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Metabolomics as a potential tool for monitoring patients with aneurysmal subarachnoid hemorrhage

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Metabolomics has evolved into a particularly useful tool to study interactions between metabolites and serves as an aid in unraveling the complexity of entire metabolomes. Nonetheless, it is increasingly viewed as a methodology with practical applications in the clinical setting, where identifying and quantifying biomarkers of interest could prove useful for diagnostics. Starting from a concise overview of the most prominent analytical techniques employed in metabolomics, herein we present a review of its application in studies of brain metabolism and cerebrovascular diseases, paying most attention to its uses in researching aneurysmal subarachnoid hemorrhage. Both animal models and human studies are considered, and metabolites identified as potential biomarkers are highlighted.

KEYWORDS

metabolomics, metabonomics, stroke, subarachnoid hemorrhage, intracranial aneurysm, neurocritical care

1. Background

Systems biology strategies to study relationships between biomolecules and biological processes in the past relied on the analysis of particular biomolecules and related pathways, with no information considering entire biological systems. The need for a complete overview of a specific set of biomolecules at any level of organization (e.g., DNA, RNA, protein, lipid, metabolite) led to the development of the omics approaches that generated an immense amount of data from biofluids, cells, tissues, or organisms. The term metabolomics was first coined by Oliver et al. (1); they defined it as the comprehensive and quantitative analysis of all metabolites of the biological system under study (2). In 1999 Nicholson et al. used the term metabonomics as 'the quantitative measurement of the dynamic multiparametric metabolic response of living systems to pathophysiological stimuli or genetic modification' (3). Due to similarities in definition, both terms are used interchangeably in the literature, with metabolomics being used more commonly.

Metabolomics relies on using analytical chemistry techniques that allow profiling and characterization of molecules in the specimen that provides data that ultimately leads to the identification of metabolites and metabolic pathways (4). This enabled metabolomics to be at the forefront of biomarkers research and helped understand subtle changes in biological systems in both physiological conditions and disease (5). As concentrations, polarities, and molecular masses of metabolites are quite diverse, the inherent problem of metabolomics is that no single analytical tool is sufficient and sensitive enough to measure the span of chemical entities present in the sample. Following the completion of the Human Genome Project, most of the human genome and corresponding transcriptome and proteome were known, and the database was made electronically available (6). On the other hand, a comprehensive investigation of the human metabolome was still lacking at that time. To address this, the Human Metabolome Project was launched in 2004 aiming at identifying and quantifying detectable molecules (>1 mM) in several bodily fluids, and made data from an electronic database called the Human Metabolome Database (HMDB) publicly available, rendering it the most comprehensive organism-specific database to date (7).

In the present work, we discuss how metabolomics has been used to study brain metabolism and cerebrovascular diseases with a particular reference to aneurysmal subarachnoid hemorrhage (aSAH). We briefly describe the most widely used tools in metabolomics research and overview their application in studying cerebrovascular diseases, with particular emphasis on potential biomarker identification in patient groups.

2. Tools to study metabolomics

For the purposes of research based on metabolomics, it is necessary to use such an analytical technique that would enable simultaneous detection and quantification of numerous metabolites, often in demanding biological matrices. Unfortunately, due to the complexity of metabolomes, which include a plethora of compounds that differ by physicochemical properties and concentration, a single analytical technique can never adequately address all metabolites in a given sample, which renders it a necessity to employ a multiplatform analysis if an extensive investigation is to be carried out to maximize metabolome coverage (8, 9). A comparison of some of the most widely employed techniques in metabolomics is presented in [Supplementary Table 1](#).

Any metabolomics experiment can either be targeted or untargeted. Targeted studies focus on quantifying a small number of metabolites (which are frequently chemically related), and usually involve extensive sample preparation procedures to optimize detection and quantification. Conversely, untargeted studies (also referred to as metabolic profiling) strive to achieve the detection of a wide range of metabolites and often result in finding new or hitherto unexpected compounds ([Figure 1](#)). Most

platforms currently used in metabolomics are based either on mass spectrometry (MS) or nuclear magnetic resonance (NMR) spectroscopy (8).

MS is based on the formation of charged species which are subsequently separated by their mass-charge ratio (m/z) (8). As most metabolite-derived species are singularly charged, mass spectrometry essentially provides information regarding molecular weights, but specificities in each platform setup provide additional clues which lead to analyte identification, often with the aid of spectral libraries and/or chromatographic parameters. Mass spectrometry is usually performed after chromatographic separation, which is the basis of the so-called hyphenated techniques, GC-MS and LC-MS (10). Metabolites (or their derivatives) that can vaporize when exposed to temperatures under 350°C can be subjected to gas chromatographic (GC) separation (8, 11). Liquid chromatographic (LC) separation is more versatile, but is generally thought of as providing lower chromatographic separation than GC, and is most adequate for non-polar metabolites (8, 12).

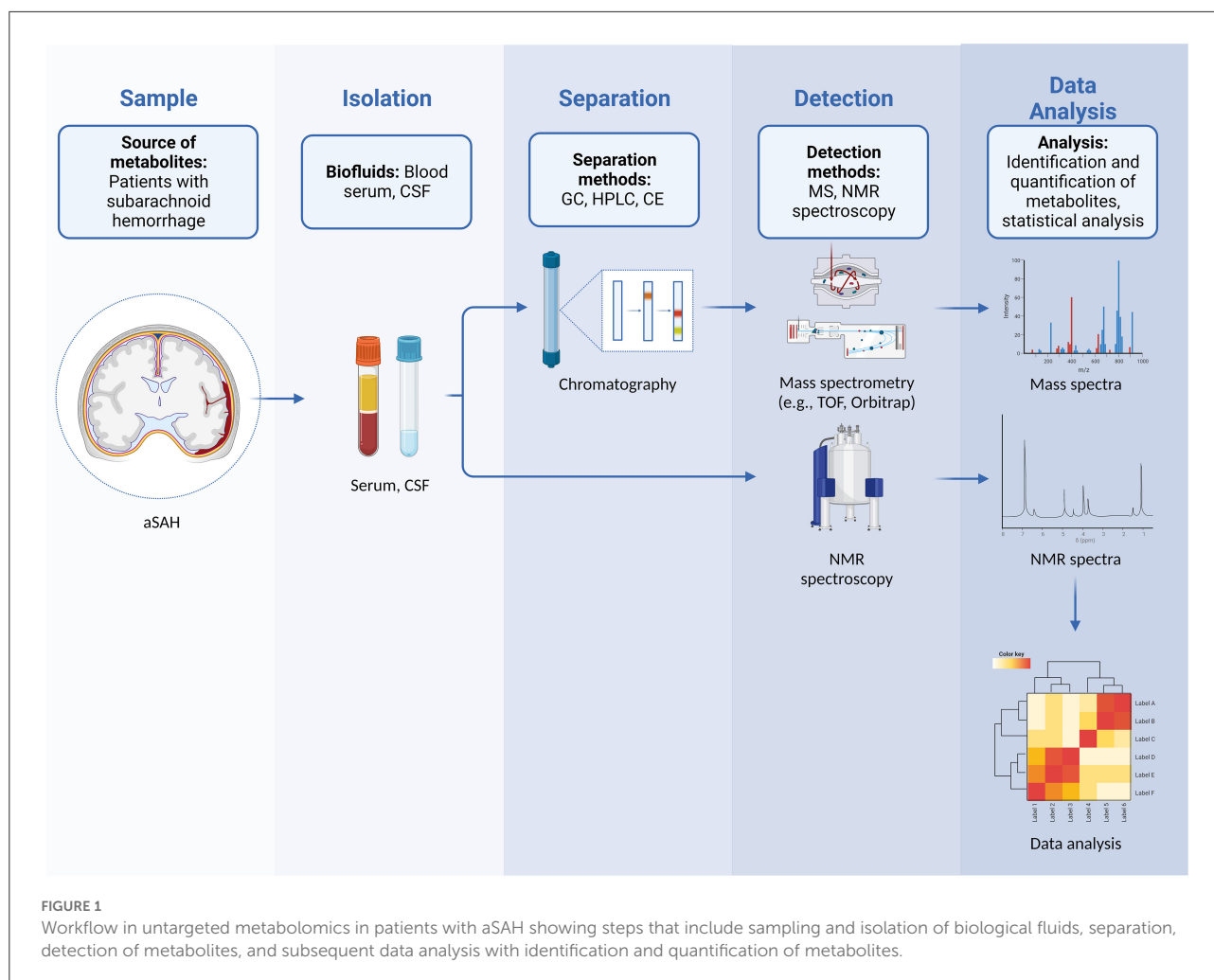
When applied in metabolomics, NMR is in a majority of cases directed at observing different hydrogen nuclei, which produce signals of specific chemical shifts and fine structure depending on their chemical environment (8, 9, 12). As biofluids contain a significant amount of water, the signal of the water hydrogen nuclei must be suppressed, or valuable data about metabolite hydrogen nuclei might be lost. Signals arising from nuclei of large molecular weight molecules can also be attenuated. Other spin-active nuclei such as ^{13}C and ^{31}P can also be studied. In the form of magnetic resonance imaging (MRI), a three-dimensional spatial mapping of metabolites is also possible (8).

3. Stroke metabolomics

Alterations in cellular metabolism preceding and during stroke induce local and systemic metabolic changes that sparked interest in investigating potential biomarkers. Such alterations may reflect the downstream function of a gene, environmental changes, and also changes in cellular signaling. Therefore, capturing the profile of circulating metabolites may provide systems-level information and insight into particular disease mechanisms, but also predict the occurrence of index events, differentiate the substrata of stroke patients, and predict the functional outcome (13).

3.1. Animal models of stroke and metabolic profile

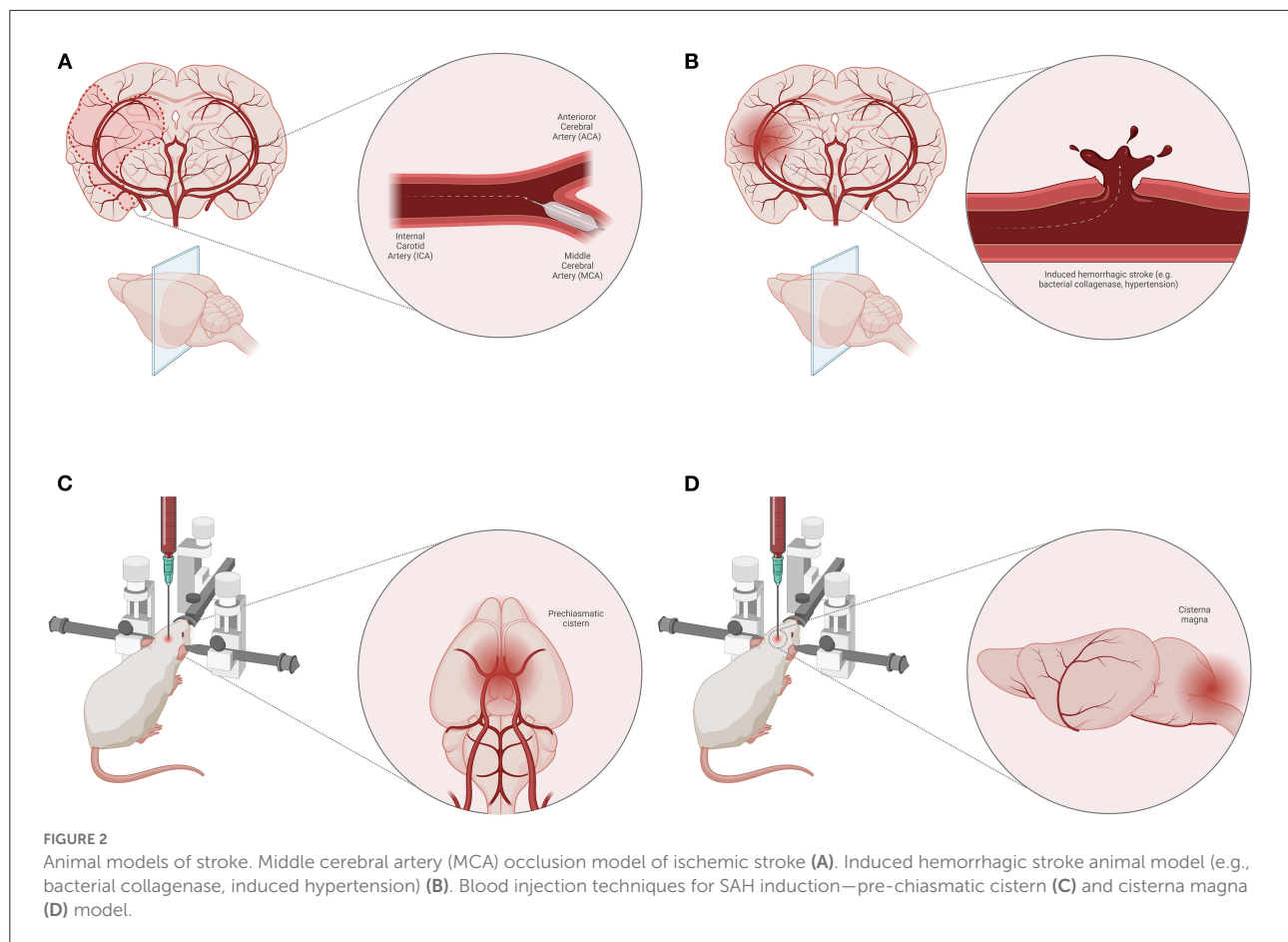
During brain damage, the changes in the metabolic profile could occur both in the brain tissue and in plasma (blood), and choosing to examine only one of the two samples could



lead to changes being overlooked, since the dysregulation in one bodily system does not mean the same would follow in another (14). However, in animal studies, the tissue samples are readily available, while in human-based studies researchers mainly depend on a blood sample and, in some cases, cerebrospinal fluid (CSF). Animal studies also enable a better understanding of how and when dysregulation occurs; e.g., blood samples can be obtained before, during, and after the animal experiments, providing further insight into metabolic pathways.

The animal models of ischemic stroke are an invaluable tool for exploring the pathophysiological mechanisms of ischemia/reperfusion (14). They have numerous advantages, such as high reproducibility, control over the ischemic area, and standardization of the experimental protocol which result in consistent conclusions regarding changes occurring at the molecular level. The most frequently used model for ischemic stroke is middle cerebral artery occlusion (Figure 2A), however, this model is not without its limitations and only moderately

mimics the pathophysiology of ischemic stroke seen in humans (14). Mainly, amino acid levels (e.g., glycine and lysine) are increased both in the brain and plasma of animals 2 days after stroke, however, the levels of methionine are only increased in brain tissue, while at the same time there is a decrease in plasma concentration of this amino acid in animals with stroke (15). In certain cases, the results from animal studies are not fully in line with those of human studies. For example, lysophosphatidylcholine, partially related with different oxidative cell damage and inflammation induction by a hitherto unknown mechanism, is found to be associated with stroke occurrence and recurrence in humans, while the same role was not observed in animals (16). However, the studies from animals and humans generally overlap and one could reach the same conclusions from the results. This is in particular true for alanine, aspartate, and glutamate metabolism which are found to be associated with a high risk for ischemic stroke in both animals and humans (17). Apart from the metabolic profile, there is a dysregulation in the cell energy which could



easily be monitored through the levels of ADP, uric acid, and lactates (17).

Spontaneous hemorrhagic stroke occurs rarely in disease-free animals, however, hemorrhagic stroke can be induced by injecting bacterial collagenase or blood into the brain parenchyma (18) (Figure 2B). This model could also be applied on spontaneously hypertensive animals (18). Although these models have some advantages, such as precise induction, and controlled volume of injecting agent, there are other disadvantages related to animals (e.g., their health status) and therapeutic approaches. In some cases, hemorrhagic stroke may be associated with poor polyamine metabolism (in genetically mutated polyamine-modulated factor 1) which results in altered blood and CSF spermine, spermidine, and putrescine levels (19). High levels of plasma homocysteine are known to cause endothelial dysfunction associated with atherosclerosis which makes it a risk factor for hemorrhagic stroke (19). Above all, one of the main risk factors for atherosclerosis and associated hemorrhagic stroke is a disturbance in lipid profile, i.e., high HDL and cholesterol, and low LDL plasma levels (19). Regarding energy depletion in animals with hemorrhagic stroke, an increase in levels of tissue serine, a ketogenic and glucogenic amino acid, is a signal of cell starvation (20). Nonetheless, an

increase in levels of phenylalanine and L-lysine could indicate a poor prognosis since they are known to increase glutamate neuron firing (21).

In animal models, subarachnoid hemorrhage (SAH) is quite hard to simulate so that it could correspond to the SAH occurring in humans. The main cause of SAH in humans is the rupture of an aneurysm, while the rodent models of SAH are defined as non-aneurysmal (22). In an attempt to do so, several techniques are performed, such as injection of blood into the subarachnoid space/cerebral ventricles or by endovascular perforation of a subarachnoid vessel. It is believed that an adequate method that models the pathophysiology in humans most closely is the intraluminal Circle of Willis perforation injury at the internal carotid artery bifurcation (23). Nonetheless, other models such as blood (homologous or heterologous) injection into pre-chiasmatic cistern (Figure 2C) and cisterna magna (Figure 2D) compromising anterior and posterior blood supply, respectively, are frequently used (22). Apart from the standard physiological parameters affected by SAH, which could be tracked in animal experiments (e.g., blood pressure, oxygen tension, cerebral perfusion pressure, etc.), contemporary studies are focused on finding specific metabolic products for SAH (22).

One of the well-known molecules associated with a more unfavorable prognosis of SAH is hemoglobin, and its increased content in CSF is found to be in correlation with cerebral ischemia and poor neurological outcome of affected animals (24). Vasospasm, a frequent complication seen in SAH patients is known to be directly related to increased concentrations of hemoglobin in CSF, as well as with bilirubin oxidation end products (BOXes), propentdyopents and Fe^{3+} ions (25). The products of energy-supplying molecules are significantly disturbed in the cerebrospinal fluid (CSF), indicating the suffering of nerve and glial cells. Often there is an increase in citrate, lactate, and pyruvate, an increased concentration of amino acids, and intensified lipid metabolism (26).

3.2. Human studies and metabolic profiles in stroke patients

3.2.1. Metabolites and stroke risk

Risk stratification prior to the index event would enable advances in prevention and therapy in patients with a particular propensity toward the development of stroke, regardless of etiology. Early metabolomic studies in elderly patients, however, limited by the number of participants, were able to identify medium- and long-chain acylcarnitines together with alanine as independent predictors of major cardiovascular events (MACE), including stroke (27). Follow-up studies on larger samples further identified a correlation between plasma ceramides and phenylacetylglutamine, a gut microbiota-derived metabolite, and increased risk through altered platelet activation and responsiveness (28, 29). Lower levels of lysine catabolites and a group of metabolites, most of them amino acids, using both targeted and untargeted approaches were found to be potential markers of thrombotic stroke (30, 31). In contrast, direct and inverse relationships between hexadecanedioate and sphingomyelin were found in ischaemic stroke, respectively (32, 33). However, one study found that none of the metabolites was consistently associated with the risk for stroke (34).

3.2.2. Metabolic signature in stroke patients

Timely diagnosis and evaluation of patients with acute stroke are essential for prompt treatment. Still, in the clinical setting, imaging modalities used for diagnosis may not reveal any changes early on and may be time-consuming, demonstrating the need for novel and sensitive biomarkers for early diagnosis. Thus far, studies were able to identify metabolomic biomarkers of transient ischemic attack (TIA) (35), ischemic (26, 36) and hemorrhagic stroke (37, 38), metabolites capable of distinguishing between the stroke types (39–41) and stages (42), but also to differentiate between stroke and stroke mimics (43). It is evident that there is strong diagnostic potential from discovering differential biomarkers. Even so, there are no

routinely used metabolic biomarkers employed in the clinical setting to date.

3.2.3. Metabolomics of aneurysmal SAH and future perspectives

Despite extensive research, predictors of outcome in patients with aSAH remain poorly understood. However, there has been a recent effort to utilize novel machine-learning methods to substantiate metabolomic biomarkers, both from blood plasma samples and the CSF, which could be potentially used in the clinical setting to foretell a functional outcome in patients with aSAH. Plasma concentrations of taurine and CSF levels of symmetric dimethylarginine, dimethylguanidino valerate, and ornithine were able to predict the 90-day functional outcome measured by the modified Rankin Scale (44, 45). Another study investigating the CSF metabolome of aSAH patients revealed differences including carbohydrate, lipid, and protein metabolism when compared to control subjects. In addition, it revealed a correlation between the pyruvate concentration and its metabolism and the severity of SAH (46, 47). Despite recent interest in providing novel insights into potential biomarkers in patients with aSAH, many questions concerning the stratification and detection of differences in patients that develop delayed cerebral ischemia, vasospasm, and other possible complications during the hospital stay and after discharge still remain unanswered, and open for future investigation.

4. Conclusion and future perspectives

There was an extensive prior effort to define metabolic profiles of stroke in both animal models and patients that may be used in the clinical setting. A recent shift of interest toward potential applications in aSAH patients, in particular, in neurocritical care, which includes risk stratification of patients and emphasis on predicting the development of early complications after the aneurysm rupture, may provide invaluable monitoring tools that will ensure better care for aSAH patients. Technological advances and increasing affordability of analytical equipment will play a pivotal role in the future implementation of omic tools, including metabolomics, not only for research purposes but for everyday patient care.

Author contributions

NL, VR, and NS reviewed literature and wrote the manuscript. NL created illustrations. VR compiled the supplementary table. All authors contributed to the critical revision of the drafted article and approved the submitted version.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fneur.2022.1101524/full#supplementary-material>

References

- Oliver SG, Winson MK, Kell DB, Baganz F. Systematic functional analysis of the yeast genome. *Trends Biotechnol.* (1998) 16:373–8. doi: 10.1016/S0167-7799(98)01214-1
- Fiehn O. Combining genomics, metabolome analysis, and biochemical modelling to understand metabolic networks. *Comp Funct Genomics.* (2001) 2:155–68. doi: 10.1002/cfg.82
- Nicholson JK, Lindon JC, Holmes E. 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica.* (1999) 29:1181–9. doi: 10.1080/004982599238047
- Moco S, Vervoort J, Bino RJ, De Vos RC, Bino R. Metabolomics technologies and metabolite identification. *Trends Analyt Chem.* (2007) 26:855–66. doi: 10.1016/j.trac.2007.08.003
- Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol.* (2016) 17:451–9. doi: 10.1038/nrm.2016.25
- Venter JC, Myers EW, Adams MD. The sequence of the human genome. *Science.* (2001) 291:1304–51. doi: 10.1126/science.1058040
- Wishart DS, Tzur D, Knox C, Eisner R, Guo AC, Young N, et al. HMDB: the human metabolome database. *Nucleic Acids Res.* (2007) 35:D521–6. doi: 10.1093/nar/gkl923
- Dunn WB, Broadhurst DI, Atherton HJ, Goodacre R, Griffin JL. Systems level studies of mammalian metabolomes: the role of mass spectrometry and nuclear magnetic resonance spectroscopy. *Chem Soc Rev.* (2011) 40:387–426. doi: 10.1039/B906712B
- Zhang A, Sun H, Wang P, Han Y, Wang X. Modern analytical techniques in metabolomics analysis. *Analyst.* (2012) 137:293–300. doi: 10.1039/C1AN15605E
- Gowda GAN, Djukovic D. Overview of mass spectrometry-based metabolomics: opportunities and challenges. In: Raftery D, editor. *Mass Spectrometry in Metabolomics: Methods and Protocols, Methods in Molecular Biology.* New York, NY: Springer Science + Business Media (2014). p. 3–12. doi: 10.1007/978-1-4939-1258-2_1
- Fancy S, Rumpel K. GC-MS-based metabolomics. In: Wang F, editor. *Methods of Pharmacology and Toxicology: Biomarker Methods in Drug Discovery and Development.* Totowa, NJ: Humana Press (2008). p. 317–340. doi: 10.1007/978-1-59745-463-6_15
- Emwas AHM. The strengths and weaknesses of NMR spectroscopy and mass spectrometry with particular focus on metabolomics research. In: Bjerrum JT, editor. *Metabonomics: Methods and Protocols.* New York, NY: Springer Science + Business Media (2015). p. 161–93. doi: 10.1007/978-1-4939-2377-9_13
- Montaner J, Ramiro L, Simats A, Tiedt S, Makris K, Jickling GC, et al. Multilevel omics for the discovery of biomarkers and therapeutic targets for stroke. *Nat Rev Neurol.* (2020) 16:247–64. doi: 10.1038/s41582-020-0350-6
- Zhang R, Meng J, Wang X, Pu L, Zhao T, Huang Y, et al. Metabolomics of ischemic stroke: insights into risk prediction and mechanisms. *Metab Brain Dis.* (2022) 37:2163–80. doi: 10.1007/s11011-022-01011-7
- Wesley UV, Bhute VJ, Hatcher JF, Palecek SP, Dempsey RJ. Local and systemic metabolic alterations in brain, plasma, and liver of rats in response to aging and ischemic stroke, as detected by nuclear magnetic resonance (NMR) spectroscopy. *Neurochem Int.* (2019) 127:113–24. doi: 10.1016/j.neuint.2019.01.025
- Law SH, Chan ML, Marathe GK, Parveen F, Chen CH, Ke LY. An updated review of lysophosphatidylcholine metabolism in human diseases. *Int J Mol Sci.* (2019) 20:1149. doi: 10.3390/ijms20051149
- Au A. Metabolomics and lipidomics of ischemic stroke. *Adv Clin Chem.* (2018) 85:31–69. doi: 10.1016/bs.acc.2018.02.002
- Jia J, Zhang H, Liang X, Dai Y, Liu L, Tan K, et al. Application of metabolomics to the discovery of biomarkers for ischemic stroke in the murine model: a comparison with the clinical results. *Mol Neurobiol.* (2021) 58:6415–26. doi: 10.1007/s12035-021-02535-2
- Alharbi BM, Tso MK, Macdonald RL. Animal models of spontaneous intracerebral hemorrhage. *Neurol Res.* (2016) 38:448–55. doi: 10.1080/01616412.2016.1144671
- Guo H, You M, Wu J, Chen A, Wan Y, Gu X, et al. Genetics of spontaneous intracerebral hemorrhage: risk and outcome. *Front Neurosci.* (2022) 16:874962. doi: 10.3389/fnins.2022.874962
- Paoli A, Bianco A, Damiani E, Bosco G. Ketogenic diet in neurodegenerative and neurodegenerative diseases. *Biomed Res Int.* (2014) 2014:474296. doi: 10.1155/2014/474296
- Wang K, Guan D, Zhao X, Qiao D, Yang Y, Cui Y. Proteomics and metabolomics of raw rhubarb and wine-processed rhubarb in the treatment of rats with intracerebral hemorrhage. *Ann Transl Med.* (2020) 8:1670. doi: 10.21037/atm-20-7831
- Leclerc JL, Garcia JM, Diller MA, Carpenter AM, Kamat PK, Hoh BL, et al. Comparison of pathophysiology in humans and rodent models of subarachnoid hemorrhage. *Front Mol Neurosci.* (2018) 11:71. doi: 10.3389/fnmol.2018.00071
- Feiler S, Friedrich B, Scholler K, Thal SC, Plesnila N. Standardized induction of subarachnoid hemorrhage in mice by intracranial pressure monitoring. *J Neurosci Methods.* (2010) 190:164–70. doi: 10.1016/j.jneumeth.2010.05.005
- Zineddine HA, Honarpisheh P, McBride D, Pandit PKT, Dienel A, Hong S-H, et al. Targeting hemoglobin to reduce delayed cerebral ischemia after subarachnoid hemorrhage. *Transl Stroke Res.* (2022) 13:725–35. doi: 10.1007/s12975-022-00995-9
- Chen Y, Galea I, Macdonald RL, Wong GKC, Zhang JH. Rethinking the initial changes in subarachnoid haemorrhage: focusing on real-time metabolism during early brain injury. *EBioMedicine.* (2022) 83:104223. doi: 10.1016/j.ebiom.2022.104223
- Zhou J, Guo P, Guo Z, Sun X, Chen Y, Feng H. Fluid metabolic pathways after subarachnoid hemorrhage. *J Neurochem.* (2022) 160:13–33. doi: 10.1111/jnc.15458
- Rizza S, Copetti M, Rossi C, Cianfarani MA, Zucchelli M, Luzzi A, et al. Metabolomics signature improves the prediction of cardiovascular events in elderly subjects. *Atherosclerosis.* (2014) 232:260–4. doi: 10.1016/j.atherosclerosis.2013.10.029
- Meeusen JW, Donato LJ, Bryant SC, Baudhuin LM, Berger PB, Jaffe AS. Plasma Ceramides. *Arterioscler Thromb Vasc Biol.* (2018) 38:1933–9. doi: 10.1161/ATVBAHA.118.311199

30. Nemet I, Saha PP, Gupta N, Zhu W, Romano KA, Skye SM, et al. A Cardiovascular disease-linked gut microbial metabolite acts via adrenergic receptors. *Cell*. (2020) 180:862–77.e22. doi: 10.1016/j.cell.2020.02.016
31. Lee Y, Khan A, Hong S, Jee SH, Park YH. A metabolomic study on high-risk stroke patients determines low levels of serum lysine metabolites: a retrospective cohort study. *Mol Biosyst*. (2017) 13:1109–20. doi: 10.1039/C6MB00732E
32. Khan A, Shin MS, Jee SH, Park YH. Global metabolomics analysis of serum from humans at risk of thrombotic stroke. *Analyst*. (2020) 145:1695–705. doi: 10.1039/C9AN02032B
33. Lind L, Salihovic S, Ganna A, Sundström J, Broeckling CD, Magnusson PK, et al. Multi-cohort metabolomics analysis discloses sphingomyelin (32:1) levels to be inversely related to incident ischemic stroke. *J Stroke Cerebrovasc Dis*. (2020) 29:104476. doi: 10.1016/j.jstrokecerebrovasdis.2019.104476
34. Sun D, Tiedt S, Yu B, Jian X, Gottesman RF, Mosley TH, et al. Prospective study of serum metabolites and risk of ischemic stroke. *Neurology*. (2019) 92:e1890–8. doi: 10.1212/WNL.0000000000007279
35. Floegel A, Kühn T, Sookthai D, Johnson T, Prehn C, Rolle-Kampczyk U, et al. Serum metabolites and risk of myocardial infarction and ischemic stroke: a targeted metabolomic approach in two German prospective cohorts. *Eur J Epidemiol*. (2018) 33:55–66. doi: 10.1007/s10654-017-0333-0
36. Purroy F, Cambray S, Mauri-Capdevila G, Jové M, Sanahuja J, Farré J, et al. Metabolomics predicts neuroimaging characteristics of transient ischemic attack patients. *EBioMedicine*. (2016) 14:131–8. doi: 10.1016/j.ebiom.2016.11.010
37. Zhou W, Li S, Sun G, Song L, Feng W, Li R, et al. Early warning of ischemic stroke based on atherosclerosis index combined with serum markers. *J Clin Endocrinol Metab*. (2022) 107:1956–64. doi: 10.1210/clinem/dgac176
38. Sun H, Zhao J, Zhong D, Li G. Potential serum biomarkers and metabolomic profiling of serum in ischemic stroke patients using UPLC/Q-TOF MS/MS. *PLoS ONE*. (2017) 12:e0189009. doi: 10.1371/journal.pone.0189009
39. Sun G, Jiang F, Hu S, Cheng H, Qu L, Tao Y, et al. Metabolomic analysis reveals potential biomarkers and serum metabolomic profiling in spontaneous intracerebral hemorrhage patients using UPLC/quadropole time-of-flight MS. *Biomed Chromatogr*. (2022) 36:e5241. doi: 10.1002/bmc.5241
40. Zhang X, Li Y, Liang Y, Sun P, Wu X, Song J, et al. Distinguishing intracerebral hemorrhage from acute cerebral infarction through metabolomics. *Rev Invest Clin*. (2017) 69:319–28. doi: 10.24875/RIC.17002348
41. Hu Z, Zhu Z, Cao Y, Wang L, Sun X, Dong J, et al. Rapid and sensitive differentiating ischemic and hemorrhagic strokes by dried blood spot based direct injection mass spectrometry metabolomics analysis. *J Clin Lab Anal*. (2016) 30:823–30. doi: 10.1002/jcla.21943
42. Lai M, Zhang X, Zhou D, Zhang X, Zhu M, Liu Q, et al. Integrating serum proteomics and metabolomics to compare the common and distinct features between acute aggressive ischemic stroke (APIS) and acute non-aggressive ischemic stroke (ANPIS). *J Proteomics*. (2022) 261:104581. doi: 10.1016/j.jprot.2022.104581
43. Sidorov EV, Xu C, Garcia-Ramiu J, Blair A, Ortiz-Garcia J, Gordon D, et al. Global metabolomic profiling reveals disrupted lipid and amino acid metabolism between the acute and chronic stages of ischemic stroke. *J Stroke Cerebrovasc Dis*. (2022) 31:106320. doi: 10.1016/j.jstrokecerebrovasdis.2022.106320
44. Tiedt S, Brandmaier S, Kollmeier H, Duering M, Artati A, Adamski J, et al. Circulating metabolites differentiate acute ischemic stroke from stroke mimics. *Ann Neurol*. (2020) 88:736–46. doi: 10.1002/ana.25859
45. Stapleton CJ, Acharjee A, Irvine HJ, Wolcott ZC, Patel AB, Kimberly WT. High-throughput metabolite profiling: identification of plasma taurine as a potential biomarker of functional outcome after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. (2019) 22:1–8. doi: 10.3171/2019.9.JNS191346
46. Koch M, Acharjee A, Ament Z, Schleicher R, Bevers M, Stapleton C, et al. Machine learning-driven metabolomic evaluation of cerebrospinal fluid: insights into poor outcomes after aneurysmal subarachnoid hemorrhage. *Neurosurgery*. (2021) 88:1003–11. doi: 10.1093/neuros/nyaa557
47. Li Y-C, Wang R, Xu M-M, Jing X-R, J-Y A, Sun R-B, et al. Aneurysmal subarachnoid hemorrhage onset alters pyruvate metabolism in poor-grade patients and clinical outcome depends on more: a cerebrospinal fluid metabolomic study. *ACS Chem Neurosci*. (2019) 10:1660–7. doi: 10.1021/acscchemneuro.8b00581