



The gastrointestinal tract microbiome and potential link to Alzheimer's disease

James M. Hill^{1,2,3}, Surjyadipta Bhattacharjee¹, Aileen I. Pogue⁴ and Walter J. Lukiw^{1,3,4,5*}

¹ LSU Neuroscience Center, Louisiana State University Health Sciences Center, New Orleans, LA, USA

² Department of Microbiology, Louisiana State University Health Sciences Center, New Orleans, LA, USA

³ Department of Ophthalmology, Louisiana State University Health Sciences Center, New Orleans, LA, USA

⁴ Alchem Biotek, Toronto, ON, Canada

⁵ Department of Neurology, Louisiana State University Health Sciences Center, New Orleans, LA, USA

*Correspondence: wlukiw@lsuhsc.edu

Edited by:

Cara Jean Westmark, University of Wisconsin, USA

Reviewed by:

Laurent Gautron, University of Texas Southwestern Medical Center, USA

Keywords: Alzheimer's disease, microbiome, genetic complexity, evolution, Bacteroidetes, Firmicutes, BMAA, BDNF

Accumulating clinical- and scientific research-based evidence is driving our increased awareness of the significance of the human microbiome (HM) to the healthy and homeostatic operation of the human central nervous system (CNS). HM communities occupy several different but distinct microbial ecosystems on and within the human body, including nasal, oral, and otic cavities, the surface of the skin and the urogenital and the gastrointestinal (GI) tracts. The complex symbiotic inter-relationship between the GI-tract microbiome and its host is strongly influenced by diet and nutrition, and when optimized can be highly beneficial to food digestion, nutrient intake, and immune health (1–6). For example, dietary composition ultimately affects the structure, organization, function, and speciation of the HM occupying the GI tract, in part by supplying multiple substrates for microbial metabolism. Typical Western diets containing high fat–cholesterol, low amounts of soluble and insoluble fiber, and sugar- and salt-enrichment not only impart deleterious nutrition but also dietary constraints on the HM. This in turn impacts the supply of microbiome-generated molecules absorbed into the systemic circulation for transport into the extensive neurovasculature of the CNS. This short communication will focus on emerging ideas concerning the contribution of the GI-tract microbiome to human neurological disease with emphasis on Alzheimer's disease (AD) wherever possible.

It is the HM of the GI tract that contains the largest reservoir of microbes in

humans, containing about 10^{14} microorganisms from at least 1000 distinct microbial species, and outnumbering human somatic cells by about 100 to 1 (1, 7). The total HM has been estimated to encode about 4×10^6 genes versus the $\sim 26,600$ genes of the human host, so again the quantity of HM genes outnumbers host genes in the order of about 150 to 1 (4). Of the 55 bacterial divisions currently identified, only two are prominent in mammalian GI-tract microbiota, including the anaerobic *Bacteroidetes* ($\sim 48\%$) and *Firmicutes* ($\sim 51\%$), with the remaining 1% of phylotypes distributed amongst the *Proteobacteria*, *Verrucomicrobia*, *Fusobacteria*, *Cyanobacteria*, *Actinobacteria*, and *Spirochetes*, with various species of fungi, protozoa, viruses, and other microorganisms making up the remainder (<http://www.genome.gov/pages/research/sequencing/seqproposals/hgmiseq.pdf>). Interestingly, microorganisms making up the smallest proportion of the HM seem to have a disproportionately large effect on host health and disease (see below). Of all GI-tract microbiota, bacterial densities of 10^{11} – 10^{12} /ml are the highest recorded density in any known microbial ecosystem of any living organism (1, 4, 7–10). There is currently expanding interest in the ability of these high density GI-tract bacteria to influence host innate-immune, neuromodulatory-, and neurotransmission-functions (3, 4, 11–14). Established pathways of GI–CNS communication and mutualism currently include the autonomic nervous system (ANS), the enteric nervous system (ENS), the

immune system, and the neuroendocrine system (15–21). Remarkably, neuronal signaling pathways along this bidirectional GI–CNS axis remain incompletely understood despite their important roles: (1) in coordinating metabolic-, nutritive-, and homeostatic-functions, and (2) in their functional disruption in chronic diseases such as anxiety, autoimmune-disease, diabetes, metabolic-syndrome, obesity, and stress-induced and progressive neuropsychiatric diseases including AD (3, 11, 12, 20, 22–24).

Here we list six specific, highly illustrative examples and recent insights into the interactive nature of the HM with a healthy, homeostatic CNS, and examples of a dysfunctional or altered HM contribution to the development of age-associated neurological disease:

- (1) studies of the ENS in germ-free “*gnotobiotic*” mice, i.e., those missing their microbiome, indicate that commensal GI-tract microbiota are critically essential for membrane electrical characteristics, including ion fluxes, action potentials, and GI-tract sensory neuron excitability, thus providing a potential mechanistic link for the initial exchange of signaling information between the GI-tract microbiome and the ANS, ENS, CNS neuroimmune–neuroendocrine systems (4, 5, 20, 23, 25);
- (2) GI-tract-abundant Gram-positive facultative anaerobic or microaerophilic *Lactobacillus*, and other *Bifidobacterium* (*Actinobacteria*) species such

as *Lactobacillus brevis* and *Bifidobacterium dentium* are capable of metabolizing glutamate to produce gamma-amino butyric acid (GABA), the major *inhibitory* neurotransmitter in the human CNS (26). Increased GI-tract GABA appears to correlate with increased CNS GABA levels, but the systemic pathways that contribute to this gut-brain linkage require additional study (3, 26). In CNS dysfunctions in GABA-mediated neuromodulatory and neurotransmission functions have been linked to the development of anxiety, behavioral deficits, epilepsy, defects in synaptogenesis, depression, and cognitive impairment including AD (16, 17, 23, 27–29). Interestingly, epileptic activities including complex partial-seizures and non-convulsive seizures are commonly associated with AD, especially in its early stages, but the contribution of GI-tract microbiome to epileptiform events via GABA modulation is not well understood (30);

- (3) the secreted, dimeric, 238 amino acid brain-derived neurotrophic factor (BDNF) essential in the maintenance and survival of neurons, has pleiotropic effects on neuronal development, differentiation, synaptogenesis, and the synaptic plasticity that underlies neuronal circuit formation and cognition, and has been found to be decreased in brains and serum from patients with anxiety, behavioral defects, schizophrenia, and AD (27, 31, 32). Interestingly, mice deficient in BDNF have altered development of GI-tract innervations including the vagus nerve, which normally serves as a major constitutive, modulatory communication pathway across the GI-CNS axis (33, 34). In experimental infection models known to lead to significant alterations in the microbiota profiles, BDNF expression was found to be reduced in the hippocampus and cortex of germ-free “*gnotobiotic*” mice, and the reduction in the expression of BDNF was found to specifically associate with increased anxiety and progressive cognitive dysfunction (20, 31, 32);
- (4) glutamate is the most abundant *excitatory* neurotransmitter in the human

CNS; the *N*-methyl-D-aspartate (NMDA) glutamate receptor, a CNS-enriched transmembrane sensor that regulates synaptic plasticity and cognition has some intriguing and potentially direct interactions with the HM; for example, the NMDA-, glutamate-targeting, glutathione-depleting, and oxidative-stress-inducing neurotoxin β -*N*-methylamino-L-alanine (BMAA), found elevated in the brains of patients with amyotrophic-lateral sclerosis (ALS), the Parkinson-dementia complex of Guam, and AD, has been hypothesized to be generated by *Cyanobacteria* of the GI-tract microbiome, and anxiety, stress, chronic intestinal inflammatory disease, or malnutrition may further induce BMAA generation to ultimately contribute to neurological dysfunction (13, 35). Interestingly, BMAA, a neurotoxic amino acid not normally incorporated into the polypeptide chains that constitute brain proteins, has been linked with intra-neuronal protein misfolding, a hallmark feature of the amyloid peptide-enriched senile plaque lesions, and resultant inflammatory neurodegeneration, that characterize AD, ALS, PD, and prion disease (21, 23, 36). These and other HM-resident *Cyanobacteria*-generated neurotoxins including saxitoxin and anatoxin- α may further contribute to neurological disease, especially over the course of aging when the intestinal epithelial barrier of the GI tract becomes significantly more permeable (13, 37);

- (5) the HM not only secretes nutritive molecules, including essential vitamins of the B and K group, but also release molecular factors that may potentially modulate or alter systemic- and CNS-amyloidosis, CNS neurochemistry, and neurotransmission. For example, HM organisms widely utilize their own naturally secreted peptides and amyloids as structural materials, adhesion molecules, and neurotoxins that ultimately function in host auto-immunity and immune-protection. The specific contribution of the HM and bacterial amyloid to protein misfolding and amyloidogenic diseases such as AD are however

not well understood, although bacterial components such as endotoxins are often found within the senile plaque lesions that characterize the AD brain (5, 21, 38). The HM further appears to condition host immunity to foreign microbes, including viral infection and xenobiotics, while regulating autoimmune responses that can impact homeostatic metabolic- and neural-signaling functions within the CNS (4, 14, 23, 39). Progressive neurological disorders such as AD have been increasingly linked to altered autoimmune and faulty innate-immune responses (12, 40, 41). An increased incidence of auto-immunity, exposure to pathogens both pre- and postnatally, and findings of antibodies to brain-specific antigens are common in disorders as diverse as anxiety, autism, depression, obsessive-compulsive disorder, schizophrenia, Parkinson's disease (PD), and AD, together suggesting that differences in exposure and genetic vulnerability toward HM-mediated auto-immunity may be significant determinants of age-related neurological disease course and outcome as humans age (14, 17, 23, 39, 42–46);

- (6) secretory products of the GI-tract microbiome and translocation of these signaling molecules via the lymphatic and systemic circulation throughout the CNS are just beginning to be identified. Recent advances in metagenomics, RNA sequencing, metatranscriptomics, metaproteomics, and metabolomics continue to clarify our perceptions of the GI-tract HM and its contribution to health and disease. Just as each individual has a unique “*stoichiometrically proportioned*” composition of microorganisms in their microbiome, individuals appear to be variably sensitive to age-related neurological disorders such as AD through the concept of “*human biochemical individuality*” (11, 16, 47). Importantly, dietary and GI-tract HM manipulation and the emergence of personalized medicine may be poised to revise and modernize our remedial efforts in the clinical management of brain disorders including AD, and the progressive transformation

to more favorable clinical outcomes (30, 48, 49).

In summary, the human GI tract is a natural habitat for large, diverse, and host-specific microbial communities including multiple species from the kingdoms of *Archaea*, *Bacteria*, the *Viruses*, and other symbiotic microbiota. How humans co-evolved with these complex microbial ecosystems, and how certain microbial species were specifically selected for mutual symbiotic benefit is of extreme interest when assessing critical HM–host interactions involving food digestion, nutrition supply and uptake, metabolic interactions, protection against pathogens and immune system development, maintenance, and dyshomeostasis in both health and disease. To cite another relevant example, abundant evidence suggests that human mitochondria originated from bacteria via endosymbiotic relationships from very early in the evolutionary history of eukaryotes, so cross-reactivity of mitochondria and host immunological responses to selective bacterial GI constituents may have deleterious effects on human mitochondrial function through molecular mimicry (4, 12, 42). This is evidenced by multiple findings in common autoimmune, inflammation-linked systemic, and neurological disorders including ALS, anxiety, diabetes, epilepsy, metabolic disease, obesity, rheumatic fever, schizophrenia, Sydenham's chorea, PD, AD, and other age-related pathologies, including transgenic animal models for these diseases (2, 4, 12, 23, 44–46, 50–54).

Lastly, since the early investigations of Koch, Metchnikoff, Pasteur, Von Leeuwenhoek, and others on the microbial basis of pathogenicity and disease transmission, Westernized societies have very successfully reduced the incidence of microbial-borne infectious disease, while an environment of autoimmune, cardiovascular, metabolic, and neuroinflammatory diseases continues to flourish. We have only recently begun to truly appreciate the potential for complex and beneficial contributions of the GI-tract HM to host genetics, phenotype, and the development and course of CNS disease. With advancement in next-generation, high throughput sequencing and metagenomic technologies our further investigations into the complex microbial ecosystems within us should yield novel

HM manipulative strategies for both the optimization of our health and the more effective clinical management of human metabolic, neuropsychiatric, and neurological disorders.

ACKNOWLEDGMENTS

This research was presented in part at the Society for Neuroscience Annual Meeting, San Diego, CA, USA, 9–13 November 2013. Research in the Lukiw laboratory involving the microbiome, microRNA, small non-coding RNA, the innate-immune response, amyloidogenesis, and neuroinflammation in AD, retinal, and prion disease, was supported through a COBRE III Pilot Award, an unrestricted grant from Research to Prevent Blindness (RPB), the Louisiana Biotechnology Research Network (LBRN), and NEI EY006311 and NIA AG038834.

REFERENCES

- Whitman WB, Coleman DC, Wiebe WJ. Prokaryotes: the unseen majority. *Proc Natl Acad Sci USA* (1998) **95**:6578–83. doi:10.1073/pnas.95.12.6578
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* (2006) **444**:1027–31. doi:10.1038/nature05414
- Bravo JA, Julio-Pieper M, Forsythe P, Kunze W, Dinan TG, Bienenstock J, et al. Communication between gastrointestinal bacteria and the nervous system. *Curr Opin Pharmacol* (2012) **12**(6):667–72. doi:10.1016/j.coph.2012.09.010
- Bhattacharjee S, Lukiw WJ. Alzheimer's disease and the microbiome. *Front Cell Neurosci* (2013) **7**:153. doi:10.3389/fncel.2013.00153
- Albenberg LG, Wu GD. Diet and the intestinal microbiome: associations, functions, and implications for health and disease. *Gastroenterology* (2014). doi:10.1053/j.gastro.2014.01.058
- Zoetendal EG, de Vos WM. Effect of diet on the intestinal microbiota and its activity. *Curr Opin Gastroenterol* (2014) **30**:189–95. doi:10.1097/MOG.0000000000000048
- Kim BS, Jeon YS, Chun J. Current status and future promise of the human microbiome. *Pediatr Gastroenterol Hepatol Nutr* (2013) **16**:71–9. doi:10.5223/pghn.2013.16.2.71
- Cammarota G, Ianiro G, Gasbarrini A. Fecal microbiota transplantation for the treatment of *Clostridium difficile* infection: a systematic review. *J Clin Gastroenterol* (2014). doi:10.1097/MCG.0000000000000046
- Manzel A, Muller DN, Hafler DA, Erdman SE, Linker RA, Kleinschewitz M. Role of “Western diet” in inflammatory autoimmune diseases. *Curr Allergy Asthma Rep* (2014) **14**:404. doi:10.1007/s11882-013-0404-6
- Pamer EG. Fecal microbiota transplantation: effectiveness, complexities, and lingering concerns. *Mucosal Immunol* (2014) **7**:210–4. doi:10.1038/mi.2013.117
- Backhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science* (2005) **307**:1915–20. doi:10.1126/science.1104816
- Boutajangout A, Wisniewski T. The innate immune system in Alzheimer's disease. *Int J Cell Biol* (2013) **2013**:576383. doi:10.1155/2013/576383
- Brenner SR. Blue-green algae or cyanobacteria in the intestinal micro-flora may produce neurotoxins such as beta-N-methylamino-L-alanine (BMAA) which may be related to development of amyotrophic lateral sclerosis, Alzheimer's disease and Parkinsons-Dementia-Complex in humans and equine motor neuron disease in horses. *Med Hypotheses* (2013) **80**(1):103–8. doi:10.1016/j.mehy.2012.10.010
- Douglas-Escobar M, Elliott E, Neu J. Effect of intestinal microbial ecology on the developing brain. *JAMA Pediatr* (2013) **167**:374–9. doi:10.1001/jamapediatrics.2013.497
- Heijtj RD, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, et al. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci USA* (2011) **108**:3047–52. doi:10.1073/pnas.1010529108
- Aziz Q, Doré J, Emmanuel A, Guarner F, Quigley EM. Gut microbiota and gastrointestinal health: current concepts and future directions. *Neurogastroenterol Motil* (2013) **25**:4–15. doi:10.1111/nmo.12046
- Camfield DA, Owen L, Scholey AB, Pipingas A, Stough C. Dairy constituents and neurocognitive health in ageing. *Br J Nutr* (2011) **106**:159–74. doi:10.1017/S0007114511000158
- Collins SM, Kassam Z, Bercik P. The adoptive transfer of behavioral phenotype via the intestinal microbiota: experimental evidence and clinical implications. *Curr Opin Microbiol* (2013) **16**(3):240–5. doi:10.1016/j.mib.2013.06.004
- Forsythe P, Kunze WA, Bienenstock J. On communication between gut microbes and the brain. *Curr Opin Gastroenterol* (2012) **28**:557–62. doi:10.1097/MOG.0b013e3283572ffa
- Foster JA, McVey Neufeld KA. Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci* (2013) **36**:305–12. doi:10.1016/j.tins.2013.01.005
- Schwartz K, Boles BR. Microbial amyloids—functions and interactions within the host. *Curr Opin Microbiol* (2013) **16**:93–9. doi:10.1016/j.mib.2012.12.001
- Lukiw WJ, Bazan NG. Survival signaling in Alzheimer's disease. *Biochem Soc Trans* (2006) **34**:1277–82. doi:10.1042/BST0341277
- Hornig M. The role of microbes and autoimmunity in the pathogenesis of neuropsychiatric illness. *Curr Opin Rheumatol* (2013) **25**:488–795. doi:10.1097/BOR.0b013e32836208de
- Udit S, Gautron L. Molecular anatomy of the gut-brain axis revealed with transgenic technologies: implications in metabolic research. *Front Neurosci* (2013) **7**:134. doi:10.3389/fnins.2013.00134
- McVey Neufeld KA, Mao YK, Bienenstock J, Foster JA, Kunze WA. The microbiome is essential for normal gut intrinsic primary afferent neuron excitability in the mouse. *Neurogastroenterol Motil* (2013) **25**(2):183–e88. doi:10.1111/nmo.12049
- Barrett E, Ross RP, O'Toole PW, Fitzgerald GF, Stanton C. γ -Aminobutyric acid production by

- culturable bacteria from the human intestine. *J Appl Microbiol* (2012) **113**:411–7. doi:10.1111/j.1365-2672.2012.05344.x
27. Mitew S, Kirkcaldie MT, Dickson TC, Vickers JC. Altered synapses and gliotransmission in Alzheimer's disease and AD model mice. *Neurobiol Aging* (2013) **34**:2341–51. doi:10.1016/j.neurobiolaging.2013.04.010
 28. Paula-Lima AC, Brito-Moreira J, Ferreira ST. Dereglulation of excitatory neurotransmission underlying synapse failure in Alzheimer's disease. *J Neurochem* (2013) **126**:191–202. doi:10.1111/jnc.12304
 29. Saulnier DM, Ringel Y, Heyman MB, Foster JA, Bercik P, Shulman RJ, et al. The intestinal microbiome, probiotics and prebiotics in neurogastroenterology. *Gut Microbes* (2013) **4**:17–27. doi:10.4161/gmic.22973
 30. Vossel KA, Beagle AJ, Rabinovici GD, Shu H, Lee SE, Naasan G, et al. Seizures and epileptiform activity in the early stages of Alzheimer disease. *JAMA Neurol* (2013) **70**:1158–66. doi:10.1001/jamaneurol.2013.136
 31. Carlino D, De Vanna M, Tongiorgi E. Is altered BDNF biosynthesis a general feature in patients with cognitive dysfunction? *Neuroscientist* (2013) **19**:345–53. doi:10.1177/1073858412469444
 32. Lu B, Nagappan G, Guan X, Nathan PJ, Wren P. BDNF-based synaptic repair as a disease-modifying strategy for neurodegenerative diseases. *Nat Rev Neurosci* (2013) **14**:401–16. doi:10.1038/nrn3505
 33. Murphy MC, Fox EA. Mice deficient in brain-derived neurotrophic factor have altered development of gastric vagal sensory innervation. *J Comp Neurol* (2010) **518**(15):2934–51. doi:10.1002/cne.22372
 34. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U S A* (2011) **108**(38):16050–5. doi:10.1073/pnas.1102999108
 35. Lakhan SE, Caro M, Hadzimidichalis N. NMDA receptor activity in neuropsychiatric disorders. *Front Psychiatry* (2013) **4**:52. doi:10.3389/fpsy.2013.00052
 36. Mulligan VK, Chakrabarty A. Protein misfolding in the late-onset neurodegenerative diseases: common themes and the unique case of amyotrophic lateral sclerosis. *Proteins* (2013) **81**:1285–303. doi:10.1002/prot.24285
 37. Tran L, Greenwood-Van Meerveld B. Age-associated remodeling of the intestinal epithelial barrier. *J Gerontol A Biol Sci Med Sci* (2013) **68**:1045–56. doi:10.1093/gerona/glt106
 38. Asti A, Gioglio L. Can a bacterial endotoxin be a key factor in the kinetics of amyloid fibril formation? *J Alzheimers Dis* (2014) **39**:169–79. doi:10.3233/JAD-131394
 39. Ball MJ, Lukiw WJ, Kammerman EM, Hill JM. Intracerebral propagation of Alzheimer's disease: strengthening evidence of a herpes simplex virus etiology. *Alzheimers Dement* (2013) **9**:169–75. doi:10.1016/j.jalz.2012.07.005
 40. Zhao Y, Bhattacharjee S, Jones BM, Dua P, Alexandrov PN, Hill JM, et al. Regulation of TREM2 expression by an NF- κ B-sensitive miRNA-34a. *Neuroreport* (2013) **24**:318–23. doi:10.1097/WNR.0b013e32835fb6b0
 41. Zhao Y, Lukiw WJ. TREM2 signaling, miRNA-34a and the extinction of phagocytosis. *Front Cell Neurosci* (2013) **7**:131. doi:10.3389/fncel.2013.00131
 42. Baum H. Mitochondrial antigens, molecular mimicry and autoimmune disease. *Biochim Biophys Acta* (1995) **1271**(1):111–21. doi:10.1016/0925-4439(95)00017-X
 43. Hill JM, Ball MJ, Neumann DM, Azcuy AM, Bhattacharjee PS, Bouhanik S, et al. The high prevalence of herpes simplex virus type 1 DNA in human trigeminal ganglia is not a function of age or gender. *J Virol* (2008) **82**:8230–4. doi:10.1128/JVI.00686-08
 44. Finegold SM, Dowd SE, Gontcharova V, Liu C, Henley KE, Wolcott RD, et al. Pyrosequencing study of fecal microflora of autistic and control children. *Anaerobe* (2010) **16**:444–53. doi:10.1016/j.anaerobe.2010.06.008
 45. Derkinderen P, Rouaud T, Lebouvier T, Bruley des Varannes S, Neunlist M, De Giorgio R. Parkinson disease: the enteric nervous system spills its guts. *Neurology* (2011) **77**:1761–7. doi:10.1212/WNL.0b013e318236ef60
 46. Cersosimo MG, Raina GB, Pecci C, Pellene A, Calandra CR, Gutiérrez C, et al. Gastrointestinal manifestations in Parkinson's disease: prevalence and occurrence before motor symptoms. *J Neurol* (2013) **260**:1332–8. doi:10.1007/s00415-012-6801-2
 47. Lukiw WJ. Variability in micro RNA (miRNA) abundance, speciation and complexity amongst different human populations and potential relevance to Alzheimer's disease (AD), "Regulatory RNAs in the Nervous System". *Front Cell Neurosci* (2013) **7**:133. doi:10.3389/fncel.2013.00133
 48. Huang Y, Mucke L. Alzheimer mechanisms and therapeutic strategies. *Cell* (2012) **148**:1204–22. doi:10.1016/j.cell.2012.02.040
 49. Souslova T, Marple TC, Spiekerman AM, Mohammad AA. Personalized medicine in Alzheimer's disease and depression. *Contemp Clin Trials* (2013) **36**:616–23. doi:10.1016/j.cct.2013.06.012
 50. Fujinami RS, von Herrath MG, Christen U, Whitton JL. Molecular mimicry, bystander activation, or viral persistence: infections and autoimmune disease. *Clin Microbiol Rev* (2006) **19**:80–94. doi:10.1128/CMR.19.1.80-94.2006
 51. Hayashi M. Anti-basal ganglia antibody. *Brain Nerve* (2013) **65**(4):377–84.
 52. Semar S, Klotz M, Letiembre M, Van Ginneken C, Braun A, Jost V, et al. Changes of the enteric nervous system in amyloid- β protein precursor transgenic mice, correlate with disease progression. *J Alzheimers Dis* (2013) **36**:7–20. doi:10.3233/JAD-120511
 53. Singh VP, Sharma J, Babu S, Rizwanulla Singla A. Role of probiotics in health and disease: a review. *J Pak Med Assoc* (2013) **63**:253–7.
 54. Tuohy KM, Fava F, Viola R. 'The way to a man's heart is through his gut microbiota' – dietary pro- and prebiotics for the management of cardiovascular risk. *Proc Nutr Soc* (2014) **4**:1–14. doi:10.1017/S0029665113003911

Received: 25 February 2014; accepted: 21 March 2014; published online: 04 April 2014.

Citation: Hill JM, Bhattacharjee S, Pogue AI and Lukiw WJ (2014) The gastrointestinal tract microbiome and potential link to Alzheimer's disease. *Front. Neurol.* **5**:43. doi: 10.3389/fneur.2014.00043

This article was submitted to *Epilepsy*, a section of the journal *Frontiers in Neurology*.

Copyright © 2014 Hill, Bhattacharjee, Pogue and Lukiw. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.