Check for updates

OPEN ACCESS

EDITED BY Mingquan Huang, Beijing Technology and Business University, China

REVIEWED BY Yongqiang Zhao,

South China Sea Fisheries Research Institute (CAFS), China Ya-Fang Shang, Hefei University of Technology, China

*CORRESPONDENCE Weijiang Sun ⊠ 000q020007@fafu.edu.cn

[†]These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Food Chemistry, a section of the journal Frontiers in Nutrition

RECEIVED 07 November 2022 ACCEPTED 05 December 2022 PUBLISHED 19 December 2022

CITATION

Wang Z, Wang Z, Dai H, Wu S, Song B, Lin F, Huang Y, Lin X and Sun W (2022) Identification of characteristic aroma and bacteria related to aroma evolution during long-term storage of compressed white tea. *Front. Nutr.* 9:1092048. doi: 10.3389/fnut.2022.1092048

COPYRIGHT

© 2022 Wang, Wang, Dai, Wu, Song, Lin, Huang, Lin and Sun. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Identification of characteristic aroma and bacteria related to aroma evolution during long-term storage of compressed white tea

Zhihui Wang^{1,2†}, Zhihua Wang^{1†}, Haomin Dai^{1,2}, Shaoling Wu^{1,2}, Bo Song^{1,2}, Fuming Lin^{1,2,3}, Yan Huang^{1,2,3}, Xingchen Lin⁴ and Weijiang Sun^{1,2*}

¹College of Horticulture, Fujian Agriculture and Forestry University, Fuzhou, China, ²Ministerial and Provincial Joint Innovation Centre for Safety Production of Cross-Strait Crops, Fujian Agriculture and Forestry University, Fuzhou, China, ³Anxi College of Tea Science, Fujian Agriculture and Forestry University, Quanzhou, China, ⁴Fujian Ming Shan Tea Industry Co., Ltd., Fuding, China

Compressed white tea (CWT) is a reprocessed tea of white tea. Long-term storage has greatly changed its aroma characteristics, but the material basis and transformation mechanism of its unique aroma are still unclear. In this study, flavor wheel, headspace gas chromatography ion mobility spectroscopy, chemometrics, and microbiomics were applied to study the flavor evolution and important aroma components during long-term storage of CWT, and core functional bacteria were screened. During long-term storage, the aroma of CWT gradually changed from sweet, fruity and floral to stale flavor, woody and herbal. A total of 56 volatile organic compounds (VOCs) were identified, 54 of which were significantly differences during storage. The alcohols content was the highest during 1-5 years of storage, the esters content was the highest during 7-13 years of storage, and the aldehydes content was the highest during 16 years of storage. Twenty-nine VOCs were identified as important aroma components, which were significantly correlated with 6 aroma sub-attributes (P < 0.05). The functional prediction of bacterial community reminded that bacterial community could participate in the transformation of VOCs during storage of CWT. Twenty-four core functional bacteria were screened, which were significantly associated with 29 VOCs. Finally, 23 characteristic differential VOCs were excavated, which could be used to identify CWT in different storage years. Taken together, these findings provided new insights into the changes in aroma characteristics during storage of CWT and increased the understanding of the mechanism of characteristic aroma formation during storage.

KEYWORDS

compressed white tea, storage, volatile compounds, GC-IMS, aroma, bacteria

1. Introduction

White tea is a kind of slightly fermented tea (1). After a long aging period, white tea can get unique aroma characteristic and then is named aged white tea. It is similar with aging process of Pu-erh tea and red wine (2, 3). However, the traditional white tea is coarse and loose, which is inconvenient for transportation and storage, so the compressed white tea (CWT) has been developed (4). CWT is a reprocessed tea product manufacture from white tea by blending, weighing, steaming, shaping, and drying. Its appearance effectively solved the problem of inconvenient transportation and storage of traditional white tea (5). Previous studies have expounded that the process of steaming and shaping further destroys the bud and leaf tissue of white tea, and has a far-reaching impact on the type and content of aroma components of CWT, making the stale flavor of CWT more show, and the characteristic aroma types such as jujube scent and herbal more prominent, while the traditional white tea retains more floral and fruity (6).

Aroma is an important factor that determines the quality of tea and affects the choice of consumers (7). When white tea is not stored, its aroma characteristics are floral, sweet, clean and refreshing, and the high proportion of alcohols and aldehydes is the basis of these aroma characteristics. Phenylethyl alcohol, γ-nonalactone, trans-β-ionone, trans-linalool oxide (furanoid), α -ionone and *cis*-3-hexenyl butyrate are considered to be the key aroma components of different types of unstored white tea (8). In the long-term storage process of white tea, the aroma will change with the extension of time (9). In the sensory, the aroma of white tea will gradually transform from clean and refreshing and floral to stale flavor, jujube scent or herbal (10). In the volatile organic compounds (VOCs), the content of alcohols decreases, hydrocarbons' increases, and aldehydes' increases first and then decreases; the aroma components of floral and fruity, such as linalool, linalool oxide, geraniol, methyl salicylate, phenylethanol, nerolidol, and citral, decreased, which make white tea clean and refreshing and floral gradually decrease or even disappear. A variety of unsaturated alkenes, mainly dihydrokiriactone, 2-methylnaphthalene, cedrene, and β -cedrene, are increased, resulting in the formation of stale flavor. Meanwhile, under the coordination of benzaldehyde, αionone, β -ionone, and geranyl acetone, the characteristics of white tea during storage, namely stale flavor with jujube scent, plumy aroma are formed (8, 11). The aroma of CWT is very different from that of traditional white tea (6). So far, the changes of aroma characteristics, important aroma components and the chemical basis of special aroma during storage of CWT are still unclear.

Headspace gas chromatography-ion mobility spectrometry (HS-GC-IMS) is an emerging technology, which can visualize VOCs without pretreatment and high sensitivity, and can better separate isomers and polymers in tea (12, 13). Nevertheless, how to match valuable information on the chemicals with sensory descriptors is still challenging. Recently, chemometrics and quantitative descriptive analysis (QDA) have been widely used to reveal the relationship between the chemical data from instrumental analysis and sensory analysis, and to recognize those chemical components that contribute significantly to food flavor (14, 15). However, there are few reports on the relationship between aroma characteristics and VOCs during storage of CWT. In addition, microorganisms may play a key role in the changes of tea metabolites during storage (3), which has been confirmed in Fu Brick tea, Liupao tea, and Pu-erh tea (16-18), and it is found that the diversity of bacteria increases gradually during the storage process of tea, mainly involved in the decomposition, transformation and degradation of small molecule compounds (17). Previous study have identified bacterial communities during storage of white tea and CWT, and performed functional prediction. It is found that bacteria can participate in 246 kyoto encyclopedia of genes and genomes (KEGG) metabolic pathways, including the metabolism of some VOCs (19, 20). Whereas, the relationship between the changes of VOCs and bacterial communities during storage of CWT are also still unknown.

Therefore, the aims of the present study were to (a) elucidate the dynamic changes in aroma characteristics and VOCs during storage of CWT, (b) elucidate the correlation between VOCs and aroma sub-attributes, and (c) identify important aroma components and core functional bacteria that contribute to the formation of characteristic aroma of CWT. This study is of significant importance for providing information in-depth to enhance the understanding of the mechanisms on aroma formation during storage of CWT.

2. Materials and methods

2.1. Experimental materials

Samples of CWT (round cake) were collected from Fujian Ruida Tea Industry Co., LTD., named A1, A3, A5, A7, A9, A10, A13, and A16, respectively. The last numbers in the nomenclature indicated the years of storage, which corresponded to the production years of 2018, 2016, 2014, 2012, 2010, 2009, 2006, and 2003. All samples were manufactured by Fujian Ruida Tea Industry Co., Ltd. in the spring with the same tea plant cultivar, plantation, and processing technology. The processing technology of CWT was that Shoumei white tea \rightarrow weighing (weight: 350.0 \pm 5.0 g) \rightarrow steam softening (steam temperature: 110° C; time: 30 s) \rightarrow shaping (round cake; pressure: 40 KN; time: $7 \min$) \rightarrow drying (temperature: $50^\circ \text{C};$ time: 48 h; water content $\leq\,$ 8%). All samples were stored in the same environment warehouse (dry and ventilated, temperature $< 35^{\circ}$ C, relative humidity < 50%). The samples of different batches prepared within 1 month each spring were regarded as biological repeats, and each sample was collected



with three biological repeats. The sample details are shown in Supplementary Table 1, and photos of dry tea, tea infusions and infused leaves of some CWT samples are shown in Figure 1.

2.2. Experimental method

2.2.1. Sensory evaluation of the aroma of CWT

Quantitative descriptive analysis (QDA) was performed by five well-trained panelists (two males and three females, aged 20-55 years) from the tea innovation team of Fujian Agriculture and Forestry University according to the Methodology for Sensory Evaluation of Tea (GB/T 23776-2018), with a little modification. Briefly, 3.0 g evenly mixed tea samples were weighed into a cylindrical cup of 150 mL, 150 mL boiling water was added, and soaked for 3 min. Then the tea infusion was poured into the evaluation bowl. After the initial evaluation of the aroma, the boiling water was added again, and the tea infusion was drained after 5 min for the second aroma evaluation. Aroma descriptors for each sample were recorded by each evaluator. After that, six descriptors with a usage rate of more than 80% were selected, namely sweet, floral, fruity, stale flavor, woody, and herbal. Descriptors were defined and their references were found based on published literature (Figure 2B) (21, 22). Finally, the tea samples were re-soaked according to the above method, and the 0-7 scale was used to evaluate the intensity of the six aroma sub-attributes of each sample, namely 0 =none, 4 =medium, and 7 =extremely. The aroma subattribute intensity value of each sample was the average value evaluated by the evaluation team. Each sample was evaluated in three biological replicates.

2.2.2. Qualitative and quantitative analysis of the VOCs by HS-GC-IMS

VOCs identification analysis was performed using HS-GC-IMS (FlavorSpec, G.A.S., Dortmund, Germany) (23). Briefly, 0.1 g of the ground CWT sample was weighed and placed into a 20 mL headspace injection bottle. The headspace injection conditions were as follows: incubation at 70° C for 15 min with revolving speed at 500 *r*/min, injection needle temperature at 90°C, and injection volume at 500 µL. Three biological replicates were performed.

GC condition: the chromatographic column was FS-SE-54-CB-1 (15 mL × 0.68 mm), and the carrier gas was N₂ (purity > 99.99%). The programmed flow rate was as follows: initially 2 mL/min holding for 10 min, linearly ramped up to 150 mL/min within 10 min, and then held for 30 min. IMS conditions: drift tube length and temperature were 98 mm and 45°C, drift gas flow rate was 150 mL/min. GC-IMS data were viewed through Laboratory Analytical Viewer (LAV), using the Reporter plugin to directly compare 2D top view and 3D spectra, and the Gallery mapping plug-in to visualize fingerprint comparisons. Normal alkanes C9—C27 (Sigma Aldrich Corporation, Saint Louis, MO, USA) were served as external references to calculate the retention index (RI) of each compound. The calculation method of RI was referred to Mao et al. (24).

2.2.3. Identification of bacterial communities

Previously, the bacterial communities of samples A1, A3, A7, A9, A10, and A16 during storage of CWT were identified (20). Primers were designed according to the V3 and V4 conserved regions of bacterial 16S rDNA for polymerase chain reaction amplification. Two-end sequencing of 2×250 bp was performed by NovaSeq PE250 sequencer (Kapa Biosciences, Woburn, MA, USA). Three biological replicates were performed.

2.3. Data analysis

One-way analysis of variance (ANOVA) with least significant difference (LSD) was performed using SPSS (version 19.0; Chicago, IL, USA). Bacterial involvement in metabolic pathways was predicted by PICRUSt2 function prediction analysis software (https://www.omicstudio.cn/tool). Principal component analysis (PCA), orthonormal partial least-squares discriminant analysis (OPLS-DA), orthonormal partial least-square variable import of project (OPLS-VIP), and two way orthogonal partial least-squares analysis (O2PLS) were performed using SIMCA (version 14.0, Umetrics, Umea, Sweden). Heat maps and correlation analysis were generated using Hiplot (https://hiplot-academic.com). The correlation network diagrams were generated using Cytoscape (version 3.9.1; Beijing, China).



3. Results and Discussion

3.1. Flavor evolution during storage of CWT

QDA was used to construct the flavor wheel of CWT during storage (Figure 2A) and formed its aroma description system (Figure 2B). During the long-term storage of CWT, the characteristics of six typical aroma sub-attributes were happened significant (P < 0.05) or extremely significant changes (P < 0.01) (Figure 2C). The intensity of sweet, floral and fruity was significantly decreased with increasing storage years, while that of stale flavor, woody, and herbal emerged a significant increasing trend (Figure 2C). The aroma intensity of CWT in the first year of storage was dominated by sweet, fruity and floral, which were 6.5, 5.1, and 3.7, respectively. During 3–5 years of storage, the intensity of stale flavor gradually

increased, and that of floral was gradually covered, with sweet, stale flavor and fruity as the dominant flavor. During 7-13 years of storage, the intensity of woody increased significantly, while that of sweet and floral decreased significantly. In this period, stale flavor, fruity and woody were the dominant flavor. After 16 years of storage, the intensity of herbal increased significantly and became the dominant flavor, with an aroma intensity of 6.6 (Figures 2A,C). In general, the change course of CWT aroma during long-term storage was as follows: sweet, fruity and floral \rightarrow sweet, fruity and stale flavor \rightarrow stale flavor, fruity and woody \rightarrow herbal. The change trend was similar to the change of white tea aroma during storage (10). The difference was that CWT had no clean and refreshing at the beginning of the storage, and the floral was weaker. After 3 years of storage, CWT had shown obvious stale flavor, and the aging speed was faster than that of white tea



There was a complex correlation between the six aroma sub-attributes (P < 0.05) (Figure 2D). Sweet, fruity and floral were significantly positively correlated with each other, and these three sub-attributes were significantly negatively correlated with woody and herbal, respectively, indicating that the aroma compounds contributing to the three sub-attributes might be similar. There was a significant positive correlation between woody and both stale flavor and herbal. There was a positive correlation between herbal and stale flavor, but the correlation did not reach a significant level (P > 0.05). Some studies have suggested that the biochemical basis of stale flavor in tea was aromatic compounds with pleasant woody and herbal (6, 25). This was similar to the conclusion of this study.

3.2. Changes of VOCs during storage of CWT

The general chromatogram of HS-GC-IMS showed that the relative abundance of VOCs in CWT during storage,

and some VOCs increased and some decreased (yellow box) (Figure 3A). A total of 56 VOCs were identified, including 5 esters, 19 aldehydes, 16 alcohols, 7 ketones, 7 heterocycles, and 2 terpenes (Figure 3B). CAS, molecular formula, RI and relative content of all VOCs are shown in Supplementary Table 2. The most abundant VOCs in CWT were alcohols, esters and aldehydes. The dominant volatile components during longterm storage were constantly changing (Figure 3B). The alcohols content was the highest during 1-5 years of storage, the esters content was the highest during 7-13 years of storage, and the aldehydes content was the highest during 16 years of storage. The content of alcohols decreased gradually during long-term storage, ranging from 24.43 to 33.44%, and reached the highest level after 1 year of storage. During storage, esters first decreased, then increased, and then decreased, with the content ranging from 23.77 to 39.76%. The content was the highest after 10 years of storage. The variation trend of aldehydes was "N", and the content of aldehydes ranged from 18.22-28.02%, with the highest content after 16 years of storage. The contents of heterocycles, ketones and terpenes were relatively low, and heterocycles' and ketones' gradually increased, while terpenes' gradually decreased. Previous study has claimed that the contents of alcohols, aldehydes and hydrocarbons in aged white tea are the highest (11). During storage of white tea, alcohols decrease continuously, hydrocarbons increase, and aldehydes increase first and then decrease (11). There were some differences with the conclusions of this study, which might be the difference between white tea and CWT. The content of esters in CWT was higher and further increased during storage, while aldehydes increased in a fluctuating manner. The increase of some esters and aldehydes might be caused by microbial metabolism, while the decreased of alcohols might be due to their own volatilization. Previous studies have shown that some alcohols gradually evaporate during tea storage (15). Some esters and aldehydes can be biosynthesized by microbial enzymes using precursors such as amino acids and fatty acids during tea fermentation (26).

The most abundant VOCs in CWT were y-butyrolactone D (dimer) (13.79-29.44%), linalool M (3.99-9.54%), ethyl benzoate (3.89-9.51%), 1-propanol D (3.32-8.55%), linalool oxide M (3.27-6.39%), 2-ethyl-6-methylpyrazine (4.18-6.18%), and γ -butyrolactone M (monomer) (2.27–3.70%) (Supplementary Table 2). Linalool and its oxides have always been considered as VOCs with high content in white tea (27). This view was also similar in CWT. The high content of γ -butyrolactone in CWT was identified for the first time. y-Lactone is the most common type of lactone, but less are identified by ordinary GC-MS (28). Xiao et al. (29) identified y-butyrolactone in Fu brick tea from different producing areas by using GC-IMS, but failed to do so by using GC-MS. Therefore, it was speculated that GC-IMS have unique advantages in identifying y-butyrolactone and its dimer. Zhu et al. (9) performed targeted identification of part of lactones in white tea with different storage times (unidentified y-butyrolactone) through enantioselective gas sterics-mass spectrometry (Es-GC-MS), it was found that some lactones accumulated with the increase of white tea storage time. In the study of wine, the content of γ -lactone is also found to be related to the aging of wine (30, 31). Therefore, long-term storage of CWT might contribute to y-butyrolactone accumulation. Among the 56 identified VOCs, 54 had significant differences in accumulation during storage (P < 0.01). Heat map analysis certified that γ -butyrolactone D increased significantly, γ butyrolactone M rised first and then decreases, and some alcohols such as linalool M and linalool oxide M decreased significantly. Methyl salicylate and ethyl benzoate decreased significantly. Heterocycles such as 2-ethyl-6-methylpyrazine and methylpyrazine M increased significantly. Aldehyde such as benzaldehyde D and 3-methylbutanal D increased significantly (Figure 3C). The differential accumulation of these VOCs changed the flavor of CWT during storage, but not all VOCs contributed to the aroma. Therefore, chemometrics should be further combined to find the important aroma components.

PCA showed the similarities and differences of samples in CWT during storage. In terms of differences, there was no coincidence among samples, indicating that VOCs were different among samples (Figure 3D), which was consistent with the conclusion of heat map analysis (Figure 3B). From the perspective of similarity, the whole storage process was divided into four stages: stage 1: A1; stage 2: A3–A5; stage 3: A7– A13, stage 4: A16 (the closer the distance, the more similar) (Figure 3D). The four stages of the storage process divided by PCA were consistent with the changes of the sensory attributes of aroma mentioned above (Figure 2A). Based on the above analysis, it could be reasonably inferred that the flavor change of CWT during storage was due to the change of VOCs content, which might be related to the volatilization of compounds and the metabolism of microorganisms during storage.

3.3. Characteristic differential VOCs of CWT in different storage years

PCA confirmed differences between CWT samples from different storage years. In order to screen the differential VOCs of CWT in different years, OPLS-DA modeling analysis was performed. Samples were effectively distinguished in the score chart (Figure 4A), and cross-validation informed that the model was not overfitted (green points were below blue points) (Figure 4B). With VIP > 1, P < 0.05 as the criterion (32), 23 differential VOCs were screened. The screening of characteristic differential VOCs is based on the variable variation trend of differential VOCs in the heat map (Figure 3B). This method has been used accurately in the screening of characteristic differential metabolites of tea from different grades and different regions (33, 34). The differential VOCs with high content in the sample of this storage year but low content in other storage years were considered as the characteristic differential VOCs of CWT of this year. Therefore, the characteristic differential VOCs were screened as follows: 2 in A1, 5 in A3, 2 in A5, 3 in A7, 3 in A9, 1 in A10, 2 in A13, and 5 in A16 (Figure 4C). These characteristic differential VOCs were differential VOCs accumulated specifically by CWT in a certain year, which might contribute significantly to the formation of its unique flavor, and could also be used to trace and identify the origin of CWT in different storage years. However, the reasons for the unique accumulation of these VOCs in different storage years needed to be further studied.

3.4. Screening of important aroma components during storage of CWT

In order to find essential variables that are the most influential on the sensory attributes of tea aroma, especially



Characteristic differential VOCs of CWT in different storage years. (A) OPLS-DA model. $R^2X = 0.999$, $R^2Y = 0.998$, $Q^2 = 0.862$. (B) Cross validation of OPLS-DA model. (C) Rising sun chart and molecular formula of characteristic differential VOCs.



when multicollinearity exists among variables, a OPLS-VIP is conducted to explain sensory attribute intensities with volatile component content as the *X* variable and sensory attribute intensity as the *Y* variable, and the VIP value of each *X* variable is calculated (24). The variables whose VIP values are >1 are considered to be the important aromatic compounds (21).

The OPLS-VIP model had a high degree of interpretation $(R^2X = 0.973, R^2Y = 0.976)$ (Figure 5A), and the closer the variables were, the greater the correlation was. Figure 5A showed that floral, fruity and sweet fragrance were the main aroma characteristics of A1, A3, and A5 (closer to each other), and stale flavor, woody and herbal were the main aroma characteristics of A7, A9, A10, A13, and A16 (closer to each other), which was consistent with the results of the above sensory analysis. Cross-validation informed that the model was not overfitted (Figure 5B). According to the principle of VIP value > 1.0, 29 volatile components were identified as aroma components that had important contributions to CWT sensory aroma (Figure 5C, yellow column). The aroma characteristics of these VOCs are relatively diversified, mainly including sweet, fruity, woody, caramel and fatty (Supplementary Table 2) (35, 36). These important aroma components acted as the aroma skeleton of CWT and interacted with other aroma components to form the unique aroma characteristics of CWT in different storage years. Linalool, linalool oxide, methyl salicylate, benzyl alcohol, 2-phenylethanol, and phenylacetaldehyde were closer to A1 and A3 (Figure 5A), and their aroma characteristics were fruit, sweet and floral (Supplementary Table 2) (35, 36). Many studies have verified that these VOCs are important aroma substances in fresh white tea (6, 11). In this study, chemometric methods were adopted to more accurately confirm the dominant contribution of these VOCs at the initial stage of CWT storage. y-Butyrolactone D was closer to A7, A9, A10, A13, and A16 (Figure 5A). The thresholds of lactone compounds are extremely low, most of which have floral, fruity, milk flavor, and coconut, and are the important aroma substances of wine and fruit (9). There are also a variety of lactones in tea, which are mainly derived from $\gamma(\delta)$ -hydroxy carboxylic acid intramolecular esterification or carotene degradation during processing and storage. y-Valerolactone, y-heptanolactone, and $\gamma(\delta)$ -decanolactone have important contributions to tea quality (28). In this study, γ -butyrolactone was found to be a important aroma component in CWT storage process for the first time. It was likely that this is an important reason for the unique aroma of CWT.

3.5. Contribution of important aroma components to aroma sub-attributes

The correlation coefficient was used to determine the unique contribution of 29 important aroma components to

six aroma sub-attributes. The 29 important aroma components were significantly correlated with six aroma sub-attributes (P < 0.05) (Figure 6). The correlation coefficient is shown in Supplementary Table 3, and the visual network diagram is shown in Figure 6.

Twelve VOCs were positively correlated with sweet, and six VOCs were negatively correlated with sweet. Six VOCs were positively correlated with fruity, and five VOCs were negatively correlated with fruity. There were nine VOCs positively correlated with floral, and six VOCs negatively correlated with floral (P < 0.05) (Figure 6). Linalool M, phenylacetaldehyde, pentanal D, pentanal M, and limonene were positively correlated with sweet, floral, and fruity. The aroma characteristics of these components are sweet, floral, fruit, citrus, lemon and cherry like (Supplementary Table 2) (35, 36). So they had a positive contribution to these aroma sub-attributes. The contents of these VOCs decreased significantly during storage (Figure 3B). (E)-2-Octenal and hexanal T were significantly negatively correlated with these three aroma sub-attributes, and the aroma characteristics of these two components are herbal, fatty and grass fragment (Supplementary Table 2) (35, 36). Therefore, it had a negative effect on sweet, floral and fruity. The contents of these two aroma components increased significantly during storage (Figure 3B). Sweet, floral and fruity had similar contribution components, which also verified the correlation analysis results among the above three (Figure 2B). The VOCs that had significant contribution to the aroma intensity of sweet, floral, and fruity also different. The VOCs with significant positive correlation with sweet also included methyl salicylate, ethyl benzoate, linalool D, linalool oxide M and linalool oxide D. The aroma characteristics of these components are mainly sweet, floral, fruity, honey-like and fresh (Supplementary Table 2). The VOCs with significant negative correlation with sweet were γ -butyrolactone D, 1-propanol D and methylpyrazine M, the aroma characteristics of these VOCs are caramel, cream, milk, fatty, ethanol-like, roasty, nutty and earthy (Supplementary Table 2). This hinted that the VOCs with caramel, fatty, roasty, and earthy were possible to reduce the sweet intensity of CWT. Benzyl alcohol (applelike) had a significant positive correlation with fruity. ybutyrolactone M (caramel) and 1-propanol M (pineapple) were significantly positively correlated with floral. The contents of these components of CWT decreased gradually during storage.

Six VOCs were positively correlated with stale flavor, and thirteen VOCs were negatively correlated with stale flavor. Six VOCs were positively correlated with woody, and eighteen VOCs were negatively correlated with woody. There were five VOCs positively correlated with herbal, and eight VOCs negatively correlated with herbal (P <0.05) (Figure 6). γ -Butyrolactone D, 1-propanol D, and 2heptanone D were positively correlated to the stale flavor and woody. The aroma characteristics of these three VOCs are carmel, cream, milk, fatty, herbal, and banana-like



Correlation network diagram of aroma sub-attributes and key aroma components. Blue represented a significant negative correlation, red represented a significant positive correlation (P < 0.05), and the darker the color, the greater the correlation.



Intensity thermogram of KEGG pathway related to bacterial involvement in VOCs metabolism. Red indicated high intensity, while blue indicated low intensity.



(Supplementary Table 2) (35, 36). The contents of these three VOCs increased gradually during storage. y-Butyrolactone M, phenylacetaldehyde, pentanal D, pentanal M, linalool M, linalool D, linalool oxide M, linalool oxide D, 1-propanol M, and limonene were negatively correlated with stale flavor and herbel. Their aroma characteristics are sweet, floral and fruity (Supplementary Table 2). The content of these VOCs decreased gradually during storage. Different with woody, methylpyrazine M showed significant positive correlations with the stale flavor. The aroma characteristic of methylpyrazine M is roasty, nutty, and earthy (Supplementary Table 2). This suggested that VOCs with roasty, earthy, nutty, and herbal might affect the strength of stale flavor. It has obvious stale flavor in the aged white tea, Pu-erh tea, and Liupao tea (19). However, there were differences in the VOCs identified to contribute to stale flavor in different tea types. In Pu-erh tea, which is methoxybenzenes (37); in Liupao tea, which is α -cedarol, β -linalool, dihydrokiristone, α -terpineol, and β -ionone (38); in aged white tea, which is cedarol, α -cedarene, and β -cedarene (6). Deng et al. (25) believed that the material basis of stale flavor in tea was mainly aromatic substances with pleasant woody and herbal. The results of this study were similar to this conclusion. Therefore, it was hypothesized that the reason for the difference in VOCs contributing to stale flavor among different tea types was the result of the interaction between these components. Different from stale flavor and wood, the VOCs that were significantly positively correlated with herbal were mainly (E)-2-octenal, (E)-2-heptenal, hexanal T, heptanal D, and methylpyrazine M. The aroma characteristics of these VOCs are herbal, fatty, grassy,

stale flavor, fruity and cilantro (Supplementary Table 2). The contents of these VOCs gradually increased during storage. The seven VOCs negatively correlated with the herbal are mainly floral, fruity, and sweet. Their contents decreased gradually during storage of CWT.

In general, during the storage process of CWT, the important VOCs that were positively correlated with sweet, fruity, and floral aroma gradually decreased, while the components that were negatively correlated with them gradually increased. However, the important VOCs that were positively correlated with stale flavor, woody, and herbal gradually increased, while those that were negatively correlated with them gradually decreased. This was the main reason why the aroma of the CWT changed from sweet, floral, and fruity to stale flavor, woody, and herbal during storage.

3.6. Functional prediction of bacterial community

Previous studies have found that microorganisms play an important role in the transformation of tea aroma during storage, among which bacteria may play a key role (17). Combined with the previous bacterial community data of CWT in different storage years (Supplementary Table 4) (20), PICRUSt2 was used to predict and analyze the function of bacteria, and acquire the information that bacteria in CWT storage process could participate in 264 KEGG metabolic pathways. Among them, 34 pathways were related to the

metabolic transformation of VOCs and aroma precursors (Figure 7). The formation of tea aroma is divided into four ways: degradation of carotenoids, oxidative degradation of lipids, maillard reaction of sugars and amino acids, and hydrolysis of glycosides (25). The 34 metabolic pathways screened by us were all related to the four pathways of tea aroma formation, mainly including amino acid and carbohydrate metabolism, fatty acid metabolism, glycosyltransferase synthesis, peroxidase synthesis, terpene degradation and secondary metabolite biosynthesis, and biodegradation. The styrene degradation, alinolenic acid metabolism, limonene and pinene degradation participated by bacteria first increased and then decreased during storage of CWT (Figure 7). The styrene content first increased and then decreased during storage, and the limonene content gradually decreased during storage (Figure 3B). α-Linolenic acid metabolism could synthesize (Z)-3-hexen-1-ol (39), and that content decreased first and then increased during storage (Figure 3B). Moreover, bacteria could participate in the metabolism of amino acid related enzymes, glycosyltransferases and peroxisome (Figure 7). These enzymes can be involved in the synthesis or degradation of tea aroma (39). Previous studies have shown that bacteria can secrete extracellular enzymes during tea storage, thereby affecting the quality of tea (17). This study also verified this conclusion through the prediction of bacterial function. In conclusion, it was reasonable to speculate that bacteria could participate in the transformation of VOCs during storage of CWT and play an important role, but not all bacteria did this. Therefore, the core functional bacteria affecting VOCs were further screened.

3.7. Screening of core functional bacteria affecting the change of VOCs

Combined with the identified VOCs content and bacterial abundance data, an O2PLS model was established to study the association between bacteria and VOCs during storage of CWT. The O2PLS model had a high degree of interpretation $(R^2X = 0.791, R^2Y = 0.797)$ (Supplementary Figure 1). Crossvalidation showed no overfitting (Supplementary Figure 2). According to the principle of VIP value > 1.0, 116 bacterial genera were confirmed to have important influence on VOCs (Supplementary Table 5). Furthermore, three conditions were considered to identify the core functional bacteria that had an important contribution to the VOCs during storage of CWT: (a) VIP value > 1.0; (b) correlation coefficient |r| > 0.8and P < 0.05; (c) the relative abundance of bacterial genera must be >1.0% (15, 16, 40, 41). Based on three criteria, 24 bacterial genera were identified as core functional bacteria. Core functional bacteria were significantly correlated with 29 VOCs (17 were important aroma components), including 12 aldehydes, 8 alcohols, 2 esters, 4 heterocycles, 2 terpenes,

and 1 ketone (Figure 8). Correlation coefficients are shown in Supplementary Table 6. It could be seen from the network diagram that the core functional bacteria were the greatest correlation with 12 aldehydes, and these aldehydes tended to increase in the storage process (Figure 3B). The aroma precursors of hexanal, heptanal, and pentanal are lipids, and the aroma precursors of 3-methylbutanal M are amino acids (12, 42). Among the core functional bacteria, Bacillus, Brevundimonas, Lactobacillus, and Enterococcus can produce a large number of various types of extracellular enzymes (43, 44). In Liupao tea, Fu brick tea, and Pu-erh tea, microorganisms have been found to affect the aroma quality of tea by secreting extracellular enzymes (15, 16, 29). Therefore, it was speculated that these core functional bacteria promoted the degradation of amino acids and fatty acids by secreting extracellular enzymes, thus making the rise of these aldehydes aroma components. Similar to aldehydes were ketones. The precursor substances of 2-heptanone M are carotenoids (42). The content of alcohols such as linalool, 1-propanol and 2-phenylethanol decreased gradually during storage. Previous studies deem that the decline of alcohols during tea storage is related to its volatilization (15). This study found that Mitochondria unclassified and Microbiaceae unclassified were significantly correlated with content change of alcohols. Moreover, these two core functional bacteria were also significantly correlated with the content change of esters (methyl salicylate and ethyl benzoate) and terpenes (limonene). At present, there was no report on the functions of these two bacteria. So it was not clear how these two bacteria affected the decline of alcohols, esters and terpenes. Oxyphotobacteria unclassified was the dominant bacteria in CWT with the abundance range of 9.02-93.14%, which increased first and then decreased during storage (Supplementary Table 4). It was positively correlated with the content changes of most alcohols, and negatively correlated with the content changes of many aldehydes and heterocycles. Oxyphotobacteria unclassified exists in a large number of food fermentation processes, and its abundance gradually decreases with the increase of the degree of fermentation (45). However, the main function of this bacterium was still unclear, and further experiments needed to be designed and studied. These findings collectively indicated that core functional bacteria made a key contribution to the characteristic aroma formation during storage of CWT.

4. Conclusion

This study was the first report on the flavor evolution and dynamic changes of VOCs of CWT during long-term storage (1–16 years), and attempted to correlate them with bacterial communities. It was of great significance for us to further understand the flavor transformation and the formation mechanism of characteristic aroma of CWT during storage. The flavor wheel and aroma description system for CWT during storage was constructed, and 29 VOCs were identified as important aroma components, among which the unique contribution of γ -butyrolactone to the aroma of CWT was first discovered. According to the flavor wheel and VOCs, the storage process of CWT was divided into four stages. The change process of aroma was sweet, fruity, and floral (1 year) \rightarrow sweet, fruity, and stale flavor $(3-5 \text{ years}) \rightarrow \text{ stale flavor, fruity, and woody}$ $(7-13 \text{ years}) \rightarrow$ herbal (16 years). The reason for the change of flavor was that the important aroma components which were positively correlated with sweet, fruity and floral gradually decrease, while the components that were positively correlated with stale flavor, woody and herbal gradually increased. The functional prediction of bacterial communities revealed that bacteria participated in 34 metabolic pathways related to VOCs transformation, and 24 bacterial genera were identified as core functional bacteria. In addition, 23 characteristic differential VOCs of CWT in different storage years were screened out, which could be used to distinguish CWT in different years. Since the standard of all VOCs could not be obtained, the absolute quantification of these VOCs was not carried out, and the aroma recombination or omission experiment was not used for verification. Therefore, only chemometrics was used in this study to screen VOCs that had an important contribution to the sensory aroma sub-attributes of CWT. Further studies should use molecular sensory science to explore the molecular sensory basis of a specific flavor of CWT.

Data availability statement

The data presented in the study are deposited in the OMIX, China National Center for Bioinformation/Beijing Institute of Genomics, Chinese Academy of Sciences, accession number OMIX002501, https://ngdc.cncb.ac.cn/omix/select-edit/OMIX002501.

Author contributions

WS, ZhihuiW, ZhihuaW, FL, YH, and XL conceived and designed the experiments. ZhihuiW, HD, and ZhihuaW performed the experiments. ZhihuiW analyzed the data and wrote the manscript. HD, SW, and BS revised the manuscript critically. WS administrated the project. All the authors read and approved the final manuscript.

References

Funding

Research This work was supported by and demonstration on quality evaluation of Fuding white tea (2020N3014), Key R&D plan of the Ministry of and Technology of China (2019YFD1001601), Science special project of China's central government to development guide local science and technology (2022L3071), and special fund for Science and Technology Innovation of Fujian Zhang Tianfu tea development foundation (FJZTF01).

Acknowledgments

We are particularly grateful to Fujian Ruida Tea Industry Co., Ltd., for providing us with experimental samples.

Conflict of interest

XL was employed by Fujian Ming Shan Tea Industry Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fnut.2022.1092048/full#supplementary-material

^{1.} Dai WD, Xie DC, Lu ML Li PL, Lv HP, Yang C, et al. Characterization of white tea metabolome: comparison against green and black tea by a nontargeted metabolomics approach. *Food Res Int.* (2017) 96:40–5. doi: 10.1016/j.foodres.2017.03.028

^{2.} Lorenzo C, Garde-Cerdán T, Pedroza MA, Alonso GL, Salinas MR. Determination of fermentative volatile compounds in aged red wines by near infrared spectroscopy. *Food Res Int.* (2009) 42:1281–6. doi: 10.1016/j.foodres.2009.03.021

3. Xue J, Yang L, Yang Y, Yan J, Ye YT, Hu CY, et al. Contrasting microbiomes of raw and ripened Pu-erh tea associated with distinct chemical profiles. *LWT Food Sci Technol.* (2020) 124:109147. doi: 10.1016/j.lwt.2020.10 9147

4. Huang Y, Liu F, Sun WJ. Research progress of white tea products and processing technology. *China Tea Process.* (2015) 6:5-9 (in Chinese).

5. Weng GF. Quality and safety evaluation of compressed white tea. *Agric Eng.* (2020) 10:50–4 (in Chinese).

6. Chen ZD, Li P, Chen XH, Yang YJ, He PM, Tu YY. Effect of compressed processing on the aroma of aged white tea. *Food Ind Sci Technol.* (2020) 41:63–9 (in Chinese).

7. Wu HT, Chen YY, Feng WZ, Shen SS, Wei YM, Jia HY, et al. Effects of three different withering treatments on the aroma of white tea. *Foods.* (2022) 11:2502. doi: 10.3390/foods11162502

8. Chen QC, Zhu Y, Yan H, Chen M, Xie DC, Wang MQ, et al. Identification of aroma composition and key odorants contributing to aroma characteristics of white teas. *Molecules*. (2020) 25:6050. doi: 10.3390/molecules25246050

9. Zhu Y, Kang SY, Han Y, Lv HP, Zhang Y, Lin Z. Enantiomeric distributions of volatile lactones and terpenoids in white teas stored for different durations. *Food Chem.* (2020) 320:126632. doi: 10.1016/j.foodchem.2020.126632

10. Fu HF. Study on Pressed Cake and Storage of Aged White Tea. Fuzhou: Fujian Agriculture and Forestry University (2019) (in Chinese).

11. Liu LY. Studies on the Quality Characteristics and Free Radical Scavenging Ability of Stored White Tea. Fuzhou: Fujian Agriculture and Forestry University (2015) (in Chinese).

12. Liu HC, Xu YJ, Wu JJ, Wen J, Yu YS, An KJ, et al. GC-IMS and olfactometry analysis on the tea aroma of Yingde black teas harvested in different seasons. *Food Res Int.* (2021) 150:110784. doi: 10.1016/j.foodres.2021.110784

13. Sun P, Xu B, Wang Y, Lin X, Chen C, Zhu J, et al. Characterization of volatile constituents and odorous compounds in peach (*Prunus persica L*) fruits of different varieties by gas chromatography–ion mobility spectrometry, gas chromatography–mass spectrometry, and relative odor activity value. *Fron Nutr.* (2022) 9:965796. doi: 10.3389/fnut.2022.965796

14. Su D, He JJ, Zhou YZ Li YL, Zhou HJ. Aroma effects of key volatile compounds in Keemun black tea at different grades: HS-SPME-GC-MS, sensory evaluation, and chemometrics. *Food Chem.* (2022) 373:131587. doi: 10.1016/j.foodchem.2021.131587

15. Li Q, Li YD, Luo Y, Xiao LZ, Wang KB, Huang JA, et al. Characterization of the key aroma compounds and microorganisms during the manufacturing process of Fu brick tea. *LWT*. (2020) 127:109355. doi: 10.1016/j.lwt.2020.109355

16. Li Q, Hong X, Zheng XX, Xu YQ, Lai XM, Teng CQ, et al. Characterization of key aroma compounds and core functional microorganisms in different aroma types of Liupao tea. *Food Res Int.* (2022) 152:110925. doi: 10.1016/j.foodres.2021.110925

17. Li J, Wu J, Xu N, Yu Y, Wu X. Dynamic evolution and correlation between microorganisms and metabolites during manufacturing process and storage of pu-erh tea. *LWT*. (2022) 158:113128. doi: 10.1016/j.lwt.2022.1 13128

18. Li J, Xu R, Zong L, Brake J, Cheng L, Wu J, et al. Dynamic evolution and correlation between metabolites and microorganisms during manufacturing process and storage of Fu Brick tea. *Metabolites.* (2021) 11:703. doi: 10.3390/metabol1100703

19. Chen JJ, Hu YF, Shen SY, Wang ZH, Zhou Z, Tang Q, et al. Analysis of bacterial diversity on the surface of stored white peony based on high-throughput sequencing. *Tea Sci.* (2020) 40:519–27 (in Chinese).

20. Wang ZH. Study on the Quality and Bacterial Community Changes of Compressed White Tea in Different Years. Fuzhou: Fujian Agriculture and Forestry University (2021) (in Chinese).

21. Mao S. Quality Analysis and Control of Congou Black Tea Based on Sensomics. Chongqing: Southwest University (2018).

22. Yue CN, Yang PX, Qin DD, Cai HL, Wang ZH Li C, et al. Identification of volatile components and analysis of aroma characteristics of Jiangxi Congou black tea. *Int J Food Prop.* (2020) 23:2160–73. doi: 10.1080/10942912.2020.18 44747

23. Wang ZH, Xue ZH, Zhu WW, Sun WJ. Analyses of volatile compounds in compressed white tea of different years based on GC-IMS. *J Food Sci Biotechnol.* (2021) 40:85–94 (in Chinese).

24. Mao S, Lu C, Li M, Ye Y, Wei X, Tong H. Identification of key aromatic compounds in Congou black tea by partial least-square regression with variable importance of projection scores and gas chromatography-mass spectrometry/gas chromatography-olfactometry. *J Sci Food Agric*. (2018) 98:5278–86. doi: 10.1002/jsfa.9066

25. Deng XJ, Huang GH, Tu Q, Zhou HJ Li YL, et al. Evolution analysis of flavoractive compounds during artificial fermentation of pu-erh tea. *Food Chem.* (2021) 357:129783. doi: 10.1016/j.foodchem.2021.129783

26. Stribny J, Gamero A, Perez-Torrado R, Querol A. Saccharomyces kudriavzevii and Saccharomyces uvarum differ from Saccharomyces cerevisiae during the production of aroma-active higher alcohols and acetate esters using their amino acidic precursors. *Int J Food Microbiol.* (2015) 205:41-6. doi: 10.1016/j.ijfoodmicro.2015.04.003

27. Qi DD, Miao AQ, Cao JX, Wang WW, Chen W, Pang S. Study on the effects of rapid aging technology on the aroma quality of white tea using GC-MS combined with chemometrics: in comparison with natural aged and fresh white tea. *Food Chem.* (2018) 265:189–99. doi: 10.1016/j.foodchem.2018.05.080

28. Yan H. Study on the Enantiomers of Volatile Lactones and Terpenoids in Fuding White Tea. Beijing: Chinese Academy of Agricultural Sciences (2019) (in Chinese).

29. Xiao Y, Huang Y, Chen Y, Xiao L, Zhang X, Yang C, et al. Discrimination and characterization of the volatile profiles of five Fu brick teas from different manufacturing regions by using HS-SPME/GC-MS and HS-GC-IMS. *Curr Res Food Sci.* (2022) 9:24. doi: 10.1016/j.crfs.2022.09.024

30. Stamatopoulos P, Brohan E, Prevost C, Siebert TE, Herderich M, Darriet P. Influence of chirality of lactones on the perception of some typical fruity notes through perceptual interaction phenomena in bordeaux dessert wines. *J Agric Food Chem.* (2016) 64:8160–7. doi: 10.1021/acs.jafc.6b03117

31. Cooke RC, Van Leeuwen KA, Capone DL, Gawel R, Elsey GM, Sefton MA. Odor detection thresholds and enantiomeric distributions of several 4alkyl substituted γ-lactones in Australian red wine. *J Agric Food Chem.* (2009) 57:2462–7. doi: 10.1021/jf8026974

32. Yue CN, Peng H, Li WJ, Tong ZF, Wang ZH, Yang PX. Untargeted metabolomics and transcriptomics reveal the mechanism of metabolite differences in spring tender shoots of tea plants of different ages. *Foods.* (2022) 11:2303. doi: 10.3390/foods11152303

33. Wang ZH, Gan S, Sun WJ, Chen ZD. Widely targeted metabolomics analysis reveals the differences of nonvolatile compounds in Oolong tea in different production areas. *Foods.* (2022) 11:1057. doi: 10.3390/foods11071057

34. Han ZS, Wen MC, Zhang HW, Zhang L, Wan XC, Ho CT, et al. based metabolomics and sensory evaluation reveal the critical compounds of different grades of Huangshan Maofeng green tea. *Food Chem.* (2022) 374:131796. doi: 10.1016/j.foodchem.2021.131796

35. Wang J, Li MR, Wang H, Huang WJ Li F, Wang LL, et al. Decoding the specific roasty aroma Wuyi rock tea (*Camellia sinensis*: Dahongpao) by the sensomics approach. *J Agric Food Chem.* (2022) 70:10571–83. doi: 10.1021/acs.jafc.2c02249

36. Guo XY, Schwab W, Ho CT, Song CK, Wan XC. Characterization of the aroma profiles of oolong tea made from three tea cultivars by both GC–MS and GC-IMS. *Food Chem.* (2022) 376:131933. doi: 10.1016/j.foodchem.2021.131933

37. Xu YQ, Wang C, Li CW, Liu SH, Zhang CX Li LW, et al. Characterization of aroma-active compounds of pu-erh tea by headspace solidphase microextraction (hs-spme) and simultaneous distillation-extraction (sde) coupled with gc-olfactometry and gc-ms. *Food Anal Methods*. (2016) 9:1188– 98. doi: 10.1007/s12161-015-0303-7

38. Wen LX, Zhang F, He MZ, Huang SH, Peng JR, Lin JW, et al. Quality characteristics of stale flavor Liupao teas and establishment for evaluation method of aroma quality. *Food Ind Sci Technol.* (2021) 42:230–6 (in Chinese).

39. Qiao DH Mi XZ, An YL, Xie H, Cao KM, Chen HR, et al. Integrated metabolic phenotypes and gene expression profiles revealed the effect of spreading on aroma volatiles formation in postharvest leaves of green tea. *Food Res Int.* (2021) 149:110680. doi: 10.1016/j.foodres.2021.110680

40. Li Q, Chai S, Li YD, Huang JA, Luo Y, Xiao LZ, et al. Biochemical components associated with microbial community shift during the pile-fermentation of primary dark tea. *Front Microbiol.* (2018) 9:1509. doi: 10.3389/fmicb.2018.01509

41. Wang J, Zhang JW, Chen Y, Yu L, Teng JW, Xia N, et al. The relationship between microbial dynamics and dominant chemical components during liupao tea processing. *Food Biosci.* (2021) 2:101315. doi: 10.1016/j.fbio.2021.101315

42. Ho CT, Zheng X, Li SM. Tea aroma formation. *Food Sci Human Wellness*. (2015) 4:9–27. doi: 10.1016/j.fshw.2015.04.001

43. Parlapani FF, Ferrocino I, Michailidou S, Argiriou A, Boziaris IS. Microbiota and volatilome profile of fresh and chill-stored deepwater rose shrimp (*parapenaeus longirostris*). *Food Res Int.* (2020) 132:109057. doi: 10.1016/j.foodres.2020.109057

44. Li Y, Nishino N. Monitoring the bacterial community of maize silage stored in a bunker silo inoculated with *Enterococcus faecium*, *Lactobacillus plantarum* and *Lactobacillus buchneri*. J Appl Microbiol. (2011) 110:1561– 70. doi: 10.1111/j.1365-2672.2011.05010.x

45. Liu YJ, Han YJ, Peng WJ, Niu QS, Fang XM, Zhao YZ, et al. Analysis of microbial community diversity in bee bread by high-throughput sequencing. *Food Sci.* (2020) 41:94–100 (in Chinese).