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Rethinking the potential role of dose painting in personalized ultra-fractionated stereotactic adaptive radiotherapy

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Fractionated radiotherapy was established in the 1920s based upon two principles: (1) delivering daily treatments of equal quantity, unless the clinical situation requires adjustment, and (2) defining a specific treatment period to deliver a total dosage. Modern fractionated radiotherapy continues to adhere to these century-old principles, despite significant advancements in our understanding of radiobiology. At UT Southwestern, we are exploring a novel treatment approach called PULSAR (Personalized Ultra-Fractionated Stereotactic Adaptive Radiotherapy). This method involves administering tumoricidal doses in a pulse mode with extended intervals, typically spanning weeks or even a month. Extended intervals permit substantial recovery of normal tissues and afford the tumor and tumor microenvironment ample time to undergo significant changes, enabling more meaningful adaptation in response to the evolving characteristics of the tumor. The notion of dose painting in the realm of radiation therapy has long been a subject of contention. The debate primarily revolves around its clinical effectiveness and optimal methods of implementation. In this perspective, we discuss two facets concerning the potential integration of dose painting with PULSAR, along with several practical considerations. If successful, the combination of the two may not only provide another level of personal adaptation ("adaptive dose painting"), but also contribute to the establishment of a timely feedback loop throughout the treatment process. To substantiate our perspective, we conducted a fundamental modeling study focusing on PET-guided dose painting, incorporating tumor heterogeneity and tumor control probability (TCP).

KEYWORDS

radiotherapy, pulsar, dose painting, TCP, PET, MRI

1 Introduction

Traditional fractionated radiotherapy upholds two fundamental principles: daily fractionation and fixed treatment duration. These two principles leverage the disparities in radiobiological behaviors between malignant tumors and healthy tissues, striking a delicate equilibrium between tumor re-sensitization, regrowth, reoxygenation, resistance development, and the benefits of normal tissue repair (1–8). The two most employed models are the linear quadratic (LQ) model and the biological effective dose (BED) (9– 11). However, a significant limitation in these models is the assumption of constant radiosensitivity throughout the treatment course. This neglects the consideration of inter- and intratumoral heterogeneity, and its dynamic evolution over time (12–17).

In addition to radiosensitivity, the efficacy of radiation therapy is influenced by its scheduling. Numerous inquiries have delved into the potential advantages of prolonging the treatment duration, employing both clinical studies and mathematical modeling. Examples include hypofractionated stereotactic radiotherapy, stereotactic radiosurgery, ramp-up scheduling, and intermittent therapy (18-29). Despite the challenges linked to testing a considerable number of permutations in terms of dose and timing combinations, multiple studies have indicated that non-uniform fractionation is at least as effective, if not superior, to standard treatment in terms of overall survival, tumor control, and time to progression (18-29). At UT Southwestern (UTSW), our focus has been on exploring Personalized, Ultra-fractionated Stereotactic Adaptive Radiotherapy (PULSAR). PULSAR leverages the precision provided by image-guided radiation therapy technologies, delivering ablative radiation doses in a pulse mode with extended intervals between pulses (up to weeks or months). Each patient's response to treatment is distinct. PULSAR provides the flexibility to adapt to the evolving characteristics of tumor, encompassing changes in size, shape, and biomarker expression. To gain a clearer understanding of the unique characteristics of PULSAR, it is beneficial to contrast it with two counterparts: conventionally fractionated radiation therapy (CFRT) and standard stereotactic body radiation therapy (SBRT). In both CFRT and standard SBRT, it is not likely for the tumor to undergo substantial changes between two closely spaced radiation doses. Additionally, frequent dosing over brief intervals may not maximize potential synergies arising from concurrent immunooncology strategies. In a recent publication that delved into PULSAR in conjunction with anti-PD-L1 checkpoint inhibitor therapy using a preclinical model, the findings clearly indicated that the efficacy of immunotherapy was contingent upon the interval between two adjacent radiation pulses (30).

We believe there is a need to include non-uniform dose delivery in PULSAR, moving towards more personalized treatments. Firstly, advancements in imaging techniques, such as positron emission tomography (PET) with novel tracers and quantitative magnetic resonance imaging (MRI), provide more comprehensive anatomical, functional, and molecular information. On-board systems, exemplified by commercially available MR-Linac and PET-Linac, offer advanced image guidance and real-time adaptability. Secondly, the intervals between radiation pulses yield valuable images that, can form a feedback loop to evaluate the outcome of non-uniform dose delivery and make timely plan adaptation, with the aid of artificial intelligence (AI) and multiomics tools. Thirdly, PULSAR, as a special form of SBRT, minimizes the number of fractions with ablative dose per fraction (6-20 Gy). The increased dose, in contrast to the standard 2 Gy used in traditional daily fractionated radiotherapy, necessitates a more meticulous examination of the varied radiosensitivity effects. This is vital to address concerns associated with underdosing.

Dose painting is a technique that entails tailoring the radiation dose delivered to a specific area within a tumor (31-36). Instead of uniformly irradiating the entire tumor volume, dose painting allows for the adjustment of radiation doses to different regions inside the tumor based on factors such as the tumor's biological/functional characteristics and response to treatment. Its ultimate objective is to enhance the therapeutic ratio by increasing the radiation dose to the most resistant or aggressive parts of the tumor, while sparing healthy surrounding tissues from unnecessary radiation exposure. By incorporating dose painting into treatment planning, radiation oncologists can devise highly individualized and targeted radiation plans for each patient. Note that there is a subtle distinction between dose painting and dose escalation. Dose painting primarily entails customizing the radiation dose distribution within the tumor based on its heterogeneous biological characteristics, while dose escalation focuses on uniformly increasing the radiation dose across the entire tumor.

2 Intratumoral heterogeneity and TCP modeling

PET, as a molecular imaging modality, offers various biomarkers for this purpose. For instance, it leverages fluorodeoxyglucose (FDG) to identify glucose metabolism and tumor malignancy, as well as fluoromisonidazole (FMISO) to detect hypoxic conditions (37, 38). It is important to note that FDG specific uptake value (SUV) alone, which measures glucose metabolism, does not fully represent tumor size and heterogeneity. Furthermore, its quantitation is influenced by factors such as inflammation and necrosis. On the other hand, FMISO PET is used to detect areas of low oxygen concentration, which are associated with increased resistance to radiation therapy. FMISO images are typically analyzed using metrics like SUV_{max}, tumor-tomuscle ratio (TMR), or tumor-blood ratio (TBR). In the case of hypoxic tumors, it often requires a 2.5- to 3-fold higher dose to achieve an equivalent cytotoxic effect compared to non-hypoxic regions (39).

Over the past few years, the research community has encountered a blend of perplexing yet promising discoveries in relation to FDG/FMISO-based radiotherapy (40–46). One major question is the degree of spatial correlation between FDG and FMISO signals. FDG uptake hinges on the upregulation of glucose transporters and glycolytic enzymes, both of which are driven by hypoxia-inducible factor-1 transcription. As a result, this

complicates the correlation between FDG-PET and FMISO-PET signals. For instance, Torwarth et al. found no correlation in 12 primary HNC tumors (41). Gagel et al. suggested that using FDG to discriminate hypoxia was not feasible, yet a strong correlation was evident between FMISO and the hypoxic condition within the range of 2.5 to 10 mmHg, as determined through polarographic needle electrode measurements (42). Nehmeh et al. demonstrated that the voxel-wise correlation between FDG and FMISO is moderate within primary tumors, but no correlation exists in lymph nodes (45). Another relevant question is whether FDG/FMISO has any prognostic value in radiotherapy. Watanabe et al. discovered that in non-small cell lung cancer (NSCLC) patients, those with both high FDG and high FMISO levels exhibited the poorest prognosis (46). FDG uptake emerged as a good predictor of short progressionfree survival (PFS) only in patients of small tumors (<2 cm). In contrast, FMISO uptake consistently acted as a better predictor of short PFS in all patients, irrespective of tumor size. In the case of head and neck squamous cell carcinoma (HNSCC) patients undergoing chemo-radiation, Lee et al. reported no dependency of treatment outcomes on the hypoxic status of tumors (43), whereas Nicolay et al. presented opposite findings (44).

Tumor heterogeneity arises from two sources: the nonuniformity of SUV and the percentage of hypoxic volume relative to total tumor volume. To gauge the potential benefits of dose painting, we developed a simple TCP model as described below. The distribution of non-uniform FMISO SUV is illustrated in Figures 1A, B, drawing from two recent publications that center on FDG/FMISO-based SBRT. One study explored the tumor nonuniformity and the quantitative relationship between FDG and FMISO in a patient cohort, encompassing 20 primary tumors and 19 metastatic lymph nodes (45). The FMISO SUV non-uniformity (per patient) ranges from 18% to 52%. In another study of NSCLC patients treated with SBRT, the FMISO PET non-uniformity (recurrent/non-recurrent groups) reaches up to SUVmax (14%/ 19%), TMR (30%/9%), and TBR (18%/5%) (46). Besides nonuniform SUV, estimating the volume percentage of the hypoxic regions necessitates a region-of-interest approach, with limited published data available. In a study with a cohort of 323 patients across 17 different studies, the ratio between the hypoxic volume and the total tumor volume was estimated to fall within the range of 34% and 42% (40). Additionally, it was observed that with each doubling of the tumor volume, the fraction of hypoxic volume increased by four percentage points (41). As a result, we evaluated the following two extremes representing the hypoxic volume fraction, 25% and 50%.

Our simplified model consists of three steps as depicted in Figures 1C-E. First, the quantitative relationship between FMISO SUV and pO_2 was established based on an empirically fitted model (organ site-specific) (47) (Equation 1 and Figure 1C). Second, the oxygen enhancement ratio (OER) as a function of pO_2 was modeled (48–50) (Equation 2 and Figure 1D). The baseline reference of the partial pressure of oxygen associated with *SUV* was arbitrarily selected to be 10 mmHg. OER reflects the difference in the required dose corresponding to a given survival fraction, between two oxygenation conditions. A hypoxia-equivalent dose can thus be derived, scaling down by the OER relative to the baseline. Third, the

TCP dose-response was modelled considering the heterogeneity of hypoxic condition, based on a logistic function (Equation 3) (51– 53). The TCP depends on two parameters, D50 (the dose corresponding to a TCP of 50%) and γ 50 (the percent change in TCP per percent change of dose at the point at D50). A constant SUV was assumed inside the hypoxic volume with the uptake of *SUV_{max}*, with the non-uniformity defined as (*SUV_{max}-SUV_{avg}*)/ *SUV_{avg}*. We subsequently combined the dose responses of both hypoxic and non-hypoxic volumes to create the overall TCP curve, using a population-averaging approach (54, 55). D50 was set to be 80 Gy. Three γ 50 values were evaluated (3, 2 and 1), representing steep, moderate, and shallow dose-response relations, respectively. Finally, the absolute change in TCP was quantitatively analyzed using two figures-of-merit (FoMs): Δ TCP₅₀ and Δ TCP₈₀.

FMISO
$$SUV = 1.05 + 6.7^{(-0.117pO_2)}$$
 (1)

$$OER(pO_2) = 1 + 1.63(1 - e^{-0.26 \ pO_2})$$
 (2)

$$TCP(D) = \frac{1}{1 + \exp(-(D - D50)/k)}$$
 $k = \frac{D50}{4\gamma_{50}}$ (3)

One important characteristic of the TCP dose-response is illustrated in Figure 1E. The heterogeneity, either hypoxic condition or other biological characteristics, results in varying radiosensitivity. Under a uniform dose delivery, this implies that some spots are under-dosed (e.g., 50% of the dose to 30% of the volume), and the TCP curve is lowered consequently. The same argument applies to a situation in which the clonogen's radiosensitivity is uniform, but dose delivery is non-uniform. A distinctive characteristic of the sigmoid curve is that the under-dose region holds greater significance than the over-dose region, while over-dosing the non-hypoxic sub-volume has a minor impact. Even when the hypoxic sub-volume constitutes a small fraction of the total tumor volume, it exerts a dominant influence on the TCP profile. If the hypoxic sub-volume can be identified through techniques like PET or MRI, dose painting strategies can thus be implemented accordingly to restore the TCP curve.

The quantitative analyses of the change in both ΔTCP_{50} and ΔTCP_{80} are summarized in Figure 2, as a function of FMISO SUV non-uniformity. All cases exhibit a linear relationship with dose non-uniformity. For a larger hypoxic sub-volume and a steeper dose-response curve, the dependency is stronger. For a relative volume of 50% and γ 50 = 3, ΔTCP_{50} and ΔTCP_{80} amount up to 24% and 54%, respectively. On the other extreme, for a relative volume of 25% and γ 50 = 1, both ΔTCP_{50} and ΔTCP_{80} are about 2%. Note that while TCP is often discussed in the context of populations to evaluate treatment strategies, it can also be applied individually to inform treatment decisions for specific patients.

3 Discussion

Below we discuss the potential needs and benefits of conjoining dose painting with PULSAR. The combination of the two may not only provide another level of personal adaptation, but also



FIGURE 1

(A) 19 HNSCC patients, primary tumors and lymph nodes (from ref 45). (B) 22 NSCLC patients, non-recurrent group (13 patients) and recurrent group (9 patients) (from ref 46). (C) Relationship between FMISO SUV and pO_2 . The solid line represents the selected baseline reference ($pO_2 = 10$ mmHg). The dotted line represents the hypoxic state, corresponding to a higher SUV and a lower OER. (D) Relationship between OER and pO_2 . (E) The TCP dose-response curve (a sigmoid function) changes along with the hypoxic condition and associated radio-sensitivity.

contribute to the establishment of a timely feedback loop throughout the whole treatment. A mechanistic model that incorporates parameters such as SUV, OER, and TCP is presented. Despite the simplicity of the model and crude borderlines, it may serve as a guideline for designing dose painting studies and interpreting experimental results. The parameters in Equations 1-3, as well as the baseline reference associated with SUV and OER, can be flexibly adjusted for different imaging protocols or diseases sites. More advanced computational modelling of the tumor microenvironment can enable the design of virtual clinical trials. Nevertheless, as stated in the Introduction section, although PULSAR is a very recent concept, dose painting has been in place for many years with limited clinical adoption. In our view, controversies surrounding PULSAR and dose painting will persist, giving rise to numerous research questions to be answered. Two unique aspects along with several practical considerations, are elaborated below.

The first aspect pertains to the role of imaging in treatment planning and the spatial extent to which dose modulation can be achieved. This aspect is also closely connected to the distinction between dose painting and dose escalation. First of all, we need more evidence to determine whether dose painting (combined with more specific biomarkers) will surpass nonselective dose escalation and achieve improved treatment outcome (37). In addition, challenges exist regarding the utilization of information on tumor heterogeneity derived from PET and MRI images. Although the FMISO uptake represents the hypoxic heterogeneity inside tumors and identifies a biological target volume, it may not be suitable for contouring purposes. The target volumes such as gross tumor volume (GTV) and clinical target volume (CTV), are routinely based on FDG PET. Note that the spatial information of heterogeneity is not reflected in Figures 1A, B, and the TCP change derived in our model assumes that the hypoxic-related heterogeneity comes from FMISO alone. However, such heterogeneity may be attenuated by the FDG-associated heterogeneity. For instance, a dose-painting strategy relying on dose escalation to the hypoxic region only, would not be advantageous if the hypoxic region extensively overlaps with the hyper-metabolic volume based on FDG-avid voxels (45). That said, it is the additional contrast between FMISO and FDG that determines the potential benefits of dose painting. In comparison with FDG-PET, the slower pharmokinetic of FMISO (~3-4 hours) and lower SUV (~1-3) may introduce additional uncertainty in quantification which should also be considered in dose painting (45, 46). Other PET tracers for imaging hypoxia can also be examined for PET-guided dose painting, such as HX4, ATSM and FAZA (56-59). Lastly, we anticipate that immuno-PET has great potential to open unique research avenues and foster the synergy between PULSAR and immunotherapy (60).

Quantitative MRI represents another valuable imaging tool in dose painting. Within tumors, subvolumes exhibiting distinct functional characteristics, such as physiology, perfusion, and oxygenation, can be recognized as diverse tumor habitats (61–64). For example, in glioblastoma, hypercellular tumor components identified through high b-value diffusion-weighted imaging (DWI) offer additional insights compared to those from dynamic contrast-enhanced (DCE) (63, 64). The hypercellular tumor volume determined by diffusion MRI well correlates with progression-free survival (61). Therefore, the thresholds of quantitative MRI imaging features can be determined empirically or established through histopathology and clinical outcome. Along with PET, they hold significant promise to provide radiation oncologists with more insights for crafting dose painting strategies.

From a physics and engineering standpoint, the spatial modulation capacity of beam delivery and factors such as finite image spatial resolution (e.g., 4-6 mm) may impose constraints on the implementation of dose painting. This is reminiscent of the situation in spatially fractionated or GRID radiation therapy, a specialized technique that administers high-dose radiation in a grid-like or checkerboard pattern to the target area, with a modulation capacity of approximately 1 cm (65). In conventional radiotherapy, the multi-leaf collimator offers spatial modulation with a resolution of ~5 mm. Analogously considering the concept of point spread function in image reconstruction, it prompts an intriguing question of whether the attainable benefits of dose painting might be attenuated, compared to the results shown in Figure 2. Additional challenges in margin setting and motion management contribute to the concerns surrounding this issue.

The second aspect pertains to the extended interval between fractions, a unique feature of PULSAR. Biology takes time. Longer intervals allow more normal tissue recovery after injury, and at the same time, provide time for tumor to undergo dramatic changes including radioresistance. PULSAR can also operate in association with immunotherapy. Compared to daily or every other day fractions, PULSAR is more likely to harness the synergy between the two, since the adaptive immune response takes time to develop and reach its full effectiveness (30, 66–70). Recently, several modeling studies have been conducted to provide mechanistic interpretation of the interaction of radiation with checkpoint blockers through ordinary differential equations, examining the dependence of the PULSAR effect as a function of dose and timing (71–73).

We recognize that the existing evidence does not entirely endorse the effectiveness of dose painting, and skepticisms persist in a number of aspects including the reproducibility of radiotracers, the need of intra-fractional PET, and the consistency of radiotracers in characterizing chronic hypoxia during radiotherapy. Notwithstanding these concerns, we envision that dose painting has potential to be integrated with PULSAR to address intratumoral heterogeneity and improve overall treatment response, targeting locoregional-resistant regions more effectively (74). Three scenarios are elaborated below.

Firstly, PULSAR minimizes the number of fractions with ablative dose per fraction (6-20 Gy). Compared to 2 Gy in traditional daily fractionated radiotherapy, the anticipated impact of dose modulation will be magnified, due to the shift toward the high-dose end in the survival curve as described by the LQ model. In other words, a large dynamic range is available for implementing dose painting and mitigating possible underdosing.

Secondly, the extended interval in PULSAR allows for more complete reoxygenation. The process of reoxygenation itself is rapid, but its cumulative impact on the hypoxic state postirradiation is complicated and may last several days. One single fraction SBRT study reported that the proportion of surviving hypoxic cells decreased over several days postirradiation (75). One plausible explanation is that the oxygen supply through small fractions of blood vessels that survived high-dose exposure, combined with a reduction of oxygen consumption due to massive cell death, causes the reoxygenation of hypoxic cells (75). In PULSAR, the extended inter-fraction time helps ensure complete reoxygenation while avoiding repopulation. Dose painting can thus be implemented based on the latest distribution of hypoxic subvolumes, improving the efficacy of radiation treatment.



 Δ TCP₅₀ and Δ TCP₈₀ as a function of SUV non-uniformity for three cases (case 1: γ 50 = 3, case 2: γ 50 = 2, case 3: γ 50 = 1), and two hypoxic volume fractions: (A) 25% and (B) 50%.

Thirdly, multiple PET or MRI images are accessible at various stages in PULSAR (pre-treatment, mid-treatment, and posttreatment), which provide valuable insights into patient response to treatment. These data can be analyzed using multiomics (radiomics, dosiomics) and AI tools. For example, delta radiomics entails the extraction of radiomic features from the same region of interest within the same patient but at different time points (76-79). In contrast to standard radiomics, which captures a static snapshot, delta radiomics focus on the phenotypic changes in tumors that occur after the treatment. It is thus reasonable to expect that delta radiomics will provide enhanced predictive capabilities compared to conventional radiomic features, making them especially suitable for PULSAR. Besides radiomics, other deep learning features such as U-Net or transformer-based models can also be used to extract specific architecture or details within the images. In the end, we can adopt a more personalized approach based on AI prediction of each patient's distinct response. Moreover, dose painting in PULSAR may go beyond pre-treatment images and incorporate posttreatment images for updating dose prescription, akin to background subtraction. The adaptive dose painting, in conjunction with a concurrent feedback loop through multiple imaging steps, allowing us to assess both the need and efficacy of dose painting in a timely manner. On a side note, the combination of PULSAR with dose painting can play a unique role in both photon and particle therapy. One promising direction can be the use of carbon or, even better, oxygen ions to overcome hypoxia independently of dose painting (80, 81).

To conclude, we foresee that the advancements in imaging technology, the emphasis on adaptive radiotherapy, the evolution of AI tools, and combined radio-immunotherapy are bolstering the demand for dose painting. The integration of dose painting into PULSAR, while a novel concept, is expected to hold substantial promise for progressing radiation therapy, initiating a shift in the approach to personalized cancer treatment.

References

1. Thames HD. Early fractionation methods and the origins of the NSD concept. *Acta Oncol.* (1988) 27:89–103. doi: 10.3109/02841868809090329

2. Pawlik TM, Keyomarsi K. Role of cell cycle in mediating sensitivity to radiotherapy. *Int J Radiat Oncol.* (2004) 59:928–42. doi: 10.1016/j.ijrobp.2004.03.005

3. Kim JJ, Tannock IF. Repopulation of cancer cells during therapy: An important cause of treatment failure. *Nat Cancer*. (2005) 5:516–25. doi: 10.1038/nrc1650

4. Wang JZ, Li XA. Impact of tumor repopulation on radiotherapy planning. Int J Radiat Oncol. (2005) 61:220–7. doi: 10.1016/j.ijrobp.2004.09.043

5. Marcu LG. Altered fractionation in radiotherapy: From radiobiological rationale to therapeutic gain. *Cancer Treat Rev.* (2010) 36:606–14. doi: 10.1016/j.ctrv.2010.04.004

6. Biau J, Chautard E, Verrelle P, Dutreix M. Altering DNA repair to improve radiation therapy: Specific and multiple pathway targeting. *Front Oncol.* (2019) 9:1009. doi: 10.3389/fonc.2019.01009

7. Enderling H, Alfonso JCL, Moros E, Caudell JJ, Harrison LB. Integrating mathematical modeling into the roadmap for personalized adaptive radiation therapy. *Trends Cancer.* (2019) 5:467–74. doi: 10.1016/j.trecan.2019.06.006

8. Brady R, Enderling H. Mathematical models of cancer: When to predict novel therapies, and when not to. *Bull Math Biol.* (2019) 81:3722–31. doi: 10.1007/s11538-019-00640-x

9. Jones B, Dale R, Deehan C, Hopkins K, Morgan D. The role of biologically effective dose (BED) in clinical oncology. *Clin Oncol.* (2001) 13:71–81. doi: 10.1053/clon.2001.9221

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Author contributions

HP: Investigation, Resources, Writing – original draft, Writing – review & editing. JD: Writing – review & editing. SJ: Writing – review & editing.

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10. Fowler JF. The linear-quadratic formula and progress in fractionated radiotherapy. Br J Radiol. (1989) 62:679–94. doi: 10.1259/0007-1285-62-740-679

11. Fowler JF. 21 years of biologically effective dose. Br J Radiol. (2010) 83:554-68. doi: 10.1259/bjr/31372149

12. Steel GG, McMillan TJ, Peacock JH. The 5Rs of radiobiology. Int J Radiat Biol. (1989) 56:1045–8. doi: 10.1080/09553008914552491

13. Lewin TD, Maini PK, Moros EG, Enderling H, Byrne HM. The evolution of tumor composition during fractionated radiotherapy: implications for outcome. *Bull Math Biol.* (2018) 80:1207–35. doi: 10.1007/s11538-018-0391-9

14. Niida A, Nagayama S, Miyano S, Mimori K. Understanding intratumor heterogeneity by combining genome analysis and mathematical modeling. *Cancer Sci.* (2018) 109:884–92. doi: 10.1111/cas.13510

15. Alfonso JCL, Berk L. Modeling the effect of intratumoral heterogeneity of radiosensitivity on tumor response over the course of fractionated radiation therapy. *Radiat Oncol.* (2019) 14:88. doi: 10.1186/s13014-019-1288-y

16. Marusyk A, Janiszewska M, Polyak K. Intratumor heterogeneity: The rosetta stone of therapy resistance. *Cancer Cell.* (2020) 37:471-84. doi: 10.1016/j.ccell.2020.03.007

17. Krisnawan VE, Stanley JA, Schwarz JK, DeNardo DG. Tumor microenvironment as a regulator of radiation therapy: New insights into stromal-mediated radioresistance. *Cancers*. (2020) 12:2916. doi: 10.3390/cancers12102916

18. Alfonso JCL, Parsai S, Joshi N, Godley A, Shah C, Koyfman SA, et al. Temporally feathered intensity-modulated radiation therapy: A planning technique to reduce normal tissue toxicity. *Med Phys.* (2018) 45:3466–74. doi: 10.1002/mp.12988

19. Navarria P, Pessina F, Cozzi L, Ascolese AM, De Rose F, Fogliata A, et al. Hypofractionated stereotactic radiotherapy alone using volumetric modulated arc therapy for patients with single, large brain metastases unsuitable for surgical resection. *Radiat Oncol.* (2016) 11:76. doi: 10.1186/s13014-016-0653-3

20. Angelov L, Mohammadi AM, Bennett EE, Abbassy M, Elson P, Chao ST, et al. Impact of 2-staged stereotactic radiosurgery for treatment of brain metastases \geq 2 cm. J Neurosurg. (2017) 129:366–82. doi: 10.3171/2017.3.JNS162532

21. Yamamoto M, Higuchi Y, Serizawa T, Kawabe T, Nagano O, Sato Y, et al. Treestage gamma knife treatment for metastatic brain tumors larger than 10 cm3: A 2institute study including re-analyses of earlier results using competing risk analysis. *J Neurosurg.* (2018), 129:77–85. doi: 10.3171/2018.7.GKS181392

22. Higuchi Y, Serizawa T, Nagano O, Matsuda S, Ono J, Sato M, et al. Three-staged stereotactic radiotherapy without whole brain irradiation for large metastatic brain tumors. *Int J Radiat Oncol Biol Phys.* (2009) 74:1543–8. doi: 10.1016/j.ijrobp.2008.10.035

23. Yu VY, Nguyen D, O'Connor D, Ruan D, Kaprealian T, Chin R, et al. Treating Glioblastoma Multiforme (GBM) with super hyperfractionated radiation therapy: Implication of temporal dose fractionation optimization including cancer stem cell dynamics. *PLoS One.* (2021) 16:e0245676. doi: 10.1371/journal.pone.0245676

24. Kuznetsov M, Kolobov A. Optimization of dose fractionation for radiotherapy of a solid tumor with account of oxygen effect and proliferative heterogeneity. *Mathematics.* (2020) 8:1204. doi: 10.3390/math8081204

25. Brüningk SC, Peacock J, Whelan CJ, Brady-Nicholls R, Yu HM, Sahebjam S, et al. Intermittent radiotherapy as alternative treatment for recurrent high grade glioma: A modeling study based on longitudinal tumor measurements. *Sci Rep.* (2021) 11:17674. doi: 10.1038/s41598-021-99507-2

26. Pérez-García VM, Bogdanska M, Martínez-González A, Belmonte-Beitia J, Schucht P, Pérez-Romasanta LA. Delay effects in the response of low-grade gliomas to radiotherapy: A mathematical model and its therapeutical implications. *Math Med Biol.* (2015) 32:307–29. doi: 10.1093/imammb/dqu009

27. Henares-Molina A, Benzekry S, Lara PC, García-Rojo M, Pérez-García VM, Martinez-Gonzalez A. Non-standard radiotherapy fractionations delay the time to Malignant transformation of low-grade gliomas. *PLoS One.* (2017) 12:e0178552. doi: 10.1371/journal.pone.0178552

28. Leder K, Pitter K, LaPlant Q, Hambardzumyan D, Ross BD, Chan TA, et al. Mathematical modeling of PDGF-driven glioblastoma reveals optimized radiation dosing schedules. *Cell.* (2014) 156:603–16. doi: 10.1016/j.cell.2013.12.029

29. Badri H, Pitter K, Holland EC, Michor F, Leder K. Optimization of radiation dosing schedules for proneural glioblastoma. *J Math Biol.* (2015) 72:1301–36. doi: 10.1007/s00285-015-0908-x

30. Moore C, Hsu CC, Chen WM, Chen BPC, Han C, Story M, et al. Personalized ultrafractionated stereotactic adaptive radiotherapy (PULSAR) in preclinical models enhances single-agent immune checkpoint blockade. *Int J Radiat Oncol Biol Phys.* (2021) 110:1306–16. doi: 10.1016/j.jirobp.2021.03.047

31. Ling C, Humm J, Larson S, Amols H, Fuks Z, Leibel S, et al. Towards multidimensional radiotherapy (MD-CRT): biological imaging and biological conformality. *Int J Radiat Oncol Biol Phys.* (2000) 47:551–60. doi: 10.1016/S0360-3016(00)00467-3

32. Bentzen SM. Theragnostic imaging for radiation oncology: dose-painting by numbers. *Lancet Oncol.* (2005) 6:112–7. doi: 10.1016/S1470-2045(05)01737-7

33. Bentzen SM, Gregoire V. Molecular imaging-based dose painting: a novel paradigm for radiation therapy prescription. *Semin Radiat Oncol.* (2011) 21:101–10. doi: 10.1016/j.semradonc.2010.10.001

34. Supiot S, Lisbona A, Paris F, Azria D, Fenoglietto P. "Dose-painting": mythe ou réalité? *Cancer Radiother*. (2010) 14:554–62. doi: 10.1016/j.canrad.2010.06.005

35. Lopez Alfonso JC, Jagiella N, Nunez L, Herrero MA, Drasdo D. Estimating dose painting effects in radiotherapy: A mathematical model. *PLoS One.* (2014) 9:e89380. doi: 10.1371/journal.pone.0089380

36. Petit SF, Breedveld S, Unkelbach J, den Hertog D, Balvert M. Robust dosepainting-by-numbers vs. nonselective dose escalation for non-small cell lung cancer patients. *Med Phys.* (2021) 48:3096–108. doi: 10.1002/mp.14840

37. James ML, Gambhir SS. A molecular imaging primer: modalities, imaging agents, and applications. *Physiol Rev.* (2012) 92:897-965. doi: 10.1152/ physrev.00049.2010

38. Lardinois D, Weder W, Hany TF, Kamel EM, Korom S, Seifert B, et al. Staging of non-small-cell lung cancer with integrated positron-emission tomography and computed tomography. *N Engl J Med.* (2003) 348:2500–7. doi: 10.1056/NEJMoa022136

39. Gray LH, Conger AD, Ebert M, Hornsey S, Scott OC. The concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol.* (1953) 26:638–48. doi: 10.1259/0007-1285-26-312-638

40. Hildingsson S, Gebre-Medhin M, Zschaeck S, Adrian G. Hypoxia in relationship to tumor volume using hypoxia PET-imaging in head & neck cancer – A scoping review. *Clin Trans Radiat Oncol.* (2022) 36:40–6. doi: 10.1016/j.ctro.2022.06.004

41. Thorwarth D, Eschmann SM, Holzner F, Paulsen F, Alber M. Combined uptake of [18F]FDG and [18F]FMISO correlates with radiation therapy outcome in head-and-

neck cancer patients. Radiother Oncol. (2006) 80:151-6. doi: 10.1016/ j.radonc.2006.07.033

42. Gagel B, Reinartz P, Dimartino E, Zimny M, Pinkawa M, Maneschi P, et al. pO (2) Polarography versus positron emission tomography ([(18)F] fluoromisonidazole, [(18)F]-2-fluoro-2'-deoxyglucose). An appraisal of radiotherapeutically relevant hypoxia. *Strahlenther Onkol.* (2004) 180:616–22. doi: 10.1007/s00066-004-1229-y

43. Lee N, Nehmeh S, Schöder H, Fury M, Chan K, Ling CC, et al. Prospective trial incorporating pre-/mid-treatment [18F]-misonidazole positron emission tomography for head-and-neck cancer patients undergoing concurrent chemoradiotherapy. *Int J Radiat Oncol Biol Phys.* (2009) 75:101–8. doi: 10.1016/j.ijrobp.2008.10.049

44. Nicolay NH, Wiedenmann N, Mix M, Weber WA, Werner M, Grosu AL, et al. Correlative analyses between tissue-based hypoxia biomarkers and hypoxia PET imaging in head and neck cancer patients during radiochemotherapy-results from a prospective trial. *Eur J Nucl Med Mol Imaging.* (2020) 47:1046–55. doi: 10.1007/s00259-019-04598-9

45. Nehmeh SA, Moussa MB, Lee N, Zanzonico P, Gönen M, Humm JL, et al. Comparison of FDG and FMISO uptakes and distributions in head and neck squamous cell cancer tumors. *Eur J Nucl Med Mol Imaging*. (2021) 11:38. doi: 10.1186/s13550-021-00767-w

46. Watanabe S, Inoue T, Okamoto S, Magota K, Takayanagi A, Sakakibara-Konishi J, et al. Combination of FDG-PET and FMISO-PET as a treatment strategy for patients undergoing early-stage NSCLC stereotactic radiotherapy. *Eur J Nucl Med Mol Imaging.* (2019) 9:104. doi: 10.1186/s13550-019-0578-6

47. Piert M, Machulla HJ, Becker G, Aldinger P, Winter E, Bares R. Dependency misonidazole uptake on oxygen delivery and tissue porcine liver. *Nucl Med Biol.* (2000) 27:693–700. doi: 10.1016/S0969-8051(00)00151-7

48. Alper T, Howward-Flanders P. Role of oxygen in modifying the radiosensitivity of E. Coli B. *Nature.* (1956) 178:978–9. doi: 10.1038/178978a0

49. Wenzl T, Wilkens JJ. Modelling of the oxygen enhancement ratio for ion beam radiation therapy. *Phys Med Biol.* (2011) 56:3251. doi: 10.1088/0031-9155/56/11/006

50. Grimes DR, Partridge M, Grimes DR, Partridge M. A mechanistic investigation of the oxygen fixation hypothesis and oxygen enhancement ratio. *BioMed Phys Eng Express.* (2015) 1:45209. doi: 10.1088/2057-1976/1/4/045209

51. Goitein M. Causes and consequences of inhomogeneous dose distributions in radiation therapy. *Int J Radiat Oncol Biol Phys.* (1986) 12(4):701–4. doi: 10.1016/0360-3016(86)90084-2

52. Niemierko A, Goitein M. Implementation of a model for estimating tumor control probability for an inhomogeneously irradiated tumor. *Radiother Oncol.* (1993) 29:140–7. doi: 10.1016/0167-8140(93)90239-5

53. Webb S, Nahum AE. A model for calculating tumour control probability in radiotherapy including the effects of inhomogeneous distributions of dose and clonogenic cell density. *Phys Med Biol.* (1993) 38:653–66. doi: 10.1088/0031-9155/38/6/001

54. Shuryak I, Carlson DJ, Brown JM, Brenner DJ. High-dose and fractionation effects in stereotactic radiation therapy: Analysis of tumor control data from 2965 patients. *Radiother Oncol.* (2015) 115:327–34. doi: 10.1016/j.radonc.2015.05.013

55. Carlone MC, Warkentin B, Stavrev P, Fallone BG. Fundamental form of a population TCP model in the limit of large heterogeneity. *Med Phys.* (2006) 33:1634–42. doi: 10.1118/1.2193690

56. Stieb S, Eleftheriou A, Warnock G, Guckenberger M, Riesterer O. Longitudinal PET imaging of tumor hypoxia during the course of radiotherapy. *Eur J Nucl Med Mol Imaging*. (2018) 45:2201–17. doi: 10.1007/s00259-018-4116-y

57. Sanduleanu S, Wiel AMAV, Lieverse RIY, Marcus D, Ibrahim A, Primakov S, et al. Hypoxia PET imaging with [18F]-HX4-A promising next-generation tracer. *Cancers (Basel).* (2020) 12:1322. doi: 10.3390/cancers12051322

58. Taylor E, Zhou J, Lindsay P, Foltz W, Cheung M, Siddiqui I, et al. Quantifying reoxygenation in pancreatic cancer during stereotactic body radiotherapy. *Sci Rep.* (2020) 10:1638. doi: 10.1038/s41598-019-57364-0

59. Singleton DC, Macann A, Wilson WR. Therapeutic targeting of the hypoxic tumour microenvironment. *Nat Rev Clin Oncol.* (2021) 18:751–72. doi: 10.1038/s41571-021-00539-4

60. Wei W, Rosenkrans ZT, Liu J, Huang G, Luo QY, Cai W. ImmunoPET: concept, design, and applications. *Chem Rev.* (2020) 120:3787–851. doi: 10.1021/acs.chemrev.9b00738

61. Law M, Young RJ, Babb JS, Peccerelli N, Chheang S, Gruber ML, et al. Gliomas: predicting time to progression or survival with cerebral blood volume measurements at dynamic susceptibility-weighted contrast-enhanced perfusion MR imaging. *Radiology*. (2008) 247:490–8. doi: 10.1148/radiol.2472070898

62. van der Heide UA, Houweling AC, Groenendaal G, Beets-Tan RG, Lambin P. Functional MRI for radiotherapy dose painting. *Magn Reson Imaging*. (2012) 30:1216–23. doi: 10.1016/j.mri.2012.04.010

63. Pramanik PP, Parmar HA, Mammoser AG, Junck LR, Kim MM, Tsien CI, et al. Hypercellularity components of glioblastoma identified by high b-value diffusion-weighted imaging. *Int J Radiat Oncol Biol Phys.* (2015) 92:811–9. doi: 10.1016/j.ijrobp.2015.02.058

64. Li Y, Kim M, Lawrence TS, Parmar H, Cao Y. Microstructure modeling of high b-value diffusion-weighted images in glioblastoma. *Tomography.* (2020) 6:34–43. doi: 10.18383/j.tom.2020.00018

65. Yan W, Khan MK, Wu X, Simone CB, Fan J, Gressen E, et al. Spatially fractionated radiation therapy: History, present and the future. *Clin Transl Radiat Oncol.* (2019) 20:30–8. doi: 10.1016/j.ctro.2019.10.004

66. Sharabi AB, Tran PT, Lim M, Drake CG, Deweese TL. Stereotactic radiation therapy combined with immunotherapy: Augmenting the role of radiation in local and systemic treatment. *Oncol (Williston Park)*. (2015) 29:331–40.

67. Gutiontov SI, Pitroda SP, Chmura SJ, Arina A, Weichselbaum RR. Cytoreduction and the optimization of immune checkpoint inhibition with radiation therapy. *Int J Radiat Oncol Biol Phys.* (2020) 108:17–26. doi: 10.1016/j.ijrobp.2019.12.033

68. Jagodinsky JC, Harari PM, Morris ZS. The promise of combining radiation therapy with immunotherapy. *Int J Radiat Oncol Biol Phys.* (2020) 108:6–16. doi: 10.1016/j.ijrobp.2020.04.023

69. Zhu S, Zhang T, Zheng L, Liu H, Song W, Liu D, et al. Combination strategies to maximize the benefits of cancer immunotherapy. *J Hematol Oncol.* (2021) 14:156. doi: 10.1186/s13045-021-01164-5

70. Gong J, Le TQ, Massarelli E, Hendifar AE, Tuli R. Radiation therapy and PD-1/ PD-L1 blockade: The clinical development of an evolving anticancer combination. *J Immunother Cancer.* (2018) 6:46. doi: 10.1186/s40425-018-0361-7

71. Friedrich T, Scholz M, Durante M. A predictive biophysical model of the combined action of radiation therapy and immunotherapy of cancer. *Int J Radiat Oncol Biol Phys.* (2022) 113:872–84. doi: 10.1016/j.ijrobp.2022.03.030

72. Rouf S, Moore C, Saha D, Nguyen D, Bleile M, Timmerman R, et al. PULSAR effect: revealing potential synergies in combined radiation therapy and immunotherapy via differential equations. arXiV doi: 10.48550/arXiv.2402.06101

73. Xing Y, Moore C, Saha D, Nguyen D, Bleile M, Jia X, et al. Mathematical modeling of the synergetic effect between radiotherapy and immunotherapy. *arXiV* doi: 10.48550/arXiv.2401.00024

74. Vitale I, Shema E, Loi S, Galluzzi L, et al. Intratumoral heterogeneity in cancer progression and response to immunotherapy. *Nat Med.* (2021) 27:212–24. doi: 10.1038/s41591-021-01233-9

75. Song CW, Griffin RJ, Lee YJ, Cho H, Seo J, Park I, et al. Reoxygenation and repopulation of tumor cells after ablative hypofractionated radiotherapy (SBRT and SRS) in murine tumors. *Radiat Res.* (2019) 192:159–68. doi: 10.1667/RR15346.1

76. Barabino E, Rossi G, Pamparino S, Fiannacca M, Caprioli S, Fedeli A, et al. Exploring response to immunotherapy in non-small cell lung cancer using delta-radiomics. *Cancers*. (2022) 14:350. doi: 10.3390/cancers14020350

77. Nardone V, Reginelli A, Guida C, Belfiore MP, Biondi M, Mormile M. Deltaradiomics increases multicentre reproducibility: A phantom study. *Med Oncol.* (2020) 37:38. doi: 10.1007/s12032-020-01359-9

78. Plautz TE, Zheng C, Noid G, Li XA. Time stability of delta-radiomics features and the impact on patient analysis in longitudinal CT images. *Med Phys.* (2019) 46:1663–76. doi: 10.1002/mp.13395

79. Abbas E, Fanni SC, Bandini C, Francischello R, Febi M, Aghakhanyan G, et al. Delta-radiomics in cancer immunotherapy response prediction: A systematic review. *Eur J Radiol.* (2023) 11:100511. doi: 10.1016/j.ejro.2023.100511

80. Sokol O, Durante M. Carbon ions for hypoxic tumors: are we making the most of them? *Cancers*. (2023) 15:4494. doi: 10.3390/cancers15184494

81. Sokol O, Scifoni E, Tinganelli W, Kraft-Weyrather W, Wiedemann J, Maier A, et al. Oxygen beams for therapy: advanced biological treatment planning and experimental verification. *Phys Med Biol.* (2017) 62:7798–813. doi: 10.1088/1361-6560/aa88a0