



Vascular Endothelium in Neonatal Sepsis: Basic Mechanisms and Translational Opportunities

Carlo Pietrasanta^{1,2,3*}, Lorenza Pagni¹, Andrea Ronchi¹, Ilaria Bottino¹, Beatrice Ghirardi¹, Guzman Sanchez-Schmitz^{3,4}, Francesco Borriello^{3,4,5,6,7}, Fabio Mosca^{1,2} and Ofer Levy^{3,4,8}

¹ Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Neonatal Intensive Care Unit, Milan, Italy, ² Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy, ³ Precision Vaccines Program, Division of Infectious Diseases, Boston Children's Hospital, Boston, MA, United States, ⁴ Harvard Medical School, Boston, MA, United States, ⁵ Division of Immunology, Boston Children's Hospital, Boston, MA, United States, ⁶ Department of Translational Medical Sciences, Center for Basic and Clinical Immunology Research (CISIR), University of Naples Federico II, Naples, Italy, ⁷ World Allergy Organisation Center of Excellence, Naples, Italy, ⁸ Broad Institute of MIT and Harvard, Cambridge, MA, United States

OPEN ACCESS

Edited by:

Amelia Licari,
University of Pavia, Italy

Reviewed by:

Oktay Kirak,
The Scripps Research Institute,
United States
Mario Barbarini,
ASST Lariana, Italy

*Correspondence:

Carlo Pietrasanta
carlo.pietrasanta@unimi.it

Specialty section:

This article was submitted to
Pediatric Immunology,
a section of the journal
Frontiers in Pediatrics

Received: 28 May 2019

Accepted: 30 July 2019

Published: 13 August 2019

Citation:

Pietrasanta C, Pagni L, Ronchi A, Bottino I, Ghirardi B, Sanchez-Schmitz G, Borriello F, Mosca F and Levy O (2019) Vascular Endothelium in Neonatal Sepsis: Basic Mechanisms and Translational Opportunities. *Front. Pediatr.* 7:340. doi: 10.3389/fped.2019.00340

Neonatal sepsis remains a major health issue worldwide, especially for low-birth weight and premature infants, with a high risk of death and devastating sequelae. Apart from antibiotics and supportive care, there is an unmet need for adjunctive treatments to improve the outcomes of neonatal sepsis. Strong and long-standing research on adult patients has shown that vascular endothelium is a key player in the pathophysiology of sepsis and sepsis-associated organ failure, through a direct interaction with pathogens, leukocytes, platelets, and the effect of soluble circulating mediators, in part produced by endothelial cells themselves. Despite abundant evidence that the neonatal immune response to sepsis is distinct from that of adults, comparable knowledge on neonatal vascular endothelium is much more limited. Neonatal endothelial cells express lower amounts of adhesion molecules compared to adult ones, and present a reduced capacity to neutralize reactive oxygen species. Conversely, available evidence on biomarkers of endothelial damage in neonates is not as robust as in adult patients, and endothelium-targeted therapeutic opportunities for neonatal sepsis are almost unexplored. Here, we summarize current knowledge on the structure of neonatal vascular endothelium, its interactions with neonatal immune system and possible endothelium-targeted diagnostic and therapeutic tools for neonatal sepsis. Furthermore, we outline areas of basic and translational research worthy of further study, to shed light on the role of vascular endothelium in the context of neonatal sepsis.

Keywords: neonatal infection, neonatal inflammation, glycocalyx, sepsis therapy, sepsis diagnosis, sepsis biomarkers, newborn

INTRODUCTION

Our knowledge of the anatomy and function of the immune system in early life have greatly improved over the past few decades. Neonatal immune responses to exogenous stimuli, like pathogens and vaccines, presents distinct features compared to adults, including:

- a) A clear prevalence of innate vs. adaptive immune mechanisms, even in cells classically considered “adaptive” such as CD8⁺ T cells (1–3);
- b) Diminished chemotaxis and phagocytic activity of polymorphonuclear granulocytes (PMNs) and of monocytes under stress conditions *in vitro* (Marodi 1980, 135 Eur J Pediatr; Fox 2005 Cytokine; Arinola 2003 Afr J Med Med Sci);
- c) Diminished pro-inflammatory activity and T-helper 1 (Th1) polarization of antigen-presenting cells (APCs) and lymphocytes, characterized by weak production of interleukin (IL)-12p70, interferon (IFN)- γ and tumor necrosis factor (TNF)- α in response to most vaccines and pattern recognition receptors (PRRs) agonists, with the partial exception of Bacille Calmette Gurerin (BCG) vaccine and Toll-like receptor (TLR)-8 agonists (4–6);
- d) Relatively high production of IL-17 and IL-23, hallmarks of Th2-Th17 polarization of immune response, of IL-6 and IL-10 (especially in preterm neonates), the latter a powerful anti-inflammatory cytokine (7).
- e) The influence of distinctive immunomodulatory blood components, such as maternal antibodies, high concentrations of adenosine, and reduced concentrations of complement (4, 8).

Despite such progress, we still lack a comprehensive grasp of the complex immune mechanisms regulating the pathophysiology of neonatal sepsis.

Basic and translational research has shed some light on features of neonatal immune response to sepsis. Nonetheless, the literature is incomplete and sometimes apparently contradictory. For example, the aforementioned lower production of proinflammatory cytokines is highly context-dependent: while pro-inflammatory/Th1-polarizing responses by isolated neonatal APCs *in vitro* to pure PRR agonists are frequently impaired, responses by the same cells to live microbes (akin to the context of sepsis) may be robust (9, 10). Furthermore, *in vivo*, clinical and experimental evidences

suggest an exaggerate susceptibility to systemic inflammatory response during the acute phase of sepsis, by both neonatal experimental animal models and human patients, especially if born preterm (11, 12).

This incomplete understanding of neonatal immune pathophysiology has also impaired the development of new therapeutic agents: sepsis therapy in neonates (and similarly in adults) has not evolved significantly from antibiotics, fluid resuscitation and vasopressors. Indeed, new preventive and therapeutic strategies are needed to limit the burden of neonatal sepsis (13).

In adult patients, the vascular endothelium (VE) has received substantial attention by scientists and clinicians, as there is increasing evidence on its role in the pathophysiology of sepsis and sepsis-associated organ damage (14, 15).

VE interacts with, and responds to leukocytes, soluble mediators, pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) involved in the pathogenesis of sepsis. It has a major role in the regulation of vascular permeability (16), in the modulation of local inflammatory responses (17) and, ultimately, in the development and progression of sepsis-associated multi-organ failure (15, 18, 19). In comparison, the role of VE in the pathophysiology of neonatal sepsis has been marginally investigated.

As summarized in **Figure 1**, we recap in this review the current evidence supporting a direct interplay between the immune system and VE during neonatal sepsis. We emphasize which research tools have been used, and others that could be used in the future to monitor endothelial function in neonates during health and sepsis, and we discuss potential immune-therapeutic interventions targeting the VE that deserve age-specific investigation in the context of neonatal sepsis.

NEONATAL IMMUNE RESPONSE TO SEPSIS

According to the most recent definition for adult patients, sepsis is a life-threatening organ dysfunction caused by a dysregulated host response to infection (20). It is by now well-established that adult sepsis criteria do not sensitively detect neonatal sepsis, reflecting a distinct age-specific host-pathogen interactions (21). Of note, there is currently no consensus definition of neonatal sepsis (22). Most datasets report an overall incidence of culture-positive early-onset neonatal sepsis (EOS) of $\sim 0.98/1,000$ live births, rising up to 1.4–4 cases/1,000 live births in very low birth weight infants (VLBWI), the more susceptible population (23, 24). Late-onset sepsis (LOS), has a more variable incidence, up to 30% in extremely low-birth weight infants (ELBWI) (25, 26).

Mortality rate is the highest for sepsis caused by Gram-negative bacteria (26) and, interestingly, more than half (50–57%) of non-survivor neonates die within the first 3 days of sepsis onset (27). This data, together with evidence linking mortality during the acute phase of neonatal sepsis to a dysregulated pro-inflammatory response (28, 29), highlight the

Abbreviations: ADAMTS13, a disintegrin and metalloproteinase with a thrombospondin type 1 motif, member 13; APACHE-II, Acute Physiology and Chronic Health Disease Classification System II; CD, cluster of differentiation; CFU, colony-forming unit; CXCL-8, C-X-C motif chemokine ligand 8; Flt-1, FLT1, fms-related tyrosine kinase 1; G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; MHC-II, major histocompatibility complex - class II; MIP-1 β , macrophage inflammatory protein 1 β . NOD-like receptor, nucleotide-binding oligomerization domain-like receptors; PAI-1, plasminogen activator inhibitor-1; RANTES, Regulated upon Activation, Normal T cell Expressed, and Secreted (chemokine); RIG-I-like receptor, retinoic acid-inducible gene I-like receptors; SOFA, Sequential Organ Failure Assessment; Tie, tyrosine kinase with immunoglobulin-like and EGF-like domains; VCAM, vascular cell adhesion molecule.

fetus (49) and the neonate (50). Nonetheless, considered the high morbidity and mortality rates of neonatal sepsis, further and stronger evidences need to be acquired before the concept of an “evolutionary favorable” tolerance strategy could be applied in this context.

The Inflammatory Response in Neonatal Sepsis

In neonates as well as in adults, the propagation of invading pathogens to the blood stream is followed by a systemic inflammatory response (19). As the contradictory data from several studies seem to indicate, measuring and deciphering systemic inflammation during neonatal sepsis is not trivial.

First, several studies *in vitro* showed that isolated neonatal immune cells, such as monocytes or dendritic cells, produce lower amounts of TNF, IL-1 β , and IFN- γ compared to adult cells when stimulated with different PRRs agonists or with microorganisms, but equal or even higher amounts of Th17-promoting cytokines (IL-23, IL-6) and IL-10 (7, 51–57). These results were also replicated with human whole blood *ex vivo* (58). In animal models of polymicrobial sepsis, neonates tend to show lower absolute plasma concentrations of TNF- α , IL-1 α/β , IL-12, GM-CSF, CCL5 (RANTES), macrophage inflammatory protein (MIP)-1 β and IFN- γ as compared to adults, when challenged with an equally-lethal dose of “cecal slurry” (59).

Conversely, some evidences support the concept of a strong systemic inflammatory response in neonates. First, IL-18 was found to be higher in both healthy and septic neonates compared to adults, and strongly correlated to morbidity and mortality in neonatal sepsis (60, 61). Second, other cytokines such as IL-6 and IL-10, the latter being an important soluble mediator for resolution of inflammation (62), seem to be produced by septic neonates in comparable or sometimes higher amounts compared to adult patients (63, 64), and several studies correlated the adverse outcomes of sepsis (i.e., septic shock or non survival) with higher concentrations of IL-10 and higher IL-10/TNF- α ratio, in neonates as well as in adults (33, 65, 66). Third, higher mortality rates and increased plasma levels of TNF, IL-6 and macrophage chemotactic protein (MCP)-1 have been reported in newborn mice challenged with a really equal dose (per gram of weight) of pure bacterial lipopolysaccharide (LPS), a TLR4 agonist, or poly(I:C), a TLR3 agonist, compared to adult mice (11). Overall, clinical and experimental studies highlight the complexity of systemic inflammatory response in neonates: notwithstanding neonatal cells show a distinct and often impaired production of cytokines and chemokines (especially for some classical pro-inflammatory cytokines) compared to adult cells, the impaired ability *in vivo* to contain locally the invading pathogen(s) leads to high bacterial loads in the blood stream and distant organs. This, ultimately, fuels an intense, systemic, acute pro-inflammatory response that seems responsible for high morbidity and mortality in the acute phase of sepsis (67).

An almost missing piece of this puzzle, at least for what concerns neonatal sepsis, is the role of VE during the systemic inflammatory response boosted by invading pathogens. VE lies at the crossroad between invading pathogen, inflammatory

response, coagulation system and organ damage (68, 69), all key components in the pathogenesis of sepsis, in adults as well as in neonates.

STRUCTURE OF THE VE IN ADULTS AND NEONATES

The endothelial wall structure, in neonates as well as in adults, is composed by a single-layer of endothelial cells (ECs), sealed by intercellular junctions and covered by the extra-cellular, intraluminal structure glycocalyx.

Structure of the Endothelial Cells Layer

Cell-to-cell junctions in the VE comprise VE-cadherins, forming adherent junctions (AJs), and tight junctions (TJs). VE-cadherins are surface adhesion glycoproteins that form a “zipper-like” structure at the base of ECs and are connected to the underlying components of the cytoskeleton, like β -catenin (70). TJs, instead, occupy the apical part of intercellular space and predominantly consist of individual protein filaments of occludin and claudin, that form the so-called “zonulae occludentes.”

Altogether, VE-cadherins and TJs seal the intercellular space between ECs and regulate the paracellular flow of fluids and solutes that occurs passively as the result of oncotic and hydrostatic pressures at both sides of endothelial layer according to the Starling’s principle. The transcellular pathway of molecules through ECs, instead, is an active process that requires either cell fenestration or a transport system through intracellular vesicles.

Besides VE-cadherins and TJs, other adhesion proteins contribute to the integrity of vascular endothelial barrier, as intercellular adhesion molecule-1 (ICAM-1, CD54), ICAM-2, platelet endothelial cell adhesion molecule (PECAM)-1 (CD31), CD34, and endoglin (70). On the apical surface, facing the vascular lumen, ECs express transmembrane glycoproteins that mediate the first contact and rolling of leukocytes, mainly P-selectin and E-selectin, and those mediating the subsequent firm adhesion and extravasation processes (immunoglobulin-like molecules as ICAM-1, ICAM-2 PECAM-1, and VCAM-1, interacting with integrins expressed on leukocytes). Leukocytes-endothelium adhesion molecules have been subject of an excellent review (71).

Several aspects of endothelium structure and physiology in neonates, especially if born preterm, are different from adults, and may play a significant role in the pathogenesis of neonatal sepsis (Figure 1).

ECs isolated from fetuses up to 22 weeks of gestational age do not express P-selectin, and intracellular storage of P-selectin in preterm neonates is less than half that of term neonates, at steady state; this, in turn, contributes to the observed defect in rolling, adhesion and crawling of neonatal neutrophils (72). Similarly defective up-regulation of inducible E-selectin, ICAM-1 and VCAM-1 upon LPS stimulation of human umbilical vein ECs (HUVECs) from preterm neonates has been noted (73, 74). Of note, it has also been shown that endothelial selectins, E-selectin and P-selectin, are expressed at least partially during fetal development in humans: in particular, P-selectin seems

expressed at adult levels since 11 weeks of gestation while, interestingly, E-selectin reaches adult levels on ECs around 32 weeks (75). Overall, endothelium-mediated adhesion and rolling of neutrophils, either at steady state or during an inflammatory process, seem to be impaired in neonates as compared to adults, and among neonates the severity of this impairment seems gestational age-dependent.

Furthermore, differences between adults and neonates exist also for what concerns endothelium metabolism: for example, HUVECs, as compared to human microvascular ECs (HMVEC) derived from adults, express higher levels of hydrogen peroxide and lower amounts of ROS-neutralizing enzymes, such as superoxide dismutase 2 (SOD2) and catalase, in response to hyperglycemia (76): this study suggests a potential epigenetic-mediated difference between neonatal and adult ECs in the capacity to neutralize ROS. Considering that both ROS and reactive nitrogen species (RNS) facilitate endothelial dysfunction, increase cell wall permeability and induce a pro-coagulant state, especially in non-physiological conditions of stress like sepsis, it is conceivable that reduced neutralizing capacity of ROS and RNS by neonatal ECs might play a role in driving endothelial metabolic dysfunction during a septic state (77).

Finally, two concepts regarding the investigation of endothelial cells *in vitro* need to be taken in consideration and should prompt more age-specific research. First, even in the context of adult sepsis, most researchers use HUVECs as a model because of their availability and practicality. If we accept that differences between neonatal and adult endothelium exist, most of the results obtained with such model might more closely reflect neonatal rather than adult physiology. Second, *in vitro* models to investigate neonatal VE should carefully replicate all age-specific microphysiological conditions, such as the quality and origin of plasma, the basement substrate and the interacting immune cells, in order to more accurately model age-specific biology that is relevant *in vivo*, thereby enhancing translational research (78).

The Glycocalyx

Above the surface of ECs, and directly in contact with blood, lies the glycocalyx, a 0.1–3 μm thick multi-layer structure of glycoproteins whose functions are to hide most of the adhesion molecules expressed by ECs, maintain the selective permeability of endothelial barrier, inhibit blood coagulation, and serve as a mechanotransducer between blood flow and the endothelial cell wall (79).

The glycocalyx is a dense network composed by two families of protein cores anchored to the EC wall, syndecans and glypicans, a few protein cores not anchored to ECs, such as perlecan and versican, and several secreted glycosaminoglycans (GAGs) attached to the protein cores, like heparan sulfate and hyaluronan (80). Glycocalyx is negatively charged and its structure does not allow the passage of proteins larger than 70 kDa. It repels most plasma proteins, including immunoglobulins, with the exception of albumin, that with a molecular weight of 67 kDa and its amphoteric nature can bind and cross, at least partially, the surface of glycocalyx: the net effect of this interaction is that, under physiological conditions, albumin itself

contributes to tighten the glycoprotein network and remains trapped on the glycocalyx, to the point that only 5% of body albumin normally leaks into the interstitial space (81). Thus, the glycocalyx significantly contributes to maintain the oncotic pressure gradient across the endothelial barrier.

Despite the growing body of literature on adult patients and experimental models on adult animals, the structure and function of glycocalyx in neonates have only been marginally explored. Over 40 years ago, it was shown that neonatal VE has a permeability to albumin that is 3 to 4 times higher than that in adults (82) and, in experimental animal models, that the fetus has a protein reflection coefficient at the endothelial interface, an index of resistance to the extravasation of proteins, that is just 70 to 90% of the adult one (83). In addition, few authors explored the shedding of glycocalyx components into the circulation during systemic inflammation states, such as sepsis, as summarized afterwards in this review. The glycocalyx structure has been studied in the context of necrotizing enterocolitis, with focus limited to the glycocalyx coating the luminal side of enterocytes (84). Studies directly investigating the biological structure of vascular endothelial glycocalyx in early life, either in the fetus or the preterm neonate, are still lacking.

INTERPLAY BETWEEN IMMUNE SYSTEM AND ENDOTHELIUM DURING NEONATAL SEPSIS

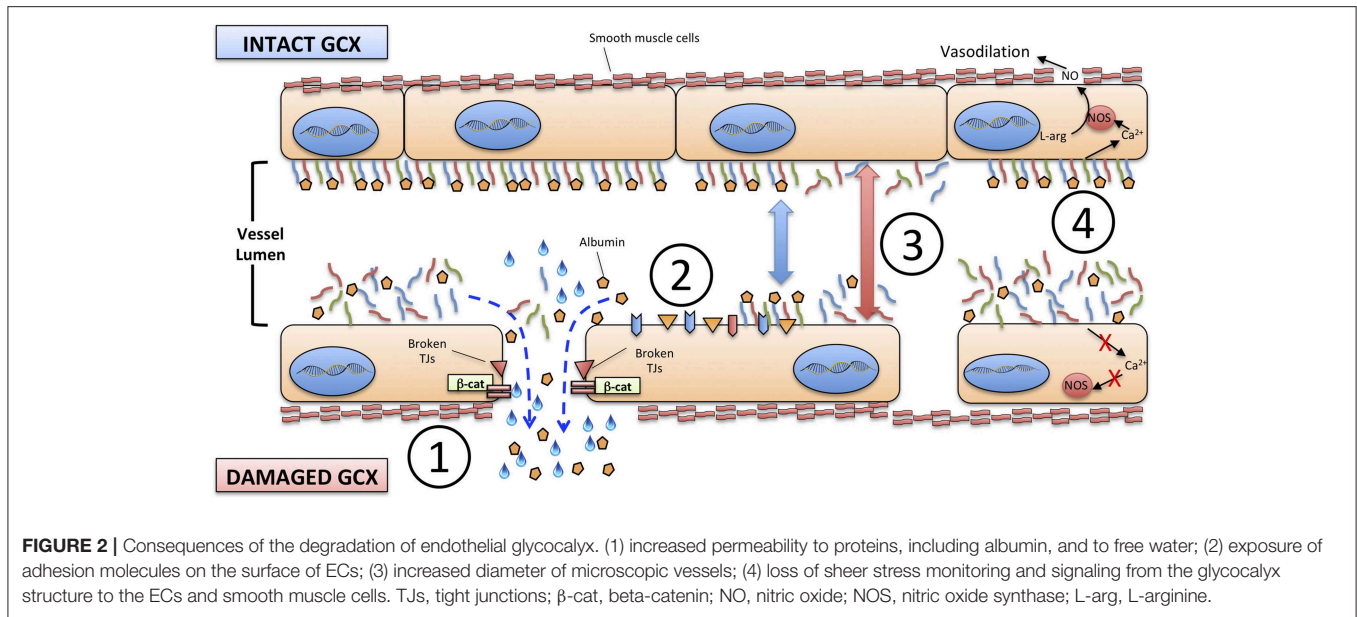
It has long been recognized that both ECs and the glycocalyx have an active role in inflammatory processes, and a deep functional connection with the immune system (85). Furthermore, the endothelium has an active role in the regulation of the coagulation process, which is another fundamental epiphenomenon of systemic inflammation and infections: thus, the triad (a) endothelium, (b) immune response, and (c) coagulation cascade need all be considered as a whole, at steady state as well as during systemic inflammation.

Glycocalyx During Neonatal Sepsis

Due to its apical position, the glycocalyx encounters circulating pathogens and blood solutes first. It has been shown that the glycocalyx is degraded in the presence of several stimuli, such as ROS, bacterial endotoxin (LPS), hyperglycemia, hypovolemia or pro-inflammatory cytokines, especially TNF- α and IL-1 β (80). This process is mediated by either direct damage to structural components or by the cleaving action of proteolytic and glycolytic enzymes, such as heparanase-I and metalloproteinases (MMPs) (86). Heparanase-I, in particular, is pre-stored in cytoplasmic granules of ECs and can be released upon stimulation of ECs by either cytokines or endotoxin.

The disruption of glycocalyx has four major consequences on VE (**Figure 2**):

- 1) An increased permeability of endothelium to proteins, including albumin, and consequently to free water;



- 2) The exposure of adhesion molecules on the surface of ECs, that favor the trapping and activation of immune cells: activated neutrophils, in particular, can also provide a determinant contribution to diffuse microvascular thrombosis, as outlined below;
- 3) The increased diameter of microscopic vessels in those areas where glycocalyx is more damaged, leading to non-homogeneous perfusion of capillary beds;
- 4) The loss of shear stress monitoring and signaling from the glycocalyx structure to the ECs, that contributes to the loss of vascular reactivity.

Robust evidence concerning the role of glycocalyx degradation in shaping the pathophysiology of neonatal sepsis is still lacking; nonetheless, its importance can be reasonably presumed when considering that the clinical course of severe neonatal sepsis and septic shock is characterized by systemic hypotension, hypoalbuminemia, diffuse edema, disseminated intravascular coagulation and, ultimately, multi-organ damage and failure (67, 87). Hypoalbuminemia and hypotension during neonatal sepsis are frequently tackled with albumin infusion, in the attempt to raise intravascular oncotic pressure despite the absence of scientific evidence, or evidence-based therapeutic individualization (88). In few septic neonates, indeed, albumin may not elicit the desired effect but, instead, worsen the water shift toward the extravascular space (89). The role of endothelial glycocalyx in these cases has yet to be explored.

Finally, circulating heparan sulfate, an end product of damaged glycocalyx, can induce a strong pro-inflammatory response and mitochondrial dysfunction in cardiomyocytes during experimental sepsis (90, 91). Myocardial dysfunction is one of the key mediators of systemic hypotension and organ hypoperfusion in the context of neonatal septic shock (92). Given the influence of glycocalyx end-products on

cardiomyocytes, it will be important to characterize how these circulating molecules affect the function of myocardium during neonatal sepsis.

Endothelial Cells in Neonatal Sepsis

ECs are directly involved in the pathophysiology of sepsis and can play several immune functions (85). ECs express a wide range of PRRs, including TLRs, RIG-I-like and NOD-like receptors (93), that directly interact with different PAMPs in the bloodstream, such as streptococcal toxins (94).

HUVECs and adult-derived ECs, like HMVECs, seem to express comparable amounts of TLRs, with high density of TLR1, 3 and 4 and the virtual absence of TLR7 and TLR8 (95). As in APCs, the activation of PRRs on ECs induces production of cytokines and chemokines. In septic neonates, high plasma concentrations of IL-1 β , IL-6, CXCL8 (IL-8), TNF α , and G-CSF are not mirrored by the elevation in the corresponding mRNA isolated from leukocytes, suggesting that cells other than leukocytes may contribute to the production of those pro-inflammatory mediators (96). Indeed, PAMPs, such as LPS, induce a strong upregulation of pro-inflammatory cytokines in ECs (85). Conversely, the effect of PAMPs on ECs is amplified by circulating pro-inflammatory cytokines that create a positive-feedback loop by inducing the upregulation of surface PRRs themselves (95).

Neonatal sepsis is characterized by high plasma levels of IL-6, CXCL8 (KC in murine models) and IL-18 (28, 60, 67, 97). Both human and experimental animal models suggest, that substantial amounts of these pro-inflammatory cytokines may be produced by activated ECs in the context of sepsis. For example, both IL-18 and IL-18R are expressed by adult atheroma-associated ECs (98), while IL-6 and CXCL8 are produced in significant amounts by ECs stimulated by LPS, TLR2 agonists

or other circulating pro-inflammatory cytokines like IL-1 (99–101). Further characterization of endothelium-derived pro-inflammatory cytokines in neonatal sepsis, as well as the effects of therapies targeting cytokine-induced endothelial activation during neonatal sepsis, is required.

The local effect of endothelium-derived cytokines is to create a gradient to guide neutrophils and monocytes to peripheral tissues. This effect on inflamed vasculature seems to be attenuated in septic neonates compared to adults and in preterm neonates compared to term ones: *in vitro* studies (42, 102) have shown how chemotaxis and transmigration of PMNs is impaired in septic neonates, because of the reduced expression of adhesion molecules (mainly E-selectin, P-selectin, and ICAM-1) on ECs and their respective ligands on leukocytes (74): this, in turn, may be due to depressed TNF- α production during systemic inflammation, as shown in experimental models of neonatal pulmonary infection (103). Interestingly, these data on inflammatory states were recently complemented by the observation that neonates present a significant endothelial activation at the steady-state (104). This seems supported by monocyte-derived TNF- α , and promotes leukocyte trafficking through peripheral tissues during the physiological neonatal period. Thus, overall, current evidences support the concept of a higher baseline activation of neonatal endothelium, possibly cytokine-mediated, followed by an impaired up-regulation of key functions (expression of adhesion molecules) during systemic inflammation (**Figure 1**).

Damaged or activated VE greatly supports the establishment of a prothrombotic environment, in order to contain the damage locally and to avoid the spread of the harmful stimulus through systemic circulation (105). In adults, disseminated intravascular coagulation is a major contributor to the establishment and progression of multi-organ failure (MOF) during severe sepsis and septic shock, together with mitochondrial dysfunction, direct cytotoxicity of bacterial toxins and cytokine-induced apoptosis (106). MOF is also typical of severe neonatal sepsis (67), and activated/damaged VE may play a significant role in its pathogenesis. In addition to promoting adhesion of leukocytes through the exposure of adhesion molecules, activated endothelia release large amounts of von Willebrand factor (VWF) promoting aggregation of platelets, granulocytes and monocytes. Granulocytes, in turn, release NETs, while activated monocytes express Tissue Factor that supports blood coagulation through fibrin formation.

The active interplay between damaged/inflamed endothelium, platelets, activated granulocytes, NETs and monocytes has been termed “immunothrombosis,” a concept that has only been recently demonstrated *in vivo* (107). The biological benefits (e.g., reduced diffusion of circulating pathogens to tissues and organs, reduced metabolic rate and oxygen consumption due to diminished perfusion) and harms (e.g., ischemic damage, shift toward anaerobic metabolism, reduced influx of immune cells) of immunothrombosis during sepsis have been the subject of a detailed review (108). These functions have been partially explored in children (109, 110). To our knowledge, the biology and systemic consequences of immunothrombosis, as well as the role of neonatal VE in this process, have not yet been

investigated in the context of neonatal sepsis. VE, platelets and leukocytes form a triad that cooperate in the establishment and progression of MOF during severe neonatal sepsis. In this context, immunothrombosis deserves a focused research effort to enlighten the pathogenesis and development of new therapeutics for neonatal sepsis.

HOW TO INVESTIGATE ENDOTHELIUM AND GLYCOCALYX DURING NEONATAL SEPSIS

Monitoring the structure and functional integrity of the VE has already been regarded, at least in adult patients, as a fundamental component of the patient’s evaluation during critical illness such as sepsis (111).

The most commonly evaluated function of VE in the context of neonatal sepsis is vasomotor regulation: albeit indirectly, the ability of endothelium to regulate vascular tone is currently assessed by visual inspection of skin color, evaluation of capillary refill (112), perfusion index (113) and less frequently, near-infrared spectroscopy (NIRS) of tissues (114). Furthermore, blood lactate is routinely evaluated in adult and neonatal sepsis as a marker of microcirculatory function and oxygen extraction by peripheral tissues (67, 115). All of the above are indirect and approximate methods, whose correlation with the real-time status of endothelial barrier or the integrity of glycocalyx is difficult to establish.

In the clinical context and experimental settings, two methodologies are currently gaining attention from physicians and scientists dealing with sepsis or septic shock in adult patients and, with some adaptation, could be applied in the context of neonatal sepsis: (1) imaging techniques, such as intravital microscopy, and (2) the quantification of biomarkers of endothelial damage.

Intravital Imaging

Before 1990, intravital microscopy was cumbersome and confined to experimental research laboratories providing little to no possibilities to translate observations and methodologies to the patient’s bedside. Over the last 30 years, hand-held, intravital microscopes became more portable and easy-to-use (15).

Three generations of devices based on different imaging technologies were subsequently introduced: orthogonally polarized spectral (OPS), sidestream dark-field (SDF) and incident dark-field (IDF) microscopes (116). Currently, only SDF and IDF imaging techniques are applied routinely to bedsides, with a growing body of literature about their utility in patients with sepsis and septic shock.

Leading scientists in the field, like Ince and co-workers, have demonstrated that intravital microscopy is able to describe the status of microcirculation both qualitatively and quantitatively during critical illness such as sepsis, and that it can effectively predict organ dysfunction and mortality (117, 118). Moreover, intravital imaging can provide a direct evaluation of tissue perfusion before and after therapeutic interventions, in adult and neonates with septic shock. In this context, it’s important to note

that not all alterations of microvascular circulation are due to endothelial damage: indeed, significant contributions can come from immunothrombosis, reduced red blood cells deformability and altered vascular tone (119, 120). Thus, some authors have tried to specifically identify features of endothelial dysfunction and activation by means of intravital microscopy (121). Nieuendorp and coll. had previously shown that it is possible to derive a measurement of glycocalyx thickness *in vivo* by measuring the distance between flowing RBCs and the endothelial wall (122), and in 2018, Uz and coll. validated a method to quantify leukocytes adhesion to endothelium and rolling using space-time diagrams derived from intravital microscopy (123).

The abovementioned technologies have been applied to neonates (124–126), and it is anticipated that they may be particularly useful in the context of neonatal sepsis and septic shock.

Neonates, especially preterm, represent an optimal setting for intravital microscopy, due to the reduced thickness of skin and mucosae. Recently, it has been shown that buccal mucosa may be the best site for evaluation of microvascular vessel density in term neonates (127), while transcutaneous evaluation seemed feasible and reproducible in healthy preterm infants (128, 129). It has also been demonstrated that persistent alterations of microcirculation, as assessed by intravital microscopy, are diagnostic of septic shock in children, with an area-under-curve (AUC) of 0.956 (95% confidence interval 0.853–1.058), and prognostic for survival (117). In preterm infants, daily acquired images of cutaneous microcirculation were useful to prove that functional small vessel density is reduced 1 day before the appearance of clinical symptoms in culture-confirmed or suspected late-onset sepsis (130). Similar results were obtained on term neonates (131), demonstrating that the proportion of microcirculation vessels with continuous flow is significantly lower in infants with suspected infection compared to controls (69 vs. 90%, p -value = 0.0003). While most of these results are preliminary and obtained on small cohorts of children and neonates, their correlation with real-time endothelial function, e.g., levels of soluble markers, is not known: further studies on the diagnostic and prognostic utility of intravital microscopy in the context of neonatal sepsis, as well as its correlation with endothelial dysfunction, are needed.

Biomarkers of Glycocalyx and Endothelium Damage

The second way to study endothelial function during sepsis, in neonates as in adults, is the quantification of soluble components of endothelial wall and glycocalyx in plasma or urine (132). A robust body of literature, including excellent reviews, exists about the use of endothelial soluble biomarkers to evaluate the course of sepsis in adult patients (133, 134). Five categories of biomarkers are currently evaluated and proven useful in adult sepsis:

- a) Proteases (e.g., MMPs);
- b) Adhesion molecules (e.g., VCAM 1-2, ICAM-1-, E-selectin);
- c) Coagulation factors (e.g., PAI-1, ADAMTS13, Von Willebrand factor or Tissue Factor);

- d) Glycocalyx end-products (e.g., syndecan, hyaluronan, endocan) and cleaving enzymes (heparanase);
- e) Endothelial growth factors and receptors (e.g., angiopoietin (Ang) –1 and –2 with their receptors Tie-1 and –2, or VEGF-1 with its cell-anchored or circulating decoy receptor Flt-1 and sFlt-1).

In adult patients, several of these biomarkers have been correlated with endothelial damage/activation or with glycocalyx degradation (frequently inter-correlated), and have shown good association with severity scores, such as SOFA, and adverse outcomes such as mortality (135, 136). This occurs in particular when sustained elevation over time of multiple analytes is present (137). Nevertheless, not a single biomarker has been proven to be the “gold standard” for diagnosis or prognostic stratification of adult septic patients.

In neonates, most published studies on soluble biomarkers of endothelial damage during sepsis have focused on E-selectin, sICAM-1 and sVCAM-1, aiming to demonstrate their early diagnostic value (138). Results have been neither consistent between studies nor straightforward to be interpreted: first, reference levels for these biomarkers in early life are not established and can be variable. For example, three different studies (96, 139, 140) have shown that sICAM-1 plasma levels tend to increase over the first month of life even in healthy neonates, thus making it difficult to establish reference ranges for sepsis. Second, enrollment criteria varied greatly, frequently including both preterm and term neonates as well as EOS and LOS. Third, definitions of sepsis diverge and are not always directly comparable. Considering these limitations, most authors have concluded that combinations of different biomarkers of endothelial damage, associated with markers of systemic inflammatory response such as C-reactive protein (CRP) or IL-6, may improved sensitivity and specificity for early diagnosis of neonatal sepsis, as compared to single measurements (141–144). Following the results obtained on adult patients, some groups extended their research to more novel endothelial biomarkers during neonatal sepsis, mainly components of degraded glycocalyx, endothelial growth factors or components of tight junctions (TJs) that shed into circulation upon endothelial damage. It has been shown that sTREM-1 (soluble triggering receptor expressed on myeloid cells-1) has a AUC of 0.97 for the diagnosis of suspected or proven neonatal sepsis, compared to 0.8 of Endocan and 0.96 of IL-6 (145). Furthermore, the correlation between plasma levels of Ang-1, Ang-2, sICAM-1, and sVCAM-1 and the occurrence of death or bacteremia in term infants admitted for suspected sepsis has been investigated. Ang-2 was shown to be higher at admission in non-survivors, as well as Ang-2: Ang-1 ratio that also correlated with bacteremia (146). Similar results were obtained by other investigators (147).

The presence of TJs components in the blood as a marker of endothelial damage has been investigated in several diseases in adult as well as in pediatric patients, including pediatric sepsis and neonatal necrotizing enterocolitis (148). Of note, no study on neonatal sepsis has been published yet. Recently, it has been demonstrated that plasma levels of occludin and

zonula occludens (ZO)-1 are correlated with both APACHE II score and SOFA score in septic adult patients, and that they are useful to identify patients at risk for MOF (149). This study highlights the interconnection between systemic inflammation, endothelial damage and organ damage during severe sepsis. The same investigational concept may be applied in the context of neonatal sepsis, especially in those populations, like ELBWI, at higher risk for severe course of sepsis, shock, MOF and death.

Overall, studies conducted on septic neonates compared to those on adults show important limitations that restrict their current utility but provide space for new research. For example, the kinetics of several biomarkers over the course of sepsis is not known for neonates, and the practical difficulty to obtain serial blood samples in small patients complicates such studies. Furthermore, most neonatal studies, with few exceptions, are focused on the diagnostic value of biomarkers, without prognostic correlation with clinical outcomes. A neonatal adaptation of the SOFA score named “nSOFA” has been recently proposed (150). The correlation between biomarkers of endothelial dysfunction, a previously validated nSOFA score and long-term outcomes of sepsis may improve the individualization of therapy, and may help to clarify the molecular mechanisms that increase the rates of long-term adverse outcomes in septic neonates.

THERAPEUTIC OPPORTUNITIES

Besides antibiotics, fluid resuscitation and cardiovascular support, we currently lack any other effective therapy to counteract an invading pathogen and the self-induced damage caused by an uncontrolled systemic inflammatory response in septic neonates. A number of potentially immunomodulatory drugs and strategies have been previously evaluated in adults as well as in neonates, or are currently under investigation (13, 19).

Aside from pure immunotherapy, an increasing attention in adult patients and in experimental models of sepsis is being given to pharmacological interventions targeted to improve the endothelial stability, or to act on the interplay between endothelial barrier, glycocalyx, and immune cells (151–153) (Figure 3). As mentioned previously, common features of severe sepsis, both in neonates and adults, include reduced systemic vascular resistance (SVR), interstitial edema, hypoalbuminemia, hypotension, and MOF (19, 92). Endothelial damage has been clearly implicated as contributing to each of these phenomena in adult patients, and to a lesser extent in neonates. These observations provide a rationale to investigate the effect of several therapeutic agents, already in use or under investigation, on the endothelial barrier of neonates in the context of neonatal sepsis.

Current Therapies That Affect Glycocalyx Integrity

Some of the therapeutic agents already in use to treat sepsis may impact the glycocalyx (152, 154). Hydrocortisone is used in the context of neonatal refractory septic shock, to improve the efficacy of vasomotor amines and counteract a possible relative adrenal insufficiency, especially in very preterm neonates (67).

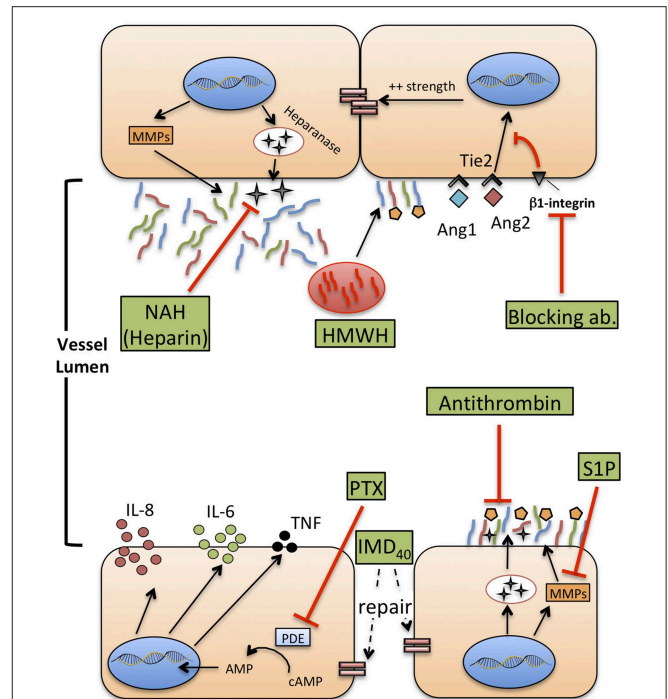


FIGURE 3 | Therapeutic agents that potentially improve or restore endothelial function during neonatal sepsis. MMPs: metalloproteases. HMWH: high molecular weight-hyaluronan. NAH, N-desulfated/re-N-acetylated heparin; S1P, sphingosine-1-phosphate (S1P); Ang1-2, Angiotensin 1-2; Tie2, TEK receptor tyrosine kinase; PTX, pentoxifylline; cAMP, cyclic-adenosine monophosphate; AMP, adenosine monophosphate; IMD40, intermedin peptide 40 kD.

In experimental models of ischemia/reperfusion, intravenous hydrocortisone reduced the shedding of syndecan-1, heparan sulfate, and hyaluronan, limiting the formation of extravascular edema (155). We are not aware of any published studies regarding the impact of steroids on the glycocalyx structure and function during sepsis, in adult or newborn patients. Another example of therapeutic intervention for sepsis that may affect glycocalyx is volume expansion with intravenous fluids, either crystalloids or colloids (in most septic newborns represented by albumin). This is frequently the first intervention to correct hypotension in presumed septic neonates, but some studies suggest that excess intravascular volume increases the shedding of glycocalyx, possibly through increased shear stress or the effect of atrial natriuretic peptide (ANP) (156, 157). Furthermore, if the glycocalyx is disrupted, a high proportion of the infused fluid flows toward the extracellular space, favoring the formation of edema and prompting the infusion of supplementary fluids in the attempt to maintain intravascular volume.

Interestingly, albumin seems to be a double edge sword in this context: as a part of the protein-glycan network forming the glycocalyx, its infusion may help restoring the integrity of a damaged glycocalyx, an effect not shared by any other colloid, such as synthetic ones (158). Albumin also carries sphingosine-1-phosphate (S1P) to the endothelial surface, and it is known

that S1P can favor glycocalyx recovery suppressing MMP activity (159). Conversely, when the damage to glycocalyx is too severe, even albumin flows out to the extracellular space, increasing the extravascular oncotic pressure and driving a further net flow of water out of the vascular lumen (160).

Hypoalbuminemia and interstitial edema are relatively common in neonates with septic shock. Nonetheless, no clinical or experimental study investigated the correlation between albumin administration, albumin extravasation and glycocalyx degradation in neonatal sepsis: more research is needed to understand the effect of current therapies on endothelial wall and glycocalyx in the context of neonatal sepsis.

S1P, Heparanase Inhibition and Hyaluronan

As mentioned previously, S1P is a natural inhibitor of syndecan-1 shedding through the inhibition of MMPs. In experimental models of sepsis, administration of fingolimod, a S1P receptor modulator, improved left ventricular systolic contractility (161). This effect may be due to a reduced shedding of glycocalyx components, similarly to what has been shown by other studies (90). No comparable evidence exists for neonatal sepsis yet.

Heparanase is a glucuronidase responsible for degradation of the glycocalyx network, and possibly mediating renal and lung injury during sepsis, as demonstrated in animal models (162). Heparin can inhibit heparanase activity and protect from shedding of glycocalyx components, even during inflammatory states such as sepsis (163). However, heparin infusion is frequently impractical in septic neonates due to its potent anticoagulant activity and high risk for hemorrhage. N-desulfated re-N-acetylated heparin (NAH), a heparin analog devoid of anticoagulant properties, prevented glycocalyx loss and neutrophil adhesion to ECs caused by endotoxemia in an animal model of sepsis, thereby reducing renal and pulmonary injury (164). This effect was confirmed also when NAH was administered hours after the onset of experimental sepsis, with a time-course that reproduces real-life intervention in septic patients, including neonates. Finally, the anti-inflammatory

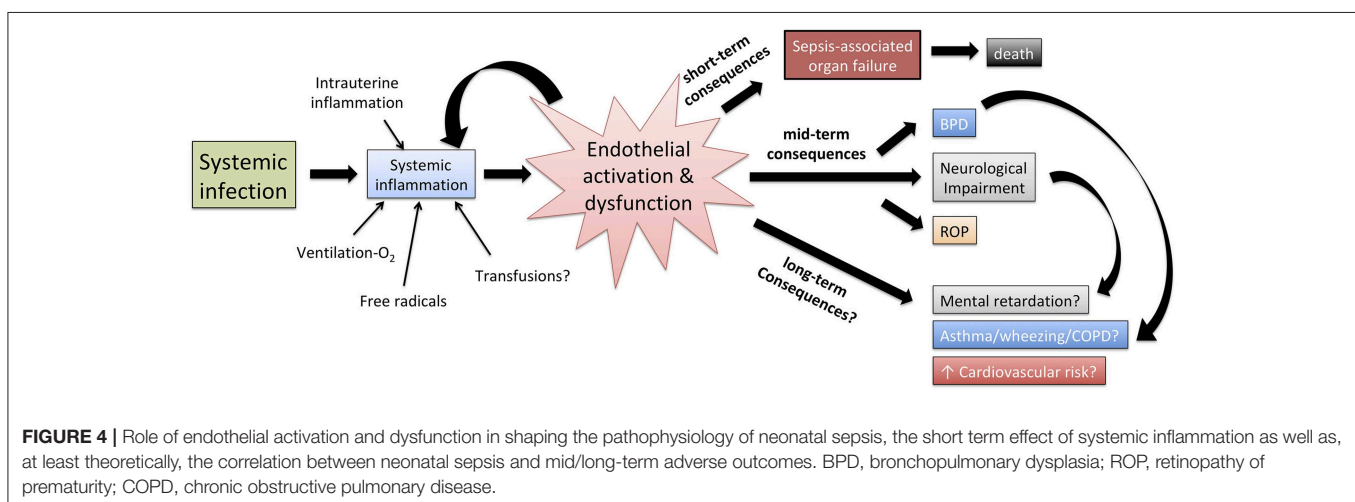
effect of high-molecular-weight hyaluronan (HMWH), a high-molecular-mass polysaccharide and component of glycocalyx, has been studied in the context of sepsis-induced lung injury (165). HMWH prevented monocyte and neutrophil infiltration into the lungs and the expression of macrophage inflammatory protein-2 mRNA.

Overall, these preliminary studies suggest that restoration of glycocalyx integrity during sepsis may represent a novel therapeutic strategy that deserves a translation into the neonatal context. S1P, NAH and hyaluronan, may represent the most promising agents worth of age-specific investigation.

The Ang1/2–Tie2 Axis

Angiopoietins are vascular growth factors that promote neo-angiogenesis interacting with their tyrosine kinase receptor, Tie2, expressed on ECs. Angiopoietin 1 and angiopoietin 2 have opposing effects on Tie2: Ang1 promotes endothelial stability and cell survival, strengthens adherens junctions through the inhibition of NF- κ B and downregulates the expression of ICAM-1 and VCAM-1, inhibiting leukocyte adhesion. Conversely, Ang2 is released during inflammatory states or hypoxia, in the “attempt” to restore a steady state environment. It promotes disruption of intercellular junctions that favor neoangiogenesis, increases the surface expression of ICAM-1 and VCAM-1, and increase paracellular permeability to fluids and proteins (166, 167).

Over the past 10 years, the Ang1/2–Tie2 axis has been targeted by new experimental therapies aiming to reduce endothelial damage during sepsis or other models of systemic inflammation. Recombinant Ang1 can increase survival and reduce the development of lung edema during systemic inflammation induced by LPS, in a murine model (168). Recently, it was also demonstrated that clustering of Ang2, induced by a monoclonal antibody, induces a more Ang-1-like response of Tie2, tightening VE-cadherin, reducing heparanase synthesis and reducing the expression of adhesion molecules by ECs (169). Finally, in a murine



model of sepsis, the block of a secondary receptor of Ang2, β -1 integrin, with a monoclonal antibody enhanced stabilization of junctions between ECs, reduced vascular leak and contraction of ECs induced by LPS and reduced mortality (170). As previously mentioned, at least two studies reported high level of Ang2 and Ang2:Ang1 ratio in septic neonates (146, 171). Overall, these studies suggest a possible benefit of Tie2 regulation in neonatal sepsis and prompt a translational approach to early life. Furthermore, Ang1 in newborns is a critical mediator of hyperoxia-induced lung injury (172) and retinopathy of prematurity (173). Accordingly, strategies directed at investigating and therapeutically regulating Ang2-Tie2 axis may have benefits in early life beyond the improvement of sepsis outcomes, and could potentially reduce neonatal sepsis-associated chronic complications.

Pentoxifylline

Pentoxifylline (PTX), a phosphodiesterase inhibitor that enhances cellular cAMP concentrations, reduces TLR-mediated and inflammasome-mediated cytokine production by neonatal immune cells, *in vitro* (174, 175) and also *in vivo* (176). Furthermore, PTX has been shown to reduce mortality of septic neonates in small-size studies, and is currently under investigation in a large trial that may provide a definitive answer on its clinical efficacy (177). Aside from the effects on cytokine production (which may anyhow involve ECs), PTX ameliorates endothelial function in different experimental models of critical illness, specifically of sepsis. For example, PTX inhibits adhesion and rolling of leukocytes on ECs (178), supports the preservation of a physiological endothelium-mediated vasodilation (179), and reduces the expression of P-selectin and ICAM-1 on mesenteric microvessels (180). Furthermore, PTX can reduce the activity of wingless integrated MMTV (Wnt)/ β -catenin pathway (181), involved in the conservation of endothelial intercellular junctions. These studies provide a rationale to further explore the action of PTX on VE during neonatal sepsis, both *in vitro*, to gain knowledge about the molecular mechanisms regulating its activity, and *in vivo*, monitoring the endothelial status and function (e.g., the quantification of endothelial biomarkers) during therapy with PTX.

Antithrombin and Intermedin

Very recently, the beneficial effect of two additional molecules on endothelial integrity has been demonstrated in experimental models of sepsis. Antithrombin infusion reduced circulating levels of syndecan-1, hyaluronan and lactate in a rat model of sepsis (182). Furthermore, antithrombin inhibited leukocyte adhesion and attenuated the reduction of blood flow through capillaries due to immunothrombosis, highlighting once more how coagulation and endothelial function are deeply interlaced in the context of sepsis. Intermedin (IMD), a member of the calcitonin family, and its peptide IMD_{40} , alleviate the increase of vascular

leakage and protect the endothelium against the effect of cytokine storm during experimental sepsis (183). The administration of IMD peptide improved the survival of septic mice, re-establishing the endothelial barrier and reducing inflammatory responses.

CONCLUSIONS

VE has emerged as a fundamental sepsis mediator interposed between the effect of the invading pathogens, the systemic immune response of the host and the multi-organ damage that ultimately leads to death. Furthermore, several long-term consequences typical of prematurity, such as bronchopulmonary dysplasia, retinopathy of prematurity and neurodevelopmental impairment, have increased incidence in previously septic neonates compared to those who never experienced sepsis. The VE may mediate some of these long-term consequences (Figure 4), but evidence supporting such a theory is limited. Finally, an improved comprehension of leukocyte-endothelium interactions in early life may enhance *in vitro* modeling of the action of vaccines and immunomodulators, to improve the efficacy of preventative strategies such as neonatal immunization (78).

Despite the distinct nature of neonatal sepsis and its heavy toll, basic and translational research on endothelial function at early stages of life, in health and disease, lags substantially behind its adult counterpart. Age-specific investigation of endothelial structure, of the interactions between ECs, glycocalyx, pathogens and immune cells, and of the effect of endothelial damage on the course of neonatal sepsis, may define a key link between sepsis and multi-organ damage. Cooperation between scientists and neonatologists is fundamental to deepen the investigation of molecular pathways and immunological mechanisms regulating endothelial dysfunction during neonatal sepsis, and inform new approaches to endothelium-targeted therapies that may significantly improve short- and long-term outcomes.

AUTHOR CONTRIBUTIONS

CP, LP, AR, and FM conceived and designed the review. CP wrote the first draft of the manuscript. CP, LP, AR, IB, BG, GS-S, FB, FM, and OL contributed to manuscript critical revision, read, and approved the submitted version.

FUNDING

The authors declare that the research was conducted without any specific funding.

ACKNOWLEDGMENTS

The authors would like to thank all the nurses and neonatologists of the Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico.

REFERENCES

- Levy O. Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol.* (2007) 7:379–90. doi: 10.1038/nri2075
- Galindo-Albarran AO, Lopez-Portales OH, Gutierrez-Reyna DY, Rodriguez-Jorge O, Sanchez-Villanueva JA, Ramirez-Pliego O, et al. CD8(+) T cells from human neonates are biased toward an innate immune response. *Cell Rep.* (2016) 17:2151–60. doi: 10.1016/j.celrep.2016.10.056
- Siefker DT, Adkins B. Rapid CD8(+) function is critical for protection of neonatal mice from an extracellular bacterial enteropathogen. *Front Pediatr.* (2016) 4:141. doi: 10.3389/fped.2016.00141
- Kollmann TR, Kampmann B, Mazmanian SK, Marchant A, Levy O. Protecting the newborn and young infant from infectious diseases: lessons from immune ontogeny. *Immunity.* (2017) 46:350–63. doi: 10.1016/j.immuni.2017.03.009
- Philbin VJ, Dowling DJ, Gallington LC, Cortés G, Tan Z, Suter EE, et al. Imidazoquinoline Toll-like receptor 8 agonists activate human newborn monocytes and dendritic cells through adenosine-refractory and caspase-1-dependent pathways. *J Allergy Clin Immunol.* (2012) 130:195–204.e9. doi: 10.1016/j.jaci.2012.02.042
- Sanchez-Schmitz G, Levy O. Development of newborn and infant vaccines. *Sci Transl Med.* (2011) 3:90ps27. doi: 10.1126/scitranslmed.3001880
- Kollmann TR, Levy O, Montgomery RR, Goriely S. Innate immune function by Toll-like receptors: distinct responses in newborns and the elderly. *Immunity.* (2012) 37:771–83. doi: 10.1016/j.immuni.2012.10.014
- Pettengill MA, van Haren SD, Levy O. Soluble mediators regulating immunity in early life. *Front Immunol.* (2014) 5:457. doi: 10.3389/fimmu.2014.00457
- Levy O, Jean-Jacques RM, Cywes C, Sisson RB, Zarembka KA, Godowski PJ, et al. Critical role of the complement system in group B streptococcus-induced tumor necrosis factor alpha release. *Infect Immun.* (2003) 71:6344–53. doi: 10.1128/IAI.71.11.6344-6353.2003
- Dowling DJ, Scott EA, Scheid A, Bergelson I, Joshi S, Pietrasanta C, et al. Toll-like receptor 8 agonist nanoparticles mimic immunomodulating effects of the live BCG vaccine and enhance neonatal innate and adaptive immune responses. *J Allergy Clin Immunol.* (2017) 140:1339–50. doi: 10.1016/j.jaci.2016.12.985
- Zhao J, Kim KD, Yang X, Auh S, Fu YX, Tang H. Hyper innate responses in neonates lead to increased morbidity and mortality after infection. *Proc Natl Acad Sci USA.* (2008) 105:7528–33. doi: 10.1073/pnas.0800152105
- Raymond SL, Stortz JA, Mira JC, Larson SD, Wynn JL, Moldawer LL. Immunological defects in neonatal sepsis and potential therapeutic approaches. *Front Pediatr.* (2017) 5:14. doi: 10.3389/fped.2017.00014
- Schuller SS, Kramer BW, Villamor E, Spittler A, Berger A, Levy O. Immunomodulation to prevent or treat neonatal sepsis: past, present, and future. *Front Pediatr.* (2018) 6:199. doi: 10.3389/fped.2018.00199
- Opal SM, van der Poll T. Endothelial barrier dysfunction in septic shock. *J Intern Med.* (2015) 277:277–93. doi: 10.1111/joim.12331
- Ince C, Mayeux PR, Nguyen T, Gomez H, Kellum JA, Ospina-Tascon GA, et al. The endothelium in sepsis. *Shock.* (2016) 45:259–70. doi: 10.1097/SHK.0000000000000473
- Claesson-Welsh L. Vascular permeability—the essentials. *Ups J Med Sci.* (2015) 120:135–43. doi: 10.3109/03009734.2015.1064501
- Schouten M, Wiersinga WJ, Levi M, van der Poll T. Inflammation, endothelium, and coagulation in sepsis. *J Leukoc Biol.* (2008) 83:536–45. doi: 10.1189/jlb.0607373
- Crouser ED, Matthay MA. Endothelial damage during septic shock: significance and implications for future therapies. *Chest.* (2017) 152:1–3. doi: 10.1016/j.chest.2017.02.016
- Hotchkiss RS, Moldawer LL, Opal SM, Reinhart K, Turnbull IR, Vincent JL. Sepsis and septic shock. *Nat Rev Dis Primers.* (2016) 2:16045. doi: 10.1038/nrdp.2016.45
- Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, et al. The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA.* (2016) 315:801–10. doi: 10.1001/jama.2016.0287
- Wynn JL. Defining neonatal sepsis. *Curr Opin Pediatr.* (2016) 28:135–40. doi: 10.1097/MOP.0000000000000315
- Wynn JL, Wong HR, Shanley TP, Bizzarro MJ, Saiman L, Polin RA. Time for a neonatal-specific consensus definition for sepsis. *Pediatr Crit Care Med.* (2014) 15:523–8. doi: 10.1097/PCC.0000000000000157
- Simonsen KA, Anderson-Berry AL, Delair SF, Davies HD. Early-onset neonatal sepsis. *Clin Microbiol Rev.* (2014) 27:21–47. doi: 10.1128/CMR.00031-13
- Schrag SJ, Farley MM, Petit S, Reingold A, Weston EJ, Pondo T, et al. Epidemiology of invasive early-onset neonatal sepsis, 2005 to 2014. *Pediatrics.* (2016) 138:2013. doi: 10.1542/peds.2016-2013
- Greenberg RG, Kandefer S, Do BT, Smith PB, Stoll BJ, Bell EF, et al. Late-onset sepsis in extremely premature infants: 2000–2011. *Pediatr Infect Dis J.* (2017) 36:774–9. doi: 10.1097/INF.0000000000001570
- Shane AL, Sanchez PJ, Stoll BJ. Neonatal sepsis. *Lancet.* (2017) 390:1770–80. doi: 10.1016/S0140-6736(17)31002-4
- Stoll BJ, Hansen NI, Sanchez PJ, Faix RG, Poindexter BB, Van Meurs KP, et al. Early onset neonatal sepsis: the burden of group B *Streptococcus* and *E. coli* disease continues. *Pediatrics.* (2011) 127:817–26. doi: 10.1542/peds.2010-2217
- Khaertynov KS, Boichuk SV, Khaiboullina SF, Anokhin VA, Andreeva AA, Lombardi VC, et al. Comparative assessment of cytokine pattern in early and late onset of neonatal sepsis. *J Immunol Res.* (2017) 2017:8601063. doi: 10.1155/2017/8601063
- Schultz C, Rott C, Temming P, Schlenke P, Moller JC, Bucsky P. Enhanced interleukin-6 and interleukin-8 synthesis in term and preterm infants. *Pediatr Res.* (2002) 51:317–22. doi: 10.1203/00006450-200203000-00009
- Delano MJ, Ward PA. The immune system's role in sepsis progression, resolution, and long-term outcome. *Immunol Rev.* (2016) 274:330–53. doi: 10.1111/imr.12499
- Hotchkiss RS, Monneret G, Payen D. Immunosuppression in sepsis: a novel understanding of the disorder and a new therapeutic approach. *Lancet Infect Dis.* (2013) 13:260–8. doi: 10.1016/S1473-3099(13)70001-X
- Mira JC, Brakenridge SC, Moldawer LL, Moore FA. Persistent inflammation, immunosuppression and catabolism syndrome. *Crit Care Clin.* (2017) 33:245–58. doi: 10.1016/j.ccc.2016.12.001
- Hibbert JE, Currie A, Strunk T. Sepsis-induced immunosuppression in neonates. *Front Pediatr.* (2018) 6:357. doi: 10.3389/fped.2018.00357
- Andrade EB, Alves J, Madureira P, Oliveira L, Ribeiro A, Cordeiro-da-Silva A, et al. TLR2-induced IL-10 production impairs neutrophil recruitment to infected tissues during neonatal bacterial sepsis. *J Immunol.* (2013) 191:4759–68. doi: 10.4049/jimmunol.1301752
- Gentile LF, Nacionales DC, Lopez MC, Vanzant E, Cuenca A, Cuenca AG, et al. Protective immunity and defects in the neonatal and elderly immune response to sepsis. *J Immunol.* (2014) 192:3156–65. doi: 10.4049/jimmunol.1301726
- Upham JW, Rate A, Rowe J, Kusel M, Sly PD, Holt PG. Dendritic cell immaturity during infancy restricts the capacity to express vaccine-specific T-cell memory. *Infect Immun.* (2006) 74:1106–12. doi: 10.1128/IAI.74.2.1106-1112.2006
- Filias A, Theodorou GL, Mouzopoulou S, Varvarigou AA, Mantagos S, Karakantza M. Phagocytic ability of neutrophils and monocytes in neonates. *BMC Pediatr.* (2011) 11:29. doi: 10.1186/1471-2431-11-29
- Melvan JN, Bagby GJ, Welsh DA, Nelson S, Zhang P. Neonatal sepsis and neutrophil insufficiencies. *Int Rev Immunol.* (2010) 29:315–48. doi: 10.3109/08830181003792803
- Yost CC, Cody MJ, Harris ES, Thornton NL, McInturf AM, Martinez ML, et al. Impaired neutrophil extracellular trap (NET) formation: a novel innate immune deficiency of human neonates. *Blood.* (2009) 113:6419–27. doi: 10.1182/blood-2008-07-171629
- Falconer AE, Carr R, Edwards SW. Impaired neutrophil phagocytosis in preterm neonates: lack of correlation with expression of immunoglobulin or complement receptors. *Biol Neonate.* (1995) 68:264–9. doi: 10.1159/000244245
- Kallman J, Schollin J, Schalen C, Erlandsson A, Kihlstrom E. Impaired phagocytosis and opsonisation towards group B streptococci in preterm neonates. *Arch Dis Child Fetal Neonatal Ed.* (1998) 78:F46–50. doi: 10.1136/fin.78.1.F46
- Nussbaum C, Sperandio M. Innate immune cell recruitment in the fetus and neonate. *J Reprod Immunol.* (2011) 90:74–81. doi: 10.1016/j.jri.2011.01.022

43. Melville JM, Moss TJ. The immune consequences of preterm birth. *Front Neurosci.* (2013) 7:79. doi: 10.3389/fnins.2013.00079
44. Azizia M, Lloyd J, Allen M, Klein N, Peebles D. Immune status in very preterm neonates. *Pediatrics.* (2012) 129:e967–74. doi: 10.1542/peds.2011-1579
45. Yagupsky P, Nolte FS. Quantitative aspects of septicemia. *Clin Microbiol Rev.* (1990) 3:269–79. doi: 10.1128/CMR.3.3.269
46. Harbeson D, Ben-Othman R, Amenyoobe N, Kollmann TR. Outgrowing the immaturity myth: the cost of defending from neonatal infectious disease. *Front Immunol.* (2018) 9:1077. doi: 10.3389/fimmu.2018.01077
47. Zhang Q, Coveney AP, Yu S, Liu JH, Li Y, Blankson S, et al. Inefficient antimicrobial functions of innate phagocytes render infant mice more susceptible to bacterial infection. *Eur J Immunol.* (2013) 43:1322–32. doi: 10.1002/eji.201243077
48. Harbeson D, Francis F, Bao W, Amenyoobe NA, Kollmann TR. Energy demands of early life drive a disease tolerant phenotype and dictate outcome in neonatal bacterial sepsis. *Front Immunol.* (2018) 9:1918. doi: 10.3389/fimmu.2018.01918
49. Burt TD. Fetal regulatory T cells and peripheral immune tolerance *in utero*: implications for development and disease. *Am J Reprod Immunol.* (2013) 69:346–58. doi: 10.1111/aji.12083
50. Levy O, Coughlin M, Cronstein BN, Roy RM, Desai A, Wessels MR. The adenosine system selectively inhibits TLR-mediated TNF- α production in the human newborn. *J Immunol.* (2006) 177:1956–66. doi: 10.4049/jimmunol.177.3.1956
51. De Kleer I, Willems F, Lambrecht B, Goriely S. Ontogeny of myeloid cells. *Front Immunol.* (2014) 5:423. doi: 10.3389/fimmu.2014.00423
52. Renneson J, Dutta B, Goriely S, Danis B, Lecomte S, Laes JF, et al. IL-12 and type I IFN response of neonatal myeloid DC to human CMV infection. *Eur J Immunol.* (2009) 39:2789–99. doi: 10.1002/eji.200939414
53. Danis B, George TC, Goriely S, Dutta B, Renneson J, Gatto L, et al. Interferon regulatory factor 7-mediated responses are defective in cord blood plasmacytoid dendritic cells. *Eur J Immunol.* (2008) 38:507–17. doi: 10.1002/eji.200737760
54. Goriely S, Van Lint C, Dadkhah R, Libin M, De Wit D, Demonte D, et al. A defect in nucleosome remodeling prevents IL-12(p35) gene transcription in neonatal dendritic cells. *J Exp Med.* (2004) 199:1011–6. doi: 10.1084/jem.20031272
55. De Wit D, Tonon S, Orlislagers V, Goriely S, Boutriaux M, Goldman M, et al. Impaired responses to toll-like receptor 4 and toll-like receptor 3 ligands in human cord blood. *J Autoimmun.* (2003) 21:277–81. doi: 10.1016/j.jaut.2003.08.003
56. Goriely S, Vincart B, Stordeur P, Vekemans J, Willems F, Goldman M, et al. Deficient IL-12(p35) gene expression by dendritic cells derived from neonatal monocytes. *J Immunol.* (2001) 166:2141–6. doi: 10.4049/jimmunol.166.3.2141
57. Dowling DJ, Levy O. Ontogeny of early life immunity. *Trends Immunol.* (2014) 35:299–310. doi: 10.1016/j.it.2014.04.007
58. van Haren SD, Ganapathi L, Bergelson I, Dowling DJ, Banks M, Samuels RC, et al. *In vitro* cytokine induction by TLR-activating vaccine adjuvants in human blood varies by age and adjuvant. *Cytokine.* (2016) 83:99–109. doi: 10.1016/j.cyto.2016.04.001
59. Wynn JL, Scumpia PO, Delano MJ, O'Malley KA, Ungaro R, Abouhamze A, et al. Increased mortality and altered immunity in neonatal sepsis produced by generalized peritonitis. *Shock.* (2007) 28:675–83. doi: 10.1097/SHK.0b013e3180556d09
60. Wynn JL, Wilson CS, Hawiger J, Scumpia PO, Marshall AF, Liu JH, et al. Targeting IL-17A attenuates neonatal sepsis mortality induced by IL-18. *Proc Natl Acad Sci USA.* (2016) 113:E2627–35. doi: 10.1073/pnas.1515793113
61. Grobmyer SR, Lin E, Lowry SF, Rivadeneira DE, Potter S, Barie PS, et al. Elevation of IL-18 in human sepsis. *J Clin Immunol.* (2000) 20:212–5. doi: 10.1023/A:1006641630904
62. Sugimoto MA, Sousa LP, Pinho V, Perretti M, Teixeira MM. Resolution of inflammation: what controls its onset? *Front Immunol.* (2016) 7:160. doi: 10.3389/fimmu.2016.00160
63. Raymond SL, Lopez MC, Baker HV, Larson SD, Efron PA, Sweeney TE, et al. Unique transcriptomic response to sepsis is observed among patients of different age groups. *PLoS ONE.* (2017) 12:e0184159. doi: 10.1371/journal.pone.0184159
64. Chen XF, Wu J, Zhang YD, Zhang CX, Chen XT, Sun JH, et al. Role of Zc3h12a in enhanced IL-6 production by newborn mononuclear cells in response to lipopolysaccharide. *Pediatr Neonatol.* (2018) 59:288–95. doi: 10.1016/j.pedneo.2017.09.006
65. Hynninen M, Pettila V, Takkunen O, Orko R, Jansson SE, Kuusela P, et al. Predictive value of monocyte histocompatibility leukocyte antigen-DR expression and plasma interleukin-4 and–10 levels in critically ill patients with sepsis. *Shock.* (2003) 20:1–4. doi: 10.1097/01.shk.0000068322.08268.b4
66. Prashant A, Vishwanath P, Kulkarni P, Sathya Narayana P, Gowdara V, Nataraj SM, et al. Comparative assessment of cytokines and other inflammatory markers for the early diagnosis of neonatal sepsis—a case control study. *PLoS ONE.* (2013) 8:e68426. doi: 10.1371/journal.pone.0068426
67. Wynn JL, Wong HR. Pathophysiology and treatment of septic shock in neonates. *Clin Perinatol.* (2010) 37:439–79. doi: 10.1016/j.clp.2010.04.002
68. Pober JS, Sessa WC. Evolving functions of endothelial cells in inflammation. *Nat Rev Immunol.* (2007) 7:803–15. doi: 10.1038/nri2171
69. Pober JS, Sessa WC. Inflammation and the blood microvascular system. *Cold Spring Harb Perspect Biol.* (2014) 7:a016345. doi: 10.1101/cshperspect.a016345
70. Dejana E, Orsenigo F, Lampugnani MG. The role of adherens junctions and VE-cadherin in the control of vascular permeability. *J Cell Sci.* (2008) 121:2115–22. doi: 10.1242/jcs.017897
71. Granger DN, Senchenkova E. *Inflammation and the Microcirculation. Integrated Systems Physiology—From Cell to Function.* San Rafael, CA: Morgan & Claypool Publishers (2010). doi: 10.4199/C00013ED1V01Y201006ISP008
72. Lorant DE, Li W, Tabatabaei N, Garver MK, Albertine KH. P-selectin expression by endothelial cells is decreased in neonatal rats and human premature infants. *Blood.* (1999) 94:600–9.
73. Wisgrill L, Muck M, Wessely I, Berger A, Spittler A, Forster-Waldl E, et al. Endothelial cells of extremely premature infants display impaired immune response after proinflammatory stimulation. *Pediatr Res.* (2018) 83:128–34. doi: 10.1038/pr.2017.202
74. Nussbaum C, Gloning A, Pruenster M, Frommhold D, Bierschenk S, Genzel-Boroviczeny O, et al. Neutrophil and endothelial adhesive function during human fetal ontogeny. *J Leukoc Biol.* (2013) 93:175–84. doi: 10.1189/jlbb.0912468
75. Auvinen K, Jalkanen S, Salmi M. Expression and function of endothelial selectins during human development. *Immunology.* (2014) 143:406–15. doi: 10.1111/imm.12318
76. Patel H, Chen J, Das KC, Kavdia M. Hyperglycemia induces differential change in oxidative stress at gene expression and functional levels in HUVEC and HMVEC. *Cardiovasc Diabetol.* (2013) 12:142. doi: 10.1186/1475-2840-12-142
77. Huet O, Dupic L, Harrois A, Duranteau J. Oxidative stress and endothelial dysfunction during sepsis. *Front Biosci.* (2011) 16:1986–95. doi: 10.2741/3835
78. Sanchez-Schmitz G, Stevens CR, Bettencourt IA, Flynn PJ, Schmitz-Abe K, Metser G, et al. Microphysiologic human tissue constructs reproduce autologous age-specific BCG and HBV primary immunization *in vitro*. *Front Immunol.* (2018) 9:2634. doi: 10.3389/fimmu.2018.02634
79. Alphonsus CS, Rodseth RN. The endothelial glycocalyx: a review of the vascular barrier. *Anaesthesia.* (2014) 69:777–84. doi: 10.1111/anae.12661
80. Schott U, Solomon C, Fries D, Bentzer P. The endothelial glycocalyx and its disruption, protection and regeneration: a narrative review. *Scand J Trauma Resusc Emerg Med.* (2016) 24:48. doi: 10.1186/s13049-016-0239-y
81. Ushiyama A, Kataoka H, Iijima T. Glycocalyx and its involvement in clinical pathophysiology. *J Intensive Care.* (2016) 4:59. doi: 10.1186/s40560-016-0182-z
82. Parving HH, Klebe JG, Ingomar CJ. Simultaneous determination of plasma volume and transcapillary escape rate with 131 I-labelled albumin and T-1824 in the newborn. *Acta Paediatr Scand.* (1973) 62:248–52. doi: 10.1111/j.1651-2227.1973.tb08100.x
83. Gold PS, Brace RA. Fetal whole-body permeability–surface area product and reflection coefficient for plasma proteins. *Microvasc Res.* (1988) 36:262–74. doi: 10.1016/0026-2862(88)90027-1

84. McElroy SJ, Weitkamp JH. Innate Immunity in the Small Intestine of the Preterm Infant. *Neoreviews*. (2011) 12:e517–e26. doi: 10.1542/neo.12-9-e517
85. Mai J, Virtue A, Shen J, Wang H, Yang XF. An evolving new paradigm: endothelial cells—conditional innate immune cells. *J Hematol Oncol*. (2013) 6:61. doi: 10.1186/1756-8722-6-61
86. Ramnath R, Foster RR, Qiu Y, Cope G, Butler MJ, Salmon AH, et al. Matrix metalloproteinase 9-mediated shedding of syndecan 4 in response to tumor necrosis factor alpha: a contributor to endothelial cell glycolocalyx dysfunction. *FASEB J*. (2014) 28:4686–99. doi: 10.1096/fj.14-252221
87. Yang C, Liu Z, Tian M, Xu P, Li B, Yang Q, et al. Relationship between serum albumin levels and infections in newborn late preterm infants. *Med Sci Monit*. (2016) 22:92–8. doi: 10.12659/MSM.895435
88. Morris I, Molloy EJ. Albumin administration in the management of neonatal hypalbuminaemia. *Arch Dis Child Fetal Neonatal Ed*. (2008) 93:F326. doi: 10.1136/adc.2007.131383
89. Shalish W, Olivier F, Aly H, Sant'Anna G. Uses and misuses of albumin during resuscitation and in the neonatal intensive care unit. *Semin Fetal Neonatal Med*. (2017) 22:328–35. doi: 10.1016/j.siny.2017.07.009
90. Martin L, Peters C, Schmitz S, Moellmann J, Martincuks A, Heussen N, et al. Soluble heparan sulfate in serum of septic shock patients induces mitochondrial dysfunction in murine cardiomyocytes. *Shock*. (2015) 44:569–77. doi: 10.1097/SHK.0000000000000462
91. Martin L, Schmitz S, De Santis R, Doemming S, Haase H, Hoeger J, et al. Peptide 19-2.5 inhibits heparan sulfate-triggered inflammation in murine cardiomyocytes stimulated with human sepsis serum. *PLoS ONE*. (2015) 10:e0127584. doi: 10.1371/journal.pone.0127584
92. Wynn J, Cornell TT, Wong HR, Shanley TP, Wheeler DS. The host response to sepsis and developmental impact. *Pediatrics*. (2010) 125:1031–41. doi: 10.1542/peds.2009-3301
93. Opitz B, Eitel J, Meixenberger K, Suttorp N. Role of Toll-like receptors, NOD-like receptors and RIG-I-like receptors in endothelial cells and systemic infections. *Thromb Haemost*. (2009) 102:1103–9. doi: 10.1160/TH09-05-0323
94. Gibson RL, Nizet V, Rubens CE. Group B streptococcal beta-hemolysin promotes injury of lung microvascular endothelial cells. *Pediatr Res*. (1999) 45:626–34. doi: 10.1203/00006450-199905010-00003
95. Khakpour S, Wilhelmsen K, Hellman J. Vascular endothelial cell Toll-like receptor pathways in sepsis. *Innate Immun*. (2015) 21:327–46. doi: 10.1177/1753425915606525
96. Berner R, Niemyer CM, Leititis JU, Funke A, Schwab C, Rau U, et al. Plasma levels and gene expression of granulocyte colony-stimulating factor, tumor necrosis factor-alpha, interleukin (IL)-1beta, IL-6, IL-8, and soluble intercellular adhesion molecule-1 in neonatal early onset sepsis. *Pediatr Res*. (1998) 44:469–77. doi: 10.1203/00006450-199810000-00002
97. Gonzalez BE, Mercado CK, Johnson L, Brodsky NL, Bhandari V. Early markers of late-onset sepsis in premature neonates: clinical, hematological and cytokine profile. *J Perinat Med*. (2003) 31:60–8. doi: 10.1515/JPM.2003.009
98. Gerdes N, Sukhova GK, Libby P, Reynolds RS, Young JL, Schonbeck U. Expression of interleukin (IL)-18 and functional IL-18 receptor on human vascular endothelial cells, smooth muscle cells, and macrophages: implications for atherogenesis. *J Exp Med*. (2002) 195:245–57. doi: 10.1084/jem.20011022
99. Sironi M, Breviario F, Proserpio P, Biondi A, Vecchi A, Van Damme J, et al. IL-1 stimulates IL-6 production in endothelial cells. *J Immunol*. (1989) 142:549–53.
100. Barnes TC, Anderson ME, Moots RJ. The many faces of interleukin-6: the role of IL-6 in inflammation, vasculopathy, and fibrosis in systemic sclerosis. *Int J Rheumatol*. (2011) 2011:721608. doi: 10.1155/2011/721608
101. Oude Nijhuis CS, Vellenga E, Daenen SM, Kamps WA, De Bont ES. Endothelial cells are main producers of interleukin 8 through Toll-like receptor 2 and 4 signaling during bacterial infection in leukopenic cancer patients. *Clin Diagn Lab Immunol*. (2003) 10:558–63. doi: 10.1128/CDLI.10.4.558-563.2003
102. Karenberg K, Hudalla H, Frommhold D. Leukocyte recruitment in preterm and term infants. *Mol Cell Pediatr*. (2016) 3:35. doi: 10.1186/s40348-016-0063-5
103. Qureshi MH, Cook-Mills J, Doherty DE, Garvy BA. TNF-alpha-dependent ICAM-1- and VCAM-1-mediated inflammatory responses are delayed in neonatal mice infected with *Pneumocystis carinii*. *J Immunol*. (2003) 171:4700–7. doi: 10.4049/jimmunol.171.9.4700
104. Bickes MS, Pirr S, Heinemann AS, Fehllhaber B, Halle S, Vollger L, et al. Constitutive TNF-alpha signaling in neonates is essential for the development of tissue-resident leukocyte profiles at barrier sites. *FASEB J*. (2019) 2019:fj201900796R. doi: 10.1096/fj.201900796R
105. Simmons J, Pittet JF. The coagulopathy of acute sepsis. *Curr Opin Anaesthesiol*. (2015) 28:227–36. doi: 10.1097/ACO.0000000000000163
106. Singer M. The role of mitochondrial dysfunction in sepsis-induced multi-organ failure. *Virulence*. (2014) 5:66–72. doi: 10.4161/viru.26907
107. Grover SP, Mackman N. Neutrophils, NETs, and immunothrombosis. *Blood*. (2018) 132:1360–1. doi: 10.1182/blood-2018-08-868067
108. Engelmann B, Massberg S. Thrombosis as an intravascular effector of innate immunity. *Nat Rev Immunol*. (2013) 13:34–45. doi: 10.1038/nri3345
109. Franchi T, Eaton S, De Coppi P, Giuliani S. The emerging role of immunothrombosis in paediatric conditions. *Pediatr Res*. (2019) 86:19–27. doi: 10.1038/s41390-019-0343-6
110. Yost CC. Pediatric immunothrombosis-understudied, but what potential! *Pediatr Res*. (2019) 86:17–8. doi: 10.1038/s41390-019-0389-5
111. Donati A, Tibboel D, Ince C. Towards integrative physiological monitoring of the critically ill: from cardiovascular to microcirculatory and cellular function monitoring at the bedside. *Crit Care*. (2013) 17(Suppl. 1):S5. doi: 10.1186/cc11503
112. Pandey A, John BM. Capillary refill time. Is it time to fill the gaps? *Med J Armed Forces India*. (2013) 69:97–8. doi: 10.1016/j.mjafi.2012.09.005
113. Mathew J, Bada Shekarappa C, Padubidri Nanyam Rao S. correlation between perfusion index and CRIB score in sick neonates admitted to a tertiary center. *J Trop Pediatr*. (2019) 65:84–9. doi: 10.1093/tropej/fmy016
114. van der Laan ME, Schat TE, Olthuis AJ, Boezen HM, Bos AF, Kooi EM. The association between multisite near-infrared spectroscopy and routine hemodynamic measurements in relation to short-term outcome in preterms with clinical sepsis. *Neonatology*. (2015) 108:297–304. doi: 10.1159/000438977
115. Garcia-Alvarez M, Marik P, Bellomo R. Sepsis-associated hyperlactatemia. *Crit Care*. (2014) 18:503. doi: 10.1186/s13054-014-0503-3
116. Massey MJ, Shapiro NI. A guide to human *in vivo* microcirculatory flow image analysis. *Crit Care*. (2016) 20:35. doi: 10.1186/s13054-016-1213-9
117. Top AP, Ince C, de Meij N, van Dijk M, Tibboel D. Persistent low microcirculatory vessel density in nonsurvivors of sepsis in pediatric intensive care. *Crit Care Med*. (2011) 39:8–13. doi: 10.1097/CCM.0b013e3181fb7994
118. De Backer D, Donadello K, Sakr Y, Ospina-Tascon G, Salgado D, Scolletta S, et al. Microcirculatory alterations in patients with severe sepsis: impact of time of assessment and relationship with outcome. *Crit Care Med*. (2013) 41:791–9. doi: 10.1097/CCM.0b013e3182742e8b
119. Lipowsky HH, Gao L, Lescanic A. Shedding of the endothelial glycocalyx in arterioles, capillaries, and venules and its effect on capillary hemodynamics during inflammation. *Am J Physiol Heart Circ Physiol*. (2011) 301:H2235–45. doi: 10.1152/ajpheart.00803.2011
120. Ince C, Boerma EC, Cecconi M, De Backer D, Shapiro NI, Duranteau J, et al. Second consensus on the assessment of sublingual microcirculation in critically ill patients: results from a task force of the European Society of Intensive Care Medicine. *Intensive Care Med*. (2018) 44:281–99. doi: 10.1007/s00134-018-5070-7
121. Cerny V, Astapenko D, Burkovskiy I, Hyspler R, Ticha A, Trevors MA, et al. Glycocalyx *in vivo* measurement. *Clin Hemorheol Microcirc*. (2017) 67:499–503. doi: 10.3233/CH-179235
122. Nieuwdorp M, Meuwese MC, Mooij HL, Ince C, Broekhuizen LN, Kastelein JJ, et al. Measuring endothelial glycocalyx dimensions in humans: a potential novel tool to monitor vascular vulnerability. *J Appl Physiol*. (2008) 104:845–52. doi: 10.1152/jappphysiol.00440.2007
123. Uz Z, van Gulik TM, Aydemirli MD, Guerci P, Ince Y, Cuppen D, et al. Identification and quantification of human microcirculatory leukocytes using handheld video microscopes at the bedside. *J Appl Physiol*. (2018) 124:1550–7. doi: 10.1152/jappphysiol.00962.2017

124. Genzel-Boroviczeny O, Christ F, Glas V. Blood transfusion increases functional capillary density in the skin of anemic preterm infants. *Pediatr Res.* (2004) 56:751–5. doi: 10.1203/01.PDR.0000141982.38959.10
125. Hiedl S, Schwepcke A, Weber F, Genzel-Boroviczeny O. Microcirculation in preterm infants: profound effects of patent ductus arteriosus. *J Pediatr.* (2010) 156:191–6. doi: 10.1016/j.jpeds.2009.08.034
126. Kroth J, Weidlich K, Hiedl S, Nussbaum C, Christ F, Genzel-boroviczeny O. Functional vessel density in the first month of life in preterm neonates. *Pediatr Res.* (2008) 64:567–71. doi: 10.1203/PDR.0b013e318184134e
127. van den Berg VJ, van Elteren HA, Buijs EA, Ince C, Tibboel D, Reiss IK, et al. Reproducibility of microvascular vessel density analysis in Sidestream dark-field-derived images of healthy term newborns. *Microcirculation.* (2015) 22:37–43. doi: 10.1111/micc.12163
128. van Elteren H, Reiss IK, de Jonge RC. Transcutaneous microcirculatory imaging in preterm neonates. *J Vis Exp.* (2015) :e53562. doi: 10.3791/53562
129. van Elteren HA, Ince C, Tibboel D, Reiss IK, de Jonge RC. Cutaneous microcirculation in preterm neonates: comparison between sidestream dark field (SDF) and incident dark field (IDF) imaging. *J Clin Monit Comput.* (2015) 29:543–8. doi: 10.1007/s10877-015-9708-5
130. Weidlich K, Kroth J, Nussbaum C, Hiedl S, Bauer A, Christ F, et al. Changes in microcirculation as early markers for infection in preterm infants—an observational prospective study. *Pediatr Res.* (2009) 66:461–5. doi: 10.1203/PDR.0b013e3181b3b1f6
131. Alba-Alejandre I, Hiedl S, Genzel-Boroviczeny O. Microcirculatory changes in term newborns with suspected infection: an observational prospective study. *Int J Pediatr.* (2013) 2013:768784. doi: 10.1155/2013/768784
132. Xing K, Murthy S, Liles WC, Singh JM. Clinical utility of biomarkers of endothelial activation in sepsis—a systematic review. *Crit Care.* (2012) 16:R7. doi: 10.1186/cc11145
133. Skibsted S, Jones AE, Puskarich MA, Arnold R, Sherwin R, Trzeciak S, et al. Biomarkers of endothelial cell activation in early sepsis. *Shock.* (2013) 39:427–32. doi: 10.1097/SHK.0b013e3182903f0d
134. Paulus P, Jennewein C, Zacharowski K. Biomarkers of endothelial dysfunction: can they help us deciphering systemic inflammation and sepsis? *Biomarkers.* (2011) 16(Suppl. 1) :S11–21. doi: 10.3109/1354750X.2011.587893
135. Mikacenic C, Hahn WO, Price BL, Harju-Baker S, Katz R, Kain KC, et al. Biomarkers of endothelial activation are associated with poor outcome in critical illness. *PLoS ONE.* (2015) 10:e0141251. doi: 10.1371/journal.pone.0141251
136. Vassiliou AG, Mastora Z, Orfanos SE, Jahaj E, Maniatis NA, Koutsoukou A, et al. Elevated biomarkers of endothelial dysfunction/activation at ICU admission are associated with sepsis development. *Cytokine.* (2014) 69:240–7. doi: 10.1016/j.cyto.2014.06.010
137. Hou PC, Filbin MR, Wang H, Ngo L, Huang DT, Aird WC, et al. Endothelial permeability and hemostasis in septic shock: results from the ProCESS trial. *Chest.* (2017) 152:22–31. doi: 10.1016/j.chest.2017.01.010
138. Zonneveld R, Martinelli R, Shapiro NI, Kuijpers TW, Plotz FB, Carman CV. Soluble adhesion molecules as markers for sepsis and the potential pathophysiological discrepancy in neonates, children and adults. *Crit Care.* (2014) 18:204. doi: 10.1186/cc13733
139. Hansen AB, Verder H, Staun-Olsen P. Soluble intercellular adhesion molecule and C-reactive protein as early markers of infection in newborns. *J Perinat Med.* (2000) 28:97–103. doi: 10.1515/JPM.2000.012
140. Phocas I, Sarandakou A, Giannaki G, Malamitsi-Puchner A, Rizos D, Zourlas PA. Soluble intercellular adhesion molecule-1 in newborn infants. *Eur J Pediatr.* (1998) 157:153–6. doi: 10.1007/s004310050788
141. Austgulen R, Arntzen KJ, Haereid PE, Aag S, Dollner H. Infections in neonates delivered at term are associated with increased serum levels of ICAM-1 and E-selectin. *Acta Paediatr.* (1997) 86:274–80. doi: 10.1111/j.1651-2227.1997.tb08889.x
142. Dollner H, Vatten L, Austgulen R. Early diagnostic markers for neonatal sepsis: comparing C-reactive protein, interleukin-6, soluble tumour necrosis factor receptors and soluble adhesion molecules. *J Clin Epidemiol.* (2001) 54:1251–7. doi: 10.1016/S0895-4356(01)00400-0
143. Figueras-Aloy J, Gomez-Lopez L, Rodriguez-Miguel JM, Salvia-Roiges MD, Jordan-Garcia I, Ferrer-Codina I, et al. Serum soluble ICAM-1, VCAM-1, L-selectin, and P-selectin levels as markers of infection and their relation to clinical severity in neonatal sepsis. *Am J Perinatol.* (2007) 24:331–8. doi: 10.1055/s-2007-981851
144. Edgar JD, Gabriel V, Gallimore JR, McMillan SA, Grant J. A prospective study of the sensitivity, specificity and diagnostic performance of soluble intercellular adhesion molecule 1, highly sensitive C-reactive protein, soluble E-selectin and serum amyloid A in the diagnosis of neonatal infection. *BMC Pediatr.* (2010) 10:22. doi: 10.1186/1471-2431-10-22
145. Saldir M, Tunc T, Cekmez F, Cetinkaya M, Kalayci T, Fidanci K, et al. Endocan and soluble triggering receptor expressed on myeloid cells-1 as novel markers for neonatal sepsis. *Pediatr Neonatol.* (2015) 56:415–21. doi: 10.1016/j.pedneo.2015.03.006
146. Wright JK, Hayford K, Tran V, Al Kibria GM, Baqui A, Manajjir A, et al. Biomarkers of endothelial dysfunction predict sepsis mortality in young infants: a matched case-control study. *BMC Pediatr.* (2018) 18:118. doi: 10.1186/s12887-018-1087-x
147. Zonneveld R, Jongman RM, Juliana A, Molema G, van Meurs M, Plotz FB. Serum concentrations of endothelial cell adhesion molecules and their shedding enzymes and early onset sepsis in newborns in Suriname. *BMJ Paediatr Open.* (2018) 2:e000312. doi: 10.1136/bmjpo-2018-000312
148. Vermette D, Hu P, Canarie MF, Funaro M, Glover J, Pierce RW. Tight junction structure, function, and assessment in the critically ill: a systematic review. *Intens Care Med Exp.* (2018) 6:37. doi: 10.1186/s40635-018-0203-4
149. Zhao GJ, Li D, Zhao Q, Lian J, Hu TT, Hong GL, et al. Prognostic value of plasma tight-junction proteins for sepsis in emergency department: an observational study. *Shock.* (2016) 45:326–32. doi: 10.1097/SHK.0000000000000524
150. Wynn JL, Polin RA. Progress in the management of neonatal sepsis: the importance of a consensus definition. *Pediatr Res.* (2018) 83:13–5. doi: 10.1038/pr.2017.224
151. Darwish I, Liles WC. Emerging therapeutic strategies to prevent infection-related microvascular endothelial activation and dysfunction. *Virulence.* (2013) 4:572–82. doi: 10.4161/viru.25740
152. Uchimido R, Schmidt EP, Shapiro NI. The glycocalyx: a novel diagnostic and therapeutic target in sepsis. *Crit Care.* (2019) 23:16. doi: 10.1186/s13054-018-2292-6
153. Delano MJ, Ward PA. Sepsis-induced immune dysfunction: can immune therapies reduce mortality? *J Clin Invest.* (2016) 126:23–31. doi: 10.1172/JCI82224
154. Martin L, Koczera P, Zechendorf E, Schuerholz T. The endothelial glycocalyx: new diagnostic and therapeutic approaches in sepsis. *BioMed Res Int.* (2016) 2016:3758278. doi: 10.1155/2016/3758278
155. Chappell D, Jacob M, Hofmann-Kiefer K, Bruegger D, Rehm M, Conzen P, et al. Hydrocortisone preserves the vascular barrier by protecting the endothelial glycocalyx. *Anesthesiology.* (2007) 107:776–84. doi: 10.1097/01.anes.0000286984.39328.96
156. Bruegger D, Schwartz L, Chappell D, Jacob M, Rehm M, Vogeser M, et al. Release of atrial natriuretic peptide precedes shedding of the endothelial glycocalyx equally in patients undergoing on- and off-pump coronary artery bypass surgery. *Basic Res Cardiol.* (2011) 106:1111–21. doi: 10.1007/s00395-011-0203-y
157. Chappell D, Bruegger D, Potzel J, Jacob M, Brettner F, Vogeser M, et al. Hypervolemia increases release of atrial natriuretic peptide and shedding of the endothelial glycocalyx. *Crit Care.* (2014) 18:538. doi: 10.1186/s13054-014-0538-5
158. Zazzeron L, Gattinoni L, Caironi P. Role of albumin, starches and gelatins versus crystalloids in volume resuscitation of critically ill patients. *Curr Opin Crit Care.* (2016) 22:428–36. doi: 10.1097/MCC.0000000000000341
159. Adamson RH, Clark JF, Radeva M, Kheirulomoom A, Ferrara KW, Curry FE. Albumin modulates S1P delivery from red blood cells in perfused microvessels: mechanism of the protein effect. *Am J Physiol Heart Circ Physiol.* (2014) 306:H1011–7. doi: 10.1152/ajpheart.00829.2013
160. Chelazzi C, Villa G, Mancinelli P, De Gaudio AR, Adembri C. Glycocalyx and sepsis-induced alterations in vascular permeability. *Crit Care.* (2015) 19:26. doi: 10.1186/s13054-015-0741-z
161. Coldewey SM, Benetti E, Collino M, Pfeilschifter J, Sponholz C, Bauer M, et al. Elevation of serum sphingosine-1-phosphate attenuates

- impaired cardiac function in experimental sepsis. *Sci Rep.* (2016) 6:27594. doi: 10.1038/srep27594
162. Lygizos MI, Yang Y, Altmann CJ, Okamura K, Hernando AA, Perez MJ, et al. Heparanase mediates renal dysfunction during early sepsis in mice. *Physiol Rep.* (2013) 1:e00153. doi: 10.1002/phy2.153
 163. Chen S, Zhang X, Sun Y, Hu Z, Lu S, Ma X. Unfractionated heparin attenuates intestinal injury in mouse model of sepsis by inhibiting heparanase. *Int J Clin Exp Pathol.* (2015) 8:4903–12.
 164. Schmidt EP, Yang Y, Janssen WJ, Gandjeva A, Perez MJ, Barthel L, et al. The pulmonary endothelial glycocalyx regulates neutrophil adhesion and lung injury during experimental sepsis. *Nat Med.* (2012) 18:1217–23. doi: 10.1038/nm.2843
 165. Liu YY, Lee CH, Dedaj R, Zhao H, Mrabat H, Sheidlin A, et al. High-molecular-weight hyaluronan—a possible new treatment for sepsis-induced lung injury: a preclinical study in mechanically ventilated rats. *Crit Care.* (2008) 12:R102. doi: 10.1186/cc6982
 166. Brindle NP, Saharinen P, Alitalo K. Signaling and functions of angiotensin-1 in vascular protection. *Circ Res.* (2006) 98:1014–23. doi: 10.1161/01.RES.0000218275.54089.12
 167. Fiedler U, Augustin HG. Angiotensins: a link between angiogenesis and inflammation. *Trends Immunol.* (2006) 27:552–8. doi: 10.1016/j.it.2006.10.004
 168. Huang ZW, Liu N, Li D, Zhang HY, Wang Y, Liu Y, et al. Angiotensin-1 modified human umbilical cord mesenchymal stem cell therapy for endotoxin-induced acute lung injury in rats. *Yonsei Med J.* (2017) 58:206–16. doi: 10.3349/ymj.2017.58.1.206
 169. Han S, Lee SJ, Kim KE, Lee HS, Oh N, Park I, et al. Amelioration of sepsis by TIE2 activation-induced vascular protection. *Sci Transl Med.* (2016) 8:335ra55. doi: 10.1126/scitranslmed.aad9260
 170. Hakanpaa L, Kiss EA, Jacquemet G, Miinalainen I, Lerche M, Guzman C, et al. Targeting beta1-integrin inhibits vascular leakage in endotoxemia. *Proc Natl Acad Sci USA.* (2018) 115:E6467–76. doi: 10.1073/pnas.1722317115
 171. Zonneveld R, Jongman R, Juliana A, Zijlmans W, Plotz F, Molema G, et al. Low serum angiotensin-1, high serum angiotensin-2, and high Ang-2/Ang-1 protein ratio are associated with early onset sepsis in surinamese newborns. *Shock.* (2017) 48:638–43. doi: 10.1097/SHK.0000000000000903
 172. Syed M, Das P, Pawar A, Aghai ZH, Kaskinen A, Zhuang ZW, et al. Hyperoxia causes miR-34a-mediated injury via angiotensin-1 in neonatal lungs. *Nat Commun.* (2017) 8:1173. doi: 10.1038/s41467-017-01349-y
 173. Liang WY, Jiang L. Relationship between expression of angiotensin-2 and retinal vascular development in hyperoxic rats. *Zhonghua Er Ke Za Zhi.* (2009) 47:204–8.
 174. Speer EM, Dowling DJ, Xu J, Ozog LS, Mathew JA, Chander A, et al. Pentoxifylline, dexamethasone and azithromycin demonstrate distinct age-dependent and synergistic inhibition of TLR- and inflammasome-mediated cytokine production in human newborn and adult blood *in vitro*. *PLoS ONE.* (2018) 13:e0196352. doi: 10.1371/journal.pone.0196352
 175. Speer EM, Diago-Navarro E, Ozog LS, Dowling DJ, Hou W, Raheel M, et al. Pentoxifylline alone or in combination with gentamicin or vancomycin inhibits live microbe-induced proinflammatory cytokine production in human cord blood and cord blood monocytes *in vitro*. *Antimicrob Agents Chemother.* (2018) 2018:62. doi: 10.1128/AAC.01462-18
 176. Lauterbach R, Pawlik D, Kowalczyk D, Ksycinski W, Helwich E, Zembala M. Effect of the immunomodulating agent, pentoxifylline, in the treatment of sepsis in prematurely delivered infants: a placebo-controlled, double-blind trial. *Crit Care Med.* (1999) 27:807–14. doi: 10.1097/00003246-199904000-00042
 177. *Intravenous Pentoxifylline as Adjunct Therapy to Improve Long-term Disability in Preterm Infants.* Trial registration ACTRN12616000405415 (2016).
 178. Unal C, Sen C, Iscen D, Dalcik H. *In vivo* observation of leukocyte-endothelium interaction in ischemia reperfusion injury with the dorsal window chamber and the effects of pentoxifylline on reperfusion injury. *J Surg Res.* (2007) 138:259–66. doi: 10.1016/j.jss.2006.05.019
 179. Wang P, Wood TJ, Ba ZF, Chaudry IH. Pentoxifylline maintains vascular endothelial cell function during hyperdynamic and hypodynamic sepsis. *Surgery.* (1996) 120:367–73. doi: 10.1016/S0039-6060(96)80311-4
 180. Nakagawa NK, Cruz RJ Jr, Aikawa P, Correia CJ, Cruz JW, Mauad T, et al. Pentoxifylline attenuates leukocyte-endothelial interactions in a two-hit model of shock and sepsis. *J Surg Res.* (2015) 193:421–8. doi: 10.1016/j.jss.2014.07.034
 181. Talar B, Gajos-Michniewicz A, Talar M, Chouaib S, Czyz M. Pentoxifylline inhibits WNT signalling in beta-cateninhigh patient-derived melanoma cell populations. *PLoS ONE.* (2016) 11:e0158275. doi: 10.1371/journal.pone.0158275
 182. Iba T, Levy JH, Hirota T, Hiki M, Sato K, Murakami T, et al. Protection of the endothelial glycocalyx by antithrombin in an endotoxin-induced rat model of sepsis. *Thromb Res.* (2018) 171:1–6. doi: 10.1016/j.thromres.2018.09.042
 183. Xiao F, Wang D, Kong L, Li M, Feng Z, Shuai B, et al. Intermedin protects against sepsis by concurrently re-establishing the endothelial barrier and alleviating inflammatory responses. *Nat Commun.* (2018) 9:2644. doi: 10.1038/s41467-018-05062-2

Conflict of Interest Statement: FB has signed a consulting agreement with Merck Sharp & Dohme Corp., a subsidiary of Merck & Co., Inc. This commercial relationship is unrelated to the current study. OL is a named inventor on patents for use of recombinant bactericidal/permeability-increasing protein (rBPI) in BPI-deficient humans including those exposed to total body irradiation.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Pietrasanta, Pugni, Ronchi, Bottino, Ghirardi, Sanchez-Schmitz, Borriello, Mosca and Levy. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.