



# Molecular Mechanisms of Antipsychotic Drug-Induced Diabetes

Jiezhong Chen<sup>1,2</sup>, Xu-Feng Huang<sup>2,3</sup>, Renfu Shao<sup>4</sup>, Chen Chen<sup>1</sup> and Chao Deng<sup>2,3\*</sup>

<sup>1</sup> School of Biomedical Sciences, The University of Queensland, St Lucia, QLD, Australia, <sup>2</sup> School of Medicine, University of Wollongong, Wollongong, NSW, Australia, <sup>3</sup> Illawarra Health and Medical Research Institute, Wollongong, NSW, Australia, <sup>4</sup> Faculty of Science, Health, Education and Engineering, GeneCology Research Centre, University of the Sunshine Coast, Maroochydore, QLD, Australia

## OPEN ACCESS

### Edited by:

Miguel López,  
Universidad de Santiago de  
Compostela, Spain

### Reviewed by:

Johan Anders Fernø,  
Haukeland University Hospital,  
Norway  
Ismael González-García,  
Universidad de Santiago de  
Compostela, Spain

### \*Correspondence:

Chao Deng  
chao@uow.edu.au

### Specialty section:

This article was submitted to  
Neuroendocrine Science,  
a section of the journal  
Frontiers in Neuroscience

**Received:** 28 August 2017

**Accepted:** 06 November 2017

**Published:** 21 November 2017

### Citation:

Chen J, Huang X-F, Shao R, Chen C  
and Deng C (2017) Molecular  
Mechanisms of Antipsychotic  
Drug-Induced Diabetes.  
*Front. Neurosci.* 11:643.  
doi: 10.3389/fnins.2017.00643

Antipsychotic drugs (APDs) are widely prescribed to control various mental disorders. As mental disorders are chronic diseases, these drugs are often used over a life-time. However, APDs can cause serious glucometabolic side-effects including type 2 diabetes and hyperglycaemic emergency, leading to medication non-compliance. At present, there is no effective approach to overcome these side-effects. Understanding the mechanisms for APD-induced diabetes should be helpful in prevention and treatment of these side-effects of APDs and thus improve the clinical outcomes of APDs. In this review, the potential mechanisms for APD-induced diabetes are summarized so that novel approaches can be considered to relieve APD-induced diabetes. APD-induced diabetes could be mediated by multiple mechanisms: (1) APDs can inhibit the insulin signaling pathway in the target cells such as muscle cells, hepatocytes and adipocytes to cause insulin resistance; (2) APD-induced obesity can result in high levels of free fatty acids (FFA) and inflammation, which can also cause insulin resistance. (3) APDs can cause direct damage to  $\beta$ -cells, leading to dysfunction and apoptosis of  $\beta$ -cells. A recent theory considers that both  $\beta$ -cell damage and insulin resistance are necessary factors for the development of diabetes. In high-fat diet-induced diabetes, the compensatory ability of  $\beta$ -cells is gradually damaged, while APDs cause direct  $\beta$ -cell damage, accounting for the severe form of APD-induced diabetes. Based on these mechanisms, effective prevention of APD-induced diabetes may need an integrated approach to combat various effects of APDs on multiple pathways.

**Keywords:** antipsychotics, diabetes mellitus, metabolic disorders, insulin resistance, obesity, pancreatic beta cell, apoptosis

## INTRODUCTION

Antipsychotic drugs (APDs) are widely prescribed to control schizophrenia and bipolar disorders, as well as other mental disorders including dementia, major depression, and even drug addiction (Fraguas et al., 2011; Leucht et al., 2013; Zhang J.-P. et al., 2013; Samara et al., 2016). Typical APDs (also called first generation APDs) such as chlorpromazine, perphenazine and haloperidol were introduced to clinics more than 60 years ago. Therapeutic effects of typical APDs are mediated largely through potent blockage of dopamine D2 receptors (Ginovart and Kapur, 2012), which also cause extra-pyramidal symptoms (EPS) side-effects (Stahl, 2003). Since the 1990s, a number of

atypical APDs (also called 2nd generation APDs) including olanzapine, clozapine and risperidone have been approved by the FDA, and are now widely used as first line APDs due to their improved tolerability and reduced EPS compared with typical APDs (Leucht et al., 2009; Zhang J.-P. et al., 2013). In addition, clozapine has better outcomes in treatment-resistant schizophrenia (Lewis et al., 2006; Samara et al., 2016). Besides blockage of D2 receptors, atypical APDs target multiple neuroreceptors such as serotonergic 5-HT<sub>2A</sub>/5-HT<sub>2C</sub>, histaminergic H<sub>1</sub> and muscarinic M<sub>3</sub> receptors (Correll, 2010). Although typical APDs have been reported causing a certain degree of metabolic disorders, atypical APDs, particularly clozapine and olanzapine, can cause much worse metabolic side-effects including body weight gain, obesity, hyperlipidaemia, insulin resistance, hyperglycaemia and diabetes (Foley and Morley, 2011; De Hert et al., 2012; Deng, 2013; Lipscombe et al., 2014; Stubbs et al., 2015). Since psychiatric patients often face chronic and even life-time APD treatment, these side-effects are major considerations in APD medication (De Hert et al., 2012; Deng, 2013). Schizophrenia patients with APD treatment have 2.5 times higher risk of developing type 2 diabetes according to a recent meta-analysis which examined 25 studies, including 145,718 individuals with schizophrenia (22.5–54.4 years) and 4,343,407 controls (Stubbs et al., 2015). It is noteworthy that over the last decade APD prescriptions in children and adolescents have sharply increased (Memarzia et al., 2014; Steinhausen, 2015). Recent studies have shown that APDs cause not only greater weight gain in children/adolescents than in adults but also significant risk of type 2 diabetes, which has been largely underestimated (Maayan and Correll, 2011; Samaras et al., 2014; Pramyothin and Khaothiar, 2015; Sohn et al., 2015). These severe side-effects have a devastating impact on life quality, and is a key risk for severe health complications, including cardiovascular disease, stroke, and premature death (Foley and Morley, 2011; Lin et al., 2014; Wu et al., 2015). Understanding how diabetes develops in patients treated with APDs and preventing APD-induced diabetes will improve medication compliance.

## ANTIPSYCHOTIC-INDUCED DIABETES

Diabetes is characterized by hyperglycaemia (fasting plasma glucose  $\geq 126$  mg/dL [7 mmol/L]) due to insufficient insulin production or insulin resistance (Kahn et al., 2014). Insulin is well known to promote glucose metabolism to produce energy. Diabetes is commonly classified into two types; type 1 is due to auto-immune damage to pancreatic  $\beta$ -cells, leading to a complete deficiency of insulin; type 2 is caused by insulin resistance, i.e., the cells do not respond to insulin stimulation, as well as  $\beta$ -cell dysfunction (Kahn et al., 2014). There is a chronic development from insulin resistance to diabetes. In the case of insulin resistance, insulin secretion from  $\beta$ -cells can increase 4-fold while  $\beta$ -cell mass can increase 2-fold (Kahn et al., 2006). Therefore, blood glucose levels can still be maintained at normal levels, however, if  $\beta$ -cells fail to increase insulin secretion, diabetes will occur.

APDs have been shown to cause both diabetes and hyperglycaemic emergencies. Chronic administration of APDs is known to cause diabetes, which has been demonstrated in both epidemiological investigations in patients and in animal studies (Boyda et al., 2010a; Sohn et al., 2015; Stubbs et al., 2015). A recent study of 307 patients with psychotic illnesses showed that olanzapine caused type 2 diabetes (17%), obesity (48%), dyslipidaemia (35%) and hypertension (32%) with mean treatment duration of 7.6 years, while other APDs also induced similar side-effects (Reed et al., 2014). Another study has shown that APDs caused 37% prediabetes and 10% diabetes (Manu et al., 2012). Furthermore, the European First-Episode Schizophrenia Trial (EUFEST) reported a 20–30% incidence rate of hyperglycemia after 1 year of treatment with olanzapine, quetiapine and ziprasidone, but no significant differences between these APDs (Fleischhacker et al., 2013). Comparison of 28,858 APD users with 14,429 controls showed that the risk of diabetes increased 3-fold in children and adolescents treated with APDs (Bobo et al., 2013). This APD-induced diabetes has been confirmed in animal models; olanzapine and clozapine have been shown to decrease the plasma level of insulin and to cause hyperglycaemia and insulin resistance in rats (Chintoh et al., 2009; Boyda et al., 2010a; Liu et al., 2017).

The major complication of diabetes is heart disease. APDs significantly increased acute myocardial infarction with adjusted odds ratio of the risk 2.52 (95% CI = 2.37–2.68) (Lin et al., 2014). Citrome et al. examined diabetes and cardiovascular disease incidence in a large number of patients who used APDs including aripiprazole, olanzapine, risperidone, quetiapine, and ziprasidone (Citrome et al., 2013). It was found that these drugs caused diabetes at similar rates but olanzapine, risperidone and quetiapine caused more cases of cardiovascular disease than aripiprazole and ziprasidone (Citrome et al., 2013).

Recent attention has also been focused on APD-induced hyperglycaemic emergencies. Patients treated with atypical APDs have ~10 times higher risk in developing diabetic ketoacidosis (Polcwiartek et al., 2016). Lipscombe and colleagues conducted a multicenter retrospective cohort study including 725,489 patients to investigate hyperglycaemic emergencies (hyperglycaemia, diabetic ketoacidosis, hyperosmolar hyperglycaemic state) experienced by new users of risperidone, olanzapine, and other typical and atypical APDs (Lipscombe et al., 2014). The results showed that hyperglycaemic emergencies were 1 per 1,000 persons in patients aged 18–65 and 2 per 1,000 persons in those older than 65. The events were much more frequent in patients with pre-existing diabetes (6–12 per 1,000 persons) (Lipscombe et al., 2014). It was similar in the risk of hyperglycaemic emergencies with initiation of olanzapine vs. risperidone.

In brief, both clinical and animal studies have shown that APDs can cause serious glucometabolic side-effects including hyperglycaemic emergency, insulin resistance, hyperglycaemia and type 2 diabetes, which is a major risk for cardiovascular disease and premature death. It is worth to note that the risk of type 2 diabetes in children and youths treated with APDs has been underestimated (Samaras et al., 2014). Understanding the underlying mechanisms will be important for preventing

and treating these side-effects and thus improving the clinical outcomes of APDs.

## MECHANISMS FOR ANTIPSYCHOTIC-INDUCED DIABETES

Over the past 10 years, a number of studies have aimed to explore the potential mechanisms underlying APD-induced diabetes. Based on recent progress, we summarized the findings into three molecular mechanisms for explaining APD-induced diabetes: (1) insulin resistance due to the direct effect of APDs, (2) APD-caused insulin resistance through obesity, and (3) APD-induced  $\beta$ -cell dysfunction and apoptosis.

### Mechanism 1: Insulin Resistance Due to Direct Effect of Antipsychotics

Insulin, secreted by pancreatic  $\beta$ -cells, is the key hormone in promotion of glucose metabolism (Kahn et al., 2014). It increases the uptake of glucose by cells and thus maintains the homeostasis of blood glucose levels. Insulin resistance refers to the situation where the target cells lose response to insulin stimulation and thus reduce glucose uptake (Kahn et al., 2014). Increased blood glucose level are mainly caused by insulin resistance in the skeletal muscles, and also in the hepatic, renal and adipose tissue (Kahn et al., 2014). Of these, the main site for glucose utilization is muscle tissue, and represents  $\sim 80\%$  of glucose consumption (Fujii et al., 2005).

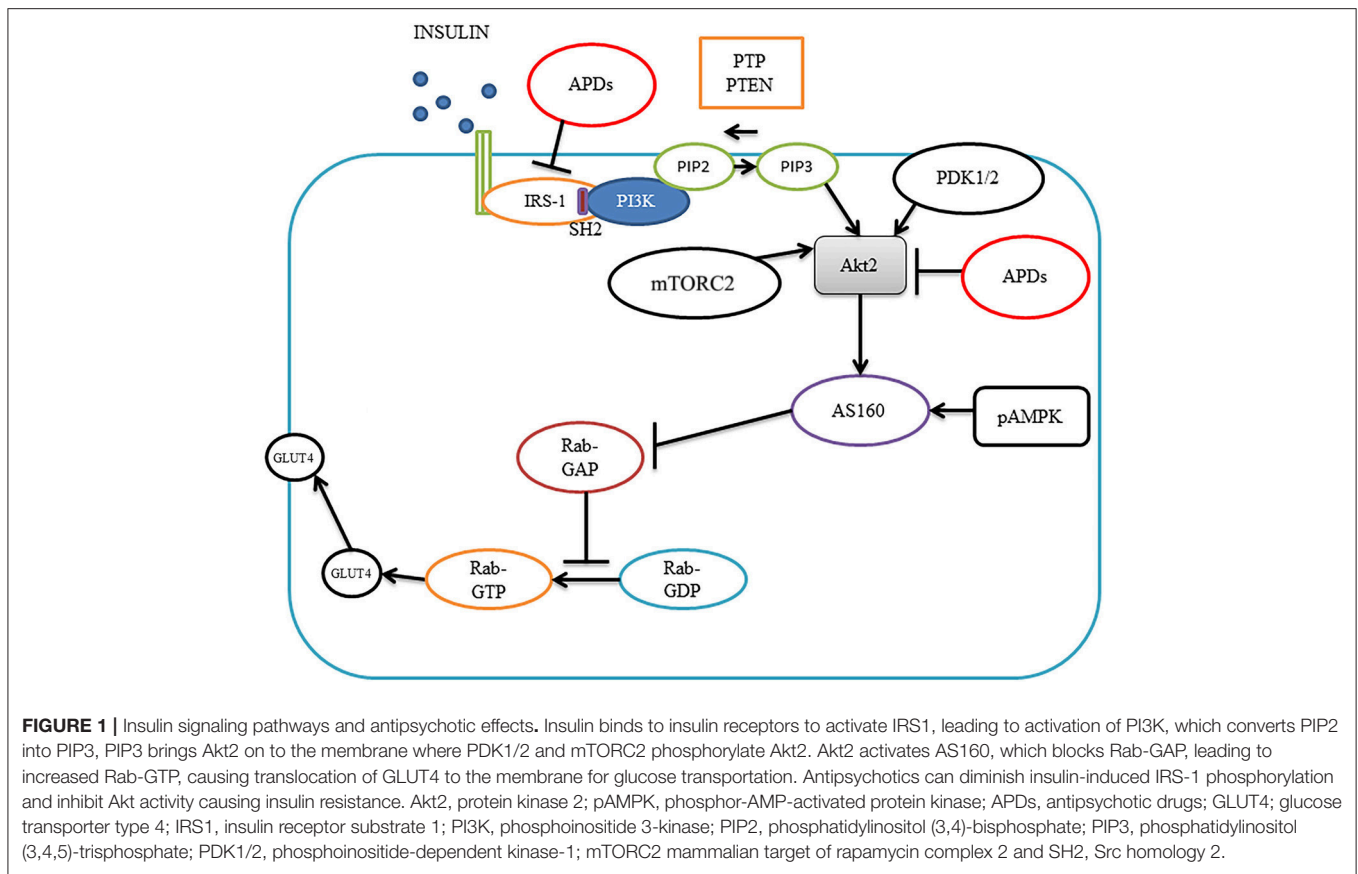
APD-induced insulin resistance could be independent of weight gain and increased food intake. It has been reported that, in patients within 3 months after initiation or switch to atypical APDs, new-onset gluco-metabolic abnormalities and diabetes was not associated with weight change and BMI (van Winkel et al., 2008). A recent study has shown that a single administration of olanzapine caused glucose metabolism change independent of obesity in healthy human subjects (Hahn et al., 2013). Aripiprazole has been shown to induce insulin resistance with metabolic changes where there is no weight gain or increase in food intake (Teff et al., 2013). Boyda et al. examined the acute effects (60, 180, or 360 min) of olanzapine, clozapine, risperidone and haloperidol on insulin resistance in rats at both low and high concentrations of clozapine (2 mg/kg; 20 mg/kg), olanzapine (1.5 mg/kg vs. 15 mg/kg), risperidone (0.5 mg/kg vs. 2.5 mg/kg) and haloperidol (0.1 mg/kg vs. 1.0 mg/kg) (Boyda et al., 2010b). Insulin resistance was evaluated through HOMA-IR (homeostasis model assessment index for insulin resistance) values in fasted rats and glucose clearance during a glucose tolerance test. Both olanzapine and clozapine produced significant dose and time dependent effects on fasting plasma glucose and insulin concentrations, HOMA-IR values, insulin resistance and glucose intolerance (Boyda et al., 2010b; Liu et al., 2017). However, risperidone and haloperidol also caused significant increases in fasting glucose and/or insulin levels at the high dose, 60 min post-drug administration (Boyda et al., 2010b).

Insulin plays an important role in glucose consumable cells via the phosphoinositide 3-kinase (PI3K)/protein kinase B (PKB, also known as Akt) pathway (**Figure 1**). Insulin can bind to

insulin receptors, leading to receptor autophosphorylation which in turn activates receptor tyrosine kinases that phosphorylate insulin receptor substrates (IRSs) including IRS1, IRS2, IRS3, IRS4, Gab1 (a member of the IRS1-like multisubstrate docking protein family) and Shc protein (Boucher et al., 2014). Phosphorylated IRSs activate PI3K by binding to its regulatory subunit via Src homology 2 (SH2) domains, increasing the conversion of phosphatidylinositol (3,4)-bisphosphate (PIP2) into phosphatidylinositol (3,4,5)-trisphosphate (PIP3). PIP3 can increase the activities of 3-phosphoinositide-dependent protein kinases 1/2 (PDK1/2). PIP3 also binds to Akt and this leads to translocation of Akt to the membrane, where Akt is phosphorylated by PDK1/2 and mammalian target of rapamycin complex 2 (mTORC2) and activated. Activated Akt increases glucose transporter type 4 (GLUT4) translocation to the membrane to increase glucose uptake via Akt substrate AS160 (Boucher et al., 2014). There are three Akt isoforms; 1, 2, and 3. The isoform regulating GLUT4 is Akt 2. GLUT4 is mainly located in adipose tissue and striated muscle (skeletal and cardiac) as well as the neurons of the hippocampus. Inhibition of Akt by siRNA has been shown to abolish the effect of insulin on glucose transport in 3T3-L1 adipocytes (Jiang et al., 2003). Insulin action is impaired in the liver, adipose tissue and muscle of Akt2 knockout mice (Cho et al., 2001). There are also two natural negative regulators in the pathway: protein tyrosine phosphatases (PTPs) and phosphatase and tension homolog deleted on chromosome 10 (PTEN); PTPs dephosphorylate insulin receptors and PTEN converts PIP3 into PIP2 (Boucher et al., 2014). Deficiencies in PTPs and PTEN have been shown to increase insulin sensitivity in mice (Boucher et al., 2014).

In type 2 diabetes patients, it has been shown that insulin-induced IRS-1 tyrosine phosphorylation, PI3K and Akt activities are decreased in skeletal muscles (Fröjdö et al., 2009). In adipose tissue, Akt2 phosphorylation is impaired in obese type 2 diabetes (Rondinone et al., 1999). In animal models, high fat-diet rats have defective insulin-induced GLUT-4 translocation in skeletal muscle which has been associated with decreased Akt activity (Tremblay et al., 2001). Therefore, abnormality of the insulin pathway in insulin target cells could be critical for the pathogenesis of diabetes.

APDs have been shown to inhibit Akt activity and thus cause insulin resistance in muscle cells (Engl et al., 2005). In L6 myotubes, olanzapine was shown to decrease glycogen content in a dose- and time- dependent fashion. Olanzapine diminished insulin-induced IRS-1 phosphorylation and abolished insulin-induced pPI3K, pAkt, and pGSK-3, while amisulpride, which does not cause diabetes, did not result in the above changes, indicating the importance of the insulin pathway in APD-induced diabetes (Engl et al., 2005). Olanzapine has been found to attenuate phosphorylation of insulin-like growth factor receptor (IGF-R) and IRS1 stimulated by insulin in wild-type fibroblast cells (Alghamdi et al., 2014). The mechanism has been considered to be olanzapine-induced membrane associated mammalian neuraminidase-3 (Neu3) and Neu 1 sialidase activity reduction. Alghamdi reported that Neu1 and matrix metalloproteinase-9 (MMP-9) could cross-talk in alliance with the neuromedin B G-protein coupled receptor (GPCR) to promote the insulin



signaling pathway (Alghamdi et al., 2014). Clozapine has been reported to reduce insulin-stimulated glucose uptake in PC12 and in L6 cells, which was accompanied with 40% decreased insulin effect on insulin receptor, 60% decrease in IRS1 tyrosine phosphorylation, and 40% decrease in insulin-stimulated Akt phosphorylation (Panariello et al., 2012). The inhibitory effect of APDs on Akt activity has also been reported in T-cells and glioblastoma cells (Shin et al., 2006; Chen M.-L. et al., 2011). However, the effects of APDs on the other elements of the insulin pathway such as PTPs and PTEN have not been studied and warrant further investigation.

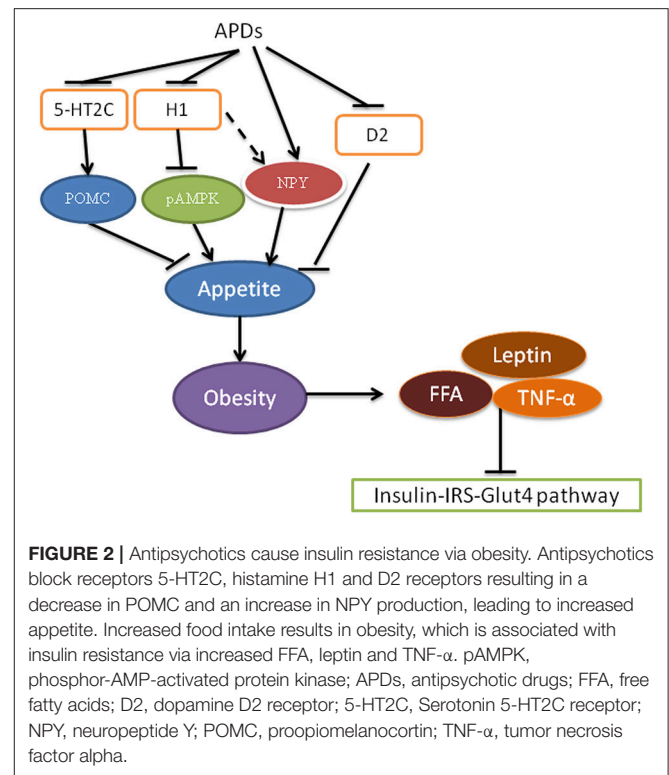
## Mechanism 2: Antipsychotic-Caused Insulin Resistance through Obesity

APDs are a well-known cause of obesity which is closely associated with diabetes. It has been repeatedly reported that APD-induced insulin resistance and diabetes are associated with the increased weight gain, BMI and intra-abdominal adiposity, particularly in patients with chronic APD treatment (Bou Khalil, 2012; Manu et al., 2012). As many as 20–50% of APD treated patients are obese and have diabetes (Deng, 2013). Animal experiments also demonstrated that APDs can cause obesity, although there are some limitations (Boyda et al., 2010a; Weston-Green et al., 2011; van der Zwaal et al., 2014). One limitation is a failure to induce weight gain by clozapine in rodents in the majority of studies, although clozapine has been well established

to cause dyslipidaemia, hyperglycaemia and insulin resistance in rodents (Boyda et al., 2010a; Liu et al., 2017), which provides a model to investigate the direct effects of clozapine on these metabolic parameters in rats without weight gain (Cooper et al., 2008; Liu et al., 2017). However, a recent study has reported that chronic clozapine treatment induced weight gain and fasting glucose levels in male rats (von Wilmsdorff et al., 2013). The most successfully established rodent model for APD-induced weight gain/obesity is by olanzapine (Weston-Green et al., 2011; van der Zwaal et al., 2014; Lord et al., 2017). Other APDs such as risperidone, sulpiride, haloperidol, and chlorpromazine have also been established to cause some degree of weight gain in rats and mice (Boyda et al., 2010a; Lian et al., 2015). There is also a sexual dimorphism in the obesogenic response to APDs, which is particularly clear in rodent models with male rats repeatedly reported less sensitive to APD-induced weight gain than females (Minet-Ringuet et al., 2006; Weston-Green et al., 2010; von Wilmsdorff et al., 2013). Although sex differences are less established in humans, females have been often observed with higher risks for weight gain and other metabolic side-effects (including increased insulin-resistance and higher plasma triglycerides) caused by APDs (Wu et al., 2007; Seeman, 2009; Weston-Green et al., 2010). Based on data from both clinical and animal studies, there are three stages in APD-caused obesity: during the first few months treatment in schizophrenia patients, APDs cause rapid weight gain (stage 1) followed by a steady

weight gain for about 1 year (stage 2) to reach a plateau and then a stage (stage 3) for maintaining heavy weight (Zipursky et al., 2005; Pai et al., 2012; Deng, 2013). Similar developmental stages of APD-induced obesity have also been observed in rat models (Huang et al., 2006; He et al., 2014). Therefore, APD-caused obesity is a chronic process.

The mechanism for APD-caused obesity is considered to be increased appetite and food intake rather than decreased energy expenditure in the early stage of obesity development (Deng, 2013). APDs can act as an antagonist on serotonin 5-HT<sub>2C</sub>, histamine H<sub>1</sub>, and dopamine D<sub>2</sub> receptors to increase appetite and thus increase food intake, leading to obesity (Figure 2; Matsui-Sakata et al., 2005; Han et al., 2008; Nasrallah, 2008; Kirk et al., 2009; Deng et al., 2010; Lian et al., 2016). It has been found that inhibition of 5-HT<sub>2C</sub> by olanzapine contributes to olanzapine-induced weight gain and hyperphagia in rodents (Kirk et al., 2009; Lord et al., 2017), while these effects were blunted in mice lacking 5-HT<sub>2C</sub> (*htr2c*-null mice) (Lord et al., 2017). 5-HT<sub>2C</sub> can promote anorexigenic proopiomelanocortin (POMC) neurons to decrease appetite (Lam et al., 2008, 2010; Lian et al., 2016). Indeed, olanzapine has been shown to decrease POMC expression in rats (Ferno et al., 2011; Weston-Green et al., 2012a; Lian et al., 2014a; Zhang et al., 2014a). Matsui-Sakata showed that H<sub>1</sub> receptor blockage is the main reason for APD-induced obesity (Matsui-Sakata et al., 2005). APDs block H<sub>1</sub> receptors and thus activate hypothalamic 5' AMP-activated protein kinase (AMPK), which stimulates appetite in rats (Kim et al., 2007; Han et al., 2008; Deng et al., 2010; He et al., 2013). It is interesting that an histamine H<sub>1</sub> agonist (2-(3-trifluoromethylphenyl) histamine) is able to reverse increased hypothalamic AMPK activation and hyperphagia induced by olanzapine in rats (He et al., 2014). Fernø and colleagues have reported a decreased hypothalamic AMPK phosphorylation by subchronic olanzapine treatment and no effects after acute treatment of olanzapine or clozapine in rats (Ferno et al., 2011), however they measured AMPK phosphorylation 20 h after the final drug treatment that might explain their failures to detect the increased AMPK activation by APDs in this study. In fact, a further study by the same research group has proven the important role of hypothalamic AMPK activation in olanzapine-induced weight gain by means of adenovirus-mediated inhibition of AMPK in the arcuate nuclei of female rats with olanzapine depot exposure (Skrede et al., 2014). Furthermore, direct intracerebroventricular infusion of olanzapine has been reported to induce hypothalamic AMPK activation and hepatic insulin resistance in rats (Martins et al., 2010). APDs have also been found to increase neuropeptide Y (NPY) expression (Lian et al., 2014a; Zhang et al., 2014a), which could be partially through blocking H<sub>1</sub> receptors and reversed by H<sub>1</sub> agonist (Lian et al., 2014a,b, 2016). The D<sub>2</sub> receptor is associated with the reward system and thus appetite (Deng, 2013). APDs can bind and block D<sub>2</sub> receptors to increase appetite (Nasrallah, 2008; Deng, 2013). Recently a study has shown that the gut microbiome plays an important role in APD-induced obesity in rats (Davey et al., 2013). In view of the importance of this microbiome in diet-induced obesity (Bäckhed et al., 2004, 2007; Turnbaugh et al., 2009), Davey et al. tested the role of this microbiome



**FIGURE 2 |** Antipsychotics cause insulin resistance via obesity. Antipsychotics block receptors 5-HT<sub>2C</sub>, histamine H<sub>1</sub> and D<sub>2</sub> receptors resulting in a decrease in POMC and an increase in NPY production, leading to increased appetite. Increased food intake results in obesity, which is associated with insulin resistance via increased FFA, leptin and TNF- $\alpha$ . pAMPK, phosphor-AMP-activated protein kinase; APDs, antipsychotic drugs; FFA, free fatty acids; D<sub>2</sub>, dopamine D<sub>2</sub> receptor; 5-HT<sub>2C</sub>, Serotonin 5-HT<sub>2C</sub> receptor; NPY, neuropeptide Y; POMC, proopiomelanocortin; TNF- $\alpha$ , tumor necrosis factor alpha.

in olanzapine-induced obesity and found that use of antibiotics resulted in decreased fat accumulation in rats treated with olanzapine (Davey et al., 2013). Regarding to energy expenditure, similar to the sedative effects of APDs on humans, it has been widely reported that APDs such as olanzapine and risperidone decreased locomotor activities in both rats and mice (Fell et al., 2007; Cope et al., 2009; Weston-Green et al., 2011; Zhang et al., 2014b; Lian et al., 2015; Lord et al., 2017). Although a recent study found an increased heat production in mice treated with olanzapine (Lord et al., 2017), a previous study reported a decrease of metabolic rate in mice with olanzapine infusion (8 mg/kg/day) but not in mice with a lower dose (4 mg/kg/day) (Coccorello et al., 2009). Furthermore, a number of studies in rats reported that olanzapine reduced core body temperature or thermogenesis of brown adipose tissue (BAT) accompanied with decreased expression of functional thermogenic proteins, uncoupling protein1 (UCP1) and peroxisome proliferator-activated receptor gamma coactivator-1 $\alpha$  (PGC-1 $\alpha$ ) of BAT (Stefanidis et al., 2009; van der Zwaal et al., 2012; Zhang et al., 2014b). Therefore, further studies are necessary to clarify the effects of APDs on thermogenesis.

Obesity is closely related to diabetes through insulin resistance and inflammation (Kahn et al., 2014; Verma and Hussain, 2017). In obesity, blood insulin levels are increased. However, in obese rats, the efficacy of insulin to stimulate Akt2 activity decreases in the muscle and liver, suggesting insulin resistance in these tissue (Kim et al., 2000). In obesity, increased blood levels of free fatty acids (FFA), tumor necrosis factor alpha (TNF-alpha) and leptin have been demonstrated to contribute to the development of type

2 diabetes (Verma and Hussain, 2017). Blood levels of FFA were increased in obesity due to an increased release from an increased size and number of adipocytes (Verma and Hussain, 2017). Increased FFA levels in turn inhibit insulin/IRS phosphorylation, leading to insulin resistance in insulin target cells such as skeletal muscle, the liver and endothelial cells (Kahn et al., 2006; Wilding, 2007). Decreased IRS phosphorylation causes decreased PI3K/Akt pathway activity and GLUT4 translocation (Wilding, 2007). Lowering the levels of FFA has been shown to increase insulin sensitivity in obese and non-obese diabetic subjects, indicating the important role of FFA in insulin resistance (Goodpaster and Coen, 2014). Exercise has been shown to reduce FFA and improve insulin sensitivity (Kwak, 2013; Malin et al., 2016).

Olanzapine has been shown to increase lipogenesis and thus increase FFA (Albaugh et al., 2011a). Recent studies showed that blood levels of FFA were significantly increased in rats after chronic administration of olanzapine (Davey et al., 2013; Liu et al., 2015, 2017). Jassim et al showed that a single injection of clozapine and olanzapine in rats increased serum FFA after 12–24 h (Jassim et al., 2012). A recent study in male rats showed that olanzapine could stimulate lipogenesis independent of weight gain, but with dosage-dependent effects (Ferno et al., 2015). All these studies support the view that APD-induced obesity could contribute to insulin resistance partially via increased blood levels of FFA.

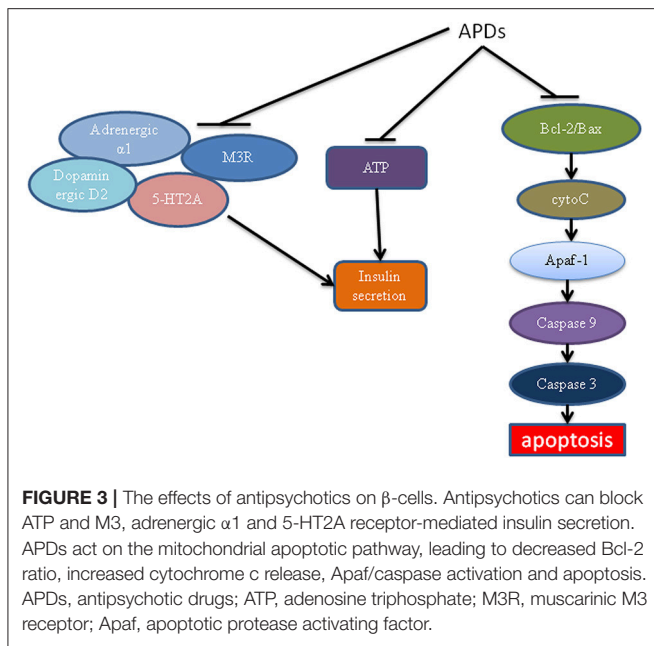
TNF-alpha, which is increased in obesity, may also play a key role in the link between obesity and diabetes (Verma and Hussain, 2017). Both cell culture and animal studies have shown TNF-alpha to interfere with the insulin pathway causing insulin resistance (Nieto-Vazquez et al., 2008). Knockout of TNF-alpha decreased obesity-caused insulin resistance (Uysal et al., 1997). In addition, Interleukin 1 (IL-1), IL-6 and leptin are also increased in obesity and have been associated with insulin resistance (Verma and Hussain, 2017). Chronic treatment (46 days) with olanzapine has been reported to increase TNF alpha expression in adipose tissue and plasma and IL-1 expression in rat plasma (Victoriano et al., 2010). In addition, administration of olanzapine and other atypical APDs has been shown to increase blood levels of leptin and ghrelin, suggesting that leptin and ghrelin could also be a link between APD-induced obesity and diabetes (Jin et al., 2008; Albaugh et al., 2011b; Zhang Q. et al., 2013). As an adipocyte-derived hormone, adiponectin controls lipid and carbohydrate metabolism, in which a low adiponectin level is associated with obesity, insulin resistance and type 2 diabetes (Jin et al., 2008). Schizophrenia patients with metabolic disorders have been reported with a low adiponectin levels (Hanssens et al., 2008; Oriot et al., 2008; Chen P.-Y. et al., 2011; Sugai et al., 2012). Accumulated evidence has shown that treatment with atypical APDs clozapine and olanzapine decreases plasma adiponectin levels associated with weight gain, high fasting glucose, hypertriglyceridemia, insulin resistance induced by these drugs (Hanssens et al., 2008; Bai et al., 2009; Wampers et al., 2012; Klemettila et al., 2014), although it should be noted that some atypical APDs (e.g., risperidone) did not affect adiponectin levels (Murashita et al., 2007; Wampers et al., 2012).

### Mechanism 3: Antipsychotic-Induced $\beta$ -Cell Dysfunction and Apoptosis

Recent studies have paid attention to the role of  $\beta$ -cells in APD-induced diabetes. Pancreatic  $\beta$ -cells are the only type of cells that secrete insulin. The pathogenesis of diabetes involves the altered mass and function of  $\beta$ -cells (Kahn et al., 2014). It has been recognized that  $\beta$ -cell damage is necessary for the pathogenesis of type 2 diabetes (Kahn et al., 2014). Insulin resistance alone is regarded as insufficient to cause diabetes. According to  $\beta$ -cell damage, the development of diabetes is classified into five stages (Weir and Bonner-Weir, 2004). Stage 1 is compensation in which insulin secretion is increased and blood glucose levels are maintained at normal levels. Stage 2 is characterized by the mild rise of blood glucose levels to 5.0–6.5 mmol/L in which there are loss of  $\beta$ -cell mass and function. Stage 3 is early decompensation, in which maintenance of a normal range of blood glucose levels does not occur due to the inability of damaged  $\beta$ -cells to secrete insulin. Stage 4 is characterized by stable decompensation and significant  $\beta$ -cell damage. Stage 5 is major decompensation where profound reduction of  $\beta$ -cell mass and ketosis occurs. Therefore, protection of  $\beta$ -cells and promotion of human  $\beta$ -cell proliferation have been investigated in preclinical models to improve diabetes (Xiao et al., 2014; Shirakawa and Kulkarni, 2016).

APDs can cause  $\beta$ -cell damage directly and indirectly. The first evidence for APD-induced  $\beta$ -cell damage was from the studies of olanzapine-induced insulin resistance in canine models in comparison with a high-fat diet (Ader et al., 2005; Bergman and Ader, 2005). It was found that there was no expected  $\beta$ -cell compensation for olanzapine-induced insulin resistance (Ader et al., 2005; Bergman and Ader, 2005). This is different from insulin resistance induced by a high-fat diet which confers  $\beta$ -cell compensation, suggesting that olanzapine can cause  $\beta$ -cell damage. Chintoh et al also demonstrated that APDs caused  $\beta$ -cell damage in a rat model (Chintoh et al., 2009). It was found that acute treatment with clozapine and olanzapine at the dosages of 10 and 3 mg/kg respectively produced insulin resistance evidenced by decreased glucose infusion, increased hepatic glucose production and decreased peripheral glucose utilization (Chintoh et al., 2009). Clozapine and olanzapine also decreased  $\beta$ -cell insulin secretion under hyperglycaemic clamp, indicating that direct  $\beta$ -cell damage accompanied clozapine- and olanzapine- induced insulin resistance. Olanzapine and clozapine have been shown to decrease insulin secretion in islet  $\beta$ -cells (Johnson et al., 2005; Weston-Green et al., 2013). In isolated rat islets, olanzapine and clozapine inhibited cholinergic-glucose induced insulin secretion but risperidone and ziprasidone did not (Johnson et al., 2005). These results may account for the fact that diabetes is common in patients treated with clozapine and olanzapine as  $\beta$ -cell damage can accelerate the development of diabetes from insulin resistance.

Studies have now shown various mechanisms for APD-caused  $\beta$ -cell dysfunction (**Figure 3**). APDs can act on several  $\beta$ -cell receptors to cause decreased insulin secretion. Binding experiments showed that APDs can bind to various receptors including dopaminergic, histaminergic, serotonergic, adrenergic



and muscarinic receptors (Guenette et al., 2013). It is known that blockade of the 5-HT2A and muscarinic M3 receptors weakens insulin response to glucose challenge, whereas antagonism at the dopamine D2 receptor increases insulin secretion (Rubí et al., 2005; Hahn et al., 2011; Weston-Green et al., 2013). Guenette et al. used a series of antagonists to screen their effects on insulin secretion including prazosin (a selective adrenergic  $\alpha 1$  antagonist), idazoxan (a selective adrenergic  $\alpha 2$  antagonist), SB242084 (a selective 5-HT2C antagonist), WAY100635 (a selective 5HT1A antagonist), and MDL100907 (a selective 5-HT2A antagonist). It was found that only prazosin and MDL100907 were associated with decreased insulin secretion (Guenette et al., 2013). Olanzapine and clozapine may bind to these receptors, blocking them to stimulate insulin secretion (Guenette et al., 2013). Interestingly, antagonist affinity with M3 has been demonstrated to be a main indicator of diabetes induced by APDs (Silvestre and Prous, 2005). APDs which have high binding affinity with M3 receptors, such as olanzapine and clozapine, have been shown to decrease insulin secretion (Johnson et al., 2005; Weston-Green et al., 2012b), providing clear evidence for the important role of M3 receptors in the direct effect of APDs on  $\beta$ -cells (Deng, 2013; Weston-Green et al., 2013).

APDs have also been linked to an increase in apoptosis of  $\beta$ -cells, leading to lower mass and thus decreased insulin secretion. Ozasa et al. demonstrated that APDs can act directly on  $\beta$ -cells to cause apoptosis (Ozasa et al., 2013). Therefore,  $\beta$ -cell mass is decreased, leading to decreased insulin secretion. Clozapine has also been shown to damage  $\beta$ -cells to cause diabetes (Best et al., 2005). The effect of APD-induced  $\beta$ -cell damage may be mediated by the mitochondrial apoptotic pathway (Figure 3). This pathway was regulated by proapoptotic proteins such as Bak, Bax, Bid, Bim, Bcl-associated death promoter (BAD), Noxa, and PUMA (Chen et al., 2013). It was also regulated by antiapoptotic

proteins such as Bcl-2, Bcl-xL, Bcl-w, Mcl-1, and A1 (Chen et al., 2013). A changed ratio of proapoptotic proteins to antiapoptotic proteins can lead to the release of cytochrome c, resulting in promotion of the formation of active apaf (apoptotic protease activating factor)/caspase-9 complex (apoptosome), leading to caspase-3 activation and apoptosis. Contreras-Shannon et al. showed that clozapine caused mitochondrial damage in several types of insulin responding cells, including cultured mouse myoblasts (C2C12), adipocytes (3T3-L1), hepatocytes (FL-83B), and monocytes (RAW 264.7) (Contreras-Shannon et al., 2013). A proteomic study has also shown that olanzapine and clozapine cause dramatic mitochondrial protein changes in the hippocampus of rats (Ji et al., 2009). Although neither study was carried out in  $\beta$ -cells, they showed APDs had a direct effect on the mitochondrial apoptotic pathway. The effect of APDs on  $\beta$ -cell mitochondria warrants detailed investigation. It may reveal a major mechanism for APD-induced diabetes.

Decreased insulin secretion caused by APDs could also be mediated by ATP. ATP is mainly produced in mitochondria and known to regulate insulin secretion (Seino, 2012). In physiological condition, glucose metabolism increases ATP production. Accumulation of ATP in  $\beta$ -cells leads to closing of the  $K_{ATP}$  channels and opening of the voltage-dependent  $Ca^{2+}$  channels, resulting in  $Ca^{2+}$  influx to trigger insulin secretion (Seino, 2012). It has been shown that treatment of insulin responsive cells with clozapine reduced ATP production. It is possible that APDs also affect ATP production in  $\beta$ -cells.

In summary, APDs can affect the  $\beta$ -cell insulin secretion function. This may be mediated by several receptors particularly the M3 receptors. APDs could also cause  $\beta$ -cell apoptosis and decrease production of ATP. Further detailed studies will be helpful to elucidate these important mechanisms.

## PREVENTIVE AND THERAPEUTIC IMPLICATIONS

As discussed above, multiple mechanisms are involved in APD-induced diabetes including increased appetite, insulin resistance and  $\beta$ -cell damage. Prevention of these changes caused by APDs could be critical for medication compliance and improving clinical outcomes. To date, dozens of drugs have been trialed with some success in partly ameliorating antipsychotic-induced metabolic side-effects. Amantadine, metformin, reboxetine, sibutramine and topiramate have been shown to be effective in reducing APD-induced weight gain in Baptista et al's examination of 25 pharmacologic weight loss intervention trials ( $n = 1,221$ ) (Baptista et al., 2008). Metformin was demonstrated to have the most promising effects on weight loss, followed by d-fenfluramine, sibutramine, topiramate, and reboxetine, in a meta-analysis of 32 placebo-controlled pharmacologic intervention trials involving 1,482 subjects (Maayan et al., 2010). Therefore, these drugs could ameliorate APD-induced insulin resistance via reducing obesity. Furthermore, the meta-analysis also showed that metformin and rosiglitazone (two antidiabetic drugs) can decrease insulin levels and insulin resistance (assessed using HOMA-IR) in patients treated with atypical APDs,

even though they have no effects on fasting glucose levels (Maayan et al., 2010). However, in two studies, metformin was found to decrease fasting glucose along with decreased insulin and HOMA-IR in chronic schizophrenia patients treated with olanzapine (Baptista et al., 2006; Chen et al., 2008). In animal experiments, metformin and glyburide have been shown to reduce olanzapine-caused hyperglycaemia (Boyda et al., 2014). These results suggest that these antidiabetic drugs may directly ameliorate insulin resistance. Although metformin outperformed other agents, the current evidence is still too limited to support it as a regular adjunctive medication to control antipsychotic-induced weight gain and metabolic abnormalities (Maayan et al., 2010). Recently, a 16 weeks randomized clinical trial examined the effects of liraglutide (a glucagon-like peptide-1 [GLP-1] receptor agonist) used in conjunction with clozapine or olanzapine treatment on prediabetes and overweight/obesity in patients with schizophrenia spectrum disorders (Larsen et al., 2017). In the patients with liraglutide co-treatment, glucose tolerance was significantly improved, and particularly 63.8% prediabetic patients developed normal glucose tolerance. In addition, liraglutide induced weight loss and reduction in a number of cardiometabolic parameters including waist circumference, systolic blood pressure, visceral fat and low-density lipoprotein (Larsen et al., 2017).

Based on the key roles of histamine H1 and 5-HT<sub>2C</sub> antagonisms in APD-induced weight gain, a number of animal and/or clinical trials targeted at these receptors have been examined recently. It has been shown that co-treatment with betahistine (an H1 agonist and H3 antagonist) could partially reduce olanzapine-induced weight gain in both schizophrenia patients and animal models (Deng et al., 2012; Poyurovsky et al., 2013; Lian et al., 2014a,b, 2016). Betahistine co-treatment can also ameliorate the increased triglyceride accumulation and non-esterified fatty acids caused by olanzapine via modulating AMPK-SREBP-1 and PPAR $\alpha$ -dependent pathways (Liu et al., 2015). A recent animal trial found that lorcaserin (a 5-HT<sub>2C</sub>-specific agonist) could not only partially reduced olanzapine-induced food intake and weight gain, but also improved glucose tolerance in female mice (Lord et al., 2017). In future, the major effort could be the elucidation of the mechanisms for APD-induced diabetes, particularly APD-induced  $\beta$ -cell damage. It is only the beginning to realize the important role of  $\beta$ -cells

in APD-induced diabetes. Since APDs may decrease insulin secretion through blocking M3 receptors on  $\beta$ -cells (Johnson et al., 2005; Weston-Green et al., 2013), it is worth to trial whether a specific M3 agonist could mitigate APD-induced diabetes. Drugs or phytochemicals that can protect  $\beta$ -cells may be used for the prevention of APD-induced diabetes. For example, resveratrol and curcumin have been demonstrated to be effective to protect  $\beta$ -cells in streptozotocin-induced damage (Su et al., 2006; Meghana et al., 2007; Palsamy and Subramanian, 2010), which may be used in APD-induced  $\beta$ -cell damage.

## CONCLUSIONS

APDs are associated with diabetes and hyperglycaemic emergency. This could be mediated by multiple mechanisms. APDs can inhibit the insulin signaling pathway in insulin sensitive cells such as muscle cells, hepatocytes and adipocytes to cause insulin resistance. APD-induced obesity can result in high levels of FFA and inflammation, which can cause insulin resistance as well. More importantly, APDs can cause direct damage to  $\beta$ -cells. This accounts for the rapid development of diabetes and severe diabetic complications. As multiple mechanisms are involved in APD-induced diabetes, preventive and therapeutic approaches may need to combine agents counteracting different mechanisms. Among them prevention of  $\beta$ -cell damage is of particular importance as diabetes would not happen without  $\beta$ -cell damage.

## AUTHOR CONTRIBUTIONS

JC and CD conceived and outlined the manuscript. JC wrote the first draft of the manuscript. CD, X-FH, RS, and CC contributed to manuscript writing, editing, synthesis of previous literature.

## FUNDING

This study was supported by a Project grant (APP1044624) from the National Health and Medical Research Council (NHMRC), Australia to CD and X-FH. The funding organization did not play a role in the design and conduct of the study. The authors have no conflicts of interest.

## REFERENCES

- Ader, M., Kim, S. P., Catalano, K. J., Ionut, V., Hucking, K., Richey, J. M., et al. (2005). Metabolic dysregulation with atypical antipsychotics occurs in the absence of underlying disease a placebo-controlled study of olanzapine and risperidone in dogs. *Diabetes* 54, 862–871. doi: 10.2337/diabetes.54.3.862
- Albaugh, V. L., Judson, J. G., She, P., Lang, C. H., Maresca, K. P., Joyal, J. L., et al. (2011a). Olanzapine promotes fat accumulation in male rats by decreasing physical activity, repartitioning energy and increasing adipose tissue lipogenesis while impairing lipolysis. *Mol. Psychiatry* 16, 569–581. doi: 10.1038/mp.2010.33
- Albaugh, V. L., Singareddy, R., Mauger, D., and Lynch, C. J. (2011b). A double blind, placebo-controlled, randomized crossover study of the acute metabolic effects of olanzapine in healthy volunteers. *PLoS ONE* 6:e22662. doi: 10.1371/journal.pone.0022662
- Alghamdi, F., Guo, M., Abdulkhalek, S., Crawford, N., Amith, S. R., and Szewczuk, M. R. (2014). A novel insulin receptor-signaling platform and its link to insulin resistance and type 2 diabetes. *Cell. Signal.* 26, 1355–1368. doi: 10.1016/j.cellsig.2014.02.015
- Bäckhed, F., Ding, H., Wang, T., Hooper, L. V., Koh, G. Y., Nagy, A., et al. (2004). The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. U.S.A.* 101, 15718–15723. doi: 10.1073/pnas.0407076101
- Bäckhed, F., Manchester, J. K., Semenkovich, C. F., and Gordon, J. I. (2007). Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc. Natl. Acad. Sci. U.S.A.* 104, 979–984. doi: 10.1073/pnas.0605374104
- Bai, Y. M., Chen, T. T., Yang, W.-S., Chi, Y.-C., Lin, C.-C., Liou, Y.-J., et al. (2009). Association of adiponectin and metabolic syndrome among patients taking atypical antipsychotics for schizophrenia: a cohort study. *Schizophr. Res.* 111, 1–8. doi: 10.1016/j.schres.2009.03.014



- Baptista, T., Elfakih, Y., Uzcategui, E., Sandia, I., Talamo, E., Araujo de Baptista, E., et al. (2008). Pharmacological management of atypical antipsychotic-induced weight gain. *CNS Drugs* 22, 477–495. doi: 10.2165/00023210-200822060-00003
- Baptista, T., Martínez, J., Lacruz, A., Rangel, N., Beaulieu, S., Serrano, A., et al. (2006). Metformin for prevention of weight gain and insulin resistance with olanzapine: a double-blind placebo-controlled trial. *Can. J. Psychiatry* 51, 192–196. doi: 10.1177/070674370605100310
- Bergman, R. N., and Ader, M. (2005). Atypical antipsychotics and glucose homeostasis. *J. Clin. Psychiatry* 66, 504–514. doi: 10.4088/JCP.v66n0414
- Best, L., Yates, A. P., and Reynolds, G. P. (2005). Actions of antipsychotic drugs on pancreatic  $\beta$ -cell function: contrasting effects of clozapine and haloperidol. *J. Psychopharmacol.* 19, 597–601. doi: 10.1177/0269881105056641
- Bobo, W. V., Cooper, W. O., Stein, C. M., Olfson, M., Graham, D., Daugherty, J., et al. (2013). Antipsychotics and the risk of type 2 diabetes mellitus in children and youth. *JAMA Psychiatry* 70, 1067–1075. doi: 10.1001/jamapsychiatry.2013.2053
- Boucher, J., Kleinridders, A., and Kahn, C. R. (2014). Insulin receptor signaling in normal and insulin-resistant states. *Cold Spring Harb. Perspect. Biol.* 6:a009191. doi: 10.1101/cshperspect.a009191
- Bou Khalil, R. (2012). Atypical antipsychotic drugs, schizophrenia, and metabolic syndrome in non-Euro-American societies. *Clin. Neuropharmacol.* 35, 141–147. doi: 10.1097/WNF.0b013e31824d5288
- Boyd, H. N., Procyshyn, R. M., Asiri, Y., Wu, C., Wang, C. K., Lo, R., et al. (2014). Antidiabetic-drug combination treatment for glucose intolerance in adult female rats treated acutely with olanzapine. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 48, 170–176. doi: 10.1016/j.pnpbp.2013.10.006
- Boyd, H. N., Tse, L., Procyshyn, R. M., Honer, W. G., and Barr, A. M. (2010a). Preclinical models of antipsychotic drug-induced metabolic side effects. *Trends Pharmacol. Sci.* 31, 484–497. doi: 10.1016/j.tips.2010.07.002
- Boyd, H. N., Tse, L., Procyshyn, R. M., Wong, D., Wu, T. K., Pang, C. C., et al. (2010b). A parametric study of the acute effects of antipsychotic drugs on glucose sensitivity in an animal model. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 34, 945–954. doi: 10.1016/j.pnpbp.2010.04.024
- Chen, C.-H., Chiu, C.-C., Huang, M.-C., Wu, T.-H., Liu, H.-C., and Lu, M.-L. (2008). Metformin for metabolic dysregulation in schizophrenic patients treated with olanzapine. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 32, 925–931. doi: 10.1016/j.pnpbp.2007.11.013
- Chen, J., Shao, R., Zhang, X. D., and Chen, C. (2013). Applications of nanotechnology for melanoma treatment, diagnosis, and theranostics. *Int. J. Nanomedicine* 8, 2677–2688. doi: 10.2147/IJN.S45429
- Chen, M.-L., Tsai, T.-C., Lin, Y.-Y., Tsai, Y.-M., Wang, L.-K., Lee, M.-C., et al. (2011). Antipsychotic drugs suppress the AKT/NF- $\kappa$ B pathway and regulate the differentiation of T-cell subsets. *Immunol. Lett.* 140, 81–91. doi: 10.1016/j.imlet.2011.06.011
- Chen, P.-Y., Huang, M.-C., Chiu, C.-C., Liu, H.-C., Lu, M.-L., and Chen, C.-H. (2011). Association of plasma retinol-binding protein-4, adiponectin, and high molecular weight adiponectin with metabolic adversities in patients with schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 35, 1927–1932. doi: 10.1016/j.pnpbp.2011.07.014
- Chintoh, A. F., Mann, S. W., Lam, L., Giacca, A., Fletcher, P., Nobrega, J., et al. (2009). Insulin resistance and secretion *in vivo*: effects of different antipsychotics in an animal model. *Schizophr. Res.* 108, 127–133. doi: 10.1016/j.schres.2008.12.012
- Cho, H., Mu, J., Kim, J. K., Thorvaldsen, J. L., Chu, Q., Crenshaw, E. B., et al. (2001). Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB $\beta$ ). *Science* 292, 1728–1731. doi: 10.1126/science.292.5522.1728
- Citrome, L., Collins, J., Nordstrom, B., Rosen, E., Baker, R., Nadkarni, A., et al. (2013). Incidence of cardiovascular outcomes and diabetes mellitus among users of second-generation antipsychotics. *J. Clin. Psychiatry* 74, 1199–1206. doi: 10.4088/JCP.13m08642
- Coccorello, R., Brina, D., Caprioli, A., Conti, R., Ghirardi, O., Schepis, F., et al. (2009). 30 days of continuous olanzapine infusion determines energy imbalance, glucose intolerance, insulin resistance, and dyslipidemia in mice. *J. Clin. Psychopharmacol.* 29, 576–583. doi: 10.1097/JCP.0b013e3181bfe13e
- Contreras-Shannon, V., Heart, D. L., Paredes, R. M., Navaira, E., Catano, G., Maffi, S. K., et al. (2013). Clozapine-induced mitochondria alterations and inflammation in brain and insulin-responsive cells. *PLoS ONE* 8:e59012. doi: 10.1371/journal.pone.0059012
- Cooper, G. D., Harrold, J. A., Halford, J. C. G., and Goudie, A. J. (2008). Chronic clozapine treatment in female rats does not induce weight gain or metabolic abnormalities but enhances adiposity: implications for animal models of antipsychotic-induced weight gain. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 32, 428–436. doi: 10.1016/j.pnpbp.2007.09.012
- Cope, M. B., Li, X., Jumbo-Lucioni, P., DiCostanzo, C. A., Jamison, W. G., Kesterson, R. A., et al. (2009). Risperidone alters food intake, core body temperature, and locomotor activity in mice. *Physiol. Behav.* 96, 457–463. doi: 10.1016/j.physbeh.2008.11.011
- Correll, C. U. (2010). From receptor pharmacology to improved outcomes: individualising the selection, dosing, and switching of antipsychotics. *Eur. Psychiatry* 25(Suppl. 2), S12–S21. doi: 10.1016/S0924-9338(10)71701-6
- Davey, K., Cotter, P., O'Sullivan, O., Crispie, F., Dinan, T., Cryan, J., et al. (2013). Antipsychotics and the gut microbiome: olanzapine-induced metabolic dysfunction is attenuated by antibiotic administration in the rat. *Transl. Psychiatry* 3:e309. doi: 10.1038/tp.2013.83
- De Hert, M., Detraux, J., van Winkel, R., Yu, W., and Correll, C. U. (2012). Metabolic and cardiovascular adverse effects associated with antipsychotic drugs. *Nat. Rev. Endocrinol.* 8, 114–126. doi: 10.1038/nrendo.2011.156
- Deng, C. (2013). Effects of antipsychotic medications on appetite, weight, and insulin resistance. *Endocrinol. Metab. Clin. North Am.* 42, 545–563. doi: 10.1016/j.ecl.2013.05.006
- Deng, C., Lian, J. M., Pai, N., and Huang, X. F. (2012). Reducing olanzapine-induced weight gain side effect by using betahistidine: a study in the rat model. *J. Psychopharmacol.* 26, 1271–1279. doi: 10.1177/0269881112449396
- Deng, C., Weston-Green, K., and Huang, X.-F. (2010). The role of histaminergic H1 and H3 receptors in food intake: a mechanism for atypical antipsychotic-induced weight gain? *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 34, 1–4. doi: 10.1016/j.pnpbp.2009.11.009
- Engl, J., Laimer, M., Niederwanger, A., Kranebitter, M., Starzinger, M., Pedrini, M., et al. (2005). Olanzapine impairs glycogen synthesis and insulin signaling in L6 skeletal muscle cells. *Mol. Psychiatry* 10, 1089–1096. doi: 10.1038/sj.mp.4001729
- Fell, M. J., Anjum, N., Dickinson, K., Marshall, K. M., Peltola, L. M., Vickers, S., et al. (2007). The distinct effects of subchronic antipsychotic drug treatment on macronutrient selection, body weight, adiposity, and metabolism in female rats. *Psychopharmacology* 194, 221–231. doi: 10.1007/s00213-007-0833-9
- Ferno, J., Ersland, K. M., Duus, I. H., Gonzalez-Garcia, I., Fossan, K. O., Berge, R. K., et al. (2015). Olanzapine depot exposure in male rats: dose-dependent lipogenic effects without concomitant weight gain. *Eur. Neuro-Psychopharmacol.* 25, 923–932. doi: 10.1016/j.euroneuro.2015.03.002
- Ferno, J., Varela, L., Skrede, S., Vazquez, M. J., Nogueiras, R., Dieguez, C., et al. (2011). Olanzapine-induced hyperphagia and weight gain associate with orexigenic hypothalamic neuropeptide signaling without concomitant AMPK phosphorylation. *PLoS ONE* 6:e20571. doi: 10.1371/journal.pone.0020571
- Fleischhacker, W. W., Siu, C. O., Bodén, R., Pappadopoulos, E., Karayal, O. N., and Kahn, R. S. (2013). Metabolic risk factors in first-episode schizophrenia: baseline prevalence and course analysed from the European First-Episode Schizophrenia Trial. *Int. J. Neuro-Psychopharmacol.* 16, 987–995. doi: 10.1017/S1461145712001241
- Foley, D. L., and Morley, K. I. (2011). Systematic review of early cardiometabolic outcomes of the first treated episode of psychosis. *Arch. Gen. Psychiatry* 68, 609–616. doi: 10.1001/archgenpsychiatry.2011.2
- Fraguas, D., Correll, C. U., Merchan-Naranjo, J., Rapado-Castro, M., Parellada, M., Moreno, C., et al. (2011). Efficacy and safety of second-generation antipsychotics in children and adolescents with psychotic and bipolar spectrum disorders: comprehensive review of prospective head-to-head and placebo-controlled comparisons. *Eur. Neuro-Psychopharmacol.* 21, 621–645. doi: 10.1016/j.euroneuro.2010.07.002
- Fröjdö, S., Vidal, H., and Pirola, L. (2009). Alterations of insulin signaling in type 2 diabetes: a review of the current evidence from humans. *Biochim. Biophys. Acta* 1792, 83–92. doi: 10.1016/j.bbadis.2008.10.019
- Fujii, N., Hirshman, M. F., Kane, E. M., Ho, R. C., Peter, L. E., Seifert, M. M., et al. (2005). AMP-activated protein kinase  $\alpha$ 2 activity is not essential for contraction- and hyperosmolarity-induced glucose transport in skeletal muscle. *J. Biol. Chem.* 280, 39033–39041. doi: 10.1074/jbc.M504208200

- Ginovart, N., and Kapur, S. (2012). Role of dopamine D(2) receptors for antipsychotic activity. *Handb. Exp. Pharmacol.* 212, 27–52. doi: 10.1007/978-3-642-25761-2\_2
- Goodpaster, B. H., and Coen, P. M. (2014). Improved mitochondrial function is linked with improved insulin sensitivity through reductions in FFA. *Diabetes* 63, 2611–2612. doi: 10.2337/db14-0277
- Guenette, M. D., Giacca, A., Hahn, M., Teo, C., Lam, L., Chintoh, A., et al. (2013). Atypical antipsychotics and effects of adrenergic and serotonergic receptor binding on insulin secretion *in vivo*: an animal model. *Schizophr. Res.* 146, 162–169. doi: 10.1016/j.schres.2013.02.023
- Hahn, M., Chintoh, A., Giacca, A., Xu, L., Lam, L., Mann, S., et al. (2011). Atypical antipsychotics and effects of muscarinic, serotonergic, dopaminergic and histaminergic receptor binding on insulin secretion *in vivo*: an animal model. *Schizophr. Res.* 131, 90–95. doi: 10.1016/j.schres.2011.06.004
- Hahn, M. K., Wolever, T. M., Arenovich, T., Teo, C., Giacca, A., Powell, V., et al. (2013). Acute effects of single-dose olanzapine on metabolic, endocrine, and inflammatory markers in healthy controls. *J. Clin. Psychopharmacol.* 33, 740–746. doi: 10.1097/JCP.0b013e31829e8333
- Han, M., Deng, C., Burne, T. H., Newell, K. A., and Huang, X.-F. (2008). Short-and long-term effects of antipsychotic drug treatment on weight gain and H1 receptor expression. *Psychoneuroendocrinology* 33, 569–580. doi: 10.1016/j.psyneuen.2008.01.018
- Hanssens, L., van Winkel, R., Wampers, M., Van Eyck, D., Scheen, A., Reginster, J.-Y., et al. (2008). A cross-sectional evaluation of adiponectin plasma levels in patients with schizophrenia and schizoaffective disorder. *Schizophr. Res.* 106, 308–314. doi: 10.1016/j.schres.2008.09.008
- He, M., Deng, C., and Huang, X.-F. (2013). The role of hypothalamic H1 receptor antagonism in antipsychotic-induced weight gain. *CNS Drugs* 27, 423–434. doi: 10.1007/s40263-013-0062-1
- He, M., Zhang, Q., Deng, C., Wang, H., Lian, J., and Huang, X.-F. (2014). Hypothalamic histamine H1 receptor-AMPK signaling time-dependently mediates olanzapine-induced hyperphagia and weight gain in female rats. *Psychoneuroendocrinology* 42, 153–164. doi: 10.1016/j.psyneuen.2014.01.018
- Huang, X.-F., Han, M., Huang, X., Zavitsanos, K., and Deng, C. (2006). Olanzapine differentially affects 5-HT2A and 2C receptor mRNA expression in the rat brain. *Behav. Brain Res.* 171, 355–362. doi: 10.1016/j.bbr.2006.03.040
- Jassim, G., Skrede, S., Vázquez, M. J., Wergedal, H., Vik-Mo, A. O., Lunder, N., et al. (2012). Acute effects of orexigenic antipsychotic drugs on lipid and carbohydrate metabolism in rat. *Psychopharmacology* 219, 783–794. doi: 10.1007/s00213-011-2397-y
- Ji, B., La, Y., Gao, L., Zhu, H., Tian, N., Zhang, M., et al. (2009). A comparative proteomics analysis of rat mitochondria from the cerebral cortex and hippocampus in response to antipsychotic medications. *J. Proteome Res.* 8, 3633–3641. doi: 10.1021/pr800876z
- Jiang, Z. Y., Zhou, Q. L., Coleman, K. A., Boese, Q., and Czech, M. P. (2003). Insulin signaling through Akt/protein kinase B analyzed by small interfering RNA-mediated gene silencing. *Proc. Natl. Acad. Sci. U.S.A.* 100, 7569–7574. doi: 10.1073/pnas.1332633100
- Jin, H., Meyer, J. M., Mudaliar, S., and Jeste, D. V. (2008). Impact of atypical antipsychotic therapy on leptin, ghrelin, and adiponectin. *Schizophr. Res.* 100, 70–85. doi: 10.1016/j.schres.2007.11.026
- Johnson, D. E., Yamazaki, H., Ward, K. M., Schmidt, A. W., Lebel, W. S., Treadway, J. L., et al. (2005). Inhibitory effects of antipsychotics on carbachol-enhanced insulin secretion from perfused rat islets: role of muscarinic antagonism in antipsychotic-induced diabetes and hyperglycemia. *Diabetes* 54, 1552–1558. doi: 10.2337/diabetes.54.5.1552
- Kahn, S. E., Cooper, M. E., and Del Prato, S. (2014). Pathophysiology and treatment of type 2 diabetes: perspectives on the past, present, and future. *Lancet* 383, 1068–1083. doi: 10.1016/S0140-6736(13)62154-6
- Kahn, S. E., Hull, R. L., and Utzschneider, K. M. (2006). Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature* 444, 840–846. doi: 10.1038/nature05482
- Kim, S. F., Huang, A. S., Snowman, A. M., Teuscher, C., and Snyder, S. H. (2007). Antipsychotic drug-induced weight gain mediated by histamine H1 receptor-linked activation of hypothalamic AMP-kinase. *Proc. Natl. Acad. Sci. U.S.A.* 104, 3456–3459. doi: 10.1073/pnas.0611417104
- Kim, Y.-B., Peroni, O. D., Franke, T. F., and Kahn, B. B. (2000). Divergent regulation of Akt1 and Akt2 isoforms in insulin target tissues of obese Zucker rats. *Diabetes* 49, 847–856. doi: 10.2337/diabetes.49.5.847
- Kirk, S. L., Glazebrook, J., Grayson, B., Neill, J. C., and Reynolds, G. P. (2009). Olanzapine-induced weight gain in the rat: role of 5-HT2C and histamine H1 receptors. *Psychopharmacology* 207, 119–125. doi: 10.1007/s00213-009-1639-8
- Klemettila, J.-P., Kampman, O., Seppala, N., Viikki, M., Hamalainen, M., Moilanen, E., et al. (2014). Cytokine and adipokine alterations in patients with schizophrenia treated with clozapine. *Psychiatry Res.* 218, 277–283. doi: 10.1016/j.psychres.2014.04.049
- Kwak, H.-B. (2013). Exercise and obesity-induced insulin resistance in skeletal muscle. *Integr. Med. Res.* 2, 131–138. doi: 10.1016/j.imr.2013.09.004
- Lam, D. D., Garfield, A. S., Marston, O. J., Shaw, J., and Heisler, L. K. (2010). Brain serotonin system in the coordination of food intake and body weight. *Pharmacol. Biochem. Behav.* 97, 84–91. doi: 10.1016/j.pbb.2010.09.003
- Lam, D. D., Przydzial, M. J., Ridley, S. H., Yeo, G. S., Rochford, J. J., O'Rahilly, S., et al. (2008). Serotonin 5-HT2C receptor agonist promotes hypophagia via downstream activation of melanocortin 4 receptors. *Endocrinology* 149, 1323–1328. doi: 10.1210/en.2007-1321
- Larsen, J. R., Vedtofte, L., Jakobsen, M. L., Jespersen, H. R., Jakobsen, M. I., Svensson, C. K., et al. (2017). Effect of liraglutide treatment on prediabetes and overweight or obesity in clozapine- or olanzapine-treated patients with schizophrenia spectrum disorder: a randomized clinical trial. *JAMA Psychiatry* 74, 719–728. doi: 10.1001/jamapsychiatry.2017.1220
- Leucht, S., Cipriani, A., Spineli, L., Mavridis, D., Orey, D., Richter, F., et al. (2013). Comparative efficacy and tolerability of 15 antipsychotic drugs in schizophrenia: a multiple-treatments meta-analysis. *Lancet* 382, 951–962. doi: 10.1016/S0140-6736(13)60733-3
- Leucht, S., Corves, C., Arbter, D., Engel, R. R., Li, C., and Davis, J. M. (2009). Second-generation versus first-generation antipsychotic drugs for schizophrenia: a meta-analysis. *Lancet* 373, 31–41. doi: 10.1016/S0140-6736(08)61764-X
- Lewis, S. W., Barnes, T. R., Davies, L., Murray, R. M., Dunn, G., Hayhurst, K. P., et al. (2006). Randomized controlled trial of effect of prescription of clozapine versus other second-generation antipsychotic drugs in resistant schizophrenia. *Schizophr. Bull.* 32, 715–723. doi: 10.1093/schbul/sbj067
- Lian, J., De Santis, M., He, M., and Deng, C. (2015). Risperidone-induced weight gain and reduced locomotor activity in juvenile female rats: the role of histaminergic and NPY pathways. *Pharmacol. Res.* 95–96, 20–26. doi: 10.1016/j.phrs.2015.03.004
- Lian, J., Huang, X.-F., Pai, N., and Deng, C. (2014a). Betahistine ameliorates olanzapine-induced weight gain through modulation of histaminergic, NPY and AMPK pathways. *Psychoneuroendocrinology* 48, 77–86. doi: 10.1016/j.psyneuen.2014.06.010
- Lian, J., Huang, X.-F., Pai, N., and Deng, C. (2014b). Preventing olanzapine-induced weight gain using betahistine: a study in a rat model with chronic olanzapine treatment. *PLoS ONE* 9:e104160. doi: 10.1371/journal.pone.0104160
- Lian, J., Huang, X.-F., Pai, N., and Deng, C. (2016). Ameliorating antipsychotic-induced weight gain by betahistine: mechanisms and clinical implications. *Pharmacol. Res.* 106, 51–63. doi: 10.1016/j.phrs.2016.02.011
- Lin, S.-T., Chen, C.-C., Tsang, H.-Y., Lee, C.-S., Yang, P., Cheng, K.-D., et al. (2014). Association between antipsychotic use and risk of acute myocardial infarction: a nationwide case-crossover study. *Circulation* 130, 235–243. doi: 10.1161/CIRCULATIONAHA.114.008779
- Lipscombe, L. L., Austin, P. C., Alessi-Severini, S., Blackburn, D. F., Blais, L., Bresee, L., et al. (2014). Atypical antipsychotics and hyperglycemic emergencies: multicentre, retrospective cohort study of administrative data. *Schizophr. Res.* 154, 54–60. doi: 10.1016/j.schres.2014.01.043
- Liu, X., Lian, J., Hu, C.-H., and Deng, C. (2015). Betahistine co-treatment ameliorates dyslipidemia induced by chronic olanzapine treatment in rats through modulation of hepatic AMPK $\alpha$ -SREBP-1 and PPAR $\alpha$ -dependent pathways. *Pharmacol. Res.* 100, 36–46. doi: 10.1016/j.phrs.2015.07.023
- Liu, X., Wu, Z., Lian, J., Hu, C.-H., Huang, X.-F., and Deng, C. (2017). Time-dependent changes and potential mechanisms of glucose-lipid metabolic disorders associated with chronic clozapine or olanzapine treatment in rats. *Sci. Rep.* 7:2762. doi: 10.1038/s41598-017-02884-w

- Lord, C. C., Wyler, S. C., Wan, R., Castorena, C. M., Ahmed, N., Mathew, D., et al. (2017). The atypical antipsychotic olanzapine causes weight gain by targeting serotonin receptor 2C. *J. Clin. Invest.* 127, 3402–3406. doi: 10.1172/JCI93362
- Maayan, L., and Correll, C. U. (2011). Weight gain and metabolic risks associated with antipsychotic medications in children and adolescents. *J. Child Adolesc. Psychopharmacol.* 21, 517–535. doi: 10.1089/cap.2011.0015
- Maayan, L., Vakhrusheva, J., and Correll, C. U. (2010). Effectiveness of medications used to attenuate antipsychotic-related weight gain and metabolic abnormalities: a systematic review and meta-analysis. *Neuropsychopharmacology* 35, 1520–1530. doi: 10.1038/npp.2010.21
- Malin, S. K., Rynders, C. A., Weltman, J. Y., Barrett, E. J., and Weltman, A. (2016). Exercise intensity modulates glucose-stimulated insulin secretion when adjusted for adipose, liver and skeletal muscle insulin resistance. *PLoS ONE* 11:e0154063. doi: 10.1371/journal.pone.0154063
- Manu, P., Correll, C. U., van Winkel, R., Wampers, M., and De Hert, M. (2012). Prediabetes in patients treated with antipsychotic drugs. *J. Clin. Psychiatry* 73, 460–466. doi: 10.4088/JCP.10m06822
- Martins, P. J. F., Haas, M., and Obici, S. (2010). Central nervous system delivery of the antipsychotic olanzapine induces hepatic insulin resistance. *Diabetes* 59, 2418–2425. doi: 10.2337/db10-0449
- Matsui-Sakata, A., Ohtani, H., and Sawada, Y. (2005). Receptor occupancy-based analysis of the contributions of various receptors to antipsychotics-induced weight gain and diabetes mellitus. *Drug Metab. Pharmacokinet.* 20, 368–378. doi: 10.2133/dmpk.20.368
- Meghana, K., Sanjeev, G., and Ramesh, B. (2007). Curcumin prevents streptozotocin-induced islet damage by scavenging free radicals: a prophylactic and protective role. *Eur. J. Pharmacol.* 577, 183–191. doi: 10.1016/j.ejphar.2007.09.002
- Memarzia, J., Tracy, D., and Giaroli, G. (2014). The use of antipsychotics in preschoolers: a veto or a sensible last option? *J. Psychopharmacol.* 28, 303–319. doi: 10.1177/0269881113519506
- Minet-Ringuet, J., Even, P. C., Goubern, M., Tome, D., and de Beaurepaire, R. (2006). Long term treatment with olanzapine mixed with the food in male rats induces body fat deposition with no increase in body weight and no thermogenic alteration. *Appetite* 46, 254–262. doi: 10.1016/j.appet.2006.01.008
- Murashita, M., Inoue, T., Kusumi, I., Nakagawa, S., Itoh, K., Tanaka, T., et al. (2007). Glucose and lipid metabolism of long-term risperidone monotherapy in patients with schizophrenia. *Psychiatry Clin. Neurosci.* 61, 54–58. doi: 10.1111/j.1440-1819.2007.01610.x
- Nasrallah, H. (2008). Atypical antipsychotic-induced metabolic side effects: insights from receptor-binding profiles. *Mol. Psychiatry* 13, 27–35. doi: 10.1038/sj.mp.4002066
- Nieto-Vazquez, I., Fernández-Veledo, S., Krämer, D. K., Vila-Bedmar, R., García-Guerra, L., and Lorenzo, M. (2008). Insulin resistance associated to obesity: the link TNF- $\alpha$ . *Arch. Physiol. Biochem.* 114, 183–194. doi: 10.1080/13813450802181047
- Oriot, P., Feys, J. L., Mertens de Wilmars, S., Misson, A., Ayache, L., Fagnart, O., et al. (2008). Insulin sensitivity, adjusted beta-cell function and adiponectinaemia among lean drug-naïve schizophrenic patients treated with atypical antipsychotic drugs: a nine-month prospective study. *Diabetes Metab.* 34, 490–496. doi: 10.1016/j.diabet.2008.03.003
- Ozasa, R., Okada, T., Nadanaka, S., Nagamine, T., Zyryanova, A., Harding, H., et al. (2013). The antipsychotic olanzapine induces apoptosis in insulin-secreting pancreatic  $\beta$  cells by blocking PERK-mediated translational attenuation. *Cell Struct. Funct.* 38, 183–195. doi: 10.1247/csf.13012
- Pai, N., Deng, C., Vella, S.-L., Castle, D., and Huang, X.-F. (2012). Are there different neural mechanisms responsible for three stages of weight gain development in anti-psychotic therapy: temporally based hypothesis. *Asian J. Psychiatr.* 5, 315–318. doi: 10.1016/j.ajp.2012.05.005
- Palsamy, P., and Subramanian, S. (2010). Ameliorative potential of resveratrol on proinflammatory cytokines, hyperglycemia mediated oxidative stress, and pancreatic  $\beta$ -cell dysfunction in streptozotocin-nicotinamide-induced diabetic rats. *J. Cell. Physiol.* 224, 423–432. doi: 10.1002/jcp.22138
- Panariello, F., Perruolo, G., Cassese, A., Giacco, F., Botta, G., Barbagallo, A. P. M., et al. (2012). Clozapine impairs insulin action by up-regulating akt phosphorylation and Ped/Pea-15 protein abundance. *J. Cell. Physiol.* 227, 1485–1492. doi: 10.1002/jcp.22864
- Polcwiartek, C., Vang, T., Bruhn, C. H., Hashemi, N., Rosenzweig, M., and Nielsen, J. (2016). Diabetic ketoacidosis in patients exposed to antipsychotics: a systematic literature review and analysis of Danish adverse drug event reports. *Psychopharmacology* 233, 3663–3672. doi: 10.1007/s00213-016-4411-x
- Poyurovsky, M., Fuchs, C., Pashinian, A., Levi, A., Weizman, R., and Weizman, A. (2013). Reducing antipsychotic-induced weight gain in schizophrenia: a double-blind placebo-controlled study of reboxetine-betahistine combination. *Psychopharmacology* 226, 615–622. doi: 10.1007/s00213-012-2935-2
- Pramyothin, P., and Khaodhjar, L. (2015). Type 2 diabetes in children and adolescents on atypical antipsychotics. *Curr. Diab. Rep.* 15:53. doi: 10.1007/s11892-015-0623-4
- Reed, R. M., Graham, K. A., Rau, S. W., Gilmore, J. H., Perkins, D. O., Peebles, S. S., et al. (2014). A naturalistic comparison of the long-term metabolic adverse effects of clozapine versus other antipsychotics for patients with psychotic illnesses. *J. Clin. Psychopharmacol.* 34, 441–445. doi: 10.1097/JCP.0000000000000159
- Rondinone, C., Carvalho, E., Wesslau, C., and Smith, U. (1999). Impaired glucose transport and protein kinase B activation by insulin, but not okadaic acid, in adipocytes from subjects with Type II diabetes mellitus. *Diabetologia* 42, 819–825. doi: 10.1007/s001250051232
- Rubi, B., Ljubicic, S., Pournourmohammadi, S., Carobbio, S., Armanet, M., Bartley, C., et al. (2005). Dopamine D2-like receptors are expressed in pancreatic beta cells and mediate inhibition of insulin secretion. *J. Biol. Chem.* 280, 36824–36832. doi: 10.1074/jbc.M505560200
- Samara, M. T., Dold, M., Gianatsi, M., Nikolakopoulou, A., Helfer, B., Salanti, G., et al. (2016). Efficacy, acceptability, and tolerability of antipsychotics in treatment-resistant schizophrenia: a network meta-analysis. *JAMA Psychiatry* 73, 199–210. doi: 10.1001/jamapsychiatry.2015.2955
- Samaras, K., Correll, C. U., Mitchell, A. J., De Hert, M., and HeAL Collaborators Healthy Active Lives for People With Severe Mental Illness (2014). Diabetes risk potentially underestimated in youth and children receiving antipsychotics. *JAMA Psychiatry* 71, 209–210. doi: 10.1001/jamapsychiatry.2013.4030
- Seeman, M. V. (2009). Secondary effects of antipsychotics: women at greater risk than men. *Schizophr. Bull.* 35, 937–948. doi: 10.1093/schbul/sbn023
- Seino, S. (2012). Cell signalling in insulin secretion: the molecular targets of ATP, cAMP and sulfonylurea. *Diabetologia* 55, 2096–2108. doi: 10.1007/s00125-012-2562-9
- Shin, S. Y., Choi, B. H., Ko, J., Kim, S. H., Kim, Y. S., and Lee, Y. H. (2006). Clozapine, a neuroleptic agent, inhibits Akt by counteracting  $Ca^{2+}$  calmodulin in PTEN-negative U-87MG human glioblastoma cells. *Cell. Signal.* 18, 1876–1886. doi: 10.1016/j.cellsig.2006.02.004
- Shirakawa, J., and Kulkarni, R. N. (2016). Novel factors modulating human beta-cell proliferation. *Diabetes Obes. Metab.* 18(Suppl. 1), 71–77. doi: 10.1111/dom.12731
- Silvestre, J. S., and Prous, J. (2005). Research on adverse drug events. I. Muscarinic M3 receptor binding affinity could predict the risk of antipsychotics to induce type 2 diabetes. *Methods Find. Exp. Clin. Pharmacol.* 27, 289–304. doi: 10.1358/mf.2005.27.5.908643
- Skrede, S., Martins, L., Berge, R. K., Steen, V. M., López, M., and Fernø, J. (2014). Olanzapine depot formulation in rat: a step forward in modelling antipsychotic-induced metabolic adverse effects. *Int. J. Neuropsychopharmacol.* 17, 91–104. doi: 10.1017/S1461145713000862
- Sohn, M., Talbert, J., Blumenschein, K., and Moga, D. C. (2015). Atypical antipsychotic initiation and the risk of type II diabetes in children and adolescents. *Pharmacoepidemiol. Drug Saf.* 24, 583–591. doi: 10.1002/pds.3768
- Stahl, S. (2003). Describing an atypical antipsychotic: receptor binding and its role in pathophysiology. *Primary Care Companion J. Clin. Psychiatry* 5(Suppl 3), 9–13.
- Stefanidis, A., Verty, A. N. A., Allen, A. M., Owens, N. C., Cowley, M. A., and Oldfield, B. J. (2009). The role of thermogenesis in antipsychotic drug-induced weight gain. *Obesity* 17, 16–24. doi: 10.1038/oby.2008.468
- Steinhausen, H.-C. (2015). Recent international trends in psychotropic medication prescriptions for children and adolescents. *Eur. Child Adolesc. Psychiatry* 24, 635–640. doi: 10.1007/s00787-014-0631-y
- Stubbs, B., Vancampfort, D., De Hert, M., and Mitchell, A. J. (2015). The prevalence and predictors of type two diabetes mellitus in people with schizophrenia: a systematic review and comparative meta-analysis. *Acta Psychiatr. Scand.* 132, 144–157. doi: 10.1111/acps.12439

- Su, H.-C., Hung, L.-M., and Chen, J.-K. (2006). Resveratrol, a red wine antioxidant, possesses an insulin-like effect in streptozotocin-induced diabetic rats. *Am. J. Physiol. Endocrinol. Metab.* 290, E1339–E1346. doi: 10.1152/ajpendo.00487.2005
- Sugai, T., Suzuki, Y., Fukui, N., Ono, S., Watanabe, J., Tsuneyama, N., et al. (2012). Dysregulation of adipocytokines related to second-generation antipsychotics in normal fasting glucose patients with schizophrenia. *J. Clin. Psychopharmacol.* 32, 390–393. doi: 10.1097/JCP.0b013e3182524393
- Teff, K. L., Rickels, M. R., Grudziak, J., Fuller, C., Nguyen, H.-L., and Rickels, K. (2013). Antipsychotic-induced insulin resistance and postprandial hormonal dysregulation independent of weight gain or psychiatric disease. *Diabetes* 62, 3232–3240. doi: 10.2337/db13-0430
- Tremblay, F., Lavigne, C., Jacques, H., and Marette, A. (2001). Defective insulin-induced GLUT4 translocation in skeletal muscle of high fat-fed rats is associated with alterations in both Akt/protein kinase B and atypical protein kinase, C. ( $\zeta/\lambda$ ) activities. *Diabetes* 50, 1901–1910. doi: 10.2337/diabetes.50.8.1901
- Turnbaugh, P. J., Hamady, M., Yatsunenko, T., Cantarel, B. L., Duncan, A., Ley, R. E., et al. (2009). A core gut microbiome in obese and lean twins. *Nature* 457, 480–484. doi: 10.1038/nature07540
- Uysal, K. T., Wiesbrock, S. M., Marino, M. W., and Hotamisligil, G. S. (1997). Protection from obesity-induced insulin resistance in mice lacking TNF- $\alpha$  function. *Nature* 389, 610–614. doi: 10.1038/39335
- van der Zwaal, E. M., Janhunen, S. K., la Fleur, S. E., and Adan, R. A. H. (2014). Modelling olanzapine-induced weight gain in rats. *Int. J. Neuropsychopharmacol.* 17, 169–186. doi: 10.1017/S146114571300093X
- van der Zwaal, E. M., Merksteijn, M., Lam, Y. K., Brans, M. A. D., Luijendijk, M. C. M., Bok, L. I. H., et al. (2012). The acute effects of olanzapine on ghrelin secretion, CCK sensitivity, meal size, locomotor activity and body temperature. *Int. J. Obes.* 36, 254–261. doi: 10.1038/ijo.2011.97
- van Winkel, R., De Hert, M., Wampers, M., Van Eyck, D., Hanssens, L., Scheen, A., et al. (2008). Major changes in glucose metabolism, including new-onset diabetes, within 3 months after initiation of or switch to atypical antipsychotic medication in patients with schizophrenia and schizoaffective disorder. *J. Clin. Psychiatry* 69, 472–479. doi: 10.4088/JCP.v69n0320
- Verma, S., and Hussain, M. E. (2017). Obesity and diabetes: an update. *Diab. Metab. Syndrome. Clin. Res. Rev.* 11, 73–79. doi: 10.1016/j.dsx.2016.06.017
- Victoriano, M., de Beaupaire, R., Naour, N., Guerre-Millo, M., Quignard-Boulangé, A., Huneau, J.-F., et al. (2010). Olanzapine-induced accumulation of adipose tissue is associated with an inflammatory state. *Brain Res.* 1350, 167–175. doi: 10.1016/j.brainres.2010.05.060
- von Wilmsdorff, M., Bouvier, M. L., Henning, U., Schmitt, A., Schneider-Axmann, T., and Gaebel, W. (2013). The Sex-dependent impact of chronic clozapine and haloperidol treatment on characteristics of the metabolic syndrome in a rat model. *Pharmacopsychiatry* 46, 1–9. doi: 10.1055/s-0032-1321907
- Wampers, M., Hanssens, L., van Winkel, R., Heald, A., Collette, J., Peuskens, J., et al. (2012). Differential effects of olanzapine and risperidone on plasma adiponectin levels over time: results from a 3-month prospective open-label study. *Eur. Neuropsychopharmacol.* 22, 17–26. doi: 10.1016/j.euroneuro.2011.03.010
- Weir, G. C., and Bonner-Weir, S. (2004). Five stages of evolving beta-cell dysfunction during progression to diabetes. *Diabetes* 53, S16–S21. doi: 10.2337/diabetes.53.suppl\_3.S16
- Weston-Green, K., Huang, X.-F., and Deng, C. (2010). Sensitivity of the female rat to olanzapine-induced weight gain—Far from the clinic? *Schizophr. Res.* 116, 299–300. doi: 10.1016/j.schres.2009.09.034
- Weston-Green, K., Huang, X.-F., and Deng, C. (2011). Olanzapine treatment and metabolic dysfunction: a dose response study in female Sprague Dawley rats. *Behav. Brain Res.* 217, 337–346. doi: 10.1016/j.bbr.2010.10.039
- Weston-Green, K., Huang, X.-F., and Deng, C. (2012a). Alterations to melanocortinergic, GABAergic and cannabinoid neurotransmission associated with olanzapine-induced weight gain. *PLoS ONE* 7:e33548. doi: 10.1371/journal.pone.0033548
- Weston-Green, K., Huang, X.-F., and Deng, C. (2013). Second generation antipsychotic-induced type 2 diabetes: a role for the muscarinic M3 receptor. *CNS Drugs* 27, 1069–1080. doi: 10.1007/s40263-013-0115-5
- Weston-Green, K., Huang, X.-F., Lian, J., and Deng, C. (2012b). Effects of olanzapine on muscarinic M3 receptor binding density in the brain relates to weight gain, plasma insulin and metabolic hormone levels. *Eur. Neuropsychopharmacol.* 22, 364–373. doi: 10.1016/j.euroneuro.2011.09.003
- Wilding, J. (2007). The importance of free fatty acids in the development of Type 2 diabetes. *Diab. Med.* 24, 934–945. doi: 10.1111/j.1464-5491.2007.02186.x
- Wu, C. S., Tsai, Y. T., and Tsai, H. J. (2015). Antipsychotic drugs and the risk of ventricular arrhythmia and/or sudden cardiac death: a nation-wide case-control study. *J. Am. Heart Assoc.* 4:e001568. doi: 10.1161/JAHA.114.001568
- Wu, R.-R., Zhao, J.-P., Zhai, J.-G., Guo, X.-F., and Guo, W.-B. (2007). Sex difference in effects of typical and atypical antipsychotics on glucose-insulin homeostasis and lipid metabolism in first-episode schizophrenia. *J. Clin. Psychopharmacol.* 27, 374–379. doi: 10.1097/JCP.0b013e3180cac8db
- Xiao, X., Gaffar, I., Guo, P., Wiersch, J., Fischbach, S., Peirish, L., et al. (2014). M2 macrophages promote beta-cell proliferation by up-regulation of SMAD7. *Proc. Natl. Acad. Sci. U.S.A.* 111, E1211–E1220. doi: 10.1073/pnas.1321347111
- Zhang, J.-P., Gallego, J. A., Robinson, D. G., Malhotra, A. K., Kane, J. M., and Correll, C. U. (2013). Efficacy and safety of individual second-generation vs. first-generation antipsychotics in first-episode psychosis: a systematic review and meta-analysis. *Int. J. Neuropsychopharmacol.* 16, 1205–1218. doi: 10.1017/S1461145712001277
- Zhang, Q., Deng, C., and Huang, X.-F. (2013). The role of ghrelin signalling in second-generation antipsychotic-induced weight gain. *Psychoneuroendocrinology* 38, 2423–2438. doi: 10.1016/j.psyneuen.2013.07.010
- Zhang, Q., He, M., Deng, C., Wang, H., Lian, J., and Huang, X.-F. (2014a). Hypothalamic ghrelin signalling mediates olanzapine-induced hyperphagia and weight gain in female rats. *Int. J. Neuropsychopharmacol.* 17, 807–818. doi: 10.1017/S1461145713001697
- Zhang, Q., Lian, J., He, M., Deng, C., Wang, H., and Huang, X.-F. (2014b). Olanzapine reduced brown adipose tissue thermogenesis and locomotor activity in female rats. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 51, 172–180. doi: 10.1016/j.pnpbp.2014.02.003
- Zipursky, R. B., Gu, H., Green, A. I., Perkins, D. O., Tohen, M. F., McEvoy, J. P., et al. (2005). Course and predictors of weight gain in people with first-episode psychosis treated with olanzapine or haloperidol. *Br. J. Psychiatry* 187, 537–543. doi: 10.1192/bjp.187.6.537

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer IG-G and handling Editor declared their shared affiliation.

Copyright © 2017 Chen, Huang, Shao, Chen and Deng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.