



Host-Based Prognostic Biomarkers to Improve Risk Stratification and Outcome of Febrile Children in Low- and Middle-Income Countries

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Specialty section:

This article was submitted to
Pediatric Infectious Diseases,
a section of the journal
Frontiers in Pediatrics

Received: 16 April 2020

Accepted: 12 August 2020

Published: 18 September 2020

Citation:

Balanza N, Erice C, Ngai M, Varo R,
Kain KC and Bassat Q (2020)
Host-Based Prognostic Biomarkers to
Improve Risk Stratification and
Outcome of Febrile Children in
Low- and Middle-Income Countries.
Front. Pediatr. 8:552083.
doi: 10.3389/fped.2020.552083

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Fever is one of the leading causes for pediatric medical consultation and the most common symptom at clinical presentation in low- and middle-income countries (LMICs). Most febrile episodes are due to self-limited infections, but a small proportion of children will develop life-threatening infections. The early recognition of children who have or are progressing to a critical illness among all febrile cases is challenging, and there are currently no objective and quantitative tools to do so. This results in increased morbidity and mortality among children with impending life-threatening infections, whilst contributing to the unnecessary prescription of antibiotics, overwhelming health care facilities, and harm to patients receiving avoidable antimicrobial treatment. Specific fever origin is difficult to ascertain and co-infections in LMICs are common. However, many severe infections share common pathways of host injury irrespective of etiology, including immune and endothelial activation that contribute to the pathobiology of sepsis (i.e., pathogen “agnostic” mechanisms of disease). Importantly, mediators of these pathways are independent markers of disease severity and outcome. We propose that measuring circulating levels of these factors can provide quantitative and objective evidence to: enable early recognition of severe infection; guide patient triage and management; enhance post-discharge risk stratification and follow up; and mitigate potential gender bias in clinical decisions. Here, we review the clinical and biological evidence supporting the clinical utility of host immune and endothelial activation biomarkers as components of novel rapid triage tests, and discuss the challenges and needs for developing and implementing such tools.

Keywords: febrile syndrome, severe infection, sepsis, risk stratification, prognostic host-biomarkers, angiotensin-2 (Ang-2), soluble triggering receptor expressed on myeloid cells 1 (sTREM-1)

INTRODUCTION

In low- and middle-income countries (LMICs), it is common to have multiple infections throughout childhood that often manifest as febrile syndromes. It is estimated that a child under five years in a LMIC may experience around six episodes of fever per year (1). Consequently, fever is one of the leading causes for medical consultation and the most common pediatric symptom at clinical presentation (2). The vast majority of febrile illnesses are uncomplicated and self-limited. However, a small proportion of these may become severe and progress to sepsis, which is defined as life-threatening organ dysfunction due to a dysregulated host response to infection and considered to be agnostic to etiology (3, 4). While severe disease/sepsis needs urgent referral/admission and adequate treatment, febrile children without critical illness can be treated conservatively and often without antimicrobials once malaria has been ruled out (5, 6). The early recognition of children who have or are progressing to sepsis is challenging, because the initial presentation of a severe infection may be subtle and non-specific (2). Delays in the recognition and treatment of severe infections/sepsis increase child mortality and neurocognitive impairment in survivors (7, 8). Moreover, unnecessary admissions, referrals, and treatment of self-limited infections results in misallocation of health resources, overloading of health care facilities, contribution to development of antimicrobial resistance, and adverse events due to nosocomial infections and exposure to unnecessary investigations and treatments.

The underlying infectious etiologies of pediatric febrile syndromes are highly diverse and differ according to community (e.g., geographical location, season, or disease control measures implemented) and individual factors (e.g., age, immunity, or concomitant conditions) (2). Despite progress in recent years, the epidemiology of fever in LMICs remains poorly characterized (9, 10). The scarcity of available diagnostic tools and laboratory services in LMICs makes it challenging to establish whether a fever is viral, bacterial or parasitic in origin. Moreover, mixed or multiple infections, and high rates of asymptomatic pathogen carriers, are frequent in these settings (5, 11). The innovative use of minimally-invasive post-mortem methods, to investigate cause of death, also supports the co-existence of multiple infections and a progressive abandonment of the dogma “one pathogen—one disease—one cause of death” (12).

International guidelines for the management of childhood illnesses (i.e., Integrated Management of Childhood Illness [IMCI], integrated Community Case Management [iCCM]), which have been shown to improve child survival, give simplified algorithms to assist health care providers in fever management (13, 14). However, these are designed in such a broad way that may result in over-diagnosis, over-treatment, and over-referral (2). Pediatric disease severity scores (e.g., Lambaréné Organ Dysfunction Score [LODS] (15), Signs of Inflammation in Child that Kill [SICK] (16)) have also been developed, but their performance is highly dependent on the capacity of health care providers to identify danger signs and the availability of clinical and laboratory data. In parallel, current rapid diagnostic tests (RDTs) are largely pathogen-based (e.g., dengue and malaria

RDTs) and, while they can identify the presence or absence of infection, they do not have the ability to differentiate between an uncomplicated infection vs. a life-threatening one. Previous studies have reported sub-optimal management of severe febrile syndromes in children, including poor triage, referral, and admission practices, in different levels of healthcare (17–20).

Additionally, once a child is admitted in a health facility, it is still difficult to clearly determine disease prognosis and thus make adequate decisions in regards to discharge. In fact, death in the first weeks after discharge remains an important and poorly acknowledged contributor to child mortality in LMICs, and can be as common or even more frequent than in-hospital mortality (21). Although triaging algorithms that identify children at increased risk of post-discharge mortality exist (22, 23), it is necessary to enhance how discharge decisions are made and identify those children that could benefit most from a follow-up visit.

Consequently, there is a compelling need to improve risk stratification tools for febrile children, for both initial management and discharge decisions. Accordingly, the World Health Organization (WHO) has highlighted the importance of developing new point-of-care (POC) tests to identify febrile patients at risk of progressing to severe disease/sepsis (2). This perspective article aims to highlight the potential of using host-biomarkers of disease severity and prognosis as objective measurements to guide critical clinical decisions. The measurement of these biomarkers using POC tests could complement clinical scores in community settings, as well as add value to them in formal health care settings.

Pathogen “Agnostic” Mechanisms of Severe Infection

Host response has an important role in determining severity and outcome of infections. Moreover, it has become well-recognized that many different types of pathogens share common host response pathways of injury (i.e., pathogen “agnostic” mechanisms of disease) (24–26). Immune and endothelial activation lay upstream of endothelial destabilization, microvascular leak, multi-organ dysfunction, and death and, therefore, these pathways have been implicated in the pathogenesis of sepsis (27–29). A healthy endothelium must be adaptive in order to interact with its changing environment, alternating between activated and quiescent states by responding to host-derived cues. During an infection, the immune response triggers endothelium activation, meaning it becomes permeable and pro-inflammatory, in order to accommodate pathogen sequestration and elimination as well as vascular remodeling and healing. However, a dysregulated host response, in either step of this cascade of events, can result in endothelial dysfunction and tissue injury (25).

Evidence suggests that mediators of immune and endothelial activation are independent and quantitative markers of disease severity and prognosis in sepsis due to multiple causes, including: malaria, bacterial infections, dengue, or Ebola (25, 26, 28, 30–34). Therefore, measuring circulating levels of mediators of these pathways could be used for disease severity

and prognostic determination to guide clinical decisions. Additionally, they represent an alternative to using acute phase proteins procalcitonin (PCT) and C-reactive protein (CRP). The performance of PCT and CRP as disease severity biomarkers has been extensively studied in high-income countries (HIC), but less is known from LMIC populations (35). In children living in LMICs, PCT and CRP concentrations have been reported to be influenced by prevalent conditions like malnutrition, coexisting malaria infection, and HIV-related immunosuppression (36–38).

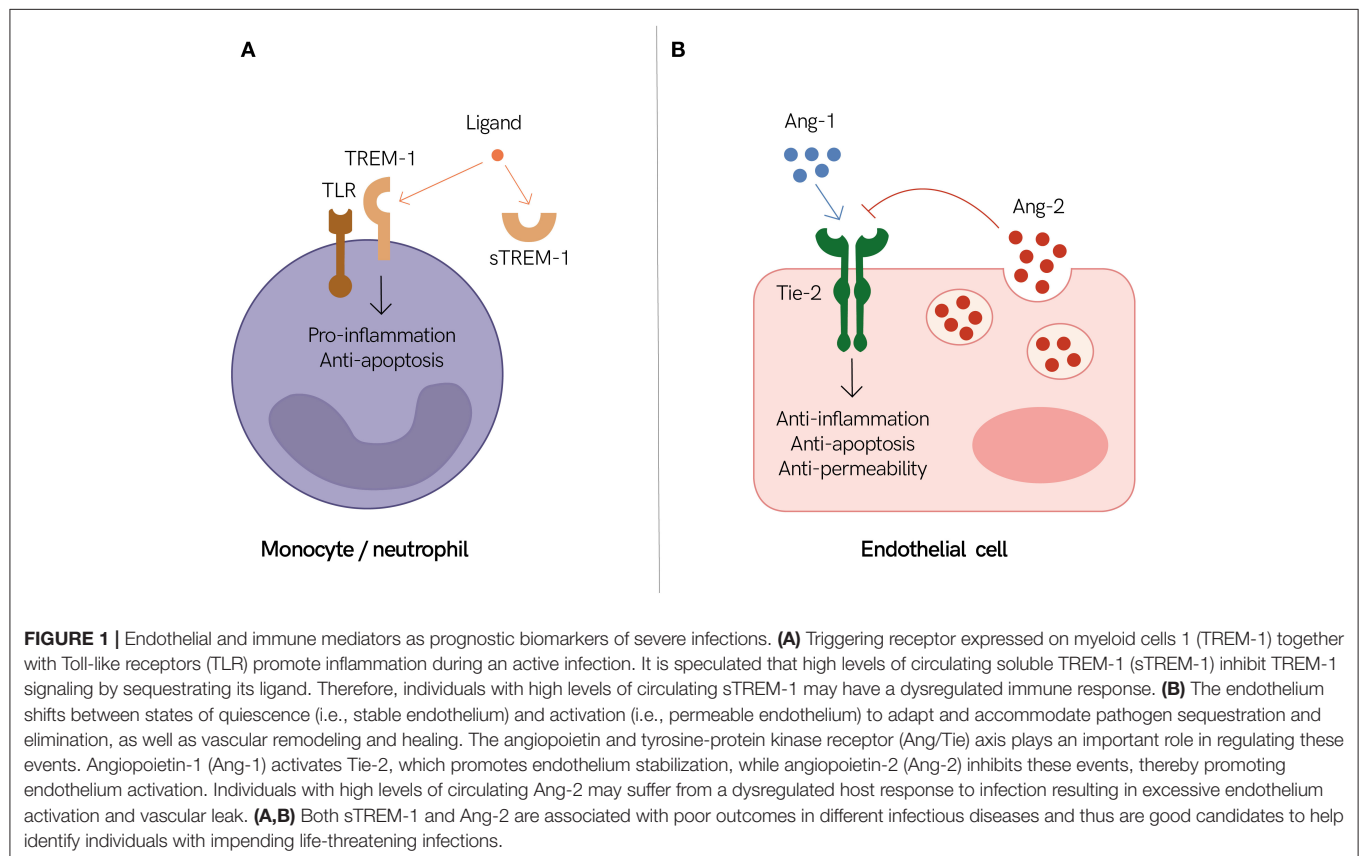
Mediators of Immune and Endothelial Activation as Prognostic and Severity Markers: Overview of Clinical Studies

A large body of literature exists for the potential use of immune and endothelial host-biomarkers as disease severity and prognostic tools. Here, we highlight and provide an overview of some of these biomarkers and concentrate on two specific promising examples (Figure 1).

Tumor necrosis factor (TNF) is the classic pro-inflammatory cytokine. Not surprisingly, elevated levels of TNF are observed in individuals with severe disease (i.e., severe malaria, bacteremia, and dengue) in comparison to individuals with uncomplicated disease (39–52). Increased TNF levels are also associated with neurological deficits and longer coma duration in children with cerebral malaria (53). Similarly, several studies have reported that individuals with severe disease and/or non-survivors have higher

levels of soluble TNF receptor 1 (sTNFR1) (30, 47, 48, 54–61). Likewise, interleukins (ILs) are key components of the immune system and circulating concentrations of IL-6, IL-8, and IL-10 have been extensively studied in disease severity and progression. Among them, IL-6 is perhaps the best characterized. Several lines of evidence indicate that high IL-6 levels are correlated with bacterial sepsis, severe malaria, severe dengue, and increased risk of dengue mortality (39, 41, 44, 46, 48, 50, 62–68). A study in febrile adults reported IL-6 to be a good predictor analyte between non-survivors and survivors (54). Interferon γ -induced protein 10 (IP-10) is an interesting pro-inflammatory chemokine, but studies of its association with disease severity have generated contradictory results. A few studies have found reduced IP-10 levels in severe dengue cases, but other observed the contrary (69–72). In the context of malaria, studies have reported an association between high IP-10 and mortality (73–75), while in febrile adults low levels were associated with death (54). Chitinase-3-like 1 (CHI3L1) has also been identified as a severity marker in different causes of sepsis, with increased levels associated with severe disease, complications and mortality (54, 76, 77).

Molecules indicative of endothelial activation and dysfunction include mediators of endothelial cell function, components of the coagulation pathway, soluble cell surface adhesion molecules, and regulators of vascular tone and permeability (25). Among them, a growing body of evidence demonstrates that elevated levels of soluble intercellular adhesion molecule-1 (sICAM-1)



are associated with disease severity and prognosis in malaria, dengue, and other causes of sepsis (72–74, 78–80). Soluble Fms-like tyrosine kinase 1 (sFlt-1) is another interesting marker of endothelial activation, which binds to vascular endothelial growth factor (VEGF). sFlt-1 has been reported to be good at discriminating between survivors and non-survivors in pediatric severe malaria (73, 74) and at predicting 28-day mortality in febrile adults (54). Moreover, other studies have shown that sFlt-1 correlates with severity in patients with bacterial sepsis (81).

sTREM-1 as a Promising Biomarker of a Dysregulated Immune Response

Triggering receptor expressed on myeloid cells 1 (TREM-1) is expressed on neutrophils and monocytes. Activation of TREM-1 signaling culminates in an enhanced pro-inflammatory response and induction of anti-apoptotic pathways. Soluble TREM-1 (sTREM-1) has the same extracellular region as TREM-1 but lacks its transmembrane region. Thus, sTREM-1 can combine with the same ligands, and it is thought to negatively regulate TREM-1 signaling pathways (Figure 1A) (82).

Several clinical studies show that increased sTREM-1 levels are correlated with disease severity, longer clinical recovery times, multi-organ dysfunction, and mortality in both children and adults (54, 61, 73, 74, 83–86). It is speculated that these adverse outcomes are reflective of an imbalance in TREM-1 signaling, resulting in excess immune effector cells death and immunosuppression (54). ROC curve analysis of a cohort study conducted in Cameroon showed that sTREM-1 had discriminatory power between children with severe malaria vs. uncomplicated malaria (AUC = 0.96 [95% CI: 0.92–0.99]) (85). This finding is complemented by a Tanzanian study in outpatient febrile adults, which found that sTREM-1 is a strong prognostic marker for 28-day mortality (AUC = 0.87 [95% CI: 0.81–0.92]) (54). Importantly, this same study showed that the mortality prognostic utility of sTREM-1 was superior to CRP and PCT ($p \leq 0.0001$). Another study conducted in rural Thailand in hospitalized patients with suspected infection also reported sTREM-1 to have strong prognostic 28-day mortality (AUC = 0.81 [95% CI: 0.77–0.85]) (61). Moreover, in these two studies, models combining sTREM-1 levels and validated clinical scoring systems (e.g., quick Sequential Organ Failure Assessment [qSOFA] and systemic Inflammatory Response Syndrome [SIRS]) had better prognostic accuracy than the clinical scoring systems alone (54, 61).

The Ang/Tie Axis as an Example of Endothelial Integrity Modulator

Microvascular function and permeability are regulated by several pathways, including the angiotensin and tyrosine-protein kinase receptor (Ang/Tie) axis. Angiotensin-1 (Ang-1) and angiotensin-2 (Ang-2) are ligands of the Tie-2 receptor, which is present on endothelial cells. Ang-1 is constitutively produced and secreted by pericytes and smooth muscle cells.

Upon Ang-1 interaction with Tie-2, stabilization and maturation of blood vessels is triggered. Ang-2 is constitutively expressed at low levels and is co-localized with von Willebrand factor (vWF) within the Weibel Palade bodies (WPB) of endothelial cells. During an inflammatory response, both the expression of Ang-2 and its release from the WPB are increased, leading to an increased Ang-2:Ang-1 ratio. Ang-2 antagonizes Ang-1 downstream signaling by binding to Tie-2, resulting in endothelial destabilization, microvascular leakage, and organ dysfunction, which are common features of sepsis due to multiple causes (Figure 1B) (26).

Several studies have linked a dysregulated Ang/Tie axis to disease severity in malaria. Low levels of Ang-1, high levels of Ang-2, high Ang-2:Ang-1 ratios, as well as high soluble Tie-2 (sTie-2) levels are consistently reported in severe malaria in comparison to uncomplicated malaria (73, 78, 87–91). Elevated levels of Ang-2 and sTie-2 are associated with mortality, and Ang-2 has also been linked to multi-organ dysfunction, respiratory distress, impaired consciousness, and acute kidney injury among other complications (73, 74, 92–95). ROC curve analyses have demonstrated that Ang-1, Ang-2, and the ratio of Ang-2:Ang-1 have strong predictive power for disease severity and adverse outcomes, and some studies indicate that these markers are superior to lactate or parasitemia (73, 89, 91, 92). Interestingly, Ang-2 levels at presentation have also been reported to be good at predicting post-discharge mortality (AUC = 0.82 [95% CI: 0.71–0.93]) (74). Moreover, Ang-2 levels in combination with clinical predictors, have been shown to improve prognostic accuracy above that of clinical predictors alone (e.g., LODS) (74, 92). Levels of Ang-1, Ang-2, and sTie-2 have also been examined in other causes of sepsis. Results are mostly comparable to the malaria studies, as elevated Ang-2 and reduced Ang-1 levels have been associated with disease severity, intensive care unit (ICU) admission, shock, organ dysfunction, and death (54, 61, 79, 96–99).

CHALLENGES AHEAD AND THE NEED TO DEVELOP A RAPID TRIAGE TOOL

In the last decades, the number of published studies on biomarkers has increased exponentially. Recent scientific advances in high throughput “omics” technologies have driven the biomarker discovery and development field forward. Despite progress in the field and investment from major stakeholders, the number of biomarkers approved for any type of clinical use is limited (100, 101). Currently, the aforementioned biomarkers of immunological and endothelial origin hold promise, but they are not used in routine clinical practice in LMICs for risk-stratification of children.

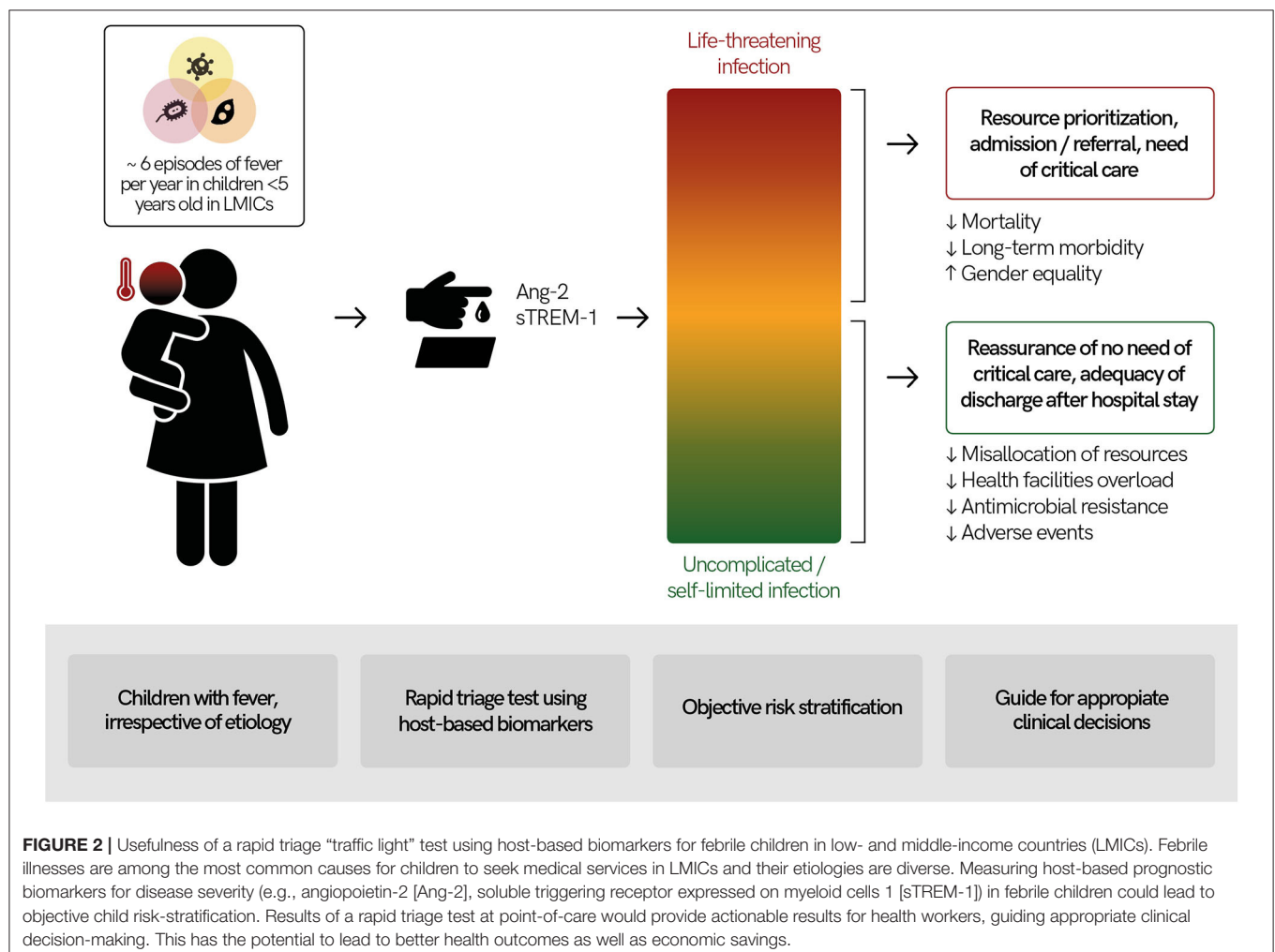
For future implementation, further research is necessary. Additional scientific evidence is needed regarding which biomarkers are most useful and which cut-off levels should be used. The ideal prognostic biomarker for severe infection/sepsis should be: capable of identifying the subset of patients at high risk of adverse outcomes before this risk is clearly evident,

correlated with disease severity over time, and useful regardless of disease microbiological etiology (25). Of the previously outlined biomarkers, sTREM-1 and Ang-2 are promising contenders to approximate this ideal. Besides, a combinational biomarker approach should also be considered, as it has been reported to enhance prognostic accuracy in some studies and it could be especially useful if biomarkers from different pathobiological pathways were measured (59, 73). Furthermore, rigorous and adequately powered randomized controlled trials will be essential to validate the clinical utility of adding biomarker measurements in guiding management decisions in both community-based and formal health care settings. Ideally, these studies should be carried out in different geographical areas in order to demonstrate reproducibility of findings and their added benefit to standard of care. If solid evidence of clinical benefit is provided, results could be used to adapt current international guidelines (i.e., iCCM, IMCI) (35).

Another crucial step is the development of rapid POC tests. This would allow measuring prognostic biomarkers at the community and peripheral level, where they could have a significant impact. These new rapid triage tests should fulfill

the WHO ASSURED criteria which include simplicity, low cost, rapidness and suitability for field use (102). Recently, a lateral flow immunoassay has been reported to detect IL-6 in a quantitative manner and similar devices need to be developed (103). Malaria RDTs are already widely implemented in Asia and Africa for fever management, where they are culturally accepted by communities (104–107). Therefore, it is expected that analogous devices would also be culturally acceptable. Another approach would be the integration of prognostic biomarker measurements into existing malaria RDTs for use in malaria endemic areas, rather than developing separate rapid triage tests, as proposed by McDonald et al. (24). In both scenarios a training phase for health care providers would be important to ensure their correct usage and interpretation of results.

Finally, cost-effectiveness studies are also necessary to ascertain affordability of using these specific biomarkers in different use case scenarios during routine clinical practice in LMICs. Currently, there are no cost-effectiveness analyses for the biomarkers we propose and therefore they need to be conducted. These will need to take into account the potential



deaths, disability, adverse effects, and resource misallocation that could be averted. Importantly, in order to make this prognostic biomarker approach cost-effective, developing inexpensive rapid triage tests is essential.

BROAD POTENTIAL IMPACT

If current barriers are overcome, rapid triage tools measuring host-biomarkers of immunological and endothelial origin have the potential to transform child fever management in LMICs. Objective risk stratification of febrile children could bring benefits for child health and optimize the use of limited health resources (Figure 2). Several studies have reported that the roll out of malaria RDTs has resulted in reduced antimalarial overuse but increased antibiotic prescription for febrile syndromes (108). Antimicrobial stewardship programs aim to reduce antimicrobial resistance by, among others, optimizing antibiotic use (109). Therefore, identification of individuals with life-threatening infections from those with self-resolving infections could help guide the safe reduction of antibiotic prescriptions.

The proposed approach could also reduce inequalities in fever management related to gender. Gender-based health disparities have been reported in LMICs, especially in south Asia, and are attributed to societal norms and family practices that favor sons (110, 111). These include differences in length of time before seeking care, frequency in consulting a health professional during illness, type of treatment given, or duration of hospital stay. This is a major driver of adverse health outcomes in girls and is hypothesized to contribute to the inequality observed in some countries' sex-stratified child mortality rates (112). A quantitative and objective triage tool indicative of severity could help mitigate gender biases when determining prioritization of clinical care, and referral/admission or discharge decisions.

THE BIGGER PICTURE

Although the immediate benefit of host-biomarkers as prognostic tools is warranted for child risk stratification in LMICs, a similar triage tool for risk-stratification could also be useful in other scenarios, such as in pediatric health care settings in HIC and could also be applied to adult patient triage. In fact, some of the evidence we discuss comes from studies in HICs or from adults in LMICs. The current coronavirus disease (COVID-19) pandemic exemplifies the urgent need for accurate and rapid triage tools for early triage of severe disease, since most cases have self-

limited courses but high-risk individuals need identification and prioritization of care and follow-up. HICs and LMICs are moving toward integrated approaches to care, and in both settings the ultimate aim is the same: better care whilst reducing costs.

CONCLUSIONS

Mediators of endothelial and immunological pathways (e.g., sTREM-1 and Ang-2) have great potential as prognostic biomarkers for disease severity. Nevertheless, their clinical utility and cost-effectiveness needs further validation by prospective studies and randomized clinical trials. Their hypothetical implementation in LMICs via POC tests could provide actionable results for frontline health care workers facing the daily challenge of visiting children with fever. The proposed approach could be highly beneficial for child health, gender equality, appropriate use of antimicrobials, and optimization of limited health resources.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

AUTHOR CONTRIBUTIONS

The manuscript was prepared with input from all authors. NB, CE, and MN contributed equally and share first co-authorship. KCK and QB share senior co-authorship. All authors read and approved the final manuscript.

FUNDING

This work was supported in part by grants from the Canadian Institutes of Health Research (CIHR FDN 148439 to KCK), the Canada Research Chairs program (KCK), and The Tesari Foundation. ISGlobal acknowledges support from the Spanish Ministry of Science and Innovation through the Centro de Excelencia Severo Ochoa 2019–2023 Program (CEX2018-000806-S) and support from the Generalitat de Catalunya through the CERCA Program. CISM is supported by the Government of Mozambique and the Spanish Agency for International Development (AECID). NB is supported by an FPU predoctoral fellowship from the Spanish Ministry of Universities (FPU18/04260).

REFERENCES

- Gething PW, Kirui VC, Alegana VA, Okiro EA, Noor AM, Snow RW. Estimating the number of paediatric fevers associated with malaria infection presenting to Africa's public health sector in (2007). *PLoS Med.* (2010) 7:e1000301. doi: 10.1371/journal.pmed.1000301
- World Health Organization. *WHO Informal Consultation on Fever Management in Peripheral Health Care Settings* (2013).
- Rudd KE, Johnson SC, Agesa KM, Shackelford KA, Tsoi D, Kievlan DR, et al. Global, regional, and national sepsis incidence and mortality, 1990–2017: analysis for the Global Burden of Disease Study. *Lancet.* (2020) 395:200–11. doi: 10.1016/S0140-6736(19)32989-7
- Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA.* (2016) 315:801–10. doi: 10.1001/jama.2016.0287
- D'Acremont V, Kilowoko M, Kyungu E, Philipina S, Sangu W, Kahama-Marro J, et al. Beyond malaria—causes of fever in outpatient Tanzanian children. *N Engl J Med.* (2014) 370:809–17. doi: 10.1056/NEJMoa1214482
- Hazir T, Nisar YB, Abbasi S, Ashraf YP, Khurshid J, Tariq P, et al. Comparison of oral amoxicillin with placebo for the treatment of world

- health organization-defined nonsevere pneumonia in children aged 2-59 months: a multicenter, double-blind, randomized, placebo-controlled trial in pakistan. *Clin Infect Dis*. (2011) 52:293–300. doi: 10.1093/cid/ciq142
7. John CC, Carabin H, Montano SM, Bangirana P, Zunt JR, Peterson PK. Global research priorities for infections that affect the nervous system. *Nature*. (2015) 527:S178–86. doi: 10.1038/nature16033
 8. Christensen SS, Eslick GD. Cerebral malaria as a risk factor for the development of epilepsy and other long-term neurological conditions: a meta-analysis. *Trans R Soc Trop Med Hyg*. (2015) 109:233–8. doi: 10.1093/trstmh/trv005
 9. Prasad N, Murdoch DR, Reyburn H, Crump JA. Etiology of severe febrile illness in low- and middle-income countries: a systematic review. *PLoS One*. (2015) 10:e0127962. doi: 10.1371/journal.pone.0127962
 10. Kiemde F, Spijker R, Mens PF, Tinto H, Boele M, Schallig HD. Aetiologies of non-malaria febrile episodes in children under 5 years in sub-Saharan Africa. *Trop Med Int Health*. (2016) 21:943–55. doi: 10.1111/tmi.12722
 11. Iroh Tam P-Y, Obaro SK, Storch G. Challenges in the etiology and diagnosis of acute febrile illness in children in low- and middle-income countries. *J Pediatr Infect Dis Soc*. (2016) 5:190–205. doi: 10.1093/jpids/piw016
 12. Raghunathan PL, Madhi SA, Breiman RF. Illuminating child mortality: discovering why children die. *Clin Infect Dis*. (2019) 69:S257–9. doi: 10.1093/cid/ciz562
 13. Gera T, Shah D, Garner P, Richardson M, Sachdev HS. Integrated management of childhood illness (IMCI) strategy for children under five. *Cochrane Database Syst Rev*. (2016) Cd010123. doi: 10.1002/14651858.CD010123.pub2
 14. Prosnitz D, Herrera S, Coelho H, Moonzwe Davis L, Zalisk K, Yourkavitch J. Evidence of Impact: iCCM as a strategy to save lives of children under five. *J Glob Health*. (2019) 9:010801. doi: 10.7189/jogh.09.010801
 15. Helbok R, Kendjo E, Issifou S, Lackner P, Newton CR, Kombila M, et al. The Lambarene Organ Dysfunction Score (LODS) is a simple clinical predictor of fatal malaria in African children. *J Infect Dis*. (2009) 200:1834–41. doi: 10.1086/648409
 16. Gupta MA, Chakrabarty A, Halstead R, Sahni M, Rangasami J, Puliylal A, et al. Validation of “Signs of Inflammation in Children that Kill” (SICK) score for immediate non-invasive assessment of severity of illness. *Ital J Pediatr*. (2010) 36:35. doi: 10.1186/1824-7288-36-35
 17. Bottger C, Bernard L, Briand V, Bougouma C, Triendebeogo J, Ridde V. Primary healthcare providers’ practices related to non-malarial acute febrile illness in Burkina Faso. *Trans R Soc Trop Med Hyg*. (2018) 111:555–63. doi: 10.1093/trstmh/try009
 18. Achan J, Tibenderana J, Kyabayinze D, Maweje H, Mugizi R, Mpeka B, et al. Case management of severe malaria—a forgotten practice: experiences from health facilities in Uganda. *PLoS One*. (2011) 6:e17053. doi: 10.1371/journal.pone.0017053
 19. Elnour FA, Alagib MEA, Bansal D, Farag E, Malik EM. Severe malaria management: current situation, challenges and lessons learned from Gezira State, Sudan. *Malar J*. (2019) 18:170. doi: 10.1186/s12936-019-2805-z
 20. Chang JL, Reyes R, Matte M, Ntaro M, Mulogo E, Wiens MO, et al. Who stays and who goes: predictors of admission among patients presenting with febrile illness and a positive malaria rapid diagnostic test in a rural Ugandan health center. *Am J Trop Med Hyg*. (2018) 99:1080–8. doi: 10.4269/ajtmh.18-0338
 21. Wiens MO, Pawluk S, Kissoon N, Kumbakumba E, Ansermino JM, Singer J, et al. Pediatric post-discharge mortality in resource poor countries: a systematic review. *PLoS One*. (2013) 8:e66698. doi: 10.1371/journal.pone.0066698
 22. Madrid L, Casellas A, Saco C, Quinto L, Siteo A, Varo R, et al. Postdischarge mortality prediction in sub-saharan Africa. *Pediatrics*. (2019) 143:e20180606. doi: 10.1542/peds.2018-0606
 23. Wiens MO, Kumbakumba E, Larson CP, Ansermino JM, Singer J, Kissoon N, et al. Postdischarge mortality in children with acute infectious diseases: derivation of postdischarge mortality prediction models. *BMJ Open*. (2015) 5:e009449. doi: 10.1136/bmjopen-2015-009449
 24. McDonald CR, Weckman A, Richard-Greenblatt M, Leligowicz A, Kain KC. Integrated fever management: disease severity markers to triage children with malaria and non-malarial febrile illness. *Malar J*. (2018) 17:353. doi: 10.1186/s12936-018-2488-x
 25. Page AV, Liles WC. Biomarkers of endothelial activation/dysfunction in infectious diseases. *Virulence*. (2013) 4:507–16. doi: 10.4161/viru.24530
 26. Leligowicz A, Richard-Greenblatt M, Wright J, Crowley VM, Kain KC. Endothelial activation: the ang/tie axis in sepsis. *Front Immunol*. (2018) 9:838. doi: 10.3389/fimmu.2018.00838
 27. Ghosh CC, David S, Zhang R, Berghelli A, Milam K, Higgins SJ, et al. Gene control of tyrosine kinase TIE2 and vascular manifestations of infections. *Proc Natl Acad Sci U S A*. (2016) 113:2472–7. doi: 10.1073/pnas.1519467113
 28. Higgins SJ, Purcell LA, Silver KL, Tran V, Crowley V, Hawkes M, et al. Dysregulation of angiopoietin-1 plays a mechanistic role in the pathogenesis of cerebral malaria. *Sci Transl Med*. (2016) 8:358ra128. doi: 10.1126/scitranslmed.aaf6812
 29. Hendrickson CM, Matthey MA. Endothelial biomarkers in human sepsis: pathogenesis and prognosis for ARDS. *Pulm Circ*. (2018) 8:2045894018769876. doi: 10.1177/2045894018769876
 30. Mikacenic C, Hahn WO, Price BL, Harju-Baker S, Katz R, Kain KC, et al. Biomarkers of endothelial activation are associated with poor outcome in critical illness. *PLoS One*. (2015) 10:e0141251. doi: 10.1371/journal.pone.0141251
 31. Hahn WO, Mikacenic C, Price BL, Harju-Baker S, Katz R, Himmelfarb J, et al. Host derived biomarkers of inflammation, apoptosis, and endothelial activation are associated with clinical outcomes in patients with bacteremia and sepsis regardless of microbial etiology. *Virulence*. (2016) 7:387–94. doi: 10.1080/21505594.2016.1144003
 32. Ghosh CC, Mukherjee A, David S, Knaus UG, Stearns-Kurosawa DJ, Kurosawa S, et al. Impaired function of the Tie-2 receptor contributes to vascular leakage and lethality in anthrax. *Proc Natl Acad Sci U S A*. (2012) 109:10024–9. doi: 10.1073/pnas.1120755109
 33. Page AV, Kotb M, McGeer A, Low DE, Kain KC, Liles WC. Systemic dysregulation of angiopoietin-1/2 in streptococcal toxic shock syndrome. *Clin Infect Dis*. (2011) 52:e157–61. doi: 10.1093/cid/cir125
 34. Page AV, Tarr PI, Watkins SL, Rajwans N, Petruzzello-Pellegrini TN, Marsden PA, et al. Dysregulation of angiopoietin 1 and 2 in *Escherichia coli* O157:H7 infection and the hemolytic-uremic syndrome. *J Infect Dis*. (2013) 208:929–33. doi: 10.1093/infdis/jit268
 35. Escadafal C, Nsanzabana C, Archer J, Chihota V, Rodriguez W, Dittrich S. New biomarkers and diagnostic tools for the management of fever in low- and middle-income countries: an overview of the challenges. *Diagnostics (Basel)*. (2017) 7:44. doi: 10.3390/diagnostics7030044
 36. Carrol ED, Mankhambo LA, Jeffers G, Parker D, Guiver M, Newland P, et al. The diagnostic and prognostic accuracy of five markers of serious bacterial infection in Malawian children with signs of severe infection. *PLoS One*. (2009) 4:e6621. doi: 10.1371/journal.pone.0006621
 37. Diez-Padriza N, Bassat Q, Machevo S, Quinto L, Morais L, Nhampossa T, et al. Procalcitonin and C-reactive protein for invasive bacterial pneumonia diagnosis among children in Mozambique, a malaria-endemic area. *PLoS One*. (2010) 5:e13226. doi: 10.1371/journal.pone.0013226
 38. Page AL, de Rekeneire N, Sayadi S, Aberrane S, Janssens AC, Dehoux M, et al. Diagnostic and prognostic value of procalcitonin and C-reactive protein in malnourished children. *Pediatrics*. (2014) 133:e363–70. doi: 10.1542/peds.2013-2112
 39. Levy A, Valero N, Espina LM, Anez G, Arias J, Mosquera J. Increment of interleukin 6, tumour necrosis factor alpha, nitric oxide, C-reactive protein and apoptosis in dengue. *Trans R Soc Trop Med Hyg*. (2010) 104:16–23. doi: 10.1016/j.trstmh.2009.06.013
 40. Kurtzhals JA, Adabayeri V, Goka BQ, Akanmori BD, Oliver-Commye JO, Nkrumah FK, et al. Low plasma concentrations of interleukin 10 in severe malarial anaemia compared with cerebral and uncomplicated malaria. *Lancet*. (1998) 351:1768–72. doi: 10.1016/S0140-6736(97)09439-7
 41. Liao B, Tang Y, Hu F, Zhou W, Yao X, Hong W, et al. Serum levels of soluble vascular cell adhesion molecules may correlate with the severity of dengue virus-1 infection in adults. *Emerg Microbes Infect*. (2015) 4:e24. doi: 10.1038/emi.2015.24
 42. Cui L, Lee YH, Thein TL, Fang J, Pang J, Ooi EE, et al. Serum metabolomics reveals serotonin as a predictor of severe dengue in the early phase of dengue fever. *PLoS Negl Trop Dis*. (2016) 10:e0004607. doi: 10.1371/journal.pntd.0004607

43. Jadhav M, Nayak M, Kumar S, Venkatesh A, Patel SK, Kumar V, et al. Clinical proteomics and cytokine profiling for dengue fever disease severity biomarkers. *OMICS*. (2017) 21:665–77. doi: 10.1089/omi.2017.0135
44. Nguyen TH, Lei HY, Nguyen TL, Lin YS, Huang KJ, Le BL, et al. Dengue hemorrhagic fever in infants: a study of clinical and cytokine profiles. *J Infect Dis*. (2004) 189:221–32. doi: 10.1086/380762
45. Othoro C, Lal AA, Nahlen B, Koech D, Orago AS, Udhayakumar V. A low interleukin-10 tumor necrosis factor- α ratio is associated with malaria anemia in children residing in a holoendemic malaria region in western Kenya. *J Infect Dis*. (1999) 179:279–82. doi: 10.1086/314548
46. Lyke KE, Burges R, Cissoko Y, Sangare L, Dao M, Diarra I, et al. Serum levels of the proinflammatory cytokines interleukin-1 beta (IL-1 β), IL-6, IL-8, IL-10, tumor necrosis factor alpha, and IL-12(p70) in Malian children with severe Plasmodium falciparum malaria and matched uncomplicated malaria or healthy controls. *Infect Immun*. (2004) 72:5630–7. doi: 10.1128/IAI.72.10.5630-5637.2004
47. Wang L, Chen RF, Liu JW, Yu HR, Kuo HC, Yang KD. Implications of dynamic changes among tumor necrosis factor- α (TNF- α), membrane TNF receptor, and soluble TNF receptor levels in regard to the severity of dengue infection. *Am J Trop Med Hyg*. (2007) 77:297–302. doi: 10.4269/ajtmh.2007.77.297
48. Arias J, Valero N, Mosquera J, Montiel M, Reyes E, Larreal Y, et al. Increased expression of cytokines, soluble cytokine receptors, soluble apoptosis ligand and apoptosis in dengue. *Virology*. (2014) 452–53:42–51. doi: 10.1016/j.virol.2013.12.027
49. Andrade BB, Reis-Filho A, Souza-Neto SM, Raffaele-Netto I, Camargo LM, Barral A, et al. Plasma superoxide dismutase-1 as a surrogate marker of vivax malaria severity. *PLoS Negl Trop Dis*. (2010) 4:e650. doi: 10.1371/journal.pntd.0000650
50. Butthep P, Chunchakan S, Yoksan S, Tangnararatchakit K, Chuansumrit A. Alteration of cytokines and chemokines during febrile episodes associated with endothelial cell damage and plasma leakage in dengue hemorrhagic fever. *Pediatr Infect Dis J*. (2012) 31:e232–8. doi: 10.1097/INF.0b013e31826fd456
51. Kinra P, Dutta V. Serum TNF alpha levels: a prognostic marker for assessment of severity of malaria. *Trop Biomed*. (2013) 30:645–53.
52. Soundravally R, Hoti SL, Patil SA, Cleetus CC, Zachariah B, Kadhiravan T, et al. Association between proinflammatory cytokines and lipid peroxidation in patients with severe dengue disease around defervescence. *Int J Infect Dis*. (2014) 18:68–72. doi: 10.1016/j.ijid.2013.09.022
53. Shabani E, Ouma BJ, Idro R, Bangirana P, Opoka RO, Park GS, et al. Elevated cerebrospinal fluid tumour necrosis factor is associated with acute and long-term neurocognitive impairment in cerebral malaria. *Parasite Immunol*. (2017) 39:e12438. doi: 10.1111/pim.12438
54. Richard-Greenblatt M, Boillat-Blanco N, Zhong K, Mbarack Z, Samaka J, Mlaganile T, et al. Prognostic accuracy of sTREM-1-based algorithms in febrile adults presenting to Tanzanian outpatient clinics. *Clin Infect Dis*. (2019) 70:1304–12. doi: 10.1093/cid/ciz419
55. Molyneux ME, Engelmann H, Taylor TE, Wirima JJ, Aderka D, Wallach D, et al. Circulating plasma receptors for tumour necrosis factor in Malawian children with severe falciparum malaria. *Cytokine*. (1993) 5:604–9. doi: 10.1016/S1043-4666(05)80011-0
56. Deloron P, Roux Lombard P, Ringwald P, Wallon M, Niyongabo T, Aubry P, et al. Plasma levels of TNF- α soluble receptors correlate with outcome in human falciparum malaria. *Eur Cytokine Netw*. (1994) 5:331–6.
57. Armah HB, Wilson NO, Sarfo BY, Powell MD, Bond VC, Anderson W, et al. Cerebrospinal fluid and serum biomarkers of cerebral malaria mortality in Ghanaian children. *Malar J*. (2007) 6:147. doi: 10.1186/1475-2875-6-147
58. Green S, Vaughn DW, Kalayanarooj S, Nimmannitya S, Suntayakorn S, Nisalak A, et al. Early immune activation in acute dengue illness is related to development of plasma leakage and disease severity. *J Infect Dis*. (1999) 179:755–62. doi: 10.1086/314680
59. Mikacenic C, Price BL, Harju-Baker S, O'Mahony DS, Robinson-Cohen C, Radella E, et al. A two-biomarker model predicts mortality in the critically ill with sepsis. *Am J Respir Crit Care Med*. (2017) 196:1004–11. doi: 10.1164/rccm.201611-2307OC
60. Anderson BJ, Calfee CS, Liu KD, Reilly JP, Kangelaris KN, Shashaty MGS, et al. Plasma sTNFR1 and IL8 for prognostic enrichment in sepsis trials: a prospective cohort study. *Crit Care*. (2019) 23:400. doi: 10.1186/s13054-019-2684-2
61. Wright SW, Lovelace-Macon L, Hantrakun V, Rudd KE, Teparrukkul P, Kosamo S, et al. sTREM-1 predicts mortality in hospitalized patients with infection in a tropical, middle-income country. *BMC Med*. (2020) 18:159. doi: 10.1186/s12916-020-01627-5
62. Guerrero CD, Arrieta AF, Ramirez ND, Rodriguez LS, Vega R, Bosch I, et al. High plasma levels of soluble ST2 but not its ligand IL-33 is associated with severe forms of pediatric dengue. *Cytokine*. (2013) 61:766–71. doi: 10.1016/j.cyto.2012.12.024
63. West TE, Wikraiphat C, Tandhavanant S, Ariyaprasert P, Suntornsut P, Okamoto S, et al. Patient characteristics, management, and predictors of outcome from severe community-onset staphylococcal sepsis in northeast Thailand: a prospective multicenter study. *Am J Trop Med Hyg*. (2017) 96:1042–9. doi: 10.4269/ajtmh.16-0606
64. Flores-Mendoza LK, Estrada-Jimenez T, Sedeno-Monge V, Moreno M, Manjarrez MDC, Gonzalez-Ochoa G, et al. IL-10 and socs3 are predictive biomarkers of dengue hemorrhagic fever. *Mediators Inflamm*. (2017) 2017:5197592. doi: 10.1155/2017/5197592
65. Brasier AR, Ju H, Garcia J, Spratt HM, Victor SS, Forshey BM, et al. A three-component biomarker panel for prediction of dengue hemorrhagic fever. *Am J Trop Med Hyg*. (2012) 86:341–8. doi: 10.4269/ajtmh.2012.11-0469
66. Suharti C, van Gorp EC, Dolmans WM, Setiati TE, Hack CE, Djokomoeljanto R, et al. Cytokine patterns during dengue shock syndrome. *Eur Cytokine Netw*. (2003) 14:172–7.
67. Bozza FA, Cruz OG, Zagne SM, Azeredo EL, Nogueira RM, Assis EF, et al. Multiplex cytokine profile from dengue patients: MIP-1 β and IFN- γ as predictive factors for severity. *BMC Infect Dis*. (2008) 8:86. doi: 10.1186/1471-2334-8-86
68. Rovira-Vallbona E, Moncunill G, Bassat Q, Aguilar R, Machevo S, Puyal L, et al. Low antibodies against Plasmodium falciparum and imbalanced pro-inflammatory cytokines are associated with severe malaria in Mozambican children: a case-control study. *Malar J*. (2012) 11:181. doi: 10.1186/1475-2875-11-181
69. Kumar Y, Liang C, Bo Z, Rajapakse JC, Ooi EE, Tannenbaum SR. Serum proteome and cytokine analysis in a longitudinal cohort of adults with primary dengue infection reveals predictive markers of DHF. *PLoS Negl Trop Dis*. (2012) 6:e1887. doi: 10.1371/journal.pntd.0001887
70. Oliveira R, Cordeiro MT, Moura P, Baptista Filho PNB, Braga-Neto UM, Marques ETAJ, et al. Serum cytokine/chemokine profiles in patients with dengue fever (DF) and dengue hemorrhagic fever (FHD) by using protein array. *J Clin Virol*. (2017) 89:39–45. doi: 10.1016/j.jcv.2017.02.007
71. Yong YK, Tan HY, Jen SH, Shankar EM, Natkunam SK, Sathar J, et al. Aberrant monocyte responses predict and characterize dengue virus infection in individuals with severe disease. *J Transl Med*. (2017) 15:121. doi: 10.1186/s12967-017-1226-4
72. Conroy AL, Gelvez M, Hawkes M, Rajwans N, Tran V, Liles WC, et al. Host biomarkers are associated with progression to dengue haemorrhagic fever: a nested case-control study. *Int J Infect Dis*. (2015) 40:45–53. doi: 10.1016/j.ijid.2015.07.027
73. Erdman LK, Dhabangi A, Musoke C, Conroy AL, Hawkes M, Higgins S, et al. Combinations of host biomarkers predict mortality among Ugandan children with severe malaria: a retrospective case-control study. *PLoS One*. (2011) 6:e17440. doi: 10.1371/journal.pone.0017440
74. Conroy AL, Hawkes M, McDonald CR, Kim H, Higgins SJ, Barker KR, et al. Host biomarkers are associated with response to therapy and long-term mortality in pediatric severe malaria. *Open Forum Infect Dis*. (2016) 3:ofw134. doi: 10.1093/ofid/ofw134
75. Wilson NO, Jain V, Roberts CE, Lucchi N, Joel PK, Singh MP, et al. CXCL4 and CXCL10 predict risk of fatal cerebral malaria. *Dis Markers*. (2011) 30:39–49. doi: 10.1155/2011/828256
76. Erdman LK, Petes C, Lu Z, Dhabangi A, Musoke C, Cserti-Gazdewich CM, et al. Chitinase 3-like 1 is induced by *Plasmodium falciparum*

- malaria and predicts outcome of cerebral malaria and severe malarial anaemia in a case-control study of African children. *Malar J.* (2014) 13:279. doi: 10.1186/1475-2875-13-279
77. Conroy AL, Hawkes MT, Elphinstone R, Opoka RO, Namasopo S, Miller C, et al. Chitinase-3-like 1 is a biomarker of acute kidney injury and mortality in paediatric severe malaria. *Malar J.* (2018) 17:82. doi: 10.1186/s12936-018-2225-5
 78. Conroy AL, Phiri H, Hawkes M, Glover S, Mallewa M, Seydel KB, et al. Endothelium-based biomarkers are associated with cerebral malaria in Malawian children: a retrospective case-control study. *PLoS One.* (2010) 5:e15291. doi: 10.1371/journal.pone.0015291
 79. Wright JK, Hayford K, Tran V, Al Kibria GM, Baqui A, Manajjir A, et al. Biomarkers of endothelial dysfunction predict sepsis mortality in young infants: a matched case-control study. *BMC Pediatr.* (2018) 18:118. doi: 10.1186/s12887-018-1087-x
 80. Mita-Mendoza NK, van de Hoef DL, Lopera-Mesa TM, Doumbia S, Konate D, Doumbouya M, et al. A potential role for plasma uric acid in the endothelial pathology of *Plasmodium falciparum* malaria. *PLoS One.* (2013) 8:e54481. doi: 10.1371/journal.pone.0054481
 81. Greco M, Palumbo C, Sicuro F, Lobreglio G. Soluble Fms-like tyrosine kinase-1 is a marker of endothelial dysfunction during sepsis. *J Clin Med Res.* (2018) 10:700–6. doi: 10.14740/jocmr3505w
 82. Klesney-Tait J, Turnbull IR, Colonna M. The TREM receptor family and signal integration. *Nat Immunol.* (2006) 7:1266–73. doi: 10.1038/ni1411
 83. Charles PE, Noel R, Massin F, Guy J, Bollaert PE, Quenot JP, et al. Significance of soluble triggering receptor expressed on myeloid cells-1 elevation in patients admitted to the intensive care unit with sepsis. *BMC Infect Dis.* (2016) 16:559. doi: 10.1186/s12879-016-1893-4
 84. Jedynak M, Siemiatkowski A, Mroczko B, Groblewska M, Milewski R, Szmitekowski M. Soluble TREM-1 serum level can early predict mortality of patients with sepsis, severe sepsis and septic shock. *Arch Immunol Ther Exp (Warsz).* (2018) 66:299–306. doi: 10.1007/s00005-017-0499-x
 85. Tahar R, Albergaria C, Zeghidour N, Ngane VF, Basco LK, Roussillon C. Plasma levels of eight different mediators and their potential as biomarkers of various clinical malaria conditions in African children. *Malar J.* (2016) 15:337. doi: 10.1186/s12936-016-1378-3
 86. Bellos I, Fitrou G, Daskalakis G, Thomakos N, Papantoniou N, Pergialiotis V. Soluble TREM-1 as a predictive factor of neonatal sepsis: a meta-analysis. *Inflamm Res.* (2018) 67:571–8. doi: 10.1007/s00011-018-1149-4
 87. Jain V, Lucchi NW, Wilson NO, Blackstock AJ, Nagpal AC, Joel PK, et al. Plasma levels of angiopoietin-1 and -2 predict cerebral malaria outcome in Central India. *Malar J.* (2011) 10:383. doi: 10.1186/1475-2875-10-383
 88. Lovegrove FE, Tangpukdee N, Opoka RO, Lafferty EI, Rajwans N, Hawkes M, et al. Serum angiopoietin-1 and -2 levels discriminate cerebral malaria from uncomplicated malaria and predict clinical outcome in African children. *PLoS One.* (2009) 4:e4912. doi: 10.1371/journal.pone.0004912
 89. Conroy AL, Lafferty EI, Lovegrove FE, Krudsood S, Tangpukdee N, Liles WC, et al. Whole blood angiopoietin-1 and -2 levels discriminate cerebral and severe (non-cerebral) malaria from uncomplicated malaria. *Malar J.* (2009) 8:295. doi: 10.1186/1475-2875-8-295
 90. Petersen JE, Mkumbaye SI, Vaaben AV, Manjurano A, Lyimo E, Kavishe RA, et al. Plasma Ang2 and ADAM17 levels are elevated during clinical malaria; Ang2 level correlates with severity and expression of EPCR-binding PfEMP1. *Sci Rep.* (2016) 6:35950. doi: 10.1038/srep35950
 91. Yeo TW, Lampah DA, Gitawati R, Tjitra E, Kenangalem E, Piera K, et al. Angiopoietin-2 is associated with decreased endothelial nitric oxide and poor clinical outcome in severe falciparum malaria. *Proc Natl Acad Sci U S A.* (2008) 105:17097–102. doi: 10.1073/pnas.0805782105
 92. Conroy AL, Glover SJ, Hawkes M, Erdman LK, Seydel KB, Taylor TE, et al. Angiopoietin-2 levels are associated with retinopathy and predict mortality in Malawian children with cerebral malaria: a retrospective case-control study*. *Crit Care Med.* (2012) 40:952–9. doi: 10.1097/CCM.0b013e3182373157
 93. Abdi AI, Fegan G, Muthui M, Kiragu E, Musyoki JN, Opiyo M, et al. *Plasmodium falciparum* antigenic variation: relationships between widespread endothelial activation, parasite PfEMP1 expression and severe malaria. *BMC Infect Dis.* (2014) 14:170. doi: 10.1186/1471-2334-14-170
 94. Prapansilp P, Medana I, Mai NT, Day NP, Phu NH, Yeo TW, et al. A clinicopathological correlation of the expression of the angiopoietin-Tie-2 receptor pathway in the brain of adults with *Plasmodium falciparum* malaria. *Malar J.* (2013) 12:50. doi: 10.1186/1475-2875-12-50
 95. Hanson J, Lee SJ, Hossain MA, Anstey NM, Charunwatthana P, Maude RJ, et al. Microvascular obstruction and endothelial activation are independently associated with the clinical manifestations of severe falciparum malaria in adults: an observational study. *BMC Med.* (2015) 13:122. doi: 10.1186/s12916-015-0365-9
 96. Ricciuto DR, dos Santos CC, Hawkes M, Toltl LJ, Conroy AL, Rajwans N, et al. Angiopoietin-1 and angiopoietin-2 as clinically informative prognostic biomarkers of morbidity and mortality in severe sepsis. *Crit Care Med.* (2011) 39:702–10. doi: 10.1097/CCM.0b013e318206d285
 97. Mankhambo LA, Banda DL, The IPD Study Group, Jeffers G, White SA, Balmer P, et al. The role of angiogenic factors in predicting clinical outcome in severe bacterial infection in Malawian children. *Crit Care.* (2010) 14:R91. doi: 10.1186/cc9025
 98. Zhang R, Rai U, Ibrahim NBM, Amazouz Y, Soo J, Conroy AL, et al. Dysregulation of angiopoietin-Tie-2 axis in Ugandan children hospitalized with pneumonia. *Cytokine.* (2020) 133:155175. doi: 10.1016/j.cyto.2020.155175
 99. Gutbier B, Neuhauß AK, Reppe K, Ehrler C, Santel A, Kaufmann J, et al. Prognostic and pathogenic role of angiopoietin-1 and -2 in pneumonia. *Am J Respir Crit Care Med.* (2018) 198:220–31. doi: 10.1164/rccm.201708-1733OC
 100. Burke HB. Predicting clinical outcomes using molecular biomarkers. *Biomark Cancer.* (2016) 8:89–99. doi: 10.4137/BIC.S33380
 101. Highsmith J. *Biomarkers: Technologies and Global Markets.* (2016).
 102. World Health Organization. *Mapping the Landscape of Diagnostics for Sexually Transmitted Infections* (2004).
 103. Huang D, Ying H, Jiang D, Liu F, Tian Y, Du C, et al. Rapid and sensitive detection of interleukin-6 in serum via time-resolved lateral flow immunoassay. *Anal Biochem.* (2020) 588:113468. doi: 10.1016/j.ab.2019.113468
 104. Misbah G. Health care worker's experience on Rapid Diagnostic Test (Rdt) for Malaria and their views on community acceptability of Rdt in Rubavu District, Rwanda: qualitative study. *Malaria Control Elimin.* (2017) 6:1000157. doi: 10.4172/2470-6965.1000157
 105. Mushi AK, Massaga JJ, Mandara CI, Mubyazi GM, Francis F, Kamugisha M, et al. Acceptability of malaria rapid diagnostic tests administered by village health workers in Pangani District, North eastern Tanzania. *Malar J.* (2016) 15:439. doi: 10.1186/s12936-016-1495-z
 106. Baiden F, Owusu-Agyei S, Okyere E, Tivura M, Adjei G, Chandramohan D, et al. Acceptability of rapid diagnostic test-based management of Malaria among caregivers of under-five children in rural Ghana. *PLoS One.* (2012) 7:e45556. doi: 10.1371/journal.pone.0045556
 107. Mukanga D, Tibenderana JK, Peterson S, Pariyo GW, Kiguli J, Waiswa P, et al. Access, acceptability and utilization of community health workers using diagnostics for case management of fever in Ugandan children: a cross-sectional study. *Malar J.* (2012) 11:121. doi: 10.1186/1475-2875-11-121
 108. Hopkins H, Bruxvoort KJ, Cairns ME, Chandler CI, Leurent B, Anshah EK, et al. Impact of introduction of rapid diagnostic tests for malaria on antibiotic prescribing: analysis of observational and randomised studies in public and private healthcare settings. *BMJ.* (2017) 356:j1054. doi: 10.1136/bmj.j1054
 109. Charani E, Smith I, Skodvin B, Perozziello A, Lucet J-C, Lescure F-X, et al. Investigating the cultural and contextual determinants of antimicrobial stewardship programmes across low-, middle- and high-income countries-A

- qualitative study. *PLoS One*. (2019) 14:e0209847. doi: 10.1371/journal.pone.0209847
110. Calu Costa J, Wehrmeister FC, Barros AJ, Victora CG. Gender bias in careseeking practices in 57 low- and middle-income countries. *J Glob Health*. (2017) 7:010418. doi: 10.7189/jogh.07.010418
111. Khera R, Jain S, Lodha R, Ramakrishnan S. Gender bias in child care and child health: global patterns. *Arch Dis Child*. (2014) 99:369–74. doi: 10.1136/archdischild-2013-303889
112. Alkema L, Chao F, You D, Pedersen J, Sawyer CC. National, regional, and global sex ratios of infant, child, and under-5 mortality and identification of countries with outlying ratios: a systematic assessment. *Lancet Glob Health*. (2014) 2:e521–e30. doi: 10.1016/S2214-109X(14)70280-3

Conflict of Interest: KCK is a named inventor on a patent “Biomarkers for early determination of a critical or life-threatening response to illness and/or treatment response” held by the University Health Network.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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