



Unrelenting Fear Under Stress: Neural Circuits and Mechanisms for the Immediate Extinction Deficit

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Therapeutic interventions for disorders of fear and anxiety rely on behavioral approaches that reduce pathological fear memories. For example, learning that threat-predictive stimuli are no longer associated with aversive outcomes is central to the extinction of conditioned fear responses. Unfortunately, fear memories are durable, long-lasting, and resistant to extinction, particularly under high levels of stress. This is illustrated by the “immediate extinction deficit,” which is characterized by a poor long-term reduction of conditioned fear when extinction procedures are attempted within hours of fear conditioning. Here, I will review recent work that has provided new insight into the neural mechanisms underlying resistance to fear extinction. Emerging studies reveal that locus coeruleus norepinephrine modulates amygdala-prefrontal cortical circuits that are critical for extinction learning. These data suggest that stress-induced activation of brain neuromodulatory systems biases fear memory at the expense of extinction learning. Behavioral and pharmacological strategies to reduce stress in patients undergoing exposure therapy might improve therapeutic outcomes.

Keywords: extinction, stress, fear conditioning, norepinephrine, locus coeruleus, amygdala, infralimbic cortex, PTSD–posttraumatic stress disorder

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INTRODUCTION

Survival depends on detecting and defending against threats in the environment. These functions, shaped over millennia by natural selection, rely on specialized neural and behavioral adaptations. For example, when a rat detects a predator, it will draw from a repertoire of species-specific defense responses that include freezing, jumping, fleeing, and biting (Bolles, 1970). The nature of this response depends on several factors, particularly the proximity of the predator and the imminence of the threat (Fanselow and Lester, 1988; Perusini and Fanselow, 2015). Once danger has passed animals disengage from defense behaviors to resume other activities. But yielding to safety in the immediate aftermath of threat is risky business. Animals are justifiably conservative when it comes to the prospect of losing their lives. A rat can afford to lose a meal or two, but it cannot afford to be caught by a predator. Maintaining a defensive posture after a dangerous encounter is highly adaptive.

Of course, animals must not only defend against present danger, but they must also learn to anticipate future threats should they survive. Decades of work have described brain circuits that

rapidly encode and store memories of an aversive experience to ward off future threats (Davis, 1992; LeDoux, 2000; Maren, 2001; Maren and Quirk, 2004; Orsini and Maren, 2012; Maren et al., 2013; Herry and Johansen, 2014; Janak and Tye, 2015; Tovote et al., 2015). Learning is not only central to mobilizing defensive behavior to potential threats, but also in selecting the specific defensive responses an animal employs in a particular situation (Moscarello and Maren, 2018). In addition to learning about the circumstances that predict danger, animals also rapidly encode escape routes and shelters used to evade threats. Memories for the spatial location of safe havens allow for rapid escape responses when threat is encountered (Vale et al., 2017; Evans et al., 2018). Of course, learning that is reinforced by threat reduction should not interfere with consolidating memories for recently experienced threats. The brain systems and cellular mechanisms dedicated to storing fear memories should be resistant to disruption soon after a dangerous encounter. Surviving future threats requires that these memories be lifelong and indelible.

Several forms of learning are recruited to encode memories for aversive events. For example, intense noxious stimuli rapidly potentiate and sensitize behavioral responses they elicit (Borszcz et al., 1989; Boulis and Davis, 1989; Gewirtz et al., 1998). Sensitization is a non-associative form of learning that is short-lived (minutes to hours) and is not stimulus-specific: behavioral responses to many different innocuous stimuli are increased by sensitization (Groves and Thompson, 1970). Associative learning, on the other hand, results in relatively specific and durable long-term memories (hours to years) of aversive events and the circumstances (including cues and contexts) that predict them (Rescorla and Holland, 1982; Wasserman and Miller, 1997; Pearce and Bouton, 2001). An example of this form of learning is Pavlovian fear conditioning, in which animals acquire conditioned fear responses (CRs, such as freezing behavior) to innocuous conditioned stimuli (CSs, such as auditory tones) paired with aversive unconditioned stimuli (USs, such as a footshock). Pavlovian fear memories are rapidly acquired, exceptionally enduring, and highly resistant to disruption once learned. Not surprisingly, this type of learning has been the subject of intense investigation. It is not only essential to the adaptive function of defensive behavior systems, but also involved in the etiology of disorders of fear and anxiety in humans.

The durability of fear memory has been underscored by decades of work on another learning process: extinction. During experimental extinction procedures, CSs are repeatedly presented to the animal in the absence of the US (Pavlov, 1927; Konorski, 1967; Rescorla, 2000; Bouton et al., 2021). Consequently, animals learn that the CS no longer predicts the US, and this leads to a reduction in the frequency and vigor of conditioned responses. But the suppression of conditional responding after extinction is labile: CRs return under several conditions (Bouton, 1993; Bouton et al., 2006, 2021). For example, the mere passage of time leads to spontaneous recovery of extinguished CRs and presenting an extinguished CS outside the extinction context leads to a renewal of conditional responding. These relapse phenomena not only demonstrate that extinction memories are labile, but also that fear memories are very durable and difficult to disrupt (Goode and Maren, 2014, 2019; Goode et al., 2017).

Over 15 years ago, we discovered more evidence for the durability of fear memory. We found that extinction procedures performed soon after fear conditioning failed to yield long-term suppression of conditioned fear (Maren and Chang, 2006; Chang and Maren, 2009). We termed this phenomenon the “immediate extinction deficit” or IED. Considerable work has confirmed this finding and the IED has now been described in several species and paradigms (Schiller et al., 2008; Woods and Bouton, 2008; Huff et al., 2009; Kim et al., 2010; Golkar and Öhman, 2012; MacPherson et al., 2013; Stafford et al., 2013; Merz et al., 2016). Several lines of evidence suggest that the IED results from the stress associated with the conditioning procedure (Maren, 2014). Consistent with this possibility, deficits in “delayed” extinction learning have been reported after a variety of acute or chronic stressors (Izquierdo et al., 2006; Miracle et al., 2006; Chauveau et al., 2012; Knox et al., 2012; Maroun et al., 2013; Hartley et al., 2014; Maren and Holmes, 2016).

From an evolutionary perspective, resistance to extinction under stress serves to protect recently acquired fear memories from disruption and functions as a critical adaptation that permits animals to survive future threats. However, the resistance of fear memories to extinction poses challenges for therapeutic interventions for fear and anxiety disorders in humans. These interventions rely on extinction-based procedures such as prolonged exposure therapy and are susceptible to relapse (Norrholm et al., 2008; Kearns et al., 2012; Vervliet et al., 2013; Craske et al., 2018). For this reason, it is essential to understand the psychological and neurobiological mechanisms underlying stress-induced impairments in fear extinction. Here, I will review recent work on the neural and behavioral mechanisms of the immediate extinction deficit. This work reveals brain systems that operate to constrain extinction learning in the immediate aftermath of an aversive experience. Understanding the nature of these constraints on extinction learning has important implications for optimizing therapeutic interventions for humans, particularly soon after trauma.

RECENT FEAR IS RESISTANT TO EXTINCTION

It has long been known that memories are labile soon after they are encoded (Lechner et al., 1999; McGaugh, 2000). A wide variety of neural insults cause severe amnesia for events occurring minutes to hours (and occasionally months to years) prior to the injury. This provides strong support for the view that newly acquired information requires the passage of time to become consolidated as a long-term memory. Decades of work have elucidated the neural mechanisms of underlying memory consolidation and have shown that a host of cellular processes, including protein synthesis, are required to stabilize long-term memories within hours of their encoding (Schafe et al., 2001; Alberini, 2005; Redondo and Morris, 2011). Beyond this initial phase of cellular consolidation, brain systems that represent memory (including fear memories) also reorganize over time (Frankland and Bontempi, 2005; Tonegawa et al., 2018; Josselyn and Tonegawa,

2020). Both cellular and systems consolidation processes represent periods during which memories are vulnerable to disruption after encoding.

Because of these vulnerabilities, we imagined that behavioral manipulations, such as extinction, might be particularly effective if delivered shortly after conditioning. To test this hypothesis, we submitted male rats to an auditory fear conditioning procedure and followed that by an extinction procedure (45 CS-alone trials) either 15 min or 24 h after conditioning (Figure 1; Maren and Chang, 2006). Critically, we included control groups at each time point that did not undergo extinction training so that we could determine the relative effectiveness of each extinction procedure. Consistent with established work, we found that delivering extinction trials 24 h after conditioning (a delayed extinction procedure) was effective at suppressing conditioned freezing. However, delivering extinction trials 15 min after conditioning (an immediate extinction procedure) resulted in poor extinction retrieval. This immediate extinction deficit persisted for several hours after conditioning and was evident even after over 200 massed extinction trials. Although conditioned freezing declined during the extinction session, this reduction was found to be short-lived and likely due to habituation—conditioned freezing showed near complete recovery within 24 h of the immediate extinction procedure (Chang and Maren, 2009).

Additional experiments suggested that the footshock stress associated with the conditioning procedure was responsible for the IED. For example, we found that delivering unsignaled

footshock immediately prior to a delayed (24 h) extinction session impaired later extinction retrieval (Maren and Chang, 2006). Similarly, limiting conditioned freezing before an immediate extinction session by reducing the number of conditioning trials or US intensity eliminates the IED (Maren and Chang, 2006; Jo and Choi, 2020). These data suggest that the non-associative sensitization of fear associated with footshock may be a factor that contributes to the IED. Consistent with context-independence of sensitization (Davis, 1989), we have observed that the IED occurs both in the conditioning context as well as in different contexts in which non-associative freezing is expressed soon after conditioning (Maren and Chang, 2006; Chang and Maren, 2009). Although the majority of work on the IED has been performed in male rats, recent work indicates that the IED is comparable in male and female rats (Binette et al., 2022).

Although several groups have observed the IED, there are some notable exceptions. Using auditory fear conditioning and extinction procedures similar to those of Maren and Chang (2006) and Ponnusamy et al. (2016) found that immediate extinction procedures produced significant decreases in freezing relative to non-extinguished controls. Interestingly, baseline freezing prior to the first extinction trial was relatively low (and no different than that in the delayed control group). This suggests that the high level of arousal that has been suggested to be requisite for the IED as not obtained in this study. Archbold et al. (2010) also failed to obtain an IED after contextual fear conditioning, although this was primarily due to particularly weak extinction retention in the

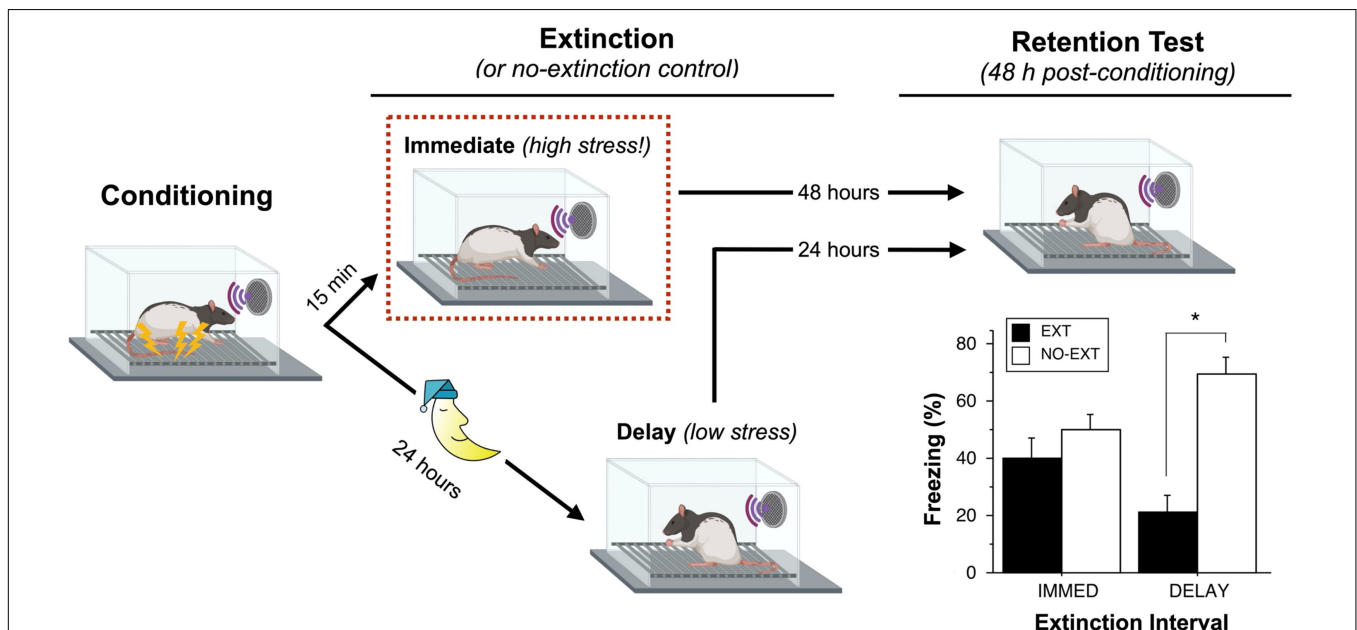


FIGURE 1 | Recent fear is resistant to extinction. Maren and Chang (2006) compared groups of rats that underwent an extinction (EXT) procedure either 15 min or 24 h after auditory fear conditioning; each of these groups was compared to a group of animals that did not undergo extinction (NO-EXT). During the extinction session, rats in the IMMED condition exhibit high levels of sensitized fear (high stress) prior to the first extinction trial, whereas rats in the DELAY condition exhibit low levels of freezing (low stress). Despite showing similar levels of within-session extinction, rats in the IMMED condition exhibit poor long-term retention of extinction relative to animals in the DELAY condition. Unlike DELAY rats, rats in the IMMED condition exhibited similar and high levels of conditioned freezing compared to non-extinguished controls. Elements of the figure were created with BioRender.com; the data shown in the plot were previously published (Maren and Chang, 2006).

delayed extinction condition. Lastly, Myers et al. (2006) also explored the effects of an immediate extinction procedure in a fear-potentiated startle paradigm in rats. Surprisingly, they found that delivering extinction trials 10 min after fear conditioning resulted in *weaker* relapse of conditioned fear. Relapse was indexed by the return of conditioned responding that is normally observed with the passage of time (spontaneous recovery), testing the CS outside the extinction context (renewal), or intervening exposure to the unconditioned stimulus (reinstatement). Based on these findings, the authors concluded that the immediate extinction procedure disrupted the consolidation of fear memory, produced long-term reductions in the expression of conditioned fear and eliminated fear relapse. However, close inspection of the data reveals that the lack of relapse observed after immediate extinction was not due to superior extinction, but just the opposite: conditioned fear was, in fact, resistant to immediate extinction. Because the animals did not acquire an extinction memory, they were unable to show relapse.

Interestingly, the IED has been observed after appetitive conditioning (Woods and Bouton, 2008) and in human fear conditioning experiments that use relatively weak unconditioned stimuli (Schiller et al., 2008; Huff et al., 2009; Merz et al., 2016) or non-emotional tasks (Merz and Wolf, 2019). These data have challenged the view that sensitization of fear in aversive conditioning experiments is necessary for the IED. However, it is possible that when extinction closely follows conditioning in the same physical context, the extinction trials are encoded within the interoceptive context of conditioning (produced by food or shock), a context that is absent at the time of extinction retrieval testing. This shift in interoceptive context might therefore lead to renewal of conditioned responding in animals that underwent immediate extinction or delayed extinction after a US reminder. However, matching the state of arousal driven by conditioning at the time of retrieval testing does not prevent expression of the IED (Woods and Bouton, 2008), although an experience with shock before extinction training reduces fear suppression as previously observed (Maren and Chang, 2006). These data suggest that the relative recency of conditioning to post-extinction retrieval testing (Devenport, 1998) does not account for the IED. An alternative idea is that the temporal gap between fear conditioning and extinction serves as an event boundary that segments and reduces interference between the two contingencies (Devenport, 1998; Dunsmoor et al., 2018). To test this hypothesis, we examined whether delivering extinction trials within seconds of conditioning (using the same ISI at which conditioning trials were delivered) would reduce the IED (Totty et al., 2019). However, we found that the continuous extinction procedure yielded a robust IED suggesting that a temporal gap between conditioning and extinction is not responsible for the IED.

Although the precise behavioral mechanisms underlying the IED are still uncertain, the existing data are most compatible with the possibility that conditioning results in emotional arousal or stress (whether aversive or appetitive) that impedes extinction learning. This conclusion is perhaps most strongly supported by recent data showing that pharmacological treatments that

limit stress-sensitive neuromodulatory circuits in the brain attenuate the IED.

STRESS MODULATES NEURAL CIRCUITS CRITICAL FOR EXTINCTION LEARNING

Extinction procedures engage multiple neural circuits that mediate not only the retrieval and expression of conditioned responses, but also the encoding of prediction errors and contextual stimuli associated with the extinction contingency (Bouton et al., 2006, 2021; Radulovic and Tronson, 2010; Maren et al., 2013). Brainstem circuits are critical for encoding prediction errors that drive extinction learning and they convey this information to forebrain circuits that encode context-dependent extinction memories (McNally et al., 2011; Iordanova et al., 2021). A critical hub for extinction learning is centered in the medial prefrontal cortex (mPFC) and its connections with the hippocampus (HPC) and amygdala (Maren and Quirk, 2004; Quirk and Mueller, 2008; Orsini and Maren, 2012; Maren et al., 2013; Giustino and Maren, 2015; Bouton et al., 2021). Specifically, projections from the infralimbic (IL) division of the mPFC to the basolateral amygdala (BLA) and amygdala intercalated cells are implicated in the acquisition and expression of extinction (Likhtik et al., 2005, 2008; Bukalo et al., 2015; Do-Monte et al., 2015; Bloodgood et al., 2018; Hagihara et al., 2021). IL projections to the HPC via the thalamic nucleus reuniens are also critical for extinction learning and retrieval (Ramanathan et al., 2018; Ramanathan and Maren, 2019). In turn, reciprocal projections within these networks are involved in extinction retrieval and the suppressing of conditional responding (Senn et al., 2014; Jin and Maren, 2015; Marek et al., 2018; Silva et al., 2021).

Considerable work indicates that both acute and chronic stressors regulate extinction learning by modulating neural activity within hippocampal-prefrontal-amygdala networks (Maren and Holmes, 2016; Singewald and Holmes, 2019). Aversive stressors activate central and peripheral hormonal and neuromodulatory systems that drive sympathetic arousal and modulate activity within this stress-sensitive network. This response is coordinated by the hypothalamic-pituitary-adrenal axis, which regulates the release of adrenal glucocorticoids, and the brainstem locus coeruleus (LC), which is the primary source for forebrain norepinephrine (NE) (Jones and Moore, 1977; Lipski and Grace, 2013; Aston-Jones and Waterhouse, 2016; Poe et al., 2020; Ross and Van Bockstaele, 2020). Not surprisingly, both of these neuromodulatory systems have been implicated in aversive conditioning and extinction (Pugh et al., 1997; Korte, 2001; Rodrigues et al., 2009; Giustino et al., 2016b; Giustino and Maren, 2018; de Quervain et al., 2019; Likhtik and Johansen, 2019). For example, it is well-established that aversive footshock increases both Fos expression and spike firing in LC neurons (Rasmussen et al., 1986; Abercrombie and Jacobs, 1987; Grant et al., 1992; Chiang and Aston-Jones, 1993; Rassnick et al., 1998), as well as increasing NE release in forebrain efferents, including the amygdala (Galvez et al., 1996; Quirarte et al., 1998) and mPFC (Gresch et al., 1994; Goldstein et al., 1996;

Jedema et al., 1999). Indeed, the role for stress-induced activation of forebrain noradrenergic activity in promoting the acquisition and consolidation of aversive learning and memory has long been appreciated (Ferry et al., 1999; Johansen et al., 2014; McGaugh, 2015; Díaz-Mataix et al., 2017; Schiff et al., 2017; Giustino and Maren, 2018). However, other forms of learning are opposed by stress, including Pavlovian extinction (Raio and Phelps, 2015; Maren and Holmes, 2016; Singewald and Holmes, 2019).

This paradox might be explained by the non-linear (inverted-U) function relating noradrenergic activity to learning and memory (Gold et al., 1975; Snyder et al., 2012). Either abnormally low or excessive activation of the LC-NE system can impair learning and memory processes, including extinction (Arnsten, 2009; Fitzgerald et al., 2015; Giustino and Maren, 2018). From an evolutionary perspective, the LC-NE system has been adapted to both promote the rapid encoding of aversive memories under stress and enable the extinction of those memories when danger has passed. By this view, excessive activation of the LC-NE system under high levels of stress promotes fear memory encoding at the expense of extinction, whereas modest levels of LC-NE system engagement under relatively low-stress conditions promotes extinction learning (Giustino et al., 2016b; Giustino and Maren, 2018).

NORADRENERGIC ANTAGONISTS RESCUE THE IMMEDIATE EXTINCTION DEFICIT

Footshock stress results in sustained LC activation and NE release in forebrain regions critical for fear conditioning and extinction. Importantly, shock-induced activation of the LC-NE system may undermine extinction learning in the immediate aftermath of fear conditioning, resulting in the IED. Consistent with this idea, there is extensive evidence that β -adrenergic receptor antagonists, such as propranolol, reduce stress-induced fear responses in both rodents and humans (Cole and Koob, 1988; Fitzgerald et al., 2014; Chalkia et al., 2019). Systemic administration of propranolol reduces the expression of conditioned freezing (Rodríguez-Romaguera et al., 2009; Fitzgerald et al., 2015; Giustino et al., 2020; Leal Santos et al., 2021) and reduces increases in conditioned fear induced by corticotropin-releasing factor (CRF) (Cole and Koob, 1988). If a high fear state associated with recent footshock opposes extinction, then pharmacologically limiting this state with systemic propranolol should attenuate the IED and facilitate extinction under stress.

To explore this possibility, we administered propranolol immediately after auditory fear conditioning in male rats undergoing an immediate extinction procedure in which CS-alone trials were presented 15 min after the last conditioning trial (Fitzgerald et al., 2015). Consistent with previous reports, we found that systemic propranolol reduced post-shock freezing in the interval prior to extinction training. In addition to a reduction in post-shock freezing, propranolol facilitated the reduction in freezing behavior that occurred during the immediate extinction

procedure. Importantly, this facilitation of extinction was long-lasting. Although control rats undergoing immediate extinction exhibited a near-complete recovery of conditioned freezing 24 h later, rats receiving propranolol exhibited a robust attenuation of conditioned freezing the following day. The facilitation of extinction under stress was not due to a propranolol-induced impairment in the consolidation of the conditioned fear memory. Rats that received immediate post-shock propranolol in the absence of extinction training showed high levels of CS-elicited freezing that were not different from vehicle-treated controls (Fitzgerald et al., 2015). This reveals that propranolol resulted in long-term decreases in freezing by augmenting extinction learning, rather than impairing the consolidation of fear conditioning. It is important to note that although propranolol administration facilitated immediate extinction, we also found that it impaired delayed extinction (Mueller et al., 2008; Uematsu et al., 2017). This suggests that noradrenergic transmission may be involved in promoting extinction learning during basal, “low stress” conditions (delayed extinction), while it undermines extinction under high levels of stress (immediate extinction). Of course, this begs the question: how does propranolol rescue the immediate extinction deficit?

Decades of work have revealed that the IL cortex, a mPFC region that is interconnected with both the HPC and amygdala, is a critical neural substrate underlying the extinction of conditioned fear (Maren and Quirk, 2004; Delgado et al., 2006; Quirk et al., 2006; Quirk and Mueller, 2008; Giustino and Maren, 2015). Neuronal activity within the IL correlates with suppression of conditioned freezing (Milad and Quirk, 2002; Chang et al., 2010; Wilber et al., 2011; Giustino et al., 2016a, 2019), electrical or optogenetic activation of the IL can facilitate extinction learning (Milad et al., 2004; Do-Monte et al., 2015) and pharmacological or optogenetic inhibition of neuronal activity in the IL impairs extinction learning (Laurent and Westbrook, 2009; Sierra-Mercado et al., 2011; Do-Monte et al., 2015). In addition, chronic stress remodels (Izquierdo et al., 2006; Miracle et al., 2006) and reduces the excitability (Wilber et al., 2011) of IL principal neurons causing impairments in forming long-term extinction memories, particularly when the stress occurs prior to fear conditioning (Chakraborty and Chattarji, 2019). Importantly, both synaptic and dendritic changes can occur within hours of an acute stressor (Chen et al., 2008; Yuen et al., 2009; Nava et al., 2017), which suggests that suppression of IL activity by footshock may cause the IED. To test this hypothesis, we explored spontaneous single-unit firing in the mPFC after auditory fear conditioning in rats. We found that conditioning induced a dramatic, but short-lived, increase in spontaneous spike firing in both the prelimbic (PL) and IL cortices (Fitzgerald et al., 2015). Interestingly, this conditioning-induced potentiation of spike firing was followed by an extended suppression of neuronal activity in the IL (Fitzgerald et al., 2015; Giustino et al., 2016a). This suppression of spontaneous firing in the IL was coincident with the emergence of post-shock freezing behavior and was maximal roughly 30 min after the final footshock (a period during which the IED is readily obtained); it was also most robust following signaled shocks (Fitzgerald et al., 2015; Giustino et al., 2016a). Suppression of spontaneous spike firing

persisted for up to 60 min (the duration of the recording session) in many IL units.

Because systemic propranolol administration rescues the IED, we hypothesized that it might promote extinction under stress by regulating post-shock spike firing in the mPFC. Consistent with this possibility, rats that received systemic propranolol injections prior to auditory fear conditioning exhibited much smaller perturbations of mPFC spike firing than vehicle-treated rats (Fitzgerald et al., 2015). Specifically, propranolol blunted both the immediate post-shock increases in PL and IL spike firing and eliminated the sustained suppression of spike firing observed in the IL during the 60-min post-shock period. These changes in spike firing were paralleled by reductions in post-shock freezing in propranolol-treated rats. Importantly, however, changes in mPFC spike firing were dissociable from freezing behavior. Auditory CSs produced dramatic increases in conditioned freezing behavior 24 h after conditioning, but this was not associated with the same increases in mPFC spike firing that were previously observed immediately after footshock (Fitzgerald et al., 2015). Together, these results suggest that stress-induced activation of the LC-NE system suppresses the activity of a key cortical substrate involved in extinction learning. The post-shock suppression of spontaneous spike firing in the IL appears to underlie, at least in part, the IED.

NEURAL CIRCUIT FOR NORADRENERGIC REGULATION OF EXTINCTION LEARNING UNDER STRESS

Systemic propranolol administration may regulate neuronal activity by direct effects on β -adrenergic receptors in IL or by affecting a brain region that regulates IL activity. For example, it is well-known that NE promotes aversive memory formation through its actions in the BLA, which, in turn, projects to and regulates IL function (Floresco and Tse, 2007; Dilgen et al., 2013; McGarry and Carter, 2016). Indeed, BLA projections to IL have been implicated in extinction learning (Senn et al., 2014). To explore whether β -adrenergic receptors in IL or BLA (or both) are involved in the IED, we made intracranial infusions of propranolol into these structures immediately after fear conditioning, a timepoint that was roughly 20 min prior to an immediate extinction procedure (Giustino et al., 2017). Unlike systemic propranolol infusions, neither IL nor BLA infusions reliably decreased post-shock freezing or freezing behavior during the immediate extinction procedure (Giustino et al., 2017). Interestingly, recent work suggests that noradrenergic regulation of post-shock freezing behavior may depend on LC projections to the central nucleus of the amygdala (CEA) (Gu et al., 2020). Importantly, intra-BLA propranolol infusions, but not infusions into the IL, promoted long-term extinction of conditioned freezing and reduced the IED. Rats treated with intra-BLA propranolol exhibited significantly lower levels of CS-evoked freezing during the extinction retrieval test 24 h after the immediate extinction

procedure. This effect was not due to an effect of BLA propranolol on the consolidation of conditioned fear memory, because immediate post-shock intra-BLA propranolol infusions had no effect on the long-term retrieval of conditioned fear in the absence of extinction (Giustino et al., 2017). Hence, local antagonism of β -adrenergic receptors in the BLA, but not the IL, rescues the IED.

The BLA receives dense noradrenergic innervation from the LC (Jones and Moore, 1977; Asan, 1998; Zhang et al., 2013). This suggests that stress-induced activation of LC-NE projections to the BLA may be critical for regulating the IED. To test this hypothesis, we explored whether footshock stress alters neuronal activity in the BLA and whether these effects are modulated by the LC and depend on noradrenergic receptors in the BLA. In contrast to our previous observations in the mPFC, fear conditioning produced dramatic and sustained increases in the spontaneous activity of BLA neuronal activity (Giustino et al., 2020). Importantly, this shock-induced increase in BLA spike firing was completely attenuated by systemic propranolol administration. To determine whether BLA spike firing is modulated by LC activity, we expressed an excitatory “designer receptor exclusively activated by designer drug” (DREADD; AAV-PR_Sx8-hM3Dq) in LC noradrenergic neurons. We then chemogenetically activated these neurons with the synthetic DREADD ligand clozapine-*N*-oxide (CNO) and coupled activation of LC with the delivery of weak footshock. Weak footshocks alone produced minimal increases in BLA spike firing that were dramatically potentiated when delivered concomitant with chemogenetic activation of LC-NE neurons (Giustino et al., 2020). This suggests that shock-elicited NE release drive increases in BLA firing in behaving rats. Although the LC-NE system has been reported to have primarily inhibitory effects on BLA spike firing (Chen and Sara, 2007), LC stimulation and iontophoretic application of NE can increase spiking in a subset of BLA neurons (Buffalari and Grace, 2007). Moreover, fear conditioning increases BLA excitability by reducing NE-driven GABAergic inhibition of BLA neurons without affecting noradrenergic facilitation of BLA excitability (Skelly et al., 2017, 2016). These results suggest that sustained footshock-induced activation of LC leads to a persistent increase in BLA excitability that underlies the IED.

Having identified a role for LC-NE activation in shock-induced increases in BLA neuronal firing, we next explored the contribution of the LC to the IED. We reasoned that an IED would not normally be induced by conditioning with weak footshocks, but that this deficit would become apparent after pairing a weak conditioning protocol with chemogenetic LC activation. Chemogenetic LC activation robustly increased conditioned freezing during the immediate extinction procedure as previously observed (Giustino et al., 2020). Consistent with our hypothesis, we also found that chemogenetic activation of the LC promoted an IED under conditions that do not typically lead to a stress-induced extinction deficit. Specifically, rats treated with vehicle after the immediate extinction procedure showed reliable reductions in conditioned freezing during a drug-free extinction retrieval test (relative to non-extinguished controls). This revealed that the weak conditioning procedure

failed to produce an IED. In contrast, CNO-treated rats undergoing the immediate extinction procedure exhibited poor extinction retrieval (high freezing) and were no different than non-extinguished controls during the drug-free retrieval test (Giustino et al., 2020). Hence, chemogenetic activation of the LC-NE system drove an extinction learning deficit under weak conditioning procedures that are not normally sufficient to yield the IED.

Finally, we sought to determine whether the induction of an IED by LC-NE activation was mediated by β -adrenergic receptors on the BLA. To this end, we infused vehicle or propranolol into the BLA prior to systemic CNO (or vehicle) injections in rats expressing excitatory DREADDs in the LC (Giustino et al., 2020). Animals then underwent a weak conditioning protocol followed by an immediate extinction procedure. The animals that received CNO infusions exhibited a robust IED as previously observed. Importantly, this CNO-induced IED was blocked by intra-BLA propranolol infusions. Propranolol infusions alone did not affect conditional freezing. Collectively, these data suggest that the IED is mediated by stress-induced activation of LC-NE afferents in the BLA, which in turn potentiates BLA firing and impedes the extinction of fear. Moreover, these results suggest that BLA projections to the IL may play a critical role in the shock-induced changes in IL spike firing that are associated with the IED. Indeed, IL principal neurons experience strong feed-forward inhibition by cortical interneurons after the activation

of excitatory BLA afferents (Floresco and Tse, 2007; Dilgen et al., 2013; McGarry and Carter, 2016). These results suggest a model whereby acute stressors activate LC-NE projections to BLA, thereby elevating BLA activity to promote the stabilization of fear memory. We propose that stress-induced increases in BLA firing yield a sustained inhibition of neuronal activity in the IL and impair extinction of fear memories (Figure 2).

CORTICOTROPIN-RELEASING FACTOR SYSTEM MODULATES NEURAL CIRCUITS UNDERLYING THE IMMEDIATE EXTINCTION DEFICIT

In addition to the strong efferent projections to the BLA, the LC-NE system also receives a potent descending projection from the CEA (Van Bockstaele et al., 1998; Ross and Van Bockstaele, 2020). Importantly, footshock increases neuronal activity in the CEA (Radulovic et al., 1998; Yu et al., 2017; Giovanniello et al., 2020) and optogenetic activation of CEA induces high tonic firing rates in LC neurons (McCall et al., 2015). Considerable work indicates that CEA-mediated increases in LC spike firing are mediated by the neuropeptide hormone, CRF (McCall et al., 2015). Hence, reciprocal projections between the LC and amygdala function to promote feed-forward excitation within these brain areas.

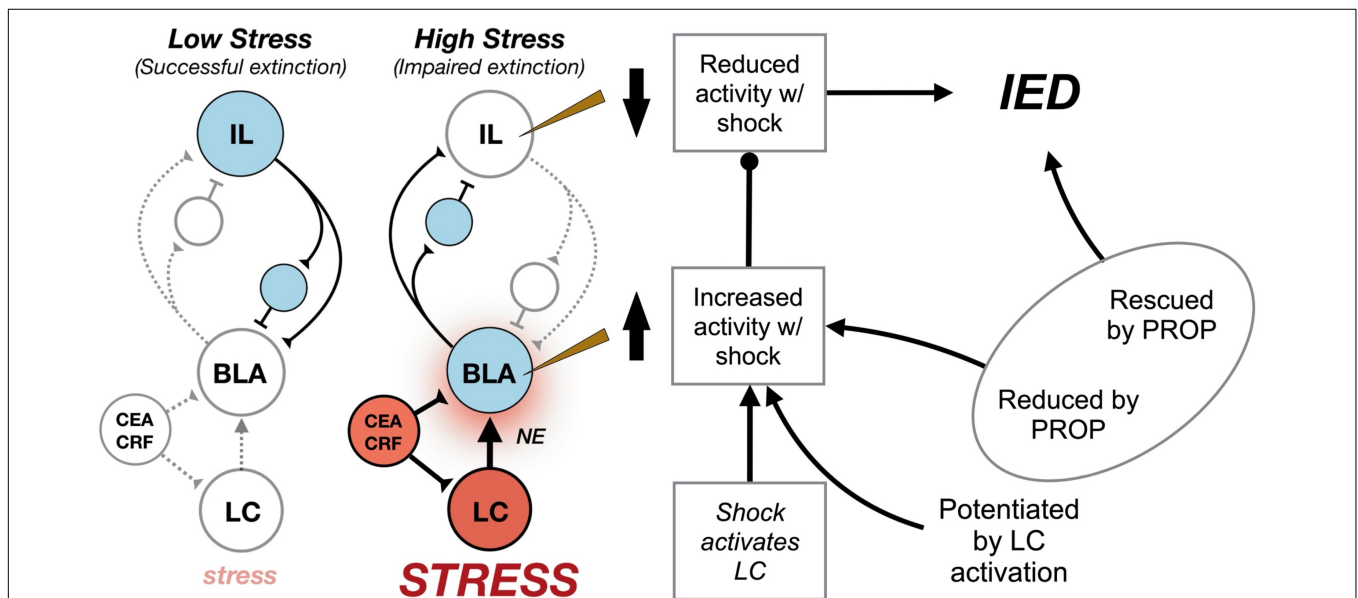


FIGURE 2 | Circuit model for the immediate extinction deficit (IED). Under basal conditions (“Low Stress”) delayed extinction procedures conducted 24 h after fear conditioning recruit infralimbic (IL) cortical circuits (blue) that mediate the acquisition and expression of extinction learning. Inhibition of conditioned fear is presumed to arise from IL-mediated excitation of inhibitory interneurons (small circles) that reduce the excitability of basolateral amygdala (BLA) principal cells representing fear memories. However, delivering extinction trials soon after fear conditioning, when animals are under extreme stress (“High Stress”) results in activation of locus coeruleus (LC) noradrenergic neurons (red) that release norepinephrine (NE) in forebrain targets, including the BLA (blue). Neurons in the central amygdala (CEA) release corticotropin-releasing hormone (CRF) in the LC and BLA to facilitate stress-induced activation of these brain areas. Consequently, fear conditioning dramatically increases spontaneous spike firing in the BLA, while decreasing spike firing in IL. We speculate that BLA decreases IL spike firing by activating IL interneurons (small circles) and driving feed-forward inhibition in IL principal cells. Shock-induced increases in BLA firing are modulated by the LC, and LC activation during weak shock enables the IED when it would not normally occur. Systemic or intra-BLA administration of the β -adrenergic antagonist, propranolol (PROP), attenuates the IED and limits shock-elicited changes in BLA and IL spike firing (Fitzgerald et al., 2015; Giustino et al., 2016a, 2017, 2020).

Specifically, LC activation drives NE-dependent increases in BLA and CEA excitability and CEA-CRF neurons (which are also excited by BLA afferents) increase the excitability of LC projection neurons (Van Bockstaele et al., 1998; Valentino and Van Bockstaele, 2008).

The excitatory effect of the CEA on tonic LC-NE firing suggests that CEA-CRF neurons may be involved in the IED. To explore this issue, Jo et al. (2020) used transgenic mouse models to either silence CEA-CRF neurons or inactivate CRF production from these cells. In both cases, silencing CRF neurons or inactivating CRF attenuated the IED: mice exhibited normal conditioning and within-session extinction of conditioned freezing but failed to exhibit long-term suppression of freezing behavior. Temporary chemogenetic inhibition of CEA-CRF neurons also facilitated immediate extinction, whereas CNO-mediated excitation of these neurons produced deficits in a normally effective delayed extinction procedure. Interestingly, the activity of CEA-CRF neurons differed under immediate and delayed extinction procedures. Recordings of calcium signals in CEA-CRF neurons (expressing GCaMP6m) revealed elevated and sustained CS-induced responses in mice undergoing immediate extinction. Interestingly, elevated CS-evoked responses in animals undergoing immediate extinction persisted during the extinction retrieval test (Jo et al., 2020). Although the spontaneous activity of CEA-CRF neurons was not quantified in non-extinguished animals, it is intriguing to speculate that footshock stress might tonically elevate the activity of these cells. Nonetheless, these data suggest that increased CS-elicited activity in CEA-CRF neurons may drive the IED. Consistent with this, Jo et al. (2020) found that optogenetic inhibition of CEA-CRF neurons during the CS also attenuated the IED. Although not directly tested, these data suggest that CEA-CRF projections to the LC may be involved in the IED.

The stress-induced activation of CEA has also been suggested to increase CRF levels in the BLA (Roozendaal et al., 2002). Moreover, the BLA has a high density of CRF receptors (De Souza et al., 1985) and CRF increases the excitability of BLA neurons (Rainnie et al., 1992). Hence, it is possible that CRF drives increases in BLA excitability after footshock stress (Giustino et al., 2020) and contributes to the IED. To test this hypothesis, Hollis et al. (2016) infused a CRFR1 antagonist into the BLA prior to an immediate extinction procedure in rats. The drug dose-dependently attenuated the IED indexed by conditioned freezing during a drug-free extinction retrieval test 24 h after extinction (Hollis et al., 2016). This effect was not due to a suppression of freezing *per se* insofar as the drug did not affect freezing during the immediate extinction session. Moreover, a no-extinction control group revealed that the drug did not exert its effect by disrupting the consolidation of fear memory. In contrast, local infusion of a CRF agonist into the BLA prior to a delayed extinction procedure results in an extinction impairment (Abiri et al., 2014; Hollis et al., 2016). Interestingly, CRF was shown to exert its actions in the BLA through the modulation of the phosphorylation state of AMPA-type glutamate receptors (Hollis et al., 2016). This suggests that an attenuation of local synaptic plasticity in the

BLA contributes to the IED. Together, these results suggest that CRF has both local and circuit effects that contribute to shock-induced increases in BLA excitability that drive the IED. Indeed, CRF may act synergistically to antagonize extinction learning soon after fear conditioning by both direct actions on BLA CRF receptors and indirect activation of LC-NE projections to the BLA. An interesting area for future work is to determine how sex differences in CRH regulation of LC function (Bangasser et al., 2010; Bangasser and Cuarenta, 2021) affect stress-induced impairments in extinction learning.

CONCLUSION

Fear conditioning is an adaptive form of learning that allows an animal to mobilize defensive behaviors in anticipation of threat. Aversive memories are rapidly encoded, long-lasting, and susceptible to relapse after conditioning. Critically, fear memories are highly resistant to extinction soon after they are encoded, resulting in an IED when extinction trials are delivered within hours of fear conditioning. A growing body of evidence reveals that the IED is mediated by the recruitment of a stress-sensitive network that prioritizes the encoding and consolidation of fear memory even when the prevailing threat has passed. Stress-induced impairments in extinction learning involve activation of the LC-NE system, which appears to regulate amygdalo-prefrontal circuits involved in extinction learning. Specifically, we propose that noradrenergic activation of the BLA recruits mPFC-projecting neurons that functionally inhibit IL principal cells involved in extinction learning. We imagine that this is supported by BLA afferents on mPFC interneurons, which provide potent feed-forward inhibition of mPFC principal cells. In addition to the LC-NE system, CRF-containing neurons in the CEA play a critical role in the IED. Descending CEA→LC projections may amplify shock-induced increases in LC excitability, potentiate NE effects on BLA principal cells, and drive feed-forward excitation of BLA neuronal activity that protects fear memory from extinction under stress.

The role for the LC-NE system in mediating resistance to extinction under stress has important implications for understanding disorders of fear and anxiety (Fitzgerald et al., 2014; Giustino et al., 2016b). Patients with post-traumatic stress disorder (PTSD), for example, exhibit dysregulated noradrenergic neurotransmission (Grillon et al., 1996; Southwick et al., 1999; Geraciotti et al., 2001; Pitman et al., 2012) and resistance to extinction (Wessa and Flor, 2007; Milad et al., 2009; VanElzaker et al., 2014). Importantly, noradrenergic hyperarousal may impede therapeutic interventions, such as prolonged exposure therapy, that rely upon extinction learning to reduce pathological fear (McCurry et al., 2020). There is evidence that treatment with propranolol after reactivating trauma memory can reduce symptom severity in patients with PTSD (Vaiva et al., 2003; Hoge et al., 2012; Brunet et al., 2018) or specific phobias (Soeter and Kindt, 2015). Potential mechanisms for these effects include the suppression of fear expression, facilitation of extinction, or impaired reconsolidation of fear memories (Grillon et al., 2004; Kindt et al., 2009;

Kroes et al., 2010, 2016; Maren, 2011; Soeter and Kindt, 2015; Chalkia et al., 2019). Preclinical work on the IED suggests that propranolol might be most effective in patients with noradrenergic hyperarousal, particularly when combined with cognitive-behavioral therapy. Ultimately, deciphering the neural circuits and cellular mechanisms underlying stress-impaired extinction offers considerable promise for understanding both adaptive regulation of emotional behavior and pathological disorders of fear and anxiety.

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AUTHOR CONTRIBUTIONS

SM wrote the manuscript.

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