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Regional contribution of vascular dysfunction in white matter dementia: clinical and neuropathological insights

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The maintenance of adequate blood supply and vascular integrity is fundamental to ensure cerebral function. A wide range of studies report vascular dysfunction in white matter dementias, a group of cerebral disorders characterized by substantial white matter damage in the brain leading to cognitive impairment. Despite recent advances in imaging, the contribution of vascular-specific regional alterations in white matter dementia has been not extensively reviewed. First, we present an overview of the main components of the vascular system involved in the maintenance of brain function, modulation of cerebral blood flow and integrity of the blood-brain barrier in the healthy brain and during aging. Second, we review the regional contribution of cerebral blood flow and bloodbrain barrier disturbances in the pathogenesis of three distinct conditions: the archetypal white matter predominant neurocognitive dementia that is vascular dementia, a neuroinflammatory predominant disease (multiple sclerosis) and a neurodegenerative predominant disease (Alzheimer's). Finally, we then examine the shared landscape of vascular dysfunction in white matter dementia. By emphasizing the involvement of vascular dysfunction in the white matter, we put forward a hypothetical map of vascular dysfunction during disease-specific progression to guide future research aimed to improve diagnostics and facilitate the development of tailored therapies.

KEYWORDS

white matter dementia, vascular dementia, multiple sclerosis, Alzheimer's disease—AD, vascular dysfunction

1. Introduction

The term white matter (WM) dementia refers to a clinical syndrome characterized by dysexecutive symptoms, such as slowed processing speed and impaired sustained attention, due to multifocal or diffuse WM damage (1). Despite their high prevalence across several conditions, WM involvement in dementia has been historically overshadowed by a focus on gray matter (GM) pathology that typically predominates in cortical dementia syndromes, such as Alzheimer's disease (AD). However, recent advances in neuroimaging have shed light on the intimate relationship between WM and cortical gray matter (CGM) and deep gray matter (DGM) regions, and how they govern vital functions, such as emotion and memory. The causes of WM dementia remain elusive, although vascular dysfunction has emerged as a key early pathophysiological contributor (2–14). Indeed, the maintenance of adequate blood supply and

vascular integrity is fundamental to ensure brain function in health. In WM dementia, growing evidence implicates vascular alterations, such as aberrant angiogenesis (7) and blood-brain barrier (BBB) impairment, and their impact on cerebral blood flow (CBF), as important culprits. Altogether, these abnormalities impact homeostasis (8, 9), brain metabolism (10, 11), and glial activation (12, 13), which contribute to neurodegeneration and the dementia syndromes that ensue. Although it is known that various neurological diseases are influenced by specific regional vascular deficits within the brain (15, 16), this angle has been poorly explored in the spectrum of WM dementia. Thus, a better understanding of the relationships, connections and differences between WM and GM vasculature may help to decipher the causes and consequences of WM dementia and are the focus of the current review.

After presenting the main components of the vascular system involved in brain maintenance in health and normal aging, we review the regional contribution of vascular changes to WM pathology (see anatomical considerations in Table 1) in the pathogenesis of three distinct conditions: the archetypal white matter predominant neurocognitive dementia that is vascular dementia, a neuroinflammatory predominant disease (multiple sclerosis, MS) and a neurodegenerative predominant disease (AD). Finally, we consider factors that contribute to vascular changes in vascular dementia, MS and AD. In so doing, we hope to highlight the importance of WM pathology in the pathogenesis of the spectrum of dementia types to guide future research.

2. The vascular system in normal brain

Blood vessels form a vital infrastructure for the supply of metabolites, oxygen and nutrients throughout the brain. The tubular structure of vessels is formed by vascular endothelial cells (ECs), surrounded by pericytes, astrocytes end-feet and extracellular matrix (ECM, forming the basal lamina), supplemented by direct interaction with neuron processes and oligodendrocytes. In that context, the term "blood-brain barrier" (BBB) encompasses a range of unique properties of brain blood vessels. The BBB tightly regulates cells, molecules and ions trafficking in and out of the parenchyma, maintaining adequate CBF and vessel permeability to ensure neuronal activity and protection against pathogens. The BBB is not a single entity: it is composed of ECs interacting with a network of brain cells (as mentioned above), that orchestrate vascular function with the activity and metabolic needs of the surrounding brain areas, termed the neurovascular unit (NVU). In this section, we briefly describe the function of these key anatomic structures in healthy brain (Figure 1) and their alteration in normal aging, commenting on the differences observed in WM and GM areas.

2.1. Vascular function in healthy brain and normal aging

2.1.1. Cerebral blood flow

By convention, CBF is defined as the volume of blood which flows at a rate of delivery through a defined quantity of brain tissue during a specific period of time (17). The brain needs an anatomically disproportionate oxygen supply, representing more than 20% of the total oxygen in the body while accounting for only 2% of the total body weight (18). Thus, CBF must modulate brain perfusion in order to supply oxygen and energy substrates essential for baseline neurological function. When neuronal activation occurs, an increase in oxygen and energy demand is followed by an increase in CBF via arterial vasodilation to supply essential metabolites, wherein glucose and its surrogates (the primary fuel source for the brain) enter the brain to complete the aerobic glycolysis (19).

With aging, cerebral oxygenation is commonly reduced. However, it is thought that global CBF remains relatively preserved (20). This

TABLE 1 Neuroanatomical considerations.

White matter (WM)	WM consists of subcortical tissues, containing myelinated axons which connect neurons and glial cells. It represents almost half of the brain's volume. The corpus callosum is the largest WM structure in the brain that connects the left and right hemisphere. WM is an essential component connecting areas of gray matter (GM) throughout the CNS and coordinates their communications.
Cortical gray matter (CGM)	The CGM (or cerebral cortex) is the outermost tissue of the brain lying on the top of the cerebrum. It is made up of folded GM defined as gyri (top areas) and sulci (deep areas). It covers subcortical WM and is divided into four lobes: frontal, parietal (where precuneus is the portion of the superior parietal lobe), temporal and occipital . The cingulate cortex located in the medial aspect of the cerebral cortex wraps the corpus callosum. The cerebral cortex contains a highly complex network of glial cells, neurons, axons and extracellular matrix proteins, and is involved in numerous functions such as emotion, memory, learning and language.
Deep gray matter (DGM)	The DGM consists of the thalamus, hippocampus, nucleus accumbens and basal ganglia. It is made up of neurons from which originate deep nerve fibers. The hippocampus is a simplified cortical structure located in the medial region of the temporal lobe, and consists of the dentate gyrus surrounded by corpus-ammonis and the subiculum. It has direct connections with the entorhinal cortex , and the amygdala. The hippocampus plays a major role in learning and memory. The thalamus is located near the center of the brain, including the hypothalamus , the subthalamus and the epithalamus. It has connections with structures of the limbic system (hippocampus, amygdala, cingulate cortex) and the cerebral cortex. The thalamus is a crucial gateway which relay the information in the CNS, playing a role in sense processing, spatial learning and memory. The basal ganglia is located deep within the cerebral hemispheres. It consists of a group of subcortical nuclei composed of the corpus striatum, the globus pallidus and the substantia nigra. These structures are connected to various brain areas, receiving many inputs from the cerebral cortex and the thalamus. Due to its large connectivity, the basal ganglia is involved in motor control, as well as rule-based learning, working memory and cognitive functions. The amygdala is located in the medial temporal lobe in front of the hippocampus. Its main connections include the hippocampus, the basal ganglia through the striatum and the cerebral cortex. The amygdala is primarily responsible of the processing and memorizing of emotional reactions.



structures apposed to the membrane of these endothelial cells, forming the basal lamina. At the interface of the peripheral circulation and brain parenchyma, the NVU is composed of various cells, such as glial cells (astrocytes end-feet in close association with vessel wall, microglia, oligodendrocytes), neurons, and perivascular pericytes, which act together to regulate CBF and clearance pathways. Importantly, blood supply matches local neuronal demand by a mechanism called neurovascular coupling, wherein activated neurons release specific effectors which activate astrocytes and pericytes, inducing the release of vasoactive mediators and subsequently regulating local CBF. In addition to their critical role in brain homeostasis, these structures form a physical barrier protecting the brain against immune infiltrates (macrophages, lymphocytes) and infiltrating pathogens, restraining the entry of harmful molecules (toxins), organisms (bacteria), and infectious agents (viruses). During aging, each component of the cerebral vasculature is impacted.

stability is ensured by homeostatic processes which modulate CBF in response to cerebral perfusion pressure changes and various external and internal physiological factors, such as vasoactive stimuli and neuronal activity changes (21–27).

2.1.2. The blood-brain barrier

2.1.2.1. Endothelial cell as the wall of the vessels

ECs form an efficient physiological wall ensuring vessel integrity. ECs are adjoined to each other by tights junctions (TJs), which modulate the paracellular transport of molecules through the BBB, and are composed of a large panel of transmembrane proteins involved in permeability regulation (claudin family) and its maintenance (occludin and Zo-1) (28). In addition, ECs modulate transcellular transport thanks to a wide range of receptor and carrier-mediated transporters, regulating the passage of ions, nutrients, energy substrates and cells (29-33).

With aging, senescence of EC combined with TJ alterations may occur, increasing BBB permeability and neurovascular uncoupling, subsequently impacting the inflammatory milieu and neuronal dysfunction (34).

2.1.2.2. The neurovascular unit

The NVU is an anatomical and functional unit encompassing glial cells (astrocytes end-feet, microglia and oligodendrocytes), mural cells (smooth muscle cells and pericytes) and neuronal processes. It plays a critical role in modulating transport of vasoactive agents and neuromodulators, helping to maintain the cerebral microenvironment and BBB integrity (35). Importantly, the NVU triggers vasodilation/vasoconstriction to match the local energy demand given

Element of vascular tree	Vascular cells	Perivascular cells
Artery	Smooth muscle: Socs3, Atf3, Id2, Csrnp1 Endothelium: Gkn3, Hey1,	Fibroblasts, mast cells, perivascular macrophages, extrinsic
	Vcam, Vwf	nerves
Arteriole (pial)	Smooth muscle: Slc26a2, Ttr, Sh3bgrl2, Tmem255b Endothelium:	Fibroblasts, mast cells, pial cells, perivascular macrophages
Arteriole (pre-capillary)	Gkn3, Hey1	Astrocytes, microglia, intrinsic nerves
Capillary	Pericytes Pdgfrb, Cspg4, Vtn Endothelium Mfsd2a, Rgcc	Astrocytes, microglia, intrinsic nerves
Venule	Smooth muscle: Sebox, Tnxb, Ggt1, Grm3 Endothelium: Slc38a5,	Astrocytes, microglia, intrinsic nerves
	Lcn2	
Vein	Smooth muscle: Sebox, Tnxb, Ggt1, Grm3 Endothelium: Slc38a5,	Fibroblasts, mast cells, perivascular macrophages, extrinsic
	Lcn2, Car4, Vwf, Vcam	nerves

TABLE 2 Heterogeneity of vascular tree [based on Schaeffer and ladecola (39)].

tight interactions between neuronal processes and vessel-related cells (36). To this end, each type of cell forming the NVU modulates a wide range of dynamic processes. Astrocytes and pericytes structurally support the BBB via their end feet and processes, respectively. Astrocytes cover 90% of the micro-vessel surface (5) and regulate homeostasis processes and neuronal activity, while pericytes have functions in CBF regulation, angiogenesis, protein clearance, and neuroinflammatory mechanisms. Microglia, which also act in concert with astrocytes, are the macrophages of the brain involved in the inflammatory response. Microglial cells interact with neurons to maintain their activity and play an important role in synaptic plasticity. Oligodendrocytes contribute to brain homeostasis by maintaining and producing myelin in both GM and WM.

The structures that form the NVU are intricately linked. With aging, each component of the NVU undergoes substantial changes, resulting in increased BBB permeability and penetration of toxic factors into the brain (37, 38). Dysregulated mechanisms include disruption of basal lamina from pericyte degeneration, release of pro-inflammatory factors from glial cells, reduced myelin repair from oligodendrocytes and reduced clearance capacity from neurons, to name a few. Each of these alterations can contribute to WM pathology seen in dementia syndromes.

2.2. Vascular heterogeneity across the normal brain and aging

2.2.1. White matter has a poorer cerebrovascular reserve compared with gray matter

The cerebral vasculature is a continuum from arteries to veins which differ across the brain (Table 2) (39). GM regions are characterized by a greater number of large vessels and capillaries compared with WM (40), particularly in DGM regions, such as the hippocampus (41, 42). This has been well described in a cohort of 42 healthy young adults wherein vascular density for both arteries and veins was found to be lower in WM compared with GM, where it correlated with cortical thickness (43). Furthermore, GM and WM structures are morphologically heterogeneous, with energy supply being proportionally linked to the extent of connectivity between brain regions. The cerebrovascular reserve in GM is related to a rich neuronal, synaptic, and glia network demonstrating concomitant higher energy use, oxygen demand, and perfusion compared to WM (44). Therefore, differences in WM and GM vasculature, including changes to the structure and function of the NVU and BBB, and how they impact CBF should be kept in mind when considering studies on healthy aging and neurodegenerative disorders. Importantly, given the often-striking regional patterns of neurodegeneration seen in dementia syndromes, regional differences in these vascular structures and CBF warrant study.

2.2.2. Cerebral blood flow is physiologically reduced in white matter

Recent advances in neuroimaging support that CBF should not be considered as a global constant. Normal CBF is considered to be around 50 mL/100 g.min⁻¹ in the healthy brain, and it is well established that CBF in WM is less compared to GM (~20 and 80 mL/100 g.min⁻¹, respectively), reflecting the poorer cerebrovascular reserve described above (17, 45). This is a consistent observation across the brain (45), as well as in the cerebellum (46). It is also well established that stress and aging lead to reductions in CBF that are more severe in WM compared to GM (47). Interestingly, WM vulnerability is linked to the degree of network integration and connectivity, as demonstrated by a study looking at CBF and glucose metabolism (48). These findings support that CBF is influenced not only by intrinsic morphological differences in WM and GM structures, but also by loss of connectivity between them that can occur with aging.

2.2.3. The blood-brain barrier is more vulnerable in periventricular WM with aging

Despite limited knowledge about region-specific BBB alterations in the healthy brain, a range of studies demonstrate vulnerability of WM and DGM to BBB and NVU damage with aging.

It is intuitive to postulate that molecular trafficking across the BBB is influenced by the distribution of the cerebrovasculature. It is well known that capillaries are more vulnerable (as the basal lamina is incomplete and pericyte coverage is lacking) in DGM (hippocampus, hypothalamus) and the surrounding WM, compared to CGM (49, 50). With aging, lower expression of TJ proteins has been reported in the periventricular WM (corpus callosum) compared with CGM, and morphological changes in ECs associate with BBB breakdown in WM but not GM following systemic inflammation (51). Prominent loss of pericytes has also been specifically observed in WM of pericyte-deficient mouse models, associating with accumulation of fibrinogen, a surrogate of BBB dysfunction (52). As fibrinogen is highly toxic for oligodendrocytes

(53), fibrinogen accumulation may also play a major role in oligodendrocyte loss leading to WM changes and cognitive decline with aging (54). Furthermore, astrocyte morphology and distribution are highly heterogenous across the central nervous system (CNS), with elongated astrocytes with few processes in WM tracts (55), in comparison with highly branched protoplasmic astrocytes in GM (56). In mice, an age-dependent increase in GFAP astrocyte subpopulations, specifically in CGM and DGM structures (frontal, temporal cortices and hippocampus), has been shown (57). These observations highlight the role of aging on vascular dysfunction in WM and GM, which may set the stage for the evolution of cognitive impairment in susceptible individuals.

3. Vascular dysfunction in vascular dementia

The entity of vascular dementia and the pathologies encompassed by the term have posed challenges over the decades (58). However, vascular dementia is likely the second most common cause of dementia (the lack of certainty of this likely reflects historical disagreement over the definition of the condition), with mean autopsy prevalence rates of 8-15% in the West and higher in parts of Asia (59). The interactions between the vasculature, WM alteration and dementia were recognized as early as the nineteenth century, where arteriosclerosis, WM atrophy and preservation of CGM were associated with a slowly progressive dementia and gait disturbance (60). More recently, vascular cognitive impairment (VCI) has been defined as "a syndrome with evidence of clinical stroke or subclinical vascular brain injury and cognitive impairment affecting at least one cognitive domain" (61), and includes the entities of post-stroke dementia, multi-infarct dementia (due to multiple cortical infarcts) and subcortical ischemic vascular dementia (SIVD) (58). In particular, the dementia associated with small vessel disease (SVD) pathology of the WM (SIVD) is characterized by a frontal dysexecutive syndrome, slowed processing speed and sustained attentional deficits (62). Another important entity linking vascular pathology and dementia is cerebral amyloid angiopathy (CAA).

In the next section, we explore the regional specific perfusion alterations and neuropathology of SVD (Figure 2), including insights from the rare monogenic subcortical arteriopathy syndromes associated with dementia, such as Cerebral Autosomal Dominant Arteriopathy with Subcortical Infarcts and Leukoencephalopathy (CADASIL).

3.1. Hypoperfusion in vascular dementia

By definition, chronic hypoperfusion is central to VCI, wherein a 20–35% reduction in global CBF has been observed (63). This is supported by metanalyses which demonstrate a relationship between severity of WM hyperintensities (WMH, the cardinal manifestation of SIVD) and reduced CBF (64, 65). In aged patients with dementia, regions with WMH had lower CBF than normal appearing white matter (NAWM) and periventricular WM had lower CBF than other WM areas (66). It was also shown that low baseline CBF in normal appearing periventricular WM can predict the progression of WMH in aged patients.

Interestingly, changes to CBF extend beyond WM, and also involve CGM. Regional differences in CBF and their relationship to WMH were studied in a cohort of MCI patients wherein VCI is commonly observed. A relationship between reduced CBF and WMH was seen in parietal, occipital and temporal CGM but not frontal CGM; these findings were not accompanied by overlying cortical atrophy implying an early role for vascular dysfunction (67). In another study, GM atrophy was observed in SIVD, in particular in prefrontal cortex, the middle and superior temporal gyri and the thalamus (68), although this study did not look at CBF. The relationship between CBF alterations and patterns of atrophy therefore warrant further study.

Taken together, these studies highlight that regional CBF changes, even in GM, are associated with WM lesions seen in SIVD, detectable even in early disease phases. What is known about the pathological substrate of these CBF changes in vascular dementia will be discussed below.

3.2. Neuropathology of vascular dysfunction in vascular dementia

Pathologically, SIVD is characterized by hyalinization of the vessel walls, fibrinoid necrosis, widening of the perivascular spaces, perivascular demyelination, tissue ischemia, and astrogliosis (69, 70). These features are influenced by hypertension, a major risk factor for SIVD. As WM is inherently vulnerable to cerebrovascular insults (71, 72), it is not surprising that prior studies have shown that the extent of subcortical WM vascular disease correlates with the presence of dementia (69, 73). Of important note, these changes in the small vessels do not occur in isolation, and have been associated with atherosclerotic disease of the larger cerebral vessels (74).

3.2.1. The blood-brain-barrier in vascular dementia

BBB permeability has been described in SVD but the results have been conflicting. Although one study reported that surrogates of BBB permeability, such as fibrinogen and IgG, in DGM and subcortical WM do not correlate with pathological measures of SVD, nor with MRI measures of leukoaraiosis in life (75), many studies have found evidence of BBB breakdown. In fact, BBB breakdown has been described in the perivascular space of DGM in SVD cases (76). A spectrum of animal and post-mortem studies support BBB dysfunction in SVD, wherein hypertension damages ECs of small cerebral vessels (77), and associates with increased fibrinogen and IgG deposition (DGM and WM) in the brain, particularly in astrocytes located in periventricular WM (78).

In addition to accumulation of toxic factors, ECs dysfunction is evidenced by reduced TJ protein expression in both rat models of SVD and deep WM of people with early stage SVD (79). Reduced endothelial nitric oxide synthase (eNOS), which results in vasoconstriction and ultimately tissue ischemia (80), is another marker of EC dysfunction that has been described in SVD. Decreased eNOS exacerbates BBB leakage, WM damage and dementia in animal models (81). Dysfunctional ECs can also secrete ECM-related proteins which disrupt TJs in DGM of animal models (82). As a consequence, BBB breakdown can lead to changes in the NVU promoting tissue



damage, in both WM and GM, mirroring CBF changes detected in imaging studies.

3.2.1.1. The neurovascular unit in vascular dementia

Although vascular pathology is thought to be the underlying mechanism for leukoaraiosis, glial dysregulation is also of interest (6). Components of the NVU may be compromised early in SIVD, including astrocytes, microglia and oligodendrocytes.

Increased reactive astrocytes are seen in WM lesions in aged patients, and are positive for fibrinogen accumulation, indicating BBB breakdown (78). Of note, the astrocytic network pattern found in SIVD is similar to that observed in MS WM demyelinated lesions. In mouse models of cerebral hypoperfusion, blockade of astrogliosis reduces demyelination and cognitive impairment (84).

Altering microglial activity can influence WM pathology. For example, microglial polarization to a less inflammatory phenotype (via fingolimod in mice) leads to preservation of both WM integrity and cognitive function (85, 86). Similarly, pioglitazone, a peroxisome proliferator-activated receptor γ agonist, ameliorates WM lesion scores and performance on the Morris water maze in stroke-prone hypertensive rats; these findings were associated with reduced proliferation of astrocytes and microglia with reduction of pro-inflammatory cytokines (87).

In rat models and human tissue of early SVD, increased numbers of oligodendrocyte progenitor cells (OPCs) have been detected and

attributed to reduced maturation into oligodendrocytes (78); interestingly, drug treatment targeting EC dysfunction was found to promote OPC differentiation into oligodendrocytes (79). This is further supported by the observation that fibrinogen extravasation across a disrupted BBB and ischemia-induced oxidative stress impede OPC differentiation and maturation (6). The conflation of these findings highlights the impact of the NVU on glial function and WM integrity relevant to cognitive function.

3.2.1.2. Insights from hereditary microangiopathies

Several genetic diseases of the small cerebral vessels are associated with dementia. These include CADASIL (due to mutations in the NOTCH3 gene) and CARASIL (cerebral autosomal recessive arteriopathy with subcortical infarcts and leukoencephalopathy, due to mutations in the HTRA1 gene) (74). These diseases may inform pathological processes involved in the much more common sporadic SIVD. NOTCH3 encodes for a transmembrane receptor expressed in pericytes and vessel smooth muscle cells and is important for their integrity. There are many identified mutations within the gene and the disease often arises from deposition of abnormal NOTCH3 protein within the vessel walls, resulting in changes similar to that seen in SIVD (88). HTRA1 (high temperature requirement a serine peptidase), mutated in CARASIL, causes loss of repression of TGF-beta, a growth factor, with resultant vascular fibrosis. A similar pathology is thus seen in CARASIL and these patients develop dementia earlier than in CADASIL, as the changes are more widespread (89). Therefore, both these conditions act as informative models of SIVD, in that they result in arteriole wall hyperplasia and resultant reduced blood flow and ischemia (89). Indeed, many of the imaging studies on CBF in cerebrovascular disease were done on cohorts of patients with CADASIL (90).

4. Vascular dysfunction in multiple sclerosis

MS is an inflammatory disorder of the CNS estimated to affect more than 2.8 million people worldwide (91). MS is characterized by inflammation, demyelination, and neurodegeneration that results in disability and reduction of quality of life (92). Changes to cognition and emotion are common features, an observation which dates back to Jean-Martin Charcot's classic descriptions (93). More recent studies suggest that cognitive impairment is present in 34% of those with clinically isolated syndrome (CIS), 50% with relapsing-remitting MS (RRMS), and 80-90% of those with progressive MS (94). Interestingly, the cognitive impairment seen in MS has several similarities to that seen in vascular dementia, namely dysexecutive symptoms, slowed processing speed, and impaired sustained attention (95). Further, it is increasingly recognized that GM alterations may be associated with vascular disturbances in MS (Figure 3) and likely play an important role in cognitive decline. Indeed, GM demyelination is frequent and extensive in MS, both in CGM and DGM structures relevant to cognitive function (e.g., thalamus, hippocampus, cerebellum) (96-98). Vascular dysfunction is also increasingly recognized as an early feature of MS pathogenesis, supported by the pathognomonic vesselassociated pathology characteristic of the disease (99).

We herein provide a summary of the main characteristics of vascular dysfunction in MS, including its regional variability.

4.1. Perfusion alterations in multiple sclerosis

The clinical pattern of MS dementia resembles that of vascular dementia with similarities in CBF hypoperfusion patterns being seen. While the formation of a new demyelinating plaque is accompanied by an increase in CBF, the MS brain gradually displays progressive reduction in CBF which correlates with the degree of atrophy and clinical disability.

Hyperperfusion has been shown to be one of the earliest events in plaque formation detectable on MRI, being present even before emergence of gadolinium-contrast enhancement and changes in diffusion (100). This relative hyperperfusion (18% increase from baseline) in active WM lesions has been attributed to vasodilation in response to an intense inflammatory response (101). This notion is supported by studies evaluating the experimental autoimmune encephalomyelitis (EAE) mouse model of MS, wherein induction of severe inflammation results in severe hypoxia (102), and increased perfusion. consequent Interestingly, oxygen administration not only attenuated hypoxia but also clinical deficits (103).

Outside the acute inflammatory phase of plaque formation, the available MRI literature consistently reports hypoperfusion across brain regions in all disease stages of MS, wherein a 20% reduction in global CBF has been reported in people with MS compared with normal aging (104). In fact, normal appearing periventricular WM and DGM and CGM areas are particularly affected (105-108) with CBF reduction of up to 50% being reported. The reduced global CBF has clinical relevance as it correlates with measures of brain atrophy, and cognitive and motoric disability (109-111), independent of vascular risk factors (109). In early disease stages, CBF hypoperfusion has been shown to preferentially occur in juxtacortical and periventricular WM areas and associate with severe demyelination with relatively sparing of CBF alterations in CGM and DGM areas. It has been suggested that secondary hypoperfusion in GM areas due to underlying WM injury may contribute to disease progression in later disease stages (112, 113) but this warrants further study. Regardless, the observation that CBF hypoperfusion occurs at the earliest disease stages in normal appearing WM areas further supports the notion that CBF alterations are not merely a downstream consequence of MS pathology but rather may be an important upstream contributor of it (112).

4.2. Neuropathology of vascular dysfunction in multiple sclerosis

Vessel-associated inflammatory demyelination is a hallmark of MS pathology, especially in WM regions. In recent years, a growing body of evidence suggests that vascular changes occur outside of WM lesions and affect the vascular tree beyond the traditionally blamed venule. The observation that vascular risk factors, which are known to impact the arterial system, associate with more severe pathology and disease progression supports this claim (114). A recent post-mortem study showed that periarterial small vessel disease outside of lesions is increased and associates with inflammatory disease activity in progressive MS (99).



demonstrated with immunohistochemistry in the primary motor cortex (frontal cortex), in non-demyelinated gray matter area (figure adapted from before renumber no (138) with permission; scale bas 500µm). (**D**) Main features of blood–brain barrier and neurovascular unit alterations in multiple sclerosis compared with healthy brain (created with BioRender.com; GM, gray matter; WM, white matter; EC, endothelial cells; MS, multiple sclerosis; NVU, neurovascular unit).

4.2.1. Pathology of the BBB and NVU in demyelinated plaques

In MS, WM lesion burden associates with the development of cognitive deficits thereby pointing to a role for vascular dysfunction in cognitive decline (115–117). MRI studies have highlighted the importance of BBB alterations in the pathogenesis of WM pathology as evidenced by the presence of gadolinium enhancement in acute lesions. BBB breakdown occurs at the earliest disease stages and associates with extensive perivascular immune cell infiltration in WM lesions (118–125). Dysfunctional ECs have been shown to propagate the inflammatory response in WM lesions by upregulating membrane receptors (e.g., TLR4) (126, 127) and inducing release of proinflammatory cytokines (e.g., IFN- γ and TNF- α) by glial cells;

these changes, in turn degrade TJs and surrounding ECM to form a vicious circle of BBB breakdown (118, 128, 129). BBB integrity is further compromised in active WM lesions by the detachment of astrocyte endfeet from the basal lamina (130). Further support for a role for astrocytes in WM lesion pathogenesis is derived from the finding of increased astrocyte-derived vasoconstrictive peptide endothelin-1 (ET-1, related to reduced CBF) in CSF of MS patients (131). The observations support early changes to BBB integrity in the pathogenesis of WM lesions.

Surprisingly, little is known about vascular dysfunction in GM lesions, despite extensive involvement of GM pathology in MS. In contrast to WM lesions, some studies report no alterations in ECs and their TJs, as well as no differences in infiltrating lymphocyte numbers

in cerebral cortical lesions compared to NAGM (132). These findings are challenged by recent neuroimaging and post-mortem studies that show loss of BBB integrity in CGM and WM lesions alike and extensive inflammatory activity in hippocampal and spinal cord GM lesions, respectively (133–137). BBB disruption in both GM and WM areas are likely substrates, which highlights the need to consider the role of vascular dysfunction in these areas and how they contribute to cognitive impairment in MS.

4.2.2. Pathology of the BBB and NVU beyond demyelination

The classical view of BBB disruption as only a feature of demyelinated lesions in MS is challenged by a body of evidence that implicates BBB breakdown in NAWM and NAGM. Extensive fibrinogen accumulation, a surrogate of BBB disruption, has been detected in NAGM areas in progressive MS; the fact that the extent of fibrinogen deposition correlates with neuronal loss, the substrate of irreversible disability, adds clinical relevance to this finding (138, 139). The mechanism by which fibrinogen, a large molecular weight protein, traverses the BBB has attracted studies evaluating the role of various constituents of the BBB in fibrinogen egress from the vasculature. A mouse model of pericyte-deficiency showing accumulation of parenchymal fibrin supports involvement of pericyte dysfunction in this process (138, 140). In support of this possibility, EAE rat models demonstrate pericyte loss in CGM capillaries with induction of PDGFRß expression associating with overexpression of TJs proteins (claudin) and reduction of clinical symptoms (141). Astrocyte activation and alterations of aquaporin-4 expression in their end feet have been also shown to contribute to vascular alterations even before the infiltration of immune cells (142, 143). The accumulation of fibrinogen in astrocyte cell bodies and processes in the glia limitans and perivascular areas of non-lesional areas further implicate a role of astrocyte function and fibrinogen accumulation (138, 139). Importantly, fibrinogen is known to initiate and propagate neurotoxic microglial activation in EAE mouse models leading to the secretion of pro-inflammatory cytokines (e.g., TNF- α , IL-1 β) that contributes to neurodegeneration (144, 145). This is supported by recent neuropathological findings in NAGM cerebral cortical areas suggesting that neurotoxic factors, opposing putative microglialassociated protective factors, are secreted by fibrinogen-stimulated microglia in people with HLA-DRB1*15 genetic status, a major risk factor for MS, further highlighting a pathologically relevant consequence of BBB compromise in the disease (146). Future work evaluating the nature and extent of BBB and NVU changes in both WM and GM within and outside lesions throughout the MS brain will help disentangle their roles in disease pathogenesis.

Overall, these findings demonstrate that changes to the BBB and NVU are important features of MS pathology even beyond areas of demyelination in both GM and WM and contribute to neurodegenerative processes relevant to cognitive impairment in MS (99).

5. Alzheimer disease and vascular dysfunction

AD is a neurodegenerative syndrome characterized by early deficits in working and episodic memory with an increased incidence

and prevalence as the population ages. There are in excess of 55 million people living with dementia worldwide and 60–70% of these have AD (147). AD is characterized by brain atrophy, neuroaxonal loss and accumulation of amyloid- β and Tau, mainly in GM structures. However, recent studies also highlight significant WM changes in AD pathology. Regional vascular abnormalities are likely important contributors to both GM and WM pathology in AD (Figure 4) with some features shared with vascular dementia and MS.

5.1. Cerebral blood flow impairment in Alzheimer's disease

Compared to normal aging, reduction of 20-40% of the global CBF is common in AD (148). In particular, it is well-established that CGM and DGM hypoperfusion, mainly in the temporal lobe, are early features of the disease. In MCI and early stages of AD, a wide range of studies show reduced CBF in DGM (such as hippocampus, thalamus and basal ganglia) (149-154) and CGM (temporal cortex) (155, 156), which often associate with progressive cognitive decline and brain atrophy. Relative hyperperfusion has also been described in early AD, which has been attributed to a transient compensatory response to local inflammatory and neurodegenerative mechanisms (157-160). With AD progression, hypoperfusion in CGM areas (estimated to be about 20 mL/100 g.min⁻¹ by MRI) (161), such as frontal and parietal cortices, relates to hypometabolism, amvloid accumulation, and cognitive decline (162-165). The extent to which CBF alterations drive or are a consequence of AD pathology requires further study.

In addition to GM, changes to CBF in WM have been described, especially at more advanced disease stages when subcortical cognitive change is a more prevalent feature. In established AD, up to 50% reduction of CBF in temporal, parietal and occipital WM areas has been shown to be more severe than in their GM counterparts (up to 40% reduction) (166, 167). These CBF changes relate to WM hyperintensities (WMH) often observed in AD (25), which variably associate with cognitive impairment (168, 169), especially when situated in periventricular and juxtaventricular areas (170). Hypoperfusion in the corpus callosum has been described in late, but not prodromal, stages of AD highlighting an important temporal element to WM involvement in AD pathogenesis (159, 171, 172). In fact, such stage-dependent regional CBF alterations in WM in AD are hypothesized to disrupt WM integrity and network connectivity between GM regions, further exacerbating cognitive decline (173-175). Studies of the brain connectome will shed further light into the relative roles of CBF changes in WM and GM at various stages of AD-related cognitive decline.

5.2. Neuropathology of vascular dysfunction in Alzheimer's disease

To date, the unequivocal diagnosis of AD relies on post-mortem detection of amyloid and Tau in characteristic brain regions. However, amyloid and tau deposition do not fully account for the progressive cognitive decline seen in AD, implying that other factors, such as vascular pathology, are contributing to AD pathology. Reports of either unchanged, reduced or increased



and neurovascular unit alterations in Alzheimer's disease compared with healthy brain (created wi matter; EC, endothelial cells; TJ, tight junction; AD, Alzheimer's disease; NVU, neurovascular unit).

vascular densities in the AD brain complicate interpretation of vascular changes in AD pathogenesis, although differences in methods, AD severity and selected controls may explain these discrepant findings. However, there is increasing recognition that loss of connectivity between WM and GM regions and vascular dysfunction are contributors to cognitive decline and may augment other aspects of AD pathology. In that context, more is known in AD than any other dementia, wherein it is suggested that vascular abnormalities affect both WM and GM through the "two-hit vascular hypothesis" (176, 177). The first hit is damage to the microcirculation independent of changes to amyloid burden, while the second hit is loss of ECs and TJs supplemented by NVU alterations that lead to impaired $A\beta$ clearance and subsequent amyloid accumulation in the AD brain.

5.2.1. Evidence of white matter pathology in Alzheimer's disease

An increasing body of evidence suggests that WM changes play an important role in AD pathophysiology, including WM degeneration and demyelination secondary to reduced numbers and function of OPCs and oligodendrocytes, disrupted connectivity of WM tracts, and amyloid-induced inflammatory damage (178–182). The finding that *ApoE4* carriers show significant WM degeneration prior to emergence of GM atrophy and cognitive dysfunction further highlights a fundamental role of WM in AD disease pathogenesis (183). Advanced MR studies show altered vascular integrity in WM areas adding further support to a role vessel-related WM changes in AD (176, 184).

Another key aspect of vascular change linked to WM damage in AD is cerebral amyloid angiopathy (CAA). CAA is characterized by the accumulation of amyloid in the wall of the cerebral vessels and occurs in up to 90% of established AD cases (185). CAA is associated with prominent ischemic WM damage and atrophy (186, 187) secondary to extravasation of immune cells and toxic factors. The accumulation of fibrinogen in neurons and vessels in areas of A β deposition (188) and the formation of A β -fibrinogen insoluble clots in the parenchyma propagate deleterious inflammatory responses, vascular dysfunction, and CAA that contribute to WM change in AD (189, 190).

5.2.2. Pathology of the blood-brain barrier in Alzheimer's disease

Macroscopic vascular changes reflect BBB alterations observed in AD. Increased numbers of fragmented vessels and branches (191, 192), vessel tortuosity as well as the presence of non-functional and degenerative capillaries are commonly observed in AD (191, 193). Intracranial atherosclerosis, especially in the hippocampus and CGM areas, mirrors the distribution of the aforementioned CBF changes (194, 195). While atherosclerosis in these eloquent areas associates with cognitive decline in AD (196), concomitant CBF alterations in subcortical WM areas and corpus callosum during disease progression may be also to blame.

In AD, BBB breakdown contributes to neurodegeneration, amyloid accumulation and neuroinflammation (71). Loss and/or degeneration of TJs, EC and pericytes and accumulation of toxic factors, such as fibrinogen and immunoglobulins (IgG), are features of BBB disruption in AD (197-202). Breakdown of the BBB in the hippocampus has been shown to precede behavioral deficits and cognitive impairment in AD animal models (203, 204) and AD patients (205), respectively, with alterations to endothelial TJs being independent of amyloid burden. Similar features are seen in periventricular WM, where shortened TJs are commonly observed (191, 206). In addition, thickening of the vascular basal lamina is thought to play an important role in AD-related neurovascular changes with alterations of vessel-associated ECM proteins being a culprit (207, 208). Other ECs alterations are postulated to be central players in AD pathogenesis, which are reviewed in detail elsewhere (209).

NVU alterations mirror the regional and temporal evolution of CBF and BBB changes seen in AD. Temporal lobe pathology is a consistent, early feature with involvement of other cortical GM and subcortical WM regions being seen in later AD stages. In a model of adult viable pericyte-deficient mice, BBB breakdown and neuronal loss occurred in both GM and WM structures in an age-dependent fashion (140). In early AD, pericytes accumulate A β in order to degrade it, while A β oligomers reduce CBF through pericyte-mediated capillary constriction, forming a vicious circle of dysfunctional clearance and energy supply (210). Pericyte alterations associate with BBB breakdown before extensive amyloid deposition in the AD hippocampus (205), and are detected in later stages and in other cortical regions when amyloid burden is severe (211, 212). Pericytes are also affected in subcortical WM during AD progression (9, 52,

213) Further studies evaluating the nature, timing, and extent of pericyte responses to AD-relevant stimuli and how they impact the NVU in GM and WM are needed.

Activation of perivascular glial cells, such as astrocytes and microglia, are thought to be secondary features of BBB breakdown and pericyte alterations in AD. Activation of perivascular astrocytes occurs in the vicinity of amyloid plaques and leads to propagation of local neuroinflammation in CGM and DGM structures in established AD (214-218). Interestingly, a specific subtype of tau-positive perivascular astrocytes in the subcortical WM associates with brain atrophy (219, 220). Microglial activation and dysfunction are linked to oxidative stress that exacerbates neurodegeneration in CGM areas, such as frontal, temporal and cingulate cortices, as well as underlying WM in AD (221, 222). In addition, microglia are involved in vascular remodeling. Microglia amplify and sustain local inflammation in response to fibrinogen accumulation in the AD entorhinal cortex (223). This is further supported by fibrinogen-mediated microglial activation in the spinal cord of an AD mouse model which exacerbates neurodegeneration and subsequent cognitive decline, independent of amyloid accumulation (224). These findings support an important role for glial activation and dysfunction in AD pathogenesis that requires further study.

5.2.3. A β accumulation impacts blood-brain barrier integrity

BBB dysfunction in CGM and DGM contributes to impaired Aβ clearance in the AD brain. Aß can reduce EC-related LRP1 activity, a protein known to regulate the clearance of several toxic factors including A\beta itself (225, 226). Fibrillar Aβ has been reported to induce pericyte loss and apoptosis in the AD hippocampus (227), while oligomeric Aß induces pericyte-mediated hypoperfusion in CGM (parietal cortex) (212). A β is known to interact with the receptor for advanced glycation end-products (RAGE), which mediates the accumulation of $A\beta$ from ECs to neurons leading to the release of proinflammatory factors and neuronal loss (225, 228, 229). In addition, ECs derived from rat cortices subjected to AB treatment show alterations in TJs through impaired Zo-1 and occludin expression (230). Importantly, the contribution of amyloids to vascular dysfunction is highlighted by the emergence of amyloidrelated imaging abnormalities (ARIA) in AD patients treated with novel amyloid-lowering monoclonal antibodies. ARIA is thought to relate to BBB disruption secondary to the mobilization of amyloids previously deposited in blood vessels with oedema and hemorrhage subtypes being described (231, 232). Altogether, these findings provide evidence that $A\beta$ can induce BBB alterations that not only reduce its own clearance but also augment subsequent neuroinflammation and neurodegeneration, mainly in CGM and DGM areas. Very little is known about the relationship between the (marginal) amyloid deposition in WM parenchyma and vascular dysfunction and should be the focus of future study.

6. The shared landscape of vascular dysfunction in WM dementia

We have discussed three diseases largely considered under separate sub-specialities of neurology: a vascular disorder (vascular dementia), an immune-mediated disorder (MS) and a neurodegenerative disorder



FIGURE 5

The shared landscape of vascular dysfunction in WM dementia. Hypothetical maps of regional vascular dysfunction in white matter dementias share similar cerebral blood flow and blood – brain barrier alterations. In vascular dementia, early vascular dysfunction is seen in temporal, parietal and occipital cortices, subcortical white matter (in particular the corpus callosum) followed by frontal cortex, and deep gray matter structures (hippocampus, hypothalamus). Similar to vascular dementia, in multiple sclerosis, early vascular dysfunction in parietal cortex (but also in frontal cortex), white matter (in particular the corpus callosum) followed by temporo-occipital cortices and deep gray matter structures (hippocampus, such and caudate nucleus) are features. In addition, vascular dementia and multiple sclerosis share similar early cognitive deficits, including impaired executive function, processing speed and attention. In Alzheimer's disease, early vascular dysfunction is commonly encountered in the temporal lobe (including cortical and deep gray matter structures), followed by subcortical white matter (in particular the corpus callosum) and frontoparietal cortices. Early cognitive deficits in Alzheimer's disease affect episodic memory, language and visuospatial processing. For each of these diseases, vascular disturbances are suggested to be linked to juxtaventricular areas, where WM damage is consistently observed.

(AD). Cognitive impairment is a unifying feature of these diseases with important similarities and differences between them. SIVD and MS both show early deficits in executive function, processing speed and attention classically termed "subcortical dementia" due to their predominant WM involvement. In contrast, AD shows early deficits in episodic memory, language and visuospatial processing consistent with what is termed a "cortical dementia" with predominant GM involvement. However, all three conditions have important WM and GM contributions at various disease stages with changes to the vasculature being a unifying culprit as highlighted in this review.

A central role for the vasculature in the pathogenesis of vascular dementia, MS, and AD is further supported by the influence of vascular risk factors upon their incidence and severity. Smoking, in particular, is an established risk factor for vascular dementia, MS and AD (233, 234). Mid-life vascular risk factors are predictive of dementia later in life (235). Whether these risk factors for vascular dysfunction are an early primary triggering step for pathological processes or whether they reduce the threshold for separate pathological processes to manifest clinically is unknown. However, the concept of one pathology triggering an accelerated manifestation of another has recently been observed in COVID-19, where infection was associated with molecular signatures of aging in the human brain (236).

In addition, genetic and "non-vascular" environmental influences are reported in WM dementias. Their impact on vascular dysfunction is poorly understood, and warrants further investigation, as it may shed light on pathological processes common to these diseases. People carrying the *ApoE4* allele are at high risk of AD and have alterations in BBB function (201), and it was recently shown that *ApoE4* may predict cognitive decline in CIS and early RRMS patients (237), however not directly linked to vascular dysfunction. Furthermore, environmental factors such as hypovitaminosis D and obesity, can influence WM dementias in terms of incidence and severity. As an example, hypovitaminosis D is a risk factor in vascular dementia, MS and AD (238–240). Interestingly, in animal models (not of the diseases discussed in this review), hypovitaminosis D affects CBF, BBB and NVU (241, 242). This finding should be explored further in the context of dementia.

The complexity of regional neuroinflammatory and neurodegenerative processes during progression (Figure 5) and the heterogeneity within and between the different WM dementias challenge our understanding of these diseases. Further, once the disease is established, the relationships between WM and GM degeneration can be distinct (243). The classical view of the regional differences in the diseases discussed in this work is that CGM and DGM damage are the drivers of AD pathology, while WM damage is the predominant process from early onset to late stages in MS and vascular dementia pathology. This review discusses a shared and fundamental role of vascular dysfunction in WM dementia which challenges such dichotomic view (Figure 5), wherein potential compensatory mechanisms early in these diseases should be explored. This is particularly true in vascular dementia, which occurs alone or as mixed dementia with comorbid AD (244). It is therefore important to consider the potential causal factors of such alterations, wherein a large spectrum of evidence points toward accelerated aging. Whether it concerns hypoperfusion, BBB permeability or dysfunctional NVU, all these features are observed in normal aging, which are exacerbated in the context of WM dementia and must be taken into account when trying to understand these diseases independently or jointly.

Despite a growing number of neuroimaging techniques to quantify WM and GM vascular properties in human brain, little is known regarding the spatial organization, the connectivity variations, and the distribution of the cerebral vasculature in different conditions, due to their relatively new development. It is complicated by the heterogeneity of inter-subject venous and arterial distributions wherein the regulation of molecule trafficking by the BBB varies in consequence. However, the emergence of new tools to quantify longitudinally vascular changes in neurodegenerative disorders, may provide an atlas of the human cerebrovascular system, which is of crucial relevance (43).

The unifying concept in WM dementia associated with vascular dementia, MS and AD, appears to lie within the fabric of the blood supply against the backdrop of an aging brain. Though WM pathology may not be the primary cause for cognitive decline, it plays an important role. The insights from the imaging and pathology studies discussed in this review can form the basis of an important public health message; identification and treatment of traditional vascular risk factors in middle age has the potential to modify

References

1. Filley CM. White matter dementia. Ther Adv Neurol Disord. (2012) 5:267–77. doi: 10.1177/1756285612454323

2. Alber J, Alladi S, Bae H-J, Barton DA, Beckett LA, Bell JM, et al. White matter hyperintensities in vascular contributions to cognitive impairment and dementia (VCID): knowledge gaps and opportunities. *Alzheimers Dement.* (2019) 5:107–17. doi: 10.1016/j.trci.2019.02.001

3. Prins ND, Scheltens P. White matter hyperintensities, cognitive impairment and dementia: an update. *Nat Rev Neurol.* (2015) 11:157–65. doi: 10.1038/nrneurol.2015.10

pathological processes in the brain later in life across a much wider spectrum of neurological diseases than perhaps is widely recognized.

Author contributions

GD conceived the concept of the review. JP, GH, and AL performed the literature review. JP, GH, AL, MP, and GD drafted the manuscript and edited for intellectual content. All authors approved the manuscript for submission.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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6. Levit A, Hachinski V, Whitehead SN. Neurovascular unit dysregulation, white matter disease, and executive dysfunction: the shared triad of vascular cognitive

^{4.} Rosenberg GA, Sullivan N, Esiri MM. White matter damage is associated with matrix metalloproteinases in vascular dementia. *Stroke.* (2001) 32:1162–8. doi: 10.1161/01.STR.32.5.1162

^{5.} Hase Y, Horsburgh K, Ihara M, Kalaria RN. White matter degeneration in vascular and other ageing-related dementias. *J Neurochem.* (2018) 144:617–33. doi: 10.1111/jnc.14271

impairment and Alzheimer disease. *Geroscience*. (2020) 42:445-65. doi: 10.1007/ s11357-020-00164-6

7. Callahan CM, Apostolova LG, Gao S, Risacher SL, Case J, Saykin AJ, et al. Novel markers of angiogenesis in the setting of cognitive impairment and dementia. *J Alzheimers Dis.* (2020) 75:959–69. doi: 10.3233/JAD-191293

8. Bronge L, Wahlund L-O. White matter lesions in dementia: an MRI study on bloodbrain barrier dysfunction. *Dement Geriatr Cogn Disord.* (2000) 11:263–7. doi: 10.1159/000017248

9. Ding R, Hase Y, Ameen-Ali KE, Ndung'u M, Stevenson W, Barsby J, et al. Loss of capillary pericytes and the blood-brain barrier in white matter in poststroke and vascular dementias and Alzheimer's disease. *Brain Pathol.* (2020) 30:1087–101. doi: 10.1111/bpa.12888

10. Kuczynski B, Targan E, Madison C, Weiner M, Zhang Y, Reed B, et al. White matter integrity and cortical metabolic associations in aging and dementia. *Alzheimers Dement*. (2010) 6:54. doi: 10.1016/j.jalz.2009.04.1228

11. Pascual B, Prieto E, Arbizu J, Marti-Climent J, Olier J, Masdeu JC. Brain glucose metabolism in vascular white matter disease with dementia: differentiation from Alzheimer disease. *Stroke*. (2010) 41:2889–93. doi: 10.1161/STROKEAHA.110.591552

12. Bandyopadhyay S. Role of neuron and glia in Alzheimer's disease and associated vascular dysfunction. *Front Aging Neurosci.* (2021) 13:653334. doi: 10.3389/fnagi.2021.653334

13. Morató L, Bertini E, Verrigni D, Ardissone A, Ruiz M, Ferrer I, et al. Mitochondrial dysfunction in central nervous system white matter disorders. *Glia*. (2014) 62:1878–94. doi: 10.1002/glia.22670

14. Smith EE, Greenberg SM. β -Amyloid, blood vessels, and brain function. *Stroke*. (2009) 40:2601–6. doi: 10.1161/STROKEAHA.108.536839

15. Ouellette J, Lacoste B. From neurodevelopmental to neurodegenerative disorders: the vascular continuum. *Front Aging Neurosci.* (2021) 13:749026. doi: 10.3389/fnagi.2021.749026

16. Sweeney MD, Kisler K, Montagne A, Toga AW, Zlokovic BV. The role of brain vasculature in neurodegenerative disorders. *Nat Neurosci.* (2018) 21:1318–31. doi: 10.1038/s41593-018-0234-x

17. Fantini S, Sassaroli A, Tgavalekos KT, Kornbluth J. Cerebral blood flow and autoregulation: current measurement techniques and prospects for noninvasive optical methods. *Neurophotonics*. (2016) 3:031411. doi: 10.1117/1.NPh.3.3.031411

18. Clarke DD, Sokoloff L. Circulation and energy metabolism of the brain. *Basic neurochemistry: molecular, cellular and medical aspects* 6th ed. (1999) https://www.ncbi. nlm.nih.gov/books/NBK20413/ (Accessed September 9, 2022).

19. Dienel GA. Brain glucose metabolism: integration of energetics with function. *Physiol Rev.* (2019) 99:949–1045. doi: 10.1152/physrev.00062.2017

20. Faraci FM, Heistad DD. Regulation of large cerebral arteries and cerebral microvascular pressure. Circ Res. (1990) 66:8–17. doi: 10.1161/01.RES.66.1.8

21. Silverman A, Petersen NH. *Physiology, cerebral autoregulation - StatPearls.* Treasure Island, FL: StatPearls Publishing (2022).

22. Bogatcheva NV, Sergeeva MG, Dudek SM, Verin AD. Arachidonic acid cascade in endothelial pathobiology. *Microvasc Res.* (2005) 69:107–27. doi: 10.1016/j. mvr.2005.01.007

23. Kontos HA, Wei EP, Raper AJ, Rosenblum WI, Navari RM, Patterson JL. Role of tissue hypoxia in local regulation of cerebral microcirculation. *Am J Phys Heart Circ Phys.* (1978) 234:H582–91. doi: 10.1152/ajpheart.1978.234.5.H582

24. Cipolla MJ. *Control of cerebral blood flow*. Morgan and Claypool Life Sciences (2009). Avalable at: https://www.ncbi.nlm.nih.gov/books/NBK53082/ (Accessed September 8, 2022).

25. Leeuwis AE, Smith LA, Melbourne A, Hughes AD, Richards M, Prins ND, et al. Cerebral blood flow and cognitive functioning in a community-based, multi-ethnic cohort: the SABRE study. *Front Aging Neurosci.* (2018) 10:279. doi: 10.3389/fnagi.2018.00279

26. Delgado MG, Bogousslavsky J. Cerebral hyperperfusion syndrome and related conditions. *Eur Neurol.* (2020) 83:453–7. doi: 10.1159/000511307

27. Kennelly SP, Lawlor BA, Kenny RA. Review: blood pressure and dementia — a comprehensive review. *Ther Adv Neurol Disord.* (2009) 2:241–60. doi: 10.1177/1756285609103483

28. Haseloff RF, Dithmer S, Winkler L, Wolburg H, Blasig IE. Transmembrane proteins of the tight junctions at the blood-brain barrier: structural and functional aspects. *Semin Cell Dev Biol.* (2015) 38:16–25. doi: 10.1016/j.semcdb.2014.11.004

29. Qi Z, Liu KJ. The interaction of zinc and the blood-brain barrier under physiological and ischemic conditions. *Toxicol Appl Pharmacol.* (2019) 364:114–9. doi: 10.1016/j.taap.2018.12.018

30. Zaragozá R. Transport of amino acids across the blood-brain barrier. *Front Physiol.* (2020) 11:973. doi: 10.3389/fphys.2020.00973

31. Pardridge WM. Blood-brain barrier carrier-mediated transport and brain metabolism of amino acids. *Neurochem Res.* (1998) 23:635–44. doi: 10.1023/A:1022482604276

32. Cohen-Kashi-Malina K, Cooper I, Teichberg VI. Mechanisms of glutamate efflux at the blood–brain barrier: involvement of glial cells. *J Cereb Blood Flow Metab*. (2012) 32:177–89. doi: 10.1038/jcbfm.2011.121

33. Begley DJ. ABC transporters and the blood-brain barrier. *Curr Pharm Des.* (2004) 10:1295–312. doi: 10.2174/1381612043384844

34. Graves SI, Baker DJ. Implicating endothelial cell senescence to dysfunction in the ageing and diseased brain. *Basic Clin Pharmacol Toxicol.* (2020) 127:102–10. doi: 10.1111/bcpt.13403

35. Kaya M, Ahishali B. Basic physiology of the blood-brain barrier in health and disease: a brief overview. *Tissue Barriers*. (2021) 9:1840913. doi: 10.1080/21688370.2020.1840913

36. Bell AH, Miller SL, Castillo-Melendez M, Malhotra A. The neurovascular unit: effects of brain insults during the perinatal period. *Front Neurosci.* (2020) 13:1452. doi: 10.3389/fnins.2019.01452

37. Cai W, Zhang K, Li P, Zhu L, Xu J, Yang B, et al. Dysfunction of the neurovascular unit in ischemic stroke and neurodegenerative diseases: an aging effect. *Ageing Res Rev.* (2017) 34:77–87. doi: 10.1016/j.arr.2016.09.006

38. Lo EH, Rosenberg GA. The neurovascular unit in health and disease. *Stroke*. (2009) 40:S2-3. doi: 10.1161/STROKEAHA.108.534404

39. Schaeffer S, Iadecola C. Revisiting the neurovascular unit. *Nat Neurosci.* (2021) 24:1198–209. doi: 10.1038/s41593-021-00904-7

40. Lu H, Law M, Johnson G, Ge Y, van Zijl PCM, Helpern JA. Novel approach to the measurement of absolute cerebral blood volume using vascular-space-occupancy magnetic resonance imaging. *Magn Reson Med.* (2005) 54:1403–11. doi: 10.1002/mrm.20705

41. Borowsky IW, Collins RC. Metabolic anatomy of brain: a comparison of regional capillary density, glucose metabolism, and enzyme activities. *J Comp Neurol.* (1989) 288:401–13. doi: 10.1002/cne.902880304

42. Wilhelm I, Nyúl-Tóth Á, Suciu M, Hermenean A, Krizbai IA. Heterogeneity of the blood-brain barrier. *Tissue Barriers*. (2016) 4:e1143544. doi: 10.1080/21688370.2016.1143544

43. Bernier M, Cunnane SC, Whittingstall K. The morphology of the human cerebrovascular system. *Hum Brain Mapp*. (2018) 39:4962–75. doi: 10.1002/hbm.24337

44. Harris JJ, Attwell D. The energetics of CNS white matter. J Neurosci. (2012) 32:356-71. doi: 10.1523/JNEUROSCI.3430-11.2012

45. Brickman AM, Zahra A, Muraskin J, Steffener J, Holland CM, Habeck C, et al. Reduction in cerebral blood flow in areas appearing as white matter hyperintensities on magnetic resonance imaging. *Psychiatry Res.* (2009) 172:117–20. doi: 10.1016/j. pscychresns.2008.11.006

46. Li X, Sarkar SN, Purdy DE, Briggs RW. Quantifying cerebellum grey matter and white matter perfusion using pulsed arterial spin labeling. *Biomed Res Int.* (2014) 2014:108691. doi: 10.1155/2014/108691

47. Ingo C, Kurian S, Higgins J, Mahinrad S, Jenkins L, Gorelick P, et al. Vascular health and diffusion properties of normal appearing white matter in midlife. *Brain Commun.* (2021) 3:fcab080. doi: 10.1093/braincomms/fcab080

48. Palombit A, Silvestri E, Volpi T, Aiello M, Cecchin D, Bertoldo A, et al. Variability of regional glucose metabolism and the topology of functional networks in the human brain. *Neuroimage*. (2022) 257:119280. doi: 10.1016/j.neuroimage.2022.119280

49. Coomber BL, Stewart PA. Morphometric analysis of CNS microvascular endothelium. *Microvasc Res.* (1985) 30:99–115. doi: 10.1016/0026-2862(85)90042-1

50. Montagne A, Barnes SR, Sweeney MD, Halliday MR, Sagare AP, Zhao Z, et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron.* (2015) 85:296–302. doi: 10.1016/j.neuron.2014.12.032

51. Stolp HB, Dziegielewska KM, Ek CJ, Habgood MD, Lane MA, Potter AM, et al. Breakdown of the blood-brain barrier to proteins in white matter of the developing brain following systemic inflammation. *Cell Tissue Res.* (2005) 320:369–78. doi: 10.1007/ s00441-005-1088-6

52. Montagne A, Nikolakopoulou AM, Zhao Z, Sagare AP, Si G, Lazic D, et al. Pericyte degeneration causes white matter dysfunction in the mouse central nervous system. *Nat Med.* (2018) 24:326–37. doi: 10.1038/nm.4482

53. Petersen MA, Ryu JK, Chang K-J, Etxeberria A, Bardehle S, Mendiola AS, et al. Fibrinogen activates BMP signaling in oligodendrocyte progenitor cells and inhibits remyelination after vascular damage. *Neuron.* (2017) 96:1003–1012.e7. doi: 10.1016/j. neuron.2017.10.008

54. Sams EC. Oligodendrocytes in the aging brain. *Neuronal Signal.* (2021) 5:NS20210008. doi: 10.1042/NS20210008

55. Matias I, Morgado J, Gomes FCA. Astrocyte heterogeneity: impact to brain aging and disease. *Front Aging Neurosci.* (2019) 11:59. doi: 10.3389/fnagi.2019.00059

56. Tabata H. Diverse subtypes of astrocytes and their development during corticogenesis. *Front Neurosci.* (2015) 9:114. doi: 10.3389/fnins.2015.00114

57. Boisvert MM, Erikson GA, Shokhirev MN, Allen NJ. The aging astrocyte transcriptome from multiple regions of the mouse brain. *Cell Rep.* (2018) 22:269–85. doi: 10.1016/j.celrep.2017.12.039

58. Skrobot OA, O'Brien J, Black S, Chen C, DeCarli C, Erkinjuntti T, et al. The vascular impairment of cognition classification consensus study. *Alzheimers Dement*. (2017) 13:624–33. doi: 10.1016/j.jalz.2016.10.007

59. Jellinger KA. The pathology of "vascular dementia": a critical update. *J Alzheimers Dis.* (2008) 14:107–23. doi: 10.3233/JAD-2008-14110

60. Farnell FJ, Globus JH. Chronic progressive vascular subcortical encephalopathy: chronic progressive subcortical encephalitis of Binswanger. *Arch Neurol Psychiatr*. (1932) 27:593–604. doi: 10.1001/archneurpsyc.1932.02230150109006

61. Gorelick PB, Scuteri A, Black SE, DeCarli C, Greenberg SM, Iadecola C, et al. Vascular contributions to cognitive impairment and dementia. *Stroke*. (2011) 42:2672–713. doi: 10.1161/STR.0b013e3182299496

62. Kalaria RN. The pathology and pathophysiology of vascular dementia. *Neuropharmacology*. (2018) 134:226–39. doi: 10.1016/j.neuropharm.2017.12.030

63. Scheel P, Puls I, Becker G, Schöning M. Volume reduction in cerebral blood flow in patients with vascular dementia. *Lancet.* (1999) 354:2137. doi: 10.1016/S0140-6736(99)04016-7

64. Shi Y, Thrippleton MJ, Makin SD, Marshall I, Geerlings MI, de Craen AJ, et al. Cerebral blood flow in small vessel disease: a systematic review and meta-analysis. J Cereb Blood Flow Metab. (2016) 36:1653. doi: 10.1177/0271678X16662891

65. Stewart CR, Stringer MS, Shi Y, Thrippleton MJ, Wardlaw JM. Associations between white matter hyperintensity burden, cerebral blood flow and transit time in small vessel disease: an updated meta-analysis. *Front Neurol.* (2021) 12:647848. doi: 10.3389/fneur.2021.647848

66. Dolui S, Tisdall D, Vidorreta M, Jacobs DR, Nasrallah IM, Bryan RN, et al. Characterizing a perfusion-based periventricular small vessel region of interest. *Neuroimage Clin.* (2019) 23:101897. doi: 10.1016/j.nicl.2019.101897

67. Kim C-M, Alvarado RL, Stephens K, Wey H-Y, Wang DJ, Leritz EC, et al. Associations between cerebral blood flow and structural and functional brain imaging measures in individuals with neuropsychologically defined mild cognitive impairment. *Neurobiol Aging*. (2020) 86:64–74. doi: 10.1016/j.neurobiolaging.2019.10.023

68. Tan L, Xing J, Wang Z, Du X, Luo R, Wang J, et al. Study of gray matter atrophy pattern with subcortical ischemic vascular disease-vascular cognitive impairment no dementia based on structural magnetic resonance imaging. *Front Aging Neurosci.* (2023) 15:1051177. doi: 10.3389/fnagi.2023.1051177

69. Esiri M, Wilcock G, Morris J. Neuropathological assessment of the lesions of significance in vascular dementia. *J Neurol Neurosurg Psychiatry*. (1997) 63:749–53. doi: 10.1136/jnnp.63.6.749

70. Raz L, Knoefel J, Bhaskar K. The neuropathology and cerebrovascular mechanisms of dementia. J Cereb Blood Flow Metab. (2016) 36:172–86. doi: 10.1038/jcbfm.2015.164

71. Iadecola C. The pathobiology of vascular dementia. *Neuron*. (2013) 80:844–66. doi: 10.1016/j.neuron.2013.10.008

72. Black S, Gao F, Bilbao J. Understanding white matter disease. Stroke. (2009) 40:S48-52. doi: 10.1161/STROKEAHA.108.537704

73. Launer LJ, Hughes TM, White LR. Microinfarcts, brain atrophy, and cognitive function: the HAAS autopsy study. *Ann Neurol.* (2011) 70:774–80. doi: 10.1002/ana.22520

74. Vinters HV, Zarow C, Borys E, Whitman JD, Tung S, Ellis WG, et al. Review: vascular dementia: clinicopathologic and genetic considerations. *Neuropathol Appl Neurobiol.* (2018) 44:247–66. doi: 10.1111/nan.12472

75. Bridges LR, Andoh J, Lawrence AJ, Khoong CHL, Poon WW, Esiri MM, et al. Blood-brain barrier dysfunction and cerebral small vessel disease (arteriolosclerosis) in brains of older people. *J Neuropathol Exp Neurol.* (2014) 73:1026–33. doi: 10.1097/NEN.00000000000124

76. Utter S, Tamboli IY, Walter J, Upadhaya AR, Birkenmeier G, Pietrzik CU, et al. Cerebral small vessel disease-induced apolipoprotein E leakage is associated with Alzheimer disease and the accumulation of amyloid β -protein in perivascular astrocytes. J Neuropathol Exp Neurol. (2008) 67:842–56. doi: 10.1097/NEN.0b013e3181836a71

77. Wardlaw J, Sandercock PAG, Dennis MS, Starr J. Is breakdown of the blood-brain barrier responsible for lacunar stroke, leukoaraiosis, and dementia? *Stroke*. (2003) 34:806–12. doi: 10.1161/01.STR.0000058480.77236.B3

78. Simpson JE, Fernando MS, Clark L, Ince PG, Matthews F, Forster G, et al. White matter lesions in an unselected cohort of the elderly: astrocytic, microglial and oligodendrocyte precursor cell responses. *Neuropathol Appl Neurobiol*. (2007) 33:410–9. doi: 10.1111/j.1365-2990.2007.00828.x

79. Rajani RM, Quick S, Ruigrok SR, Graham D, Harris SE, Verhaaren BFJ, et al. Reversal of endothelial dysfunction reduces white matter vulnerability in cerebral small vessel disease in rats. *Sci Transl Med.* (2018) 10:eaam9507. doi: 10.1126/scitranslmed.aam9507

80. Quick S, Moss J, Rajani RM, Williams A. A vessel for change: endothelial dysfunction in cerebral small vessel disease. *Trends Neurosci.* (2021) 44:289–305. doi: 10.1016/j.tins.2020.11.003

81. An L, Shen Y, Chopp M, Zacharek A, Venkat P, Chen Z, et al. Deficiency of endothelial nitric oxide synthase (eNOS) exacerbates brain damage and cognitive deficit in a mouse model of vascular dementia. *Aging Dis.* (2021) 12:732. doi: 10.14336/AD.2020.0523

82. Yang Y, Estrada EY, Thompson JF, Liu W, Rosenberg GA. Matrix metalloproteinasemediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *J Cereb Blood Flow Metab.* (2007) 27:697–709. doi: 10.1038/sj.jcbfm.9600375 83. Roseborough AD, Rasheed B, Jung Y, Nishimura K, Pinsky W, Langdon KD, et al. Whitehead SN. Microvessel stenosis, enlarged perivascular spaces, and fibrinogen deposition are associated with ischemic periventricular white matter hyperintensities. *Brain Pathology*. (2022) 32:e13017. doi: 10.1111/bpa.13017

84. Liu Q, Bhuiyan MIH, Liu R, Song S, Begum G, Young CB, et al. Attenuating vascular stenosis-induced astrogliosis preserves white matter integrity and cognitive function. *J Neuroinflammation*. (2021) 18:1–21. doi: 10.1186/s12974-021-02234-8

85. Lee KM, Bang JH, Han J-S, Kim BY, Lee IS, Kang HW, et al. Cardiotonic pill attenuates white matter and hippocampal damage via inhibiting microglial activation and downregulating ERK and p38 MAPK signaling in chronic cerebral hypoperfused rat. BMC Complement Altern Med. (2013) 13:334. doi: 10.1186/1472-6882-13-334

86. Qin C, Fan W-H, Liu Q, Shang K, Murugan M, Wu L-J, et al. Fingolimod protects against ischemic white matter damage by modulating microglia toward M2 polarization via STAT3 pathway. *Stroke.* (2017) 48:3336–46. doi: 10.1161/STROKEAHA.117. 018505

87. Lan L-F, Zheng L, Yang X, Ji X-T, Fan Y-H, Zeng J-S. Peroxisome proliferatoractivated receptor- γ agonist pioglitazone ameliorates white matter lesion and cognitive impairment in hypertensive rats. *CNS Neurosci Ther.* (2015) 21:410–6. doi: 10.1111/ cns.12374

88. Chabriat H, Joutel A, Dichgans M, Tournier-Lasserve E, Bousser M-G. CADASIL. Lancet Neurol. (2009) 8:643-53. doi: 10.1016/S1474-4422(09)70127-9

89. Tikka S, Baumann M, Siitonen M, Pasanen P, Pöyhönen M, Myllykangas L, et al. CADASIL and CARASIL. *Brain Pathol.* (2014) 24:525. doi: 10.1111/bpa.12181

90. De Guio F, Duering M, Fazekas F, De Leeuw F-E, Greenberg SM, Pantoni L, et al. Brain atrophy in cerebral small vessel diseases: extent, consequences, technical limitations and perspectives: the HARNESS initiative. *J Cereb Blood Flow Metab*. (2020) 40:231–45. doi: 10.1177/0271678X19888967

91. Walton C, King R, Rechtman L, Kaye W, Leray E, Marrie RA, et al. Rising prevalence of multiple sclerosis worldwide: insights from the Atlas of MS, third edition. *Mult Scler.* (2020) 26:1816–21. doi: 10.1177/1352458520970841

92. DeLuca GC, Yates RL, Beale H, Morrow SA. Cognitive impairment in multiple sclerosis: clinical, radiologic and pathologic insights. *Brain Pathol.* (2015) 25:79–98. doi: 10.1111/bpa.12220

93. Filley CM, Franklin GM, Heaton RK, Rosenberg NL. White matter dementia: clinical disorders and implications. *Cogn Behav Neurol.* (1988) 1:239.

94. DeLuca J, Chiaravalloti ND, Sandroff BM. Treatment and management of cognitive dysfunction in patients with multiple sclerosis. *Nat Rev Neurol.* (2020) 16:319–32. doi: 10.1038/s41582-020-0355-1

95. Chiaravalloti ND, DeLuca J. Cognitive impairment in multiple sclerosis. Lancet Neurol. (2008) 7:1139-51. doi: 10.1016/S1474-4422(08)70259-X

96. Gilmore CP, Donaldson I, Bö L, Owens T, Lowe J, Evangelou N. Regional variations in the extent and pattern of grey matter demyelination in multiple sclerosis: a comparison between the cerebral cortex, cerebellar cortex, deep grey matter nuclei and the spinal cord. *J Neurol Neurosurg Psychiatry.* (2009) 80:182–7. doi: 10.1136/jnnp.2008.148767

97. Vercellino M, Plano F, Votta B, Mutani R, Giordana MT, Cavalla P. Grey matter pathology in multiple sclerosis. *J Neuropathol Exp Neurol.* (2005) 64:1101–7. doi: 10.1097/01.jnen.0000190067.20935.42

98. Papadopoulos D, Dukes S, Patel R, Nicholas R, Vora A, Reynolds R. Substantial Archaeocortical atrophy and neuronal loss in multiple sclerosis. *Brain Pathol.* (2009) 19:238–53. doi: 10.1111/j.1750-3639.2008.00177.x

99. Geraldes R, Esiri MM, DeLuca GC, Palace J. Age-related small vessel disease: a potential contributor to neurodegeneration in multiple sclerosis. *Brain Pathol.* (2017) 27:707–22. doi: 10.1111/bpa.12460

100. Wuerfel J, Bellmann-Strobl J, Brunecker P, Aktas O, McFarland H, Villringer A, et al. Changes in cerebral perfusion precede plaque formation in multiple sclerosis: a longitudinal perfusion MRI study. *Brain*. (2004) 127:111–9. doi: 10.1093/brain/awh007

101. Ingrisch M, Sourbron S, Morhard D, Ertl-Wagner B, Kümpfel T, Hohlfeld R, et al. Quantification of perfusion and permeability in multiple sclerosis: dynamic contrastenhanced MRI in 3D at 3T. *Investig Radiol.* (2012) 47:252–8. doi: 10.1097/ RLI.0b013e31823bfc97

102. Johnson TW, Wu Y, Nathoo N, Rogers JA, Wee Yong V, Dunn JF. Gray matter hypoxia in the brain of the experimental autoimmune encephalomyelitis model of multiple sclerosis. *PLoS One.* (2016) 11:e0167196. doi: 10.1371/journal.pone.0167196

103. Davies AL, Desai RA, Bloomfield PS, McIntosh PR, Chapple KJ, Linington C, et al. Neurological deficits caused by tissue hypoxia in neuroinflammatory disease. *Ann Neurol.* (2013) 74:815–25. doi: 10.1002/ana.24006

104. D'haeseleer M, Hostenbach S, Peeters I, Sankari SE, Nagels G, de Keyser J, et al. Cerebral Hypoperfusion: a new pathophysiologic concept in multiple sclerosis? *J Cereb Blood Flow Metab.* (2015) 35:1406–10. doi: 10.1038/jcbfm.2015.131

105. Brooks DJ, Leenders KL, Head G, Marshall J, Legg NJ, Jones T. Studies on regional cerebral oxygen utilisation and cognitive function in multiple sclerosis. *J Neurol Neurosurg Psychiatry*. (1984) 47:1182–91. doi: 10.1136/jnnp.47.11.1182

106. Sun X, Tanaka M, Kondo S, Okamoto K, Hirai S. Clinical significance of reduced cerebral metabolism in multiple sclerosis: a combined PET and MRI study. *Ann Nucl Med.* (1998) 12:89–94. doi: 10.1007/BF03164835

107. Law M, Saindane AM, Ge Y, Babb JS, Johnson G, Mannon LJ, et al. Microvascular abnormality in relapsing-remitting multiple sclerosis: perfusion MR imaging findings in normal-appearing white matter. *Radiology*. (2004) 231:645–52. doi: 10.1148/radiol.2313030996

108. D'haeseleer M, Cambron M, Vanopdenbosch L, de Keyser J. Vascular aspects of multiple sclerosis. Lancet Neurol. (2011) 10:657–66. doi: 10.1016/S1474-4422(11)70105-3

109. Jakimovski D, Bergsland N, Dwyer MG, Traversone J, Hagemeier J, Fuchs TA, et al. Cortical and deep gray matter perfusion associations with physical and cognitive performance in multiple sclerosis patients. *Front Neurol.* (2020) 11:700. doi: 10.3389/ fneur.2020.00700

110. Debernard L, Melzer TR, Stockum SV, Graham C, Wheeler-Kingshott CA, Dalrymple-Alford JC, et al. Reduced grey matter perfusion without volume loss in early relapsing-remitting multiple sclerosis. *J Neurol Neurosurg Psychiatry*. (2014) 85:544–51. doi: 10.1136/jnnp-2013-305612

111. Varga AW, Johnson G, Babb JS, Herbert J, Grossman RI, Inglese M. White matter hemodynamic abnormalities precede sub-cortical gray matter changes in multiple sclerosis. *J Neurol Sci.* (2009) 282:28–33. doi: 10.1016/j.jns.2008.12.036

112. Saindane AM, Law M, Ge Y, Johnson G, Babb JS, Grossman RI. Correlation of diffusion tensor and dynamic perfusion MR imaging metrics in normal-appearing corpus callosum: support for primary Hypoperfusion in multiple sclerosis. *Am J Neuroradiol.* (2007) 28:767–72.

113. Wakita H, Tomimoto H, Akiguchi I, Matsuo A, Lin J-X, Ihara M, et al. Axonal damage and demyelination in the white matter after chronic cerebral hypoperfusion in the rat. *Brain Res.* (2002) 924:63–70. doi: 10.1016/S0006-8993(01)03223-1

114. Marrie RA, Rudick R, Horwitz R, Cutter G, Tyry T, Campagnolo D, et al. Vascular comorbidity is associated with more rapid disability progression in multiple sclerosis. *Neurology.* (2010) 74:1041–7. doi: 10.1212/WNL.0b013e3181d6b125

115. Papadopoulou A, Müller-Lenke N, Naegelin Y, Kalt G, Bendfeldt K, Kuster P, et al. Contribution of cortical and white matter lesions to cognitive impairment in multiple sclerosis. *Mult Scler*. (2013) 19:1290–6. doi: 10.1177/1352458513475490

116. Zhang X, Zhang F, Huang D, Wu L, Ma L, Liu H, et al. Contribution of gray and White matter abnormalities to cognitive impairment in multiple sclerosis. *Int J Mol Sci.* (2017) 18:46. doi: 10.3390/ijms18010046

117. Engl C, Tiemann L, Grahl S, Bussas M, Schmidt P, Pongratz V, et al. Cognitive impairment in early MS: contribution of white matter lesions, deep grey matter atrophy, and cortical atrophy. *J Neurol.* (2020) 267:2307–18. doi: 10.1007/s00415-020-09841-0

118. Alvarez JI, Cayrol R, Prat A. Disruption of central nervous system barriers in multiple sclerosis. *Biochim Biophys Acta*. (2011) 1812:252–64. doi: 10.1016/j. bbadis.2010.06.017

119. Estes ML, Rudick RA, Barnett GH, Ransohoff RM. Stereotactic biopsy of an active multiple sclerosis lesion: immunocytochemical analysis and Neuropathologic correlation with magnetic resonance imaging. *Arch Neurol.* (1990) 47:1299–303. doi: 10.1001/archneur.1990.00530120043008

120. Lassmann H. Multiple sclerosis pathology. Cold Spring Harb Perspect Med. (2018) 8:a028936. doi: 10.1101/cshperspect.a028936

121. Ortiz GG, Pacheco-Moisés FP, Macías-Islas MÁ, Flores-Alvarado LJ, Mireles-Ramírez MA, González-Renovato ED, et al. Role of the blood-brain barrier in multiple sclerosis. *Arch Med Res.* (2014) 45:687–97. doi: 10.1016/j.arcmed.2014.11.013

122. Kirk J, Plumb J, Mirakhur M, McQuaid S. Tight junctional abnormality in multiple sclerosis white matter affects all calibres of vessel and is associated with bloodbrain barrier leakage and active demyelination. *J Pathol.* (2003) 201:319–27. doi: 10.1002/path.1434

123. Lund H, Krakauer M, Skimminge A, Sellebjerg F, Garde E, Siebner HR, et al. Blood-brain barrier permeability of normal appearing white matter in relapsing-remitting multiple sclerosis. *PLoS One.* (2013) 8:e56375. doi: 10.1371/journal. pone.0056375

124. Cramer SP, Simonsen H, Frederiksen JL, Rostrup E, Larsson HBW. Abnormal blood-brain barrier permeability in normal appearing white matter in multiple sclerosis investigated by MRI. *Neuroimage Clin.* (2014) 4:182–9. doi: 10.1016/j.nicl.2013.12.001

125. Lassmann H. Pathogenic mechanisms associated with different clinical courses of multiple sclerosis. *Front Immunol.* (2019) 9:3116. doi: 10.3389/fimmu.2018.03116

126. Duperray A, Barbe D, Raguenez G, Weksler BB, Romero IA, Couraud P-O, et al. Inflammatory response of endothelial cells to a human endogenous retrovirus associated with multiple sclerosis is mediated by TLR4. *Int Immunol.* (2015) 27:545–53. doi: 10.1093/intimm/dxv025

127. Asadzadeh Manjili F, Yousefi-Ahmadipour A, Kazemi Arababadi M. The roles played by TLR4 in the pathogenesis of multiple sclerosis; a systematic review article. *Immunol Lett.* (2020) 220:63–70. doi: 10.1016/j.imlet.2020.02.004

128. Spencer JI, Bell JS, DeLuca GC. Vascular pathology in multiple sclerosis: reframing pathogenesis around the blood-brain barrier. *J Neurol Neurosurg Psychiatry*. (2018) 89:42–52. doi: 10.1136/jnnp-2017-316011

129. Bö L, Dawson TM, Wesselingh S, Möurk S, Choi S, Kong PA, et al. Induction of nitric oxide synthase in demyelinating regions of multiple sclerosis brains. *Ann Neurol.* (1994) 36:778–86. doi: 10.1002/ana.410360515

130. Prineas JW, Lee S. Multiple sclerosis: destruction and regeneration of astrocytes in acute lesions. *J Neuropathol Exp Neurol*. (2019) 78:140–56. doi: 10.1093/jnen/nly121

131. Castellazzi M, Lamberti G, Resi MV, Baldi E, Caniatti LM, Galante G, et al. Increased levels of endothelin-1 in cerebrospinal fluid are a marker of poor visual recovery after optic neuritis in multiple sclerosis patients. *Dis Markers*. (2019) 2019:9320791. doi: 10.1155/2019/9320791

132. van Horssen J, Brink BP, de Vries HE, van der Valk P, Bø L. The blood-brain barrier in cortical multiple sclerosis lesions. *J Neuropathol Exp Neurol.* (2007) 66:321–8. doi: 10.1097/nen.0b013e318040b2de

133. Parra Corral MA, Govindarajan ST, Stefancin P, Bangiyev L, Coyle PK, Duong TQ. Characterization of gray-matter multiple sclerosis lesions using double inversion recovery, diffusion, contrast-enhanced, and volumetric MRI. *Mult Scler Relat Disord.* (2019) 31:74–81. doi: 10.1016/j.msard.2019.03.021

134. Gilmore CP, Bö L, Owens T, Lowe J, Esiri MM, Evangelou N. Spinal cord gray matter demyelination in multiple sclerosis-a novel pattern of residual plaque morphology. *Brain Pathol.* (2006) 16:202–8. doi: 10.1111/j.1750-3639.2006.00018.x

135. Geurts JJG, Bö L, Roosendaal SD, Hazes T, Daniëls R, Barkhof F, et al. Extensive hippocampal demyelination in multiple sclerosis. *J Neuropathol Exp Neurol.* (2007) 66:819–27. doi: 10.1097/nen.0b013e3181461f54

136. Huitinga I, De Groot CJA, Van der Valk P, Kamphorst W, Tilders FJH, Swaab DF. Hypothalamic lesions in multiple sclerosis. *J Neuropathol Exp Neurol.* (2001) 60:1208–18. doi: 10.1093/jnen/60.12.1208

137. Vercellino M, Masera S, Lorenzatti M, Condello C, Merola A, Mattioda A, et al. Demyelination, inflammation, and neurodegeneration in multiple sclerosis deep gray matter. J Neuropathol Exp Neurol. (2009) 68:489–502. doi: 10.1097/ NEN.0b013e3181a19a5a

138. Yates RL, Esiri MM, Palace J, Jacobs B, Perera R, DeLuca GC. Fibrin(ogen) and neurodegeneration in the progressive multiple sclerosis cortex. *Ann Neurol.* (2017) 82:259–70. doi: 10.1002/ana.24997

139. Davalos D, Mahajan KR, Trapp BD. Brain fibrinogen deposition plays a key role in MS pathophysiology – yes. *Mult Scler.* (2019) 25:1434–5. doi: 10.1177/1352458519852723

140. Bell RD, Winkler EA, Sagare AP, Singh I, LaRue B, Deane R, et al. Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron.* (2010) 68:409–27. doi: 10.1016/j.neuron.2010.09.043

141. Grygorowicz T, Dąbrowska-Bouta B, Strużyńska L. Administration of an antagonist of P2X7 receptor to EAE rats prevents a decrease of expression of claudin-5 in cerebral capillaries. *Purinergic Signal.* (2018) 14:385–93. doi: 10.1007/s11302-018-9620-9

142. Chanaday NL, Roth GA. Microglia and astrocyte activation in the frontal cortex of rats with experimental autoimmune encephalomyelitis. *Neuroscience*. (2016) 314:160–9. doi: 10.1016/j.neuroscience.2015.11.060

143. Ponath G, Ramanan S, Mubarak M, Housley W, Lee S, Sahinkaya FR, et al. Myelin phagocytosis by astrocytes after myelin damage promotes lesion pathology. *Brain*. (2017) 140:399–413. doi: 10.1093/brain/aww298

144. Davalos D, Kyu Ryu J, Merlini M, Baeten KM, Le Moan N, Petersen MA, et al. Fibrinogen-induced perivascular microglial clustering is required for the development of axonal damage in neuroinflammation. *Nat Commun.* (2012) 3:1227. doi: 10.1038/ ncomms2230

145. Ryu JK, Petersen MA, Murray SG, Baeten KM, Meyer-Franke A, Chan JP, et al. Blood coagulation protein fibrinogen promotes autoimmunity and demyelination via chemokine release and antigen presentation. *Nat Commun.* (2015) 6:8164. doi: 10.1038/ ncomms9164

146. Yates RL, Pansieri J, Li Q, Bell JS, Yee SA, Palace J, et al. The influence of HLA-DRB1*15 on the relationship between microglia and neurons in multiple sclerosis normal appearing cortical grey matter. *Brain Pathol.* 32:e13041. doi: 10.1111/ bpa.13041

147. World Alzheimer Report (2022). *Life after diagnosis: Navigating treatment, care and support.* Available at: https://www.alzint.org/resource/world-alzheimer-report-2022/ (Accessed March 29, 2023).

148. Asllani I, Habeck C, Scarmeas N, Borogovac A, Brown TR, Stern Y. Multivariate and Univariate analysis of continuous arterial spin labeling perfusion MRI in Alzheimer's disease. J Cereb Blood Flow Metab. (2008) 28:725–36. doi: 10.1038/sj.jcbfm.9600570

149. Johnson KA, Jones K, Holman BL, Becker JA, Spiers PA, Satlin A, et al. Preclinical prediction of Alzheimer's disease using SPECT. *Neurology*. (1998) 50:1563–71. doi: 10.1212/WNL.50.6.1563

150. Rodriguez G, Vitali P, Calvini P, Bordoni C, Girtler N, Taddei G, et al. Hippocampal perfusion in mild Alzheimer's disease. *Psychiatry Res Neuroimaging*. (2000) 100:65–74. doi: 10.1016/S0925-4927(00)00071-8

151. Ruitenberg A, den Heijer T, Bakker SLM, van Swieten JC, Koudstaal PJ, Hofman A, et al. Cerebral hypoperfusion and clinical onset of dementia: the Rotterdam study. *Ann Neurol.* (2005) 57:789–94. doi: 10.1002/ana.20493

152. Loskutova N, Honea RA, Brooks WM, Burns JM. Reduced limbic and hypothalamic volumes correlate with bone density in early Alzheimer's disease. *J Alzheimers Dis.* (2010) 20:313–22. doi: 10.3233/JAD-2010-1364

153. Callen DJA, Black SE, Gao F, Caldwell CB, Szalai JP. Beyond the hippocampus: MRI volumetry confirms widespread limbic atrophy in AD. *Neurology*. (2001) 57:1669–74. doi: 10.1212/WNL.57.9.1669 154. Ishii M, Iadecola C. Metabolic and non-cognitive manifestations of Alzheimer's disease: the hypothalamus as both culprit and target of pathology. *Cell Metab.* (2015) 22:761–76. doi: 10.1016/j.cmet.2015.08.016

155. Herholz K, Schopphoff H, Schmidt M, Mielke R, Eschner W, Scheidhauer K, et al. Direct comparison of spatially normalized PET and SPECT scans in Alzheimer's disease. *J Nucl Med.* (2002) 43:21–6.

156. Matsuda H. Role of neuroimaging in Alzheimer's disease, with emphasis on brain perfusion SPECT. J Nucl Med. (2007) 48:1289–300. doi: 10.2967/jnumed.106.037218

157. Alsop DC, Casement M, de Bazelaire C, Fong T, Press DZ. Hippocampal hyperperfusion in Alzheimer's disease. *NeuroImage*. (2008) 42:1267–74. doi: 10.1016/j. neuroimage.2008.06.006

158. Chen W, Song X, Beyea S, D'Arcy R, Zhang Y, Rockwood K. Advances in perfusion magnetic resonance imaging in Alzheimer's disease. *Alzheimers Dement*. (2011) 7:185–96. doi: 10.1016/j.jalz.2010.04.004

159. Ding B, Ling H, Zhang Y, Huang J, Zhang H, Wang T, et al. Pattern of cerebral hyperperfusion in Alzheimer's disease and amnestic mild cognitive impairment using voxel-based analysis of 3D arterial spin-labeling imaging: initial experience. *Clin Interv Aging*. (2014) 9:493–500. doi: 10.2147/CIA.S58879

160. Alexopoulos P, Sorg C, Förschler A, Grimmer T, Skokou M, Wohlschläger A, et al. Perfusion abnormalities in mild cognitive impairment and mild dementia in Alzheimer's disease measured by pulsed arterial spin labeling MRI. *Eur Arch Psychiatry Clin Neurosci.* (2012) 262:69–77. doi: 10.1007/s00406-011-0226-2

161. Benedictus MR, Leeuwis AE, Binnewijzend MAA, Kuijer JPA, Scheltens P, Barkhof F, et al. Lower cerebral blood flow is associated with faster cognitive decline in Alzheimer's disease. *Eur Radiol.* (2017) 27:1169–75. doi: 10.1007/s00330-016-4450-z

162. Hirono N, Mori E, Ishii K, Ikejiri Y, Imamura T, Shimomura T, et al. Frontal lobe hypometabolism and depression in Alzheimer's disease. *Neurology*. (1998) 50:380–3. doi: 10.1212/WNL.50.2.380

163. Harwood DG, Sultzer DL, Feil D, Monserratt L, Freedman E, Mandelkern MA. Frontal lobe hypometabolism and impaired insight in Alzheimer disease. *Am J Geriatr Psychiatry*. (2005) 13:934–41. doi: 10.1097/00019442-200511000-00003

164. Ahmadi K, Pereira JB, Berron D, Vogel J, Ingala S, Strandberg OT, et al. Cerebral hypoperfusion is a late pathological event in the course of Alzheimer's disease. *medRxiv*. (2021). doi: 10.1101/2021.07.02.21259911

165. Weijs RWJ, Shkredova DA, Brekelmans ACM, Thijssen DHJ, Claassen JAHR. Longitudinal changes in cerebral blood flow and their relation with cognitive decline in patients with dementia: current knowledge and future directions. *Alzheimers Dement.* 19:532–48. doi: 10.1002/alz.12666

166. Siennicki-Lantz A, Lilja B, Rosén I, Elmståhl S. Cerebral blood flow in white matter is correlated with systolic blood pressure and EEG in senile dementia of the Alzheimer type. *Dement Geriatr Cogn Disord*. (1998) 9:29–38. doi: 10.1159/000017019

167. Tohgi H, Yonezawa H, Takahashi S, Sato N, Kato E, Kudo M, et al. Cerebral blood flow and oxygen metabolism in senile dementia of Alzheimer's type and vascular dementia with deep white matter changes. *Neuroradiology*. (1998) 40:131–7. doi: 10.1007/s002340050553

168. Wharton SB, Simpson JE, Brayne C, Ince PG. Age-associated white matter lesions: the MRC cognitive function and ageing study. *Brain Pathol.* (2014) 25:35–43. doi: 10.1111/bpa.12219

169. McAleese KE, Miah M, Graham S, Hadfield GM, Walker L, Johnson M, et al. Frontal white matter lesions in Alzheimer's disease are associated with both small vessel disease and AD-associated cortical pathology. *Acta Neuropathol.* (2021) 142:937–50. doi: 10.1007/s00401-021-02376-2

170. Li R-R, He Y-S, Liu M, Nie Z-Y, Huang L-H, Lu Z, et al. Analysis of correlation between cerebral perfusion and KIM score of white matter lesions in patients with Alzheimer's disease. *Neuropsychiatr Dis Treat*. (2019) 15:2705–14. doi: 10.2147/NDT. S207069

171. Badji A, de la Colina AN, Boshkovski T, Sabra D, Karakuzu A, Robitaille-Grou M-C, et al. A cross-sectional study on the impact of arterial stiffness on the corpus callosum, a key white matter tract implicated in Alzheimer's disease. J Alzheimers Dis. (2020) 77:591–605. doi: 10.3233/JAD-200668

172. Meguro K, Constans J-M, Shimada M, Yamaguchi S, Ishizaki J, Ishii H, et al. Corpus callosum atrophy, white matter lesions, and frontal executive dysfunction in normal aging and Alzheimer's disease. A community-based study: the Tajiri project. *Int Psychogeriatr.* (2003) 15:9–25. doi: 10.1017/S104161020300872X

173. Rose SE, Chen F, Chalk JB, Zelaya FO, Strugnell WE, Benson M, et al. Loss of connectivity in Alzheimer's disease: an evaluation of white matter tract integrity with colour coded MR diffusion tensor imaging. *J Neurol Neurosurg Psychiatry*. (2000) 69:528–30. doi: 10.1136/jnnp.69.4.528

174. Gu Y, Lin Y, Huang L, Ma J, Zhang J, Xiao Y, et al. Abnormal dynamic functional connectivity in Alzheimer's disease. *CNS Neurosci Ther.* (2020) 26:962. doi: 10.1111/ cns.13387

175. Yezhuvath US, Uh J, Cheng Y, Martin-Cook K, Weiner M, Diaz-Arrastia R, et al. Forebrain-dominant deficit in cerebrovascular reactivity in Alzheimer's disease. *Neurobiol Aging.* (2012) 33:75–82. doi: 10.1016/j.neurobiolaging.2010.02.005

176. Sachdev PS, Zhuang L, Braidy N, Wen W. Is Alzheimer's a disease of the white matter? *Curr Opin Psychiatry*. (2013) 26:244. doi: 10.1097/YCO.0b013e32835ed6e8

177. Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci.* (2011) 12:723–38. doi: 10.1038/nrn3114

178. Agosta F, Pievani M, Sala S, Geroldi C, Galluzzi S, Frisoni GB, et al. White matter damage in Alzheimer disease and its relationship to gray matter atrophy. *Radiology*. (2011) 258:853–63. doi: 10.1148/radiol.10101284

179. Cardenas VA, Tosun D, Chao LL, Fletcher PT, Joshi S, Weiner MW, et al. Voxelwise co-analysis of macro- and microstructural brain alteration in mild cognitive impairment and Alzheimer's disease using anatomical and diffusion MRI. *J Neuroimaging*. (2014) 24:435–43. doi: 10.1111/jon.12002

180. Wang Q, Wang Y, Liu J, Sutphen CL, Cruchaga C, Blazey T, et al. Quantification of white matter cellularity and damage in preclinical and early symptomatic Alzheimer's disease. *Neuroimage Clin.* (2019) 22:101767. doi: 10.1016/j.nicl.2019.101767

181. Ota M, Sato N, Kimura Y, Shigemoto Y, Kunugi H, Matsuda H. Changes of myelin Organization in Patients with Alzheimer's disease shown by q-space myelin map imaging. *Dement Geriatr Cogn Dis Extra*. (2019) 9:24–33. doi: 10.1159/000493937

182. Papuć E, Rejdak K. The role of myelin damage in Alzheimer's disease pathology. Arch Med Sci. (2018) 16:345–51. doi: 10.5114/aoms.2018.76863

183. Heise V, Filippini N, Ebmeier KP, Mackay CE. The APOE ϵ 4 allele modulates brain white matter integrity in healthy adults. *Mol Psychiatry*. (2011) 16:908–16. doi: 10.1038/mp.2010.90

184. Nasrabady SE, Rizvi B, Goldman JE, Brickman AM. White matter changes in Alzheimer's disease: a focus on myelin and oligodendrocytes. *Acta Neuropathol Commun.* (2018) 6:22. doi: 10.1186/s40478-018-0515-3

185. Love S, Miners S, Palmer J, Chalmers K, Kehoe P. Insights into the pathogenesis and pathogenicity of cerebral amyloid angiopathy. *Front Biosci.* (2009) 14:4778–92. doi: 10.2741/3567

186. Fotiadis P, Reijmer YD, Van Veluw SJ, Martinez-Ramirez S, Karahanoglu FI, Gokcal E, et al. White matter atrophy in cerebral amyloid angiopathy. *Neurology*. (2020) 95:e554–62. doi: 10.1212/WNL.000000000010017

187. Charidimou A, Boulouis G, Haley K, Auriel E, van Etten ES, Fotiadis P, et al. White matter hyperintensity patterns in cerebral amyloid angiopathy and hypertensive arteriopathy. *Neurology*. (2016) 86:505–11. doi: 10.1212/WNL.00000000002362

188. Bian Z, Yamashita T, Shi X, Feng T, Yu H, Hu X, et al. Accelerated accumulation of fibrinogen peptide chains with A β deposition in Alzheimer's disease (AD) mice and human AD brains. *Brain Res.* (2021) 1767:147569. doi: 10.1016/j.brainres.2021.147569

189. Paul J, Strickland S, Melchor JP. Fibrin deposition accelerates neurovascular damage and neuroinflammation in mouse models of Alzheimer's disease. *J Exp Med.* (2007) 204:1999–2008. doi: 10.1084/jem.20070304

190. Cajamarca SA, Norris EH, van der Weerd L, Strickland S, Ahn HJ. Cerebral amyloid angiopathy-linked β-amyloid mutations promote cerebral fibrin deposits via increased binding affinity for fibrinogen. *Proc Natl Acad Sci U S A*. (2020) 117:14482–92. doi: 10.1073/pnas.1921327117

191. Baloyannis SJ, Baloyannis IS. The vascular factor in Alzheimer's disease: a study in Golgi technique and electron microscopy. *J Neurol Sci.* (2012) 322:117–21. doi: 10.1016/j.jns.2012.07.010

192. Kitaguchi H, Ihara M, Saiki H, Takahashi R, Tomimoto H. Capillary beds are decreased in Alzheimer's disease, but not in Binswanger's disease. *Neurosci Lett.* (2007) 417:128–31. doi: 10.1016/j.neulet.2007.02.021

193. Fischer VW, Siddiqi A, Yusufaly Y. Altered angioarchitecture in selected areas of brains with Alzheimer's disease. *Acta Neuropathol.* (1990) 79:672–9. doi: 10.1007/BF00294246

194. Roher AE, Esh C, Kokjohn TA, Kalback W, Luehrs DC, Seward JD, et al. Circle of Willis atherosclerosis is a risk factor for sporadic Alzheimer's disease. *Arterioscler Thromb Vasc Biol.* (2003) 23:2055–62. doi: 10.1161/01.ATV.0000095973.42032.44

195. Chalmers K, Wilcock G, Love S. Contributors to white matter damage in the frontal lobe in Alzheimer's disease. *Neuropathol Appl Neurobiol.* (2005) 31:623–31. doi: 10.1111/j.1365-2990.2005.00678.x

196. Arvanitakis Z, Capuano AW, Leurgans SE, Bennett DA, Schneider JA. Relation of cerebral vessel disease to Alzheimer's disease dementia and cognitive function in elderly people: a cross-sectional study. *Lancet Neurol.* (2016) 15:934–43. doi: 10.1016/S1474-4422(16)30029-1

197. Sengillo JD, Winkler EA, Walker CT, Sullivan JS, Johnson M, Zlokovic BV. Deficiency in mural vascular cells coincides with blood–brain barrier disruption in Alzheimer's disease. *Brain Pathol.* (2013) 23:303–10. doi: 10.1111/bpa.12004

198. Zipser BD, Johanson CE, Gonzalez L, Berzin TM, Tavares R, Hulette CM, et al. Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. *Neurobiol Aging*. (2007) 28:977–86. doi: 10.1016/j.neurobiolaging.2006.05.016

199. Miners JS, Schulz I, Love S. Differing associations between A β accumulation, hypoperfusion, blood–brain barrier dysfunction and loss of PDGFRB pericyte marker in the precuneus and parietal white matter in Alzheimer's disease. *J Cereb Blood Flow Metab.* (2018) 38:103–15. doi: 10.1177/0271678X17690761

200. Wisniewski HM, Kozlowski PB. Evidence for blood-brain barrier changes in senile dementia of the Alzheimer type (SDAT). *Ann N Y Acad Sci.* (1982) 396:119–29. doi: 10.1111/j.1749-6632.1982.tb26848.x

201. Montagne A, Nation DA, Sagare AP, Barisano G, Sweeney MD, Chakhoyan A, et al. APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature.* (2020) 581:71–6. doi: 10.1038/s41586-020-2247-3

202. Fu L, Jiang G, Weng H, Dick GM, Chang Y, Kassab GS. Cerebrovascular miRNAs correlate with the clearance of A β through perivascular route in younger 3xTg-AD mice. *Brain Pathol.* (2020) 30:92–105. doi: 10.1111/bpa.12759

203. Sagare AP, Bell RD, Zhao Z, Ma Q, Winkler EA, Ramanathan A, et al. Pericyte loss influences Alzheimer-like neurodegeneration in mice. *Nat Commun.* (2013) 4:2932. doi: 10.1038/ncomms3932

204. Webster SJ, Bachstetter AD, Nelson PT, Schmitt FA, Van Eldik LJ. Using mice to model Alzheimer's dementia: an overview of the clinical disease and the preclinical behavioral changes in 10 mouse models. *Front Genet.* (2014) 5:88. doi: 10.3389/fgene.2014.00088

205. Nation DA, Sweeney MD, Montagne A, Sagare AP, D'Orazio LM, Pachicano M, et al. Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat Med.* (2019) 25:270–6. doi: 10.1038/s41591-018-0297-y

206. Claudio L. Ultrastructural features of the blood-brain barrier in biopsy tissue from Alzheimer's disease patients. Acta Neuropathol. (1995) 91:6–14. doi: 10.1007/s004010050386

207. Magaki S, Tang Z, Tung S, Williams CK, Lo D, Yong WH, et al. The effects of cerebral amyloid angiopathy on integrity of the blood-brain barrier. *Neurobiol Aging*. (2018) 70:70–7. doi: 10.1016/j.neurobiolaging.2018.06.004

208. Lepelletier F-X, Mann DMA, Robinson AC, Pinteaux E, Boutin H. Early changes in extracellular matrix in Alzheimer's disease. *Neuropathol Appl Neurobiol.* (2017) 43:167–82. doi: 10.1111/nan.12295

209. Di Marco LY, Venneri A, Farkas E, Evans PC, Marzo A, Frangi AF. Vascular dysfunction in the pathogenesis of Alzheimer's disease — a review of endotheliummediated mechanisms and ensuing vicious circles. *Neurobiol Dis.* (2015) 82:593–606. doi: 10.1016/j.nbd.2015.08.014

210. Korte N, Nortley R, Attwell D. Cerebral blood flow decrease as an early pathological mechanism in Alzheimer's disease. *Acta Neuropathol.* (2020) 140:793–810. doi: 10.1007/s00401-020-02215-w

211. Ma Q, Zhao Z, Sagare AP, Wu Y, Wang M, Owens NC, et al. Blood-brain barrierassociated pericytes internalize and clear aggregated amyloid- β 42 by LRP1-dependent apolipoprotein E isoform-specific mechanism. *Mol Neurodegener*. (2018) 13:57. doi: 10.1186/s13024-018-0286-0

212. Nortley R, Korte N, Izquierdo P, Hirunpattarasilp C, Mishra A, Jaunmuktane Z, et al. Amyloid β oligomers constrict human capillaries in Alzheimer's disease via signaling to pericytes. *Science*. (2019) 365:eaav9518. doi: 10.1126/science.aav9518

213. Kapoor A, Nation DA. Platelet-derived growth factor-BB and white matter hyperintensity burden in APOE4 carriers. *Cereb Circ Cogn Behav.* (2022) 3:100131. doi: 10.1016/j.cccb.2022.100131

214. Argaw AT, Asp L, Zhang J, Navrazhina K, Pham T, Mariani JN, et al. Astrocytederived VEGF-A drives blood-brain barrier disruption in CNS inflammatory disease. *J Clin Invest*. (2012) 122:2454–68. doi: 10.1172/JCI60842

215. Simpson JE, Ince PG, Lace G, Forster G, Shaw PJ, Matthews F, et al. Astrocyte phenotype in relation to Alzheimer-type pathology in the ageing brain. *Neurobiol Aging*. (2010) 31:578–90. doi: 10.1016/j.neurobiolaging.2008.05.015

216. Meda L, Baron P, Scarlato G. Glial activation in Alzheimer's disease: the role of A β and its associated proteins. *Neurobiol Aging*. (2001) 22:885–93. doi: 10.1016/S0197-4580(01)00307-4

217. Spohr T. Glia in health and disease BoD - Books on Demand (2020). doi: 10.5772/intechopen.82593

218. Wyss-Coray T, Masliah E, Mallory M, McConlogue L, Johnson-Wood K, Lin C, et al. Amyloidogenic role of cytokine TGF- β 1 in transgenic mice and in Alzheimer's disease. *Nature*. (1997) 389:603–6. doi: 10.1038/39321

219. Munoz DG, Woulfe J, Kertesz A. Argyrophilic thorny astrocyte clusters in association with Alzheimer's disease pathology in possible primary progressive aphasia. *Acta Neuropathol.* (2007) 114:347–57. doi: 10.1007/s00401-007-0266-x

220. Kovacs GG, Robinson JL, Xie SX, Lee EB, Grossman M, Wolk DA, et al. Evaluating the patterns of aging-related tau astrogliopathy unravels novel insights into brain aging and neurodegenerative diseases. *J Neuropathol Exp Neurol.* (2017) 76:270–88. doi: 10.1093/jnen/nlx007

221. Woollacott IOC, Toomey CE, Strand C, Courtney R, Benson BC, Rohrer JD, et al. Microglial burden, activation and dystrophy patterns in frontotemporal lobar degeneration. *J Neuroinflammation*. (2020) 17:234. doi: 10.1186/s12974-020-01907-0

222. Davies DS, Ma J, Jegathees T, Goldsbury C. Microglia show altered morphology and reduced arborization in human brain during aging and Alzheimer's disease. *Brain Pathol.* (2017) 27:795–808. doi: 10.1111/bpa.12456

223. Ryu JK, McLarnon JG. A leaky blood–brain barrier, fibrinogen infiltration and microglial reactivity in inflamed Alzheimer's disease brain. *J Cell Mol Med.* (2009) 13:2911–25. doi: 10.1111/j.1582-4934.2008.00434.x

224. Merlini M, Rafalski VA, Coronado PRE, Gill TM, Ellisman M, Muthukumar G, et al. Fibrinogen induces microglia-mediated spine elimination and cognitive impairment in an Alzheimer's disease model. *Neuron.* (2019) 101:1099–1108.e6. doi: 10.1016/j.neuron.2019.01.014

225. Donahue JE, Flaherty SL, Johanson CE, Duncan JA, Silverberg GD, Miller MC, et al. RAGE, LRP-1, and amyloid-beta protein in Alzheimer's disease. *Acta Neuropathol.* (2006) 112:405–15. doi: 10.1007/s00401-006-0115-3

226. Owen JB, Sultana R, Aluise CD, Erickson MA, Price TOBu G, et al. Oxidative modification to LDL receptor-related protein 1 in hippocampus from subjects with Alzheimer disease: implications for A β accumulation in AD brain. *Free Radic Biol Med.* (2010) 49:1798–803. doi: 10.1016/j.freeradbiomed.2010.09.013

227. Schultz N, Brännström K, Byman E, Moussaud S, Nielsen HM, Bank TNBet al. Amyloid-beta 1-40 is associated with alterations in NG2+ pericyte population *ex vivo* and *in vitro*. *Aging Cell*. (2018) 17:e12728. doi: 10.1111/acel.12728

228. Deane R, Du Yan S, Submamaryan RK, LaRue B, Jovanovic S, Hogg E, et al. RAGE mediates amyloid- β peptide transport across the blood-brain barrier and accumulation in brain. *Nat Med.* (2003) 9:907–13. doi: 10.1038/nm890

229. Yan SD, Chen X, Fu J, Chen M, Zhu H, Roher A, et al. RAGE and amyloid-β peptide neurotoxicity in Alzheimer's disease. *Nature*. (1996) 382:685–91. doi: 10.1038/382685a0

230. Marco S, Skaper SD. Amyloid β -peptide1–42 alters tight junction protein distribution and expression in brain microvessel endothelial cells. *Neurosci Lett.* (2006) 401:219–24. doi: 10.1016/j.neulet.2006.03.047

231. Filippi M, Cecchetti G, Spinelli EG, Vezzulli P, Falini A, Agosta F. Amyloid-related imaging abnormalities and β -amyloid-targeting antibodies: a systematic review. *JAMA Neurol.* (2022) 79:291–304. doi: 10.1001/jamaneurol.2021.5205

232. Sperling RA, Jack CR, Black SE, Frosch MP, Greenberg SM, Hyman BT, et al. Amyloid related imaging abnormalities (ARIA) in amyloid modifying therapeutic trials: recommendations from the Alzheimer's association research roundtable workgroup. *Alzheimers Dement.* (2011) 7:367–85. doi: 10.1016/j.jalz.2011.05.2351

233. Durazzo TC, Mattsson N, Weiner MW. Smoking and increased Alzheimer's disease risk: a review of potential mechanisms. *Alzheimers Dement*. (2014) 10:S122–45. doi: 10.1016/j.jalz.2014.04.009

234. Wingerchuk DM. Smoking: effects on multiple sclerosis susceptibility and disease progression. *Ther Adv Neurol Disord*. (2012) 5:13–22. doi: 10.1177/1756285611425694

235. McGrath ER, Beiser AS, O'Donnell A, Himali JJ, Pase MP, Satizabal CL, et al. Determining vascular risk factors for dementia and dementia risk prediction across mid- to later life. *Neurology*. (2022) 99:e142–53. doi: 10.1212/WNL.000000000200521

236. Mavrikaki M, Lee JD, Solomon IH, Slack FJ. Severe COVID-19 induces molecular signatures of aging in the human brain. *medRxiv*. (2021). doi: 10.1101/2021.11.24. 21266779

237. Engel S, Graetz C, Salmen A, Muthuraman M, Toenges G, Ambrosius B, et al. Is APOE ε4 associated with cognitive performance in early MS? *Neurol Neuroimmunol Neuroinflamm*. (2020) 7:e728. doi: 10.1212/NXI.00000000000728

238. Munger KL, Zhang SM, O'Reilly E, Hernán MA, Olek MJ, Willett WC, et al. Vitamin D intake and incidence of multiple sclerosis. *Neurology*. (2004) 62:60–5. doi: 10.1212/01.WNL.0000101723.79681.38

239. Littlejohns TJ, Henley WE, Lang IA, Annweiler C, Beauchet O, Chaves PHM, et al. Vitamin D and the risk of dementia and Alzheimer disease. *Neurology.* (2014) 83:920–8. doi: 10.1212/WNL.00000000000755

240. Prabhakar P, Chandra SR, Supriya M, Issac TG, Prasad C, Christopher R. Vitamin D status and vascular dementia due to cerebral small vessel disease in the elderly Asian Indian population. *J Neurol Sci.* (2015) 359:108–11. doi: 10.1016/j.jns.2015.10.050

241. Pál É, Hricisák L, Lékai Á, Nagy D, Fülöp Á, Erben RG, et al. Ablation of vitamin D signaling compromises cerebrovascular adaptation to carotid artery occlusion in mice. *Cells.* (2020) 9:1457. doi: 10.3390/cells9061457

242. Takahashi S, Maeda T, Sano Y, Nishihara H, Takeshita Y, Shimizu F, et al. Active form of vitamin D directly protects the blood–brain barrier in multiple sclerosis. *Clin Exp Neuroimmunol.* (2017) 8:244–54. doi: 10.1111/cen3.12398

243. Jang H, Kwon H, Yang J-J, Hong J, Kim Y, Kim KW, et al. Correlations between gray matter and White matter degeneration in pure Alzheimer's disease, pure subcortical vascular dementia, and mixed dementia. *Sci Rep.* (2017) 7:9541. doi: 10.1038/ s41598-017-10074-x

244. Jaul E, Meiron O. Systemic and disease-specific risk factors in vascular dementia: diagnosis and prevention. *Front Aging Neurosci.* (2017) 9:333. doi: 10.3389/fnagi.2017.00333