci. Biotechnol. Biochem.* 77, 389–391. doi: 10.1271/bbb.120722
- Corzo-Martínez, M., Corzo, N., and Villamiel, M. (2007). Biological properties of onions and garlic. *Trends Food Sci. Technol.* 18, 609–625. doi: 10.1016/j.tifs.2007.07.011
- Czank, C., Cassidy, A., Zhang, Q., Morrison, D. J., Preston, T., and Kroon, P. A., et al. (2013). Human metabolism and elimination of the anthocyanin, cyanidin-3-glucoside: a ¹³C-tracer study. *Am. J. Clin. Nutr.* 97, 995–1003. doi: 10.3945/ajcn.112.049247
- da Silva Pinto, M., Kwon, Y. I., Apostolidis, E., Lajolo, F. M., Genovese, M. I., and Shetty, K. (2008). Functionality of bioactive compounds in Brazilian strawberry (*Fragaria ananassa* Duch.) cultivars: evaluation of hyperglycemia and hypertension potential using *in vitro* models. *J. Agric. Food Chem.* 56, 4386–4392. doi: 10.1021/jf0732758
- D'alessandro, L. G., Kriaa, K., Nikov, I., and Dimitrov, K. (2012). Ultrasound assisted extraction of polyphenols from black chokeberry. *Separ. Purif. Technol.* 93, 42–47. doi: 10.1016/j.seppur.2012.03.024
- Day, C. (2007). Metabolic syndrome, or What you will: definitions and epidemiology. *Diabetes Vasc. Dis. Res.* 4, 32–38. doi: 10.3132/dvdr.2007.003
- Defuria, J., Bennett, G., Strissel, K. J., Perfield, J. W., Milbury, P. E., and Greenberg, A. S., et al. (2009). Dietary blueberry attenuates whole-body insulin resistance in high fat-fed mice by reducing adipocyte death and its inflammatory sequelae. *J. Nutr.* 139, 1510–1516. doi: 10.3945/jn.109.105155
- Deng, G.-F., Xu, X.-R., Zhang, Y., Li, D., Gan, R.-Y., and Li, H.-B. (2013). Phenolic compounds and bioactivities of pigmented rice. *Crit. Rev. Food Sci. Nutr.* 53, 296–306. doi: 10.1080/10408398.2010.529624
- Desroches, S., and Lamarche, B. (2007). The evolving definitions and increasing prevalence of the metabolic syndrome. *Appl. Physiol. Nutr. Metab.* 32, 23–32. doi: 10.1139/h06-095
- Di Gregorio, G. B., Yao-Borengasser, A., Rasouli, N., Varma, V., Lu, T., Miles, L. M., Ranganathan, G., et al. (2005). Expression of CD68 and macrophage chemoattractant protein-1 genes in human adipose and muscle tissues. *Diabetes* 54, 2305–2313. doi: 10.2337/diabetes.54.8.2305
- Dreiseitel, A., Schreier, P., Oehme, A., Locher, S., Rogler, G., and Piberger, H., et al. (2008). Inhibition of proteasome activity by anthocyanins and anthocyanidins. *Biochem. Biophys. Res. Commun.* 372, 57–61. doi: 10.1016/j.bbrc.2008.04.140
- Du, C., Shi, Y., Ren, Y., Wu, H., Yao, F., and Wei, J., et al. (2015). Anthocyanins inhibit high-glucose-induced cholesterol accumulation and inflammation by activating LXR α pathway in HK-2 cells. *Drug Des. Devel. Ther.* 9, 5099–5113. doi: 10.2147/DDDT.S90201
- Eldin, I. M. T., Ahmed, E. M., and Abd, E. H. (2010). Preliminary study of the clinical hypoglycemic effects of *Allium cepa* (red onion) in type 1 and type 2 diabetic patients. *Environ. Health Insights* 4, 71–77. doi: 10.4137/EHI.S5540
- Elks, C. M., Terrebonne, J. D., Ingram, D. K., and Stephens, J. M. (2015). Blueberries improve glucose tolerance without altering body composition in obese postmenopausal mice. *Obesity* 23, 573–580. doi: 10.1002/oby.20926
- Esposito, K., Marfella, R., Ciotola, M., Di Palo, C., Giugliano, F., Giugliano, G., et al. (2004). Effect of a Mediterranean-style diet on endothelial dysfunction and markers of vascular inflammation in the metabolic syndrome: a randomized trial. *JAMA* 292, 1440–1446. doi: 10.1001/jama.292.12.1440
- Fang, J. (2015). Classification of fruits based on anthocyanin types and relevance to their health effects. *Nutrition* 31, 1301–1306. doi: 10.1016/j.nut.2015.04.015
- Farzaei, F., Morovati, M. R., Farjadmand, F., and Farzaei, M. H. (2017). A mechanistic review on medicinal plants used for diabetes mellitus in traditional persian medicine. *J. Evid. Based Complementary Altern. Med.* 22, 944–955. doi: 10.1177/2156587216686461
- Farzaei, M. H., El-Senduny, F. F., Momtaz S., Parvizi F, Iranpanah A, Tewari D, et al. (2018). An update on dietary consideration in inflammatory bowel disease: anthocyanins and more. *Expert. Rev. Gastroenterol. Hepatol.* 12, 1007–1024. doi: 10.1080/17474124.2018.1513322
- Favaro-Trindade, C., Santana, A., Monterrey-Quintero, E., Trindade, M., and Netto, F. (2010). The use of spray drying technology to

- reduce bitter taste of casein hydrolysate. *Food Hydrocoll.* 24, 336–340. doi: 10.1016/j.foodhyd.2009.10.012
- Felgines, C., Texier, O., Besson, C., Fraisse, D., Lamaison, J.-L., and Remesy, C. (2002). Blackberry anthocyanins are slightly bioavailable in rats. *J. Nutr.* 132, 1249–1253. doi: 10.1093/jn/132.6.1249
- Fernandes, I., Faria, A., Calhau, C., De Freitas, V., and Mateus, N. (2014). Bioavailability of anthocyanins and derivatives. *J. Funct. Foods* 7, 54–66. doi: 10.1016/j.jff.2013.05.010
- Ferrars, R., Czank, C., Zhang, Q., Botting, N., Kroon, P., Cassidy, A., et al. (2014). The pharmacokinetics of anthocyanins and their metabolites in humans. *Br. J. Pharmacol.* 171, 3268–3282. doi: 10.1111/bph.12676
- Feshani, A. M., Kouhsari, S. M., and Mohammadi, S. (2011). Vaccinium arctostaphylos, a common herbal medicine in Iran: molecular and biochemical study of its antidiabetic effects on alloxan-diabetic Wistar rats. *J. Ethnopharmacol.* 133, 67–74. doi: 10.1016/j.jep.2010.09.002
- Festa, A., D'agostino R. Jr, Williams, K., Karter, A., Mayer-Davis, E., Tracy, R., et al. (2001). The relation of body fat mass and distribution to markers of chronic inflammation. *Int. J. Obes. Relat. Metab. Disord.* 25, 1407–1415. doi: 10.1038/sj.ijo.0801792
- Flores, F. P., Singh, R. K., Kerr, W. L., Pegg, R. B., and Kong, F. (2013). Antioxidant and enzyme inhibitory activities of blueberry anthocyanins prepared using different solvents. *J. Agric. Food Chem.* 61, 4441–4447. doi: 10.1021/jf400429f
- Folmer, V., Soares, J. C., and Rocha, J. (2002). Oxidative stress in mice is dependent on the free glucose content of the diet. *Int. J. Biochem. Cell Biol.* 34, 1279–1285. doi: 10.1016/S1357-2725(02)00065-1
- Ford, E. S., Giles, W. H., and Dietz, W. H. (2002). Prevalence of the metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. *JAMA* 287, 356–359. doi: 10.1001/jama.287.3.356
- Frejngel, S., and Zdunczyk, Z. (2008). Chokeberry polyphenols reduce prooxidative influence of oxidized fats in rat diets. *Pol. J. Vet. Sci.* 11, 125–132.
- Friedman, J. M., and Halaas, J. L. (1998). Leptin and the regulation of body weight in mammals. *Nature* 395, 763–770. doi: 10.1038/27376
- Furukawa, S., Fujita, T., Shimabukuro, M., Iwaki, M., Yamada, Y., and Nakajima, Y., et al. (2017). Increased oxidative stress in obesity and its impact on metabolic syndrome. *J. Clin. Invest.* 114, 1752–1761. doi: 10.1172/JCI121625
- Gil, M. I., Tomás-Barberán, F. A., Hess-Pierce, B., Holcroft, D. M., and Kader, A. A. (2000). Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *J. Agric. Food Chem.* 48, 4581–4589. doi: 10.1021/jf000404a
- Gluckman, P. D., and Hanson, M. A. (2004). The developmental origins of the metabolic syndrome. *Trends Endocrinol. Metab.* 15, 183–187. doi: 10.1016/j.tem.2004.03.002
- González-Barrio, R., Borges, G., Mullen, W., and Crozier, A. (2010). Bioavailability of anthocyanins and ellagitannins following consumption of raspberries by healthy humans and subjects with an ileostomy. *J. Agric. Food Chem.* 58, 3933–3939. doi: 10.1021/jf100315d
- González-Barrio, R., Edwards, C. A., and Crozier, A. (2011). Colonic catabolism of ellagitannins, ellagic acid, and raspberry anthocyanins: *in vivo* and *in vitro* studies. *Drug Metab. Dispos.* 39, 1680–1688. doi: 10.1124/dmd.111.039651
- Grewal, S. A., Beinwal, M., Pandita, D., Sekhon, S. B., and Lather, V. (2016). Recent Updates on peroxisome proliferator-activated receptor δ agonists for the treatment of metabolic syndrome. *Med. Chem.* 12, 3–21. doi: 10.2174/1573406411666150525105826
- Grundy, S. M., Brewer, H. B., Cleeman, J. I., Smith, S. C., and Lenfant, C. (2004). Definition of metabolic syndrome. *Circulation* 109, 433–438. doi: 10.1161/01.CIR.0000111245.75752.C6
- Guilder, G. P., Hoetzer, G. L., Greiner, J. J., Stauffer, B. L., and Desouza, C. A. (2006). Influence of metabolic syndrome on biomarkers of oxidative stress and inflammation in obese adults. *Obesity* 14, 2127–2131. doi: 10.1038/oby.2006.248
- Gurrola-Diaz, C. M., García-López, P. M., Sánchez-Enríquez, S., Troyo-Sanromán, R., Andrade-González, I., and Gómez-Leyva, J. F. (2010). Effects of *Hibiscus sabdariffa* extract powder and preventive treatment (diet) on the lipid profiles of patients with metabolic syndrome (MeSy). *Phytomedicine* 17, 500–505. doi: 10.1016/j.phymed.2009.10.014
- Guzmán-Gerónimo, R. I., Alarcon-Zavaleta, T. M., Oliart-Ros, R. M., Meza-Alvarado, J. E., Herrera-Meza, S., and Chavez-Servia, J. L. (2017). Blue maize extract improves blood pressure, lipid profiles, and adipose tissue in high-sucrose diet-induced metabolic syndrome in rats. *J. Med. Food* 20, 110–115. doi: 10.1089/jmf.2016.0087
- Hämäläinen, M., Nieminen, R., Vuorela, P., Heinonen, M., and Moilanen, E. (2007). Anti-inflammatory effects of flavonoids: genistein, kaempferol, quercetin, and daidzein inhibit STAT-1 and NF- κ B activations, whereas flavone, isorhamnetin, naringenin, and pelargonidin inhibit only NF- κ B activation along with their inhibitory effect on iNOS expression and NO production in activated macrophages. *Mediators Inflamm.* 2007:45673. doi: 10.1155/2007/45673
- Hansel, B., Giral, P., Nobecourt, E., Chantepie, S., Bruckert, E., Chapman, M. J., et al. (2004). Metabolic syndrome is associated with elevated oxidative stress and dysfunctional dense high-density lipoprotein particles displaying impaired antioxidant activity. *J. Clin. Endocrinol. Metab.* 89, 4963–4971. doi: 10.1210/jc.2004-0305
- Hassimotto, N. M. A., Genovese, M. I., and Lajolo, F. M. (2008). Absorption and metabolism of cyanidin-3-glucoside and cyanidin-3-rutinoside extracted from wild mulberry (*Morus nigra* L.) in rats. *Nutr. Res.* 28, 198–207. doi: 10.1016/j.nutres.2007.12.012
- Hawley, S. A., Gadalla, A. E., Olsen, G. S., and Hardie, D. G. (2002). The antidiabetic drug metformin activates the AMP-activated protein kinase cascade via an adenine nucleotide-independent mechanism. *Diabetes* 51, 2420–2425. doi: 10.2337/diabetes.51.8.2420
- He, J., Magnuson, B. A., Lala, G., Tian, Q., Schwartz, S. J., and Giusti, M. M. (2006). Intact anthocyanins and metabolites in rat urine and plasma after 3 months of anthocyanin supplementation. *Nutr. Cancer* 54, 3–12. doi: 10.1207/s15327914nc5401_2
- He, Y., Li, X., Wang, J., Yang, Q., Yao, B., Zhao, Y., et al. (2017). Synthesis, characterization and evaluation cytotoxic activity of silver nanoparticles synthesized by Chinese herbal *Cornus officinalis* via environment friendly approach. *Environ. Toxicol. Pharmacol.* 56, 56–60. doi: 10.1016/j.etap.2017.08.035
- Hidalgo, J., Flores, C., Hidalgo, M., Perez, M., Yañez, A., Quiñones, L., et al. (2014). Delphinol® standardized Maqui berry extract reduces postprandial blood glucose increase in individuals with impaired glucose regulation by novel mechanism of sodium glucose cotransporter inhibition. *Panminerva Med.* 56, 1–7.
- Hotamisligil, G. S., Shargill, N. S., and Spiegelman, B. M. (1993). Adipose expression of tumor necrosis factor- α : direct role in obesity-linked insulin resistance. *Science* 259, 87–91. doi: 10.1126/science.7678183
- Hu, C., Zawistowski, J., Ling, W., and Kitts, D. D. (2003). Black rice (*Oryza sativa* L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems. *J. Agric. Food Chem.* 51, 5271–5277. doi: 10.1021/jf034466n
- Huang, H. P., Ou, T. T., and Wang, C. J. (2013). Mulberry (??? Sang Shèn Zi) and its bioactive compounds, the chemoprevention effects and molecular mechanisms *in vitro* and *in vivo*. *J. Tradit. Complement. Med.* 3, 7–15. doi: 10.4103/2225-4110.106535
- Huang, T. W., Chang, C. L., Kao, E. S., and Lin, J. H., (2015). Effect of *Hibiscus sabdariffa* extract on high fat diet-induced obesity and liver damage in hamsters. *Food Nutr. Res.* 59:29018. doi: 10.3402/fnr.v59.29018
- Huang, W.-Y., Liu, Y.-M., Wang, J., Wang, X.-N., and Li, C.-Y. (2014). Anti-inflammatory effect of the blueberry anthocyanins malvidin-3-glucoside and malvidin-3-galactoside in endothelial cells. *Molecules* 19, 12827–12841. doi: 10.3390/molecules190812827
- Ichiyanagi, T., Rahman, M. M., Kashiwada, Y., Ikeshiro, Y., Shida, Y., Hatano, Y., et al. (2004). Absorption and metabolism of delphinidin 3-O- β -D-glucopyranoside in rats. *Free Radic. Biol. Med.* 36, 930–937. doi: 10.1016/j.freeradbiomed.2004.01.005
- Ichiyanagi, T., Shida, Y., Rahman, M. M., Hatano, Y., and Konishi, T. (2005). Extended glucuronidation is another major path of cyanidin 3-O- β -D-glucopyranoside metabolism in rats. *J. Agric. Food Chem.* 53, 7312–7319. doi: 10.1021/jf051002b
- Iwaki, M., Matsuda, M., Maeda, N., Funahashi, T., Matsuzawa, Y., Makishima, M., et al. (2003). Induction of adiponectin, a fat-derived antidiabetic and antiatherogenic factor, by nuclear receptors. *Diabetes* 52, 1655–1663. doi: 10.2337/diabetes.52.7.1655

- Jeon, Y. D., Kang, S. H., Moon, K. H., Lee, J. H., Kim, D. G., Kim, W., et al. (2018). The effect of aronia berry on type 1 diabetes *in vivo* and *in vitro*. *J. Med. Food* 21, 244–253. doi: 10.1089/jmf.2017.3939
- Johnson, M. H., Lucius, A., Meyer, T., and Gonzalez De Mejia, E. (2011). Cultivar evaluation and effect of fermentation on antioxidant capacity and *in vitro* inhibition of α -amylase and α -glucosidase by highbush blueberry (*Vaccinium corombosum*). *J. Agric. Food Chem.* 59, 8923–8930. doi: 10.1021/jf201720z
- Jung, J. Y., Lim, Y., Moon, M. S., Kim, J. Y., and Kwon, O. (2011). Onion peel extracts ameliorate hyperglycemia and insulin resistance in high fat diet/streptozotocin-induced diabetic rats. *Nutr. Metab.* 8:18. doi: 10.1186/1743-7075-8-18
- Jurgonski, A., Juśkiewicz, J., and Zdunczyk, Z. (2008). Ingestion of black chokeberry fruit extract leads to intestinal and systemic changes in a rat model of prediabetes and hyperlipidemia. *Plant Foods Hum. Nutr.* 63, 176–182. doi: 10.1007/s11130-008-0087-7
- Kalea, A. Z., Clark, K., Schuschke, D. A., and Klimis-Zacas, D. J. (2009). Vascular reactivity is affected by dietary consumption of wild blueberries in the Sprague-Dawley rat. *J. Med. Food* 12, 21–28. doi: 10.1089/jmf.2008.0078
- Kalt, W., McDonald, J. E., Vinqvist-Tymchuk, M. R., Liu, Y., and Fillmore, S. A. (2017). Human anthocyanin bioavailability: effect of intake duration and dosing. *Food Funct.* 8, 4563–4569. doi: 10.1039/C7FO01074E
- Kamiloglu, S., Capanoglu, E., Grootaert, C., and Van Camp, J. (2015). Anthocyanin absorption and metabolism by human intestinal Caco-2 cells—A review. *Int. J. Mol. Sci.* 16, 21555–21574. doi: 10.3390/ijms160921555
- Kaplan, M., Hayek, T., Raz, A., Coleman, R., Dornfeld, L., Vaya, J., et al. (2001). Pomegranate juice supplementation to atherosclerotic mice reduces macrophage lipid peroxidation, cellular cholesterol accumulation and development of atherosclerosis. *J. Nutr.* 131, 2082–2089. doi: 10.1093/jn/131.8.2082
- Kastorini, C.-M., Milionis, H. J., Esposito, K., Giugliano, D., Goudevenos, J. A., and Panagiotakos, D. B. (2011). The effect of Mediterranean diet on metabolic syndrome and its components: a meta-analysis of 50 studies and 534,906 individuals. *J. Am. Coll. Cardiol.* 57, 1299–1313. doi: 10.1016/j.jacc.2010.09.073
- Kaul, K., Tarr, J. M., Ahmad, S. I., Kohner, E. M., and Chibber, R. (2013). Introduction to diabetes mellitus. *Adv. Exp. Med. Biol.* 771, 1–11.
- Kay, C. D. (2006). Aspects of anthocyanin absorption, metabolism and pharmacokinetics in humans. *Nutr. Res. Rev.* 19, 137–146. doi: 10.1079/NRR2005116
- Kay, C. D., Pereira-Caro, G., Ludwig, I. A., Clifford, M. N., and Crozier, A. (2017). Anthocyanins and flavanones are more bioavailable than previously perceived: a review of recent evidence. *Annu. Rev. Food Sci. Technol.* 8, 155–180. doi: 10.1146/annurev-food-030216-025636
- Kim, B., Ku, C. S., Pham, T. X., Park, Y., Martin, D. A., Xie, L., et al. (2013). Aronia melanocarpa (chokeberry) polyphenol-rich extract improves antioxidant function and reduces total plasma cholesterol in apolipoprotein E knockout mice. *Nutr. Res.* 33, 406–413. doi: 10.1016/j.nutres.2013.03.001
- Kim, H., Simbo, S., Fang, C., McAlister, L., Roque-Andrade, A., Banerjee, N., et al. (2018). Açai (*Euterpe oleracea* Mart.) beverage consumption improves biomarkers for inflammation but not glucose-or lipid-metabolism in individuals with metabolic syndrome in a randomized, double-blinded, placebo-controlled clinical trial. *Food Funct.* 9, 3097–3103. doi: 10.1039/C8FO00595H
- Kim, H. J., Tsoy, I., Park, J. M., Chung, J. I., Shin, S. C., and Chang, K. C. (2006). Anthocyanins from soybean seed coat inhibit the expression of TNF- α -induced genes associated with ischemia/reperfusion in endothelial cell by NF- κ B dependent pathway and reduce rat myocardial damages incurred by ischemia and reperfusion *in vivo*. *FEBS Lett.* 580, 1391–1397. doi: 10.1016/j.febslet.2006.01.062
- Kolovou, G. D., Anagnostopoulou, K. K., Salpea, K. D., and Mikhailidis, D. P. (2007). The prevalence of metabolic syndrome in various populations. *Am. J. Med. Sci.* 333, 362–371. doi: 10.1097/MAJ.0b013e318065c3a1
- Kudva, Y. C., and Butler, P. C. (1997). “Insulin secretion in type II diabetes mellitus,” in *Clinical Research in Diabetes and Obesity* (Totowa, NJ: Humana Press), 119–136. doi: 10.1007/978-1-4757-3906-0_7
- Kulling, S. E., and Rawel, H. M. (2008). Chokeberry (*Aronia melanocarpa*)—A review on the characteristic components and potential health effects. *Planta Med.* 74, 1625–1634. doi: 10.1055/s-0028-1088306
- Kumar, B., Arora, V., Kuhad, A., and Chopra, K. (2012). *Vaccinium myrtillus* ameliorates unpredictable chronic mild stress induced depression: possible involvement of nitric oxide pathway. *Phytother. Res.* 26, 488–497. doi: 10.1002/ptr.3584
- Kumari, K., and Augusti, K. (2002). Antidiabetic and antioxidant effects of S-methyl cysteine sulfoxide isolated from onions (*Allium cepa* Linn) as compared to standard drugs in alloxan diabetic rats. *Indian J. Exp. Biol.* 40, 1005–1009.
- Kusano, S., and Abe, H. (2000). Antidiabetic activity of white skinned sweet potato (*Ipomoea batatas* L.) in obese Zucker fatty rats. *Biol. Pharm. Bull.* 23, 23–26. doi: 10.1248/bpb.23.23
- Kusano, S., Abe, H., and Tamura, H. (2001). Isolation of antidiabetic components from white-skinned sweet potato (*Ipomoea batatas* L.). *Biosci. Biotechnol. Biochem.* 65, 109–114. doi: 10.1271/bbb.65.109
- Lakka, H.-M., Laaksonen, D. E., Lakka, T. A., Niskanen, L. K., Kumpusalo, E., Tuomilehto, J., et al. (2002). The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *JAMA* 288, 2709–2716. doi: 10.1001/jama.288.21.2709
- Lakka, T. A., and Laaksonen, D. E. (2007). Physical activity in prevention and treatment of the metabolic syndrome. *Appl. Physiol. Nutr. Metab.* 32, 76–88. doi: 10.1139/h06-113
- Lara-Castro, C., Luo, N., Wallace, P., Klein, R. L., and Garvey, W. T. (2006). Adiponectin multimeric complexes and the metabolic syndrome trait cluster. *Diabetes* 55, 249–259. doi: 10.2337/diabetes.55.01.06.db05-1105
- Lau, F. C., Bielinski, D. F., and Joseph, J. A. (2007). Inhibitory effects of blueberry extract on the production of inflammatory mediators in lipopolysaccharide-activated BV2 microglia. *J. Neurosci. Res.* 85, 1010–1017. doi: 10.1002/jnr.21205
- Lee, J. S., Kim, Y. R., Park, J. M., Kim, Y. E., Baek, N. I., and Hong, E. K. (2015). Cyanidin-3-glucoside isolated from mulberry fruits protects pancreatic β -cells against glucotoxicity-induced apoptosis. *Mol. Med. Rep.* 11, 2723–2728. doi: 10.3892/mmr.2014.3078
- Leyva, F., Godstrand, I. F., Ghatei, M., Proudler, A. J., Aldis, S., and Walton, C. (1998). Hyperleptinemia as a component of a metabolic syndrome of cardiovascular risk. *Arterioscler. Thromb. Vasc. Biol.* 18, 928–933. doi: 10.1161/01.ATV.18.6.928
- Lietti, A., Cristoni, A., and Picci, M. (1976). Studies on *Vaccinium myrtillus* anthocyanosides. I. Vasoprotective and antiinflammatory activity. *Arzneimittelforschung* 26, 829–832.
- Liu, Y., Tan, D., Shi, L., Liu, X., Zhang, Y., Tong, C., et al. (2015). Blueberry anthocyanins-enriched extracts attenuate cyclophosphamide-induced cardiac injury. *PLoS ONE* 10:e0127813. doi: 10.1145/2818302
- Louwen, F., Ritter, A., Kreis, N.n, and Yuan, J. (2018). Insight into the development of obesity: functional alterations of adipose-derived mesenchymal stem cells. *Obes. Rev.* 19, 888–904. doi: 10.1111/obr.12679
- Ludvik, B., Hanefeld, M., and Pacini, G. (2008). Improved metabolic control by *Ipomoea batatas* (Caiapo) is associated with increased adiponectin and decreased fibrinogen levels in type 2 diabetic subjects. *Diabetes Obes. Metab.* 10, 586–592. doi: 10.1111/j.1463-1326.2007.00752.x
- Ludvik, B., Neuffer, B., and Pacini, G. (2004). Efficacy of *Ipomoea batatas* (Caiapo) on diabetes control in type 2 diabetic subjects treated with diet. *Diabetes Care* 27, 436–440.
- Ludwig, I. A., Mena, P., Calani, L., Borges, G., Pereira-Caro, G., and Bresciani, L., et al. (2015). New insights into the bioavailability of red raspberry anthocyanins and ellagitannins. *Free Radic. Biol. Med.* 89, 758–769. doi: 10.1016/j.freeradbiomed.2015.10.400
- Maddux, B. A., See, W., Lawrence, J. C., Goldfine, A. L., Goldfine, I. D., and Evans, J. L. (2001). Protection against oxidative stress—induced insulin resistance in rat L6 muscle cells by micromolar concentrations of α -lipoic acid. *Diabetes* 50, 404–410. doi: 10.2337/diabetes.50.2.404
- Maiese, K. (2015). New insights for oxidative stress and diabetes mellitus. *Oxid. Med. Cell. Longev.* 2015:875961. doi: 10.1155/2015/875961
- Markakis, P. (2012). *Anthocyanins as Food Colors*. Elsevier.
- Martineau, L. C., Couture, A., Spoor, D., Benhaddou-Andaloussi, A., Harris, C., Meddah, B., et al. (2006). Anti-diabetic properties of the Canadian lowbush blueberry *Vaccinium angustifolium* Ait. *Phytomedicine* 13, 612–623. doi: 10.1016/j.phymed.2006.08.005
- Martinez-Abundis, E., Mendez-Del Villar, M., Perez-Rubio, K. G., Zuniga, L. Y., Cortez-Navarrete, M., and Ramirez-Rodriguez, A., et al. (2016). Novel

- nutraceutical therapies for the treatment of metabolic syndrome. *World J. Diabetes* 7, 142–152. doi: 10.4239/wjd.v7.i7.142
- Marvasti, T. B., and Adeli, K. (2010). Pharmacological management of metabolic syndrome and its lipid complications. *DARU J. Pharm. Sci.* 18:146.
- Maskimov, M. L., Svistunova, A. A., Tarasov, V. V., Chubarev, V. N., Barreto, G. E., Dralova, O. V., et al. (2016). Approaches for the development of drugs for treatment of obesity and metabolic syndrome. *Curr. Pharm. Des.* 22, 895–903. doi: 10.2174/1381612822666151209153047
- Mathew, P., and Augusti, K. (1975). Hypoglycaemic effects of onion, *Allium cepa* Linn. on diabetes mellitus—a preliminary report. *Indian J. Physiol. Pharmacol.* 19, 213–217.
- Matsui, T., Ebuchi, S., Kobayashi, M., Fukui, K., Sugita, K., Terahara, N., et al. (2002). Anti-hyperglycemic effect of diacylated anthocyanin derived from Ipomoea batatas cultivar Ayamurasaki can be achieved through the α -glucosidase inhibitory action. *J. Agric. Food Chem.* 50, 7244–7248. doi: 10.1021/jf025913m
- Matsuzawa, Y., Funahashi, T., and Nakamura, T. (1999). Molecular mechanism of metabolic syndrome X: contribution of adipocytokines-adipocyte derived bioactive substances. *Ann. N.Y. Acad. Sci.* 892, 146–154. doi: 10.1111/j.1749-6632.1999.tb07793.x
- Merone, L., and McDermott, R. (2017). Nutritional anti-inflammatories in the treatment and prevention of type 2 diabetes mellitus and the metabolic syndrome. *Diabetes Res. Clin. Pract.* 127, 238–253. doi: 10.1016/j.diabres.2017.02.019
- Mirshekar, M., Roghani, M., Khalili, M., Baluchnejadmojarad, T., and Moazzen, S. A. (2010). Chronic oral pelargonidin alleviates streptozotocin-induced diabetic neuropathic hyperalgesia in rat: involvement of oxidative stress. *Iran. Biomed. J.* 14, 33–39.
- Montague, C. T., and O'rahilly, S. (2000). The perils of portliness: causes and consequences of visceral adiposity. *Diabetes* 49, 883–888. doi: 10.2337/diabetes.49.6.883
- Morazzoni, P., and Bombardelli, E. (1996). *Vaccinium myrtillus* L. *Fitoterapia* 67, 3–29.
- Morrison, C., Ribnicky, D., Chang, J., Dunville, K., Forney, L., Stewart, L., et al. (2015). *In vivo* effects of dietary quercetin and quercetin-rich red onion extract on skeletal muscle mitochondria, metabolism, and insulin sensitivity. *Genes Nutr.* 10:2. doi: 10.1007/s12263-014-0451-1
- Mueller, D., Jung, K., Winter, M., Rogoll, D., Melcher, R., and Kulozik, U., et al. (2018). Encapsulation of anthocyanins from bilberries—Effects on bioavailability and intestinal accessibility in humans. *Food Chem.* 248, 217–224. doi: 10.1016/j.foodchem.2017.12.058
- Munasinghe, P. E., and Katara, R. (2016). Maladaptive autophagy in diabetic heart disease. *Int. J. Clin. Exp. Physiol.* 3:155.
- Nayak, C. A., and Rastogi, N. K. (2010). Effect of selected additives on microencapsulation of anthocyanin by spray drying. *Drying Technol.* 28, 1396–1404. doi: 10.1080/07373937.2010.482705
- Niemann, B., Rohrbach, S., Miller, M. R., Newby, D. E., Fuster, V., and Kovacic, J. C. (2017). Oxidative stress and cardiovascular risk: obesity, diabetes, smoking, and pollution: part 3 of a 3-part series. *J. Am. Coll. Cardiol.* 70, 230–251. doi: 10.1016/j.jacc.2017.05.043
- Olas, B., Wachowicz, B., Nowak, P., Kedzierska, M., Tomczak, A., and Stochma, A., et al. (2008). Studies on antioxidant properties of polyphenol-rich extract from berries of *Aronia melanocarpa* in blood platelets. *Acta Physiol. Pol.* 59:823.
- Passamonti, S., Vrhovsek, U., Vanzo, A., and Mattivi, F. (2003). The stomach as a site for anthocyanins absorption from food 1. *FEBS Lett.* 544, 210–213. doi: 10.1016/S0014-5793(03)00504-0
- Pergola, C., Rossi, A., Dugo, P., Cuzzocrea, S., and Sautebin, L. (2006). Inhibition of nitric oxide biosynthesis by anthocyanin fraction of blackberry extract. *Nitric Oxide* 15, 30–39. doi: 10.1016/j.niox.2005.10.003
- Pojer, E., Mattivi, F., Johnson, D., and Stockley, C. S. (2013). The case for anthocyanin consumption to promote human health: a review. *Comprehens. Rev. Food Sci. Food Safety* 12, 483–508. doi: 10.1111/1541-4337.12024
- Prior, R. L., Lazarus, S. A., Cao, G., Muccitelli, H., and Hammerstone, J. F. (2001). Identification of procyanidins and anthocyanins in blueberries and cranberries (*Vaccinium* spp.) using high-performance liquid chromatography/mass spectrometry. *J. Agric. Food Chem.* 49, 1270–1276. doi: 10.1021/jf001211q
- Prior, R. L., Wilkes, S., Rogers, T., Khanal, R. C., Wu, X., and Hager, T. J., et al. (2010). Dietary black raspberry anthocyanins do not alter development of obesity in mice fed an obesogenic high-fat diet. *J. Agric. Food Chem.* 58, 3977–3983. doi: 10.1021/jf9030772
- Qin, B., and Anderson, R. A. (2012). An extract of chokeberry attenuates weight gain and modulates insulin, adipogenic and inflammatory signalling pathways in epididymal adipose tissue of rats fed a fructose-rich diet. *Br. J. Nutr.* 108, 581–587. doi: 10.1017/S000711451100599X
- Qin, Y., Xia, M., Ma, J., Hao, Y., Liu, J., Mou, H., et al. (2009). Anthocyanin supplementation improves serum LDL and HDL-cholesterol concentrations associated with the inhibition of cholesteryl ester transfer protein in dyslipidemic subjects. *Am. J. Clin. Nutr.* 90, 485–492. doi: 10.3945/ajcn.2009.27814
- Reho, J. J., and Rahmouni, K. (2017). Oxidative and inflammatory signals in obesity-associated vascular abnormalities. *Clin. Sci.* 131, 1689–1700. doi: 10.1042/CS20170219
- Renaud, S. D., and De Lorgeril, M. (1992). Wine, alcohol, platelets, and the French paradox for coronary heart disease. *Lancet* 339, 1523–1526. doi: 10.1016/0140-6736(92)91277-F
- Robert, P., Gorena, T., Romero, N., Sepulveda, E., Chavez, J., and Saenz, C. (2010). Encapsulation of polyphenols and anthocyanins from pomegranate (*Punica granatum*) by spray drying. *Int. J. Food Sci. Technol.* 45, 1386–1394. doi: 10.1111/j.1365-2621.2010.02270.x
- Rojo, L. E., Ribnicky, D., Logendra, S., Poulev, A., Rojas-Silva, P., Kuhn, P., et al. (2012). *In vitro* and *in vivo* anti-diabetic effects of anthocyanins from Maqui Berry (*Aristotelia chilensis*). *Food Chem.* 131, 387–396. doi: 10.1016/j.foodchem.2011.08.066
- Romeo, G. R., Lee, J., and Shoelson, S. E. (2012). Metabolic syndrome, insulin resistance, and roles of inflammation—mechanisms and therapeutic targets. *Arterioscler. Thromb. Vasc. Biol.* 32, 1771–1776. doi: 10.1161/ATVBAHA.111.241869
- Routray, W., and Orsat, V. (2011). Blueberries and their anthocyanins: factors affecting biosynthesis and properties. *Comprehens. Rev. Food Sci. Food Safety* 10, 303–320. doi: 10.1111/j.1541-4337.2011.00164.x
- Roy, M., Sen, S., and Chakraborti, A. S. (2008). Action of pelargonidin on hyperglycemia and oxidative damage in diabetic rats: implication for glycation-induced hemoglobin modification. *Life Sci.* 82, 1102–1110. doi: 10.1016/j.lfs.2008.03.011
- Russell, J. C., and Proctor, S. D. (2006). Small animal models of cardiovascular disease: tools for the study of the roles of metabolic syndrome, dyslipidemia, and atherosclerosis. *Cardiovasc. Pathol.* 15, 318–330. doi: 10.1016/j.carpath.2006.09.001
- Samadder, A., Abraham, S. K., and Khuda-Bukhsh, A. R. (2016). Nanopharmaceutical approach using pelargonidin towards enhancement of efficacy for prevention of alloxan-induced DNA damage in L6 cells via activation of PARP and p53. *Environ. Toxicol. Pharmacol.* 43, 27–37. doi: 10.1016/j.etap.2016.02.010
- Samadder, A., Tarafdar, D., Abraham, S. K., Ghosh, K., and Khuda-Bukhsh, A. R. (2017). Nano-pelargonidin protects hyperglycemic-induced L6 cells against mitochondrial dysfunction. *Planta Med.* 83, 468–475. doi: 10.1055/s-0043-100017
- Scazzocchio, B., Vari, R., Filesi, C., D'archivio, M., Santangelo, C., Giovannini, C., Iacovelli, A., et al. (2011). Cyanidin-3-O- β -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR γ activity in human omental adipocytes. *Diabetes* 60, 2234–2244. doi: 10.2337/db10-1461
- Schmeichel, A. M., Schmelzer, J. D., and Low, P. A. (2003). Oxidative injury and apoptosis of dorsal root ganglion neurons in chronic experimental diabetic neuropathy. *Diabetes* 52, 165–171. doi: 10.2337/diabetes.52.1.165
- Schreckinger, M. E., Wang, J., Yousef, G., Lila, M. A., and Gonzalez De Mejia, E. (2010). Antioxidant capacity and *in vitro* inhibition of adipogenesis and inflammation by phenolic extracts of *Vaccinium floribundum* and *Aristotelia chilensis*. *J. Agric. Food Chem.* 58, 8966–8976. doi: 10.1021/jf100975m
- Schröder, H. (2007). Protective mechanisms of the Mediterranean diet in obesity and type 2 diabetes. *J. Nutr. Biochem.* 18, 149–160. doi: 10.1016/j.jnutbio.2006.05.006
- Schubert, S. Y., Lansky, E. P., and Neeman, I. (1999). Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids. *J. Ethnopharmacol.* 66, 11–17. doi: 10.1016/S0378-8741(98)00222-0
- Seeram, N. P., Henning, S. M., Zhang, Y., Suchard, M., Li, Z., and Heber, D. (2006). Pomegranate juice ellagitannin metabolites are present in human

- plasma and some persist in urine for up to 48 hours. *J. Nutr.* 136, 2481–2485. doi: 10.1093/jn/136.10.2481
- Seeram, N. P., Schutzki, R., Chandra, A., and Nair, M. G. (2002). Characterization, quantification, and bioactivities of anthocyanins in Cornus species. *J. Agric. Food Chem.* 50, 2519–2523. doi: 10.1021/jf0115903
- Sellappan, S., Akoh, C. C., and Krewer, G. (2002). Phenolic compounds and antioxidant capacity of Georgia-grown blueberries and blackberries. *J. Agric. Food Chem.* 50, 2432–2438. doi: 10.1021/jf011097r
- Seymour, E. M., Singer, A. A., Kirakosyan, A., Urcuyo-Llanes, D. E., Kaufman, P. B., and Bolling, S. F. (2008). Altered hyperlipidemia, hepatic steatosis, and hepatic peroxisome proliferator-activated receptors in rats with intake of tart cherry. *J. Med. Food* 11, 252–259. doi: 10.1089/jmf.2007.658
- Shamim, U. (2009). *Studies on the Mechanism of Action of Plant Derived Polyphenolic Compounds (Mobilization of Nuclear Copper by Plant Polyphenols and Oxidative Dna Breakage: Implications For an Anticancer Mechanism)*. Doctoral dissertation, Aligarh Muslim University.
- Shimomura, I., Funahashi, T., Takahashi, M., Maeda, K., Kotani, K., and Nakamura, T. Y. et al. (1996). Enhanced expression of PAI-1 in visceral fat: possible contributor to vascular disease in obesity. *Nat. Med.* 2, 800–803. doi: 10.1038/nm0796-800
- Shirode, A. B., Bharali, D. J., Nallanthighal, S., Coon, J. K., Mousa, S. A., and Reliene, R. (2015). Nanoencapsulation of pomegranate bioactive compounds for breast cancer chemoprevention. *Int. J. Nanomed.* 10, 475–484. doi: 10.2147/IJN.S65145
- Simeonov, S., Botushanov, N., Karahanian, E., Pavlova, M., Husianitis, H., and Troev, D. (2002). Effects of Aronia melanocarpa juice as part of the dietary regimen in patients with diabetes mellitus. *Folia Med. (Plovdiv)*. 44, 20–23.
- Skates, E., Overall, J., Dezege, K., Wilson, M., Esposito, D., Lila, M. A., et al. (2017). Berries containing anthocyanins with enhanced methylation profiles are more effective at ameliorating high fat diet-induced metabolic damage. *Food Chem. Toxicol.* 111, 445–453. doi: 10.1016/j.fct.2017.11.032
- Skoczynska, A., Jedrychowska, I., Poreba, R., Affelska-Jercha, A., Turczyn, B., Wojakowska, A., et al. (2007). Influence of chokeberry juice on arterial blood pressure and lipid parameters in men with mild hypercholesterolemia. *Pharmacol. Rep.* 59, 177–182.
- Sodagari, H. R., Farzaei, M. H., Bahramsoltani, R., Abdolghaffari, A. H., Mahmoudi, M., and Rezaei, N. (2015). Dietary anthocyanins as a complementary medicinal approach for management of inflammatory bowel disease. *Expert Rev. Gastroenterol. Hepatol.* 9, 807–820. doi: 10.1586/17474124.2015.1002086
- Spiegelman, B. M., and Flier, J. S. (2001). Obesity and the regulation of energy balance. *Cell* 104, 531–543. doi: 10.1016/S0092-8674(01)00240-9
- Stalmach, A., Clifford, M. N., Williamson, G., and Crozier, A. (2012). “Phytochemicals in coffee and the bioavailability of chlorogenic acids,” in *Teas, Cocoa and Coffee: Plant Secondary Metabolites and Health*, eds A. Crozier, H. Ashihara, and F. Tomás-Barbèran (West Sussex: Blackwell Publishing Ltd), 143–168.
- Steed, L., and Truong, V. D. (2008). Anthocyanin content, antioxidant activity, and selected physical properties of flowable purple fleshed sweetpotato purees. *J. Food Sci.* 73, S215–S221. doi: 10.1111/j.1750-3841.2008.00774.x
- Steppan, C. M., Bailey, S. T., Bhat, S., Brown, E. J., Banerjee, R. R., Wright, C. M., et al. (2001). The hormone resistin links obesity to diabetes. *Nature* 409, 307–312. doi: 10.1038/35053000
- Sundaram, R. K., Bhaskar, A., Vijayalingam, S., Viswanathan, M., Mohan, R., and Shanmugasundaram, K. R. (1996). Antioxidant status and lipid peroxidation in type II diabetes mellitus with and without complications. *Clin. Sci.* 90, 255–260. doi: 10.1042/cs0900255
- Suzuki, R., Tanaka, M., Takanashi, M., Hussain, A., Yuan, B., Toyoda, H., et al. (2011). Anthocyanidins-enriched bilberry extracts inhibit 3T3-L1 adipocyte differentiation via the insulin pathway. *Nutr. Metab.* 8:14. doi: 10.1186/1743-7075-8-14
- Taniguchi, C. M., Emanuelli, B., and Kahn, C. R. (2006). Critical nodes in signalling pathways: insights into insulin action. *Nat. Rev. Mol. Cell Biol.* 7, 85–96. doi: 10.1038/nrm1837
- Thibado, S. P., Thornthwaite, J. T., Ballard, T. K., and Goodman, B. T. (2018). Anticancer effects of Bilberry anthocyanins compared with NutraNanoSphere encapsulated Bilberry anthocyanins. *Mol. Clin. Oncol.* 8, 330–335. doi: 10.3892/mco.2017.1520
- Tonelli, C., Recupero, G. R., Berniakovich, I., Petroni, K., Titta, L., Giorgio, M., et al. (2009). Blood orange juice inhibits fat accumulation in mice. *Int. J. Obes.* 34, 578–588. doi: 10.1038/ijo.2009.266
- Toshima, S., Hasegawa, A., Kurabayashi, M., Itabe, H., Takano, T., Sugano, J. et al. (2000). Circulating oxidized low density lipoprotein levels. *Arterioscler. Thromb. Vasc. Biol.* 20, 2243–2247. doi: 10.1161/01.ATV.20.10.2243
- Tsang, C., Smail, N. F., Almoosawi, S., Mcdougall, G. J. M., and Al-Dujaili, E. A. S. (2018). Antioxidant rich potato improves arterial stiffness in healthy adults. *Plant Foods Hum. Nutr.* 73, 203–208. doi: 10.1007/s11130-018-0673-2
- Tsuda, T. (2012). Dietary anthocyanin-rich plants: biochemical basis and recent progress in health benefits studies. *Mol. Nutr. Food Res.* 56, 159–170. doi: 10.1002/mnfr.201100526
- Tsuda, T., Horio, F., Uchida, K., Aoki, H., and Osawa, T. (2003). Dietary cyanidin 3-O-β-D-glucoside-rich purple corn color prevents obesity and ameliorates hyperglycemia in mice. *J. Nutr.* 133, 2125–2130. doi: 10.1093/jn/133.7.2125
- Tymchuk, M. R., Liu, Y., and Fillmore, S. A. (2017). Human anthocyanin bioavailability: effect of intake duration and dosing. *Food Funct.* 8, 4563–4569. doi: 10.1039/c7fo01074e
- Unger, R. H. (2003). Minireview: weapons of lean body mass destruction: the role of ectopic lipids in the metabolic syndrome. *Endocrinology* 144, 5159–5165. doi: 10.1210/en.2003-0870
- Valcheva-Kuzmanova, S., and Belcheva, A. (2006). Current knowledge of *Aronia melanocarpa* as a medicinal plant. *Folia Med.* 48, 11–17.
- Vasconcelos, L. C. D. S., Sampaio, F. C., Sampaio, M. C. C., Pereira, M. D. S. V., Higinio, J. S., and Peixoto, M. H. P. (2006). Minimum inhibitory concentration of adherence of *Punica granatum* Linn (pomegranate) gel against *S. mutans*, *S. mitis* and *C. albicans*. *Braz. Dental J.* 17, 223–227. doi: 10.1590/S0103-64402006000300009
- Vendrame, S., Daugherty, A., Kristo, A. S., Riso, P., and Klimis-Zacas, D. (2013). Wild blueberry (*Vaccinium angustifolium*) consumption improves inflammatory status in the obese Zucker rat model of the metabolic syndrome. *J. Nutr. Biochem.* 24, 1508–1512. doi: 10.1016/j.jnutbio.2012.12.010
- Vlachopoulos, C., Aznaouridis, K., and Stefanadis, C. (2010). Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. *J. Am. Coll. Cardiol.* 55, 1318–1327. doi: 10.1016/j.jacc.2009.10.061
- Vroegrijk, I. O., Van Diepen, J. A., Van Den Berg, S., Westbroek, I., Keizer, H., Gambelli, L., et al. (2011). Pomegranate seed oil, a rich source of punicic acid, prevents diet-induced obesity and insulin resistance in mice. *Food Chem. Toxicol.* 49, 1426–1430. doi: 10.1016/j.fct.2011.03.037
- Wallace, A. M., McMahon, A. D., Packard, C. J., Kelly, A., Shepherd, J., and Gaw, A. (2001). Plasma leptin and the risk of cardiovascular disease in the west of Scotland coronary prevention study (WOSCOPS). *Circulation* 104, 3052–3056. doi: 10.1161/hc5001.101061
- Wang, H., Nair, M. G., Strasburg, G. M., Chang, Y.-C., Booren, A. M., Gray, J. I., et al. (1999). Antioxidant and antiinflammatory activities of anthocyanins and their aglycon, cyanidin, from tart cherries. *J. Nat. Prod.* 62, 294–296. doi: 10.1021/nj980501m
- Wang, S. Y., and Lin, H.-S. (2000). Antioxidant activity in fruits and leaves of blackberry, raspberry, and strawberry varies with cultivar and developmental stage. *J. Agric. Food Chem.* 48, 140–146. doi: 10.1021/jf9908345
- World Health Organization (2012). *10 Facts About Diabetes*. World Health Organization Available online at: <http://www.who.int/diabetes/en> (Accessed October 10, 2009).
- Wu, X., and Prior, R. L. (2005). Systematic identification and characterization of anthocyanins by HPLC-ESI-MS/MS in common foods in the United States: fruits and berries. *J. Agric. Food Chem.* 53, 2589–2599. doi: 10.1021/jf048068b
- Xiao, D., Sandhu, A., Huang, Y., Park, E., Edrington, I., and Burton-Freeman, B. M. (2017). The effect of dietary factors on strawberry anthocyanins oral bioavailability. *Food Funct.* 8, 3970–3979. doi: 10.1039/c7fo00885f

- Yan, F., Dai, G., and Zheng, X. (2016). Mulberry anthocyanin extract ameliorates insulin resistance by regulating PI3K/AKT pathway in HepG2 cells and db/db mice. *J. Nutr. Biochem.* 36, 68–80. doi: 10.1016/j.jnutbio.2016.07.004
- Yang, F., Li, H., Li, F., Xin, Z., Zhao, L., Zheng, Y., et al. (2010). Effect of nano-packing on preservation quality of fresh strawberry (*Fragaria ananassa* Duch. cv Fengxiang) during storage at 4°C. *J. Food Sci.* 75, C236–C240. doi: 10.1111/j.1750-3841.2010.01520.x
- Yang, L., Ling, W., Du, Z., Chen, Y., Li, D., Deng, S., et al. (2017). Effects of anthocyanins on cardiometabolic health: a systematic review and meta-analysis of randomized controlled trials. *Adv. Nutr.* 8, 684–693. doi: 10.3945/an.116.014852
- Yue, Q., Xu, L., Xiang, G., Yu, X., and Yao, Y. (2018). Characterization of gene expression profile, phenolic composition and antioxidant capacity in red-fleshed grape berries and their wines. *J. Agric. Food Chem.* 66, 7190–7199. doi: 10.1021/acs.jafc.8b01323
- Zhang, Y., Lian, F., Zhu, Y., Xia, M., Wang, Q., Ling, W., et al. (2010). Cyanidin-3-O- β -glucoside inhibits LPS-induced expression of inflammatory mediators through decreasing I κ B α phosphorylation in THP-1 cells. *Inflamm. Res.* 59, 723–730. doi: 10.1007/s00011-010-0183-7
- Zheng, W., and Wang, S. Y. (2003). Oxygen radical absorbing capacity of phenolics in blueberries, cranberries, chokeberries, and lingonberries. *J. Agric. Food Chem.* 51, 502–509. doi: 10.1021/jf020728u
- Zhou, F. H., Deng, X. J., Chen, Y. Q., Ya, F. L., Zhang, X. D., Song, F., et al. (2017). Anthocyanin cyanidin-3-glucoside attenuates platelet granule release in mice fed high-fat diets. *J. Nutr. Sci. Vitaminol.* 63, 237–243. doi: 10.3177/jnsv.63.237
- Zhou, G., Myers, R., Li, Y., Chen, Y., Shen, X., Fenyk-Melody, J., et al. (2001). Role of AMP-activated protein kinase in mechanism of metformin action. *J. Clin. Invest.* 108, 1167–1174. doi: 10.1172/JCI13505

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Naseri, Farzaei, Haratipour, Nabavi, Habtemariam, Farzaei, Khodarahmi, Tewari and Momtaz. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.