

Obstacles, Research Progress, and Prospects of Oral Delivery of Bioactive Peptides: A Comprehensive Review

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Provisional

Obstacles, Research Progress, and Prospects of Oral Delivery of Bioactive Peptides: A Comprehensive Review

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11 peptide transports₅.

12 **Abstract**

13 Bioactive peptides hold significant potential for enhancing human health, however, their limited
14 oral bioavailability poses a substantial barrier to their widespread use in the food and pharmaceutical
15 industries. This article reviews the key factors influencing the absorption efficiency of oral bioactive
16 peptides, including issues related to bitter taste perception, challenges in gastrointestinal
17 environmental stability, and limitations in transmembrane transport. Furthermore, it highlights the
18 latest technologies, such as osmotic technology, chemical modification, and advanced delivery
19 systems, and discusses their advantages in enhancing the stability of bioactive peptides and
20 facilitating intestinal absorption. In addition, the application and challenges of common delivery
21 systems such as liposomes, emulsions, polymer nanoparticles, and hydrogels in oral bioactive peptide
22 delivery are also discussed. This paper aims to provide a theoretical foundation for scientific research
23 and practical applications of oral delivery of bioactive peptides, thereby promoting the further
24 development of bioactive peptides in the context of human health.

25 **1 Introduction**

26 Bioactive peptides are a class of compounds composed of natural amino acids arranged in
27 various combinations, sequences, and spatial conformations. These peptides exhibit diverse
28 physiological activities that are beneficial to the body's functions. Typically, bioactive peptides range
29 in size from 2 to 20 amino acid residues and have smaller molecular weights compared to proteins,
30 but their bioactivity is often greater than that of proteins (1). Traditional protein digestion theory
31 suggests that proteins can only be absorbed and utilized after being broken down into amino acids
32 upon entering the body (2). However, recent studies have demonstrated that small-molecule peptides
33 are absorbed more readily than proteins. Absorption channels for bioactive peptides exist in the small
34 intestine, allowing these peptides to be directly absorbed and utilized by the body, with an absorption

35 rate that surpasses that of proteins and amino acids. The bioactivity of bioactive peptides is reflected
36 in various aspects, exhibiting regulatory functions such as antihypertensive, antihyperlipidemic,
37 antihyperglycemic, anti-cholesterol, antiviral, and anticancer effects (3).

38 Although bioactive peptides have the potential to become functional foods and even drugs, their
39 low bioavailability and low activity caused by oral administration are an urgent problem to be solved.
40 The biological activity of a bioactive peptide depends largely on its chemical structure, including
41 amino acid composition, molecular weight, amino acid sequence, and peptide spatial conformation
42 (4). Oral administration of bioactive peptides need to overcome multiple barriers (such as complex
43 enzymatic decomposition in the gastrointestinal tract, changes in pH, adsorption of small intestinal
44 mucus, obstruction of small intestinal mucosal cells, etc.) before they can be absorbed and utilized by
45 the human body. These barriers may cause changes in the sequence and spatial structure of bioactive
46 peptides, resulting in the loss of biological activity of bioactive peptides. Furthermore, these barriers
47 can hinder the absorption and utilization of bioactive peptides, significantly decreasing the amount
48 that enters systemic circulation and performs biological functions in targeted areas.

49 Currently, various strategies have been developed to enhance the bioavailability of bioactive
50 peptides in the human body. These strategies include chemical structure modifications, permeation
51 enhancers, and colloidal delivery systems, such as liposomes, emulsions, biopolymer nanoparticles,
52 and hydrogels. Each of these approaches has its own advantages and disadvantages. For instance,
53 chemical modifications can significantly improve the stability of bioactive peptides; however, they
54 may alter the original chemical structure of the peptides, potentially affecting their biological activity
55 and even leading to the production of harmful substances (9). Although intestinal permeation
56 enhancers (PEs) show good absorption-promoting effects, excessive use can compromise the
57 integrity of the intestinal barrier, and the stability of permeation enhancers in the gastrointestinal tract
58 also requires careful consideration by researchers (10). Encapsulating bioactive peptides using
59 colloidal delivery systems is considered the most promising approach, as it can mask bitterness and
60 overcome many challenges encountered during oral administration, but there are still some problems
61 such as low encapsulation efficiency, poor stability, and poor targeting(11).

62 In summary, improving the bioavailability of orally delivered bioactive peptides requires a
63 thorough analysis of the advantages and limitations of current delivery strategies. Unfortunately, to
64 date, there remains a lack of systematic collation and comprehensive reviews addressing these issues
65 in the relevant literature. Therefore, this review comprehensively examines the challenges associated
66 with the oral delivery of bioactive peptides, introduces the advantages and disadvantages of existing
67 oral delivery systems, and summarizes the future development trends of these systems. The aim of
68 this review is to provide a valuable reference for subsequent studies on bioactive peptide delivery
69 systems through this in-depth analysis.

70 **2 Obstacles to oral administration of bioactive**

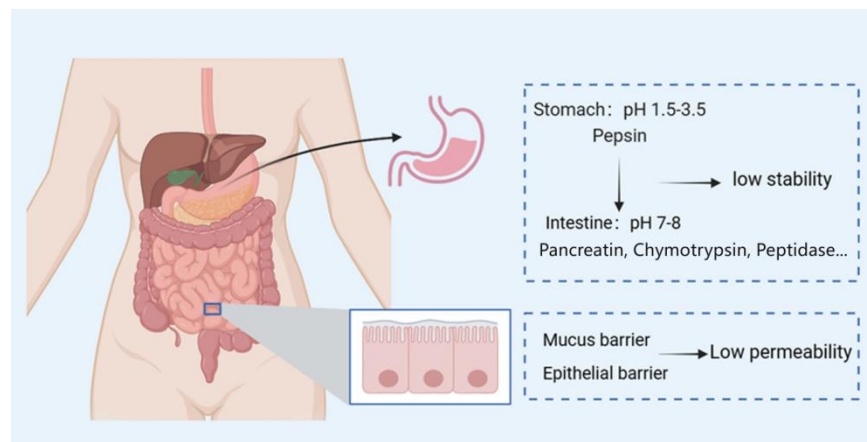
71 The oral delivery of bioactive peptides presents several challenges. First, some bioactive
72 peptides may possess a pronounced bitter taste, which can significantly impact patients' acceptance of
73 oral administration. Second, the digestive tolerance of bioactive peptides within the gastrointestinal
74 tract poses another major challenge for their oral delivery. The variable pH gradient and the complex
75 digestive enzyme system of the gastrointestinal tract can severely affect both the structural integrity
76 and the functional stability of bioactive peptides. Additionally, the intricate defense system formed
77 by the mucus layer, epithelial cells, and microbial community in the gastrointestinal tract is a critical
78 factor limiting the oral bioavailability of these peptides. Furthermore, the unique physicochemical

79 and structural properties of bioactive peptides can also significantly influence their efficacy in oral
80 delivery.

81 2.1 Bitter taste barrier.

82 Bioactive peptides from natural sources are very limited, so most bioactive peptides are
83 produced by enzymatic hydrolysis of proteins. However, proteolysis can not only produce
84 biologically active peptides, but also produce some peptides with a pronounced bitter taste. Generally
85 speaking, bitter taste in food products is not accepted by consumers. The bitterness produced by the
86 hydrolysis process limits the application of active peptides in the food industry, so how to reduce the
87 bitterness is an extremely important issue. The bitter taste of peptides is related to hydrophobic amino
88 acids (12) and their relative molecular masses (13). As early as 1997, Henriksen (14) extracted
89 peptides with molecular weights less than 4000 Da from dried sausages, graded the extracts for
90 sensory evaluation, and found that the higher the intensity of bitterness, the higher the concentration
91 of hydrophobic amino acids in the extracts. Myong et al. (15) extracted bitter peptides from
92 commercially available soy protein hydrolysates. The analysis showed that the bitterness of soy
93 peptides was mainly associated with the presence of medium molecular weight peptides in the range
94 of 1000–4000 Da, and the bitterness of peptide fractions less than 1000 Da was lower than that of
95 high molecular weight fractions.

96 Since the middle of the 20th century, the research on the removal of the bitterness of short
97 protein peptides has gradually increased, and the most common method is masking. Bertelsen et al.
98 (16) used a variety of masking agents for removing bitterness from soy protein hydrolysates, among
99 which xylitol, sucrose, and α -maltodextrin had significant debittering effects. In addition, bitterness
100 can also be removed by destroying the structure of bitter peptides by enzymatic hydrolysis (17),
101 which is widely used in industry because of its high efficiency and no loss of nitrogen. Lei et al. (18)
102 used aminopeptidase to hydrolyze soybean protein isolate with a bitterness value of 3.6 to reduce its
103 bitterness value to 0.4. reducing its bitterness value to 0.4. It is worth noting that the plastein reaction,
104 the reaction in which protease promotes the formation of a gel-like substance from high-
105 concentration protein hydrolyzate under suitable conditions, is an effective debittering method
106 (19,20). Peptide condensation during plastein reactions can help reduce the bitterness intensity of
107 polypeptides. However, the plastein reaction is not yet applied in industry and needs further
108 exploration.



109

110

Figure 1. Gastrointestinal disorders affecting peptide absorption.

111 2.2 Barriers of orally administered bioactive peptides in the gastrointestinal tract.

112 **Biochemical barrier.** Two major types of biochemical barriers exist for orally administered
113 peptides: variable pH and gastrointestinal proteases (Figure 1). Orally administered bioactive
114 peptides travel through the oral cavity to the stomach, then to the duodenum, jejunum, ileum, and
115 finally to the colon and rectum (21). Although digestion begins in the oral cavity, due to the
116 extremely short oral action time, the oral cavity not typically cited as a major factor hindering the
117 absorption and utilization of orally administered bioactive peptides. The main factors affecting the
118 absorption and utilization of oral bioactive peptides mainly come from the stomach and small
119 intestine. The first thing to overcome when taking bioactive peptides orally is the variable pH of the
120 gastrointestinal tract. The pH value of gastric juice is 1.5-3.5, that of the duodenum is about 5-6, and
121 that of the jejunum and terminal ileum rises to 7-8 (22). Variable pH gradients have a great impact on
122 the physiological efficacy of some bioactive peptides. The antioxidant activity of the pentapeptide
123 ATSHH from whitefish protein will show a significant decrease trend under acidic conditions (pH=2)
124 (23).

125 In addition, after the bioactive peptides reach the stomach, they will stimulate the gastric
126 mucosa to secrete pepsin from the gastric lining cells. Pepsin can hydrolyze the polypeptide with
127 aromatic residues such as phenylalanine, tryptophan, and tyrosine. Bioactive peptides hydrolyzed by
128 pepsin will lose their inherent biological activity. After the bioactive peptide enters the small
129 intestine through the stomach, the trypsin and chymotrypsin present in the small intestine will also
130 specifically hydrolyze the peptide chain (24). The hydrolysis of the above enzymes will change the
131 structure and activity of the bioactive peptide. Li et al. (25) performed in vitro simulated digestion
132 experiments on rice protein hydrolyzate and found that the anti-hypertensive IC₅₀ (half maximal
133 inhibitory concentration) value of rice protein increased from 140 to 180 µg/mL in the presence of
134 digestive enzymes (pepsin and pancreatic enzymes), indicating that the anti-hypertensive activity of
135 rice protein hydrolyzate was significantly reduced. In addition, after the bioactive peptides reach the
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145 that the anti-hypertensive activity of rice protein hydrolyzate was significantly reduced.

146 **Mucus and epithelial barrier.** After bioactive peptides are digested in the stomach and
147 successfully reach the small intestine, the intestinal mucus layer covering the intestinal surface is one
148 of the main factors limiting the bioavailability of oral bioactive peptides. The intestinal mucus layer
149 is a kind of intelligent hydrogel with high viscoelasticity and adhesiveness, which contains highly
150 branched polysaccharides and negatively charged mucin (26). The intestinal mucus layer plays a
151 protective role by forming a sieve-like structure on itself. This structure can effectively prevent 10-
152 200 nm particles from passing through the mesh, and has the function of selectively transmitting
153 nutrients (27). Mucin, glycolipids, and glycoproteins in the mucus layer act as both barriers and
154 transmit signals (28). When bioactive peptides reach the intestinal mucus layer, their further diffusion
155 may be affected by mucin adhesion.

156 After bioactive peptides pass through the mucus layer and reach the surface of epithelial cells,
157 the epithelial cells located under the mucus are another major factor limiting the bioavailability of
158 oral bioactive peptides. The small intestine epithelial cells are a continuous monolayer that separates
159 the intestinal lumen from the underlying lamina propria. There is a tight junction (TJ) between
160 adjacent epithelial cells, which only allows small molecules such as water and ions to pass through.
161 In addition, the small intestine cell membrane acts as a barrier to prevent extracellular substances
162 from freely entering and exiting the cells by selectively absorbing nutrients (29). Based on the above
163 reasons, the small intestinal epithelium is impermeable. Bioactive peptides need to pass through the
164 TJ or intestinal epithelial cell membrane to reach the bloodstream and ultimately bind to the target to
165 exert physiological activity. However, most bioactive peptides cannot effectively penetrate intestinal
166 epithelial cells due to the lack of targeted carrier proteins on the intestinal epithelial cell membrane,
167 which seriously affects the bioavailability of bioactive peptides.

168 **2.3 Physical and chemical properties of peptides.**

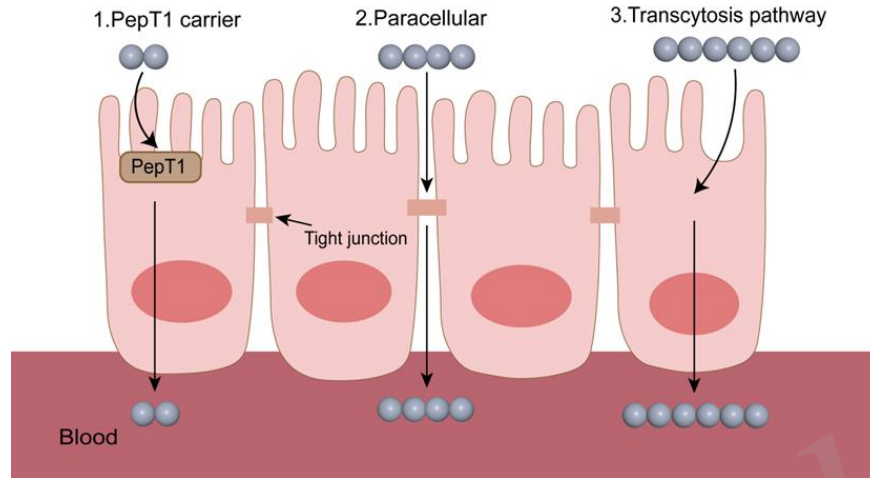
169 The physicochemical properties of peptides are one of the important factors affecting the
170 bioavailability of orally delivered active peptides. The molecular weight and structural characteristics
171 of the peptides can affect their absorption. Compared with short-chain peptides with smaller
172 molecular weights, long-chain peptides are more sensitive to gastrointestinal proteases, which results
173 in long-chain peptides being more easily degraded and absorbed by gastrointestinal digestive
174 enzymes (30). Research by Chen and Li (31) showed that the stability of casein-derived peptides with
175 different molecular weights varies in simulated gastrointestinal tracts. Peptides with a molecular
176 weight greater than 3 kDa are more likely to be degraded during gastric digestion than peptides with
177 molecular weights less than 3 kDa (31). In addition, studies have shown that some short peptides
178 with smaller molecular weights can be transported across intestinal cells through peptide transporters
179 expressed in the intestine, while oligopeptides can be passively transported and absorbed into the
180 body through hydrophobic regions or tight junctions of membrane epithelial cells (32). However,
181 long-chain peptides typically need to be absorbed through endocytosis. Therefore, short-chain
182 peptides are more easily absorbed and utilized by the body.

183 In addition, the structural characteristics of peptides also play a crucial role in the stability of
184 oral bioactive peptides. The amino acids sequence and structure of bioactive peptides can affect the
185 stability of peptides during digestion, thereby affecting their bioavailability. Savoie et al. (33) found
186 that high levels of proline and glutamic acid in peptide sequences can enhance the resistance of
187 peptides to pepsin and trypsin. Udenigwe (34) research showed that bioactive peptides with a higher
188 β -sheet structure ratio are more sensitive to heat treatment. In addition, the charge of the peptide has
189 been shown to affect the transport of peptides. For example, peptides with neutral amino acid
190 residues can be preferentially recognized by oligopeptide transporter 1 (PepT1) (35). PepT1 is a
191 transporter protein present on the brush like border membrane of the small intestine epithelium. The
192 research of Wang and Li (36) showed that in addition to PepT1 mediated transport pathway,
193 bioactive peptides can also cross small intestinal epithelial cells through endocytic transport and
194 paracellular transport. For example, positively charged hydrophobic antioxidant casein peptides can
195 be transported via endocytosis, whereas negatively charged hydrophilic peptides need to be
196 transported via paracellular pathways.

197 **2.4 Absorption mechanism of peptides.**

198 After successfully overcoming multiple obstacles such as the variable pH environment of the
199 gastrointestinal tract, enzymatic hydrolysis by gastrointestinal digestive enzymes, and adhesion/pre-
200 cleavage of the intestinal mucus layer, bioactive peptides still need to overcome the obstruction of the

201 small intestinal epithelial cells to enter the blood circulation system, which is the prerequisite for the
 202 physiological functions of bioactive peptides in vivo. There are three main modes of transmembrane
 203 transport of bioactive peptides (Figure 2): vector transport, cell bypass transport, and endocytosis
 204 transport (37).



205

206

Figure 2. Several transmembrane transport pathways of bioactive peptides.

207 **Carrier-mediated transport pathway.** The carrier-mediated transport pathway primarily relies
 208 on oligopeptide transporters (38). The important feature of transporters is that they can select
 209 peptides. Transporters have been found to recognize and transport over 8,000 different peptides (39).
 210 There are two main types of transporters: PepT1 and PepT2. Both PepT1 and PepT2 can be used for
 211 the transport of dipeptides and tripeptides (40). Currently, there are more studies on PepT1 than
 212 PepT2 on the transport of polypeptides. PepT1 is mainly expressed in intestinal epithelial cells and is
 213 responsible for the transport and absorption of bioactive peptides. As mentioned in the section on the
 214 physicochemical properties of peptides, the charge of peptides affects the mode of transport, and
 215 PepT1 preferentially recognizes peptides with neutral charge and high hydrophobicity, and
 216 preferentially binds residues rich in non-polar amino acids. Fan et al. (41) studied the transport
 217 modes of IW, IWH, and IWHHT peptides in Caco-2 cells, which further verified that PepT1
 218 preferred to select small peptides with high hydrophobicity. **Table 1 summarizes the transport**
 219 **pathways of different bioactive peptides through the Caco-2 cell model, aiming to provide a solid**
 220 **experimental basis for subsequent research and product development.**

221

Table 1 Transport pathway of bioactive peptides through Caco-2 cells

| Bioactive Peptides | Function | Source | Transport pathways | Ref |
|--------------------|------------------------------------|-----------------------------|--------------------|------|
| IRW | Anti-hypertensive, Anti-oxidant | Ovotransferrin | PepT1, TJs | (42) |
| IPP, LKP | Anti-hypertensive | Bovine milk β -casein | PepT1, TJs | (43) |
| VPP | Anti-hypertensive | Fermented milk | TJs | (44) |
| IQW | Anti-hypertensive | Ovotransferrin | PepT1, TJs | (45) |

| | | | | |
|-------------------------|--------------------|----------------------------------|-------------------|------|
| | Anti-hypertensive | | | |
| LSW | | Soybean protein | PepT1, TJs | (46) |
| | Anti-inflammatory | | | |
| YPI | Anti-hypertensive | Whey protein | PepT1 | (47) |
| IW | Anti-hypertensive | Myogenic fibers of hens | PepT1 | (41) |
| IWH | Anti-hypertensive | Myogenic fibers of hens | PepT1, TJs | (41) |
| IWHHT | Anti-hypertensive | Myogenic fibers of hens | TJs | (41) |
| RVPSL | Anti-hypertensive | Ovotransferrin | TJs | (48) |
| VLPVP | Anti-hypertensive | Genetic engineering isolation | TJs | (49) |
| HLPLP | Anti-hypertensive | β -casein | TJs | (50) |
| VY | Anti-hyperglycemic | Black bean sauce | PepT1, TJs | (51) |
| VPLVM | Anti-hyperglycemic | Broccoli | PepT1, TJs | (52) |
| LPEW | Anti-hypertensive | Fermented milk | Transcytosis | (53) |
| GLLLPH | Anti-oxidant | Corn Gluten | TJs, Transcytosis | (54) |
| YFCLT | Anti-oxidant | Corn Gluten | TJs, Transcytosis | (54) |
| LAPSLPKPKPD | Anti-hypertensive | Egg yolk protein | Transcytosis | (55) |
| β -casein 193-209 | Immunomodulatory | Bovine milk β -casein | Transcytosis | (63) |
| YWDHNNPQIR | Anti-oxidant | Canola protein | Transcytosis | (64) |

222 **Paracellular transport pathway.** The paracellular transport pathway is currently the most
 223 reported passive absorption pathway for bioactive peptides with more than tripeptides (56). The
 224 driving force for oligopeptide transport comes from the electrochemical gradient formed by protons
 225 as high-energy electrons are transferred along the respiratory chain, and the diffusion process does
 226 not require a carrier or energy consumption (57). The paracellular transport pathway is mediated
 227 through the TJ between epithelial cells, a tight biological barrier with selective permeability (58). It
 228 has been shown that TJ tends to transport negatively charged peptides and is selective for positively
 229 charged peptides (59), and bioactive peptides with small hydrophilic molecular weights are more
 230 inclined to this transport mode (60). In general, when the molecular diameter of a bioactive peptide
 231 exceeds 15 Å, the peptide cannot undergo paracellular transport. However, it is still possible for
 232 bioactive peptides with larger molecular sizes to diffuse through TJ if their structures have high
 233 conformational flexibility (61). **Chiasma has successfully developed an oral formulation of octreotide,**
 234 **named Mycapssa®, utilizing its innovative Transient Permeation Enhancer (TPE™) technology. In**
 235 **this approach, sodium caprate serves as an osmotic enhancer, inducing the reversible opening of tight**
 236 **junctions between intestinal epithelial cells to facilitate the paracellular transport of peptides. The**
 237 **successful development of Mycapssa® not only strongly confirms the feasibility of the paracellular**
 238 **transport strategy for the oral delivery of peptide drugs but also paves the way for further research**
 239 **into the oral delivery of bioactive peptides (62).**

240 **Endocytic transport pathway.** Endocytic transport is an energy-dependent transcellular
 241 transport pathway and is the main transport pathway for long-chain peptides. In this pathway,
 242 bioactive peptides are transported into cells through the formation of vesicles formed by invagination
 243 of the cell membrane (63). Bioactive peptides with smaller molecules can enter the blood circulation
 244 through carrier transport and paracellular pathways, while most large molecule peptides need to be

245 transported through endocytosis. The study by Regazzo et al. (64) showed that 17-peptide (casein
 246 193-209) can be completely absorbed by the Caco-2 cell monolayer model, and its absorption
 247 process is mainly carried out through endocytosis transport. The first step in endocytic transport is
 248 the interaction of polypeptides with the cell membranes. Since the cell membrane is composed of a
 249 lipid bilayer, endocytic transport is considered an ideal pathway for the transport of lipophilic
 250 peptides. The anti-oxidant peptide YWDHNNPQIR is transported across the Caco-2 cell monolayer
 251 via endocytosis, primarily because it is composed of hydrophobic amino acids (65). **Xiao et al. have**
 252 **innovatively designed and prepared a hybrid liposome system named mExos@DSPE-Hyd-PMPC.**
 253 **This system significantly improves drug encapsulation efficiency and enhances endocytic transport**
 254 **efficacy by effectively integrating functional liposomes with milk-derived exosomes (mExos).**
 255 **Notably, this hybrid liposome exhibits adaptive surface characteristics, enabling it to intelligently**
 256 **adjust its physicochemical properties based on the pH microenvironment of the intestinal mucosal**
 257 **surface. This adaptability facilitates a more efficient endocytic transport process (66).**

258 **Notably, research has demonstrated that the hydrophilicity and charge state of bioactive peptides**
 259 **play a significant role in their transport within the body (67). The charge can influence the**
 260 **interactions of bioactive peptides with cell membranes, transport carriers, and other molecules in the**
 261 **gastrointestinal environment. Table 2 summarizes the relationship between various transport**
 262 **mechanisms and the properties of peptides. However, it is important to emphasize that hydrophilicity**
 263 **and charge state are not the only factors determining the transport pathways of bioactive peptides.**
 264 **The transport pathways are also influenced by several other factors, including molecular weight,**
 265 **peptide structure, hydrophobicity, the gastrointestinal environment, and the selection of transport**
 266 **carriers.**

267 Table 2 Relationship between different transport modes and peptide properties

| Characteristics Transport pathways | Peptide molecular size | Water affinity | Electric charge |
|---------------------------------------|-------------------------|-------------------|--------------------------|
| PepT1 | Dipeptide or tripeptide | Hydrophobic | Neutral charge |
| TJs | Short-chain peptides | Hydrophilic | Negative charge |
| Transcytosis | Long-chain peptides | Hydrophobic | Positive electric charge |

268 3 Oral delivery systems for bioactive peptides

269 As mentioned above, the oral administration of bioactive peptides encounters numerous barriers
 270 in the human body, which significantly diminish their bioavailability. Therefore, the development of
 271 effective oral delivery systems to enhance the bioavailability of bioactive peptides is imperative. An
 272 ideal oral delivery system should ensure that the bioactive peptide maintains its integrity before
 273 reaching the site of absorption and promotes targeted release at the desired site of absorption.
 274 Currently, several prominent oral delivery technologies have been extensively studied and applied to
 275 overcome the barriers associated with bioactive peptides delivery in the human body. These oral
 276 delivery technologies include permeation promotion technologies, chemical structural modifications,
 277 colloidal delivery systems, etc.

278 3.1 Permeation promotion technology.

279 One of the biggest obstacles to oral administration of bioactive peptides is the poor permeability
 280 of intestinal epithelial cells to bioactive peptides. Permeation enhancers (PEs) are substances that can

281 temporarily increase intestinal permeability and promote the penetration of bioactive peptides
 282 through the intestinal epithelium (65). Currently, over 250 substances have been investigated in
 283 clinical research as PEs for the oral delivery of bioactive peptides, such as surfactants, fatty acids,
 284 bile salts, and cell-penetrating peptides (68).Based on their mechanisms of action, PEs are mainly
 285 divided into two categories (69).The first category mainly acts on the TJ between epithelial cells and
 286 achieves paracellular transport of bioactive peptides by opening the TJ between epithelial cells. The
 287 second category is to promote the transmembrane transport of bioactive peptides by increasing the
 288 permeability of the cell membrane. Table 3 lists some typical PEs and their respective mechanisms of
 289 action. It is worth noting that some specific PEs can act on both pathways at the same time, such as
 290 sodium decanoate, bile salts and chitosan. In addition, although PEs are generally considered safe
 291 and non-toxic, the additive dosage of PEs still needs to be strictly controlled when using them.
 292 Excessive use of PES can cause excessive changes in the permeability of intestinal epithelial cells,
 293 which will eventually induce local inflammation or long-term damage to intestinal epithelium
 294 (75).For example, calcium chelators can cause Ca^{2+} depletion in the body, thereby damaging actin
 295 filaments, altering adherens junctions and reducing cell adhesion (76).

296 Table 3 Typical PEs for three different mechanisms

| Categories | Mechanism | PEs | Ref |
|------------|---|-------------------------|------|
| 1 | Opens the paracellular pathway to facilitate transcellular transport | EDTA | (70) |
| | | Citric Acid | (70) |
| 2 | Increasing cell membrane permeability to facilitate transcellular transport | SNAC | (71) |
| | | Bile salts | (72) |
| 3 | Simultaneous enhancement of both pathways | Sodium Caprate (C10) | (73) |
| | | Chitosan | (74) |

297 Cell-penetrating peptides (CPPs), as an important branch of penetration enhancers, are mainly
 298 polypeptides ranging from 5 to 30 amino acids, which transport bioactive peptides across the
 299 membrane by penetrating the cell membrane or endocytosis (77). Currently, researchers have
 300 designed or identified more than 100 peptides that can effectively promote the transport of biological
 301 macromolecules across cell membranes. In practical applications, nucleotides, bioactive peptides, and
 302 other biologically active substances are prone to lose their activity in the systemic circulation.
 303 Encapsulating such substances in nanoparticles can greatly enhance their stability in vivo. However,
 304 the presence of the cell membrane hinders the uptake of bioactive substances by target cells. CPPs
 305 provide researchers with a new direction of exploration. Studies have shown that combining CPPs
 306 with nanoparticles can further enhance the transcellular delivery of bioactive peptides and effectively
 307 improve the uptake of bioactive substances by target cells. Knoll et al. (78) developed a new type of
 308 CPP-modified nanostructured lipid-based carrier, and experimental results demonstrated that this
 309 new type of coated nanocarrier can improve the uptake of bioactive substances by cells. The in vivo
 310 toxicity of CPPs is not yet fully understood, but a small number of published animal studies and
 311 several CPP formulations approved for clinical trials demonstrate the general safety profile of CPP
 312 molecules at study doses (79). Nevertheless, no CPP-encapsulated drugs have entered clinical trials,
 313 and further research is needed to evaluate their in vivo delivery effects.

314 **3.2 Chemical structural modifications.**

315 Bioactive peptides are a type of molecules that are relatively easy to modify in chemical
316 structure. Chemical modification can significantly improve the stability of bioactive peptides. The
317 more commonly used chemical modification methods are PEGylation and cyclization (80).
318 PEGylation is a chemical modification technique that involves the covalent attachment of
319 polyethylene glycol (PEG) molecules to biological macromolecules, such as proteins and peptides.
320 This process aims to optimize the physicochemical properties and biological characteristics of these
321 biomolecules. For bioactive peptides, the incorporation of PEG can significantly enhance their water
322 solubility, thereby improving their solubility in physiological environments, which is essential for
323 effective absorption and distribution. Furthermore, PEG, being an inert polymer, effectively protects
324 peptide drugs from enzymatic degradation, leading to a substantial increase in the retention rate and
325 bioavailability of bioactive peptides. Additionally, the increase in molecular weight resulting from
326 PEGylation reduces the renal clearance rate of peptide drugs, thereby prolonging their half-life in the
327 body and decreasing the frequency of administration (81). Zhou et al. (82) demonstrated that when
328 the HM-3 peptide was modified with methoxy-PEG-aldehyde, its half-time was extended by 5.86
329 times in male SD rats. Wang et al. (83) similarly showed that after pegylation, the CPU-HM peptide
330 exhibited higher in vivo activity and a longer half-time.

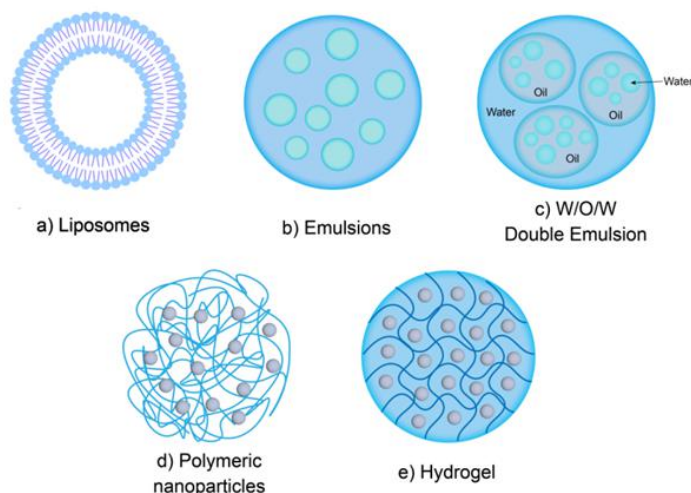
331 Cyclization is another commonly used method for chemical modification of bioactive peptides.
332 By creating a cyclic structure, cyclization eliminates the exposed N- and C-terminals in peptide
333 molecules, rendering them less susceptible to enzymatic degradation (84). Desmopressin is an
334 analogue obtained by cyclization of vasopressin, which is more resistant to enzymatic degradation
335 than vasopressin (85). Similarly, cyclized opioids exhibit longer half-life and higher metabolic
336 stability (86). In addition, cyclic structural peptides have better permeability than linear structural
337 peptides. The cyclic structure is more compact than the linear structure, which reduces the collision
338 of the cyclic structure peptide in the solution and ultimately allows it to pass through the epithelial
339 barrier faster (87).

340 In addition to debittering, the plastein reaction mentioned above also provides a feasible method
341 for the modification of peptides. Studies have shown that plastein reactions can enhance the activity
342 of angiotensin-converting enzyme (ACE) inhibitory peptides. Song et al. (88) used plastein reactions
343 to modify hazelnut peptides, and the results showed that the ACE inhibition rate of the modified
344 products was significantly improved. Similarly, Jiang et al. (89) employed plastein reaction to modify
345 ACE inhibitory peptides derived from sea cucumbers, and found that the modified peptide showed
346 significantly enhanced thermal stability, and the thermal transition temperature of the modified
347 peptide increased from 120°C to 134°C. These studies indicate that plastein reaction is a promising
348 strategy to induce structural modifications to improve the biological activity of peptides. However,
349 the application of plastein reactions in peptide modification is not immature at present, and research
350 on peptide sequence changes after plastein reactions is relatively limited. Regardless, when
351 modifying the chemical structure of bioactive peptides to improve their bioavailability, it is necessary
352 to pay attention that the modification process cannot affect the original functions of the bioactive
353 peptides and to avoid the generation of harmful substances.

354 **3.3 Colloidal delivery system.**

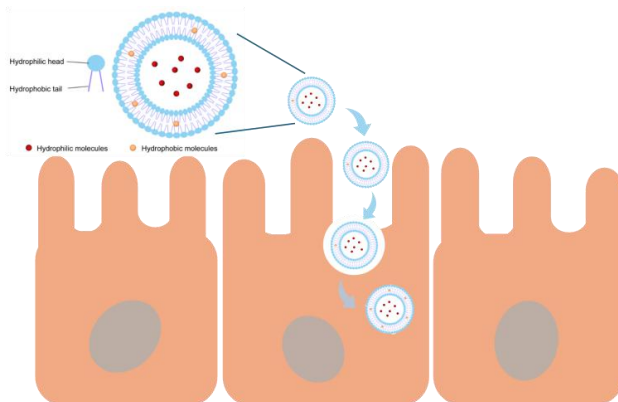
355 Due to the susceptibility of bioactive peptides to loss of physiological activity under different
356 pH values and the action of digestive enzymes in the body, using a delivery system to encapsulate
357 bioactive peptides can effectively eliminate the bitter taste while improving the stability of peptides
358 in systemic circulation. Colloidal delivery systems have been widely applied in the delivery of

359 bioactive peptides. Common colloidal delivery systems include liposomes, emulsions, polymer
360 nanoparticles, and hydrogels, as illustrated in Figure 3.



361
362 **Figure 3.** Colloidal delivery system structure.

363 **Liposomes.** Liposomes are a kind of spherical closed vesicle formed by concentric phospholipid
364 molecules linked end to end through hydrophobic interactions, which can protect the loaded materials
365 from being broken down by enzymes and improve their bioavailability in the body (Figure 3a) (90).
366 Gong et al. (91) the bioavailability of peanut peptides was effectively improved after being
367 encapsulated in nanoliposomes. The main reason is that the nanoliposomes prepared in this study
368 exhibited good stability under different pH conditions and different morphologies, which allows the
369 peanut peptides encapsulated in the nanoliposomes to retain a relatively complete structure and high
370 ACE inhibitory activity. Compared with other delivery systems, liposomes have the advantages of
371 easy encapsulation, large encapsulation capacity, and minimal residual organic solvents. Liposomes
372 can encapsulate both hydrophobic and hydrophilic bioactive peptides. Hydrophobic peptides can be
373 embedded within the phospholipid bilayer, while hydrophilic peptides can be encapsulated in the
374 aqueous core (92).



375
376 **Figure 4.** Liposomes deliver substances into cells through vesicle-based delivery.

377 However, liposomes also have some limitations. Firstly, The phospholipid membrane of
378 liposomes is sensitive to adverse factors such as high temperature, enzymes, and ionic strength.
379 These adverse factors may cause the liposomes to decompose during storage or before reaching the
380 small intestine, causing the bioactive peptides wrapped inside to leak out in advance (93). To
381 overcome this limitation, researchers have found that surface modification of liposomes with
382 polymers such as chitosan, pectin, and polyethylene glycol can effectively improve the stability and

383 sustained release ability of liposomes (94). Ramezanzade et al. (95) developed a novel composite
384 nano-carrier of triphosphorus sodium cross-linked chitosan coated liposomes, and differential
385 scanning calorimetry showed that this composite nano-carrier had better thermal stability than
386 ordinary liposomes. Wu et al. (96) used sodium alginate (SA) to coated liposomes containing DPP-
387 IV inhibitory collagen peptides and found that compared with uncoated liposomes, SA-coated
388 collagen peptide liposomes exhibited higher storage stability, gastrointestinal stability and
389 transcellular permeability. Secondly, due to the large size structure of liposomes, they may not be
390 absorbed by intestinal epithelial cells, and the penetration mechanism of liposomes is not yet clear.
391 Therefor the best approach is to choose vesicles as small as possible for the delivery of active
392 substances, with particle diameters below 100-200 nm (Figure 4) (97). Additionally, cationic charged
393 liposomes are often chosen to deliver bioactive substances because they are more easily attracted to
394 the negatively charged mucus layer. Cuomo et al. (98) employed liposomes for the oral delivery of
395 all-trans-retinoic acid and observed that cationic liposomes could interact with saliva in the oral
396 cavity, which carries a net negative charge. Importantly, when cationic liposomes were coated with
397 mucoproteins from oral saliva, the charge on the cationic surface interaction changed from positive to
398 negative. This prevented the liposomes from being attracted to the negatively charged mucus layer
399 during other stages of digestion, providing further protection for the loaded molecules.

400 **Emulsion.** An emulsion is a thermodynamically unstable colloidal dispersion formed by two
401 immiscible liquids (usually oil and water), in which one liquid is dispersed as small droplets in the
402 other liquid (99). According to their structural characteristics, emulsions can be divided into single-
403 layer emulsions (water-in-oil, oil-in-water) (Figure 3b) and multi-layer emulsions (water-in-oil-in-
404 water, oil-in-water-in-oil) (Figure 3c) (100). As a complex multi-phase system, multi-layer emulsion
405 has various system types, among which W1/O/W2 is the most commonly used in food. The main
406 structural state of W1/O/W2 type emulsions is that small water droplets (internal water phase, W1
407 phase) are trapped in larger oil droplets, and are subsequently dispersed in the external water phase
408 (W2 phase). Multi-layer emulsions are complex multiphase systems. W1/O/W2 type is more
409 common in food, where small water droplets (inner aqueous phase, W1 phase) are trapped in larger
410 oil droplets, which are then dispersed in the outer aqueous phase (W2 phase) (101). Like singlelayer
411 emulsions, the formation of multilayer emulsions also requires the addition of emulsifiers. Previous
412 studies have found that the type of emulsifier can affect the stability of multilayer emulsions. Yeon-Ji
413 Jo et al. (102) found that the hydrophilic and lipophilic balance value of the emulsifier can
414 significantly affect the stability of W1/O/W2 emulsion loaded with collagen peptides, and emulsifiers
415 with significant amphiphilicity can make W1/O/W2 emulsion more stable. Ying et al. (103) used
416 polyglycerol ricinoleate and modified starch as emulsifiers to successfully prepare an emulsion
417 system with a soybean peptide encapsulation rate of more than 80%. The results of in vitro simulated
418 gastrointestinal digestion showed that the emulsion system showed strong resistance to the
419 decomposition of pepsin, and the retention rate of soybean peptide was higher than 70% after
420 simulated gastric digestion. In some cases, even with the addition of emulsifiers, the properties of
421 multilayer emulsions are still not stable enough. This is because the system has two interfaces with a
422 large interfacial area, making the multiphase structure prone to destruction during storage (104).
423 Currently, there are various methods to stabilize the structure of multiple emulsions. One effective
424 method to improve the stability of multiple emulsions is to add proteins or polysaccharides to limit
425 the movement of components. For example, the addition of gelatin to multiple emulsions could
426 significantly improves their stability (105). Furthermore, studies have shown that emulsion delivery
427 systems not only improve the gastrointestinal stability of peptides, but also have the characteristics of
428 masking the bitter taste of bioactive peptides (102). Gao et al. (106) used water-in-oil high internal
429 phase emulsions (W/O HIPE) to encapsulate bitter peptides and found that W/O HIPE had a
430 significant masking effect on the bitter taste of peptides.

431 Although both single-layer emulsions and multi-layer emulsions need to be stabilized by adding
432 emulsifiers, some synthetic low molecular weight surfactants still need to be considered for their
433 potential harm to the human body (107). Specifically, surfactants with a high HLB (Hydrophilic-
434 Lipophilic Balance) value may disrupt the skin barrier due to their strong interfacial activity, which
435 can increase the skin's permeability to harmful substances, leading to skin irritation and even
436 triggering allergic reactions and skin inflammation. Secondly, during the preparation of emulsions,
437 although surfactants are renowned for their emulsifying properties, there is also a risk of causing
438 emulsion instability, such as phase separation, coalescence, or creaming. These instability
439 phenomena not only affect the appearance and texture of the product but may also compromise its
440 actual efficacy. Moreover, the interactions between surfactants and bioactive ingredients may lead to
441 structural changes in the bioactive components, resulting in the loss of their original functions, which
442 is crucial for maintaining the integrity of bioactive ingredients. Surfactants may interfere with the
443 permeability and retention time of bioactive components, thereby affecting their distribution and
444 metabolism within the organism, ultimately reducing their bioavailability and therapeutic effects
445 (107). Therefore, researchers have been on the way to seek other safer methods to stabilize the
446 emulsion structure. At this time, a special emulsion, Pickering emulsion, came into the the attention
447 of researchers. Cai et al. (108) found that the natural Pickering emulsion system formed by composite
448 nanoparticles that interacted/conjugated antimicrobial peptide Parasin I with chitosan significantly
449 improved the stability and antibacterial activity of Parasin I. The solid particles in Picorling
450 emulsions are irreversibly adsorbed on the surface of the emulsion droplets and play a role in
451 stabilizing the emulsion system. This characteristic of Picorling emulsion avoids the use of
452 surfactants, so its advantage is that there is no need to consider the safety of surfactants in food
453 systems (109). In view of the characteristics and high safety of Pickering emulsions, it has a large
454 application space in the field of bioactive substance delivery, but its specific mechanism of action
455 and application characteristics still require further extensive research.

456 **Polymer nanoparticles.** Polymer nanoparticles are solid colloidal particles with an average
457 particle size ranging from 10 to 1000 nm (Figure 3d). Polymer nanoparticle delivery system is a kind
458 of system that uses natural, semi-synthetic or synthetic polymer nanoparticles as delivery carriers to
459 load bioactive substances through non-covalent methods such as electrostatic adsorption,
460 hydrophobic interaction, hydrogen bonding and so on (110). Compared to lipid-based carriers and
461 emulsions, polymer nanoparticles have a simple preparation process, smaller system size, better
462 stability which can protect bioactive peptides from being decomposed in harsh gastrointestinal
463 environments (111), thereby improving the oral bioavailability of bioactive peptides. Additionally,
464 high lipid intake may induce obesity and cardiovascular diseases (112), while the commonly used
465 materials of polymer nanoparticles are proteins, polysaccharides and their composite derivatives,
466 such as gelatin, sodium alginate, chitosan, and their derivatives, etc. Thus, polymer nanoparticles are
467 more healthier and easier to be accepted by consumers. Currently, various polymer nanoparticle
468 delivery systems have been designed and applied to bioactive peptides delivery. Zhu et al. (113) used
469 lysozyme-xanthan gum nanoparticles as carriers of selenium-containing peptides and prepared
470 lysozyme-xanthan gum-selenopeptide composite nanoparticles. In vitro release test results showed
471 that the composite nanoparticles successfully delayed the release of selenium-containing peptides and
472 improved their in vitro antioxidant activity. Uhl et al. (114) developed a surface-modified PLA
473 nanoparticles that can be loaded with liraglutide, which increased the oral bioavailability of
474 liraglutide by 4.5-fold.

475 Some polymers can reversibly open TJs between intestinal epithelial cells, help bioactive
476 peptides to be transported through the paracellular pathway, and promote the penetration and
477 absorption of bioactive peptides, such as chitosan and its derivatives (115). In addition, chitosan also

478 has good degradability and is one of the commonly used materials for constructing polymer
479 nanoparticle delivery systems (116). Auwal et al. (117) used sodium tripolyphosphate cross-linked
480 chitosan nanoparticles as the carrier to encapsulate ACE-inhibitory peptides, and found that not only
481 the physical and chemical stability of the peptides was significantly improved in vitro, but also the
482 ACE inhibitory effect of the peptides was significantly improved after simulated gastrointestinal
483 digestion. Han et al. (118) prepared a pH-sensitive complex through the electrostatic self-assembly of
484 chitosan derivative N-trimethyl chitosan, peanut peptide, and sodium alginate. This complex
485 exhibited a regular spherical shape with good stability, and the highest entrapment efficiency for
486 peanut peptide reached 91%.

487 **Hydrogel.** Hydrogel is a highly crosslinked hydrophilic polymer with a three-dimensional
488 network structure and abundant pores that can absorb and retain a large amount of water (119)(Figure
489 3e). A hydrogel system is a very effective delivery system for bioactive peptides, which can be
490 prepared by mixing bioactive peptides with a solution containing biopolymer molecules before gel
491 formation, or also by loading bioactive peptides into a microgel after microgel formation (120). Ma
492 et al. (121) developed a novel type of fish skin gelatin-based hydrogel that successfully loaded
493 codfish peptides after gel formation and exhibited good mechanical properties and biocompatibility.
494 Because different types of materials have greatly different molecular and physicochemical properties,
495 the physical and chemical differences of materials have a greater impact on the encapsulation effect
496 of the system. Therefore, when preparing hydrogels, materials need to be selected according to
497 specific purposes and applications. Protein and polysaccharide are commonly used materials for the
498 preparation of ingestible food-grade microgels. Huang et al. (122) used the emulsion template
499 method to successfully loaded ACE inhibitory peptides into biopolymer microgels composed of
500 chitosan and alginate, which effectively reduced the in vitro release rate of ACE-inhibitory peptides.
501 Ma et al. (123) used hydrogel made of alginate and chitosan to contain sericin with anti-inflammatory
502 activity, and animal experiments showed that sericin loaded by hydrogel could more effectively
503 alleviate ulcerative colitis in mice. These experimental results indicate that hydrogels have great
504 potential in oral delivery systems.

505 In addition, pH, temperature and other stimuli will lead to the morphological changes of some
506 polymer hydrogels, which will eventually lead to the phase transition of hydrogels (124). The
507 hydrogels with this phenomenon are called smart hydrogels, which can respond to environmental
508 stimuli, also known as environmentally responsive hydrogels. Environmentally responsive hydrogels
509 can make corresponding shrinkage and swelling changes when single or multiple changes occur in
510 external temperature, pH, light, electric field, salinity and other conditions, ultimately achieving
511 targeted release of bioactive peptides (125). **The environmental responsiveness of smart hydrogels
512 shows important application potential and value in the field of substance delivery. Specifically, some
513 temperature responsive smart hydrogels can exhibit different morphologies through corresponding
514 phase transitions at elevated or low temperatures depending on the ambient temperature. This
515 temperature responsiveness allows the hydrogel to adjust the position and rate of drug release in
516 response to fluctuations in body temperature or environmental temperature, resulting in precise
517 delivery of internal embedding. For example, Chuang et al. (126) cleverly designed a thermosensitive
518 hydrogel based on the fact that tumor tissue is slightly hotter than normal tissue. This hydrogel will
519 precisely undergo phase transition and release the embedded drug in the high temperature
520 environment of the tumor site, allowing effective tumor treatment with minimal drug damage to
521 normal tissues. In addition, there are some ph-responsive smart hydrogels that can adjust their
522 morphology or properties according to changes in environmental pH, a property that enables the
523 embedded material to respond to release in a specific pH environment, such as the slightly acidic
524 environment of tumor tissue or the acidic environment of the stomach. Xie et al. (127) designed a ph-**

525 sensitive hydrogel that expands and releases drugs in the acidic environment of the stomach, which
 526 could facilitate precision treatment of gastric ulcer sites. In addition to the temperature and pH
 527 response, some smart hydrogels can undergo morphological changes upon the induction of light,
 528 which are called photoresponsive hydrogels. In the treatment of skin diseases, Hu et al. (128) use
 529 photosensitive hydrogels to deliver drugs precisely to lesions, which can significantly reduce the
 530 damage of drugs to surrounding normal tissues and improve the accuracy and safety of treatment.

531 Due to their unique environmental responsiveness, smart hydrogels have the ability to precisely
 532 regulate the drug release process, which makes them show broad application prospects in the field of
 533 drug delivery. Similarly, with appropriate design and preparation strategies, smart hydrogels are also
 534 suitable for quantitative, timed, and site-directed delivery of bioactive peptides. Ye et al. (129) found
 535 that the pH-responsive carboxymethyl cellulose/polyvinyl alcohol hydrogel effectively prevented the
 536 release of soy peptides in the stomach and could basically achieve the directional release of soy
 537 peptides in the intestine. This precise delivery strategy not only enhances the retention rate of
 538 bioactive peptides but also significantly improves their bioavailability, thereby optimizing
 539 therapeutic effects. In addition, it needs to be acknowledged that although smart hydrogels can
 540 effectively control the directional release of bioactive peptides, because the human body environment
 541 is complex and changeable, the changes and safety of smart hydrogels in the body need to be further
 542 studied.

543 Another, it needs to be acknowledged that hydrogels also have some disadvantages that are
 544 difficult to avoid. Typically, hydrogels are very porous and have weak structural strength, which
 545 allows bioactive peptides (especially small peptides) to easily diffuse out of them. At present, some
 546 studies have shown that improving the capture rate of bioactive peptides by hydrogels by ensuring
 547 that the pores are small enough or enhancing the interaction between bioactive peptides and the
 548 biopolymer network within the microgel (130). Two polymers with complementary properties can
 549 form a double crosslinked hydrogel to increase the stability of the hydrogel (131). Chen et al. (132)
 550 successfully prepared strong gelatin hydrogels by dual-crosslinking gelatin with transglutaminase and
 551 carrageenan, which improved the mechanical properties and thermal stability of gelatin hydrogel. In
 552 addition, since hydrogels are mostly hydrophilic substances, they have certain limitations when
 553 embedding hydrophobic substances. Studies have found that polymerizing hydrogels with
 554 nanoparticles, micelles and cyclodextrins can significantly improve the encapsulation rate of
 555 hydrophobic substances in hydrogels. Mohammad Ali et al. (133) successfully encapsulated a β -
 556 cyclodextrin inclusion complex containing glycyrrhizic acid and thyme essential oil into alginate
 557 hydrogel beads, increasing the peptide encapsulation rate to 89%. However, there are few reports on
 558 the use of this technology in bioactive peptide entrapment, and further investigation is required. In
 559 summary, with the further development of smart hydrogel delivery systems, more innovative
 560 breakthroughs will be achieved in the application of smart hydrogels in the delivery of bioactive
 561 peptides.

562 Table 4 Advantages and disadvantages of four delivery systems

| Categories | Advantages | Disadvantages |
|------------------|----------------------|------------------------|
| Liposomes | Adjustable structure | Lack of stability |
| | Surface modifiabl | High production cost |
| Emulsion | High bioavailability | Structural instability |

| | | |
|------------------------------|--|---|
| Polymer nanoparticles | Structural stability Surface modifiable | Complex preparation Potential toxicity |
| Hydrogel | Biocompatible Controlled release | Mechanical strength |

563

564 **4 Conclusions and outlook**

565 Bioactive peptides have garnered significant attention from researchers due to their diverse
 566 physiological activities. However, the bioavailability of orally delivered bioactive peptides is
 567 severely restricted by the natural barriers of the gastrointestinal digestive system, as well as the
 568 physical and chemical properties of the peptides themselves. To enhance the stability and
 569 bioavailability of oral bioactive peptides within the gastrointestinal environment, various strategies
 570 have been explored, including chemical structure modification, the use of penetration enhancers, and
 571 colloidal delivery systems (such as liposomes, emulsions, biopolymer nanoparticles, and hydrogels).
 572 Nevertheless, each strategy presents distinct limitations in practical applications.

573 **4.1 Limitations of Delivery Strategies**

574 Although chemical modification can effectively enhance the stability of bioactive peptides,
 575 alterations in their chemical structure may reduce biological activity or even result in the formation of
 576 harmful substances. PEs possess a strong ability to promote absorption; however, inappropriate use
 577 can compromise the integrity of the intestinal barrier and significantly impact intestinal health.
 578 Liposomes, which mimic the structure of biological membranes, facilitate interactions with cell
 579 membranes, thereby offering substantial advantages in improving drug bioavailability and targeting.
 580 Nevertheless, liposomes exhibit poor structural stability and are susceptible to external factors that
 581 can lead to rupture, fusion, and leakage of their contents. Additionally, the drug loading capacity of
 582 liposomes is often suboptimal due to limitations related to molecular size, charge, and
 583 hydrophobicity. Emulsions can effectively enhance the solubility and stability of drugs, but they face
 584 challenges such as poor dispersion stability and low bioavailability. Polymeric nanoparticles have
 585 garnered considerable attention due to their controllable particle size, excellent stability, and
 586 biocompatibility. However, improvements are still needed in their drug loading capacity, drug release
 587 efficiency, and targeting capabilities. Smart hydrogels exhibit high environmental responsiveness;
 588 however, their stability within the digestive system and the controlled release of embedded materials
 589 restrict their practical applications.

590 **4.2 Future Research Trends**

591 Recent research indicates that a single delivery system is insufficient to overcome all delivery
 592 challenges. As a result, hybrid delivery systems that combine various delivery methods are
 593 anticipated to emerge as a major research focus in oral delivery moving forward. With consumers
 594 increasingly prioritizing safety and health, the main research emphasis for the oral delivery of
 595 bioactive peptides will be on discovering natural, edible, and biocompatible materials that have low
 596 toxicity to serve as delivery carriers. Moreover, current design approaches for oral delivery systems
 597 mainly concentrate on overcoming the gastrointestinal barrier, while the targeting features of these
 598 systems have not been thoroughly investigated. As a result, a key area of research in the oral delivery
 599 of bioactive peptides will focus on creating targeted homeostasis within these systems. Additionally,
 600 most existing data on the oral delivery of bioactive peptides has come from in vitro or animal studies,

601 with a lack of relevant clinical data. To effectively evaluate the impact of oral delivery systems for
602 bioactive peptides on human health, clinical studies are necessary to determine if prolonged use of
603 these systems could result in unexpected side effects in vivo. With ongoing technological
604 advancements, it is expected that new hybrid delivery systems will be developed, leading to
605 improved delivery of bioactive peptides.

606 **5 Conflict of Interest**

607 *The authors declare that the research was conducted in the absence of any commercial or financial*
608 *relationships that could be construed as a potential conflict of interest.*

609 **6 Author Contributions**

610 Conceptualization, Songmin Cao and Wangang Zhang; methodology, Songmin Cao; software, Xinyu
611 Wang; writing—original draft preparation, Zeyao Yang & Xinyu wang; writing—review and
612 editing, Lujuan Xing, Ruiming Luo, Wangang Zhang. All authors have read and agreed to the
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- 979 **12 Data Availability Statement**
- 980 Not applicable.

Figure 01.JPEG

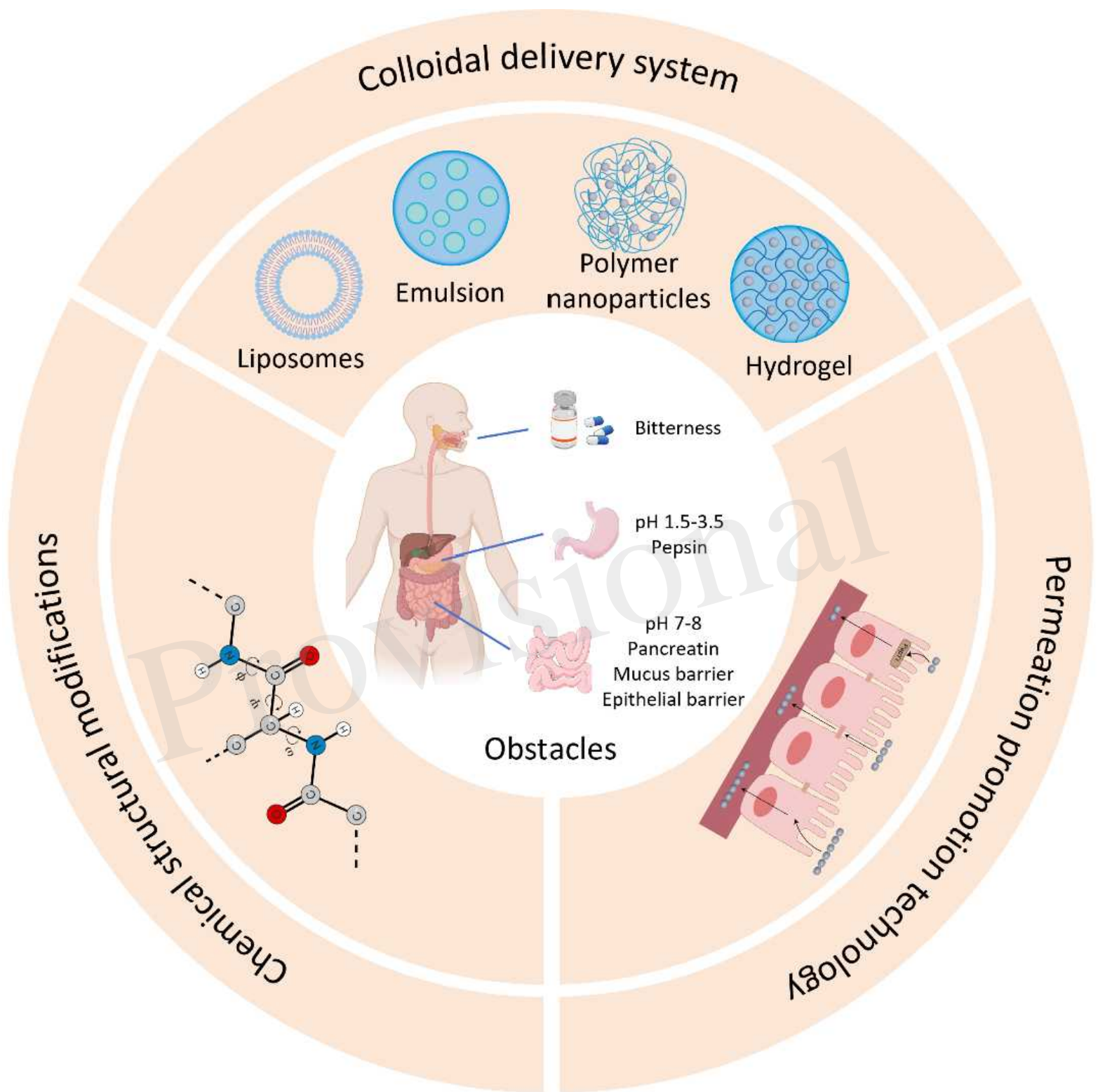


Figure 02.JPEG

