



Infection and Immunometabolism in the Central Nervous System: A Possible Mechanistic Link Between Metabolic Imbalance and Dementia

Noriko Shinjyo^{1,2*} and Kiyoshi Kita^{1,3}

¹ School of Tropical Medicine and Global Health, Nagasaki University, Nagasaki, Japan, ² Laboratory of Immune Homeostasis, WPI Immunology Frontier Research Center, Osaka University, Suita, Japan, ³ Department of Host-Defense Biochemistry, Institute of Tropical Medicine, Nagasaki University, Nagasaki, Japan

OPEN ACCESS

Edited by:

Agnes Lumi Nishimura,
Queen Mary University of London,
United Kingdom

Reviewed by:

Vijay Kumar,
The University of Tennessee Health
Science Center (UTHSC),
United States

Alberto Camacho-Morales,
Autonomous University of Nuevo
León, Mexico

*Correspondence:

Noriko Shinjyo
nshinjyo@ifrec.osaka-u.ac.jp

Specialty section:

This article was submitted to
Cellular Neuropathology,
a section of the journal
Frontiers in Cellular Neuroscience

Received: 26 August 2021

Accepted: 12 October 2021

Published: 02 November 2021

Citation:

Shinjyo N and Kita K (2021)
Infection and Immunometabolism
in the Central Nervous System: A
Possible Mechanistic Link Between
Metabolic Imbalance and Dementia.
Front. Cell. Neurosci. 15:765217.
doi: 10.3389/fncel.2021.765217

Metabolic syndromes are frequently associated with dementia, suggesting that the dysregulation of energy metabolism can increase the risk of neurodegeneration and cognitive impairment. In addition, growing evidence suggests the link between infections and brain disorders, including Alzheimer's disease. The immune system and energy metabolism are in an intricate relationship. Infection triggers immune responses, which are accompanied by imbalance in cellular and organismal energy metabolism, while metabolic disorders can lead to immune dysregulation and higher infection susceptibility. In the brain, the activities of brain-resident immune cells, including microglia, are associated with their metabolic signatures, which may be affected by central nervous system (CNS) infection. Conversely, metabolic dysregulation can compromise innate immunity in the brain, leading to enhanced CNS infection susceptibility. Thus, infection and metabolic imbalance can be intertwined to each other in the etiology of brain disorders, including dementia. Insulin and leptin play pivotal roles in the regulation of immunometabolism in the CNS and periphery, and dysfunction of these signaling pathways are associated with cognitive impairment. Meanwhile, infectious complications are often comorbid with diabetes and obesity, which are characterized by insulin resistance and leptin signaling deficiency. Examples include human immunodeficiency virus (HIV) infection and periodontal disease caused by an oral pathogen *Porphyromonas gingivalis*. This review explores potential interactions between infectious agents and insulin and leptin signaling pathways, and discuss possible mechanisms underlying the relationship between infection, metabolic dysregulation, and brain disorders, particularly focusing on the roles of insulin and leptin.

Keywords: infection, immunometabolism, glia, neuroinflammation, dementia, leptin, insulin

INTRODUCTION

Dementia is a general term for debilitating conditions, in which progressive and long-lasting loss of mental ability impairs cognition and simple daily activities. Alzheimer's disease (AD), the most common form of dementia, is a neurodegenerative disorder characterized by cognitive decline associated with the accumulation of β -amyloid (A β) plaques and neurofibrillary tangles in the brain (Ferri et al., 2005). Amyloid cascade hypothesis is a model postulating a linear pathway initiated by A β deposition, eventually leading to neuroinflammation and neuronal loss. Although amyloid

cascade hypothesis has provided the theoretical framework for the research direction and advanced the knowledge and understanding of AD pathology at molecular levels in the last decades, it remains controversial whether A β is the cause of the pathogenesis (De Strooper and Karran, 2016).

While the exact role for A β in dementia etiology remains unclear, several elements have been suggested to increase the risk of cognitive decline. Among those suggested are impaired glycemic control [e.g., metabolic syndromes (MetS) including diabetes] and infection (Ott et al., 1999; Peila et al., 2002; Arvanitakis et al., 2004; Biessels et al., 2006; Irie et al., 2008; Biessels and Despa, 2018). Largely due to modern lifestyle and diet (i.e., the lack of exercise and food with high sugar and carbohydrate), overweight, obesity, and associated metabolic disorders are widespread epidemics. Obesity is a common risk factor for many chronic disorders, such as type 2 diabetes (T2DM) and cardiovascular diseases (Haslam and James, 2005). In addition, evidence suggests that metabolic disturbance can cause neurodegenerative disorders, including AD (Whitmer et al., 2005; Kivipelto et al., 2006; Mejido et al., 2020), possibly via blood-brain barrier (BBB) disruption and neuroinflammation (Pugazhenthil et al., 2017). The association may also be compounded and influenced by other factors, such as age (Bos et al., 2017; Deckers et al., 2017; Whitmer et al., 2018), indicating the complexity of mechanisms underlying the link between metabolic disturbances and cognitive decline. Meanwhile, the relationship between infections and dementia, especially late-onset AD, has been repeatedly suggested over the past three decades (Ashraf et al., 2019). The aim of this review is to overview and discuss how these two contributory causes may be intertwined in the etiology of neurodegenerative disorders, particularly focusing on the interaction between infectious agents and two metabolic hormones, namely insulin and leptin, in the brain and periphery.

METABOLIC DISTURBANCE AND COGNITIVE IMPAIRMENT

Insulin and Leptin – Linking Metabolic Syndromes, Infection, and Cognitive Impairment

Energy homeostasis is regulated by intricate interactions between the peripheral organs and central regulatory system in the brain, where insulin and leptin play crucial roles (Boucsein et al., 2021) (**Figure 1**). Insulin and leptin resistance, as well as dysregulation of related pathways, are associated with obesity and metabolic disorders (Francisco et al., 2019; Gruzdeva et al., 2019) and central nervous system (CNS) dysfunctions (Cereda et al., 2007; Arnold et al., 2018; Kellar and Craft, 2020). Growing evidence suggests that insulin and leptin play significant physiological roles in cognition (Paz-Filho et al., 2008; Morrison, 2009; Gray et al., 2014; Arnold et al., 2018), and these signaling pathways may be promising therapeutic targets to alleviate cognitive impairment accompanied by obesity and MetS (Mejido et al., 2020). On

the other hand, obesity, visceral adiposity in particular, is frequently associated with immune dysregulation and infection susceptibility (Hamdy et al., 2006; Conde et al., 2010; Kumari et al., 2019; Obradovic et al., 2021), while infection increases the risk of neurodegeneration and dementia (Heneka et al., 2020; Shinjo et al., 2021), suggesting the link between MetS, infection, and cognitive impairment.

Insulin Insulin and Dementia

Insulin is a peptide hormone composed of 51 amino acids, generated from the precursor proinsulin through a series of processing (Rahman et al., 2021). Primarily produced by β cells of the pancreatic islets, insulin is the main anabolic hormone that regulates the energy metabolism throughout the body, i.e., promoting glucose uptake into the liver, fat, and muscle cells (White, 2003). In addition, acting through common receptors (insulin receptor [InsR] and IGF-1 receptor [IGF-1R]), insulin and insulin-like growth factors (IGFs) regulate proliferation and survival of various cell types throughout the body during development, in adulthood, and in aging processes (Nakae et al., 2001; Richardson et al., 2004). InsR and IGF-1R are tyrosine kinase receptors that can be present as homodimers (InsR/InsR, IGF-1R/IGF-1R) or heterodimer (InsR/IGF-1R), sharing the majority of downstream pathways, namely phosphoinositide 3-kinase (PI3K), serine threonine kinase Akt, glycogen synthase kinase 3 β (GSK3 β), and mammalian target of rapamycin (mTOR) (O'Neill et al., 2012). InsR and IGF-1R are highly expressed in the CNS, including the hippocampus and hypothalamus, and central insulin regulates peripheral energy metabolism (O'Neill et al., 2012; Tiedemann et al., 2017; Barrios et al., 2021; Scherer et al., 2021). Furthermore, it is now evident that insulin and IGF-1 play important roles in higher-order brain functions, such as memory and cognitive processing, and in neuroprotection in response to brain injury (Stewart and Rotwein, 1996; Gerozissis, 2003; Fernandez and Torres-Aleman, 2012), indicating pleiotropic roles for insulin/IGF signaling in the CNS.

Insulin crosses the BBB using a saturable transporter, which is regulated by insulin itself and altered by a number of factors including hyperglycemia and diabetes (Banks et al., 2012). In addition, the choroid plexus, a highly vascularized tissue responsible for the production of cerebrospinal fluid (CSF) at the interface of the CNS and periphery, releases insulin and IGFs (Salehi et al., 2009; Ziegler et al., 2012; Mazucanti et al., 2019; Dani et al., 2021). In the parenchyma, microglia generate IGF-1 during development (Włodarczyk et al., 2017) as well as in adulthood (Myhre et al., 2019), while astrocytes can produce insulin, which is negatively regulated by A β and bacterial lipopolysaccharide (LPS) (Takano et al., 2018). InsR and IGF-1R are expressed on neurons and non-neuronal cells (Shaughnessy et al., 2020), and are involved in the regulation of synaptic plasticity (Dyer et al., 2016), astroglial energy metabolism, microglial inflammatory phenotypes (Haas et al., 2020), as well as the self-renewal and maintenance of neural stem cells (Ziegler et al., 2015), indicating crucial roles of insulin through multiple targets and mechanisms in the brain parenchyma (Gabbouj et al., 2019). Although the hypothalamus is the most

well-studied target region regulating systemic energy metabolism (Mitchell and Begg, 2021), specific inactivation of InsR and IGF-1R in the hippocampus and amygdala led to increased anxiety and cognitive impairment in animal models (Soto et al., 2019), indicating the significance of extrahypothalamic and non-metabolic roles for insulin. Moreover, hippocampal microglia responded to insulin treatment in young rats but not in the aged group (Haas et al., 2020), suggesting that microglia develop insulin resistance during aging. Indeed, experimental evidence suggests the link between impaired insulin signaling in the CNS and cognitive impairment. Intracerebral streptozotocin (STZ) induces AD-like brain pathology in mice, which has been widely used as a model for sporadic AD (Lester-Coll et al., 2006). In mixed AD and diabetic mice using APP^{swe}/PS1^{dE9} (APP/PS1) transgenic AD model or triple-transgenic model of AD (3xTg-AD) with either STZ treatment or InsR deficiency (*db/db*), genetic background and dysfunctional insulin signaling cooperatively exacerbated CNS inflammation and AD pathology (Hierro-Bujalance et al., 2020; Imamura et al., 2020; Sankar et al., 2020). In addition, intranasal insulin treatment can alleviate AD pathogenesis and cognitive impairment via reduced neuroinflammation and enhanced neural plasticity (Chen Y. et al., 2014; Guo et al., 2017), possibly via the actions through receptors on microglia and astrocytes (Spielman et al., 2015). Although what roles insulin/IGF signaling play in the CNS could be context-dependent, i.e., either beneficial (Carro et al., 2003; Sukhanov et al., 2007; Tien et al., 2017) or damaging (Labandeira-Garcia et al., 2017), fine-tuning of these signaling pathways is crucial to maintaining brain functions. Considering the anti-inflammatory effects of insulin and IGF-1 on microglia (Labandeira-Garcia et al., 2017; Shaughnessy et al., 2020) and observations that age-related changes of microglia toward pro-inflammatory phenotypes are implicated in brain aging and neurodegenerative disorders (Gemma et al., 2010; Chowen and Garcia-Segura, 2020), microglial insulin/IGF resistance may underlie chronic inflammation in the brain, which is associated with dementia (Lutshumba et al., 2021). Consequently, dysregulated insulin/IGF signaling and insulin resistance in the CNS have been linked to an increased risk of dementia, including AD, which is sometimes referred to as type 3 diabetes (Biessels et al., 2006; Whitmer et al., 2008; Zemva and Schubert, 2011; Banks et al., 2012; Smolina et al., 2015; Biessels and Despa, 2018; Kuo et al., 2018; Shinjo et al., 2020).

Insulin as an Immunomodulator – A Possible Link Between Metabolic Syndromes and Infection Susceptibility

Acute and chronic infections induce insulin resistance (Yki-Järvinen et al., 1989; Fernández-Real et al., 2006), whereas metabolic imbalance (e.g., higher body fat mass and diabetes) is associated with higher infection susceptibility (Rayfield et al., 1982; Fernández-Real et al., 2007), indicating bidirectional interactions between the immune system and energy metabolism in human body. Chronic low-grade inflammation and immune dysregulation likely mediate the mutual and possibly synergetic relationship. The strong interaction between the immune and

metabolic pathways is apparently rooted in their common evolutionary origin, as represented by the fat body in *Drosophila*, which senses both infectious and metabolic stresses and perform the functions of the liver, adipose tissue, and immune system (Hotamisligil, 2017). Insulin plays a key role in such evolutionarily conserved immunometabolism, partly through the interactions with tumor necrosis factor (TNF) receptor (Uysal et al., 1997) and Toll-like receptor (TLR) pathways (DiAngelo et al., 2009; Hotamisligil, 2017), as well as through modulating metabolic pathways in immune cells (van Niekerk et al., 2020). Through metabolic regulation, insulin alleviates the harmful effects of hyperglycemia (Sun et al., 2014). For example, due to its glucose-lowering effect, insulin exerts anti-inflammatory effects through modulating the release of inflammatory mediators. As glucose is pro-inflammatory, insulin deficiency activates inflammatory reactions in the body, leading to the release of inflammatory mediators including reactive oxygen species and pro-inflammatory cytokines from leukocytes (Mohanty et al., 2000; Esposito et al., 2002). In addition, insulin directly activates phagocytic and bactericidal activity of immune cells and diabetes-induced infection susceptibility is partly mediated by impaired immune responses due to the lack of insulin signaling, as shown in rodent studies (Yano et al., 2012).

Leptin

Leptin as a Metabolic Hormone

Leptin is a 16 kDa polypeptide that regulates metabolic balance and fat storage. Mainly produced by the white adipose tissue (WAT), leptin acts via leptin receptor (LepR) in the brain and plays a pivotal role in the control of appetite and energy expenditure. There are six LepRs (LepRa to LepRf), with identical extracellular N-terminal domain and distinct intracellular C-terminal regions generated by alternative splicing of *db* (Lee et al., 1996). LepRb, the long isoform with high ligand affinity, is the major isoform expressed in the brain and activates intracellular pathways, including JAK/STAT, ERK/MAPK, and IRS/PI3K (Allison and Myers, 2014). The hypothalamic nuclei, the regulatory center of energy homeostasis, are highly enriched with LepRb (Elmquist et al., 1998; Balthasar et al., 2004; Leshan et al., 2009), and leptin exerts its effects through the action in the hypothalamus to regulate food intake and energy metabolism (Friedman, 2019). Leptin enters the hypothalamus through fenestrated capillary and acts on LepR expressed on neurons in arcuate nucleus (ARC), dorsomedial hypothalamus (DMH), and ventromedial hypothalamus (VMH), enabling a feedback mechanism to maintain energy balance, thereby preventing obesity and metabolic disorders (Pandit et al., 2017). In addition, leptin can cross BBB using a saturable transport system (Banks and Farrell, 2003), as well as the blood-CSF barrier (choroid plexus epithelia) (Merino et al., 2006; Dietrich et al., 2008). LepRb in extrahypothalamic brain regions also plays significant roles in the regulation of energy metabolism (Scott et al., 2009). For example, LepRb in the ventral tegmental area (VTA) regulates energy balance via mesolimbic dopaminergic system (Fulton et al., 2006; Hommel et al., 2006), suggesting that leptin targets multiple brain regions and cellular components. Consequently, dysregulation of leptin signaling results in obesity, diabetes, and

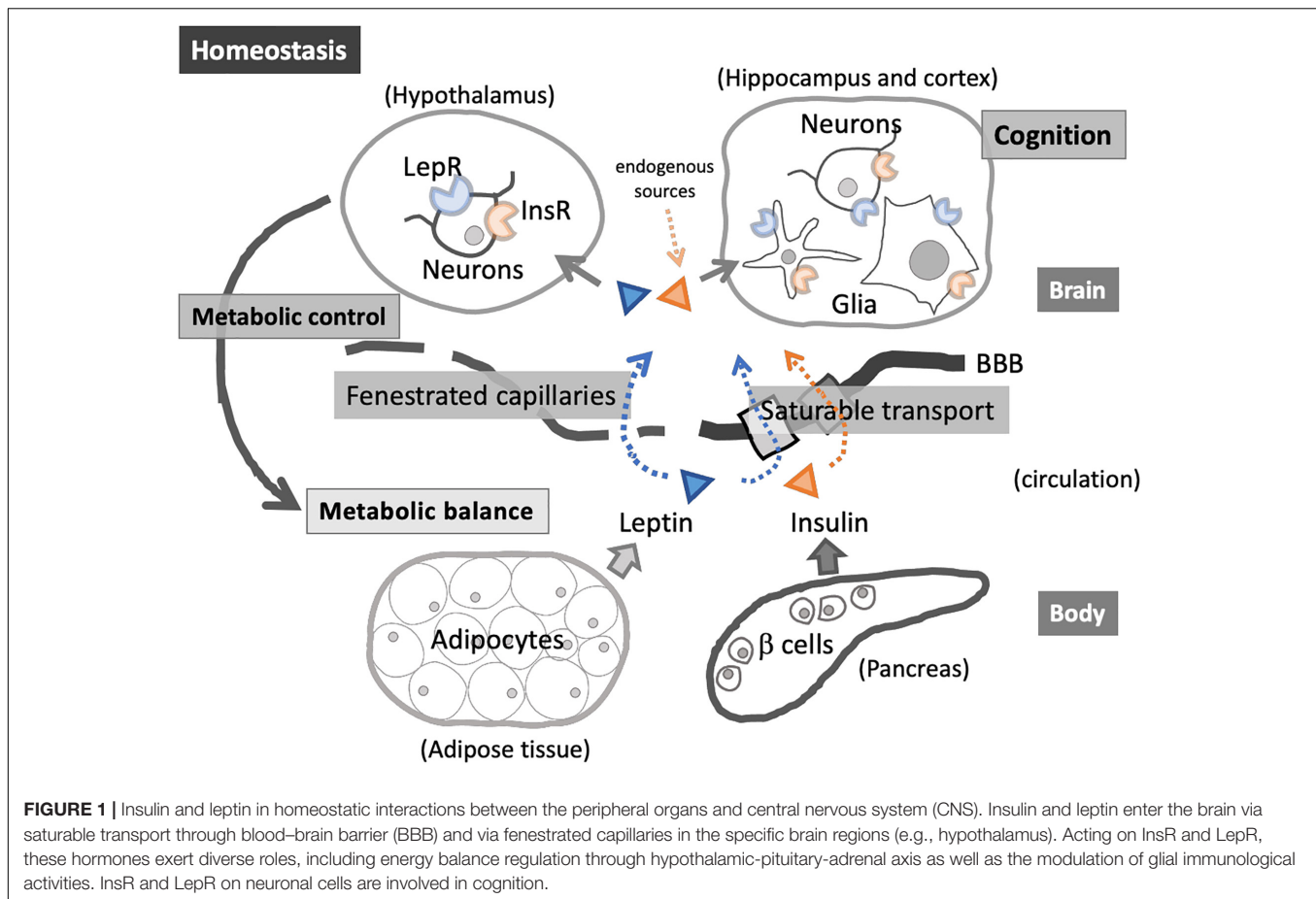


FIGURE 1 | Insulin and leptin in homeostatic interactions between the peripheral organs and central nervous system (CNS). Insulin and leptin enter the brain via saturable transport through blood–brain barrier (BBB) and via fenestrated capillaries in the specific brain regions (e.g., hypothalamus). Acting on InsR and LepR, these hormones exert diverse roles, including energy balance regulation through hypothalamic–pituitary–adrenal axis as well as the modulation of glial immunological activities. InsR and LepR on neuronal cells are involved in cognition.

associated comorbidities (Myers et al., 2008; Wasim et al., 2016; Fischer et al., 2020). Leptin resistance is the condition where diminished leptin sensitivity occurs, resulting in a defect in satiety detection despite high leptin levels, which has been linked to obesity (Izquierdo et al., 2019). Leptin deficient (*ob/ob*) and LepR deficient (*db/db*) mice, carrying mutations in leptin (*ob*) and LepR (*db*) genes, respectively, exhibit excessive eating, develop obesity and diabetes, and are widely used as animal models of T2DM (Chen et al., 1996; Ninomiya et al., 2002; Gautron and Elmquist, 2011). Growing evidence suggests the contribution of leptin resistance to neurodegeneration in AD (Bonda et al., 2014). Serum leptin levels showed negative correlation with cognitive decline in the elderly (Holden et al., 2009). In the brain autopsy, CSF leptin levels were significantly higher in AD compared to control and mild cognitive impairment cases, and CSF leptin concentration was correlated with pathological neurofibrillary tangle burden (Bonda et al., 2014), suggesting that leptin resistance develops during AD progression.

Leptin as an Adipokine

Adipokines are the adipose tissue-derived factors that affect whole body homeostasis in autocrine and paracrine manners, targeting a number of biological processes such as glucose metabolism, lipid metabolism, insulin sensitivity, as well as immune response (Fasshauer and Blüher, 2015). Adipokines

include leptin, adiponectin, vaspin, fibroblast growth factor 21 (FGF21), and many more, each exerting specific biological effects, and mediate diverse actions throughout the body (Fasshauer and Blüher, 2015). Originally identified as an adipocyte-derived hormone that regulates neuroendocrine axis, leptin is one of the most studied adipokines linking the immune system and energy metabolism (Abella et al., 2017; Jiménez-Cortegana et al., 2021).

Indeed, leptin belongs to the family of long-chain helical cytokines and has similarity to IL-6, IL-12, and granulocyte colony-stimulating factor (G-CSF). LepR is a type I cytokine receptor (La Cava and Matarese, 2004). Although neurons are the most well-established cellular targets of leptin, LepR is also expressed by non-neuronal cells throughout the body. Importantly, most immune cells, including hematopoietic bone-marrow precursors, monocytes/macrophages, and lymphocytes, express LepR, suggesting that leptin directly modulate immune responses and inflammation (La Cava and Matarese, 2004; Procaccini et al., 2012, 2017). Adipose tissue and lymphoid organs are often anatomically associated, and the contiguity between adipocytes and lymphoid cells supports the functional interactions (Matarese et al., 2002). For example, leptin affects thymic function and growth/survival of bone-marrow CD34⁺ precursors and CD4⁺ T cells (Lord et al., 1998; Howard et al., 1999; Martín-Romero and Sánchez-Margalet, 2001; Papanthassoglou et al., 2006; Cohen et al., 2017), and

modulates both innate and adaptive immunity through diverse mechanisms (Maurya et al., 2018), such as enhancement of neutrophil oxidative burst (Mancuso et al., 2002; Caldefie-Chezet et al., 2003), phagocytosis by monocytes/macrophages (Faggioni et al., 1999; Fantuzzi and Faggioni, 2000; Sánchez-Margalet et al., 2003), and cytotoxic activity of natural killer (NK) cells (Tian et al., 2002), as well as mobilization of macrophages, lymphocytes (Abella et al., 2017), and neutrophils (Souza-Almeida et al., 2018).

Leptin deficiency and resistance are associated with increased susceptibility to infectious diseases, including bacterial (e.g., *Mycobacterium tuberculosis* and *Streptococcus pneumoniae*), viral [e.g., coronaviruses, influenza A virus, and human immunodeficiency virus (HIV)], and parasitic (e.g., *Trypanosoma brucei*, *Trypanosoma cruzi*, and *Entamoeba histolytica*) infections (Amole et al., 1985; Sánchez-Pozo et al., 2003; Wieland et al., 2005; Hsu et al., 2007; Nagajyothi et al., 2010; Tschöp et al., 2010; Vedantama and Viswanathan, 2012; Zhang et al., 2013; Radigan et al., 2014; Altı et al., 2018; Guglielmi et al., 2021), while infection with certain pathogens, such as *Plasmodium* spp. and *Toxoplasma gondii*, can cause dysregulated leptin secretion, independently of adiposity (Pulido-Mendez et al., 2002; Baltaci and Mogulkoc, 2012). The significant association between obesity-induced inflammation (meta-inflammation) and the severity of infectious disease has been highlighted by COVID-19 pandemic (Huizinga et al., 2020; Rebello et al., 2020). It is notable that *Streptococcus pneumoniae* is a major cause of meningitis, which potentially leads to persistent cognitive disability (Yau et al., 2018), and *M. tuberculosis* can disseminate into the brain and induce CNS tuberculosis, a cascade of inflammatory responses that can potentially cause brain damage (Leonard, 2017; Davis et al., 2019). HIV-positive individuals frequently suffer neurocognitive disorders (HIV-associated neurocognitive disorders, HAND), and influenza A virus (H1N1) have been associated with neurological manifestation in both young and adult patients (Cárdenas et al., 2014; Wilking et al., 2014), with some developing permanent sequelae (Cárdenas et al., 2014), suggesting that defective leptin signaling could affect brain functions through increased infection susceptibility. While the mechanistic link between leptin resistance and infection susceptibility is complex and multifactorial (Maurya et al., 2018; Rebello et al., 2020), leptin's actions through immune cells in the periphery and CNS, and dysregulation thereof, likely play a significant role.

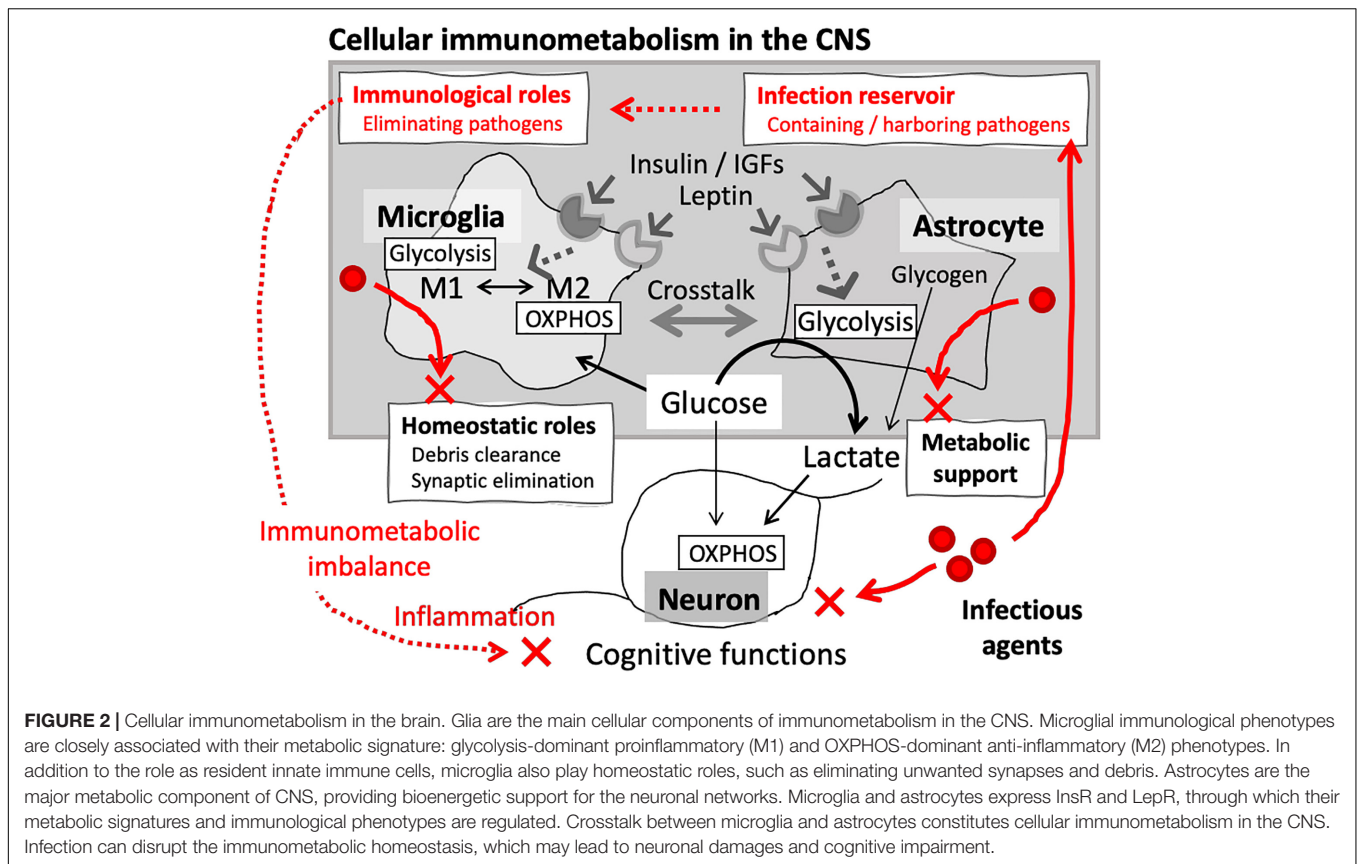
The Roles for Leptin and Insulin in Cellular Immunometabolism

Cellular energy metabolism is mainly driven by glycolysis, tricarboxylic acid (TCA) cycle, fatty acid oxidation, and oxidative phosphorylation (OXPHOS). Glutaminolysis, the conversion of glutamine to glutamate, is activated to fuel TCA cycle when glucose availability is limited. Glutaminolysis also plays a crucial role in the brain, where glutamate functions as a major neurotransmitter (Wang et al., 2017). In addition, in the absence of glucose, such as insulin-induced hypoglycemia, microglia utilize glutamine as an alternative fuel to support

their immunological functions (Bernier et al., 2020). Upon stimulation by pathogen-derived molecules and endogenous ligands, immune cells undergo metabolic reprogramming into alternative modes of energy metabolism (O'Neill et al., 2016; Gaber et al., 2017; Hotamisligil, 2017; Próchnicki and Latz, 2017), which can be largely classified into the pro-inflammatory phenotypes dominated by glycolysis (similar to the Warburg effect in cancer cells), and anti-inflammatory phenotypes characterized by TCA cycle, fatty acid oxidation, and OXPHOS (O'Neill et al., 2016). Such bioenergetic shifts determine the properties of various immune cell populations, including macrophages (Van den Bossche et al., 2017), neutrophils (Curi et al., 2020), and T cells (O'Neill et al., 2016; Spadaro et al., 2017; Balyan et al., 2020), where insulin/IGFs and leptin come into play. For example, insulin regulates T cell's metabolic reprogramming, thereby shaping adaptive immunity (Tsai et al., 2018). InsR-deficient T cells showed compromised responses to antigens *in vitro*, and T cell-specific InsR knockout in mice led to reduced antigen-specific immunity to influenza virus infection *in vivo* (Tsai et al., 2018), suggesting that InsR signaling reinforces metabolic reprogramming required for T cell activation. Such immunometabolic changes of T cells are likely mediated by mTOR, a key regulator of cellular homeostasis including protein synthesis and autophagy (Rao et al., 2010; Chi, 2012; Martin et al., 2021). mTOR also mediates insulin-induced alteration of metabolic rates and immune responses in myeloid cells (Ratter et al., 2021), and mTOR mediates age-associated microglial priming and neurodegeneration (Keane et al., 2021), suggesting that insulin-induced activation of mTOR pathway plays a role in immunometabolic imbalance in the CNS. IGF-1R signaling is essential for the anti-inflammatory polarization of macrophages upon metabolic stress as well as helminth infection (Labandeira-Garcia et al., 2017; Spadaro et al., 2017), and IGF-2 instructs macrophage precursor cells to become anti-inflammatory through metabolic pre-programming toward OXPHOS (Du et al., 2019). Leptin also induces immunometabolic changes in immune cells, including macrophages and T cells (Cohen et al., 2017; Boutens et al., 2018; Monteiro et al., 2019). Activation of leptin signaling pathways (JAK/STAT and IRS/PI3K) leads to intracellular metabolic changes, such as increased glucose uptake and glycolytic activity and reduced OXPHOS, associated with pro-inflammatory phenotype of macrophages and T cells (Cohen et al., 2017; Boutens et al., 2018; Monteiro et al., 2019). It is conceivable that immunometabolic imbalance due to dysregulated insulin/IGF and leptin signaling pathways underlie immunodeficiency and infection susceptibility in nutritional imbalance, including malnutrition and obesity.

Brain-Resident Immune Cells and Immunometabolism Microglia and Astrocytes

The brain is the most energy-demanding organ, consuming glucose at a disproportionately high rate compared to the rest of the body. Glucose metabolism in the brain is founded on intricate interactions between the cellular components, i.e., neurons and glia. Neurons are highly aerobic and heavily



dependent on OXPHOS, while astrocytes and oligodendrocytes, the major neuroglial populations, are predominantly glycolytic. These cell-type specific metabolic profiles support the overall power system in the CNS: lactate generation via glycolysis in neuroglia fueling OXPHOS in neurons (Fünfschilling et al., 2012; Jha and Morrison, 2018; Rosko et al., 2019) (Figure 2). To protect such elaborate systems, the brain harbors resident innate immune cells, namely resident macrophage populations: microglia, perivascular macrophages, meningeal macrophages, and choroid plexus macrophages. The former three, including microglia, have embryonic origins and are maintained through local self-renewal while choroid plexus macrophages are replenished via constant supply of bone marrow-derived monocytes (Prinz et al., 2017). These brain-resident macrophage populations play significant roles in the brain physiology, as well as in defense against infection (Ransohoff and Brown, 2012; Heneka et al., 2015; Herz et al., 2017; Kierdorf and Prinz, 2017).

While aberrant activation of resident macrophages contributes to CNS pathology via inflammation (De Strooper and Karran, 2016; Herz et al., 2017), evidence is growing that brain-resident macrophages play homeostatic roles in the brain. Residing in the brain parenchyma, microglia play pivotal roles in the development and maintenance of neuronal networks. The most notable roles of microglia include complement-dependent synaptic pruning during CNS development (Schafer et al., 2012) and synaptic reorganization throughout life (Herz et al., 2017). In addition, microglia-derived neurotrophic factors, such

as brain-derived neurotrophic factor (BDNF), promote learning-dependent synaptic formation (Parkhurst et al., 2013) and protect neurons from brain injuries (Madinier et al., 2009). Furthermore, CNS macrophages are the major component of glymphatic system, which plays a crucial role in the clearance of waste products and toxic materials, including A β (Gordleeva et al., 2020). In the context of AD pathogenesis, microglial activation is a double-edged sword; it can facilitate the clearance of A β and tau, while potentially inducing neuroinflammation leading to neuronal damages associated with AD (Lue et al., 2010; Leyns and Holtzman, 2017; Shippy and Ulland, 2020).

Meanwhile, growing evidence suggests that astrocytes are part of the innate immunity in the brain (Sofroniew, 2020). Responding to the changing microenvironment and diverse stimuli (e.g., microbial infections, exposure to environmental toxins, tumor formation, and neurodegenerative diseases), astrocytes undergo morphological and functional changes and influence the outcome of a number of brain disorders (Drögemüller et al., 2008; Soung and Klein, 2018; Katsouri et al., 2020; Sofroniew, 2020). In addition, crosstalk between astrocytes and microglia in the hippocampus is responsible for sensitivity to insults (Lana et al., 2020), pointing to the importance of communication between astroglia and brain-resident macrophages in cognitive dysfunctions (Liddelov et al., 2017; Linnerbauer et al., 2020). Furthermore, astrocytes and microglia participate in A β clearance and protection of synaptic connectivity in AD models, suggesting highly context-dependent

immunological roles of these glial cell populations against AD development (Davis et al., 2021).

Glial Cells and Central Nervous System Immunometabolism

Microglia and astrocytes, also called macroglia, are major resident non-neuronal cells in the brain. As described above, evidence suggests that the interplay between neurons and these glial population plays pivotal roles in the brain, including the hippocampus, in physiological and pathological conditions (Lana et al., 2021). While neuronal cells express InsR/IGF-1R at high levels and many studies have focused on the roles of neuronal InsR/IGF-1R (Frölich et al., 1998), microglia and astrocytes also express InsR and IGF-1R (Shaughnessy et al., 2020). Insulin and IGFs support brain functions through regulating astroglial glucose metabolism (Sonnewald et al., 1996; Fernandez et al., 2018). InsR signaling modulates astroglial glucose uptake and bioenergetics (Heni et al., 2011), and specific ablation of astroglial InsR led to altered mood and cognition in mice (González-García et al., 2021). In addition, insulin can modulate inflammatory responses of astrocytes (Spielman et al., 2015), suggesting the immunometabolic regulation of astrocytes by insulin. IGF-1R can also modulate astroglial metabolic and immunological signatures (Hernandez-Garzón et al., 2016), and astroglial IGF-1R signaling mediates synaptic plasticity of cortical inhibitory neurons (Noriega-Prieto et al., 2021). These data suggest that impairment in InsR/IGF-1R signaling pathways may disrupt the metabolic network between glia and neurons, as well as immunological roles of astrocytes, eventually leading CNS dysfunctions. Microglial InsR/IGF-1R also mediate the effects of insulin. Low-dose insulin exerted pro-inflammatory effects on microglial cells *in vitro* (Spielman et al., 2015), while it was anti-inflammatory at a higher concentration (Brabazon et al., 2018), suggesting the variable roles of microglial InsR/IGF-1R signaling. In addition, microglia are the major source of IGF-1 in the brain, and microglial IGF-1 was found to be increased in AD model mice compared to wild-type (Myhre et al., 2019). These data suggest that insulin and IGFs play crucial roles in immunometabolism via the phenotypic regulation of glial cells, potentially mediating the link between bioenergetics and immunity in the brain.

Microglia (Tang et al., 2007; André et al., 2017; Fujita and Yamashita, 2019), and astrocytes (Naranjo et al., 2020; Pratap and Holsinger, 2020) express LepR. Leptin can enhance microglial pro-inflammatory responses, including IL-6 production through a mechanism involving insulin receptor substrate-1 (IRS-1), PI3K, and Akt (Tang et al., 2007; André et al., 2017; Fujita and Yamashita, 2019), IL-1 β release via a caspase 1-independent mechanism (Pinteaux et al., 2007), and lipopolysaccharide (LPS)-induced pro-inflammatory responses (Lafrance et al., 2010). On the other hand, in a spinal cord injury model, leptin reduced microglial inflammatory responses, while inducing neuroprotective phenotypes, Fernández-Martos et al. (2012). In myeloid cell-specific LepR deficient mice, hypothalamic microglia exhibited less ramified morphology and impaired phagocytic capacity (Gao et al., 2018), suggesting that leptin directly regulates homeostatic microglial phenotypes

(Davis et al., 2017). Considering multifaceted physiological roles of microglia and potential harm (i.e., neuroinflammation) caused by their aberrant activation (Colonna and Butovsky, 2017; Li and Barres, 2018), it is conceivable that impaired leptin signaling in microglia could lead to significant homeostatic imbalance in the CNS. On the other hand, specific LepR depletion in astrocytes (GFAP-LepR $^{-/-}$) resulted in impaired neurotransmission in the hippocampus, suggesting that leptin regulates hippocampal plasticity via astroglial LepR, possibly by regulating the glucose and glutamate up-take capacity (Naranjo et al., 2020).

Other Central Nervous System-Resident Immune Cells

In addition to resident macrophages, the CNS harbors multiple leukocyte populations, including dendritic cells (DCs) and T lymphocytes. As a professional antigen-presenting cells, DCs play an essential role in the regulation of adaptive immunity. Although initially believed to be absent in the healthy brain parenchyma, only appearing in response to aging, injury, and infections (Fischer and Reichmann, 2001), studies have shown the presence of DCs within the healthy steady-state brain (Bulloch et al., 2008; Proding et al., 2011). Upon activation, brain DCs can migrate out of the brain and induce T-cell homing into the CNS where antigen-specific immune responses may take place (Karman et al., 2004). Correlation between aging and the accumulation of DCs in the CNS (Kaunzner et al., 2012) suggests potential involvement of brain DCs in immunological changes and inflammation associated with aging. In response to immunological stimuli, such as viral and parasitic infections, T cells infiltrate into the brain, where these cells play beneficial roles by eliminating pathogens. In addition, studies have shown that long-lived memory T cells are established after CNS infections (Wakim et al., 2012; Landrith et al., 2017; Mockus et al., 2018), and these tissue-resident memory CD8 cells can provide frontline defense against re-infection (Netherby-Winslow et al., 2021). However, persistence of these cells can also cause neuronal damages (Ai and Klein, 2020; Ghazanfari et al., 2021).

Although it is unknown how these CNS resident immune cells respond to and regulated by leptin and insulin, InsR is expressed on the surface of activated T cells (Helderman et al., 1978). Acting through InsR, insulin enhances and maintains T cell functions after immunological challenges (Helderman, 1984) and potentially induces anti-inflammatory polarization in the periphery (Viardot et al., 2007). These data suggest that insulin may play an immunomodulatory role via actions on T cell populations in the brain.

Glia as the Target and Reservoir of Infectious Agents

Growing evidence supports the infectious etiology of dementia, and a number of causative agents have been proposed, including viruses (e.g., Herpes simplex virus type 1 (HSV-1), cytomegalovirus (CMV)) and bacteria (e.g., *Chlamydomphila pneumoniae*, spirochetes, *Helicobacter pylori*, *Porphyromonas gingivalis*) (Harris and Harris, 2015; Sochocka et al., 2017; Nazareth et al., 2021). Suggested mechanisms include persistent inflammation caused by chronic infection, leading to a vicious cycle of neuroinflammation and neurodegeneration, as well

as BBB disruption, which may further enhance the entry of infectious agents and proinflammatory mediators from the periphery (Sochocka et al., 2017) (**Figure 3**).

Some infectious agents can enter and persistently inhabit the brain, directly causing chronic activation of resident immune cells (Sochocka et al., 2017). For example, *Chlamydia pneumoniae*, obligate intracellular bacteria, are frequently found in microglia and astrocytes in the brain of AD patients (Balin et al., 1998; Nazareth et al., 2021). Microglia and astrocytes are also the target of some viruses, such as CMV (Cheeran et al., 2001) and HIV (Avalos et al., 2017; Valdebenito et al., 2021). In addition, *T. gondii*, a neurotropic protozoan parasite causing a lifelong CNS infection, has been implicated in various neurological disorders, including dementia (Nayeri Chegeni et al., 2019). *T. gondii* is an obligate intracellular parasite, potentially infecting all nucleated cell types. Chronic *Toxoplasma* infection is linked to microglial activation and persistent neuroinflammation (Li et al., 2019), and CNS energy metabolism is significantly skewed toward glycolysis upon infection (Hargrave et al., 2019), suggesting that *T. gondii* interferes with host immunometabolism in the brain. Indeed, it has been shown that *Toxoplasma* infection may lead to reduced IGF-1 signaling in the brain (El Saftawy et al., 2020). On the other hand, *In vitro*, high glucose and up-regulation of glycolysis in the host cell had a significant impact on the life-stage conversion of *T. gondii* (i.e., from fast-growing tachyzoite stage into dormant bradyzoite stage) (Weilhammer et al., 2012), indicating that host cell metabolism could modulate the virulence of intracellular pathogens. Intriguingly, *T. gondii* infection can enhance A β clearance by increasing phagocytic activity of recruited monocytes (Möhle et al., 2016), suggesting that the ultimate outcome may depend on the extent and the aggressiveness of the infection. Of note, SARS-CoV-2 is frequently found in astroglia and microglia, rather than neurons, in the autopsy brain, indicating potential involvement of glial activation in the neurological manifestation of COVID-19 (Solomon, 2021). These data suggest that infectious agents could produce immunometabolic imbalance in the CNS via affecting microglial and astroglial phenotypes (**Figure 2**), which may lead to chronic inflammation and an increased risk of neurodegenerative disorders (Stephenson et al., 2018).

INSULIN AND LEPTIN MEDIATING THE LINK BETWEEN INFECTION AND NEURODEGENERATIVE DISORDERS

Infectious Etiology of Dementia

Recently, it has been shown that A β , a hallmark of AD, is an antimicrobial peptide, part of innate immunity to protect the host from various infectious agents (Soscia et al., 2010; Kumar et al., 2016; Gosztyła et al., 2018), suggesting the necessity to revisit the role of amyloid plaque formation in AD pathogenesis. Above mentioned neurotropic viruses, such as HSV-1, HSV-2 and CMV, certain bacterial species, including spirochetes and *P. gingivalis*, fungi (e.g., *Candida albicans*), and neurotropic parasite (e.g.,

T. gondii), may cause disruption of brain functions via chronic inflammation and immune dysregulation in the brain (Sochocka et al., 2017; Shinjyo et al., 2021). The antimicrobial hypothesis for AD proposes that A β generation and amyloid plaque formation are not the major culprit in AD pathogenesis, but rather a defense against such infectious agents (Moir et al., 2018; Iqbal et al., 2020; Fulop et al., 2021).

While metabolic disorders and infection may independently increase the risk of dementia, obesity and metabolic disorders may indirectly increase the risk of infection-induced neurological disorders, by enhanced susceptibility to infection and infection-induced complications via immune dysregulation (Shah and Hux, 2003; Muller et al., 2005; Knapp, 2013; Tamara and Tahapary, 2020). Conversely, obesity can occur as a result of infection (Pasarica and Dhurandhar, 2007; Dhurandhar, 2011), and infection-induced chronic low-grade inflammation may lead to insulin resistance (Fernández-Real et al., 2006), suggesting bi-directional interactions between infection and metabolic disorders. Furthermore, several pathogens, such as HSV, CMV, *H. pylori*, and *P. gingivalis*, are implicated in both metabolic disorders and AD (Jeon et al., 2012; Harris and Harris, 2015; Fleck-Derderian et al., 2017; Beydoun et al., 2018; Lövheim et al., 2018; Mei et al., 2020; Baradaran et al., 2021; Costa et al., 2021; Salem et al., 2021), supporting the potential mechanistic link between infection, MetS, and dementia etiology.

Metabolic Imbalance Causing Central Nervous System Infection Susceptibility

Studies showing possible interactions between metabolic imbalance and infection in dementia etiology, particularly focusing on data highlighting the roles for insulin and leptin, are listed in **Table 1**.

West Nile Virus

West Nile virus (WNV) is a single-stranded RNA virus, genetically related to the Japanese encephalitis virus (JEV). Transmitted by mosquitoes, WNV potentially causes life-threatening encephalitis or meningitis, especially in the elderly (Alli et al., 2021). In addition, it can cause persistent cognitive impairments (Murray et al., 2014; Vittor et al., 2020). While CNS infiltration of lymphocytes, including CD8⁺ T cells, is essential in eliminating viruses, persistent inflammation can cause synaptic loss and neuronal death, impairing cognitive ability (Garber et al., 2019; Vittor et al., 2020). Of note, diabetes is a frequent comorbidity of severe WNV diseases and considered a risk factor for developing WNV encephalitis (Badawi et al., 2018). In *db/db* obesity model, WNV infection-induced leukocyte infiltration into the brain was significantly lower, suggesting that obesity compromises protection against viral infection in the brain (Kumar et al., 2014). In particular, infiltration of CD8⁺ T cells was significantly reduced in obese mice, which was associated with higher viral load and enhanced inflammatory responses in the brain (Kumar et al., 2014), suggesting a role of leptin signaling in protecting the brain via leukocyte recruitment (Rummel et al., 2010).

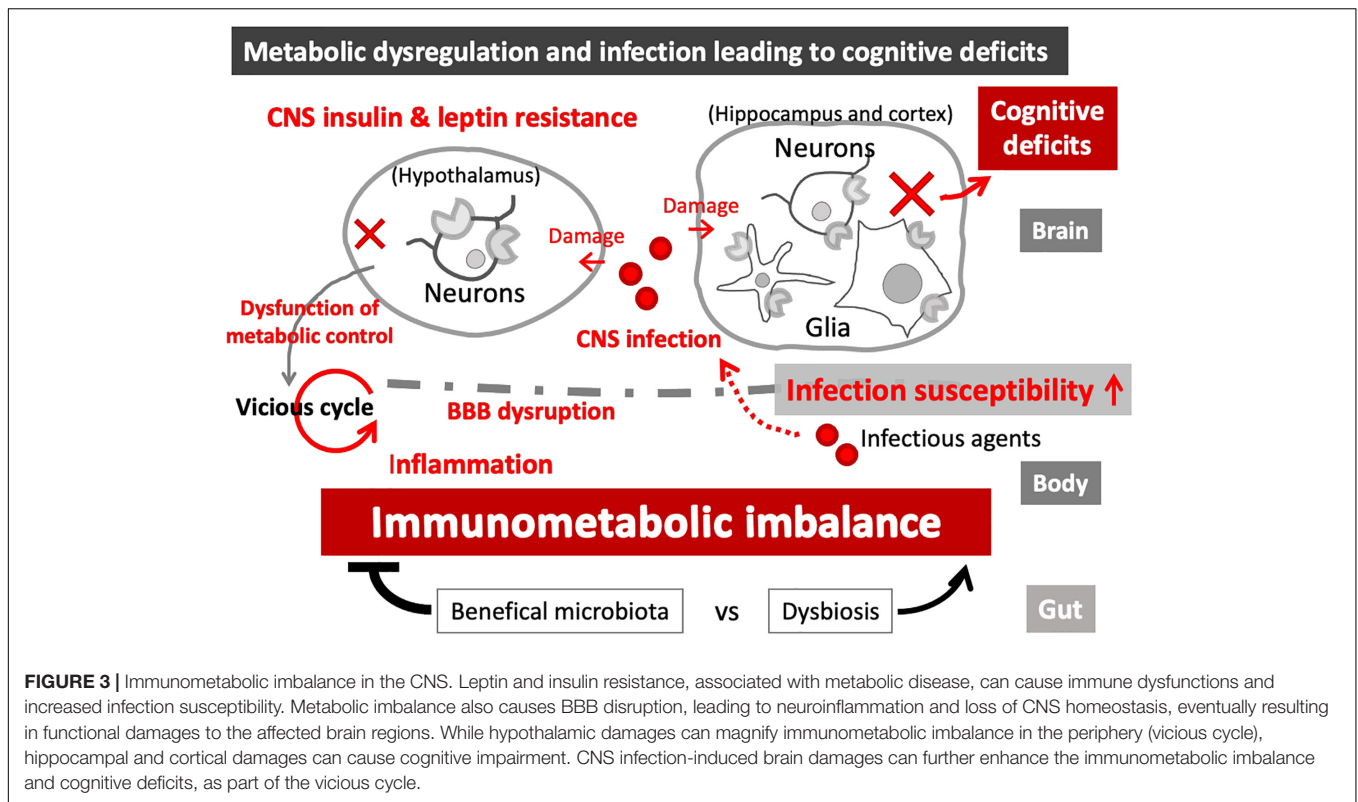


FIGURE 3 | Immunometabolic imbalance in the CNS. Leptin and insulin resistance, associated with metabolic disease, can cause immune dysfunctions and increased infection susceptibility. Metabolic imbalance also causes BBB disruption, leading to neuroinflammation and loss of CNS homeostasis, eventually resulting in functional damages to the affected brain regions. While hypothalamic damages can magnify immunometabolic imbalance in the periphery (vicious cycle), hippocampal and cortical damages can cause cognitive impairment. CNS infection-induced brain damages can further enhance the immunometabolic imbalance and cognitive deficits, as part of the vicious cycle.

Porphyromonas gingivalis

Porphyromonas gingivalis is a periodontal disease-causing Gram-negative bacteria found in the oral cavity. Evidence suggests a strong association between *P. gingivalis* infection and sporadic AD (Kanagasingam et al., 2020), and its presence has been identified in the brain of AD patients (Dominy et al., 2019). *P. gingivalis* infection can trigger inflammation both in the periphery and CNS in affected individuals, leading to cognitive decline (Dominy et al., 2019; Kanagasingam et al., 2020). In mice, oral *P. gingivalis* infection led to brain colonization and AD-like pathogenesis, including complement activation and A β_{1-42} formation (Poole et al., 2015; Dominy et al., 2019), suggesting potential mechanistic links between periodontal disease and AD. A systematic review of pre-clinical studies also found that *P. gingivalis* infection induced inflammatory responses and tissue degeneration in the brain, which were associated with cognitive impairment (Costa et al., 2021).

Periodontitis is also associated with obesity (Suvan et al., 2011; Keller et al., 2015; Nascimento et al., 2015; Khan et al., 2018). High fat diet (HFD) significantly enhanced systemic inflammation induced by periodontal pathogens in rodents (Virto et al., 2018). *P. gingivalis* infection up-regulated the expression of genes associated with insulin/IGF-1 signaling and induced inflammatory responses in the brain of *db/db* mice (Virto et al., 2018), suggesting that metabolic imbalance due to dysregulated insulin and leptin signaling pathways may exacerbate the outcome of periodontitis and associated inflammation in the brain.

Human Immunodeficiency Virus

Despite the effectiveness of antiretroviral therapy in saving the lives of many from acquired immunodeficiency syndrome (AIDS), HIV-positive individuals frequently suffer neurocognitive disorders (HAND) (Antinori et al., 2007; Foley et al., 2011). While inflammation and brain atrophy due to persistent viral presence in the CNS may partly provide explanations (Pemberton and Brew, 2001; Woods et al., 2010; Ances et al., 2012; Brew and Barnes, 2019), exact mechanisms underlying HAND remain unclear.

Metabolic syndromes, including diabetes, is prevalent in HIV-infected individuals (Calza et al., 2011; Paik and Kotler, 2011), and insulin resistance was associated with lower cognitive scores in a HIV-1 cohort (Valcour et al., 2005, 2006), suggesting that HIV infection may increase the risk of cognitive impairment via metabolic dysregulation. In fact, in a murine model of HAND, intranasal insulin administration restored hippocampal dendritic integrity and cognitive functions, independently of HIV burden in the brain (Kim et al., 2019). In primary human neurons and microglia *in vitro*, insulin suppressed infection-induced inflammatory responses and HIV-1 growth in microglia, and prevented infection-induced neuronal death (Mamik et al., 2016). In addition, in a feline HIV model *in vivo*, intranasal insulin enhanced the preservation of cortical neurons and improved cognitive performance (Mamik et al., 2016), suggesting that impaired insulin signaling in the CNS may underlie cognitive impairment in HIV positive individuals.

TABLE 1 | The roles of insulin and leptin in central nervous system (CNS) infection.

Pathogen/host	Study design and outcome measures	Findings	Proposed direction of causality and/or mechanism	References
Borna disease virus (BDV)				
BDV/mouse	<u>Animal model</u> : virus-induced obesity model using intracerebral injection of BDV-1 in rats. <u>Outcome measures</u> : neuropathology including inflammatory infiltrates and neurodegeneration in the hypothalamus, hippocampal shrinkage, and leptin resistance	BDV infection-induced obesity was associated with: 1. hypothalamic inflammation. 2. Hippocampal involution and microglial activation in the neocortex. 3. Preferential infection of glutamatergic sites, while sparing GABAergic areas, causing (anabolic appetite-stimulating) GABAergic predominance and fat accumulation 4. Leptin resistance in the brain.	CNS infection → Hypothalamic inflammation, neurotransmitter imbalance (GABA predominance), obesity, and leptin resistance.	Gosztonyi et al., 2020
BDV/rat	<u>Animal model</u> : BDV infection models using two different strains: <i>BDV-ob</i> (obesity-inducing) and <i>BDV-bi</i> (no obesity-inducing effect). <u>Outcome measures</u> : Mononuclear infiltrates into the brain, astrogliosis, and neuronal death.	1. <i>BDV-ob</i> infection and mononuclear infiltrates were restricted to certain brain areas including hypothalamus, hippocampus, and amygdala. Particularly severe infiltration in the median eminence of hypothalamus. (<i>BDV-ob</i> infection was observed evenly throughout the brain.) 2. Mononuclear infiltrates, astrogliosis, and neuronal death in the hippocampus of <i>BDV-ob</i> infected brain.	CNS infection → Hypothalamic inflammation and disruption of neuroendocrine system → Obesity	Herden et al., 2000
Canine distemper virus (CDV)				
CDV/mouse	<u>Metabolic disturbance</u> : obesity induced by intracerebral CDV infection. <u>Outcome measures</u> : Plasma insulin and lipid composition.	Hyperinsulinemia and triglyceride accumulation in CDV-induced obesity mice	CDV infection → Hyperinsulinemia and obesity	Bernard et al., 1988
CDV/mouse	<u>Animal model</u> : obesity induced by intracerebral infection with CDV. <u>Outcome measures</u> : Leptin and LepR expression.	Functional LepR was specifically downregulated in the hypothalamus of obese mice.	CNS infection → Leptin resistance in the hypothalamus. → Obesity.	Bernard et al., 1999
Human immunodeficiency virus (HIV)				
HIV/human	<u>Subjects</u> : patients with HIV-1 infection (n203, Cohort study). <u>Outcome measures</u> : association between HIV-associated dementia (HAND) and diabetes .	HAND was significantly associated with diabetes (odds ratio 5.43, 1.66–17.70), which was not fully explained by age or coexisting vascular risk factors.	Diabetes ↔ HAND	Valcour et al., 2005
HIV/mouse	<u>Animal model</u> : HIV model in mice using EcoHIV, with vs. without intranasal insulin treatment (daily for 9 days). <u>Outcome measures</u> : cognitive functions, hippocampal neuronal integrity, and the expression of genes associated with brain functions.	1. Infected mice exhibited cognitive impairment. 2. Intranasal insulin restored cognitive functions, hippocampal dendritic integrity, and gene expressions. 3. The beneficial effect of intranasal insulin was independent of HIV burden in the brain.	Infection → Cognitive impairment. Central insulin treatment → Restoration of brain functions	Kim et al., 2019
HIV/human FIV/cat	1. <u>Human <i>ex vivo</i> model</u> : (1) brain autopsy of patients with HIV/AIDS. (2) HIV-1 infection in primary human neurons and microglia, treated with insulin . <u>Outcome measures</u> : neuroinflammation and neuronal death. 2. <u>Animal <i>in vivo</i> HAND model</u> : feline retrovirus (FIV) intracranial infection in cats. Insulin intranasal treatment for 6 weeks compared to PBS treatment. <u>Outcome measures</u> : morphological changes in the brain, neuroinflammation, neuronal survival, neurobehavioral performance.	1-1. Increased neuroinflammatory gene expression in the brain of HIV/AIDS. 1-2. Insulin treatment suppressed HIV-1 growth and reduced infection-induced CSCL10 and <i>IL-6</i> expression in HIV-infected microglia. 1-3. Insulin treatment prevented HIV-1 infection-induced neuronal death. 2-1. Insulin treatment enhanced the preservation of cortical neurons, and improved neurobehavioral performance in FIV-infected cats.	Infection → Cognitive impairment. Central insulin treatment → Restoration of brain functions.	Mamik et al., 2016

(Continued)

TABLE 1 | (Continued)

Pathogen/host	Study design and outcome measures	Findings	Proposed direction of causality and/or mechanism	References
<i>Porphyromonas gingivalis</i>				
<i>P. gingivalis</i> /mouse	Animal model: <i>db/db</i> mouse infected with <i>P. gingivalis</i> (W83, p.o.). Outcome measures: neuroinflammation in the hippocampus; mRNA levels for genes associated with insulin signaling.	1. Infection induced reactive microglia and astrocytes. 2. Infection enhanced the expression of insulin signaling pathway genes (e.g., InsR , Igf1 , Irs , and Gsk3β). 3. Pro-inflammatory genes were also up-regulated.	Infection → Disruption of insulin signaling pathway and inflammation in the brain.	Bahar et al., 2021
<i>Toxoplasma gondii</i>				
<i>T. gondii</i> /rat	Animal model: <i>T. gondii</i> infection model in rats. Outcome measures: plasma leptin levels	Plasma leptin levels increased in chronic <i>T. gondii</i> infection.	CNS infection → Increase in plasma leptin (metabolic imbalance)	Baltaci and Mogulkoc, 2012
West Nile virus (WNV)				
WNV/mouse	Animal model: <i>db/db</i> mouse infected with WNV. Outcome measures: leukocyte infiltration and neuroinflammation/neuronal damage.	Infection-induced leukocyte infiltration into the brain was reduced, while neuroinflammation/neuronal death was enhanced, in <i>db/db</i> mice.	LepR dysfunction → Increased CNS infection susceptibility	Kumar et al., 2014

Relevant keywords, such as *insulin* and *leptin*, are highlighted in bold.

Central Nervous System Infections Causing Metabolic Imbalance – Borna Disease Virus and Canine Distemper Virus

Borna disease virus (BDV) is a neurotropic RNA virus infecting a broad host spectrum including humans. Borna disease was originally observed as infectious diseases in domestic animals in the nineteenth century. The confirmation of human infection has begun since the 1980s, mainly in neuropsychiatric patients (Hatalski et al., 1997; Schwemmler, 2001; Taieb et al., 2001). BDV causes persistent infection in the brain and neurobehavioral deficits associated with neuroinflammation. In an experimental infection model in rodents, it takes a biphasic course characterized by hyperactivity associated with inflammatory lesions in the brain during the first acute stage, followed by the development of varying symptoms, including obesity, depending on viral strains and affected brain regions (Narayan et al., 1983; Herden et al., 2000). Intracerebral infection of rats with a BDV variant induced obesity without neurological signs, which is correlated with severe mononuclear cell infiltration into the hypothalamus, suggesting that infection-induced neuroendocrine dysregulations caused the development of obesity (Herden et al., 2000). It was also suggested that BDV infection-induced neuroinflammation and neurotransmitter imbalance underlie the dysfunction of hypothalamus and leptin resistance, leading to obesity (Gosztonyi et al., 2020).

Canine distemper virus (CDV) is an RNA virus closely related to measles virus, infecting a wide range of host species (Martinez-Gutierrez and Ruiz-Saenz, 2016). CDV causes canine distemper, a severe systemic disease in dogs, presenting a variety of symptoms including neurologic disorders (Martella et al., 2008). In a virus CDV infection-induced obesity model in mice using intracerebral infection, hyperinsulinemia and

alteration in leptin signaling were observed (Bernard et al., 1988, 1999). In this model, infection caused hyperinsulinemia and obesity, while CDV showed tropism for the hypothalamus. Obesity developed in up to 30% of the surviving mice (Bernard et al., 1988). In addition, functional LepR was specifically down-regulated in the hypothalamus of infected obese mice (Bernard et al., 1999), suggesting that CDV infection in the brain induced leptin resistance in the hypothalamus, which led to obesity, thereby increasing the risk of MetS-associated cognitive impairment (Figure 3).

Peripheral Infection and Cognitive Impairment – Potential Involvement of Insulin and Glycemic Control

Peripheral infection-induced metabolic disturbance can lead to CNS dysfunction (Table 2). Sepsis-associated encephalopathy (SAE) is a brain disease secondary to peripheral infection without overt CNS infection, occurring up to 50–70% of sepsis cases (Chen Q. et al., 2014). After initial response to eliminate pathogens, systemic inflammation and increased BBB permeability occur, causing severe encephalopathy. While SAE is partly reversible, it can lead to persistent neurocognitive deficits, increasing the risk of dementia later in life (Widmann and Heneka, 2014; Seidel et al., 2020). Disrupted glycemic control is frequently encountered in sepsis patients (Hirasawa et al., 2009), and it has been suggested that higher infection susceptibility in patients with hyperglycemia is associated with disease severity (Koh et al., 2012). Experimental sepsis in rodents induced cognitive deficits accompanied by hyperglycemia (Huang et al., 2020). BBB disruption, microglial activation, oxidative damage and inflammation in the hippocampus, cortex and cerebrum occurred in those animal models (Michels et al., 2015; Sonnevile et al., 2015; Huang et al., 2020), suggesting glial activation and neuroinflammation underlie SAE-induced cognitive impairment.

In addition, the experimental sepsis induced more severe brain damages, including microglial activation and neuronal death, in hyperglycemic mice compared to insulin-treated mice (Sonneville et al., 2015), suggesting that poor glycemic control renders CNS more vulnerable to neuroinflammation, and insulin may protect the brain from sepsis-induced neuroinflammation and neuronal damages (Hache et al., 2015).

In addition, acute inflammation in the periphery (LPS challenge) induced metabolic changes in the CNS in rodents, and the metabolic imbalance in the brain was associated impaired cognition (Kealy et al., 2020). In addition, LPS-induced acute peripheral inflammation produced hypoglycemia in blood and CSF, and caused severe cognitive impairment selectively in those mice with brain disease (chronic neurodegeneration model induced by prion infection), which was mimicked by insulin and alleviated by glucose administration (Kealy et al., 2020). These data suggest that acute peripheral inflammation can cause neuroinflammation and cognitive deficits via disruption of glycemic control, whereas chronic neuroinflammation, including infection-induced neurodegenerative disorders, renders the brain more vulnerable to metabolic imbalance (Figure 3).

Other Pathogens Affecting Metabolic Balance – Indirect Link to Cognition

‘Infatobesity’ is a concept that proposes the infectious etiology of obesity (Pasarica and Dhurandhar, 2007; van Ginneken et al., 2009). Considering the link between MetS and cognitive decline (Mejido et al., 2020), infatobesity can be an indirect cause of dementia. In addition to BDV and CDV, there are several viruses potentially causing obesity (Dhurandhar, 2001; Atkinson, 2007). Rous-associated virus 7 (RAV-7), a retrovirus causing avian leukosis, can induce stunting and obesity in chickens, which is associated with hyperlipidemia and increased insulin levels, as well as immune dysregulation (Carter and Smith, 1984). Adenovirus 36 (Ad-36) is a human adenovirus associated with obesity (Esposito et al., 2012). Ad-36 infection in adipocytes reduced leptin production while inducing proliferation, differentiation, and lipid accumulation in adipocytes (Vangipuram et al., 2007; van Ginneken et al., 2009), suggesting viral infection of fat cells can directly cause adipogenesis and obesity. Reduced leptin release caused by infection may also compromise CNS-mediated control of energy homeostasis, further enhancing the risk of MetS-associated cognitive impairment (Whitmer et al., 2005; Kivipelto et al., 2006; Mejido et al., 2020).

Gut Microbiome

While the concept of microbial infection and pathogenicity has dominated the mainstream of microbiology, only a small fraction of microorganism are inherently pathogenic. In particular, the gut is inhabited by microbiota, a collection of microorganisms including bacteria, archaea, viruses, and fungi, consisting of at least 1,000 distinct species. Commensal bacterial and fungal species are involved in the regulation/dysregulation of energy homeostasis and immune responses, through extracting and metabolizing nutrients, regulating peripheral and central insulin sensitivity (Schertzer and Lam, 2021), and contributing to

intestinal immune control via bidirectional communication with immune cells (Khan et al., 2021). Disruption of the symbiotic relationship between the host and microbiota leads to chronic inflammation and insulin resistance (Patterson et al., 2016). Obesity is associated with altered gut microbial composition in mice and humans, and the trait is transmissible as colonization of germ-free mice with microbiota from obese mice led to a significant increase in body fat compared to colonization with microbiota from lean mice (Turnbaugh et al., 2006), suggesting that gut microbiome is part of the host metabolic system actively regulating energy balance (Bäckhed et al., 2004). It has also been suggested that microbiome plays a crucial role in the communication between the gut and brain (microbiota-gut-brain axis), which is essential for the regulation of energy homeostasis (Romaní-Pérez et al., 2021; van Son et al., 2021) as well as the development and functions of the nervous system (Chen et al., 2021; Gwak and Chang, 2021). Consequently, altered gut microbiota (dysbiosis) has been implicated in a number of chronic inflammatory diseases, such as diabetes (Zawada et al., 2020; Rodriguez and Delzenne, 2021) and neurodegenerative disorders, including AD (Jiang et al., 2017; Chen et al., 2021; Leblhuber et al., 2021; Romanenko et al., 2021). Furthermore, these indigenous microbiota play a critical role in host defense against infection, through stimulating mucosal immune defenses (e.g., of antimicrobial peptides and IgA release) and limiting resource availability/niche opportunity for invading microbes (Libertucci and Young, 2019). Consequently, dysbiosis has been associated with infection susceptibility (Lazar et al., 2018; Libertucci and Young, 2019), which may further accelerate immunometabolic imbalance.

Probiotics are living microorganisms that provide health benefits by improving or restoring the composition of gut microbiota. It has been shown that probiotics can reduce leptin secretion and improve hypothalamic leptin and insulin resistance in high fat diet-induced obesity models in rodents (Al-muzafar and Amin, 2017; Bagarolli et al., 2017; Ji et al., 2018; Cheng and Liu, 2020). Of note, probiotics may prevent AD pathogenesis by improving glucose metabolism. Triple transgenic AD model mice (3xTg-AD) exhibited significant metabolic impairment [increased glycated hemoglobin [HbA1c] in the serum, accumulation of advanced glycation end products (AGE), and impaired glucose uptake due to decreased glucose transporter levels in the brain], and all these parameters were normalized by oral treatment with probiotics (lactic acid bacteria and bifidobacteria) (Bonfili et al., 2020). These data suggest that beneficial gut microbiome can reduce AD pathogenesis via the restoration of metabolic balance (Figure 3).

DISCUSSION AND CONCLUSION

Metabolic diseases and infection are important risk factors for dementia (Whitmer et al., 2005; Kivipelto et al., 2006; Ashraf et al., 2019; Mejido et al., 2020). Metabolic dysregulation enhances infection susceptibility via immune dysfunction, whereas infection, both in the CNS and periphery, can disrupt metabolic balance (Figure 3). In the CNS, infection-induced

TABLE 2 | The roles of insulin in peripheral infection-induced CNS damage.

Pathogen or disease/host	Study design and outcome measures	Findings	Proposed direction of causality and/or mechanism	References
Sepsis/rat	<u>Animal model</u> : surgically induced polymicrobial sepsis model in rats. Glucose treatment, insulin treatment compared to control. <u>Outcome measures</u> : Blood glucose, behavioral deficits, brain activity (EEG), BBB permeability, glial activation and inflammation in the cerebrum.	1. Sepsis induced hyperglycemia. 2. Glucose treatment led to a decline in survival rate, reduced brain activity, increased BBB permeability, and enhanced microglial and astroglial activation and inflammatory responses in the cerebrum. 3. Glycemic control (insulin treatment) inhibited inflammatory responses and restored BBB integrity and brain activity to near normal.	Peripheral infection →Hyperglycemia. →Glial activation and neuroinflammation. → Cognitive dysfunction. Note: Insulin help restore brain functions by preventing BBB disruption and neuroinflammation.	Huang et al., 2020
Sepsis/mouse	<u>Animal model</u> : surgically induced polymicrobial sepsis model in mice. Mice with moderate hyperglycemia were compared to control (normoglycemia). <u>Outcome measures</u> : neuronal damages, glial activation, and cell death in the hippocampus and frontal cortex.	(A) In hyperglycemic mice (compared to normoglycemia mice), sepsis induced: 1. Higher neuronal damage in frontal cortex. 2. Microglial activation in frontal cortex and hippocampus. 3. More apoptotic cells in frontal cortex. (B) Insulin prevented the above damages	Hyperglycemia →Enhanced infection (sepsis)-induced brain damage. Note: Insulin prevents hyperglycemia-induced susceptibility to sepsis-induced brain damages	Sonneville et al., 2015
Sepsis/mouse	<u>Animal model</u> : surgically induced polymicrobial sepsis model in mice. <u>Outcome measures</u> : cognitive functions, hippocampal synaptic plasticity, and hippocampal insulin signaling in post-septic mice compared to control.	1. Post-septic mice exhibited cognitive impairment, which was accompanied by reduced synaptic plasticity and disrupted insulin signaling in the hippocampus. 2. Treatment with a GLP-1 receptor agonist (insulinotropic) or GSK3 β inhibitor (insulin signaling downstream) rescued cognition.	Peripheral infection →Disruption of hippocampal insulin signaling. → Impaired synaptic plasticity and cognitive deficits.	Neves et al., 2018
Sepsis/rat	<u>Animal model</u> : sepsis model induced by LPS (i.p.) in rats. Insulin treatment: continuous intravenous infusion for 6h after LPS stimulation. Compared to control (saline). <u>Outcome measures</u> : Inflammatory cytokines and oxidative stress in the cortex, hippocampus, and hypothalamus.	In the brain regions (cortex, hippocampus, and hypothalamus): 1. Insulin alleviated sepsis-induced inflammatory response (IL-1 β , IL-6, and TNF- α). 2. Insulin suppressed oxidative damage while restoring antioxidants (SOD and GSH).	Peripheral infection →Inflammation and oxidative stress in the brain. Note: Insulin treatment lowered sepsis-induced inflammation in the brain	Chen Q. et al., 2014
Systemic inflammation/mouse	<u>Animal model</u> : LPS (i.p.) challenge in chronic neurodegeneration model (ME7 prion infection) in mice. Treated with glucose and insulin . <u>Outcome measures</u> : blood glucose, cognitive performance	LPS challenge induced hypoglycemia and acute cognitive impairment in mice with brain disease (ME7 prion infection), which was mitigated by glucose and mimicked by insulin .	Peripheral infection →Metabolic imbalance in the CNS. →Cognitive impairment. Note: Brain disease (prion disease) makes the CNS more vulnerable to peripheral inflammation	Kealy et al., 2020

Relevant keywords, such as *insulin* and *leptin*, are highlighted in bold.

damage in the regions associated with metabolic control, such as hypothalamus, can disrupt whole-body energy metabolism. In the periphery, infection-induced immune reactions may persist and disarray immunometabolism, resulting in chronic inflammation and increased BBB permeability (Gustafson et al., 2007; Kanoski et al., 2010; Montagne et al., 2015), which can disrupt the CNS integrity, cognitive ability, and central energy control (Montagne et al., 2015). In addition, infection-induced dysregulation of glucose metabolism can lead to glial activation and neuroinflammation (Huang et al., 2020; Bahar et al., 2021). The vicious cycle may further advance the systemic imbalance, while beneficial gut microbiome can restore the balance of immunometabolism. Furthermore, although evidence is still lacking, infectious agents can potentially disrupt the immunometabolism in the brain by infecting microglia and astrocytes. Thus, multidirectional interactions between metabolic imbalance and infection at cellular and

systemic levels likely occur during the development of dementia and neurodegenerative disorders. Combined and multifactorial impacts due to these interactions can further exacerbate brain dysfunction. As the key regulators of energy metabolism and immune responses, insulin and leptin play significant roles in those intricate interactions. It should also be noted that other metabolic hormones and adipokines, such as glucagon and adiponectin, similarly take part in immunometabolism, and dysregulation of these molecules are also implicated in neurodegenerative disorders, including AD (Talbot and Wang, 2014; Grieco et al., 2019; Kim et al., 2020). In addition, growing evidence suggests the central actions of thyroid hormones (THs) (Capelli et al., 2021); THs exert immunomodulatory roles in the brain by inducing microglial phenotypic changes (Mallat et al., 2002) and affecting the expression of chemokines (Davis et al., 2016). Thyroid dysfunctions are possibly associated with AD pathogenesis (Figueroa et al., 2021). Future research

may unveil the interactions between infection and these metabolic hormones.

While the potential therapeutic benefit of insulin in treating dementia has been increasingly recognized (Frölich et al., 1998; Kim et al., 2019; Kellar and Craft, 2020), it is relatively unexplored how the modulation of metabolic regulators, including insulin and leptin, can impact infection-induced neurological disorders. Considering the diverse roles for these molecules play in the immunometabolic network, including the CNS immunometabolism (Larabee et al., 2020), it is plausible that infection-induced damages to the brain can be alleviated by modulating these signaling pathways. On the other hand, it is largely unresolved how glia - pathogen interactions can impact immunometabolism in the CNS and what roles leptin and insulin may play in the interactions. How these relationships may relate to the etiology of dementia is also an unanswered question. Further research is warranted to understand the mechanisms

underlying the interrelationship between infection, metabolic disorders, and dementia.

AUTHOR CONTRIBUTIONS

NS wrote the first draft of the manuscript. KK critically assessed the content. NS and KK jointly finalized the manuscript. Both authors contributed to the article and approved the submitted version.

FUNDING

This study was supported by the Nagasaki University “Doctoral Program for World-leading Innovative and Smart Education” for Global Health, “Global Health Elite Programme for Building a Healthier World”.

REFERENCES

- Abella, V., Scotece, M., Conde, J., Pino, J., Gonzalez-Gay, M., Gómez-Reino, J., et al. (2017). Leptin in the interplay of inflammation, metabolism and immune system disorders. *Nat. Rev. Rheumatol.* 13, 100–109. doi: 10.1038/nrrheum.2016.209
- Ai, S., and Klein, R. (2020). Update on T cells in the virally infected brain: friends and foes. *Curr. Opin. Neurol.* 33, 405–412. doi: 10.1097/WCO.0000000000000825
- Alli, A., Ortiz, J., Atoot, A., Atoot, A., and Millhouse, P. (2021). Management of West Nile encephalitis: an uncommon complication of West Nile virus. *Cureus* 13:e13183. doi: 10.7759/cureus.13183
- Allison, M., and Myers, M. J. (2014). 20 years of leptin: connecting leptin signaling to biological function. *J. Endocrinol.* 223, T25–T35. doi: 10.1530/JOE-14-0404
- Al-muzafar, H., and Amin, K. (2017). Probiotic mixture improves fatty liver disease by virtue of its action on lipid profiles, leptin, and inflammatory biomarkers. *BMC Complement. Altern. Med.* 17:43. doi: 10.1186/s12906-016-1540-z
- Alti, D., Sambamurthy, C., and Kalangi, S. (2018). Emergence of leptin in infection and immunity: scope and challenges in vaccines formulation. *Front. Cell Infect. Microbiol.* 8:147. doi: 10.3389/fcimb.2018.00147
- Amole, B., Wittner, M., Hewlett, D., and Tanowitz, H. (1985). Trypanosoma brucei: infection in murine diabetes. *Exp. Parasitol.* 60, 342–347. doi: 10.1016/0014-4894(85)90040-90042
- Ances, B., Ortega, M., Vaida, F., Heaps, J., and Paul, R. (2012). Independent effects of HIV, aging, and HAART on brain volumetric measures. *J. Acquir. Immune Defic. Syndr.* 59, 469–477. doi: 10.1097/QAI.0b013e318249db17
- André, C., Guzman-Quevedo, O., Rey, C., Rémus-Borel, J., Clark, S., Castellanos-Jankiewicz, A., et al. (2017). Inhibiting microglia expansion prevents diet-induced hypothalamic and peripheral inflammation. *Diabetes Metab. Res. Rev.* 66, 908–919. doi: 10.2337/db16-0586
- Antinori, A., Arendt, G., Becker, J., Brew, B., Byrd, D., Cherner, M., et al. (2007). Updated research nosology for HIV-associated neurocognitive disorders. *Neurology* 69, 1789–1799. doi: 10.1212/01.WNL.0000287431.88658.8b
- Arnold, S., Arvanitakis, Z., Macauley-Rambach, S., Koenig, A., Wang, H., Ahima, R., et al. (2018). Brain insulin resistance in type 2 diabetes and Alzheimer disease: concepts and conundrums. *Nat. Rev. Neurol.* 14, 168–181. doi: 10.1038/nrneuro.2017.185
- Arvanitakis, Z., Wilson, R., Bienias, J., Evans, D., and Bennett, D. (2004). Diabetes mellitus and risk of Alzheimer disease and decline in cognitive function. *Arch. Neurol.* 61, 661–666. doi: 10.1001/archneur.61.5.661
- Ashraf, G., Tarasov, V., Makhmutová, A., Chubarev, V., Avila-Rodriguez, M., Bachurin, S., et al. (2019). The possibility of an infectious etiology of Alzheimer disease. *Mol. Neurobiol.* 56, 4479–4491. doi: 10.1007/s12035-018-1388-y
- Atkinson, R. (2007). Viruses as an etiology of obesity. *Mayo Clin. Proc.* 82, 1192–1198. doi: 10.4065/82.10.1192
- Avalos, C., Abreu, C., Queen, S., Li, M., Price, S., Shirk, E., et al. (2017). Brain macrophages in simian immunodeficiency virus-infected, antiretroviral-suppressed macaques. *mBio* 8:e01186-17. doi: 10.1128/mBio.01186-17
- Bäckhed, F., Ding, H., Wang, T., Hooper, L., Koh, G., Nagy, A., et al. (2004). The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. U.S.A.* 101, 15718–15723. doi: 10.1073/pnas.0407076101
- Badawi, A., Velummailum, R., Ryoo, S., Senthinathan, A., Yaghoubi, S., Vasileva, D., et al. (2018). Prevalence of chronic comorbidities in dengue fever and West Nile virus: a systematic review and meta-analysis. *PLoS One* 13:e0200200. doi: 10.1371/journal.pone.0200200
- Bagaroli, R., Tobar, N., Oliveira, A., Araújo, T., Carvalho, B., Rocha, G., et al. (2017). Probiotics modulate gut microbiota and improve insulin sensitivity in DIO mice. *J. Nutr. Biochem.* 50, 16–25. doi: 10.1016/j.jnutbio.2017.08.00
- Bahar, B., Kanagasingham, S., Tambuwala, M., Aljabali, A., Dillon, S., Doaei, S., et al. (2021). Porphyromonas gingivalis (W83) infection induces Alzheimer's disease-like pathophysiology in obese and diabetic mice. *J. Alzheimers Dis.* 82, 1259–1275. doi: 10.3233/JAD-210465
- Balin, B., Gérard, H., Arking, E., Appelt, D., Branigan, P., Abrams, J., et al. (1998). Identification and localization of *Chlamydia pneumoniae* in the Alzheimer's brain. *Med. Microbiol. Immunol.* 187, 23–42. doi: 10.1007/s004300050071
- Baltaci, A., and Mogulkoc, R. (2012). Plasma leptin levels in rats with induced *Toxoplasma gondii* infection. *Bratisl Lek List.* 113, 67–69. doi: 10.4149/bll_2012_016
- Balthasar, N., Coppari, R., McMinn, J., Liu, S., Lee, C., Tang, V., et al. (2004). Leptin receptor signaling in POMC neurons is required for normal body weight homeostasis. *Neuron* 42, 983–991. doi: 10.1016/j.neuron.2004.06.004
- Balyan, R., Gautam, N., and Gascoigne, N. (2020). The Ups and downs of metabolism during the lifespan of a T Cell. *Int. J. Mol. Sci.* 21:7972. doi: 10.3390/ijms21217972
- Banks, W., and Farrell, C. (2003). Impaired transport of leptin across the blood-brain barrier in obesity is acquired and reversible. *Am. J. Physiol. Endocrinol. Metab.* 285, E10–E15. doi: 10.1152/ajpendo.00468.2002
- Banks, W., Owen, J., and Erickson, M. (2012). Insulin in the brain: there and back again. *Pharmacol. Ther.* 136, 82–93. doi: 10.1016/j.pharmthera.2012.07.006
- Baradaran, A., Dehghanbanadaki, H., Naderpour, S., Pirkashani, L., Rajabi, A., Rashti, R., et al. (2021). The association between *Helicobacter pylori* and obesity: a systematic review and meta-analysis of case-control studies. *Clin. Diabetes Endocrinol.* 7:15. doi: 10.1186/s40842-021-00131-w
- Barrios, V., López-Villar, E., Frago, L., Canelles, S., Díaz-González, F., Burgos-Ramos, E., et al. (2021). Cerebral insulin bolus revokes the changes in hepatic

- lipid metabolism induced by chronic central leptin infusion. *Cells* 10:581. doi: 10.3390/cells10030581
- Bernard, A., Cohen, R., Khuth, S., Vedrine, B., Verlaeten, O., Akaoka, H., et al. (1999). Alteration of the leptin network in late morbid obesity induced in mice by brain infection with canine distemper virus. *J. Virol.* 73, 7317–7327. doi: 10.1128/JVI.73.9.7317-7327.1999
- Bernard, A., Zwingelstein, G., Meister, R., and Wild, T. (1988). Hyperinsulinemia induced by canine distemper virus infection of mice and its correlation with the appearance of obesity. *Comp. Biochem. Physiol. B* 91, 691–696. doi: 10.1016/0305-0491(88)90193-90199
- Bernier, L., York, E., Kamyabi, A., Choi, H., Weiling, N., and MacVicar, B. (2020). Microglial metabolic flexibility supports immune surveillance of the brain parenchyma. *Nat. Commun.* 11:1559. doi: 10.1038/s41467-020-15267-z
- Beydoun, M., Beydoun, H., Elbejjani, M., Dore, G., and Zonderman, A. (2018). *Helicobacter pylori* seropositivity and its association with incident all-cause and Alzheimer's disease dementia in large national surveys. *Alzheimers Dement* 14, 1148–1158. doi: 10.1016/j.jalz.2018.04.009
- Biessels, G., and Despa, F. (2018). Cognitive decline and dementia in diabetes mellitus: mechanisms and clinical implications. *Nat. Rev. Endocrinol.* 14, 591–604.
- Biessels, G., Staekenborg, S., Brunner, E., Brayne, C., and Scheltens, P. (2006). Risk of dementia in diabetes mellitus: a systematic review. *Lancet Neurol.* 5, 64–74. doi: 10.1016/S1474-4422(05)70284-70282
- Bonda, D., Stone, J., Torres, S., Siedlak, S., Perry, G., Kryscio, R., et al. (2014). Dysregulation of leptin signaling in Alzheimer disease: evidence for neuronal leptin resistance. *J. Neurochem.* 128, 162–172. doi: 10.1111/jnc.12380
- Bonfili, L., Cecarini, V., Gogoi, O., Berardi, S., Scarpona, S., Angeletti, M., et al. (2020). Gut microbiota manipulation through probiotics oral administration restores glucose homeostasis in a mouse model of Alzheimer's disease. *Neurobiol. Aging* 87, 35–43. doi: 10.1016/j.neurobiolaging.2019.11.004
- Bos, I., Vos, S., Frölich, L., Kornhuber, J., Wiltfang, J., Maier, W., et al. (2017). The frequency and influence of dementia risk factors in prodromal Alzheimer's disease. *Neurobiol. Aging* 56, 33–40. doi: 10.1016/j.neurobiolaging.2017.03.034
- Boucsein, A., Kamstra, K., and Tups, A. (2021). Central signalling cross-talk between insulin and leptin in glucose and energy homeostasis. *J. Neuroendocr.* 33:e12944. doi: 10.1111/jne.12944
- Boutens, L., Hooiveld, G., Dhingra, S., Cramer, R., Netea, M., and Stienstra, R. (2018). Unique metabolic activation of adipose tissue macrophages in obesity promotes inflammatory responses. *Diabetologia* 61, 942–953. doi: 10.1007/s00125-017-4526-4526
- Brabazon, F., Bermudez, S., Shaughness, M., Khayrullina, G., and Byrnes, K. (2018). The effects of insulin on the inflammatory activity of BV2 microglia. *PLoS One* 13:e0201878. doi: 10.1371/journal.pone.0201878
- Brew, B., and Barnes, S. (2019). The impact of HIV central nervous system persistence on pathogenesis. *AIDS* 33, S113–S121. doi: 10.1097/QAD.0000000000002251
- Bulloch, K., Miller, M., Gal-Toth, J., Milner, T., Gottfried-Blackmore, A., Waters, E., et al. (2008). CD11c/EYFP transgene illuminates a discrete network of dendritic cells within the embryonic, neonatal, adult, and injured mouse brain. *J. Comp. Neurol.* 508, 687–710. doi: 10.1002/cne.21668
- Caldefie-Chezet, F., Poulin, A., and Vasson, M. (2003). Leptin regulates functional capacities of polymorphonuclear neutrophils. *Free Radic. Res.* 37, 809–814. doi: 10.1080/1071576031000097526
- Calza, L., Masetti, G., Piergentili, B., Trapani, F., Cascavilla, A., Manfredi, R., et al. (2011). Prevalence of diabetes mellitus, hyperinsulinaemia and metabolic syndrome among 755 adult patients with HIV-1 infection. *Int. J. STD AIDS* 22, 43–45. doi: 10.1258/ijisa.2010.010256
- Capelli, V., Diéguez, C., Mittag, J., and López, M. (2021). Thyroid wars: the rise of central actions. *Trends Endocrinol. Metab.* 32, 659–671. doi: 10.1016/j.tem.2021.05.006
- Cárdenas, G., Soto-Hernández, J., Díaz-Alba, A., Ugalde, Y., Mérida-Puga, J., Rosetti, M., et al. (2014). Neurological events related to influenza A (H1N1) pdm09. *Influ. Other Respir. Viruses* 8, 339–346. doi: 10.1111/irv.12241
- Carro, E., Trejo, J., Núñez, A., and Torres-Aleman, I. (2003). Brain repair and neuroprotection by serum insulin-like growth factor. *Mol. Neurobiol.* 27, 153–162.
- Carter, J., and Smith, R. (1984). Specificity of avian leukosis virus-induced hyperlipidemia. *J. Virol.* 50, 301–308. doi: 10.1128/JVI.50.2.301-308.1984
- Cereda, E., Sansone, V., Meola, G., and Malavazos, A. (2007). Increased visceral adipose tissue rather than BMI as a risk factor for dementia. *Age Ageing* 36, 488–491. doi: 10.1093/ageing/afm096
- Cheeran, M., Hu, S., Yager, S., Gekker, G., Peterson, P., and Lokensgard, J. (2001). Cytomegalovirus induces cytokine and chemokine production differentially in microglia and astrocytes. *J. Neurovirol.* 7, 135–147. doi: 10.1080/13550280120508799
- Chen, H., Charlat, O., Tartaglia, L., Woolf, E., Weng, X., Ellis, S., et al. (1996). Evidence that the diabetes gene encodes the leptin receptor: identification of a mutation in the leptin receptor gene in db/db mice. *Cell* 84, 491–495. doi: 10.1016/s0092-8674(00)81294-81295
- Chen, Q., Yu, W., Shi, J., Shen, J., Gao, T., Zhang, J., et al. (2014). Insulin alleviates the inflammatory response and oxidative stress injury in cerebral tissues in septic rats. *J. Inflamm.* 11:18. doi: 10.1186/1476-9255-11-18
- Chen, Y., Zhao, Y., Dai, C., Liang, Z., Run, X., Iqbal, K., et al. (2014). Intranasal insulin restores insulin signaling, increases synaptic proteins, and reduces Abeta level and microglia activation in the brains of 3xTg-AD mice. *Exp. Neurol.* 261, 610–619. doi: 10.1016/j.expneurol.2014.06.004
- Chen, Y., Xu, J., and Chen, Y. (2021). Regulation of neurotransmitters by the gut microbiota and effects on cognition in neurological disorders. *Nutrients* 13:2099. doi: 10.3390/nu13062099
- Cheng, Y., and Liu, J. (2020). Effect of *Lactobacillus rhamnosus* GG on energy metabolism, leptin resistance, and gut microbiota in mice with diet-induced obesity. *Nutrients* 12:2557. doi: 10.3390/nu12092557
- Chi, H. (2012). Regulation and function of mTOR signalling in T cell fate decisions. *Nat. Rev. Immunol.* 12, 325–338. doi: 10.1038/nri3198
- Chowen, J., and Garcia-Segura, L. (2020). Microglia, neurodegeneration and loss of neuroendocrine control. *Prog. Neurobiol.* 184:101720. doi: 10.1016/j.neurobio.2019.101720
- Cohen, S., Danzaki, K., and MacIver, N. (2017). Nutritional effects on T-cell immunometabolism. *Eur. J. Immunol.* 47, 225–235. doi: 10.1002/eji.201646423
- Colonna, M., and Butovsky, O. (2017). Microglia function in the central nervous system during health and neurodegeneration. *Annu. Rev. Immunol.* 35, 441–468. doi: 10.1146/annurev-immunol-051116-052358
- Conde, J., Scotete, M., Gómez, R., Gómez-Reino, J., Lago, F., and Gualillo, O. (2010). At the crossroad between immunity and metabolism: focus on leptin. *Expert Rev. Clin. Immunol.* 6, 801–808. doi: 10.1586/eci.10.48
- Costa, M., de Araújo, I., da Rocha, Alves, L., da Silva, R., Dos Santos, et al. (2021). Relationship of *Porphyromonas gingivalis* and Alzheimer's disease: a systematic review of pre-clinical studies. *Clin. Oral Investig.* 25, 797–806. doi: 10.1007/s00784-020-03764-w
- Curi, R., Levada-Pires, A., Silva, E., Poma, S., Zambonato, R., Domenech, P., et al. (2020). The critical role of cell metabolism for essential neutrophil functions. *Cell Physiol. Biochem.* 54, 629–647. doi: 10.33594/000000245
- Dani, N., Herbst, R., McCabe, C., Green, G., Kaiser, K., Head, J., et al. (2021). A cellular and spatial map of the choroid plexus across brain ventricles and ages. *Cell* 184, 3056–3074. doi: 10.1016/j.cell.2021.04.003
- Davis, A., Rohlwick, U., Proust, A., Figaji, A., and Wilkinson, R. (2019). The pathogenesis of tuberculous meningitis. *J. Leukoc. Biol.* 105, 267–280. doi: 10.1002/JLB.MR0318-102R
- Davis, B., Salinas-Navarro, M., Cordeiro, M., Moons, L., and De Groef, L. (2017). Characterizing microglia activation: a spatial statistics approach to maximize information extraction. *Sci. Rep.* 7:1576. doi: 10.1038/s41598-017-01747-1748
- Davis, N., Mota, B., Stead, L., Palmer, E., Lombardero, L., Rodríguez-Puertas, R., et al. (2021). Pharmacological ablation of astrocytes reduces A β degradation and synaptic connectivity in an ex vivo model of Alzheimer's disease. *J. Neuroinflamm.* 18:73. doi: 10.1186/s12974-021-02117-y
- Davis, P., Glinksky, G., Lin, H., and Mousa, S. (2016). Actions of thyroid hormone analogues on chemokines. *J. Immunol. Res.* 2016:3147671. doi: 10.1155/2016/3147671
- De Strooper, B., and Karran, E. (2016). The cellular phase of Alzheimer's disease. *Cell* 164, 603–615. doi: 10.1016/j.cell.2015.12.056
- Deckers, K., Van Bostel, M., Verhey, F., and Köhler, S. (2017). Obesity and cognitive decline in adults: effect of methodological choices and confounding by age in a longitudinal study. *J. Nutr. Heal. Aging* 21, 546–553. doi: 10.1007/s12603-016-0757-753

- Dhurandhar, N. (2001). Infectobesity: obesity of infectious origin. *J. Nutr.* 131, 2794S–2797S. doi: 10.1093/jn/131.10.2794S
- Dhurandhar, N. (2011). A framework for identification of infections that contribute to human obesity. *Lancet Infect. Dis.* 11, 963–969. doi: 10.1016/S1473-3099(11)70274-70272
- DiAngelo, J., Bland, M., Bambina, S., Cherry, S., and Birnbaum, M. (2009). The immune response attenuates growth and nutrient storage in *Drosophila* by reducing insulin signaling. *Proc. Natl. Acad. Sci. U S A.* 106, 20853–20858. doi: 10.1073/pnas.0906749106
- Dietrich, M., Spuch, C., Antequera, D., Rodal, I., de Yébenes, J., Molina, J., et al. (2008). Megalin mediates the transport of leptin across the blood-CSF barrier. *Neurobiol. Aging* 29, 902–912. doi: 10.1016/j.neurobiolaging.2007.01.008
- Dominy, S., Lynch, C., Ermini, F., Benedyk, M., Marczyk, A., Konradi, A., et al. (2019). Porphyromonas gingivalis in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci. Adv.* 5:eau3333. doi: 10.1126/sciadv.aau3333
- Drögemüller, K., Helmuth, U., Brun, A., Sakowicz-Burkiewicz, M., Gutmann, D., Mueller, W., et al. (2008). Astrocyte gp130 expression is critical for the control of *Toxoplasma encephalitis*. *J. Immunol.* 181, 2683–2693. doi: 10.4049/jimmunol.181.4.2683
- Du, L., Lin, L., Li, Q., Liu, K., Huang, Y., Wang, X., et al. (2019). IGF-2 preprograms maturing macrophages to acquire oxidative phosphorylation-dependent anti-inflammatory properties. *Cell Metab.* 29, 1363–1375. doi: 10.1016/j.cmet.2019.01.006
- Dyer, A., Vahdatpour, C., Sanfelici, A., and Tropea, D. (2016). The role of Insulin-Like Growth Factor 1 (IGF-1) in brain development, maturation and neuroplasticity. *Neuroscience* 325, 89–99. doi: 10.1016/j.neuroscience.2016.03.056
- El Saftawy, E., Amin, N., Sabry, R., El-Anwar, N., Shash, R., Elsebaie, E., et al. (2020). Can *Toxoplasma gondii* pave the road for dementia? *J. Parasitol. Res.* 2020:8859857. doi: 10.1155/2020/8859857
- Elmqvist, J., Bjørbaek, C., Ahima, R., Flier, J., and Saper, C. (1998). Distributions of leptin receptor mRNA isoforms in the rat brain. *J. Comp. Neurol.* 395, 535–547.
- Esposito, K., Nappo, F., Marfella, R., Giugliano, G., Giugliano, F., Ciotola, M., et al. (2002). Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation* 106, 2067–2072. doi: 10.1161/01.cir.0000034509.14906.ae
- Esposito, S., Preti, V., Consolo, S., Nazzari, E., and Principi, N. (2012). Adenovirus 36 infection and obesity. *J. Clin. Virol.* 55, 95–100. doi: 10.1016/j.jcv.2012.06.009
- Faggioni, R., Fantuzzi, G., Gabay, C., Moser, A., Dinarello, C., Feingold, K., et al. (1999). Leptin deficiency enhances sensitivity to endotoxin-induced lethality. *Am. J. Physiol.* 276, R136–R142. doi: 10.1152/ajpregu.1999.276.1.R136
- Fantuzzi, G., and Faggioni, R. (2000). Leptin in the regulation of immunity, inflammation, and hematopoiesis. *J. Leukoc. Biol.* 68, 437–446.
- Fasshauer, M., and Blüher, M. (2015). Adipokines in health and disease. *Trends Pharmacol. Sci.* 36, 461–470. doi: 10.1016/j.tips.2015.04.014
- Fernandez, A., Hernandez, E., Guerrero-Gomez, D., Miranda-Vizuete, A., and Torres Aleman, I. (2018). A network of insulin peptides regulate glucose uptake by astrocytes: potential new druggable targets for brain hypometabolism. *Neuropharmacology* 136, 216–222. doi: 10.1016/j.neuropharm.2017.08.034
- Fernandez, A., and Torres-Aleman, I. (2012). The many faces of insulin-like peptide signalling in the brain. *Nat. Rev. Neurosci.* 13, 225–239.
- Fernández-Martos, C., González, P., and Rodríguez, F. (2012). Acute leptin treatment enhances functional recovery after spinal cord injury. *PLoS One* 7:e35594. doi: 10.1371/journal.pone.0035594
- Fernández-Real, J., Ferri, M., Vendrell, J., and Ricart, W. (2007). Burden of infection and fat mass in healthy middle-aged men. *Obes (Silver Spring)* 15, 245–252. doi: 10.1038/oby.2007.541
- Fernández-Real, J., López-Bermejo, A., Vendrell, J., Ferri, M., Recasens, M., and Ricart, W. (2006). Burden of infection and insulin resistance in healthy middle-aged men. *Diabetes Care* 29, 1058–1064. doi: 10.2337/diacare.2951058
- Ferri, C., Prince, M., Brayne, C., Brodaty, H., Fratiglioni, L., Ganguli, M., et al. (2005). Global prevalence of dementia: a *Delphi consensus* study. *Lancet* 366, 2112–2117. doi: 10.1016/S0140-6736(05)67889-67880
- Figuerola, P., Ferreira, A., Britto, L., Doussoulin, A., and Torrão, A. (2021). Association between thyroid function and Alzheimer's disease: a systematic review. *Metab. Brain Dis.* 36, 1523–1543. doi: 10.1007/s11011-021-00760-761
- Fischer, A., Cannon, B., and Nedergaard, J. (2020). Leptin: is it thermogenic? *Endocr Rev.* 41, 232–260. doi: 10.1210/edrv/bnz016
- Fischer, H., and Reichmann, G. (2001). Brain dendritic cells and macrophages/microglia in central nervous system inflammation. *J. Immunol.* 166, 2717–2726. doi: 10.4049/jimmunol.166.4.2717
- Fleck-Derderian, S., McClellan, W., and Wojcicki, J. (2017). The association between cytomegalovirus infection, obesity, and metabolic syndrome in U.S. adult females. *Obes (Silver Spring)* 25, 626–633. doi: 10.1002/oby.21764
- Foley, J., Wright, M., Gooding, A., Ettenhofer, M., Kim, M., Choi, M., et al. (2011). Operationalization of the updated diagnostic algorithm for classifying HIV-related cognitive impairment and dementia. *Int. Psychogeriatr.* 23, 835–843. doi: 10.1017/S1041610210002085
- Francisco, V., Ruiz-Fernández, C., Pino, J., Mera, A., González-Gay, M., Gómez, R., et al. (2019). Adipokines: linking metabolic syndrome, the immune system, and arthritic diseases. *Biochem. Pharmacol.* 165, 196–206. doi: 10.1016/j.bcp.2019.03.030
- Friedman, J. (2019). Leptin and the endocrine control of energy balance. *Nat. Metab.* 1, 754–764. doi: 10.1038/s42255-019-0095-y
- Frölich, L., Blum-Degen, D., Bernstein, H., Engelsberger, S., Humrich, J., Laufer, S., et al. (1998). Brain insulin and insulin receptors in aging and sporadic Alzheimer's disease. *J. Neural Transm. (Vienna)*. 105, 423–438. doi: 10.1007/s007020050068
- Fujita, Y., and Yamashita, T. (2019). The effects of leptin on glial cells in neurological diseases. *Front. Neurosci.* 13:828. doi: 10.3389/fnins.2019.00828
- Fulop, T., Tripathi, S., Rodrigues, S., Desroches, M., Bunt, T., Eiser, A., et al. (2021). Targeting impaired antimicrobial immunity in the brain for the treatment of Alzheimer's disease. *Neuropsychiatr Dis. Treat* 17, 1311–1339. doi: 10.2147/NDT.S264910
- Fulton, S., Pissios, P., Manchon, R., Stiles, L., Frank, L., Pothos, E. N., et al. (2006). Leptin regulation of the mesoaccumbens dopamine pathway. *Neuron* 51, 811–822. doi: 10.1016/j.neuron.2006.09.006
- Fünfschilling, U., Supplie, L., Mahad, D., Boretius, S., Saab, A., Edgar, J., et al. (2012). Glycolytic oligodendrocytes maintain myelin and long-term axonal integrity. *Nature* 485, 517–521. doi: 10.1038/nature11007
- Gabbouj, S., Ryhänen, S., Marttinen, M., Wittrahm, R., Takalo, M., Kemppainen, S., et al. (2019). Altered insulin signaling in Alzheimer's disease brain - special emphasis on PI3K-Akt pathway. *Front. Neurosci.* 13:629. doi: 10.3389/fnins.2019.00629
- Gaber, T., Strehl, C., and Buttgerit, F. (2017). Metabolic regulation of inflammation. *Nat. Rev. Rheumatol.* 13, 267–279. doi: 10.1038/nrrheum.2017.37
- Gao, Y., Vidal-Itriago, A., Milanova, I., Korpel, N., Kalsbeek, M., Tom, R., et al. (2018). Deficiency of leptin receptor in myeloid cells disrupts hypothalamic metabolic circuits and causes body weight increase. *Mol. Metab.* 7, 155–160. doi: 10.1016/j.molmet.2017.11.003
- Garber, C., Soung, A., Vollmer, L., Kanmogne, M., Last, A., Brown, J., et al. (2019). T cells promote microglia-mediated synaptic elimination and cognitive dysfunction during recovery from neuropathogenic flaviviruses. *Nat. Neurosci.* 22, 1276–1288. doi: 10.1038/s41593-019-0427-y
- Gautron, L., and Elmqvist, J. (2011). Sixteen years and counting: an update on leptin in energy balance. *J. Clin. Invest.* 121, 2087–2093. doi: 10.1172/JCI45888
- Gemma, C., Bachstetter, A., and Bickford, P. (2010). Neuron-Microglia dialogue and hippocampal neurogenesis in the aged brain. *Aging Dis.* 1, 232–244.
- Gerozissis, K. (2003). Brain insulin: regulation, mechanisms of action and functions. *Cell Mol. Neurobiol.* 23, 873–874.
- Ghazanfari, N., Gregory, J., Devi, S., Fernandez-Ruiz, D., Beattie, L., Mueller, S., et al. (2021). CD8(+) and CD4(+) T cells infiltrate into the brain during *Plasmodium berghei* ANKA infection and form long-term resident memory. *J. Immunol.* 207, 1578–1590. doi: 10.4049/jimmunol.2000773
- González-García, I., Gruber, T., and García-Cáceres, C. (2021). Insulin action on astrocytes: from energy homeostasis to behaviour. *J. Neuroendocr.* 33:e12953. doi: 10.1111/jne.12953
- Gordleeva, S., Kanakov, O., Ivanchenko, M., Zaikin, A., and Franceschi, C. (2020). Brain aging and garbage cleaning: modelling the role of sleep, lymphatic system, and microglia senescence in the propagation of inflammation. *Semin. Immunopathol.* 42, 647–665. doi: 10.1007/s00281-020-00816-x

- Gosztonyi, G., Ludwig, H., Bode, L., Kao, M., Sell, M., Petrusz, P., et al. (2020). Obesity induced by Borna disease virus in rats: key roles of hypothalamic fast-acting neurotransmitters and inflammatory infiltrates. *Brain Struct. Funct.* 225, 1459–1482. doi: 10.1007/s00429-020-02063-2060
- Gosztyla, M., Brothers, H., and Robinson, S. (2018). Alzheimer's amyloid-beta is an antimicrobial peptide: a review of the evidence. *J. Alzheimers Dis.* 62, 1495–1506. doi: 10.3233/JAD-171133
- Gray, S., Meijer, R., and Barrett, E. (2014). Insulin regulates brain function, but how does it get there? *Diabetes* 63, 3992–3997. doi: 10.2337/db14-0340
- Grieco, M., Giorgi, A., Gentile, M., D'Erme, M., Morano, S., Maras, B., et al. (2019). Glucagon-Like Peptide-1: a focus on neurodegenerative diseases. *Front. Neurosci.* 13:1112. doi: 10.3389/fnins.2019.01112
- Gruzdeva, O., Borodkina, D., Uchasova, E., Dyleva, Y., and Barbarash, O. (2019). Leptin resistance: underlying mechanisms and diagnosis. *Diab. Metab. Syndr. Obes* 12, 191–198. doi: 10.2147/DMSO.S182406
- Guglielmi, V., Colangeli, L., D'Adamo, M., and Sbraccia, P. (2021). Susceptibility and severity of viral infections in obesity: lessons from influenza to COVID-19. does leptin play a role? *Int. J. Mol. Sci.* 22:3183. doi: 10.3390/ijms22063183
- Guo, Z., Chen, Y., Mao, Y., Zheng, T., Jiang, Y., Yan, Y., et al. (2017). Long-term treatment with intranasal insulin ameliorates cognitive impairment, tau hyperphosphorylation, and microglial activation in a streptozotocin-induced Alzheimer's rat model. *Sci. Rep.* 7:45971. doi: 10.1038/srep45971
- Gustafson, D., Karlsson, C., Skoog, I., Rosengren, L., Lissner, L., and Blennow, K. (2007). Mid-life adiposity factors relate to blood-brain barrier integrity in late life. *J. Int. Med.* 262, 643–650. doi: 10.1111/j.1365-2796.2007.01869.x
- Gwak, M., and Chang, S. (2021). Gut-Brain connection: microbiome, gut barrier, and environmental sensors. *Immune Netw.* 21:e20. doi: 10.4110/in.2021.21.e20
- Haas, C., de Carvalho, A., Muller, A., Eggen, B., and Portela, L. (2020). Insulin activates microglia and increases COX-2/IL-1beta expression in young but not in aged hippocampus. *Brain Res.* 1741:146884. doi: 10.1016/j.brainres.2020.146884
- Hache, G., Osuchowski, M., and Thiemermann, C. (2015). Does insulin protect the brain in mice and man with sepsis? *Shock* 44:287. doi: 10.1097/SHK.0000000000000423
- Hamdy, O., Porrmatikul, S., and Al-Ozairi, E. (2006). Metabolic obesity: the paradox between visceral and subcutaneous fat. *Curr. Diab. Rev.* 2, 367–373. doi: 10.2174/1573399810602040367
- Hargrave, K., Woods, S., Millington, O., Chalmers, S., Westrop, G., and Roberts, C. (2019). Multi-Omics studies demonstrate *Toxoplasma gondii*-induced metabolic reprogramming of murine dendritic cells. *Front. Cell Infect. Microbiol.* 9:309. doi: 10.3389/fcimb.2019.00309
- Harris, S., and Harris, E. (2015). Herpes simplex virus type 1 and other pathogens are key causative factors in sporadic Alzheimer's disease. *J. Alzheimers Dis.* 48, 319–353. doi: 10.3233/JAD-142853
- Haslam, D., and James, W. (2005). Obesity. *Lancet* 366, 1197–1209. doi: 10.1016/S0140-6736(05)67483-67481
- Hatalski, C., Lewis, A., and Lipkin, W. (1997). Borna disease. *Emerg. Infect. Dis.* 3, 129–135. doi: 10.3201/eid0302.970205
- Helderman, J. (1984). Acute regulation of human lymphocyte insulin receptors. analysis by the glucose clamp. *J. Clin. Invest.* 74, 1428–1435. doi: 10.1172/JCI111554
- Helderman, J., Reynolds, T., and Strom, T. (1978). The insulin receptor as a universal marker of activated lymphocytes. *Eur. J. Immunol.* 8, 589–595. doi: 10.1002/eji.1830080810
- Heneka, M., Carson, M., El Khoury, J., Landreth, G., Brosseron, F., Feinstein, D., et al. (2015). Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14, 388–405. doi: 10.1016/S1474-4422(15)70016-70015
- Heneka, M., Golenbock, D., Latz, E., Morgan, D., and Brown, R. (2020). Immediate and long-term consequences of COVID-19 infections for the development of neurological disease. *Alzheimers Res. Ther.* 12:69. doi: 10.1186/s13195-020-00640-643
- Heni, M., Hennige, A., Peter, A., Siegel-Axel, D., Ordelheide, A., Krebs, N., et al. (2011). Insulin promotes glycogen storage and cell proliferation in primary human astrocytes. *PLoS One* 6:e21594. doi: 10.1371/journal.pone.0021594
- Herden, C., Herzog, S., Richt, J., Nessler, A., Christ, M., Failing, K., et al. (2000). Distribution of borna disease virus in the brain of rats infected with an obesity-inducing virus strain. *Brain Pathol.* 10, 39–48. doi: 10.1111/j.1750-3639.2000.tb00241.x
- Hernandez-Garzón, E., Fernandez, A., Perez-Alvarez, A., Genis, L., Bascuñana, P., Fernandez de la Rosa, R., et al. (2016). The insulin-like growth factor I receptor regulates glucose transport by astrocytes. *Glia* 64, 1962–1971. doi: 10.1002/glia.23035
- Herz, J., Filiano, A., Smith, A., Yogev, N., and Kipnis, J. (2017). Myeloid cells in the central nervous system. *Immunity* 46, 943–956. doi: 10.1016/j.immuni.2017.06.007
- Hierro-Bujalance, C., Del Marco, A., José Ramos-Rodríguez, J., Infante-García, C., Bella Gomez-Santos, S., Herrera, M., et al. (2020). Cell proliferation and neurogenesis alterations in Alzheimer's disease and diabetes mellitus mixed murine models. *J. Neurochem.* 154, 673–692. doi: 10.1111/jnc.14987
- Hirasawa, H., Oda, S., and Nakamura, M. (2009). Blood glucose control in patients with severe sepsis and septic shock. *World J. Gastroenterol.* 15, 4132–4136. doi: 10.3748/wjg.15.4132
- Holden, K., Lindquist, K., Tylavsky, F., Rosano, C., Harris, T., and Yaffe, K. (2009). Serum leptin level and cognition in the elderly: findings from the Health ABC Study. *Neurobiol. Aging* 30, 1483–1489. doi: 10.1016/j.neurobiolaging.2007.11.024
- Hommel, J., Trinko, R., Sears, R., Georgescu, D., Liu, Z., Gao, X., et al. (2006). Leptin receptor signaling in midbrain dopamine neurons regulates feeding. *Neuron* 51, 801–810. doi: 10.1016/j.neuron.2006.08.023
- Hotamisligil, G. (2017). Inflammation, metaflammation and immunometabolic disorders. *Nature* 542, 177–185. doi: 10.1038/nature21363
- Howard, J., Lord, G., Matarese, G., Vendetti, S., Ghatei, M., Ritter, M., et al. (1999). Leptin protects mice from starvation-induced lymphoid atrophy and increases thymic cellularity in ob/ob mice. *J. Clin. Invest.* 104, 1051–1059. doi: 10.1172/JCI6762
- Hsu, A., Aronoff, D., Phipps, J., Goel, D., and Mancuso, P. (2007). Leptin improves pulmonary bacterial clearance and survival in ob/ob mice during *Pneumococcal pneumonia*. *Clin. Exp. Immunol.* 150, 332–339. doi: 10.1111/j.1365-2249.2007.03491.x
- Huang, C., Lue, J., Cheng, T., and Tsai, Y. (2020). Glycemic control with insulin attenuates sepsis-associated encephalopathy by inhibiting glial activation via the suppression of the nuclear factor kappa B and mitogen-activated protein kinase signaling pathways in septic rats. *Brain Res.* 1738:146822. doi: 10.1016/j.brainres.2020.146822
- Huizinga, G., Singer, B., and Singer, K. (2020). The Collision of Meta-Inflammation and SARS-CoV-2 Pandemic Infection. *Endocrinology* 161:bqaa154. doi: 10.1210/endo/bqaa154
- Imamura, T., Yanagihara, Y., Ohyagi, Y., Nakamura, N., Inuma, K., Yamasaki, R., et al. (2020). Insulin deficiency promotes formation of toxic amyloid-beta42 conformer co-aggregating with hyper-phosphorylated tau oligomer in an Alzheimer's disease model. *Neurobiol. Dis.* 137:104739. doi: 10.1016/j.nbd.2020.104739
- Iqbal, U., Zeng, E., and Pasinetti, G. (2020). The use of antimicrobial and antiviral drugs in Alzheimer's disease. *Int. J. Mol. Sci.* 21:4920. doi: 10.3390/ijms21144920
- Irie, F., Fitzpatrick, A., Lopez, O., Kuller, L., Peila, R., Newman, A., et al. (2008). Enhanced risk for Alzheimer disease in persons with type 2 diabetes and APOE epsilon4: the cardiovascular health study cognition study. *Arch. Neurol.* 65, 89–93. doi: 10.1001/archneurol.2007.29
- Izquierdo, A., Crujeiras, A., Casanueva, F., and Carreira, M. (2019). Leptin, obesity, and leptin resistance: where are we 25 years later? *Nutrients* 11:2704. doi: 10.3390/nu11112704
- Jeon, C., Haan, M., Cheng, C., Clayton, E., Mayeda, E., Miller, J., et al. (2012). *Helicobacter pylori* infection is associated with an increased rate of diabetes. *Diab. Care* 35, 520–525. doi: 10.2337/dc11-1043
- Jha, M., and Morrison, B. (2018). Glia-neuron energy metabolism in health and diseases: new insights into the role of nervous system metabolic transporters. *Exp. Neurol.* 309, 23–31. doi: 10.1016/j.expneurol.2018.07.009
- Ji, Y., Park, S., Park, H., Hwang, E., Shin, H., Pot, B., et al. (2018). Modulation of active gut microbiota by Lactobacillus rhamnosus GG in a diet induced obesity murine model. *Front. Microbiol.* 9:710. doi: 10.3389/fmicb.2018.00710
- Jiang, C., Li, G., Huang, P., Liu, Z., and Zhao, B. (2017). The gut microbiota and Alzheimer's disease. *J. Alzheimers Dis.* 58, 1–15. doi: 10.3233/JAD-161141
- Jiménez-Cortegana, C., López-Saavedra, A., Sánchez-Jiménez, F., Pérez-Pérez, A., Castiñeiras, J., Virizuela-Echaburu, J., et al. (2021). Leptin, both bad and good actor in Cancer. *Biomolecules* 11:913. doi: 10.3390/biom11060913

- Kanagasigam, S., Chukkappalli, S., Welbury, R., and Singhrao, S. (2020). *Porphyromonas gingivalis* is a strong risk factor for Alzheimer's disease. *J. Alzheimers Dis. Rep.* 4, 501–511. doi: 10.3233/ADR-200250
- Kanoski, S., Zhang, Y., Zheng, W., and Davidson, T. (2010). The effects of a high-energy diet on hippocampal function and blood-brain barrier integrity in the rat. *J. Alzheimers Dis.* 21, 207–219. doi: 10.3233/JAD-2010-091414
- Karman, J., Ling, C., Sandor, M., and Fabry, Z. (2004). Initiation of immune responses in brain is promoted by local dendritic cells. *J. Immunol.* 173, 2353–2361. doi: 10.4049/jimmunol.173.4.2353
- Katsouri, L., Birch, A., Renziehausen, A., Zach, C., Aman, Y., Steeds, H., et al. (2020). Ablation of reactive astrocytes exacerbates disease pathology in a model of Alzheimer's disease. *Glia* 68, 1017–1030. doi: 10.1002/glia.23759
- Kaunzner, U., Miller, M., Gottfried-Blackmore, A., Gal-Toth, J., Felger, J., McEwen, B., et al. (2012). Accumulation of resident and peripheral dendritic cells in the aging CNS. *Neurobiol. Aging* 33, 681–693.e1. doi: 10.1016/j.neurobiolaging.2010.06.007
- Kealy, J., Murray, C., Griffin, E., Lopez-Rodriguez, A., Healy, D., Tortorelli, L., et al. (2020). Acute inflammation alters brain energy metabolism in mice and humans: role in suppressed spontaneous activity, impaired cognition, and delirium. *J. Neurosci.* 40, 5681–5696. doi: 10.1523/JNEUROSCI.2876-19.2020
- Keane, L., Antignano, I., Riechers, S., Zollinger, R., Dumas, A., Offermann, N., et al. (2021). mTOR-dependent translation amplifies microglia priming in aging mice. *J. Clin. Invest.* 131:e132727. doi: 10.1172/JCI132727
- Kellar, D., and Craft, S. (2020). Brain insulin resistance in Alzheimer's disease and related disorders: mechanisms and therapeutic approaches. *Lancet Neurol.* 19, 758–766. doi: 10.1016/S1474-4422(20)30231-30233
- Keller, A., Rohde, J., Raymond, K., and Heitmann, B. (2015). Association between periodontal disease and overweight and obesity: a systematic review. *J. Periodontol.* 86, 766–776. doi: 10.1902/jop.2015.140589
- Khan, S., Barrington, G., Bettiol, S., Barnett, T., and Crocombe, L. (2018). Is overweight/obesity a risk factor for periodontitis in young adults and adolescents: a systematic review. *Obes. Rev.* 19, 852–883. doi: 10.1111/obr.12668
- Khan, S., Luck, H., Winer, S., and Winer, D. (2021). Emerging concepts in intestinal immune control of obesity-related metabolic disease. *Nat. Commun.* 12:2598. doi: 10.1038/s41467-021-22727-22727
- Kierdorf, K., and Prinz, M. (2017). Microglia in steady state. *J. Clin. Invest.* 127, 3201–3209. doi: 10.1172/JCI90602
- Kim, B., Kelschenbach, J., Borjabad, A., Hadas, E., He, H., Potash, M., et al. (2019). Intranasal insulin therapy reverses hippocampal dendritic injury and cognitive impairment in a model of HIV-associated neurocognitive disorders in EcoHIV-infected mice. *AIDS* 33, 973–984. doi: 10.1097/QAD.0000000000002150
- Kim, J., Barua, S., Jeong, Y., and Lee, J. (2020). Adiponectin: the potential regulator and therapeutic target of obesity and Alzheimer's disease. *Int. J. Mol. Sci.* 21:6419. doi: 10.3390/ijms21176419
- Kivipelto, M., Ngandu, T., Laatikainen, T., Winblad, B., Soininen, H., and Tuomilehto, J. (2006). Risk score for the prediction of dementia risk in 20 years among middle aged people: a longitudinal, population-based study. *Lancet Neurol.* 5, 735–741. doi: 10.1016/S1474-4422(06)70537-70533
- Knapp, S. (2013). Diabetes and infection: is there a link?-A mini-review. *Gerontology* 59, 99–104. doi: 10.1159/000345107
- Koh, G., Peacock, S., van der Poll, T., and Wiersinga, W. (2012). The impact of diabetes on the pathogenesis of sepsis. *Eur. J. Clin. Microbiol. Infect. Dis.* 31, 379–388. doi: 10.1007/s10096-011-1337-1334
- Kumar, D., Choi, S., Washicosky, K., Eimer, W., Tucker, S., Ghofrani, J., et al. (2016). Amyloid- β peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci. Transl. Med.* 8:340ra72. doi: 10.1126/scitranslmed.aaf1059
- Kumar, M., Roe, K., Nerurkar, P., Orillo, B., Thompson, K., Verma, S., et al. (2014). Reduced immune cell infiltration and increased pro-inflammatory mediators in the brain of Type 2 diabetic mouse model infected with West Nile virus. *J. Neuroinflamm.* 11:80. doi: 10.1186/1742-2094-11-80
- Kumari, R., Kumar, S., and Kant, R. (2019). An update on metabolic syndrome: metabolic risk markers and adipokines in the development of metabolic syndrome. *Diabetes Metab. Syndr.* 13, 2409–2417. doi: 10.1016/j.dsx.2019.06.005
- Kuo, C., Lu, C., Chang, Y., and Li, C. (2018). Population-Based cohort study on dementia risk in patients with type 1 diabetes mellitus. *Neuroepidemiology* 50, 57–62. doi: 10.1159/000486719
- La Cava, A., and Matarese, G. (2004). The weight of leptin in immunity. *Nat. Rev. Immunol.* 4, 371–379. doi: 10.1038/nri1350
- Labandeira-Garcia, J., Costa-Besada, M., Labandeira, C., Villar-Cheda, B., and Rodríguez-Perez, A. (2017). Insulin-Like growth factor-1 and neuroinflammation. *Front. Aging Neurosci.* 9:365. doi: 10.3389/fnagi.2017.00365
- Lafrance, V., Inoue, W., Kan, B., and Luheshi, G. (2010). Leptin modulates cell morphology and cytokine release in microglia. *Brain Behav. Immun.* 24, 358–365. doi: 10.1016/j.bbi.2009.11.003
- Lana, D., Ugolini, F., and Giovannini, M. (2020). Space-Dependent glia-neuron interplay in the hippocampus of transgenic models of β -amyloid deposition. *Int. J. Mol. Sci.* 21:9441. doi: 10.3390/ijms21249441
- Lana, D., Ugolini, F., Nosi, D., Wenk, G., and Giovannini, M. (2021). The emerging role of the interplay among astrocytes, microglia, and neurons in the hippocampus in health and disease. *Front. Aging Neurosci.* 13:651973. doi: 10.3389/fnagi.2021.651973
- Landrith, T., Sureshchandra, S., Rivera, A., Jang, J., Rais, M., Nair, M., et al. (2017). CD103+ CD8 T cells in the Toxoplasma-infected brain exhibit a tissue-resident memory transcriptional profile. *Front. Immunol.* 8:335. doi: 10.3389/fimmu.2017.00335
- Larabee, C., Neely, O., and Domingos, A. (2020). Obesity: a neuroimmunometabolic perspective. *Nat. Rev. Endocrinol.* 16, 30–43. doi: 10.1038/s41574-019-0283-286
- Lazar, V., Ditu, L., Pircalabioru, G., Gheorghe, I., Curutiu, C., Holban, A., et al. (2018). Aspects of gut microbiota and immune system interactions in infectious diseases, immunopathology, and cancer. *Front. Immunol.* 9:1830. doi: 10.3389/fimmu.2018.01830
- Leblhuber, F., Ehrlich, D., Steiner, K., Geisler, S., Fuchs, D., Lanser, L., et al. (2021). The immunopathogenesis of Alzheimer's disease is related to the composition of gut microbiota. *Nutrients* 13:361. doi: 10.3390/nu13020361
- Lee, G., Proenca, R., Montez, J., Carroll, K., Darvishzadeh, J., Lee, J., et al. (1996). Abnormal splicing of the leptin receptor in diabetic mice. *Nature* 378, 632–635. doi: 10.1038/379632a0
- Leonard, J. (2017). Central nervous system tuberculosis. *Microbiol. Spectr.* 5:11. doi: 10.1128/microbiolspec.TNMI7-0044-2017
- Leshan, R., Louis, G., Jo, Y., Rhodes, C., Münzberg, H., and Myers, M. J. (2009). Direct innervation of GnRH neurons by metabolic- and sexual odorant-sensing leptin receptor neurons in the hypothalamic ventral premammillary nucleus. *J. Neurosci.* 29, 3138–3147. doi: 10.1523/JNEUROSCI.0155-09.2009
- Lester-Coll, N., Rivera, E., Soscia, S., Doiron, K., Wands, J., and de la Monte, S. (2006). Intracerebral streptozotocin model of type 3 diabetes: relevance to sporadic Alzheimer's disease. *J. Alzheimers Dis.* 9, 13–33. doi: 10.3233/jad-2006-9102
- Leyns, C., and Holtzman, D. (2017). Glial contributions to neurodegeneration in tauopathies. *Mol. Neurodegener.* 12:50. doi: 10.1186/s13024-017-0192-x
- Li, Q., and Barres, B. (2018). Microglia and macrophages in brain homeostasis and disease. *Nat. Rev. Immunol.* 18, 225–242. doi: 10.1038/nri.2017.125
- Li, Y., Severance, E., Viscidi, R., Yolken, R., and Xiao, J. (2019). Persistent toxoplasma infection of the brain induced neurodegeneration associated with activation of complement and microglia. *Infect. Immun.* 87:e00139-19. doi: 10.1128/IAI.00139-119
- Libertucci, J., and Young, V. (2019). The role of the microbiota in infectious diseases. *Nat. Microbiol.* 4, 35–45. doi: 10.1038/s41564-018-0278-274
- Liddelov, S., Guttenplan, K., Clarke, L., Bennett, F., Bohlen, C., Schirmer, L., et al. (2017). Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541, 481–487. doi: 10.1038/nature21029
- Linnerbauer, M., Wheeler, M., and Quintana, F. (2020). Astrocyte crosstalk in CNS inflammation. *Neuron* 108, 608–622. doi: 10.1016/j.neuron.2020.08.012
- Lord, G., Matarese, G., Howard, J., Baker, R., Bloom, S., and Lechler, R. (1998). Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. *Nature* 394, 897–901. doi: 10.1038/29795
- Lövheim, H., Olsson, J., Weidung, B., Johansson, A., Eriksson, S., Hallmans, G., et al. (2018). Interaction between cytomegalovirus and herpes simplex virus Type 1 associated with the risk of Alzheimer's disease development. *J. Alzheimers Dis.* 61, 939–945. doi: 10.3233/JAD-161305

- Lue, L., Kuo, Y., Beach, T., and Walker, D. (2010). Microglia activation and anti-inflammatory regulation in Alzheimer's disease. *Mol. Neurobiol.* 41, 115–128. doi: 10.1007/s12035-010-8106-8108
- Lutshumba, J., Nikolajczyk, B., and Bachstetter, A. (2021). Dysregulation of systemic immunity in aging and dementia. *Front. Cell Neurosci.* 15:652111. doi: 10.3389/fncel.2021.652111
- Madinier, A., Bertrand, N., Mossiat, C., Prigent-Tessier, A., Beley, A., Marie, C., et al. (2009). Microglial involvement in neuroplastic changes following focal brain ischemia in rats. *PLoS One* 4:e8101. doi: 10.1371/journal.pone.0008101
- Mallat, M., Lima, F., Gervais, A., Colin, C., and Moura Neto, V. (2002). New insights into the role of thyroid hormone in the CNS: the microglial track. *Mol. Psychiatry* 7, 7–8. doi: 10.1038/sj.mp.4000988
- Mamik, M., Asahchop, E., Chan, W., Zhu, Y., Branton, W., McKenzie, B., et al. (2016). Insulin treatment prevents neuroinflammation and neuronal injury with restored neurobehavioral function in models of HIV/AIDS neurodegeneration. *J. Neurosci.* 36, 10683–10695. doi: 10.1523/JNEUROSCI.1287-16.2016
- Mancuso, P., Gottschalk, A., Phare, S., Peters-Golden, M., Lukacs, N., and Huffnagle, G. (2002). Leptin-deficient mice exhibit impaired host defense in Gram-negative pneumonia. *J. Immunol.* 168, 4018–4024. doi: 10.4049/jimmunol.168.8.4018
- Martella, V., Elia, G., and Buonavoglia, C. (2008). Canine distemper virus. *Vet. Clin. North Am. Small Anim. Pr.* 38, 787–797. doi: 10.1016/j.cvsm.2008.02.007
- Martin, D., Torrance, B., Haynes, L., and Bartley, J. (2021). Targeting aging: lessons learned from immunometabolism and cellular senescence. *Front. Immunol.* 12:714742. doi: 10.3389/fimmu.2021.714742
- Martinez-Gutierrez, M., and Ruiz-Saenz, J. (2016). Diversity of susceptible hosts in canine distemper virus infection: a systematic review and data synthesis. *BMC Vet. Res.* 12:78. doi: 10.1186/s12917-016-0702-z
- Martín-Romero, C., and Sánchez-Margalet, V. (2001). Human leptin activates PI3K and MAPK pathways in human peripheral blood mononuclear cells: possible role of Sam68. *Cell Immunol.* 212, 83–91. doi: 10.1006/cimm.2001.1851
- Matàrese, G., La Cava, A., Sanna, V., Lord, G., Lechler, R., Fontana, S., et al. (2002). Balancing susceptibility to infection and autoimmunity: a role for leptin? *Trends Immunol.* 23, 182–187. doi: 10.1016/s1471-4906(02)02188-2189
- Maurya, R., Bhattacharya, P., Dey, R., and Nakhasi, H. (2018). Leptin functions in infectious diseases. *Front. Immunol.* 9:2741. doi: 10.3389/fimmu.2018.02741
- Mazucanti, C., Liu, Q., Lang, D., Huang, N., O'Connell, J., Camandola, S., et al. (2019). Release of insulin produced by the choroid plexus is regulated by serotonergic signaling. *JCI Insight* 4:e131682. doi: 10.1172/jci.insight.131682
- Mei, F., Xie, M., Huang, X., Long, Y., Lu, X., Wang, X., et al. (2020). *Porphyromonas gingivalis* and its systemic impact: current status. *Pathogens* 9:944. doi: 10.3390/pathogens9110944
- Mejido, D., Peny, J., Vieira, M., Ferreira, S., and De Felice, F. (2020). Insulin and leptin as potential cognitive enhancers in metabolic disorders and Alzheimer's disease. *Neuropharmacology* 171:108115. doi: 10.1016/j.neuropharm.2020.108115
- Merino, B., Díez-Fernández, C., Ruiz-Gayo, M., and Somoza, B. (2006). Choroid plexus epithelial cells co-express the long and short form of the leptin receptor. *Neurosci. Lett.* 393, 269–272. doi: 10.1016/j.neulet.2005.10.003
- Michels, M., Vieira, A., Vuolo, F., Zapelini, H., Mendonça, B., Mina, F., et al. (2015). The role of microglia activation in the development of sepsis-induced long-term cognitive impairment. *Brain Behav. Immun.* 43, 54–59. doi: 10.1016/j.bbi.2014.07.002
- Mitchell, C., and Begg, D. (2021). The regulation of food intake by insulin in the central nervous system. *J. Neuroendocrinol.* 33:e12952. doi: 10.1111/jne.12952
- Mockus, T., Shwetank, Lauver, M., Ren, H., Netherby, C., Salameh, T., et al. (2018). CD4 T cells control development and maintenance of brain-resident CD8 T cells during polyomavirus infection. *PLoS Pathog.* 14:e1007365. doi: 10.1371/journal.ppat.1007365
- Mohanty, P., Hamouda, W., Garg, R., Aljada, A., Ghanim, H., and Dandona, P. (2000). Glucose challenge stimulates reactive oxygen species (ROS) generation by leucocytes. *J. Clin. Endocrinol. Metab.* 85, 2970–2973. doi: 10.1210/jcem.85.8.6854
- Möhle, L., Israel, N., Paarmann, K., Krohn, M., Pietkiewicz, S., Müller, A., et al. (2016). Chronic *Toxoplasma gondii* infection enhances beta-amyloid phagocytosis and clearance by recruited monocytes. *Acta Neuropathol. Commun.* 4:25. doi: 10.1186/s40478-016-0293-298
- Moir, R., Lathe, R., and Tanzi, R. (2018). The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement.* 14, 1602–1614. doi: 10.1016/j.jalz.2018.06.3040
- Montagne, A., Barnes, S., Sweeney, M., Halliday, M., Sagare, A., Zhao, Z., et al. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85, 296–302. doi: 10.1016/j.neuron.2014.12.032
- Monteiro, L., Pereira, J., Palhinha, L., and Moraes-Vieira, P. (2019). Leptin in the regulation of the immunometabolism of adipose tissue-macrophages. *J. Leukoc. Biol.* 106, 703–716. doi: 10.1002/JLB.MR1218-478R
- Morrison, C. (2009). Leptin signaling in brain: a link between nutrition and cognition? *Biochim Biophys. Acta* 1792, 401–408. doi: 10.1016/j.bbadis.2008.12.004
- Muller, L., Gorter, K., Hak, E., Goudzwaard, W., Schellevis, F., Hoepelman, A., et al. (2005). Increased risk of common infections in patients with type 1 and type 2 diabetes mellitus. *Clin. Infect. Dis.* 41, 281–288. doi: 10.1086/431587
- Murray, K., Garcia, M., Rahbar, M., Martinez, D., Khuwaja, S., Arafat, R., et al. (2014). Survival analysis, long-term outcomes, and percentage of recovery up to 8 years post-infection among the Houston West Nile virus cohort. *PLoS One* 9:e102953. doi: 10.1371/journal.pone.0102953
- Myers, M., Cowley, M., and Münzberg, H. (2008). Mechanisms of leptin action and leptin resistance. *Annu. Rev. Physiol.* 70, 537–556. doi: 10.1146/annurev.physiol.70.113006.100707
- Myhre, C., Thygesen, C., Villadsen, B., Vollerup, J., Ilkjaer, L., Krohn, K., et al. (2019). Microglia express insulin-like growth factor-1 in the hippocampus of aged APP swe/PS1 ΔE9 transgenic mice. *Front. Cell Neurosci.* 13:308. doi: 10.3389/fncel.2019.00308
- Nagajyothi, F., Zhao, D., Machado, F., Weiss, L., Schwartz, G., Desruisseaux, M., et al. (2010). Crucial role of the central leptin receptor in murine *Trypanosoma cruzi* infection. *J. Infect. Dis.* 202, 1104–1113. doi: 10.1086/656189
- Nakae, J., Kido, Y., and Accili, D. (2001). Distinct and overlapping functions of insulin and IGF-I receptors. *Endocr. Rev.* 22, 818–835. doi: 10.1210/edrv.22.6.0452
- Naranjo, V., Contreras, A., Merino, B., Plaza, A., Lorenzo, M., García-Cáceres, C., et al. (2020). Specific deletion of the astrocyte leptin receptor induces changes in hippocampus glutamate metabolism, synaptic transmission and plasticity. *Neuroscience* 447, 182–190. doi: 10.1016/j.neuroscience.2019.10.005
- Narayan, O., Herzog, S., Frese, K., Scheefers, H., and Rott, R. (1983). Behavioral disease in rats caused by immunopathological responses to persistent borna virus in the brain. *Science* 220, 1401–1403. doi: 10.1126/science.6602380
- Nascimento, G., Leite, F., Do, L., Peres, K., Correa, M., Demarco, F., et al. (2015). Is weight gain associated with the incidence of periodontitis? a systematic review and meta-analysis. *J. Clin. Periodontol.* 42, 495–505. doi: 10.1111/jcpe.12417
- Nayeri Chegeni, T., Sarvi, S., Moosazadeh, M., Sharif, M., Aghayan, S., Amouei, A., et al. (2019). Is *Toxoplasma gondii* a potential risk factor for Alzheimer's disease? a systematic review and meta-analysis. *Microb. Pathog.* 137:103751. doi: 10.1016/j.micpath.2019.103751
- Nazareth, L., Walkden, H., Chacko, A., Delbaz, A., Shelper, T., Armitage, C., et al. (2021). *Chlamydia muridarum* can invade the central nervous system via the olfactory and trigeminal nerves and infect peripheral nerve glial cells. *Front. Cell Infect. Microbiol.* 10:607779. doi: 10.3389/fcimb.2020.607779
- Netherby-Winslow, C., Ayers, K., and Lukacher, A. (2021). Balancing inflammation and central nervous system homeostasis: T cell receptor signaling in antiviral brain T(RM) formation and function. *Front. Immunol.* 11:624144. doi: 10.3389/fimmu.2020.624144
- Neves, F., Marques, P., Barros-Aragão, F., Nunes, J., Venancio, A., Cozachenko, D., et al. (2018). Brain-Defective insulin signaling is associated to late cognitive impairment in post-septic mice. *Mol. Neurobiol.* 55, 435–444. doi: 10.1007/s12035-016-0307-303
- Ninomiya, Y., Shigemura, N., Yasumatsu, K., Ohta, R., Sugimoto, K., Nakashima, K., et al. (2002). Leptin and sweet taste. *Vitam Horm.* 64, 221–248. doi: 10.1016/s0083-6729(02)64007-64005
- Noriega-Prieto, J., Maglio, L., Zegarra-Valdivia, J., Pignatelli, J., Fernandez, A., Martinez-Rachadell, L., et al. (2021). Astrocytic IGF-IRs induce adenosine-mediated inhibitory downregulation and improve sensory discrimination. *J. Neurosci.* 41, 4768–4781. doi: 10.1523/JNEUROSCI.0005-21.2021

- Obradovic, M., Sudar-Milovanovic, E., Soskic, S., Essack, M., Arya, S., Stewart, A., et al. (2021). Leptin and obesity: role and clinical implication. *Front. Endocrinol.* 12:585887. doi: 10.3389/fendo.2021.585887
- O'Neill, C., Kiely, A., Coakley, M., Manning, S., and Long-Smith, C. (2012). Insulin and IGF-1 signalling: longevity, protein homeostasis and Alzheimer's disease. *Biochem. Soc. Trans.* 40, 721–727. doi: 10.1042/BST20120080
- O'Neill, L., Kishton, R., and Rathmell, J. (2016). A guide to immunometabolism for immunologists. *Nat. Rev. Immunol.* 16, 553–565. doi: 10.1038/nri.2016.70
- Ott, A., Stolk, R., van Harskamp, F., Pols, H., Hofman, A., and Breteler, M. (1999). Diabetes mellitus and the risk of dementia: the rotterdam study. *Neurology* 53, 1937–1942. doi: 10.1212/wnl.53.9.1937
- Paik, I., and Kotler, D. (2011). The prevalence and pathogenesis of diabetes mellitus in treated HIV-infection. *Best Pr. Res. Clin. Endocrinol. Metab.* 25, 469–478. doi: 10.1016/j.beem.2011.04.003
- Pandit, R., Beerens, S., and Adan, R. (2017). Role of leptin in energy expenditure: the hypothalamic perspective. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 312, R938–R947. doi: 10.1152/ajpregu.00045.2016
- Papathanassoglou, E., El-Hashimi, K., Li, X., Matarese, G., Strom, T., and Mantzoros, C. (2006). Leptin receptor expression and signaling in lymphocytes: kinetics during lymphocyte activation, role in lymphocyte survival, and response to high fat diet in mice. *J. Immunol.* 176, 7745–7752. doi: 10.4049/jimmunol.176.12.7745
- Parkhurst, C., Yang, G., Ninan, I., Savas, J., Yates, J. III, Lafaille, J., et al. (2013). Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell* 155, 1596–1609. doi: 10.1016/j.cell.2013.11.030
- Pasarica, M., and Dhurandhar, N. (2007). Infectobesity: obesity of infectious origin. *Adv. Food Nutr. Res.* 52, 61–102. doi: 10.1016/S1043-4526(06)52002-52009
- Patterson, E., Ryan, P., Cryan, J., Dinan, T., Ross, R., Fitzgerald, G., et al. (2016). Gut microbiota, obesity and diabetes. *Postgr. Med J.* 92, 286–300. doi: 10.1136/postgradmedj-2015-133285
- Paz-Filho, G., Babikian, T., Asarnow, R., Delibasi, T., Esposito, K., Erol, H., et al. (2008). Leptin replacement improves cognitive development. *PLoS One* 3:e3098. doi: 10.1371/journal.pone.0003098
- Peila, R., Rodriguez, B., Launer, L., Honolulu-Asia Aging, and Study. (2002). Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: the honolulu-asia aging study. *Diabetes Metab. Res. Rev.* 51, 1256–1262. doi: 10.2337/diabetes.51.4.1256
- Pemberton, L., and Brew, B. (2001). Cerebrospinal fluid S-100beta and its relationship with AIDS dementia complex. *J. Clin. Virol.* 22, 249–253. doi: 10.1016/s1386-6532(01)00196-192
- Pinteaux, E., Inoue, W., Schmidt, L., Molina-Holgado, F., Rothwell, N., and Luheshi, G. (2007). Leptin induces interleukin-1beta release from rat microglial cells through a caspase 1 independent mechanism. *J. Neurochem.* 102, 826–833. doi: 10.1111/j.1471-4159.2007.04559.x
- Poole, S., Singhrao, S., Chukkappalli, S., Rivera, M., Velsko, I., Kesavalu, L., et al. (2015). Active invasion of *Porphyromonas gingivalis* and infection-induced complement activation in ApoE^{-/-} mice brains. *J. Alzheimers Dis.* 43, 67–80. doi: 10.3233/JAD-140315
- Pratap, A., and Holsinger, R. (2020). Altered brain leptin and leptin receptor expression in the 5XFAD mouse model of Alzheimer's disease. *Pharm* 13:401. doi: 10.3390/ph13110401
- Prinz, M., Erny, D., and Hagemeyer, N. (2017). Ontogeny and homeostasis of CNS myeloid cells. *Nat. Immunol.* 18, 385–392. doi: 10.1038/ni.3703
- Procaccini, C., Jirillo, E., and Matarese, G. (2012). Leptin as an immunomodulator. *Mol. Asp. Med.* 33, 35–45. doi: 10.1016/j.mam.2011.10.012
- Procaccini, C., La Rocca, C., Carbone, F., De Rosa, V., Galgani, M., and Matarese, G. (2017). Leptin as immune mediator: interaction between neuroendocrine and immune system. *Dev. Comp. Immunol.* 66, 120–129. doi: 10.1016/j.dci.2016.06.006
- Próchnicki, T., and Latz, E. (2017). Inflammasomes on the crossroads of innate immune recognition and metabolic control. *Cell Metab.* 26, 71–93. doi: 10.1016/j.cmet.2017.06.018
- Prodinger, C., Bunse, J., Krüger, M., Schiefenhövel, F., Brandt, C., Laman, J., et al. (2011). CD11c-expressing cells reside in the juxtavascular parenchyma and extend processes into the glia limitans of the mouse nervous system. *Acta Neuropathol.* 121, 445–458. doi: 10.1007/s00401-010-0774-y
- Pugazhenth, S., Qin, L., and Reddy, P. H. (2017). Common neurodegenerative pathways in obesity, diabetes, and Alzheimer's disease. *Biochim Biophys. Acta Mol. Basis Dis.* 1863, 1037–1045. doi: 10.1016/j.bbdis.2016.04.017
- Pulido-Mendez, M., De Sanctis, J., and Rodriguez-Acosta, A. (2002). Leptin and leptin receptors during malaria infection in mice. *Folia Parasitol.* 49, 249–251. doi: 10.14411/fp.2002.046
- Radigan, K., Morales-Nebreda, L., Soberanes, S., Nicholson, T., Nigdeliogl, R., Cho, T., et al. (2014). Impaired clearance of influenza a virus in obese, leptin receptor deficient mice is independent of leptin signaling in the lung epithelium and macrophages. *PLoS One* 9:e108138. doi: 10.1371/journal.pone.0108138
- Rahman, M., Hossain, K., Das, S., Kundu, S., Adegoke, E., Rahman, M., et al. (2021). Role of insulin in health and disease: an update. *Int. J. Mol. Sci.* 22:6403. doi: 10.3390/ijms22126403
- Ransohoff, R., and Brown, M. (2012). Innate immunity in the central nervous system. *J. Clin. Invest.* 122, 1164–1171. doi: 10.1172/JCI58644
- Rao, R., Li, Q., Odunsi, K., and Shrikant, P. (2010). The mTOR kinase determines effector versus memory CD8⁺ T cell fate by regulating the expression of transcription factors T-bet and Eomesodermin. *Immunity* 32, 67–78. doi: 10.1016/j.immuni.2009.10.010
- Ratter, J., van Heck, J., Rooijackers, H., Jansen, H., van Poppel, P., Tack, C., et al. (2021). Insulin acutely activates metabolism of primary human monocytes and promotes a proinflammatory phenotype. *J. Leukoc Biol.* doi: 10.1002/JLB.3AB0120-019RR Online ahead of print.
- Rayfield, E., Ault, M., Keusch, G., Brothers, M., Nechemias, C., and Smith, H. (1982). Infection and diabetes: the case for glucose control. *Am. J. Med.* 72, 439–450. doi: 10.1016/0002-9343(82)90511-3
- Rebello, C., Kirwan, J., and Greenway, F. (2020). Obesity, the most common comorbidity in SARS-CoV-2: is leptin the link? *Int. J. Obes (Lond)* 44, 1810–1817. doi: 10.1038/s41366-020-0640-645
- Richardson, A., Liu, F., Adamo, M., Van Remmen, H., and Nelson, J. (2004). The role of insulin and insulin-like growth factor-I in mammalian ageing. *Best Pr. Res. Clin. Endocrinol. Metab.* 18, 393–406. doi: 10.1016/j.beem.2004.02.002
- Rodriguez, J., and Delzenne, N. (2021). Modulation of the gut microbiota-adipose tissue-muscle interactions by prebiotics. *J. Endocrinol.* 249, R1–R23. doi: 10.1530/JOE-20-0499
- Romanenko, M., Kholin, V., Koliada, A., and Vaiserman, A. (2021). Nutrition, gut microbiota, and Alzheimer's disease. *Front. Psychiatry* 12:712673. doi: 10.3389/fpsy.2021.712673
- Romani-Pérez, M., Bullich-Vilarrubias, C., López-Almela, I., Liébana-García, R., Olivares, M., and Sanz, Y. (2021). The microbiota and the gut-brain axis in controlling food intake and energy homeostasis. *Int. J. Mol. Sci.* 22:5830. doi: 10.3390/ijms22115830
- Rosko, L., Smith, V., Yamazaki, R., and Huang, J. (2019). Oligodendrocyte bioenergetics in health and disease. *Neuroscientist* 25, 334–343. doi: 10.1177/1073858418793077
- Rummel, C., Inoue, W., Poole, S., and Luheshi, G. (2010). Leptin regulates leukocyte recruitment into the brain following systemic LPS-induced inflammation. *Mol. Psychiatry* 15, 523–534. doi: 10.1038/mp.2009.98
- Salehi, Z., Mashayekhi, F., Naji, M., and Pandamooz, S. (2009). Insulin-like growth factor-1 and insulin-like growth factor binding proteins in cerebrospinal fluid during the development of mouse embryos. *J. Clin. Neurosci.* 16, 950–953. doi: 10.1016/j.jocn.2008.09.018
- Salem, D., Salem, N., and Hendawy, S. (2021). Association between *Toxoplasma gondii* infection and metabolic syndrome in obese adolescents: a possible immune-metabolic link. *Parasitol. Int.* 83:102343. doi: 10.1016/j.parint.2021.102343
- Sánchez-Margalet, V., Martín-Romero, C., Santos-Alvarez, J., Goberna, R., Najib, S., and Gonzalez-Yanes, C. (2003). Role of leptin as an immunomodulator of blood mononuclear cells: mechanisms of action. *Clin. Exp. Immunol.* 133, 11–19. doi: 10.1046/j.1365-2249.2003.02190.x
- Sánchez-Pozo, C., Rodríguez-baño, J., Domínguez-Castellano, A., Muniain, M., Goberna, R., and Sánchez-Margalet, V. (2003). Leptin stimulates the oxidative burst in control monocytes but attenuates the oxidative burst in monocytes from HIV-infected patients. *Clin. Exp. Immunol.* 134, 464–469. doi: 10.1111/j.1365-2249.2003.02321.x
- Sankar, S., Infante-Garcia, C., Weinstock, L., Ramos-Rodriguez, J., Hierro-Bujalance, C., Fernandez-Ponce, C., et al. (2020). Amyloid beta and diabetic

- pathology cooperatively stimulate cytokine expression in an Alzheimer's mouse model. *J. Neuroinflamm.* 17:38. doi: 10.1186/s12974-020-1707-x
- Schafer, D., Lehrman, E., Kautzman, A., Koyama, R., Mardinly, A., Yamasaki, R., et al. (2012). Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron* 74, 691–705. doi: 10.1016/j.neuron.2012.03.026
- Scherer, T., Sakamoto, K., and Buettner, C. (2021). Brain insulin signalling in metabolic homeostasis and disease. *Nat. Rev. Endocrinol.* 17, 468–483. doi: 10.1038/s41574-021-00498-x
- Schertzer, J., and Lam, T. (2021). Peripheral and central regulation of insulin by the intestine and microbiome. *Am. J. Physiol. Endocrinol. Metab.* 320, E234–E239. doi: 10.1152/ajpendo.00547.2020
- Schwemme, M. (2001). Borna disease virus infection in psychiatric patients: are we on the right track? *Lancet Infect. Dis.* 1, 46–52. doi: 10.1016/S1473-3099(01)00021-24
- Scott, M., Lachey, J., Sternson, S., Lee, C., Elias, C., Friedman, J., et al. (2009). Leptin targets in the mouse brain. *J. Comp. Neurol.* 514, 518–532. doi: 10.1002/cne.22025
- Seidel, G., Gaser, C., Götz, T., Günther, A., and Hamzei, F. (2020). Accelerated brain ageing in sepsis survivors with cognitive long-term impairment. *Eur. J. Neurosci.* 52, 4395–4402. doi: 10.1111/ejn.14850
- Shah, B., and Hux, J. (2003). Quantifying the risk of infectious diseases for people with diabetes. *Diabetes Care* 26, 510–513. doi: 10.2337/diacare.26.2.510
- Shaughness, M., Acs, D., Brabazon, F., Hockenbury, N., and Byrnes, K. (2020). Role of insulin in neurotrauma and neurodegeneration: a review. *Front. Neurosci.* 14:547175. doi: 10.3389/fnins.2020.547175
- Shinjo, N., Kagaya, W., and Pekna, M. (2021). Interaction between the complement system and infectious agents - a potential mechanistic link to neurodegeneration and dementia. *Front. Cell Neurosci.* 15:710390. doi: 10.3389/fncel.2021.710390
- Shinjo, N., Parkinson, J., Bell, J., Katsuno, T., and Bligh, A. (2020). Berberine for prevention of dementia associated with diabetes and its comorbidities: a systematic review. *J. Int. Med.* 18, 125–151. doi: 10.1016/j.joim.2020.01.004
- Shippy, D., and Ulland, T. (2020). Microglial immunometabolism in Alzheimer's disease. *Front. Cell Neurosci.* 14:56446. doi: 10.3389/fncel.2020.56446
- Smolina, K., Wotton, C., and Goldacre, M. (2015). Risk of dementia in patients hospitalised with type 1 and type 2 diabetes in England, 1998–2011: a retrospective national record linkage cohort study. *Diabetologia* 58, 942–950. doi: 10.1007/s00125-015-3515-x
- Sochocka, M., Zwolińska, K., and Leszek, J. (2017). The infectious etiology of Alzheimer's disease. *Curr. Neuropharmacol.* 15, 996–1009. doi: 10.2174/1570159X15666170313122937
- Sofroniew, M. (2020). Astrocyte reactivity: subtypes, states, and functions in CNS innate immunity. *Trends Immunol.* 41, 758–770. doi: 10.1016/j.it.2020.07.004
- Solomon, T. (2021). Neurological infection with SARS-CoV-2 - the story so far. *Nat. Rev. Neurol.* 17, 65–66. doi: 10.1038/s41582-020-00453-w
- Sonneville, R., Derese, I., Marques, M., Langouche, L., Derde, S., Chatre, L., et al. (2015). Neuropathological correlates of hyperglycemia during prolonged polymicrobial sepsis in mice. *Shock* 44, 245–251. doi: 10.1097/SHK.0000000000000403
- Sonnwald, U., Wang, A., Schousboe, A., Erikson, R., and Skottner, A. (1996). New aspects of lactate metabolism: IGF-I and insulin regulate mitochondrial function in cultured brain cells during normoxia and hypoxia. *Dev. Neurosci.* 18, 443–448. doi: 10.1159/000111439
- Soscia, S., Kirby, J., Washicosky, K., Tucker, S., Ingelsson, M., Hyman, B., et al. (2010). The Alzheimer's disease-associated amyloid beta-protein is an antimicrobial peptide. *PLoS One* 5:e9505. doi: 10.1371/journal.pone.0009505
- Soto, M., Cai, W., Konishi, M., and Kahn, C. (2019). Insulin signaling in the hippocampus and amygdala regulates metabolism and neurobehavior. *Proc. Natl. Acad. Sci. U S A.* 116, 6379–6384. doi: 10.1073/pnas.1817391116
- Soung, A., and Klein, R. (2018). Viral encephalitis and neurologic diseases: focus on astrocytes. *Trends Mol. Med.* 24, 950–962. doi: 10.1016/j.molmed.2018.09.001
- Souza-Almeida, G., D'Avila, H., Almeida, P., Luna-Gomes, T., Liechocki, S., Walzog, B., et al. (2018). Leptin mediates in vivo neutrophil migration: involvement of tumor necrosis factor-alpha and CXCL1. *Front. Immunol.* 9:111. doi: 10.3389/fimmu.2018.00111
- Spadaro, O., Camell, C., Bosurgi, L., Nguyen, K., Youm, Y., Rothlin, C., et al. (2017). IGF1 shapes macrophage activation in response to immunometabolic challenge. *Cell Rep.* 19, 225–234. doi: 10.1016/j.celrep.2017.03.046
- Spielman, L., Bahniwal, M., Little, J., Walker, D., and Klegeris, A. (2015). Insulin modulates in vitro secretion of cytokines and cytotoxins by human glial cells. *Curr. Alzheimer Res.* 12, 684–693. doi: 10.2174/1567205012666150710104428
- Stephenson, J., Nutma, E., van der Valk, P., and Amor, S. (2018). Inflammation in CNS neurodegenerative diseases. *Immunology* 154, 204–219. doi: 10.1111/imm.12922
- Stewart, C., and Rotwein, P. (1996). Growth, differentiation, and survival: multiple physiological functions for insulin-like growth factors. *Physiol. Rev.* 76, 1005–1026. doi: 10.1152/physrev.1996.76.4.1005
- Sukhanov, S., Higashi, Y., Shai, S., Vaughn, C., Mohler, J., Li, Y., et al. (2007). IGF-1 reduces inflammatory responses, suppresses oxidative stress, and decreases atherosclerosis progression in ApoE-deficient mice. *Arter. Thromb. Vasc. Biol.* 27, 2684–2690. doi: 10.1161/ATVBAHA.107.156257
- Sun, Q., Li, J., and Gao, F. (2014). New insights into insulin: the anti-inflammatory effect and its clinical relevance. *World J. Diabetes* 5, 89–96. doi: 10.4239/wjdv.5.12.89
- Suvan, J., D'Aiuto, F., Moles, D., Petrie, A., and Donos, N. (2011). Association between overweight/obesity and periodontitis in adults: a systematic review. *Obes Rev.* 12, e381–e404. doi: 10.1111/j.1467-789X.2010.00808.x
- Taieb, O., Baleyte, J., Mazet, P., and Fillet, A. (2001). Borna disease virus and psychiatry. *Eur. Psychiatry* 16, 3–10. doi: 10.1016/s0924-9338(00)00529-20
- Takano, K., Koarashi, K., Kawabe, K., Itakura, M., Nakajima, H., Moriyama, M., et al. (2018). Insulin expression in cultured astrocytes and the decrease by amyloid beta. *Neurochem. Int.* 119, 171–177. doi: 10.1016/j.neuint.2017.10.017
- Talbot, K., and Wang, H. (2014). The nature, significance, and glucagon-like peptide-1 analog treatment of brain insulin resistance in Alzheimer's disease. *Alzheimers Dement.* 10, S12–S25. doi: 10.1016/j.jalz.2013.12.007
- Tamara, A., and Tahapary, D. (2020). Obesity as a predictor for a poor prognosis of COVID-19: a systematic review. *Diab. Metab. Syndr.* 14, 655–659. doi: 10.1016/j.dsx.2020.05.020
- Tang, C., Lu, D., Yang, R., Tsai, H., Kao, M., Fu, W., et al. (2007). Leptin-induced IL-6 production is mediated by leptin receptor, insulin receptor substrate-1, phosphatidylinositol 3-kinase, Akt, NF-kappaB, and p300 pathway in microglia. *J. Immunol.* 179, 1292–1302. doi: 10.4049/jimmunol.179.2.1292
- Tian, Z., Sun, R., Wei, H., and Gao, B. (2002). Impaired natural killer (NK) cell activity in leptin receptor deficient mice: leptin as a critical regulator in NK cell development and activation. *Biochem. Biophys. Res. Commun.* 298, 297–302. doi: 10.1016/s0006-291x(02)02462-2462
- Tiedemann, L., Schmid, S., Hettel, J., Giesen, K., Francke, P., Büchel, C., et al. (2017). Central insulin modulates food valuation via mesolimbic pathways. *Nat. Commun.* 8:16052. doi: 10.1038/ncomms16052
- Tien, L., Lee, Y., Pang, Y., Lu, S., Lee, J., Tseng, C., et al. (2017). Neuroprotective effects of intranasal IGF-1 against neonatal lipopolysaccharide-induced neurobehavioral deficits and neuronal inflammation in the substantia nigra and locus coeruleus of juvenile rats. *Dev. Neurosci.* 39, 443–459. doi: 10.1159/000477898
- Tsai, S., Clemente-Casares, X., Zhou, A., Lei, H., Ahn, J., Chan, Y., et al. (2018). Insulin receptor-mediated stimulation boosts T cell immunity during inflammation and infection. *Cell Metab.* 28, 922–934. doi: 10.1016/j.cmet.2018.08.003
- Tschöp, J., Nogueiras, R., Haas-Lockie, S., Kasten, K., Castañeda, T., and Huber, N. (2010). CNS leptin action modulates immune response and survival in sepsis. *J. Neurosci.* 30, 6036–6047. doi: 10.1523/JNEUROSCI.4875-09.2010
- Turnbaugh, P., Ley, R., Mahowald, M., Magrini, V., Mardis, E., and Gordon, J. (2006). An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* 444, 1027–1031. doi: 10.1038/nature05414
- Uysal, K., Wiesbrock, S., Marino, M., and Hotamisligil, G. (1997). Protection from obesity-induced insulin resistance in mice lacking TNF-alpha function. *Nature* 389, 610–614. doi: 10.1038/39335
- Valcour, V., Sacktor, N., Paul, R., Watters, M., Selnes, O., Shiramizu, B., et al. (2006). Insulin resistance is associated with cognition among HIV-1-infected patients: the Hawaii Aging With HIV cohort. *J. Acquir. Immune Defic. Syndr.* 43, 405–410. doi: 10.1097/01.qai.0000243119.67529.f5
- Valcour, V., Shikuma, C., Shiramizu, B., Williams, A., Watters, M., Poff, P., et al. (2005). Diabetes, insulin resistance, and dementia among HIV-1-infected

- patients. *J. Acquir. Immune Defic. Syndr.* 38, 31–36. doi: 10.1097/00126334-200501010-200501016
- Valdebenito, S., Castellano, P., Ajasin, D., and Eugenin, E. (2021). Astrocytes are HIV reservoirs in the brain: a cell type with poor HIV infectivity and replication but efficient cell-to-cell viral transfer. *J. Neurochem.* 158, 429–443. doi: 10.1111/jnc.15336
- Van den Bossche, J., O'Neill, L., and Menon, D. (2017). Macrophage immunometabolism: where are we (going)? *Trends Immunol.* 38, 395–406. doi: 10.1016/j.it.2017.03.001
- van Ginneken, V., Sitnyakowsky, L., and Jeffery, J. (2009). "Infectobesity: viral infections (especially with human adenovirus-36: Ad-36) may be a cause of obesity. *Med. Hypotheses* 72, 383–388. doi: 10.1016/j.mehy.2008.11.034
- van Niekerk, G., Christowitz, C., Conradie, D., and Engelbrecht, A. (2020). Insulin as an immunomodulatory hormone. *Cytokine Growth. Factor. Rev.* 52, 34–44. doi: 10.1016/j.cytogfr.2019.11.006
- van Son, J., Koekkoek, L., La Fleur, S., Serlie, M., and Nieuwdorp, M. (2021). The role of the gut microbiota in the gut-brain axis in obesity: mechanisms and future implications. *Int. J. Mol. Sci.* 22:2993. doi: 10.3390/ijms2206299
- Vangipuram, S., Yu, M., Tian, J., Stanhope, K., Pasarica, M., Havel, P., et al. (2007). Adipogenic human adenovirus-36 reduces leptin expression and secretion and increases glucose uptake by fat cells. *Int. J. Obes (Lond)* 31, 87–96. doi: 10.1038/sj.ijo.0803366
- Vedantama, G., and Viswanathan, V. (2012). Leptin signaling protects the gut from *Entamoeba histolytica* infection. *Gut Microbes* 3, 2–3. doi: 10.4161/gmic.19424
- Viardot, A., Grey, S., Mackay, F., and Chisholm, D. (2007). Potential antiinflammatory role of insulin via the preferential polarization of effector T cells toward a T helper 2 phenotype. *Endocrinology* 148, 346–353. doi: 10.1210/en.2006-2686
- Virto, L., Cano, P., Jiménez-Ortega, V., Fernández-Mateos, P., González, J., Esquifino, A., et al. (2018). Obesity and periodontitis: an experimental study to evaluate periodontal and systemic effects of comorbidity. *J. Periodontol.* 89, 176–185. doi: 10.1902/jop.2017.170355
- Vittor, A., Long, M., Chakrabarty, P., Aycock, L., Kollu, V., and DeKosky, S. (2020). West Nile virus-induced neurologic sequelae—relationship to neurodegenerative cascades and dementias. *Curr. Trop. Med. Rep.* 7, 25–36. doi: 10.1007/s40475-020-00200-207
- Wakim, L., Woodward-Davis, A., Liu, R., Hu, Y., Villadangos, J., Smyth, G., et al. (2012). The molecular signature of tissue resident memory CD8 T cells isolated from the brain. *J. Immunol.* 189, 3462–3471. doi: 10.4049/jimmunol.1201305
- Wang, Y., Li, Y., Zhao, R., Wu, B., Lanoha, B., Tong, Z., et al. (2017). Glutaminase C overexpression in the brain induces learning deficits, synaptic dysfunctions, and neuroinflammation in mice. *Brain Behav. Immun.* 66, 135–145. doi: 10.1016/j.bbi.2017.06.007
- Wasim, M., Awan, F., Najam, S., Khan, A., and Khan, H. (2016). Role of leptin deficiency, inefficiency, and leptin receptors in obesity. *Biochem. Genet.* 54, 565–572. doi: 10.1007/s10528-016-9751-z
- Weilhammer, D., Iavarone, A., Villegas, E., Brooks, G., Sinai, A., and Sha, W. (2012). Host metabolism regulates growth and differentiation of *Toxoplasma gondii*. *Int. J. Parasitol.* 42, 947–959. doi: 10.1016/j.ijpara.2012.07.011
- White, M. (2003). Insulin signaling in health and disease. *Science* 302, 1710–1711. doi: 10.1126/science.1092952
- Whitmer, R., Gunderson, E., Barrett-Connor, E., Quesenberry, C. J., and Yaffe, K. (2005). Obesity in middle age and future risk of dementia: a 27 year longitudinal population based study. *BMJ* 330:1360. doi: 10.1136/bmj.38446.466238.E0
- Whitmer, R., Gunderson, E., Barrett-Connor, E., Quesenberry, C. J., and Yaffe, K. (2018). Obesity trajectories and risk of dementia: 28 years of follow-up in the Whitehall II Study. *Alzheimers Dement.* 14, 178–186. doi: 10.1016/j.jalz.2017.06.2637
- Whitmer, R., Gustafson, D., Barrett-Connor, E., Haan, M., Gunderson, E., and Yaffe, K. (2008). Central obesity and increased risk of dementia more than three decades later. *Neurology* 71, 1057–1064. doi: 10.1212/01.wnl.0000306313.89165.ef
- Widmann, C., and Heneka, M. (2014). Long-term cerebral consequences of sepsis. *Lancet Neurol.* 13, 630–636. doi: 10.1016/S1474-4422(14)70017-70011
- Wieland, C., Florquin, S., Chan, E., Leemans, J., Weijer, S., Verbon, A., et al. (2005). Pulmonary *Mycobacterium tuberculosis* infection in leptin-deficient ob/ob mice. *Int. Immunol.* 17, 1399–1408. doi: 10.1093/intimm/dxh317
- Wilking, A., Elliott, E., Garcia, M., Murray, K., and Munoz, F. (2014). Central nervous system manifestations in pediatric patients with influenza A H1N1 infection during the 2009 pandemic. *Pediatr. Neurol.* 51, 370–376. doi: 10.1016/j.pediatrneurol.2014.04.026
- Wlodarczyk, A., Holtman, I., Krueger, M., Yogev, N., Bruttger, J., Khoroshi, R., et al. (2017). A novel microglial subset plays a key role in myelinogenesis in developing brain. *EMBO J.* 36, 3292–3308. doi: 10.15252/embj.201696056
- Woods, S., Iudicello, J., Dawson, M., Weber, E., Grant, I., and Letendre, S. (2010). HIV-associated deficits in action (verb) generation may reflect astrocytosis. *J. Clin. Exp. Neuropsychol.* 32, 522–527. doi: 10.1080/13803390903264130
- Yano, H., Kinoshita, M., Fujino, K., Nakashima, M., Yamamoto, Y., Miyazaki, H., et al. (2012). Insulin treatment directly restores neutrophil phagocytosis and bactericidal activity in diabetic mice and thereby improves surgical site *Staphylococcus aureus* infection. *Infect. Immun.* 80, 4409–4416. doi: 10.1128/IAI.00787-12
- Yau, B., Hunt, N., Mitchell, A., and Too, L. (2018). Blood-Brain barrier pathology and CNS outcomes in *Streptococcus pneumoniae* meningitis. *Int. J. Mol. Sci.* 19:3555. doi: 10.3390/ijms19113555
- Yki-Järvinen, H., Sammalkorpi, K., Koivisto, V., and Nikkilä, E. (1989). Severity, duration, and mechanisms of insulin resistance during acute infections. *J. Clin. Endocrinol. Metab.* 69, 317–323. doi: 10.1210/jcem-69-2-317
- Zawada, A., Rychter, A., Ratajczak, A., Lisiecka-Masian, A., Dobrowolska, A., and Krela-Kazmierczak, I. (2020). Does gut-microbiome interaction protect against obesity and obesity-associated metabolic disorders? *Microorganisms* 9:18. doi: 10.3390/microorganisms9010018
- Zemva, J., and Schubert, M. (2011). Central insulin and insulin-like growth factor-1 signaling: implications for diabetes associated dementia. *Curr. Diab. Rev.* 7, 356–366. doi: 10.2174/157339911797415594
- Zhang, A., To, K., Li, C., Lau, C., Poon, V., Chan, C., et al. (2013). Leptin mediates the pathogenesis of severe 2009 pandemic influenza A (h1N1) infection associated with cytokine dysregulation in mice with diet-induced obesity. *J. Infect. Dis.* 207, 1270–1280. doi: 10.1093/infdis/jit031
- Ziegler, A., Levison, S., and Wood, T. (2015). Insulin and IGF receptor signalling in neural-stem-cell homeostasis. *Nat. Rev. Endocrinol.* 11, 161–170. doi: 10.1038/nrendo.2014.208
- Ziegler, A., Schneider, J., Qin, M., Tyler, W., Pintar, J., Fraidenaich, D., et al. (2012). IGF-II promotes stemness of neural restricted precursors. *Stem Cells* 30, 1265–1276. doi: 10.1002/stem.1095

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Shinjo and Kita. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.