



Emerging Roles of Neuronal Ca²⁺ Sensor-1 in Cardiac and Neuronal Tissues: A Mini Review

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The EF-hand calcium (Ca²⁺)-binding protein, neuronal Ca²⁺ sensor-1 (NCS-1/frequenin), is predominantly expressed in neuronal tissues and plays a crucial role in neuronal functions, including synaptic transmission and plasticity. NCS-1 has diverse functional roles, as elucidated in the past 15 years, which include the regulation of phosphatidylinositol 4-kinase IIIβ (PI-4K-β) and several ion channels such as voltage-gated K⁺ and Ca²⁺ channels, the D2 dopamine receptors, and inositol 1,4,5-trisphosphate receptors (InsP₃Rs). Functional analyses demonstrated that NCS-1 enhances exocytosis and neuronal survival after injury, as well as promotes learning and memory in mice. NCS-1 is also expressed in the heart including the Purkinje fibers (PFs) of the conduction system. NCS-1 interacts with K_v4 K⁺ channels together with dipeptidyl peptidase-like protein-6 (DPP-6), and this macromolecule then composes the transient outward current in PFs and contributes to the repolarization of PF action potential, thus being responsible for idiopathic arrhythmia. Moreover, NCS-1 expression was reported to be significantly high at the immature stage and at hypertrophy in adults. That report demonstrated that NCS-1 positively regulates cardiac contraction in immature hearts by increasing intracellular Ca²⁺ signals through interaction with InsP₃Rs. With the related signals, NCS-1 activates nuclear Ca²⁺ signals, which would be a mechanism underlying hormone-induced cardiac hypertrophy. Furthermore, NCS-1 contributes to stress tolerance in cardiomyocytes by activating mitochondrial detoxification pathways, with a key role in Ca²⁺-dependent pathways. In this review, we will discuss recent findings supporting the functional significance of NCS-1 in the brain and heart and will address possible underlying molecular mechanisms.

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NCS-1 AND ITS INTERACTING PROTEINS

Intracellular calcium (Ca²⁺) is a versatile second messenger that regulates diverse cellular processes, including neurotransmission, muscle contraction, and signal transduction. Changes in intracellular Ca²⁺ are transduced by multiple proteins, with a key role of a large family of EF-hand Ca²⁺-binding proteins that act as Ca²⁺ sensors or Ca²⁺ buffers. Ca²⁺-buffer proteins chelate Ca²⁺ and

often terminate Ca^{2+} signals (Ikura, 1996). In contrast, the binding of Ca^{2+} to Ca^{2+} -sensor proteins causes a large conformational change, which consequently transduces the Ca^{2+} signal into various cellular functional changes, by regulating specific target proteins. Calmodulin is one of the best-characterized Ca^{2+} -sensor proteins and is involved in many aspects of Ca^{2+} signaling. Neuronal calcium sensor-1 (NCS-1) is the mammalian homolog of the *Drosophila* frequenin protein, which belongs to the larger NCS protein family that includes NCS-1, visinin-like proteins, recoverin, guanylate cyclase-activating proteins, and potassium channel-interacting proteins (KChIPs). The structure of NCS-1 is shown in the **Figure 1A**, indicating that it is a small (22 kDa) Ca^{2+} -binding protein containing 4 EF-hand motifs; of these, 3 (EF2-4) bind to Ca^{2+} . Unlike ubiquitously expressed calmodulin, NCS-1 is predominantly expressed in the brain and cardiac tissues, suggesting its specialized roles in these tissues. The Ca^{2+} -binding affinity of NCS-1 is significantly higher than that of calmodulin (K_d values of ~ 300 nM vs. ~ 10 μM , respectively). Although both Ca^{2+} -binding proteins can operate within the physiological range of Ca^{2+} levels (~ 100 nM to ~ 1 – 5 μM), the above data suggest that NCS-1 may be more sensitive to small changes in intracellular Ca^{2+} .

The functional roles of NCS-1 are still being elucidated. Currently, known functions include the regulation of diverse target proteins, including phosphatidylinositol 4-kinase (PI-4K), voltage- and ligand-gated ion channels, and interleukin-1 receptor accessory protein like-1 (IL1RAPL1). Here, we summarize the current understanding of NCS-1 regulation of some of these target proteins and how it affects brain and cardiac functions (**Table 1**). Even though NCS-1 has a well-characterized function in these organ systems, it should be pointed out that NCS-1 may have more diverse functions in human physiology and disease, such as a potential role in oncogenesis (Jerng et al., 2004). Such roles should be the focus of future studies.

Phosphatidylinositol 4-Kinase

Role in Exocytosis and Secretion

PI-4K III β (PI-4K- β) catalyzes the synthesis of phosphatidylinositol 4-phosphate, which is a late limiting step in the synthesis of phosphatidylinositol 4,5-bisphosphate, an important lipid regulator of many cellular functions including exocytosis. A yeast homolog of NCS-1 and PI-4-K interact, and NCS-1-induced activation of PI-4-K is required for yeast survival (Hendricks et al., 1999). Structural support for interaction was obtained from a recent NMR structure of Ca^{2+} -bound yeast NCS-1 (Ncs1) in complex with an N-terminal yeast PI-4-K (Pik1) fragment (Strahl et al., 2007). This interaction was also detected in neuroendocrine cells (Koizumi et al., 2002; Scalettar et al., 2002; Rajebhosale et al., 2003; de Barry et al., 2006), neurons (Taverna et al., 2002; Zheng et al., 2005) and other cell types including pancreatic beta cells (Gromada et al., 2005) and mast cells (Kapp-Barnea et al., 2003), and was shown to facilitate exocytosis and secretion in these cells (**Table 1**). However, previous reports have suggested no direct interaction in neurons (Bartlett et al., 2000).

This contradiction may be explained by the presence of newly discovered PI-4-K β regulators, calneurons. While calneurons interact with PI-4K- β at low Ca^{2+} levels to inhibit its enzyme activity, NCS-1 binds to PI-4K- β at high Ca^{2+} levels to activate it (Mikhaylova et al., 2009), suggesting that calneurons and NCS-1 compete for PI-4-K- β interaction depending on intracellular Ca^{2+} levels. Thus, when the intracellular Ca^{2+} level is low, its interaction might be difficult to be detected.

Voltage-Gated K_V4 K^+ Channels

Regulation of Excitability in the Brain

The V7 *Drosophila* mutant that overexpresses NCS-1 has a phenotype of hyperactivity, which results in the proposal that NCS-1 facilitates neurotransmission, possibly by regulating the activities of ion channels (Pongs et al., 1993; Poulain et al., 1994). Indeed, we found that NCS-1 is a Ca^{2+} -sensitive regulatory component of a native K^+ current (Nakamura et al., 2001; **Table 1**). In the brain and heart, rapidly inactivating (A-type) voltage-gated K^+ currents control cellular excitability. Although the pore-forming alpha-subunits of these channels are considered to be K_V4 channels (Serôdio et al., 1994; Fiset et al., 1997; Nakamura et al., 1997), the kinetic properties of K_V4 channels differ from native A-type currents, suggesting the presence of regulatory subunits. KChIPs, a member of the NCS protein subfamily, were initially reported as a specific K_V4 regulatory subunit (An et al., 2000). Because NCS-1 modulates K_V4 currents similar to KChIPs, by increasing current amplitude and slowing the inactivation time course, and NCS-1 physically interacts with $\text{K}_V4.2$ in mouse brain, it was identified as a regulator of A-type K^+ currents in neurons (Nakamura et al., 2001; **Table 1**).

Involvement in Cardiac Arrhythmia

This interaction and activation also occurs in adult mouse cardiomyocytes (Guo et al., 2002) and in zebrafish heart (Nakamura and Coetzee, 2008), which lacks KChIPs. The differential regulation of K_V4 channels by NCS-1 and KChIPs in specific tissues and cell types was an unaddressed topic, and this was clearly demonstrated in the report by Nattel's group (Xiao et al., 2013). Purkinje fibers (PFs) show an unusual form of transient outward K^+ current I_{to} with slow recovery kinetics and TEA sensitivity compared with ventricular I_{to} , suggesting a distinct molecular composition. This group found that NCS-1 and DPP6, which were also reported to be auxiliary subunits of K_V4 K^+ channels (Jerng et al., 2004), are preferentially enriched in PFs, while KChIP2, an essential subunit of ventricular $\text{K}_V4.3$ is weakly expressed. Moreover, NCS-1 slowed inactivation kinetics of $\text{K}_V4.3$, while DPP6 increased its current amplitude, thus increasing the I_{to} -mediated K^+ efflux (**Figure 1Ba**), which would accelerate PF repolarization and shortening of action potentials (**Figure 1Bb**; similar computer simulation was reported by Xiao et al., 2013). Thus, overexpression of K_V4 auxiliary subunits may result in steep transmural repolarization gradients in PFs with adjacent ventricular tissues that induces coupled ectopic activity, and potentially leads to lethal arrhythmias (**Figure 1Bc**). NCS-1

also interacts with the anti-cancer drug taxol (Boehmerle et al., 2006), and is involved in the regulation of taxol-induced cardiac arrhythmia (Zhang et al., 2010). Thus, NCS-1 can be a potential target for anti-arrhythmic therapy.

Voltage-Gated Ca^{2+} Channels

Neurotransmitter Release and Neurite Elongation

NCS-1 regulates voltage-gated Ca^{2+} channels. Published data, however, are somewhat inconsistent with reports demonstrating

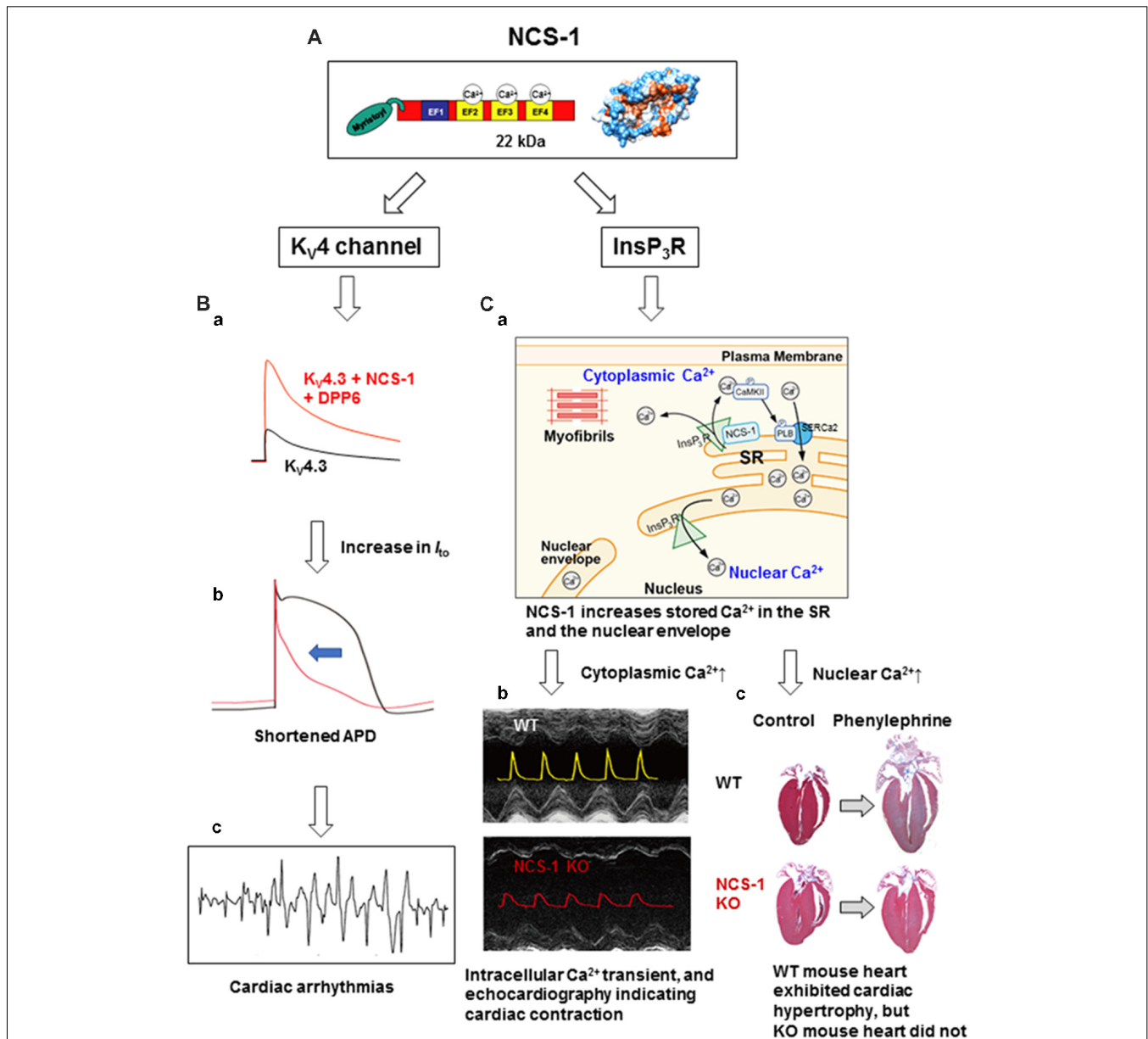


FIGURE 1 | The structure and cardiac functions of NCS-1. **(A)** The structure of NCS-1 (PDB: 1G8I). **(B)** Cartoons demonstrate that NCS-1 and DPP6, both are auxiliary subunits of K_V4 channels in Purkinje fiber (PF), slows inactivation kinetics of K_V4 current and increases the current amplitude, respectively, thus increase I_{to} -mediated K^+ efflux **(Ba)**. This would accelerate PF repolarization and shortening of APD **(Bb)**, and may lead to cardiac arrhythmias **(Bc)**. The same concepts of the traces in **(B)** were originally reported by Xiao et al. (2013). **(C)** NCS-1 also interacts with IP_3Rs on the SR and increases local Ca^{2+} . This activates CaMKII, followed by CaMKII-dependent phosphorylation of PLB that enhances the Ca^{2+} -pump activity of SERCA2, resulting in the increase in SR Ca^{2+} content **(Ca)**. This increases the global Ca^{2+} transient and contraction in the immature heart. NCS-1 deficiency results in a smaller Ca^{2+} -transient and contraction **(Cb)**; the composite figure of echocardiograms and Ca^{2+} transients are based on data from Nakamura et al., (2011). NCS-1 also increases nuclear Ca^{2+} levels because the SR and the nuclear envelope are interconnected **(Ca)**. NCS-1-mediated increase in nuclear Ca^{2+} signal can promote hormone-induced cardiac hypertrophy, whereas NCS-1 deficiency prevents progression of hypertrophy **(Cc)**; adapted from Nakamura et al., 2011). Phenylephrine is an agonist of α_1 -adrenergic receptor. For further details, please refer to the text. APD, action potential duration; CaMKII, calcium/calmodulin-dependent protein kinase II; DPP6, dipeptidyl peptidase-like protein 6; EF, EF-hand; InsP_3R , inositol 3,4,5-trisphosphate receptor; I_{to} , transient outward K^+ current; KO, knock-out; K_V , voltage-dependent potassium channel; NCS-1, neuronal Ca^{2+} sensor-1; PLB, phospholamban; SERCA2, sarcoplasmic/endoplasmic reticulum calcium ATPase 2; SR, sarcoplasmic reticulum; WT, wild type.

both positive and negative effects (**Table 1**). NCS-1 was described to inhibit P/Q-type Ca^{2+} channels and regulates autocrine pathways in adrenal chromaffin cells (Weiss et al., 2000; Weiss and Burgoyne, 2001) and N-type Ca^{2+} channels in PC12 cells, which reduces neurite elongation (Gambino et al., 2007). Other

studies, in contrast, have demonstrated positive regulation of N-type Ca^{2+} channels, causing glial cell line-derived neurotrophic factor (GDNF)-induced enhancement of neurotransmitter release in motoneurons (Wang et al., 2001). Activation of P/Q-type Ca^{2+} channels by NCS-1 causes

TABLE 1 | Neuronal Ca^{2+} sensor-1 (NCS-1) interacting proteins, known functions of NCS-1 mediated by those proteins and involvement in various diseases.

Interacting proteins	Molecular functions	Physiological roles and involvement in various diseases	References
Phosphatidylinositol 4-kinase III β (PI-4K- β)	Activation	Required for yeast survival Stimulation of exocytosis in neuroendocrine cells Facilitation of synaptic transmission in neurons Regulation of glucose-induced insulin secretion in pancreatic β cells Controlling exocytosis and inflammatory reactions in mast cells	Hendricks et al. (1999) Koizumi et al. (2002), Scalettar et al. (2002), Rajebhosale et al. (2003) and de Barry et al. (2006) Taverna et al. (2002) and Zheng et al. (2005) Gromada et al. (2005) Kapp-Barnea et al. (2003)
Voltage-gated K_v4 K^+ channels	Increase in K_v4 -mediated K^+ efflux	Increasing K_v4 current amplitude and slowing inactivation time course of A-type current in neurons Regulation of I_{to} in cardiomyocytes	Nakamura et al. (2001) Guo et al. (2002) and Nakamura and Coetzee (2008) Xiao et al. (2013)
Voltage-gated Ca^{2+} channels	Inhibition (P/Q-type) Inhibition (N-type) Activation (N-type) Activation (P/Q-type) Activation	Regulation of autocrine pathways in adrenal chromaffin cells Reduction of neurite elongation in PC12 cells Enhancement of GDNF-induced neurotransmitter release in motoneurons Activity-dependent synaptic facilitation in nerve terminals Enhancement of neurotransmission and nerve terminal growth in <i>Drosophila</i> Preservation of dopamine signaling	Weiss et al. (2000) and Weiss and Burgoyne (2001) Gambino et al. (2007) Wang et al. (2001) Tsujiimoto et al. (2002) Dason et al. (2009) Kabbani et al. (2002)
D2 dopamine receptor	Inhibition of D2 receptor phosphorylation, reduction of the agonist-mediated internalization of the receptor	Promotion of exploration, synaptic plasticity, and rapid acquisition of spatial memory in mice overexpressing NCS-1 in dentate gyrus Involvement in schizophrenia and bipolar disorder Augmentation of learning and memory in mice	Saab et al. (2009) Koh et al. (2003) and Bai et al. (2004) Saab et al. (2009), Mun et al. (2015) and Nakamura et al. (2017)
Inositol 1,4,5-trisphosphate receptors (InsP ₃ Rs)	Increase in InsP ₃ R channel activity	Enhancement of InsP ₃ R-mediated Ca^{2+} -signaling Regulation of neurite outgrowth in cultured neurons Involvement in long-term depression Involvement in bipolar disorder Involvement in Taxol-induced Ca^{2+} -oscillation and neuropathy Promotion of Ca^{2+} signaling and contraction in immature heart Regulation of cardiac hypertrophy Nuclear Ca^{2+} regulation in cardiomyocytes	Schlecker et al. (2006) Iketani et al. (2009) Jo et al. (2008) Schlecker et al. (2006) Boehmerle et al. (2006, 2007) and Blachford et al. (2009) Nakamura et al. (2011) Nakamura et al. (2011) Nakao et al. (2015)

activity-dependent synaptic facilitation in nerve terminals (Tsujiimoto et al., 2002). In *Drosophila*, NCS-1 enhances neurotransmission and nerve terminal growth, by functionally interacting with the $\alpha 1$ subunit of the voltage-gated Ca^{2+} channel (Dason et al., 2009). The possible reason for the apparent contradictory findings is that the effects may be cell type-specific and/or mediated by accessory proteins, such as a β -subunit (Rousset et al., 2003), or dependent on other interacting proteins, such as IL1RAPL1 that cooperatively regulates the N-type Ca^{2+} channel *via* NCS-1 (Gambino et al., 2007). In addition, regulation of the Ca^{2+} channel by Ca^{2+} influx through the channel should be considered. Well-characterized examples are Ca^{2+} -dependent inactivation (Standen and Stanfield, 1982) and facilitation (Dolphin, 1996) of Ca^{2+} channels regulated by other Ca^{2+} -binding proteins, such as calmodulin (Budde et al., 2002; Christel and Lee, 2012). Future research should aim to understand the regulatory mechanisms of Ca^{2+} channels that involve NCS-1.

D2 Dopamine Receptor

Role in Synaptic Plasticity and Psychiatric Illness

Dopamine plays an important role in the reward system of the brain. Disorders of the dopamine system result in several psychiatric and neurological conditions. Dopamine transmission is regulated by dopamine receptor-interacting proteins (DRIP), including NCS-1, calycon, and DARPP-32. NCS-1 directly interacts with the D2 dopamine receptor, inhibits D2 receptor phosphorylation, and reduces the agonist-mediated internalization of the receptor (Kabbani et al., 2002), indicating that NCS-1 preserves dopamine signaling (Table 1). Indeed, modest NCS-1 overexpression in the dentate gyrus in mice promotes exploration, synaptic plasticity, and rapid acquisition of spatial memory (Saab et al., 2009). NCS-1 is upregulated in the prefrontal cortex of patients with schizophrenia and bipolar disorder (Koh et al., 2003; Bai et al., 2004). Because the levels of other DRIPs were also changed in patients with schizophrenia (Bai et al., 2004; Souza et al., 2008), DRIP signaling is possibly involved in psychiatric disorders. Furthermore, recent findings indicate the N-terminal 60 residues of NCS-1 are responsible for binding to the D2 receptor (Woll et al., 2011). Such knowledge would provide an opportunity to screen for drugs that can specifically interrupt the NCS-1-D2 dopamine receptor interaction and thus prevent psychiatric disorders.

Role in Learning and Memory and Possible Mechanism

Several studies have demonstrated that NCS-1 modulates learning and memory. For example, deletion or reduction of NCS-1 resulted in dysfunction of learning and memory in *Caenorhabditis elegans* (Gomez et al., 2001), as well as in mice (Mun et al., 2015), whereas mice overexpressing NCS-1 rapidly acquire spatial memory (Saab et al., 2009). Thus, NCS-1 affects neurophysiology, possibly through various interacting proteins. A mechanism underlying NCS-1-mediated learning and memory was further investigated (Nakamura et al., 2017). *Ncs1*^{-/-} mice exhibited impaired spatial learning and memory function in the

Morris Water Maze test, with slight changes in their exercise activity or a structural change in the hippocampus. However, the levels of brain-derived neurotrophic factor (BDNF), a key regulator of memory function, and dopamine were decreased. Furthermore, phosphorylation of Ca^{2+} /calmodulin-dependent protein kinase II- α (CaMKII- α), which regulates long-term potentiation, and BDNF levels were decreased, suggesting that CaMKII- α signaling that increases BDNF production is at least partly involved in NCS-1-mediated learning and memory function.

Inositol 1,4,5-Trisphosphate Receptors

Role in Neuronal Pathogenesis

Ca^{2+} signaling *via* inositol 1,4,5-trisphosphate receptors (InsP₃Rs) regulates cellular function and is involved in pathogenesis (Table 1). NCS-1 physically interacts with InsP₃R1 and enhances InsP₃R-mediated Ca^{2+} signaling in rat brains. Indeed, physical/functional interaction of these proteins was directly demonstrated in an *in vitro* experiment showing that the addition of NCS-1 to InsP₃R1 in the lipid bilayer increased InsP₃R channel activity (Schlecker et al., 2006). This interaction was also detected at the growth cone region of neurites in cultured neurons, and indicated to be crucial for neurite outgrowth (Iketani et al., 2009). Metabotropic glutamate receptor-mediated cis also mediated by NCS-1/InsP₃R interaction (Jo et al., 2008). Pathologically, NCS-1/InsP₃R1 interaction is believed to be involved in bipolar disorder (Schlecker et al., 2006) because lithium, a medical drug for bipolar disorder, inhibited the NCS-1-induced enhancement of InsP₃R function. NCS-1/InsP₃R interaction is also considered to mediate neuropathy (Boehmerle et al., 2006, 2007; Blachford et al., 2009), as paclitaxel (taxol), a chemotherapeutic agent used for the treatment of solid cancers, modulates the expression/function of NCS-1, and hence InsP₃R1-mediated Ca^{2+} signaling.

Enhancement of Immature Heart Contraction and Hypertrophy

NCS-1/InsP₃R interaction is also detected in the heart and is crucial for contraction at the immature stage and cardiac hypertrophy in adult (Nakamura et al., 2011; Table 1 and Figure 1C). A high expression of NCS-1 was found in the immature heart (Nakamura et al., 2003, 2011), but its function at this stage was unknown. Using *Ncs1*^{-/-} mice, Nakamura et al demonstrated that NCS-1 contributes to an increase in contraction and Ca^{2+} signaling, specifically at the immature stage (Figure 1Cb). Intracellular Ca^{2+} levels and the sarcoplasmic reticulum (SR) Ca^{2+} content were significantly lower in *Ncs1*^{-/-} myocytes at the neonatal stage than in wild-type cells. Mechanistically, the interaction of NCS-1 with InsP₃R increases InsP₃R-dependent Ca^{2+} signaling, followed by the activation of CaMKII-dependent pathways, and promotes SR Ca^{2+} pump *via* the phosphorylation of phospholamban (PLB), which ultimately induce increase in the SR Ca^{2+} content and global Ca^{2+} transient, thus cardiomyocyte contraction (Figures 1Ca,b). The importance of crosstalk among NCS-1, InsP₃Rs, and CaMKII in the immature hearts was evident

by the high expression of all three proteins in immature hearts (Nakamura et al., 2011). In the neonatal mouse heart, the structure and function of SR are immature. Nonetheless, it is considered a primary source of Ca^{2+} necessary for muscle contractions, suggesting the existence of factors missing during development and promoting SR-dependent excitation-contraction (E-C) coupling in the postnatal stages. NCS-1 may act as one of these missing factors. Numbers of molecules which levels are high at the immature stage are often up-regulated in the disease conditions, such as cardiac hypertrophy. NCS-1 is also highly expressed during the early stages of hypertrophy in the adult heart and promotes the progression of hypertrophy, at least in part, through InsP_3R activation (Nakamura et al., 2011; **Figure 1C**). A possible molecular mechanism is suggested in the next section.

Regulation of Nuclear Ca^{2+} Signals

The aforementioned data indicate that NCS-1 can discretely regulate different types of Ca^{2+} signaling pathways in the heart (i.e., regulation of E-C coupling in immature heart and changes in gene expression in the adult heart). Recent evidence suggests that gene transcription is regulated by nuclear Ca^{2+} signals. However, the mechanisms underlying nuclear Ca^{2+} regulation and its relationship to cytoplasmic Ca^{2+} regulation have not been completely solved. Using a subcellular-specific, fluorescent protein-based Ca^{2+} indicator GECO (Zhao et al., 2011; Nakao et al., 2015) confirmed the following: (1) nuclear Ca^{2+} transients were elicited by both electrical and receptor stimulations (with insulin-like growth factor-1, IGF-1) in neonatal mouse ventricular myocytes; and (2) receptor stimulation-elicited nuclear Ca^{2+} transients were mainly mediated by InsP_3Rs . Furthermore, based on the evidence that IGF-1-elicited nuclear Ca^{2+} transient was significantly diminished in *Ncs1*^{-/-} cardiomyocytes, NCS-1 is involved in the receptor stimulation-induced nuclear Ca^{2+} regulation through interaction with InsP_3Rs (Nakao et al., 2015; **Table 1**). This may contribute to NCS-1-mediated hypertrophy, which was described above. A possible mechanism underlying a dual effect of NCS-1 on nuclear Ca^{2+} signals and E-C coupling is that NCS-1 increases the Ca^{2+} content of SR (Nakamura et al., 2011) that is interconnected to the nuclear envelope (Wu and Bers, 2006) and consequently may increase nuclear Ca^{2+} (Nakao et al., 2015; **Figure 1Ca**).

Other Functions of NCS-1 With Unknown Interacting Proteins

Enhancement of Neuronal Survival After Injury

Physical or chemical injury and genetic abnormalities can result in neuronal degeneration, which may underlie human neurodegenerative disorders, such as Alzheimer's disease and Parkinson's disease. Both intrinsic and extrinsic factors, including neurotrophic factors, can activate the anti-apoptotic process to rescue neuronal cell death. The signaling pathway leading to cell survival remains unresolved. In this regard, NCS-1 was found to be a novel Ca^{2+} -dependent survival-promoting factor upregulated in injured neurons (Nakamura et al., 2006), based on the following observations. (1) NCS-1 expression

increases in injured neurons; (2) NCS-1 overexpression diminished various stress-induced neuronal cell death in culture; and (3) the dominant negative EF-hand NCS-1 mutant (E120Q) accelerated cell death. Mechanistically, the expression level of NCS-1 in neuron is increased by GDNF, a neurotrophic factor upregulated by neuronal injury, and NCS-1 mediates GDNF survival signaling *via* the activation of the Akt pathway.

Role in Stress Tolerance in Cardiomyocytes

Not only in neurons, NCS-1 also plays a key role in protecting cardiomyocytes against stress through the activation of mitochondrial detoxification pathways (Nakamura et al., 2016). Excessive stress induces cytosolic Ca^{2+} overload and cell death. In contrast, mild forms of stress lead to physiologically relevant changes in Ca^{2+} , which activate Ca^{2+} -dependent survival pathways by binding to Ca^{2+} -sensor proteins. As one such protein, NCS-1 was found to play important roles in Ca^{2+} -dependent survival signaling. *Ncs1*^{-/-} myocytes were more susceptible to oxidative and metabolic stress, and cellular ATP levels, mitochondrial respiration and biosynthesis were significantly reduced in these cells. In wild-type myocytes, mild oxidative stress increased the mitochondrial proton leak, which exerted a protective effect by inhibiting the production of reactive oxygen species. However, this response was diminished in *Ncs1*^{-/-} cardiomyocytes, thus resulting in cell death. Similar susceptibility was also observed in *Ncs1*^{-/-} hearts subjected to ischemia-reperfusion injury. In these hearts, molecules regulating Ca^{2+} -dependent survival pathways, such as Akt and PGC-1 α , which promote mitochondrial biogenesis and function, were significantly downregulated compared to wild-type hearts. These data demonstrate a novel role of NCS-1 that contributes to stress tolerance in cardiomyocytes, partly by the activation of Ca^{2+} -dependent survival pathways. NCS-1 may also participate in cardioprotection by mediating receptor-signaling pathways. For example, NCS-1 associates with, and modulates, adenosine receptor activity (Navarro et al., 2012). Given the central role of adenosine in mediating the protective effects of ischemic preconditioning (Cohen and Downey, 2008), it is entirely possible that the cardioprotective effects of NCS-1 is partially mediated by this pathway.

CONCLUSION

Recently, studies have elucidated new roles of NCS-1 in physiology and pathophysiology. In this review, we have mainly focused on NCS-1 in the neuronal system and heart. Our particular interest is the emerging theme that NCS-1 directly regulates the function of several ion channels that permeate Ca^{2+} (e.g., several types of voltage-gated Ca^{2+} channels, ionotropic dopamine receptors, and InsP_3Rs), suggesting a general role of Ca^{2+} influx *via* the channel that binds to NCS-1 and consequently regulates channel functions and/or downstream Ca^{2+} -dependent signaling, which affect various neuronal and cardiac functions. Furthermore, many established roles of NCS-1 are related to protective responses of cells against exogenous stress that leads to mild increases in cytosolic Ca^{2+} . This suggest

that intracellular Ca^{2+} as a determinant of cell survival and cell death, and Ca^{2+} -sensor proteins, such as NCS-1, may serve as a switch to proceed the signal. We believe that this review provides intriguing observations and compels researchers to conduct detailed investigations and extend their studies on NCS-1 and its important regulatory proteins.

AUTHOR CONTRIBUTIONS

TN conducted most part of research, wrote, organized, and finalized the article. SN did some experiments and wrote some part of the article. SW contributed to the discussion on all part

REFERENCES

- An, W. F., Bowlby, M. R., Betty, M., Cao, J., Ling, H. P., Mendoza, G., et al. (2000). Modulation of A-type potassium channels by a family of calcium sensors. *Nature* 403, 553–556. doi: 10.1038/35000592
- Bai, J., He, F., Novikova, S. I., Undie, A. S., Dracheva, S., Haroutunian, V., et al. (2004). Abnormalities in the dopamine system in schizophrenia may lie in altered levels of dopamine receptor-interacting proteins. *Biol. Psychiatry* 56, 427–440. doi: 10.1016/j.biopsych.2004.06.022
- Bartlett, S. E., Reynolds, A. J., Weible, M., Jeromin, A., Roder, J., and Hendry, I. A. (2000). PtdIns 4-kinase β and neuronal calcium sensor-1 co-localize but may not directly associate in mammalian neurons. *J. Neurosci. Res.* 62, 216–224. doi: 10.1002/1097-4547(20001015)62:2<216::AID-JNR6>3.0.CO;2-A
- Blachford, C., Celić, A., Petri, E. T., and Ehrlich, B. E. (2009). Discrete proteolysis of neuronal calcium sensor-1 (NCS-1) by mu-calpain disrupts calcium binding. *Cell Calcium* 46, 257–262. doi: 10.1016/j.ceca.2009.08.002
- Boehmerle, W., Splittgerber, U., Lazarus, M. B., Mckenzie, K. M., Johnston, D. G., Austin, D. J., et al. (2006). Paclitaxel induces calcium oscillations via an inositol 1,4,5-trisphosphate receptor and neuronal calcium sensor 1-dependent mechanism. *Proc. Natl. Acad. Sci. U S A* 103, 18356–18361. doi: 10.1073/pnas.0607240103
- Boehmerle, W., Zhang, K., Sivula, M., Heidrich, F. M., Lee, Y., Jordt, S. E., et al. (2007). Chronic exposure to paclitaxel diminishes phosphoinositide signaling by calpain-mediated neuronal calcium sensor-1 degradation. *Proc. Natl. Acad. Sci. U S A* 104, 11103–11108. doi: 10.1073/pnas.0701546104
- Budde, T., Meuth, S., and Pape, H. C. (2002). Calcium-dependent inactivation of neuronal calcium channels. *Nat. Rev. Neurosci.* 3, 873–883. doi: 10.1038/nrn959
- Christel, C., and Lee, A. (2012). Ca^{2+} -dependent modulation of voltage-gated Ca^{2+} channels. *Biochim. Biophys. Acta* 1820, 1243–1252. doi: 10.1016/j.bbagen.2011.12.012
- Cohen, M. V., and Downey, J. M. (2008). Adenosine: trigger and mediator of cardioprotection. *Basic Res. Cardiol.* 103, 203–215. doi: 10.1007/s00395-007-0687-7
- Dason, J. S., Romero-Pozuelo, J., Marin, L., Iyengar, B. G., Klose, M. K., Ferrus, A., et al. (2009). Frequentin/NCS-1 and the Ca^{2+} -channel α_1 -subunit co-regulate synaptic transmission and nerve-terminal growth. *J. Cell Sci.* 122, 4109–4121. doi: 10.1242/jcs.055095
- de Barry, J., Janoshazi, A., Dupont, J. L., Procksch, O., Chasserot-Golaz, S., Jeromin, A., et al. (2006). Functional implication of neuronal calcium sensor-1 and phosphoinositol 4-kinase- β interaction in regulated exocytosis of PC12 cells. *J. Biol. Chem.* 281, 18098–18111. doi: 10.1074/jbc.m509842200
- Dolphin, A. C. (1996). Facilitation of Ca^{2+} current in excitable cells. *Trends Neurosci.* 19, 35–43. doi: 10.1016/0166-2236(96)81865-0
- Fiset, C., Clark, R. B., Shimoni, Y., and Giles, W. R. (1997). Shal-type channels contribute to the Ca^{2+} -independent transient outward K^+ current in rat ventricle. *J. Physiol.* 500, 51–64. doi: 10.1113/jphysiol.1997.sp021998
- Gambino, F., Pavlowsky, A., Begle, A., Dupont, J. L., Bahi, N., Courjaret, R., et al. (2007). IL1-receptor accessory protein-like 1 (IL1RAPL1), a protein involved in cognitive functions, regulates N-type Ca^{2+} -channel and neurite elongation. *Proc. Natl. Acad. Sci. U S A* 104, 9063–9068. doi: 10.1073/pnas.0701133104
- Gomez, M., De Castro, E., Guarin, E., Sasakura, H., Kuhara, A., Mori, I., et al. (2001). Ca^{2+} signaling via the neuronal calcium sensor-1 regulates associative learning and memory in *C. elegans*. *Neuron* 30, 241–248. doi: 10.1016/s0896-6273(01)00276-8
- Gromada, J., Bark, C., Smidt, K., Efanov, A. M., Janson, J., Mandic, S. A., et al. (2005). Neuronal calcium sensor-1 potentiates glucose-dependent exocytosis in pancreatic β cells through activation of phosphatidylinositol 4-kinase β . *Proc. Natl. Acad. Sci. U S A* 102, 10303–10308. doi: 10.1073/pnas.0504487102
- Guo, W., Malin, S. A., Johns, D. C., Jeromin, A., and Nerbonne, J. M. (2002). Modulation of Kv4-encoded K^+ currents in the mammalian myocardium by neuronal calcium sensor-1. *J. Biol. Chem.* 277, 26436–26443. doi: 10.1074/jbc.m201431200
- Hendricks, K. B., Wang, B. Q., Schnieders, E. A., and Thorner, J. (1999). Yeast homologue of neuronal frequenin is a regulator of phosphatidylinositol-4-OH kinase. *Nat. Cell Biol.* 1, 234–241. doi: 10.1038/12058
- Iketani, M., Imaizumi, C., Nakamura, F., Jeromin, A., Mikoshiba, K., Goshima, Y., et al. (2009). Regulation of neurite outgrowth mediated by neuronal calcium sensor-1 and inositol 1,4,5-trisphosphate receptor in nerve growth cones. *Neuroscience* 161, 743–752. doi: 10.1016/j.neuroscience.2009.04.019
- Ikura, M. (1996). Calcium binding and conformational response in EF-hand proteins. *Trends Biochem. Sci.* 21, 14–17. doi: 10.1016/s0968-0004(06)80021-6
- Jerng, H. H., Pfaffinger, P. J., and Covarrubias, M. (2004). Molecular physiology and modulation of somatodendritic A-type potassium channels. *Mol. Cell. Neurosci.* 27, 343–369. doi: 10.1016/j.mcn.2004.06.011
- Jo, J., Heon, S., Kim, M. J., Son, G. H., Park, Y., Henley, J. M., et al. (2008). Metabotropic glutamate receptor-mediated LTD involves two interacting Ca^{2+} sensors, NCS-1 and PICK1. *Neuron* 60, 1095–1111. doi: 10.1016/j.neuron.2008.10.050
- Kabbani, N., Negyessy, L., Lin, R., Goldman-Rakic, P., and Levenson, R. (2002). Interaction with neuronal calcium sensor NCS-1 mediates desensitization of the D2 dopamine receptor. *J. Neurosci.* 22, 8476–8486. doi: 10.1523/jneurosci.22-19-08476.2002
- Kapp-Barnea, Y., Melnikov, S., Shefler, I., Jeromin, A., and Sagi-Eisenberg, R. (2003). Neuronal calcium sensor-1 and phosphatidylinositol 4-kinase β regulate IgE receptor-triggered exocytosis in cultured mast cells. *J. Immunol.* 171, 5320–5327. doi: 10.4049/jimmunol.171.10.5320
- Koh, P. O., Undie, A. S., Kabbani, N., Levenson, R., Goldman-Rakic, P. S., and Lidow, M. S. (2003). Up-regulation of neuronal calcium sensor-1 (NCS-1) in the prefrontal cortex of schizophrenic and bipolar patients. *Proc. Natl. Acad. Sci. U S A* 100, 313–317. doi: 10.1073/pnas.232693499
- Koizumi, S., Rosa, P., Willars, G. B., Challiss, R. A., Taverna, E., Francolini, M., et al. (2002). Mechanisms underlying the neuronal calcium sensor-1-evoked enhancement of exocytosis in PC12 cells. *J. Biol. Chem.* 277, 30315–30324. doi: 10.1074/jbc.m201132200
- Mikhaylova, M., Reddy, P. P., Munsch, T., Landgraf, P., Suman, S. K., Smalla, K. H., et al. (2009). Calneurons provide a calcium threshold for trans-Golgi network to plasma membrane trafficking. *Proc. Natl. Acad. Sci. U S A* 106, 9093–9098. doi: 10.1073/pnas.0903001106
- Mun, H. S., Saab, B. J., Ng, E., Mcgirr, A., Lipina, T. V., Gondo, Y., et al. (2015). Self-directed exploration provides a Ncs1-dependent learning bonus. *Sci. Rep.* 5:17697. doi: 10.1038/srep17697
- Nakamura, T. Y., and Coetzee, W. A. (2008). Functional and pharmacological characterization of a Shal-related K^+ channel subunit in Zebrafish. *BMC Physiol.* 8:2. doi: 10.1186/1472-6793-8-2

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- Nakamura, T. Y., Coetzee, W. A., Vega-Saenz De Miera, E., Artman, M., and Rudy, B. (1997). Modulation of K_V4 channels, key components of rat ventricular transient outward K^+ current, by PKC. *Am. J. Physiol.* 273, H1775–H1786. doi: 10.1152/ajpheart.1997.273.4.h1775
- Nakamura, T. Y., Jeromin, A., Mikoshiba, K., and Wakabayashi, S. (2011). Neuronal calcium sensor-1 promotes immature heart function and hypertrophy by enhancing Ca^{2+} signals. *Circ. Res.* 109, 512–523. doi: 10.1161/CIRCRESAHA.111.248864
- Nakamura, T. Y., Jeromin, A., Smith, G., Kurushima, H., Koga, H., Nakabeppu, Y., et al. (2006). Novel role of neuronal Ca^{2+} sensor-1 as a survival factor up-regulated in injured neurons. *J. Cell Biol.* 172, 1081–1091. doi: 10.1083/jcb.200508156
- Nakamura, T. Y., Nakao, S., and Wakabayashi, S. (2016). Neuronal Ca^{2+} sensor-1 contributes to stress tolerance in cardiomyocytes via activation of mitochondrial detoxification pathways. *J. Mol. Cell. Cardiol.* 99, 23–34. doi: 10.1016/j.yjmcc.2016.08.013
- Nakamura, T. Y., Nakao, S., Nakajo, Y., Takahashi, J. C., Wakabayashi, S., and Yanamoto, H. (2017). Possible signaling pathways mediating neuronal calcium sensor-1-dependent spatial learning and memory in mice. *PLoS One* 12:e0170829. doi: 10.1371/journal.pone.0170829
- Nakamura, T. Y., Pountney, D. J., Ozaita, A., Nandi, S., Ueda, S., Rudy, B., et al. (2001). A role for frequenin, a Ca^{2+} -binding protein, as a regulator of K_V4K^+ -currents. *Proc. Natl. Acad. Sci. U S A* 98, 12808–12813. doi: 10.1073/pnas.221168498
- Nakamura, T. Y., Sturm, E., Pountney, D. J., Orenzoff, B., Artman, M., and Coetzee, W. A. (2003). Developmental expression of NCS-1 (frequenin), a regulator of K_V4K^+ channels, in mouse heart. *Pediatr. Res.* 53, 554–557. doi: 10.1203/01.PDR.0000057203.72435.C9
- Nakao, S., Wakabayashi, S., and Nakamura, T. Y. (2015). Stimulus-dependent regulation of nuclear Ca^{2+} signaling in cardiomyocytes: a role of neuronal calcium sensor-1. *PLoS One* 10:e0125050. doi: 10.1371/journal.pone.0125050
- Navarro, G., Hradsky, J., Lluís, C., Casado, V., McCormick, P. J., Kreutz, M. R., et al. (2012). NCS-1 associates with adenosine A_{2A} receptors and modulates receptor function. *Front. Mol. Neurosci.* 5:53. doi: 10.3389/fnmol.2012.00053
- Pongs, O., Lindemeier, J., Zhu, X. R., Theil, T., Engelkamp, D., Krah-Jentgens, I., et al. (1993). Frequenin—a novel calcium-binding protein that modulates synaptic efficacy in the *Drosophila* nervous system. *Neuron* 11, 15–28. doi: 10.1016/0896-6273(93)90267-u
- Poulain, C., Ferrús, A., and Mallart, A. (1994). Modulation of type A K^+ current in *Drosophila* larval muscle by internal Ca^{2+} ; effects of the overexpression of frequenin. *Pflugers Arch.* 427, 71–79. doi: 10.1007/bf00585944
- Rajebhosale, M., Greenwood, S., Vidugiriene, J., Jeromin, A., and Hilfiker, S. (2003). Phosphatidylinositol 4-OH kinase is a downstream target of neuronal calcium sensor-1 in enhancing exocytosis in neuroendocrine cells. *J. Biol. Chem.* 278, 6075–6084. doi: 10.1074/jbc.M204702200
- Roussel, M., Cens, T., Gavarini, S., Jeromin, A., and Charnet, P. (2003). Down-regulation of voltage-gated Ca^{2+} channels by neuronal calcium sensor-1 is β subunit-specific. *J. Biol. Chem.* 278, 7019–7026. doi: 10.1074/jbc.M209537200
- Saab, B. J., Georgiou, J., Nath, A., Lee, F. J., Wang, M., Michalon, A., et al. (2009). NCS-1 in the dentate gyrus promotes exploration, synaptic plasticity, and rapid acquisition of spatial memory. *Neuron* 63, 643–656. doi: 10.1016/j.neuron.2009.08.014
- Scalettar, B. A., Rosa, P., Taverna, E., Francolini, M., Tsuboi, T., Terakawa, S., et al. (2002). Neuronal calcium sensor-1 binds to regulated secretory organelles and functions in basal and stimulated exocytosis in PC12 cells. *J. Cell Sci.* 115, 2399–2412.
- Schlecker, C., Boehmerle, W., Jeromin, A., DeGray, B., Varshney, A., Sharma, Y., et al. (2006). Neuronal calcium sensor-1 enhancement of $InsP_3$ receptor activity is inhibited by therapeutic levels of lithium. *J. Clin. Invest.* 116, 1668–1674. doi: 10.1172/jci22466
- Serôdio, P., Kentros, C., and Rudy, B. (1994). Identification of molecular components of A-type channels activating at subthreshold potentials. *J. Neurophysiol.* 72, 1516–1529. doi: 10.1152/jn.1994.72.4.1516
- Souza, B. R., Motta, B. S., Rosa, D. V., Torres, K. C., Castro, A. A., Comim, C. M., et al. (2008). DARPP-32 and NCS-1 expression is not altered in brains of rats treated with typical or atypical antipsychotics. *Neurochem. Res.* 33, 533–538. doi: 10.1007/s11064-007-9470-2
- Standen, N. B., and Stanfield, P. R. (1982). A binding-site model for calcium-channel inactivation that depends on calcium entry. *Proc. R. Soc. Lond. B Biol. Sci.* 217, 101–110. doi: 10.1098/rspb.1982.0097
- Strahl, T., Huttner, I. G., Lusin, J. D., Osawa, M., King, D., Thorner, J., et al. (2007). Structural insights into activation of phosphatidylinositol 4-kinase (Pik1) by yeast frequenin (Frq1). *J. Biol. Chem.* 282, 30949–30959. doi: 10.1074/jbc.M705499200
- Taverna, E., Francolini, M., Jeromin, A., Hilfiker, S., Roder, J., and Rosa, P. (2002). Neuronal calcium sensor 1 and phosphatidylinositol 4-OH kinase β interact in neuronal cells and are translocated to membranes during nucleotide-evoked exocytosis. *J. Cell Sci.* 115, 3909–3922. doi: 10.1242/jcs.00072
- Tsujimoto, T., Jeromin, A., Saitoh, N., Roder, J. C., and Takahashi, T. (2002). Neuronal calcium sensor 1 and activity-dependent facilitation of P/Q-type calcium currents at presynaptic nerve terminals. *Science* 295, 2276–2279. doi: 10.1126/science.1068278
- Wang, C. Y., Yang, F., He, X., Chow, A., Du, J., Russell, J. T., et al. (2001). Ca^{2+} binding protein frequenin mediates GDNF-induced potentiation of Ca^{2+} channels and transmitter release. *Neuron* 32, 99–112. doi: 10.1016/s0896-6273(01)00434-2
- Weiss, J. L., Archer, D. A., and Burgoyne, R. D. (2000). Neuronal Ca^{2+} sensor-1/frequenin functions in an autocrine pathway regulating Ca^{2+} channels in bovine adrenal chromaffin cells. *J. Biol. Chem.* 275, 40082–40087. doi: 10.1074/jbc.M008603200
- Weiss, J. L., and Burgoyne, R. D. (2001). Voltage-independent inhibition of P/Q-type Ca^{2+} channels in adrenal chromaffin cells via a neuronal Ca^{2+} sensor-1-dependent pathway involves Src family tyrosine kinase. *J. Biol. Chem.* 276, 44804–44811. doi: 10.1074/jbc.m103262200
- Woll, M. P., De Cotiis, D. A., Bewley, M. C., Tacelosky, D. M., Levenson, R., and Flanagan, J. M. (2011). Interaction between the D2 dopamine receptor and neuronal calcium sensor-1 analyzed by fluorescence anisotropy. *Biochemistry* 50, 8780–8791. doi: 10.1021/bi200637e
- Wu, X., and Bers, D. M. (2006). Sarcoplasmic reticulum and nuclear envelope are one highly interconnected Ca^{2+} store throughout cardiac myocyte. *Circ. Res.* 99, 283–291. doi: 10.1161/01.res.0000233386.02708.72
- Xiao, L., Koopmann, T. T., Ördög, B., Postema, P. G., Verkerk, A. O., Iyer, V., et al. (2013). Unique cardiac purkinje fiber transient outward current β -subunit composition a potential molecular link to idiopathic ventricular fibrillation. *Circ. Res.* 112, 1310–1322. doi: 10.1161/circresaha.112.300227
- Zhang, K., Heidrich, F. M., Degray, B., Boehmerle, W., and Ehrlich, B. E. (2010). Paclitaxel accelerates spontaneous calcium oscillations in cardiomyocytes by interacting with NCS-1 and the $InsP_3R$. *J. Mol. Cell. Cardiol.* 49, 829–835. doi: 10.1016/j.yjmcc.2010.08.018
- Zhao, Y. X., Araki, S., Wu, J., Teramoto, T., Chang, Y. F., Nakano, M., et al. (2011). An expanded palette of genetically encoded Ca^{2+} indicators. *Science* 333, 1888–1891. doi: 10.1126/science.1208592
- Zheng, Q., Bobich, J. A., Vidugiriene, J., McFadden, S. C., Thomas, F., Roder, J., et al. (2005). Neuronal calcium sensor-1 facilitates neuronal exocytosis through phosphatidylinositol 4-kinase. *J. Neurochem.* 92, 442–451. doi: 10.1111/j.1471-4159.2004.02897.x

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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