



# Accelerated Ovarian Failure as a Unique Model to Study Peri-Menopause Influence on Alzheimer's Disease

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Despite decades of extensive research efforts, efficacious therapies for Alzheimer's disease (AD) are lacking. The multi-factorial nature of AD neuropathology and symptomatology has taught us that a single therapeutic approach will most likely not fit all. Women constitute ~70% of the affected AD population, and pathology and rate of symptoms progression are 2–3 times higher in women than men. Epidemiological data suggest that menopausal estrogen loss may be causative of the more severe symptoms observed in AD women, however, results from clinical trials employing estrogen replacement therapy are inconsistent. AD pathological hallmarks—amyloid  $\beta$  ( $A\beta$ ), neurofibrillary tangles (NFTs), and chronic gliosis—are laid down during a 20-year prodromal period before clinical symptoms appear, which coincides with the menopause transition (peri-menopause) in women (~45–54-years-old). Peri-menopause is marked by widely fluctuating estrogen levels resulting in periods of irregular hormone-receptor interactions. Recent studies showed that peri-menopausal women have increased indicators of AD phenotype (brain  $A\beta$  deposition and hypometabolism), and peri-menopausal women who used hormone replacement therapy (HRT) had a reduced AD risk. This suggests that neuroendocrine changes during peri-menopause may be a trigger that increases risk of AD in women. Studies on sex differences have been performed in several AD rodent models over the years. However, it has been challenging to study the menopause influence on AD due to lack of optimal models that mimic the human process. Recently, the rodent model of accelerated ovarian failure (AOF) was developed, which uniquely recapitulates human menopause, including a transitional peri-AOF period with irregular estrogen fluctuations and a post-AOF stage with low estrogen levels. This model has proven useful in hypertension and cognition studies with wild type animals. This review article will highlight the molecular mechanisms by which peri-menopause may influence the female brain vulnerability to AD and AD risk factors, such as hypertension and apolipoprotein E (APOE) genotype. Studies on these biological mechanisms together with the use of the AOF model have the potential to shed light on key molecular pathways underlying AD pathogenesis for the development of precision medicine approaches that take sex and hormonal status into account.

**Keywords:** Alzheimer's disease, menopause, peri-menopause, accelerated ovarian failure, ovariectomy (OVX), reproductive senescence

## INTRODUCTION

Alzheimer's disease (AD) is an irreversible, progressive neurodegenerative disorder and a leading cause of mortality (Alzheimer's Association, 2016; Scheltens et al., 2016). AD accounts for approximately 60%–80% of dementia cases in older adults with an average age at onset of around 65 years of age (Alzheimer's Association, 2016). Symptoms are characterized by memory loss and impairment in other non-memory aspects of cognitive function such as word-finding, vision/spatial issues, and impaired reasoning or judgment. This loss of cognitive functioning along with loss of behavioral abilities are extremely debilitating for AD patients and dramatically interfere with their daily life (Scheltens et al., 2016).

Neuropathologically, AD is characterized by progressive deposition of parenchymal senile plaques comprised of amyloid- $\beta$  (A $\beta$ ) protein, intracellular neurofibrillary tangles (NFTs) of abnormal phosphorylated tau, and chronic gliosis primarily in the hippocampus and neocortex (Price et al., 1991; Braak and Braak, 1997; Montine et al., 2012; Au et al., 2016; Scheltens et al., 2016; Lane et al., 2018), brain areas critical for learning and memory. This leads to synaptic damage, cognitive impairment, and neurodegeneration (Canter et al., 2016; Scheltens et al., 2016).

Cause(s) of AD are still unknown but several gene mutations, as well as other genetic and modifiable risk factors, have been identified. Age is the major risk factor for sporadic AD. As the number of aging Americans rapidly increases, so does the incidence of AD. An estimated 47 million people are currently living with AD worldwide, with 5.8 million people only in the US. This number is projected to triple by 2,050 to over 16 million (Alzheimer's Association, 2016; Deb et al., 2017; Nebel et al., 2018). The ongoing demographic changes as well as the lack of treatments and preventive strategies will contribute not only to the rising of AD prevalence, but also to the increasing medical costs associated with the disease, which will exceed \$1 trillion (Alzheimer's Association, 2016; Deb et al., 2017; Nebel et al., 2018).

Although remarkable research advancements were made in the past decade, the complexity of the cellular mechanisms underlying AD pathogenesis and neuropathology as well as the heterogeneity of symptoms among AD patients caused many clinical therapeutic trials to fail. Understanding the multifactorial nature of AD pathogenesis and pathology is a crucial step to develop efficacious prevention and treatment strategies, and to optimize care and reduce high costs associated with the disease.

Women constitute nearly 70% of the affected AD population (Fisher et al., 2018), ~3.5 million Americans aged 65 or older. AD prevalence is 2–3 times higher in post-menopausal women than men, even after controlling for lifespan (Alzheimer's Association, 2016; Laws et al., 2016; Scheltens et al., 2016; Fisher et al., 2018). Furthermore, women present with a faster pathology progression and greater memory impairment than men (Alzheimer's Association, 2016; Laws et al., 2016; Fisher et al., 2018). Despite the established vulnerability, the biological mechanisms underlying the increased risk of AD in

women are largely unknown. Clinical data suggest that loss of estrogen at menopause may be a main factor influencing susceptibility to AD, but results from clinical trials employing estrogen replacement therapy in post-menopausal women are inconsistent (Alzheimer's Association, 2016; Fisher et al., 2018). Nevertheless, results from recent clinical studies have suggested that hormone replacement therapy (HRT) initiated during peri-menopause may lower the risk of dementia and have cognitive benefits (Henderson et al., 2005; Whitmer et al., 2011; Shao et al., 2012).

Pre-clinical rodent models have proven to be important for understanding the wide range of estrogen effects that occur during human menopause. Rodent models have been constructed to replicate different elements of human menopause. These include natural aging, ovariectomy (OVX) and hormone replacement, and genetic models (Marques-Lopes et al., 2018). Although the current rodent models of menopause have and will continue to inform our understanding of the role of both estrogen and HRT in menopause, they fail to adequately recapitulate the human menopause process. The intact aging model fails to achieve very low estrogen levels, and the OVX model lacks a stage mimicking peri-menopause. Among the pre-clinical models, the innovative accelerated ovarian failure (AOF) using the chemical 4-vinylcyclohexene diepoxide (VCD), uniquely recapitulates hormonal changes that occur during human menopause, including estrous acyclicity and fluctuation (as in human peri-menopause), followed by undetectable, estrogen levels (as in human post-menopause). These model also allow for the dissociation of the effects of hormone levels from the effects of aging in young animals (reviewed in Van Kempen et al., 2011).

Over the last 20 years, striking advancements have been made in our understanding of AD sex dimorphism. The overarching goal of this review article is to highlight recent advancements in understanding the molecular mechanisms by which menopause may influence the female brain vulnerability to AD in view of the rodent models of menopause available. Studies on these biological mechanisms in combination with the use of the AOF model have the potential to shed light on key signaling pathways with the potential to improve diagnosis, prevention or stabilization of risk factors, and clinical outcomes.

## THE HUMAN MENOPAUSE PROCESS

Menopause, or cessation of menstrual cycling, is a uniquely human process and marks the beginning of women's reproductive senescence (Walker and Herndon, 2008; Alberts et al., 2013). The majority of women enter menopause *via* a gradual and irreversible process (peri-menopause) of reduction in ovarian function and decline in estrogen levels followed by a decrease in estrogen receptor (ER) expression over several years. Duration of peri-menopause is ~5 years, between ages of 45 and 54, which is followed by amenorrhea and then post-menopause (Practice Committee of the American Society for Reproductive Medicine, 2008; Butler and Santoro, 2011; Harlow et al., 2012). Peri-menopause is marked by irregular

estrous cycles and fluctuations in ovarian hormones, but not loss of estrogen levels which are typical of post-menopause (Harsh et al., 2007). This period of irregular hormone-receptor imbalance contributes to the physiological and psychological symptoms associated with menopause (Morrison et al., 2006; Practice Committee of the American Society for Reproductive Medicine, 2008). Although clinically menopause is primarily defined as reproductive senescence, biological changes occur that significantly alter brain function which is reflected by a range of neurological symptoms including depression, insomnia, hypertension, and cognitive dysfunction (Morrison et al., 2006; Practice Committee of the American Society for Reproductive Medicine, 2008).

With current age of menopause transition around 50 years and life expectancy getting close to 80 years, it is estimated that by 2,060 20%–30% of women in the US will be in post-menopause, meaning that a woman will spend approximately 1/3 of her life in post-menopause (U.S. Census Bureau, 2017). Mounting evidences indicate that menopause constitutes a risk factor for developing AD and newly published work suggests that peri-menopause, as a neurological transition state, may be the key time for the female brain susceptibility to the disease (Brinton et al., 2015).

## MENOPAUSE INFLUENCE ON CLINICAL PRESENTATION OF ALZHEIMER'S DISEASE

There is epidemiological, clinical, and biological evidence that sex dimorphism influences the onset, progression, and clinical manifestation of AD (Mazure and Swendsen, 2016; Fisher et al., 2018; Nebel et al., 2018). It is recognized that aging is the major risk factor for AD (Mielke et al., 2014; Alzheimer's Association, 2016). Although, women live longer than men, they carry an increased life-long risk of being diagnosed with AD even after adjusting for age and lifespan (Barron and Pike, 2012; Vest and Pike, 2013; Alzheimer's Association, 2016). AD women tend to exhibit a broader spectrum of dementia-related behavioral symptoms and experience greater cognitive deterioration than men in the progression of the disease (Schmidt et al., 2008; Chapman et al., 2011; Mazure and Swendsen, 2016; Nebel et al., 2018). Although men with AD have a shorter survival time (Burns et al., 1991; Todd et al., 2013; Kua et al., 2014; Wattmo et al., 2014a,b), a meta-analysis of neurocognitive data from 15 published studies revealed that women with AD showed a consistent worse mental deterioration than men with the disease, even when at the same stage of the condition (Irvine et al., 2012). Moreover, AD pathology appears more likely to be clinically expressed as dementia in women than in men (Barnes et al., 2005; Lin and Doraiswamy, 2014).

There are multiple biological hypotheses by which female sex may affect AD. As recently reviewed by the Society for Women's Health Research Interdisciplinary Network on AD, these could be represented by: (1) genetic factors that have a stronger effect in women [i.e., apolipoprotein E (APOE) genotype]; (2) risk factors that differentially affect men and women (i.e., hypertension); or

(3) biological events that are uniquely experienced by women (i.e., menopause, pregnancy; Nebel et al., 2018).

Clinical data suggest that neurological consequences of menopause may trigger more severe symptoms and pathology observed in women with AD compared to men (Barnes et al., 2005; Hua et al., 2010; Skup et al., 2011; Hall et al., 2012; Holland et al., 2013; Lin et al., 2015; Ball and Chen, 2016; Filon et al., 2016; Sundermann et al., 2016a,b, 2017; Jack et al., 2017; Koran et al., 2017; Fisher et al., 2018; Buckley et al., 2019; reviewed in Li and Singh, 2014; McCarrey and Resnick, 2015; Georgakis et al., 2016; Hampel et al., 2018). In support of this, early surgical menopause was associated in women with a 2-fold increase in dementia risk, faster rate of cognitive decline, and more AD pathology (Bove et al., 2014; Fisher et al., 2018). Randomized clinical trials and observational studies found menopause, and particularly the peri-menopause phase, associated with decline in memory function, and increase risk of mild cognitive impairment and dementia (Joffe et al., 2006; Morrison et al., 2006; Greendale et al., 2009; Epperson et al., 2013). Findings from two longitudinal studies reported that peri-menopause was associated with decreased cognitive performance compared to not only pre-menopause, but also post-menopause (Greendale et al., 2009; Epperson et al., 2013), suggesting that the memory impairment was temporary during menopause transition. Recent cross-sectional studies, however, found impairment in verbal memory also in post-menopausal women (Jacobs et al., 2016, 2017; Rentz et al., 2017).

AD pathological hallmarks—A $\beta$  plaques, NFTs, chronic gliosis, and neurodegeneration (Hampel et al., 2018)—are laid down during a prodromal period beginning ~20 years before clinical symptoms appear, which coincides with the time of peri-menopause in women (Jack et al., 2013; Dubois et al., 2016).

Clinical findings suggest that neurological changes during peri-menopause increase the female brain vulnerability to AD. Accordingly, two recent amyloid-PET and magnetic resonance imaging (MRI) studies on cognitively normal women reported increased indicators of AD phenotype—A $\beta$  deposition in frontal and temporal cortex, hypometabolism, and reduced brain volume in AD-vulnerable regions—starting at peri-menopause (Mosconi et al., 2017b) compared to pre-menopause and male sex (Mosconi et al., 2017a,b). This suggests that irregular estrogen fluctuations and neuroendocrine changes unique to peri-menopause, rather than the loss of estrogen at post-menopause, may be the triggering events that increase the susceptibility to AD risk accelerating AD neuropathology and cognitive decline in women.

Observational data link the use of HRT with lower AD risk in women, but clinical trials employing HRT have produced many inconsistent results, largely due to certain caveats such as hormone formulation, timing of therapy, and dose or route of hormone administration as well as studies limited to post-menopausal women (McCarrey and Resnick, 2015; Georgakis et al., 2016).

Ovaries produce and secrete several types of estrogens, including estrone (E1), 17 $\alpha$ - and 17 $\beta$ -estradiol (E2), and estriol (E3). 17 $\beta$ -estradiol is the most abundant and potent female gonadal hormone based on binding activity to ERs

(Folmar et al., 2002; Blaustein, 2008; Koebele and Bimonte-Nelson, 2015). Most human studies, but not all, have used the HRT formulation consisting in Conjugated Equine Estrogens (CEE; Hersh et al., 2004), which upon absorption and metabolism are primarily converted into 17 $\beta$ -estradiol and equilin (Sitruk-Ware, 2002; Bhavnani, 2003).

The initial results from controlled clinical trials, including the Women's Health Initiative (WHI), WHI Memory Study (WHIMS), and WHI Study of Cognitive Aging (WHISCA) found that HRT in women may lead to no, or even adverse, effects on cognition and AD risk (Shumaker et al., 2003, 2004; Maki and Henderson, 2012, 2016; Gurney et al., 2014; Hampel et al., 2018). Administration of CEE and progestin medroxy-progesterone acetate after a prolonged period of hypogonadism or menopause diminishes the neuroprotective effect of HRT and enhances neuroinflammation (Shumaker et al., 2003, 2004). Data from the recent WHI publication on the effect of HRT on mortality reported lower risk of dying from AD and dementia for women receiving estrogen, but not estrogen in combination with progestin (Manson et al., 2017). To date, there is no evidence to support that HRT initiated during early post-menopause confer cognitive benefit, but it appears to be safe for cognitive function (Espeland et al., 2013; Gleason et al., 2015; Georgakis et al., 2016; Henderson et al., 2016). However, recent studies have shown that HRT initiated during peri-menopause lowers the risk of AD and has cognitive benefits, especially to hippocampal-mediated memory processes (Henderson et al., 2005; Whitmer et al., 2011; Shao et al., 2012). In the Cache County Memory Study, peri-menopausal women who used HRT had a reduced risk of AD later in life (Zandi et al., 2002; Shao et al., 2012). Data from the WHIMS-Young study and the Kronos Early Estrogen Prevention Study (KEEPS), which only enrolled early menopausal women, support the beneficial effect of HRT on cognitive function early in the menopause transition process (McCarrey and Resnick, 2015). Therefore, the "Window of opportunity" hypothesis was formulated that the beneficial effect of estrogen on cognition and AD depends on women's age and stage of menopause, and HRT started during post-menopause when a new hormone-receptor equilibrium is already achieved may disturb the established balance (Resnick and Henderson, 2002; Alzheimer's Association, 2016; Pines, 2016; Pike, 2017; Fisher et al., 2018). Similarly, the "Healthy Cell Bias of estrogen action" hypothesis has been proposed (Brinton, 2008; Gillies and McArthur, 2010). The healthy cell bias postulates that, as cognitive health declines over women's lifetime, so are the benefits of HRT treatment on the brain. Specifically, the effects of HRT on cognitive function may progress from beneficial to neutral or deleterious over time. In this scenario, estrogen is neuroprotective if neurons are healthy at time of HRT administration. In contrast, estrogen exposure negatively affects neurological functions if neuronal health is compromised. This may explain the initial results from the different WHI studies where HRT administered later in life (and after menopause) had neutral or adverse effects on cognition. Since it is more likely that neurons during peri-menopause are healthier than at post-menopause, the

window of opportunity and healthy cell bias hypotheses are inter-related. Yet, the critical cellular and molecular mechanisms underlying the influence of peri-menopause are still awaiting a clear understanding.

## MOLECULAR MECHANISMS FOR THE EFFECT OF PERI-MENOPAUSE ON AD BRAIN

Estrogens exert their physiological and pathological actions mainly through nuclear estrogen receptors (nERs—ER $\alpha$ , ER $\beta$ ), and membrane estrogen receptors (mERs) which include ER $\alpha$ , ER $\beta$ , G-protein coupled receptor 30 (GPER1 aka GPER30), ER-X, Gq-coupled membrane receptor (Gq-mER; Milner et al., 2001, 2005; McEwen et al., 2012; Hara et al., 2015; Korol and Pisani, 2015; McEwen and Milner, 2017). Each brain region possesses a specific ERs gene expression profile, and activity of each ER is differentially regulated by estrogen, which may account for the sex brain dimorphism (Waters et al., 2009; Mitterling et al., 2010; Li and Singh, 2014). Acting *via* genomic and non-genomic signaling pathways, estrogen has long been known to regulate brain function in both males and females *via* multiple mechanisms of action (Cui et al., 2013). For a review of estrogen actions on the brain, see Spencer et al. (2008); Gillies and McArthur (2010); Luine (2014); Frick (2015); Koebele and Bimonte-Nelson (2017) and McEwen and Milner (2017). In the interest of space, this review article will only discuss biological mechanisms that are related to menopause and AD. For a review of studies on sex differences in mouse models of AD, see Gillies and McArthur (2010); Dubal et al. (2012) and Li et al. (2014).

Changes in estrogen levels during menopause have been associated with mitochondrial dysfunction, synaptic decay, and neuroinflammation. Localization of ERs directly within mitochondria suggests that estrogen may regulate **mitochondrial activity** *via* genomic mechanism, but also affect its function directly by modulating mitochondrial respiration and mitochondrial DNA transcription (Klinge, 2017). This is particularly relevant in the brain as mitochondrial dysfunction plays a role in aging and neurodegenerative diseases, specifically in AD (Barja, 2004; Cantuti-Castelvetri et al., 2005; Kujoth et al., 2007). In the 3xTgAD mouse model of AD, Dr. Brinton lab showed that there is a significant correlation between decreased mitochondrial and glucose metabolism and A $\beta$  load in the hippocampus of aged or ovariectomized OVX females with respect to intact females (Brinton, 2008; Yao et al., 2010, 2012; Ding et al., 2013a,b). However, the hypometabolism in the hippocampus of aged and OVX females was observed across all ages, and in both 3xTgAD and non-transgenic mice suggesting that these mechanisms may influence AD, but are not exclusive to it (Yao et al., 2012; Ding et al., 2013a). Accordingly, pre-clinical and human brain imaging studies showed that A $\beta$  and tau pathology precede metabolic changes and cognitive deficits in AD (Landau et al., 2012; Jack et al., 2013), supporting the idea that mechanisms other than hypometabolism may act during menopause to accelerate the initiation of AD.



Another possible mechanism for menopause effect on AD may be through the estrogen modulation of **neuronal excitability and synaptic plasticity** (Spencer et al., 2008; Spencer-Segal et al., 2012; Baudry et al., 2013; Arevalo et al., 2015; Hara et al., 2015; Waters et al., 2015), which are major mechanisms underlying learning and mnemonic processes. Indeed, high levels of 17 $\beta$ -estradiol during the proestrus phase of the estrous cycle increases hippocampal excitability, long term potentiation (LTP), and remodels dendritic spines in female rodents (Woolley and McEwen, 1992; McEwen et al., 2001; Spencer et al., 2008; Mukai et al., 2010; Broestl et al., 2018). This suggests that irregular estrogen fluctuations during peri-menopause and the following decline in estrogen levels may profoundly impact neuronal activity in females. The effects of menopause transition on neuronal activity may also be related to the estrogen action on neurotrophins, including brain-derived neurotrophic factor (BDNF; Spencer et al., 2008, 2010; Spencer-Segal et al., 2011; Wei et al., 2017), which is an important regulator of synaptic plasticity in the brain (Fisher et al., 2018). BDNF is neuroprotective against A $\beta$ -induced cell death, and its levels are reduced in hippocampus and serum of individuals with AD (Laske et al., 2006a,b; Arancibia et al., 2008; Tapia-Arancibia et al., 2008; Nagahara et al., 2009; Nagahara and Tuszynski, 2011). Interestingly, recent studies on carriers of the BDNF single-nucleotide polymorphism Val66Met showed an increase in AD risk in women but not in men, and that women respond differently to estrogen in hippocampal-related working memory tasks compared to non-carrier controls (Wei et al., 2017, 2018; Fisher et al., 2018). Serum levels of BDNF significantly decrease in menopause women and during aging. In rodent models, aging and OVX induce a significant reduction in hippocampal BDNF expression which is ameliorated by HRT.

Other than affecting neurophysiology in higher cognitive brain regions—hippocampus, frontal cortex, basal forebrain, and striatum—estrogen also influences the neuropathology of AD including seeding of parenchymal A $\beta$  plaques, intracellular NFTs, and chronic inflammation (Brinton, 2008; Hirata-Fukae et al., 2008; Lee et al., 2014; Au et al., 2016; McEwen and Milner, 2017; Merlo et al., 2017; Fisher et al., 2018).

Amyloid precursor protein (APP) is processed by two competing pathways, the non-amyloidogenic pathway *via*  $\alpha$ -secretase, and the amyloidogenic *via*  $\beta$ -secretase (BACE1), which produces the **toxic  $\beta$ -APPs and A $\beta$ 40/A $\beta$ 42 peptides** (Baranello et al., 2015). Estrogen can reduce A $\beta$  deposits by favoring the non-amyloidogenic pathway and reducing BACE1 levels, promoting A $\beta$  glial phagocytosis, and regulating the major enzymes involved in A $\beta$  degradation (Jaffe et al., 1994; Xu et al., 1998, 2006; Li et al., 2000; Manthey et al., 2001; Levin-Allerhand et al., 2002; Joffe et al., 2006; Yao et al., 2007; Liang et al., 2010; Zhao et al., 2011; Lee et al., 2014). In hippocampus and prefrontal cortex (PFC) of AD patients, estrogen loss exacerbates deposition of A $\beta$  and NFTs, which induces synaptic dysfunction (Pike, 2017). Following OVX, levels of A $\beta$  peptide and A $\beta$  plaque burden are increased in different transgenic mouse models of AD in females compared to

males, and are rescued by estradiol treatment (Callahan et al., 2001; Levin-Allerhand et al., 2002; Zheng et al., 2002; Carroll et al., 2007, 2010). In the 3xTgAD mouse model there is a significant correlation between decreased mitochondrial and glucose metabolism and A $\beta$  load in the hippocampus of aged or OVX females with respect to intact females (Yao et al., 2010, 2012; Ding et al., 2013a; Ding et al., 2013b).

Deposition of NTFs **containing hyperphosphorylated tau protein** may also be an early event during the prodromal phase of AD, which correlates significantly with cognitive symptoms later in life (Nelson et al., 2009; Jack et al., 2013; Wang and Mandelkow, 2016). In the presence of brain tau pathology, women have an increased risk of AD compared to men (Barnes et al., 2005). In mouse models overexpressing mutant tau, females show higher levels of tau pathology and more severe cognitive deficits compared to males (Asuni et al., 2007; Yue et al., 2011; Buccarello et al., 2017), although the animal's ages spanned from young to old (5–15 months). This discrepancy may be explained by the mouse models used or by the concurrence of estrogen-independent mechanisms. Most pre-clinical studies agree that estradiol reduces levels of hyperphosphorylated tau *via* ERs, though opposing roles for ER $\alpha$  and ER $\beta$  were reported (Merlo et al., 2017).

Several lines of evidence support the pivotal role of **chronic inflammation** in the initiation and progression of AD (Frautschy et al., 1998; Benzing et al., 1999; Parachikova et al., 2007; Kraft et al., 2013; Christensen and Pike, 2015; Heneka et al., 2015; Matarin et al., 2015; Au et al., 2016; Song et al., 2016; Villa et al., 2016; Wang et al., 2016; Czirr et al., 2017; Jevtic et al., 2017; Liu et al., 2017). An increase in density of resting microglia precedes A $\beta$  plaque formation, and increased activated microglia is observed around A $\beta$  plaques in the brain of transgenic models of AD (Heneka et al., 2015; Wang et al., 2015; Manocha et al., 2016; Jevtic et al., 2017). Inflammation is another risk factor for AD that varies by sex (Hanamsagar and Bilbo, 2016). The expression of genes related to inflammation increases with age more in the forebrain of menopausal women than men and pre-menopausal women (Sárvári et al., 2012; Christensen and Pike, 2015). Acting *via* ER $\alpha$  and ER $\beta$  on glial cells, estrogen regulates glial response to neurotoxins and promotes A $\beta$  removal (Li et al., 2000; Blurton-Jones and Tuszynski, 2001; Au et al., 2016; Villa et al., 2016; Merlo et al., 2017). At menopause, estrogen level fluctuations affect conversion of microglia phenotype from resting state to reactive state (Sárvári et al., 2012). In rodents, aging or OVX induce changes in ER $\alpha$  and ER $\beta$  levels on astrocytes which leads to impaired neurotrophic responses to estrogen (Rozovsky et al., 2002; McAsey et al., 2006; Morgan and Finch, 2015). Accordingly, reactive astrogliosis and microglia reactivity marker genes are up-regulated in OVX female mice (Sárvári et al., 2012).

Furthermore, besides directly influencing brain structures regulating cognitive function, hormonal and neurological changes during menopause transition may indirectly influence other risk factors that differentially affect men and women, such as APOE genotype and hypertension (see also paragraph on

the use of VCD mouse model to study hypertension; Nebel et al., 2018). For instance, the most well-established genetic risk factors for late onset AD is the presence of a common allele  $\epsilon 4$  in the **APOE gene** (Corder et al., 1993; Coon et al., 2007), which encodes for a lipid-binding protein crucial in triglycerides and cholesterol transport to neurons (Puglielli et al., 2003; Bu, 2009; Leduc et al., 2010). APOE4 genotype increases risk of developing AD over the other alleles APOE2 and APOE3 by 5–10 years (Noguchi et al., 1993; Ossenkoppele et al., 2015; Fisher et al., 2018), and was reported to increase A $\beta$  deposition and oligomer formation, as well as phosphorylated tau within neurons (Riedel et al., 2016; Fisher et al., 2018). The APOE4 allele shows sex-dependent effects whereby it is a stronger risk factor for AD in female than male carriers of same age (Farrer et al., 1997; Mortensen and Hogh, 2001; Beydoun et al., 2012; Altmann et al., 2014; Neu et al., 2017). APOE4 women also show a sharper decline in cognitive function and have a greater A $\beta$  brain pathology than APOE4 men (Mortensen and Hogh, 2001; Corder et al., 2004; Beydoun et al., 2012; Fisher et al., 2018). *In vitro* and *in vivo*, activation of ER $\alpha$  up-regulates levels of ApoE mRNA and protein, whereas selective ER agonists down-regulate ApoE mRNA and protein levels in rat hippocampal neurons (Wang et al., 2006). Altogether, this may suggest that HRT could constitute a possible approach to provide therapeutic benefits and/or reduce the risk of developing AD in APOE4 carriers. However, studies on the use of estrogen formulations have provided inconclusive results thus far highlighting a complex interaction between estrogen, APOE and AD (Depypere et al., 2016; Riedel et al., 2016).

In conclusion, there is enough evidence to support that widely fluctuating changes in estrogen-ER response network may underlie the increased susceptibility of female brains to AD conferred by menopause transition. In many women, the brain compensates for these changes during peri-menopause. However, it is likely that in the presence of other AD risk factors for some women this adaptive compensation is diminished, which gives rise to the increased the susceptibility to AD pathogenesis.

## RODENT MODELS OF MENOPAUSE

Despite continuous advances in understanding AD pathophysiology, it has been challenging to systematically evaluate the biological mechanisms underlying the influence of menopause on AD in the human population, and clinical research continues to have many inconsistent results and unresolved issues. Pre-clinical rodent models of menopause have proven useful to start teasing out the neurological changes that during menopause may increase the female brain vulnerability to AD. The disparity between animal study results and epidemiological and clinical data has highlighted the need to address the strengths and weaknesses of current animal models of menopause and their use to predict therapeutic outcomes.

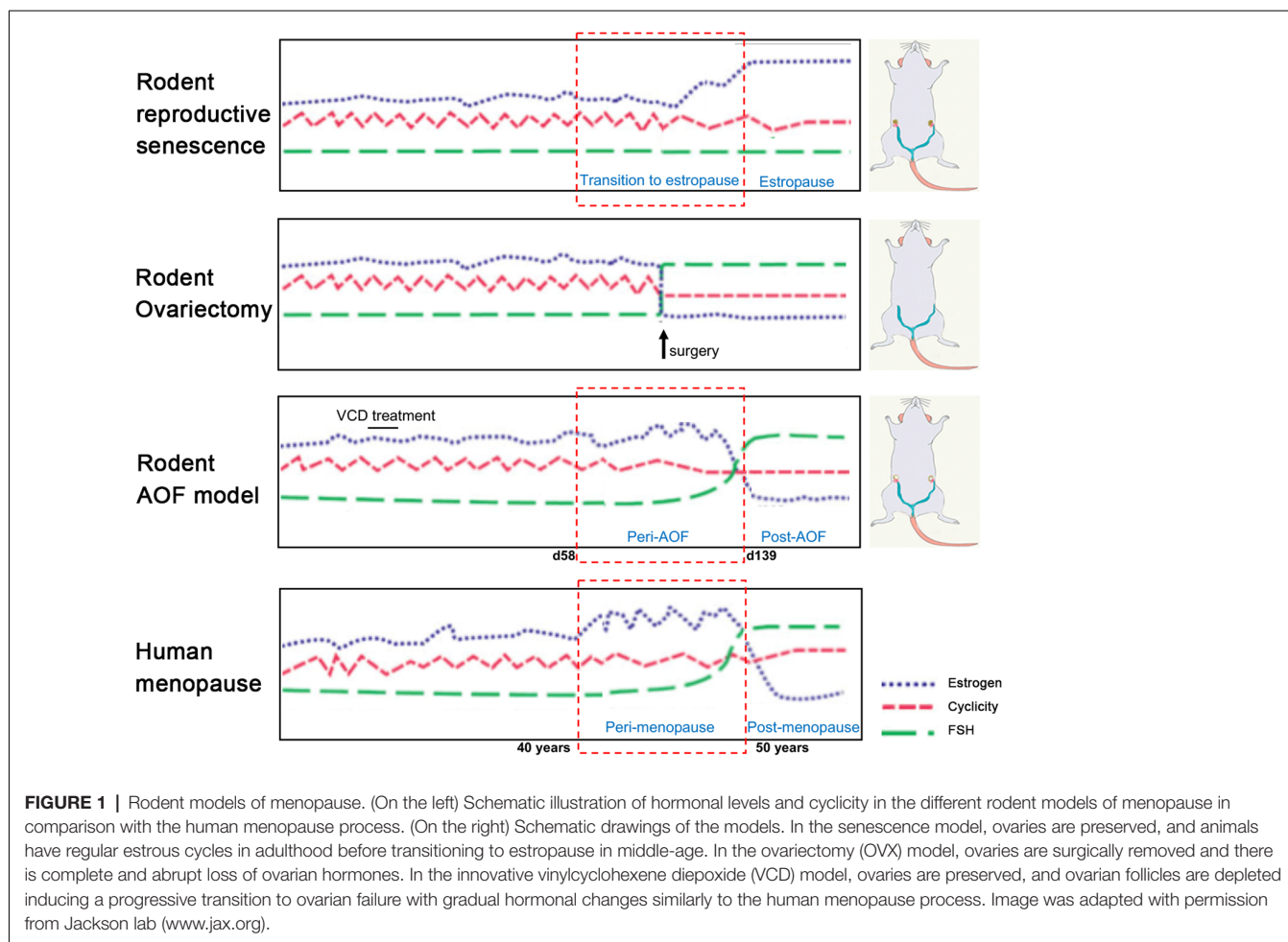
The most employed models, ovary-intact aging (reproductive senescence) and ovariectomy present several drawbacks that limit our understanding of the neurological underpinning of menopause transition. These models will be discussed in

the following paragraphs in view of their application in AD research and compared to the more recently developed model of AOF (Figure 1).

## Reproductive Senescence

The ovary is the main site of female sex hormone production. The mammalian ovary has two major functions: (1) differentiation, maturation, and release of oocytes for fertilization (McGee and Hsueh, 2000); and (2) production and sequential secretion of the hormones estrogen, progesterone, and inhibin A and B that regulate the hypothalamus–pituitary–gonadal axis in a repetitive process of follicle development, ovulation, corpus luteum formation and regression during the menstrual/estrous cycle (Peters et al., 1975; Hirshfield, 1991). At birth, the mammalian ovary carries a finite number of immature oocyte-containing follicles, which represent a limited pool of germ cells for reproduction. Following follicular maturation, oocytes are released during ovulation to be fertilized. Most follicles do not reach maturity and go through cell death named atresia. Physiologically, menopause occurs following depletion of all follicles in the ovary (McGee and Hsueh, 2000).

Women have a menstrual cycle, whereas female rodents have an estrous cycle which occurs every 4–5 days and consists of proestrus, estrous, metestrus, diestrus phases that involve similar hormonal fluctuations seen in the human cycle (Goldman et al., 2007). Similar to women, rodents experience natural hormonal fluctuations that emerge at middle age (9–12 months of age) which involves irregular cycles with prolonged diestrus phases and possible absence of ovulation (Maffucci and Gore, 2006; Downs and Wise, 2009; Finch, 2014). Around 1 year of age, differently from the human menopause, rodents transition into a state of extended estrous phases (estropause) with cessation of reproductive cycles. Eventually, at about 16–18 months of age, rodents transition into an acyclic, anestrus state involving persistent estrous and complete halt of ovulation (Chakraborty and Gore, 2004; Maffucci and Gore, 2006). Stages of pseudopregnancy with irregular ovulation and dysregulation of estrogen levels are also common during the rodent estropause. Interestingly, the ovary also produces testosterone. However, levels of ovarian testosterone are not impacted by menopause transition, unlike estrogens and progestins, but declines slowly with age (Burger, 2002; Davis and Wahlin-Jacobsen, 2015). Research conducted over the past 20 years in the ovary-intact aging model supports the notion that female brain vulnerability to AD is associated with loss of ovarian hormones after menopause. For instance, several groups reported an increase in A $\beta$  deposition in the brains of 1-year-old, and older, females compared to age-matched males and young females in rodent models of AD (Zheng et al., 2002; Wang et al., 2003). Using the aging model, the Brinton group has provided gene expression and proteomic evidence of an early shift in mitochondrial and lipid metabolism in the hippocampus of females tested starting at ~9 months of age compared to age-matched males and young females in the 3xTgAD mouse model (Yao et al., 2009; Ding et al., 2013a; Zhao et al., 2016). However, this does not rule out the possibility of an influence of the aging process itself and high estrogen levels in the elderly rodent systems used.



In conclusion, the rodent reproductive senescence model is characterized by multiple features found in the human menopause including: (1) retention of ovarian tissue; (2) irregular cycling and steroid hormone fluctuations; and (3) irregular fertility. However, because rodents do not exhibit significant depletion in the pool of ovarian follicles at the onset of reproductive senescence, circulating ovarian hormone levels do not present a drastic reduction (Dubal et al., 2012). This rodent model of aging contrasts from the human menopause, where levels of estrogens are very low or undetectable (Chakraborty and Gore, 2004).

### Ovariectomy Model

Ovariectomy, or bilateral surgical removal of the ovaries, is a well-established model of menopause (Olson and Bruce, 1986; Maffucci and Gore, 2006). In this model, rodents can be OVXed at different ages that correlate with different life stages. For example, females may be OVXed at 2–6 months of age with regular estrous cycles, 10 months at the beginning of the acyclicity, and at 18 months of age at the beginning of persistent estrous. This is an ideal model to study the effect of ovarian hormones deficit and influence of exogenous hormonal treatments on the brain. Experimental interventions may occur

either at the time of OVX or commence once  $17\beta$ -estradiol has reached a low to non-detectable level in the plasma, which typically occurs within 1–2 weeks (Marques-Lopes et al., 2018).

Using this model, many laboratories have studied the effect of estrogen loss, and  $17\beta$ -estradiol and other hormonal formulations, on cognitive function in normal wild type and AD rodents (Diaz Brinton, 2012; Li et al., 2014; Koebele and Bimonte-Nelson, 2016). For instance, administration of  $17\beta$ -estradiol to OVX females have shown to increase performance in spatial memory behavioral tasks (Koebele and Bimonte-Nelson, 2016; Koebele et al., 2017). As mentioned above, in the 3xTgAD mouse model of AD, there is a significant correlation between decreased mitochondrial and glucose metabolism and  $A\beta$  load in the hippocampus of OVX females with respect to intact females (Yao et al., 2009, 2012; Ding et al., 2013a,b). The shift in energy metabolism was associated with an increase in  $A\beta$  deposition in OVX females compared to males and ameliorated by treatment with  $17\beta$ -estradiol (Callahan et al., 2001; Levin-Allerhand and Smith, 2002; Zheng et al., 2002; Yue et al., 2005; Yao et al., 2012; Ding et al., 2013b; Lee et al., 2014).

In the OVX model, some shortcomings may influence the interpretation of data: (1) age of mice at OVX; (2) dose,



duration, and mode of administration of hormonal treatment; and (3) differential animal hormonal responsiveness depending of time elapsed since OVX (Gibbs, 1997; Rapp et al., 2003; Maffucci and Gore, 2006; Scharfman et al., 2007). For instance, several studies have addressed these issues and support that estrogen sensitivity decreases when greater time has lapsed since OVX or with increasing age making it difficult to compare across experiments (Morrison et al., 2006; Smith et al., 2010; Koebele et al., 2017).

While the OVX model provides insights into the role of estrogen and other gonadal hormones separately from aging as a confounding factor, this has two drawbacks. First, in addition to estrogens, OVX depletes other hormones that have important roles in menopause and can impact brain function (Maffucci and Gore, 2006; Rocca et al., 2011). Second, since the majority of menopausal women have intact ovaries, the abrupt loss of gonadal hormones models surgical menopause, but not natural transitional menopause. This does not allow to shed light on the progressive changes in estrogen responsiveness that occur when the natural cycle of circulating estrogens are disrupted at peri-menopause (Nejat and Chervenak, 2010; Shuster et al., 2010; Van Kempen et al., 2011). In support of this, Koebele and Bimonte-Nelson (2017) have shown that in rats, the model of ovarian hormone loss influences spatial memory performance and response to estrogen therapy, supporting the importance of studying surgical and transitional menopause independently (Acosta et al., 2009, 2010).

## The Accelerated Ovarian Failure (AOF) Model

Until recently, aging and OVX have been the primary rodent models of menopause to examine effects of ovarian hormone loss on cognition and AD in females (Koebele and Bimonte-Nelson, 2016). Although these models have furthered our understanding of estrogen actions on the central nervous system, as reported above, the rodent aging and OVX do not fully recapitulate the transitional events of human menopause (Van Kempen et al., 2011). The lack of an adequate model that includes a progressive transition through menopause has hindered studies on how irregular hormonal changes during peri-menopause impact the female brain susceptibility to AD, and the identification of the molecular mechanisms underlying this phenomenon that can be targeted to prevent or delay the disease. Lack of an optimal model that could also dissociate the effect of menopause from aging has limited the studies on the timing of hormonal replacement therapies and how these can influence disease outcomes.

In early 2000s, the Hoyer group developed the innovative model of AOF that successfully replicates the human peri- and post-menopause stages, including irregular estrous fluctuations at peri-AOF (Mayer et al., 2004, 2005; Williams, 2005; Van Kempen et al., 2011, 2014), and acyclicity at post-AOF stage with very low estrogen levels, and follicle-stimulating hormone (FSH) and luteinizing hormone (LH) levels raising accordingly (Mayer et al., 2002; Van Kempen et al., 2011; Brooks et al., 2016; Marques-Lopes et al., 2018). In this model, 15-day administration of low doses of the 4-VCD (130–160 mg/Kg in

0.5% dimethyl sulfoxide in sesame oil; i.p.) induces selective depletion of ovarian primary and primordial follicles (Lohff et al., 2006; Acosta et al., 2009; Van Kempen et al., 2011, 2014; Brooks et al., 2016; Koebele and Bimonte-Nelson, 2016). For a mechanistic review, see Hoyer and Sipes (1996, 2007) and Hoyer et al. (2001). Following VCD treatment, mature follicles deplete with normal estrous cycles (Flaws et al., 1994; Mayer et al., 2002; Lohff et al., 2006) resulting in a progressive transition to acyclicity and ovarian failure (Mayer et al., 2004). In addition to reducing confounds associated with surgical manipulations, the AOF model maintains the presence of ovarian tissue and presents a peri-AOF stage which importantly parallels human peri-menopause (Mayer et al., 2004; Rivera et al., 2009). Another advantage of this model is that it allows for the longitudinal study of the effects of hormone levels dissociated from the effects of aging in young animals (Danilovich and Ram Sairam, 2006; Koebele and Bimonte-Nelson, 2016). The drawback of this model is that VCD is toxic at higher doses (National Toxicology Program, 1986). However, in a laboratory setting, administration of low doses of VCD does not negatively affect peripheral tissues, including liver and kidney function and organ weights (Devine et al., 2001; Hoyer et al., 2001; Mayer et al., 2005; Haas et al., 2007; Sahambi et al., 2008; Wright et al., 2008; Frye et al., 2012; Van Kempen et al., 2014), and does not appear to cross the blood–brain barrier (Van Kempen et al., 2011, 2014). Work from the Milner group established that this model is suitable for longitudinal studies (Van Kempen et al., 2014), and does not have any direct effects on induction of inflammation markers in brain areas inside (e.g., hippocampus and hypothalamus) and outside (e.g., circumventricular organs) the blood–brain barrier (Van Kempen et al., 2014). Based on the assessment of ovarian follicle depletion and responses of plasticity markers in brain areas known to be estrogen responsive, timepoints for pre-AOF (~25 days after first injection), peri-AOF (>58 days post-injection), and post-AOF (>139 days post-injection) stages have been established corresponding to the human pre-, peri-, and post-menopause, respectively (Mayer et al., 2004; Lohff et al., 2006; Van Kempen et al., 2011, 2014; Marques-Lopes et al., 2018; **Figure 2**). Length of peri-AOF period and beginning of post-AOF can be manipulated by varying the duration of VCD treatment (Brooks et al., 2016).

Several laboratories have implemented this model to study cognition, cardiovascular disease, insulin resistance, atherosclerosis, bone loss, anxiety (Van Kempen et al., 2014), and hypertension (see next paragraph). A fairly recent summary of VCD doses and organs evaluated in studies applying the AOF model can be found in Van Kempen et al. (2011) and Marques-Lopes et al. (2018). A single study using the AOF has been reported in Tg2576 mouse model of AD. Golub et al. (2008) treated female mice with VCD at 60–75 days of age to induce AOF, and found no significant improvement of estrogen replacement in cognitive performance or A $\beta$  plaque load in the hippocampus and anterior cortex of post-AOF females. The study was affected by low power due to small sample size and measured the effect of estrogen only on late post-AOF mice at 15 months



of age, thus missing the opportunity to quantify the influence of peri-AOF on AD phenotype and introducing aging as a variable during data collection. Therefore, the influence of peri-AOF on neuropathology and phenotype in AD model has not been tested and cannot be ruled out. Nevertheless, the AOF models has been used to study cognitive changes in both mice and rats, which suggested a need for caution in extrapolating data on cognitive function from OVX models to a VCD model, and demonstrated the importance of studying transitional and surgical menopause independently (Acosta et al., 2009, 2010; Koebele and Bimonte-Nelson, 2017; Koebele et al., 2017).

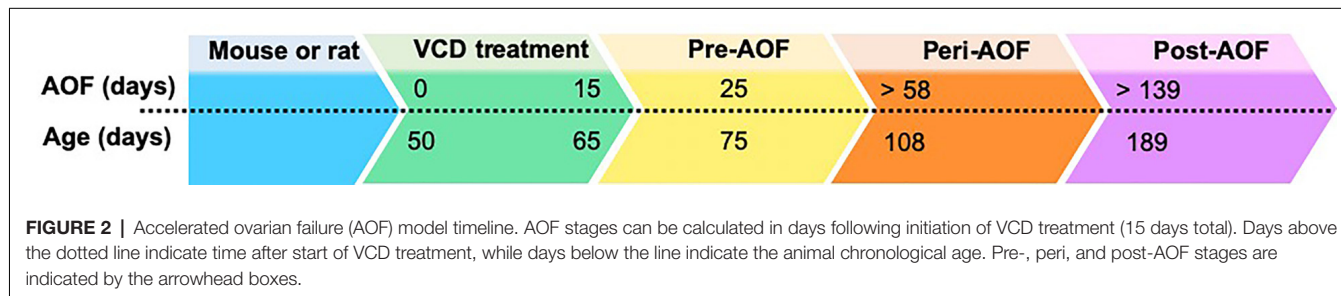
In summary, studies employing VCD-treated animals highlight that this model of AOF is extremely useful for modeling biological mechanisms and disease states associated with peri- and post-menopause, and that studies employing the VCD model will be particularly valuable to validate the most promising findings generated in the aged and OVX models of AD.

## Lessons From Hypertension Studies as an Illustration of Rodent AOF Model Utilization

Although works employing the AOF in AD animals are lacking, this model has been used to study sex dimorphism in other diseases and in risk factors of AD, such as the study of the influence of menopause transition on chronic elevated blood pressure (hypertension; Van Kempen et al., 2011; Iadecola, 2014; Wiesmann et al., 2017; Marques-Lopes et al., 2018). Given that some AD risk factors, like hypertension, are modifiable, determining the extent to which sex differences contribute to differential vulnerability may present opportunities for the development of more targeted and efficacious therapies. A large body of literature supports the deleterious role of hypertension in midlife as a risk factor for AD in both men and women (Feldstein, 2012; Davey, 2014a,b; Iadecola, 2014; Iadecola et al., 2016; Kehoe, 2018). There is both clinical and pre-clinical evidence that hypertension induces cerebrovascular damage that causes the reduction of A $\beta$  removal from the brain, and activates chronic inflammation, factors that accelerate the neuropathology and cognitive impairment in AD (Zlokovic, 2011; Carnevale et al., 2012; Shah et al., 2012; Attems and Jellinger, 2014; Kruyer et al., 2015; Faraco et al., 2016).

Sex differences in blood pressure control have been extensively reported (Lima et al., 2012; Sandberg and Ji, 2012). Men with hypertension seem to have a higher risk of developing AD than pre-menopausal women, but this relationship is inverted during the menopause transition and postmenopausal women have a higher risk of AD than age-matched men (Burt et al., 1995a,b; Zanchetti et al., 2005; Yanes and Reckelhoff, 2011; Lima et al., 2012). Notably, estrogen influence on blood pressure involves regulation of a number of brain regions including the ventrolateral medulla, nucleus of solitary tract, and paraventricular nucleus of the hypothalamus (McEwen et al., 2012). Within this circuitry, there are many potential sites by which estrogen can interact with renin-angiotensin system and molecular signaling pathways critical for regulation of brain cardiovascular circuits (McEwen et al., 2012; Maranon et al., 2014; McEwen and Milner, 2017).

It is now established that young female mice are protected from slow pressor Angiotensin II (AngII)-induced hypertension compared to age-matched males (Girouard et al., 2009; Xue et al., 2009, 2013, 2014; Marques-Lopes et al., 2014, 2015, 2017; Van Kempen et al., 2015). An increasing number of studies using either the senescence or OVX models have shown that in “post-menopause-like” state females are more susceptible to AngII-induced hypertension to a magnitude similar to that observed in males (Tiwari et al., 2009; Capone et al., 2012; Sandberg and Ji, 2012; Coleman et al., 2013; Marques-Lopes et al., 2014, 2015). These studies provide a first indication that estrogen may play a critical role in protection against hypertension. Interestingly, by using the AOF model, work from Dr. Milner group showed for the first time that peri-AOF is the critical time for the susceptibility to AngII-induced hypertension (Marques-Lopes et al., 2014, 2017; Van Kempen et al., 2016). While the data obtained employing the aging and OVX models only suggested that estrogen plays a role in hypertension in rodents, in the studies by the Milner group, the use of the AOF model uniquely allowed the testing of the hypothesis that irregular estrogen fluctuations during AOF transition, rather than loss of estrogen at post-AOF, may be responsible for the observed increase in susceptibility to AngII-induced hypertension. Furthermore, it was shown that estradiol activation of ER $\beta$  in the paraventricular nucleus of the hypothalamus (PVN) attenuates the glutamate-induced increase in blood pressure (Gingerich and Krukoff, 2006). To better understand the link between estrogen and hypertension, Milner et al. studied the changes in subcellular localization of the glutamate N-Methyl-D-aspartate (NMDA)



receptor subunit GluN1 in ER $\beta$ -positive neurons of the PVN following slow pressor AngII treatment in males and females using both the senescence and AOF models (Marques-Lopes et al., 2014, 2017). In both models, AngII-treated young females, males, and peri-AOF females show decreased total density of GluN1 in ER $\beta$  dendrites of PVN, whereas aged females and post-AOF females had an increase in total density. However, in males and peri-AOF females, plasmalemmal affiliation of GluN1 was increased, while it was unchanged in post-AOF females. As for the studies by Acosta et al. (2009, 2010) on the influence of the menopause model used in cognitive tasks, these findings indicate that distinct neurobiological processes underlie AngII-induced hypertension in aging and AOF, which may arise from the differences in estrogen levels between aged and AOF female mice.

## CONCLUSION

The multi-factorial nature of AD neuropathology and symptomatology has taught us that a single therapeutic approach will most likely not fit all. Studying the effects of sex differences and menopause will lead to the development of novel targeted precision medicine approaches that take sex and hormonal status into account.

Although substantial advances in medicine and research for AD have been made over the past years, the degree to which

human studies can understand the neurological mechanisms of menopause in AD is limited. Rodent models of menopause have proven very useful to start dissecting the key molecular mechanisms underlying the influence of menopause transition on AD. In particular, the innovative model of AOF can provide a valuable approach to the study of physiological changes that more closely parallel to the ones of human menopause. The advantages of the AOF as a model of transitional menopause make it an ideal choice for studies of menopause and HRT in mouse AD models.

## AUTHOR CONTRIBUTIONS

RM performed literature search, prepared and reviewed the manuscript.

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## REFERENCES

- Acosta, J. I., Mayer, L. P., Braden, B. B., Nonnenmacher, S., Mennenga, S. E., and Bimonte-Nelson, H. A. (2010). The cognitive effects of conjugated equine estrogens depend on whether menopause etiology is transitional or surgical. *Endocrinology* 151, 3795–3804. doi: 10.1210/en.2010-0055
- Acosta, J. I., Mayer, L., Talboom, J. S., Tsang, C. W., Smith, C. J., Enders, C. K., et al. (2009). Transitional versus surgical menopause in a rodent model: etiology of ovarian hormone loss impacts memory and the acetylcholine system. *Endocrinology* 150, 4248–4259. doi: 10.1210/en.2008-1802
- Alberts, S. C., Altmann, J., Brockman, D. K., Cords, M., Fedigan, L. M., Pusey, A., et al. (2013). Reproductive aging patterns in primates reveal that humans are distinct. *Proc. Natl. Acad. Sci. U S A* 110, 13440–13445. doi: 10.1073/pnas.1311857110
- Altmann, A., Tian, L., Henderson, V. W., Greicius, M. D., and Alzheimer's Disease Neuroimaging Initiative Investigators. (2014). Sex modifies the APOE-related risk of developing Alzheimer disease. *Ann. Neurol.* 75, 563–573. doi: 10.1002/ana.24135
- Alzheimer's Association (2016). 2016 Alzheimer's disease facts and figures. *Alzheimers Dement.* 12, 459–509. doi: 10.1016/j.jalz.2016.03.001
- Arancibia, S., Silhol, M., Moulriere, F., Meffre, J., Hollinger, I., Maurice, T., et al. (2008). Protective effect of BDNF against  $\beta$ -amyloid induced neurotoxicity *in vitro* and *in vivo* in rats. *Neurobiol. Dis.* 31, 316–326. doi: 10.1016/j.nbd.2008.05.012
- Arevalo, M. A., Azcoitia, I., Gonzalez-Burgos, I., and Garcia-Segura, L. M. (2015). Signaling mechanisms mediating the regulation of synaptic plasticity and memory by estradiol. *Horm. Behav.* 74, 19–27. doi: 10.1016/j.yhbeh.2015.04.016
- Asuni, A. A., Boutajangout, A., Quartermain, D., and Sigurdsson, E. M. (2007). Immunotherapy targeting pathological tau conformers in a tangle mouse model reduces brain pathology with associated functional improvements. *J. Neurosci.* 27, 9115–9129. doi: 10.1523/jneurosci.2361-07.2007
- Attems, J., and Jellinger, K. A. (2014). The overlap between vascular disease and Alzheimer's disease—lessons from pathology. *BMC Med.* 12:206. doi: 10.1186/s12916-014-0206-2
- Au, A., Feher, A., McPhee, L., Jessa, A., Oh, S., and Einstein, G. (2016). Estrogens, inflammation and cognition. *Front. Neuroendocrinol.* 40, 87–100. doi: 10.1016/j.yfrne.2016.01.002
- Ball, J. D., and Chen, X. (2016). Shifts in endocrine homeostasis and preventive hormone replacement therapy: extending the Women's Health Initiative globally. *Glob. Health Res. Policy* 1:9. doi: 10.1186/s41256-016-0009-4
- Baranello, R. J., Bharani, K. L., Padmaraju, V., Chopra, N., Lahiri, D. K., Greig, N. H., et al. (2015). Amyloid- $\beta$  protein clearance and degradation (ABCD) pathways and their role in Alzheimer's disease. *Curr. Alzheimer Res.* 12, 32–46. doi: 10.2174/1567205012666141218140953
- Barja, G. (2004). Free radicals and aging. *Trends Neurosci.* 27, 595–600. doi: 10.1016/j.tins.2004.07.005
- Barnes, L. L., Wilson, R. S., Bienias, J. L., Schneider, J. A., Evans, D. A., and Bennett, D. A. (2005). Sex differences in the clinical manifestations of Alzheimer disease pathology. *Arch. Gen. Psychiatry* 62, 685–691. doi: 10.1001/archpsyc.62.6.685
- Barron, A. M., and Pike, C. J. (2012). Sex hormones, aging, and Alzheimer's disease. *Front. Biosci.* 4, 976–997. doi: 10.2741/e434
- Baudry, M., Bi, X., and Aguirre, C. (2013). Progesterone-estrogen interactions in synaptic plasticity and neuroprotection. *Neuroscience* 239, 280–294. doi: 10.1016/j.neuroscience.2012.10.051
- Benzing, W. C., Wujek, J. R., Ward, E. K., Shaffer, D., Ashe, K. H., Younkin, S. G., et al. (1999). Evidence for glial-mediated inflammation in aged APP(SW) transgenic mice. *Neurobiol. Aging* 20, 581–589. doi: 10.1016/s0197-4580(99)00065-2
- Beydoun, M. A., Boueiz, A., Abougergi, M. S., Kitner-Triolo, M. H., Beydoun, H. A., Resnick, S. M., et al. (2012). Sex differences in the association of the apolipoprotein E epsilon 4 allele with incidence of dementia, cognitive impairment, and decline. *Neurobiol. Aging* 33, 720.e4–731.e4. doi: 10.1016/j.neurobiolaging.2010.05.017
- Bhavnani, B. R. (2003). Estrogens and menopause: pharmacology of conjugated equine estrogens and their potential role in the prevention of neurodegenerative diseases such as Alzheimer's. *J. Steroid Biochem. Mol. Biol.* 85, 473–482. doi: 10.1016/s0960-0760(03)00220-6

- Blaustein, J. D. (2008). An estrogen by any other name. *Endocrinology* 149, 2697–2698. doi: 10.1210/en.2008-0396
- Blurton-Jones, M., and Tuszynski, M. H. (2001). Reactive astrocytes express estrogen receptors in the injured primate brain. *J. Comp. Neurol.* 433, 115–123. doi: 10.1002/cne.1129
- Bove, R., Secor, E., Chibnik, L. B., Barnes, L. L., Schneider, J. A., Bennett, D. A., et al. (2014). Age at surgical menopause influences cognitive decline and Alzheimer pathology in older women. *Neurology* 82, 222–229. doi: 10.1212/wnl.000000000000033
- Braak, H., and Braak, E. (1997). Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol. Aging* 18, 377–379. doi: 10.1016/s0197-4580(97)00051-1
- Brinton, R. D. (2008). The healthy cell bias of estrogen action: mitochondrial bioenergetics and neurological implications. *Trends Neurosci.* 31, 529–537. doi: 10.1016/j.tins.2008.07.003
- Brinton, R. D., Yao, J., Yin, F., Mack, W. J., and Cadenas, E. (2015). Perimenopause as a neurological transition state. *Nat. Rev. Endocrinol.* 11, 393–405. doi: 10.1038/nrendo.2015.82
- Broestl, L., Worden, K., Moreno, A. J., Davis, E. J., Wang, D., Garay, B., et al. (2018). Ovarian cycle stages modulate Alzheimer-related cognitive and brain network alterations in female mice. *eNeuro* 5:ENEURO.0132-17.2018. doi: 10.1523/eneuro.0132-17.2018
- Brooks, H. L., Pollow, D. P., and Hoyer, P. B. (2016). The VCD mouse model of menopause and perimenopause for the study of sex differences in cardiovascular disease and the metabolic syndrome. *Physiology* 31, 250–257. doi: 10.1152/physiol.00057.2014
- Bu, G. (2009). Apolipoprotein E and its receptors in Alzheimer's disease: pathways, pathogenesis and therapy. *Nat. Rev. Neurosci.* 10, 333–344. doi: 10.1038/nrn2620
- Buccarello, L., Grignaschi, G., Castaldo, A. M., Di Giancamillo, A., Domeneghini, C., Melcangi, R. C., et al. (2017). Sex impact on tau-aggregation and postsynaptic protein levels in the P301L mouse model of tauopathy. *J. Alzheimers Dis.* 56, 1279–1292. doi: 10.3233/jad-161087
- Buckley, R. F., Mormino, E. C., Rabin, J. S., Hohman, T. J., Landau, S., Hanseeuw, B. J., et al. (2019). Sex differences in the association of global amyloid and regional tau deposition measured by positron emission tomography in clinically normal older adults. *JAMA Neurol.* 76, 542–551. doi: 10.1001/jamaneurol.2018.4693
- Burger, H. G. (2002). Androgen production in women. *Fertil. Steril.* 77, S3–S5. doi: 10.1016/s0015-0282(02)02985-0
- Burns, A., Lewis, G., Jacoby, R., and Levy, R. (1991). Factors affecting survival in Alzheimer's disease. *Psychol. Med.* 21, 363–370. doi: 10.1017/S0033291700020468
- Burt, V. L., Cutler, J. A., Higgins, M., Horan, M. J., Labarthe, D., Whelton, P., et al. (1995a). Trends in the prevalence, awareness, treatment and control of hypertension in the adult US population. Data from the health examination surveys, 1960 to 1991. *Hypertension* 26, 60–69. doi: 10.1161/01.hyp.26.1.60
- Burt, V. L., Whelton, P., Roccella, E. J., Brown, C., Cutler, J. A., Higgins, M., et al. (1995b). Prevalence of hypertension in the US adult population. Results from the Third National Health and Nutrition Examination Survey, 1988–1991. *Hypertension* 25, 305–313. doi: 10.1161/01.hyp.25.3.305
- Butler, L., and Santoro, N. (2011). The reproductive endocrinology of the menopausal transition. *Steroids* 76, 627–635. doi: 10.1016/j.steroids.2011.02.026
- Callahan, M. J., Lipinski, W. J., Bian, F., Durham, R. A., Pack, A., and Walker, L. C. (2001). Augmented senile plaque load in aged female  $\beta$ -amyloid precursor protein-transgenic mice. *Am. J. Pathol.* 158, 1173–1177. doi: 10.1016/s0002-9440(10)64064-3
- Canter, R. G., Penney, J., and Tsai, L. H. (2016). The road to restoring neural circuits for the treatment of Alzheimer's disease. *Nature* 539, 187–196. doi: 10.1038/nature20412
- Cantuti-Castelvetri, I., Lin, M. T., Zheng, K., Keller-McGandy, C. E., Betensky, R. A., Johns, D. R., et al. (2005). Somatic mitochondrial DNA mutations in single neurons and glia. *Neurobiol. Aging* 26, 1343–1355. doi: 10.1016/j.neurobiolaging.2004.11.008
- Capone, C., Faraco, G., Peterson, J. R., Coleman, C., Anrather, J., Milner, T. A., et al. (2012). Central cardiovascular circuits contribute to the neurovascular dysfunction in angiotensin II hypertension. *J. Neurosci.* 32, 4878–4886. doi: 10.1523/jneurosci.6262-11.2012
- Carnevale, D., Mascio, G., Ajmone-Cat, M. A., D'Andrea, I., Cifelli, G., Madonna, M., et al. (2012). Role of neuroinflammation in hypertension-induced brain amyloid pathology. *Neurobiol. Aging* 33, 205.e19–205.e29. doi: 10.1016/j.neurobiolaging.2010.08.013
- Carroll, J. C., Rosario, E. R., Chang, L., Stanczyk, F. Z., Oddo, S., LaFerla, F. M., et al. (2007). Progesterone and estrogen regulate Alzheimer-like neuropathology in female 3xTg-AD mice. *J. Neurosci.* 27, 13357–13365. doi: 10.1523/jneurosci.2718-07.2007
- Carroll, J. C., Rosario, E. R., Villamagna, A., and Pike, C. J. (2010). Continuous and cyclic progesterone differentially interact with estradiol in the regulation of Alzheimer-like pathology in female 3xTransgenic-Alzheimer's disease mice. *Endocrinology* 151, 2713–2722. doi: 10.1210/en.2009-1487
- Chakraborty, T. R., and Gore, A. C. (2004). Aging-related changes in ovarian hormones, their receptors, and neuroendocrine function. *Exp. Biol. Med.* 229, 977–987. doi: 10.1177/153537020422901001
- Chapman, R. M., Mapstone, M., McCrary, J. W., Gardner, M. N., Porsteinsson, A., Sandoval, T. C., et al. (2011). Predicting conversion from mild cognitive impairment to Alzheimer's disease using neuropsychological tests and multivariate methods. *J. Clin. Exp. Neuropsychol.* 33, 187–199. doi: 10.1080/13803395.2010.499356
- Christensen, A., and Pike, C. J. (2015). Menopause, obesity and inflammation: interactive risk factors for Alzheimer's disease. *Front. Aging Neurosci.* 7:130. doi: 10.3389/fnagi.2015.00130
- Coleman, C. G., Wang, G., Faraco, G., Marques Lopes, J., Waters, E. M., Milner, T. A., et al. (2013). Membrane trafficking of NADPH oxidase p47(phox) in paraventricular hypothalamic neurons parallels local free radical production in angiotensin II slow-pressor hypertension. *J. Neurosci.* 33, 4308–4316. doi: 10.1523/jneurosci.3061-12.2013
- Coon, K. D., Myers, A. J., Craig, D. W., Webster, J. A., Pearson, J. V., Lince, D. H., et al. (2007). A high-density whole-genome association study reveals that APOE is the major susceptibility gene for sporadic late-onset Alzheimer's disease. *J. Clin. Psychiatry* 68, 613–618. doi: 10.4088/jcp.v68.n0419
- Corder, E. H., Ghebremedhin, E., Taylor, M. G., Thal, D. R., Ohm, T. G., and Braak, H. (2004). The biphasic relationship between regional brain senile plaque and neurofibrillary tangle distributions: modification by age, sex and APOE polymorphism. *Ann. N Y Acad. Sci.* 1019, 24–28. doi: 10.1196/annals.1297.005
- Corder, E. H., Saunders, A. M., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., Small, G. W., et al. (1993). Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261, 921–923. doi: 10.1126/science.8346443
- Cui, J., Shen, Y., and Li, R. (2013). Estrogen synthesis and signaling pathways during aging: from periphery to brain. *Trends Mol. Med.* 19, 197–209. doi: 10.1016/j.molmed.2012.12.007
- Czirr, E., Castello, N. A., Mosher, K. I., Castellano, J. M., Hinkson, I. V., Lucin, K. M., et al. (2017). Microglial complement receptor 3 regulates brain A $\beta$  levels through secreted proteolytic activity. *J. Exp. Med.* 214, 1081–1092. doi: 10.1084/jem.20162011
- Danilovich, N., and Ram Sairam, M. (2006). Recent female mouse models displaying advanced reproductive aging. *Exp. Gerontol.* 41, 117–122. doi: 10.1016/j.exger.2005.10.010
- Davey, D. A. (2014a). Alzheimer's disease and vascular dementia: one potentially preventable and modifiable disease? Part II: management, prevention and future perspective. *Neurodegener. Dis. Manag.* 4, 261–270. doi: 10.2217/nmt.14.14
- Davey, D. A. (2014b). Alzheimer's disease and vascular dementia: one potentially preventable and modifiable disease. Part I: pathology, diagnosis and screening. *Neurodegener. Dis. Manag.* 4, 253–259. doi: 10.2217/nmt.14.13
- Davis, S. R., and Wahlin-Jacobsen, S. (2015). Testosterone in women—the clinical significance. *Lancet Diabetes Endocrinol.* 3, 980–992. doi: 10.1016/s2213-8587(15)00284-3
- Deb, A., Thornton, J. D., Sambamoorthi, U., and Innes, K. (2017). Direct and indirect cost of managing Alzheimer's disease and related dementias in the United States. *Expert Rev. Pharmacoecon. Outcomes Res.* 17, 189–202. doi: 10.1080/14737167.2017.1313118



- Depypere, H., Vierin, A., Weyers, S., and Sieben, A. (2016). Alzheimer's disease, apolipoprotein E and hormone replacement therapy. *Maturitas* 94, 98–105. doi: 10.1016/j.maturitas.2016.09.009
- Devine, P. J., Sipes, I. G., and Hoyer, P. B. (2001). Effect of 4-vinylcyclohexene diepoxide dosing in rats on GSH levels in liver and ovaries. *Toxicol. Sci.* 62, 315–320. doi: 10.1093/toxsci/62.2.315
- Diaz Brinton, R. (2012). Minireview: translational animal models of human menopause: challenges and emerging opportunities. *Endocrinology* 153, 3571–3578. doi: 10.1210/en.2012-1340
- Ding, F., Yao, J., Rettberg, J. R., Chen, S., and Brinton, R. D. (2013a). Early decline in glucose transport and metabolism precedes shift to ketogenic system in female aging and Alzheimer's mouse brain: implication for bioenergetic intervention. *PLoS One* 8:e79977. doi: 10.1371/journal.pone.0079977
- Ding, F., Yao, J., Zhao, L., Mao, Z., Chen, S., and Brinton, R. D. (2013b). Ovariectomy induces a shift in fuel availability and metabolism in the hippocampus of the female transgenic model of familial Alzheimer's. *PLoS One* 8:e59825. doi: 10.1371/journal.pone.0059825
- Downs, J. L., and Wise, P. M. (2009). The role of the brain in female reproductive aging. *Mol. Cell. Endocrinol.* 299, 32–38. doi: 10.1016/j.mce.2008.11.012
- Dubal, D. B., Broestl, L., and Worden, K. (2012). Sex and gonadal hormones in mouse models of Alzheimer's disease: what is relevant to the human condition? *Biol. Sex Differ.* 3:24. doi: 10.1186/2042-6410-3-24
- Dubois, B., Hampel, H., Feldman, H. H., Scheltens, P., Aisen, P., Andrieu, S., et al. (2016). Preclinical Alzheimer's disease: definition, natural history, and diagnostic criteria. *Alzheimers Dement.* 12, 292–323. doi: 10.1016/j.jalz.2016.02.002
- Epperson, C. N., Sammel, M. D., and Freeman, E. W. (2013). Menopause effects on verbal memory: findings from a longitudinal community cohort. *J. Clin. Endocrinol. Metab.* 98, 3829–3838. doi: 10.1210/jc.2013-1808
- Espeland, M. A., Shumaker, S. A., Leng, I., Manson, J. E., Brown, C. M., LeBlanc, E. S., et al. (2013). Long-term effects on cognitive function of postmenopausal hormone therapy prescribed to women aged 50 to 55 years. *JAMA Intern. Med.* 173, 1429–1436. doi: 10.1001/jamainternmed.2013.7727
- Faraco, G., Park, L., Zhou, P., Luo, W., Paul, S. M., Anrather, J., et al. (2016). Hypertension enhances A $\beta$ -induced neurovascular dysfunction, promotes  $\beta$ -secretase activity and leads to amyloidogenic processing of APP. *J. Cereb. Blood Flow Metab.* 36, 241–252. doi: 10.1038/jcbfm.2015.79
- Farrer, L. A., Cupples, L. A., Haines, J. L., Hyman, B., Kukull, W. A., Mayeux, R., et al. (1997). Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer disease meta analysis consortium. *JAMA* 278, 1349–1356. doi: 10.1001/jama.278.16.1349
- Feldstein, C. A. (2012). Association between chronic blood pressure changes and development of Alzheimer's disease. *J. Alzheimers Dis.* 32, 753–763. doi: 10.3233/jad-2012-120613
- Filon, J. R., Intorcchia, A. J., Sue, L. I., Vazquez Arreola, E., Wilson, J., Davis, K. J., et al. (2016). Gender differences in alzheimer disease: brain atrophy, histopathology burden and cognition. *J. Neuropathol. Exp. Neurol.* doi: 10.1093/jnen/nlw047 [Epub ahead of print].
- Finch, C. E. (2014). The menopause and aging, a comparative perspective. *J. Steroid Biochem. Mol. Biol.* 142, 132–141. doi: 10.1016/j.jsbmb.2013.03.010
- Fisher, D. W., Bennett, D. A., and Dong, H. (2018). Sexual dimorphism in predisposition to Alzheimer's disease. *Neurobiol. Aging* 70, 308–324. doi: 10.1016/j.neurobiolaging.2018.04.004
- Flaws, J. A., Doerr, J. K., Sipes, I. G., and Hoyer, P. B. (1994). Destruction of preantral follicles in adult rats by 4-vinyl-1-cyclohexene diepoxide. *Reprod. Toxicol.* 8, 509–514. doi: 10.1016/0890-6238(94)90033-7
- Folmar, L. C., Hemmer, M. J., Denslow, N. D., Kroll, K., Chen, J., Cheek, A., et al. (2002). A comparison of the estrogenic potencies of estradiol, ethynylestradiol, diethylstilbestrol, nonylphenol and methoxychlor *in vivo* and *in vitro*. *Aquat. Toxicol.* 60, 101–110. doi: 10.1016/s0166-445x(01)00276-4
- Frautschy, S. A., Yang, F., Irrizarry, M., Hyman, B., Saido, T. C., Hsiao, K., et al. (1998). Microglial response to amyloid plaques in APPsw transgenic mice. *Am. J. Pathol.* 152, 307–317.
- Frick, K. M. (2015). Molecular mechanisms underlying the memory-enhancing effects of estradiol. *Horm. Behav.* 74, 4–18. doi: 10.1016/j.yhbeh.2015.05.001
- Frye, J. B., Lukefahr, A. L., Wright, L. E., Marion, S. L., Hoyer, P. B., and Funk, J. L. (2012). Modeling perimenopause in Sprague-Dawley rats by chemical manipulation of the transition to ovarian failure. *Comp. Med.* 62, 193–202.
- Georgakis, M. K., Kalogirou, E. I., Diamantaras, A. A., Daskalopoulou, S. S., Munro, C. A., Lyketsos, C. G., et al. (2016). Age at menopause and duration of reproductive period in association with dementia and cognitive function: a systematic review and meta-analysis. *Psychoneuroendocrinology* 73, 224–243. doi: 10.1016/j.psyneuen.2016.08.003
- Gibbs, R. B. (1997). Effects of estrogen on basal forebrain cholinergic neurons vary as a function of dose and duration of treatment. *Brain Res.* 757, 10–16. doi: 10.1016/s0006-8993(96)01432-1
- Gillies, G. E., and McArthur, S. (2010). Estrogen actions in the brain and the basis for differential action in men and women: a case for sex-specific medicines. *Pharmacol. Rev.* 62, 155–198. doi: 10.1124/pr.109.002071
- Gingerich, S., and Krukoff, T. L. (2006). Estrogen in the paraventricular nucleus attenuates L-glutamate-induced increases in mean arterial pressure through estrogen receptor  $\beta$  and NO. *Hypertension* 48, 1130–1136. doi: 10.1161/01.hyp.0000248754.67128.ff
- Girouard, H., Wang, G., Gallo, E. F., Anrather, J., Zhou, P., Pickel, V. M., et al. (2009). NMDA receptor activation increases free radical production through nitric oxide and NOX2. *J. Neurosci.* 29, 2545–2552. doi: 10.1523/jneurosci.0133-09.2009
- Gleason, C. E., Dowling, N. M., Wharton, W., Manson, J. E., Miller, V. M., Atwood, C. S., et al. (2015). Effects of hormone therapy on cognition and mood in recently postmenopausal women: findings from the randomized, controlled KEEPS-cognitive and affective study. *PLoS Med.* 12:e1001833; discussion e1001833. doi: 10.1371/journal.pmed.1001833
- Goldman, J. M., Murr, A. S., and Cooper, R. L. (2007). The rodent estrous cycle: characterization of vaginal cytology and its utility in toxicological studies. *Birth Defects Res. B Dev. Reprod. Toxicol.* 80, 84–97. doi: 10.1002/bdrb.20106
- Golub, M. S., Germann, S. L., Mercer, M., Gordon, M. N., Morgan, D. G., Mayer, L. P., et al. (2008). Behavioral consequences of ovarian atrophy and estrogen replacement in the APPsw mouse. *Neurobiol. Aging* 29, 1512–1523. doi: 10.1016/j.neurobiolaging.2007.03.015
- Greendale, G. A., Huang, M. H., Wight, R. G., Seeman, T., Luetters, C., Avis, N. E., et al. (2009). Effects of the menopause transition and hormone use on cognitive performance in midlife women. *Neurology* 72, 1850–1857. doi: 10.1212/wnl.0b013e3181a71193
- Gurney, E. P., Nachtigall, M. J., Nachtigall, L. E., and Naftolin, F. (2014). The Women's Health Initiative trial and related studies: 10 years later: a clinician's view. *J. Steroid Biochem. Mol. Biol.* 142, 4–11. doi: 10.1016/j.jsbmb.2013.10.009
- Haas, J. R., Christian, P. J., and Hoyer, P. B. (2007). Effects of impending ovarian failure induced by 4-vinylcyclohexene diepoxide on fertility in C57BL/6 female mice. *Comp. Med.* 57, 443–449.
- Hall, J. R., Johnson, L. A., Barber, R. C., Vo, H. T., Winter, A. S., O'Bryant, S. E., et al. (2012). Biomarkers of basic activities of daily living in Alzheimer's disease. *J. Alzheimers Dis.* 31, 429–437. doi: 10.3233/JAD-2012-111481
- Hampel, H., Vergallo, A., Giorgi, F. S., Kim, S. H., Depypere, H., Graziani, M., et al. (2018). Precision medicine and drug development in Alzheimer's disease: the importance of sexual dimorphism and patient stratification. *Front. Neuroendocrinol.* 50, 31–51. doi: 10.1016/j.yfrne.2018.06.001
- Hanamsagar, R., and Bilbo, S. D. (2016). Sex differences in neurodevelopmental and neurodegenerative disorders: focus on microglial function and neuroinflammation during development. *J. Steroid Biochem. Mol. Biol.* 160, 127–133. doi: 10.1016/j.jsbmb.2015.09.039
- Hara, Y., Waters, E. M., McEwen, B. S., and Morrison, J. H. (2015). Estrogen effects on cognitive and synaptic health over the lifecourse. *Physiol. Rev.* 95, 785–807. doi: 10.1152/physrev.00036.2014
- Harlow, S. D., Gass, M., Hall, J. E., Lobo, R., Maki, P., Rebar, R. W., et al. (2012). Executive summary of the stages of reproductive aging workshop + 10: addressing the unfinished agenda of staging reproductive aging. *J. Clin. Endocrinol. Metab.* 97, 1159–1168. doi: 10.1210/jc.2011-3362



- Harsh, V., Schmidt, P. J., and Rubinow, D. R. (2007). The menopause transition: the next neuroendocrine frontier. *Expert Rev. Neurother.* 7, S7–S10. doi: 10.1586/14737175.7.11s.s7
- Henderson, V. W., Benke, K. S., Green, R. C., Cupples, L. A., Farrer, L. A., and Group, M. S. (2005). Postmenopausal hormone therapy and Alzheimer's disease risk: interaction with age. *J. Neurol. Neurosurg. Psychiatry* 76, 103–105. doi: 10.1136/jnnp.2003.024927
- Henderson, V. W., St John, J. A., Hodis, H. N., McCleary, C. A., Stanczyk, F. Z., Shoupe, D., et al. (2016). Cognitive effects of estradiol after menopause: a randomized trial of the timing hypothesis. *Neurology* 87, 699–708. doi: 10.1212/wnl.0000000000002980
- Heneka, M. T., Carson, M. J., El Khoury, J., Landreth, G. E., Brosseron, F., Feinstein, D. L., et al. (2015). Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14, 388–405. doi: 10.1016/S1474-4422(15)70016-5
- Hersh, A. L., Stefanick, M. L., and Stafford, R. S. (2004). National use of postmenopausal hormone therapy: annual trends and response to recent evidence. *JAMA* 291, 47–53. doi: 10.1001/jama.291.1.47
- Hirata-Fukae, C., Li, H. F., Hoe, H. S., Gray, A. J., Minami, S. S., Hamada, K., et al. (2008). Females exhibit more extensive amyloid, but not tau, pathology in an Alzheimer transgenic model. *Brain Res.* 1216, 92–103. doi: 10.1016/j.brainres.2008.03.079
- Hirshfield, A. N. (1991). Development of follicles in the mammalian ovary. *Int. Rev. Cytol.* 124, 43–101. doi: 10.1016/s0074-7696(08)61524-7
- Holland, D., Desikan, R. S., Dale, A. M., McEvoy, L. K., and Alzheimer's Disease Neuroimaging Initiative (2013). Higher rates of decline for women and apolipoprotein E epsilon4 carriers. *AJNR Am. J. Neuroradiol.* 34, 2287–2293. doi: 10.3174/ajnr.a3601
- Hoyer, P. B., Devine, P. J., Hu, X., Thompson, K. E., and Sipes, I. G. (2001). Ovarian toxicity of 4-vinylcyclohexene diepoxide: a mechanistic model. *Toxicol. Pathol.* 29, 91–99. doi: 10.1080/019262301301418892
- Hoyer, P. B., and Sipes, I. G. (1996). Assessment of follicle destruction in chemical-induced ovarian toxicity. *Annu. Rev. Pharmacol. Toxicol.* 36, 307–331. doi: 10.1146/annurev.pa.36.040196.001515
- Hoyer, P. B., and Sipes, I. G. (2007). Development of an animal model for oototoxicity using 4-vinylcyclohexene: a case study. *Birth Defects Res. B Dev. Reprod. Toxicol.* 80, 113–125. doi: 10.1002/bdrb.20103
- Hua, X., Hibar, D. P., Lee, S., Toga, A. W., Jack, C. R. Jr., Weiner, M. W., et al. (2010). Sex and age differences in atrophic rates: an ADNI study with  $n = 1368$  MRI scans. *Neurobiol. Aging* 31, 1463–1480. doi: 10.1016/j.neurobiolaging.2010.04.033
- Iadecola, C. (2014). Hypertension and dementia. *Hypertension* 64, 3–5. doi: 10.1161/HYPERTENSIONAHA.114.03040
- Iadecola, C., Yaffe, K., Biller, J., Bratzke, L. C., Faraci, F. M., Gorelick, P. B., et al. (2016). Impact of hypertension on cognitive function: a scientific statement from the american heart association. *Hypertension* 68, e67–e94. doi: 10.1161/hyp.0000000000000053
- Irvine, K., Laws, K. R., Gale, T. M., and Kondel, T. K. (2012). Greater cognitive deterioration in women than men with Alzheimer's disease: a meta analysis. *J. Clin. Exp. Neuropsychol.* 34, 989–998. doi: 10.1080/13803395.2012.712676
- Jack, C. R. Jr., Knopman, D. S., Jagust, W. J., Petersen, R. C., Weiner, M. W., Aisen, P. S., et al. (2013). Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol.* 12, 207–216. doi: 10.1016/s1474-4422(12)70291-0
- Jack, C. R. Jr., Wiste, H. J., Weigand, S. D., Therneau, T. M., Knopman, D. S., Lowe, V., et al. (2017). Age-specific and sex-specific prevalence of cerebral  $\beta$ -amyloidosis, tauopathy and neurodegeneration in cognitively unimpaired individuals aged 50–95 years: a cross-sectional study. *Lancet Neurol.* 16, 435–444. doi: 10.1016/s1474-4422(17)30077-7
- Jacobs, E. G., Weiss, B., Makris, N., Whitfield-Gabrieli, S., Buka, S. L., Klibanski, A., et al. (2017). Reorganization of functional networks in verbal working memory circuitry in early midlife: the impact of sex and menopausal status. *Cereb. Cortex* 27, 2857–2870. doi: 10.1093/cercor/bhw127
- Jacobs, E. G., Weiss, B. K., Makris, N., Whitfield-Gabrieli, S., Buka, S. L., Klibanski, A., et al. (2016). Impact of sex and menopausal status on episodic memory circuitry in early midlife. *J. Neurosci.* 36, 10163–10173. doi: 10.1523/JNEUROSCI.0951-16.2016
- Jaffe, A. B., Toran-Allerand, C. D., Greengard, P., and Gandy, S. E. (1994). Estrogen regulates metabolism of Alzheimer amyloid  $\beta$  precursor protein. *J. Biol. Chem.* 269, 13065–13068.
- Jevtic, S., Sengar, A. S., Salter, M. W., and McLaurin, J. (2017). The role of the immune system in Alzheimer disease: etiology and treatment. *Ageing Res. Rev.* 40, 84–94. doi: 10.1016/j.arr.2017.08.005
- Joffe, H., Hall, J. E., Gruber, S., Sarmiento, I. A., Cohen, L. S., Yurgelun-Todd, D., et al. (2006). Estrogen therapy selectively enhances prefrontal cognitive processes: a randomized, double-blind, placebo-controlled study with functional magnetic resonance imaging in perimenopausal and recently postmenopausal women. *Menopause* 13, 411–422. doi: 10.1097/01.gme.0000189618.48774.7b
- Kehoe, P. G. (2018). The coming of age of the angiotensin hypothesis in Alzheimer's disease: progress toward disease prevention and treatment? *J. Alzheimers Dis.* 62, 1443–1466. doi: 10.3233/jad-171119
- Klinge, C. M. (2017). Estrogens regulate life and death in mitochondria. *J. Bioenerg. Biomembr.* 49, 307–324. doi: 10.1007/s10863-017-9704-1
- Koebele, S. V., and Bimonte-Nelson, H. A. (2015). Trajectories and phenotypes with estrogen exposures across the lifespan: what does Goldilocks have to do with it? *Horm. Behav.* 74, 86–104. doi: 10.1016/j.yhbeh.2015.06.009
- Koebele, S. V., and Bimonte-Nelson, H. A. (2016). Modeling menopause: the utility of rodents in translational behavioral endocrinology research. *Maturitas* 87, 5–17. doi: 10.1016/j.maturitas.2016.01.015
- Koebele, S. V., and Bimonte-Nelson, H. A. (2017). The endocrine-brain-aging triad where many paths meet: female reproductive hormone changes at midlife and their influence on circuits important for learning and memory. *Exp. Gerontol.* 94, 14–23. doi: 10.1016/j.exger.2016.12.011
- Koebele, S. V., Mennenga, S. E., Hiroi, R., Quihuis, A. M., Hewitt, L. T., Poisson, M. L., et al. (2017). Cognitive changes across the menopause transition: a longitudinal evaluation of the impact of age and ovarian status on spatial memory. *Horm. Behav.* 87, 96–114. doi: 10.1016/j.yhbeh.2016.10.010
- Koran, M. E. I., Wagener, M., Hohman, T. J., and Alzheimer's Neuroimaging Initiative (2017). Sex differences in the association between AD biomarkers and cognitive decline. *Brain Imaging Behav.* 11, 205–213. doi: 10.1007/s11682-016-9523-8
- Korol, D. L., and Pisani, S. L. (2015). Estrogens and cognition: friends or foes?: an evaluation of the opposing effects of estrogens on learning and memory. *Horm. Behav.* 74, 105–115. doi: 10.1016/j.yhbeh.2015.06.017
- Kraft, A. W., Hu, X., Yoon, H., Yan, P., Xiao, Q., Wang, Y., et al. (2013). Attenuating astrocyte activation accelerates plaque pathogenesis in APP/PS1 mice. *FASEB J.* 27, 187–198. doi: 10.1096/fj.12-208660
- Krueyer, A., Soplop, N., Strickland, S., and Norris, E. H. (2015). Chronic hypertension leads to neurodegeneration in the TgSwDI mouse model of Alzheimer's disease. *Hypertension* 66, 175–182. doi: 10.1161/HYPERTENSIONAHA.115.05524
- Kua, E. H., Ho, E., Tan, H. H., Tsoi, C., Thng, C., and Mahendran, R. (2014). The natural history of dementia. *Psychogeriatrics* 14, 196–201. doi: 10.1111/psygy.12053
- Kujoth, G. C., Bradshaw, P. C., Haroon, S., and Prolla, T. A. (2007). The role of mitochondrial DNA mutations in mammalian aging. *PLoS Genet.* 3:e24. doi: 10.1371/journal.pgen.0030024
- Landau, S. M., Mintun, M. A., Joshi, A. D., Koeppe, R. A., Petersen, R. C., Aisen, P. S., et al. (2012). Amyloid deposition, hypometabolism and longitudinal cognitive decline. *Ann. Neurol.* 72, 578–586. doi: 10.1002/ana.23650
- Lane, C. A., Hardy, J., and Schott, J. M. (2018). Alzheimer's disease. *Eur. J. Neurol.* 25, 59–70. doi: 10.1111/ene.13439
- Laske, C., Stransky, E., Leyhe, T., Eschweiler, G. W., Schott, K., Langer, H., et al. (2006a). Decreased brain-derived neurotrophic factor (BDNF)- and  $\beta$ -thromboglobulin ( $\beta$ -TG)- blood levels in Alzheimer's disease. *Thromb. Haemost.* 96, 102–103. doi: 10.1160/th06-03-0173
- Laske, C., Stransky, E., Leyhe, T., Eschweiler, G. W., Wittorf, A., Richartz, E., et al. (2006b). Stage-dependent BDNF serum concentrations in Alzheimer's disease. *J. Neural Transm.* 113, 1217–1224. doi: 10.1007/s00702-005-0397-y

- Laws, K. R., Irvine, K., and Gale, T. M. (2016). Sex differences in cognitive impairment in Alzheimer's disease. *World J. Psychiatry* 6, 54–65. doi: 10.5498/wjp.v6.i1.54
- Leduc, V., Jasmin-Belanger, S., and Poirier, J. (2010). APOE and cholesterol homeostasis in Alzheimer's disease. *Trends Mol. Med.* 16, 469–477. doi: 10.1016/j.molmed.2010.07.008
- Lee, J. H., Jiang, Y., Han, D. H., Shin, S. K., Choi, W. H., and Lee, M. J. (2014). Targeting estrogen receptors for the treatment of Alzheimer's disease. *Mol. Neurobiol.* 49, 39–49. doi: 10.1007/s12035-013-8484-9
- Levin-Allerhand, J. A., Lominska, C. E., Wang, J., and Smith, J. D. (2002). 17 $\alpha$ -estradiol and 17 $\beta$ -estradiol treatments are effective in lowering cerebral amyloid- $\beta$  levels in A $\beta$ PPSWE transgenic mice. *J. Alzheimers Dis.* 4, 449–457. doi: 10.3233/jad-2002-4601
- Levin-Allerhand, J. A., and Smith, J. D. (2002). Ovariectomy of young mutant amyloid precursor protein transgenic mice leads to increased mortality. *J. Mol. Neurosci.* 19, 163–166. doi: 10.1007/s12031-002-0027-1
- Li, R., Cui, J., and Shen, Y. (2014). Brain sex matters: estrogen in cognition and Alzheimer's disease. *Mol. Cell. Endocrinol.* 389, 13–21. doi: 10.1016/j.mce.2013.12.018
- Li, R., Shen, Y., Yang, L. B., Lue, L. F., Finch, C., and Rogers, J. (2000). Estrogen enhances uptake of amyloid  $\beta$ -protein by microglia derived from the human cortex. *J. Neurochem.* 75, 1447–1454. doi: 10.1046/j.1471-4159.2000.0751447.x
- Li, R., and Singh, M. (2014). Sex differences in cognitive impairment and Alzheimer's disease. *Front. Neuroendocrinol.* 35, 385–403. doi: 10.1016/j.yfrne.2014.01.002
- Liang, K., Yang, L., Yin, C., Xiao, Z., Zhang, J., Liu, Y., et al. (2010). Estrogen stimulates degradation of  $\beta$ -amyloid peptide by up-regulating neprilysin. *J. Biol. Chem.* 285, 935–942. doi: 10.1074/jbc.m109.051664
- Lima, R., Wofford, M., and Reckelhoff, J. F. (2012). Hypertension in postmenopausal women. *Curr. Hypertens. Rep.* 14, 254–260. doi: 10.1007/s11906-012-0260-0
- Lin, K. A., Choudhury, K. R., Rathakrishnan, B. G., Marks, D. M., Petrella, J. R., Doraiswamy, P. M., et al. (2015). Marked gender differences in progression of mild cognitive impairment over 8 years. *Alzheimers Dement.* 1, 103–110. doi: 10.1016/j.trci.2015.07.001
- Liu, C. C., Hu, J., Zhao, N., Wang, J., Wang, N., Cirrito, J. R., et al. (2017). Astrocytic LRP1 mediates brain A $\beta$  clearance and impacts amyloid deposition. *J. Neurosci.* 37, 4023–4031. doi: 10.1523/JNEUROSCI.3442-16.2017
- Lin, K. A., and Doraiswamy, P. M. (2014). When mars versus venus is not a cliché: gender differences in the neurobiology of Alzheimer's disease. *Front. Neurol.* 5:288. doi: 10.3389/fneur.2014.00288
- Lohff, J. C., Christian, P. J., Marion, S. L., and Hoyer, P. B. (2006). Effect of duration of dosing on onset of ovarian failure in a chemical-induced mouse model of perimenopause. *Menopause* 13, 482–488. doi: 10.1097/01.gme.00000191883.59799.2e
- Luine, V. N. (2014). Estradiol and cognitive function: past, present and future. *Horm. Behav.* 66, 602–618. doi: 10.1016/j.yhbeh.2014.08.011
- Maffucci, J. A., and Gore, A. C. (2006). "Age-related changes in hormones and their receptors in animal models of female reproductive senescence," in *Handbook of Models for Human Aging*, ed. P. M. Conn (London: Elsevier), 533–552.
- Maki, P. M., and Henderson, V. W. (2012). Hormone therapy, dementia and cognition: the Women's Health Initiative 10 years on. *Climacteric* 15, 256–262. doi: 10.3109/13697137.2012.660613
- Maki, P. M., and Henderson, V. W. (2016). Cognition and the menopause transition. *Menopause* 23, 803–805. doi: 10.1097/gme.0000000000000681
- Manocha, G. D., Floden, A. M., Rausch, K., Kulas, J. A., McGregor, B. A., Rojanathammanee, L., et al. (2016). APP regulates microglial phenotype in a mouse model of Alzheimer's disease. *J. Neurosci.* 36, 8471–8486. doi: 10.1523/jneurosci.4654-15.2016
- Manson, J. E., Aragaki, A. K., Rossouw, J. E., Anderson, G. L., Prentice, R. L., LaCroix, A. Z., et al. (2017). Menopausal hormone therapy and long-term all-cause and cause-specific mortality: the women's health initiative randomized trials. *JAMA* 318, 927–938. doi: 10.1001/jama.2017.11217
- Manthey, D., Heck, S., Engert, S., and Behl, C. (2001). Estrogen induces a rapid secretion of amyloid  $\beta$  precursor protein via the mitogen-activated protein kinase pathway. *Eur. J. Biochem.* 268, 4285–4291. doi: 10.1046/j.1432-1327.2001.02346.x
- Maranon, R. O., Lima, R., Mathbout, M., do Carmo, J. M., Hall, J. E., Roman, R. J., et al. (2014). Postmenopausal hypertension: role of the sympathetic nervous system in an animal model. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 306, R248–R256. doi: 10.1152/ajpregu.00490.2013
- Marques-Lopes, J., Lynch, M. K., Van Kempen, T. A., Waters, E. M., Wang, G., Iadecola, C., et al. (2015). Female protection from slow-pressor effects of angiotensin II involves prevention of ROS production independent of NMDA receptor trafficking in hypothalamic neurons expressing angiotensin 1A receptors. *Synapse* 69, 148–165. doi: 10.1002/syn.21800
- Marques-Lopes, J., Tesfaye, E., Israilov, S., Van Kempen, T. A., Wang, G., Glass, M. J., et al. (2017). Redistribution of NMDA receptors in estrogen-receptor- $\beta$ -containing paraventricular hypothalamic neurons following slow-pressor angiotensin ii hypertension in female mice with accelerated ovarian failure. *Neuroendocrinology* 104, 239–256. doi: 10.1159/000446073
- Marques-Lopes, J., Van Kempen, T. A., and Milner, T. A. (2018). "Rodent models of ovarian failure," in *Conn's Handbook of Models for Human Aging*, ed. J. L. P. M. Conn (London: Elsevier Inc.), 831–844.
- Marques-Lopes, J., Van Kempen, T., Waters, E. M., Pickel, V. M., Iadecola, C., and Milner, T. A. (2014). Slow-pressor angiotensin II hypertension and concomitant dendritic NMDA receptor trafficking in estrogen receptor  $\beta$ -containing neurons of the mouse hypothalamic paraventricular nucleus are sex and age dependent. *J. Comp. Neurol.* 522, 3075–3090. doi: 10.1002/cne.23569
- Matarin, M., Salih, D. A., Yasvoina, M., Cummings, D. M., Guelfi, S., Liu, W., et al. (2015). A genome-wide gene-expression analysis and database in transgenic mice during development of amyloid or tau pathology. *Neurochem. Int.* 10, 633–644. doi: 10.1016/j.celrep.2014.12.041
- Mayer, L. P., Devine, P. J., Dyer, C. A., and Hoyer, P. B. (2004). The follicle-deplete mouse ovary produces androgen. *Biol. Reprod.* 71, 130–138. doi: 10.1095/biolreprod.103.016113
- Mayer, L. P., Dyer, C. A., Eastgard, R. L., Hoyer, P. B., and Banka, C. L. (2005). Atherosclerotic lesion development in a novel ovary-intact mouse model of perimenopause. *Arterioscler. Thromb. Vasc. Biol.* 25, 1910–1916. doi: 10.1161/01.atv.0000175767.46520.6a
- Mayer, L. P., Pearsall, N. A., Christian, P. J., Devine, P. J., Payne, C. M., McCuskey, M. K., et al. (2002). Long-term effects of ovarian follicular depletion in rats by 4-vinylcyclohexene diepoxide. *Reprod. Toxicol.* 16, 775–781. doi: 10.1016/s0890-6238(02)00048-5
- Mazure, C. M., and Swendsen, J. (2016). Sex differences in Alzheimer's disease and other dementias. *Lancet Neurol.* 15, 451–452. doi: 10.1016/S1474-4422(16)00067-3
- McAsey, M. E., Cady, C., Jackson, L. M., Li, M., Randall, S., Nathan, B. P., et al. (2006). Time course of response to estradiol replacement in ovariectomized mice: brain apolipoprotein E and synaptophysin transiently increase and glial fibrillary acidic protein is suppressed. *Exp. Neurol.* 197, 197–205. doi: 10.1016/j.expneurol.2005.09.008
- McCarney, A. C., and Resnick, S. M. (2015). Postmenopausal hormone therapy and cognition. *Horm. Behav.* 74, 167–172. doi: 10.1016/j.yhbeh.2015.04.018
- McEwen, B., Akama, K., Alves, S., Brake, W. G., Bulloch, K., Lee, S., et al. (2001). Tracking the estrogen receptor in neurons: implications for estrogen-induced synapse formation. *Proc. Natl. Acad. Sci. U S A* 98, 7093–7100. doi: 10.1073/pnas.121146898
- McEwen, B. S., Akama, K. T., Spencer-Segal, J. L., Milner, T. A., and Waters, E. M. (2012). Estrogen effects on the brain: actions beyond the hypothalamus via novel mechanisms. *Behav. Neurosci.* 126, 4–16. doi: 10.1037/a0026708
- McEwen, B. S., and Milner, T. A. (2017). Understanding the broad influence of sex hormones and sex differences in the brain. *J. Neurosci. Res.* 95, 24–39. doi: 10.1002/jnr.23809
- McGee, E. A., and Hsueh, A. J. (2000). Initial and cyclic recruitment of ovarian follicles. *Endocr. Rev.* 21, 200–214. doi: 10.1210/edrv.21.2.0394
- Merlo, S., Spampinato, S. F., and Sortino, M. A. (2017). Estrogen and Alzheimer's disease: Still an attractive topic despite disappointment from early clinical results. *Eur. J. Pharmacol.* 817, 51–58. doi: 10.1016/j.ejphar.2017.05.059
- Mielke, M. M., Vemuri, P., and Rocca, W. A. (2014). Clinical epidemiology of Alzheimer's disease: assessing sex and gender differences. *Clin. Epidemiol.* 6, 37–48. doi: 10.2147/ceip.s37929

- Milner, T. A., Ayoola, K., Drake, C. T., Herrick, S. P., Tabori, N. E., McEwen, B. S., et al. (2005). Ultrastructural localization of estrogen receptor  $\beta$  immunoreactivity in the rat hippocampal formation. *J. Comp. Neurol.* 491, 81–95. doi: 10.1002/cne.20724
- Milner, T. A., McEwen, B. S., Hayashi, S., Li, C. J., Reagan, L. P., and Alves, S. E. (2001). Ultrastructural evidence that hippocampal  $\alpha$  estrogen receptors are located at extranuclear sites. *J. Comp. Neurol.* 429, 355–371. doi: 10.1002/1096-9861(20010115)429:3<355::aid-cne1>3.3.co;2-r
- Mitterling, K. L., Spencer, J. L., Dziedzic, N., Shenoy, S., McCarthy, K., Waters, E. M., et al. (2010). Cellular and subcellular localization of estrogen and progesterone receptor immunoreactivities in the mouse hippocampus. *J. Comp. Neurol.* 518, 2729–2743. doi: 10.1002/cne.22361
- Montine, T. J., Phelps, C. H., Beach, T. G., Bigio, E. H., Cairns, N. J., Dickson, D. W., et al. (2012). National institute on aging-Alzheimer's association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. *Acta Neuropathol.* 123, 1–11. doi: 10.1007/s00401-011-0910-3
- Morgan, T. E., and Finch, C. E. (2015). Astrocytic estrogen receptors and impaired neurotrophic responses in a rat model of perimenopause. *Front. Aging Neurosci.* 7:179. doi: 10.3389/fnagi.2015.00179
- Morrison, J. H., Brinton, R. D., Schmidt, P. J., and Gore, A. C. (2006). Estrogen, menopause and the aging brain: how basic neuroscience can inform hormone therapy in women. *J. Neurosci.* 26, 10332–10348. doi: 10.1523/JNEUROSCI.3369-06.2006
- Mortensen, E. L., and Hogh, P. (2001). A gender difference in the association between APOE genotype and age-related cognitive decline. *Neurology* 57, 89–95. doi: 10.1212/wnl.57.1.89
- Mosconi, L., Berti, V., Quinn, C., McHugh, P., Petrongolo, G., Osorio, R. S., et al. (2017a). Perimenopause and emergence of an Alzheimer's bioenergetic phenotype in brain and periphery. *PLoS One* 12:e0185926. doi: 10.1371/journal.pone.0185926
- Mosconi, L., Berti, V., Quinn, C., McHugh, P., Petrongolo, G., Varsavsky, I., et al. (2017b). Sex differences in Alzheimer risk: brain imaging of endocrine vs chronological aging. *Neurology* 89, 1382–1390. doi: 10.1212/WNL.0000000000004425
- Mukai, H., Kimoto, T., Hojo, Y., Kawato, S., Murakami, G., Higo, S., et al. (2010). Modulation of synaptic plasticity by brain estrogen in the hippocampus. *Biochim. Biophys. Acta* 1800, 1030–1044. doi: 10.1016/j.bbagen.2009.11.002
- Nagahara, A. H., and Tuszynski, M. H. (2011). Potential therapeutic uses of BDNF in neurological and psychiatric disorders. *Nat. Rev. Drug Discov.* 10, 209–219. doi: 10.1038/nrd3366
- Nagahara, A. H., Merrill, D. A., Coppola, G., Tsukada, S., Schroeder, B. E., Shaked, G. M., et al. (2009). Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nat. Med.* 15, 331–337. doi: 10.1038/nm.1912
- National Toxicology Program (1986). NTP toxicology and carcinogenesis studies of 4-vinylcyclohexene (CAS No. 100–40–3) in F344/N rats and B6C3F1 mice (Gavage Studies). *Natl. Toxicol. Program. Tech. Rep. Ser.* 303, 1–190.
- Nebel, R. A., Aggarwal, N. T., Barnes, L. L., Gallagher, A., Goldstein, J. M., Kantarci, K., et al. (2018). Understanding the impact of sex and gender in Alzheimer's disease: a call to action. *Alzheimers Dement.* 14, 1171–1183. doi: 10.1016/j.jalz.2018.04.008
- Nejat, E. J., and Chervenak, J. L. (2010). The continuum of ovarian aging and clinicopathologies associated with the menopausal transition. *Maturitas* 66, 187–190. doi: 10.1016/j.maturitas.2010.02.017
- Nelson, P. T., Braak, H., and Markesbery, W. R. (2009). Neuropathology and cognitive impairment in Alzheimer disease: a complex but coherent relationship. *J. Neuropathol. Exp. Neurol.* 68, 1–14. doi: 10.1097/nen.0b013e3181919a48
- Neu, S. C., Pa, J., Kukull, W., Beekly, D., Kuzma, A., Gangadharan, P., et al. (2017). Apolipoprotein E genotype and sex risk factors for Alzheimer disease: a meta-analysis. *JAMA Neurol.* 74, 1178–1189. doi: 10.1001/jamaneurol.2017.2188
- Noguchi, S., Murakami, K., and Yamada, N. (1993). Apolipoprotein E genotype and Alzheimer's disease. *Lancet* 342:737. doi: 10.1016/0140-6736(93)91728-5
- Olson, M. E., and Bruce, J. (1986). Ovariectomy, ovariectomy and orchidectomy in rodents and rabbits. *Can. Vet. J.* 27, 523–527.
- Ossenkoppe, R., Jansen, W. J., Rabinovici, G. D., Knol, D. L., van der Flier, W. M., van Berckel, B. N., et al. (2015). Prevalence of amyloid PET positivity in dementia syndromes: a meta-analysis. *JAMA* 313, 1939–1949. doi: 10.1001/jama.2015.4669
- Parachikova, A., Agadjanyan, M. G., Cribbs, D. H., Blurton-Jones, M., Perreau, V., Rogers, J., et al. (2007). Inflammatory changes parallel the early stages of Alzheimer disease. *Neurobiol. Aging* 28, 1821–1833. doi: 10.1016/j.neurobiolaging.2006.08.014
- Peters, H., Byskov, A. G., Himelstein-Braw, R., and Faber, M. (1975). Follicular growth: the basic event in the mouse and human ovary. *J. Reprod. Fertil.* 45, 559–566. doi: 10.1530/jrf.0.0450559
- Pike, C. J. (2017). Sex and the development of Alzheimer's disease. *J. Neurosci. Res.* 95, 671–680. doi: 10.1002/jnr.23827
- Pines, A. (2016). Alzheimer's disease, menopause and the impact of the estrogenic environment. *Climacteric* 19, 430–432. doi: 10.1080/13697137.2016.1201319
- Practice Committee of the American Society for Reproductive Medicine (2008). The menopausal transition. *Fertil Steril* 90, S61–S65. doi: 10.1016/j.fertnstert.2008.08.095
- Price, J. L., Davis, P. B., Morris, J. C., and White, D. L. (1991). The distribution of tangles, plaques and related immunohistochemical markers in healthy aging and Alzheimer's disease. *Neurobiol. Aging* 12, 295–312. doi: 10.1016/0197-4580(91)90006-6
- Pugliese, L., Tanzi, R. E., and Kovacs, D. M. (2003). Alzheimer's disease: the cholesterol connection. *Nat. Neurosci.* 6, 345–351. doi: 10.1038/nn0403-345
- Rapp, P. R., Morrison, J. H., and Roberts, J. A. (2003). Cyclic estrogen replacement improves cognitive function in aged ovariectomized rhesus monkeys. *J. Neurosci.* 23, 5708–5714. doi: 10.1523/JNEUROSCI.23-13-057.08.2003
- Rentz, D. M., Weiss, B. K., Jacobs, E. G., Cherkertzian, S., Klubanski, A., Remington, A., et al. (2017). Sex differences in episodic memory in early midlife: impact of reproductive aging. *Menopause* 24, 400–408. doi: 10.1097/gme.0000000000000771
- Resnick, S. M., and Henderson, V. W. (2002). Hormone therapy and risk of Alzheimer disease: a critical time. *JAMA* 288, 2170–2172. doi: 10.1001/jama.288.17.2170
- Riedel, B. C., Thompson, P. M., and Brinton, R. D. (2016). Age, APOE and sex: triad of risk of Alzheimer's disease. *J. Steroid Biochem. Mol. Biol.* 160, 134–147. doi: 10.1016/j.jsbmb.2016.03.012
- Rivera, Z., Christian, P. J., Marion, S. L., Brooks, H. L., and Hoyer, P. B. (2009). Steroidogenic capacity of residual ovarian tissue in 4-vinylcyclohexene diepoxide-treated mice. *Biol. Reprod.* 80, 328–336. doi: 10.1095/biolreprod.108.070359
- Rocca, W. A., Grossardt, B. R., and Shuster, L. T. (2011). Oophorectomy, menopause, estrogen treatment, and cognitive aging: clinical evidence for a window of opportunity. *Brain Res.* 1379, 188–198. doi: 10.1016/j.brainres.2010.10.031
- Rozovsky, I., Hoving, S., Anderson, C. P., O'Callaghan, J., and Finch, C. E. (2002). Equine estrogens induce apolipoprotein E and glial fibrillary acidic protein in mixed glial cultures. *Neurosci. Lett.* 323, 191–194. doi: 10.1016/s0304-3940(02)00146-5
- Sahambi, S. K., Visser, J. A., Themmen, A. P., Mayer, L. P., and Devine, P. J. (2008). Correlation of serum anti-Mullerian hormone with accelerated follicle loss following 4-vinylcyclohexene diepoxide-induced follicle loss in mice. *Reprod. Toxicol.* 26, 116–122. doi: 10.1016/j.reprotox.2008.07.005
- Sandberg, K., and Ji, H. (2012). Sex differences in primary hypertension. *Biol. Sex Differ.* 3:7. doi: 10.1186/2042-6410-3-7
- Sárvári, M., Hrabovszky, E., Kalló, I., Solymosi, N., Likó, I., Berchtold, N., et al. (2012). Menopause leads to elevated expression of macrophage-associated genes in the aging frontal cortex: rat and human studies identify strikingly similar changes. *J. Neuroinflammation* 9:264. doi: 10.1186/1742-2094-9-264
- Scharfman, H. E., Hintz, T. M., Gomez, J., Stormes, K. A., Barouk, S., Malthankar-Phatak, G. H., et al. (2007). Changes in hippocampal function of ovariectomized rats after sequential low doses of estradiol to simulate the preovulatory estrogen surge. *Eur. J. Neurosci.* 26, 2595–2612. doi: 10.1111/j.1460-9568.2007.05848.x



- Scheltens, P., Blennow, K., Breteler, M. M., de Strooper, B., Frisoni, G. B., Salloway, S., et al. (2016). Alzheimer's disease. *Lancet* 388, 505–517. doi: 10.1016/S0140-6736(15)01124-1
- Schmidt, R., Kienbacher, E., Benke, T., Dal-Bianco, P., Delazer, M., Ladurner, G., et al. (2008). Sex differences in Alzheimer's disease. *Neuropsychiatr* 22, 1–15.
- Shah, N. S., Vidal, J. S., Masaki, K., Petrovitch, H., Ross, G. W., Tilley, C., et al. (2012). Midlife blood pressure, plasma  $\beta$ -amyloid, and the risk for Alzheimer disease: the Honolulu Asia Aging Study. *Hypertension* 59, 780–786. doi: 10.1161/hypertensionaha.111.178962
- Shao, H., Breitner, J. C., Whitmer, R. A., Wang, J., Hayden, K., Wengreen, H., et al. (2012). Hormone therapy and Alzheimer disease dementia: new findings from the Cache County Study. *Neurology* 79, 1846–1852. doi: 10.1212/WNL.0b013e318271f823
- Shumaker, S. A., Legault, C., Kuller, L., Rapp, S. R., Thal, L., Lane, D. S., et al. (2004). Conjugated equine estrogens and incidence of probable dementia and mild cognitive impairment in postmenopausal women: Women's Health Initiative Memory Study. *JAMA* 291, 2947–2958. doi: 10.1001/jama.291.24.2947
- Shumaker, S. A., Legault, C., Rapp, S. R., Thal, L., Wallace, R. B., Ockene, J. K., et al. (2003). Estrogen plus progestin and the incidence of dementia and mild cognitive impairment in postmenopausal women: the Women's Health Initiative Memory Study: a randomized controlled trial. *JAMA* 289, 2651–2662. doi: 10.1001/jama.289.20.2651
- Shuster, L. T., Rhodes, D. J., Gostout, B. S., Grossardt, B. R., and Rocca, W. A. (2010). Premature menopause or early menopause: long-term health consequences. *Climacteric* 65, 161–166. doi: 10.1016/j.maturitas.2009.08.003
- Sitruk-Ware, R. (2002). Hormonal replacement therapy. *Rev. Endocr. Metab. Disord.* 3, 243–256. doi: 10.1023/A:1020028510797
- Skup, M., Zhu, H., Wang, Y., Giovanello, K. S., Lin, J. A., Shen, D., et al. (2011). Sex differences in grey matter atrophy patterns among AD and aMCI patients: results from ADNI. *Neuroimage* 56, 890–906. doi: 10.1016/j.neuroimage.2011.02.060
- Smith, C. C., Vedder, L. C., Nelson, A. R., Bredemann, T. M., and McMahon, L. L. (2010). Duration of estrogen deprivation, not chronological age, prevents estrogen's ability to enhance hippocampal synaptic physiology. *Proc. Natl. Acad. Sci. U S A* 107, 19543–19548. doi: 10.1073/pnas.1009307107
- Song, F., Qian, Y., Peng, X., Han, G., Wang, J., Bai, Z., et al. (2016). Perturbation of the transcriptome: implications of the innate immune system in Alzheimer's disease. *Curr. Opin. Pharmacol.* 26, 47–53. doi: 10.1016/j.coph.2015.09.015
- Spencer, J. L., Waters, E. M., Milner, T. A., Lee, F. S., and McEwen, B. S. (2010). BDNF variant Val66Met interacts with estrous cycle in the control of hippocampal function. *Proc. Natl. Acad. Sci. U S A* 107, 4395–4400. doi: 10.1073/pnas.0915105107
- Spencer, J. L., Waters, E. M., Romeo, R. D., Wood, G. E., Milner, T. A., and McEwen, B. S. (2008). Uncovering the mechanisms of estrogen effects on hippocampal function. *Front. Neuroendocrinol.* 29, 219–237. doi: 10.1016/j.yfrne.2007.08.006
- Spencer-Segal, J. L., Tsuda, M. C., Mattei, L., Waters, E. M., Romeo, R. D., Milner, T. A., et al. (2012). Estradiol acts via estrogen receptors  $\alpha$  and  $\beta$  on pathways important for synaptic plasticity in the mouse hippocampal formation. *Neuroscience* 202, 131–146. doi: 10.1016/j.neuroscience.2011.11.035
- Spencer-Segal, J. L., Waters, E. M., Bath, K. G., Chao, M. V., McEwen, B. S., and Milner, T. A. (2011). Distribution of phosphorylated TrkB receptor in the mouse hippocampal formation depends on sex and estrous cycle stage. *J. Neurosci.* 31, 6780–6790. doi: 10.1523/JNEUROSCI.0910-11.2011
- Sundermann, E. E., Biegion, A., Rubin, L. H., Lipton, R. B., Landau, S., Maki, P. M., et al. (2017). Does the female advantage in verbal memory contribute to underestimating Alzheimer's disease pathology in women versus men? *J. Alzheimers Dis.* 56, 947–957. doi: 10.3233/jad-160716
- Sundermann, E. E., Biegion, A., Rubin, L. H., Lipton, R. B., Mowrey, W., Landau, S., et al. (2016a). Better verbal memory in women than men in MCI despite similar levels of hippocampal atrophy. *Neurology* 86, 1368–1376. doi: 10.1212/wnl.0000000000002570
- Sundermann, E. E., Maki, P. M., Rubin, L. H., Lipton, R. B., Landau, S., Biegion, A., et al. (2016b). Female advantage in verbal memory: evidence of sex-specific cognitive reserve. *Neurology* 87, 1916–1924. doi: 10.1212/wnl.0000000000003288
- Tapia-Arancibia, L., Aliaga, E., Silhol, M., and Arancibia, S. (2008). New insights into brain BDNF function in normal aging and Alzheimer disease. *Brain Res. Rev.* 59, 201–220. doi: 10.1016/j.brainresrev.2008.07.007
- Tiwari, S., Li, L., Riaz, S., Halagappa, V. K., and Ecelbarger, C. M. (2009). Sex and age result in differential regulation of the renal thiazide-sensitive NaCl cotransporter and the epithelial sodium channel in angiotensin II-infused mice. *Am. J. Nephrol.* 30, 554–562. doi: 10.1159/000252776
- Todd, S., Barr, S., Roberts, M., and Passmore, A. P. (2013). Survival in dementia and predictors of mortality: a review. *Int. J. Geriatr. Psychiatry* 28, 1109–1124. doi: 10.1002/gps.3946
- U.S. Census Bureau. (2017). *2017 National Population Projections Tables*. Suitland, MD: U.S. Census Bureau.
- Van Kempen, T. A., Dodos, M., Woods, C., Marques-Lopes, J., Justice, N. J., Iadecola, C., et al. (2015). Sex differences in NMDA GluN1 plasticity in rostral ventrolateral medulla neurons containing corticotropin-releasing factor type 1 receptor following slow-pressor angiotensin II hypertension. *Neuroscience* 307, 83–97. doi: 10.1016/j.neuroscience.2015.08.029
- Van Kempen, T. A., Gorecka, J., Gonzalez, A. D., Soeda, F., Milner, T. A., and Waters, E. M. (2014). Characterization of neural estrogen signaling and neurotrophic changes in the accelerated ovarian failure mouse model of menopause. *Endocrinology* 155, 3610–3623. doi: 10.1210/en.2014-1190
- Van Kempen, T. A., Milner, T. A., and Waters, E. M. (2011). Accelerated ovarian failure: a novel, chemically induced animal model of menopause. *Brain Res.* 1379, 176–187. doi: 10.1016/j.brainres.2010.12.064
- Van Kempen, T. A., Narayan, A., Waters, E. M., Marques-Lopes, J., Iadecola, C., Glass, M. J., et al. (2016). Alterations in the subcellular distribution of NADPH oxidase p47(phox) in hypothalamic paraventricular neurons following slow-pressor angiotensin II hypertension in female mice with accelerated ovarian failure. *J. Comp. Neurol.* 524, 2251–2265. doi: 10.1002/cne.23944
- Vest, R. S., and Pike, C. J. (2013). Gender, sex steroid hormones, and Alzheimer's disease. *Horm. Behav.* 63, 301–307. doi: 10.1016/j.yhbeh.2012.04.006
- Villa, A., Vegeto, E., Poletti, A., and Maggi, A. (2016). Estrogens, neuroinflammation, and neurodegeneration. *Endocr. Rev.* 37, 372–402. doi: 10.1210/er.2016-1007
- Walker, M. L., and Herndon, J. G. (2008). Menopause in nonhuman primates? *Biol. Reprod.* 79, 398–406. doi: 10.1095/biolreprod.108.068536
- Wang, Y., Cella, M., Mallinson, K., Ulrich, J. D., Young, K. L., Robinette, M. L., et al. (2015). TREM2 lipid sensing sustains the microglial response in an Alzheimer's disease model. *Cell* 160, 1061–1071. doi: 10.1016/j.cell.2015.01.049
- Wang, J. M., Irwin, R. W., and Brinton, R. D. (2006). Activation of estrogen receptor  $\alpha$  increases and estrogen receptor  $\beta$  decreases apolipoprotein E expression in hippocampus *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. U S A* 103, 16983–16988. doi: 10.1073/pnas.0608128103
- Wang, Y., and Mandelkow, E. (2016). Tau in physiology and pathology. *Nat. Rev. Neurosci.* 17, 5–21. doi: 10.1038/nrn.2015.1
- Wang, J., Tanila, H., Puolivali, J., Kadish, I., and van Groen, T. (2003). Gender differences in the amount and deposition of amyloid  $\beta$  in APPsw and PS1 double transgenic mice. *Neurobiol. Dis.* 14, 318–327. doi: 10.1016/j.nbd.2003.08.009
- Wang, Y., Ulland, T. K., Ulrich, J. D., Song, W., Tzaferis, J. A., Hole, J. T., et al. (2016). TREM2-mediated early microglial response limits diffusion and toxicity of amyloid plaques. *J. Exp. Med.* 213, 667–675. doi: 10.1084/jem.20151948
- Waters, E. M., Mitterling, K., Spencer, J. L., Mazid, S., McEwen, B. S., and Milner, T. A. (2009). Estrogen receptor  $\alpha$  and  $\beta$  specific agonists regulate expression of synaptic proteins in rat hippocampus. *Brain Res.* 1290, 1–11. doi: 10.1016/j.brainres.2009.06.090
- Waters, E. M., Thompson, L. I., Patel, P., Gonzales, A. D., Ye, H. Z., Filardo, E. J., et al. (2015). G-protein-coupled estrogen receptor 1 is anatomically positioned to modulate synaptic plasticity in the mouse hippocampus. *J. Neurosci.* 35, 2384–2397. doi: 10.1523/JNEUROSCI.1298-14.2015
- Wattmo, C., Londos, E., and Minthon, L. (2014a). Response to cholinesterase inhibitors affects lifespan in Alzheimer's disease. *BMC Neurol.* 14:173. doi: 10.1186/s12883-014-0173-4



- Wattmo, C., Londos, E., and Minthon, L. (2014b). Risk factors that affect life expectancy in Alzheimer's disease: a 15-year follow-up. *Dement. Geriatr. Cogn. Disord.* 38, 286–299. doi: 10.1159/000362926
- Wei, S. M., Baller, E. B., Kohn, P. D., Kippenhan, J. S., Kolachana, B., Soldin, S. J., et al. (2018). Brain-derived neurotrophic factor Val<sup>66</sup>Met genotype and ovarian steroids interactively modulate working memory-related hippocampal function in women: a multimodal neuroimaging study. *Mol. Psychiatry* 23, 1066–1075. doi: 10.1038/mp.2017.72
- Wei, Y. C., Wang, S. R., and Xu, X. H. (2017). Sex differences in brain-derived neurotrophic factor signaling: functions and implications. *J. Neurosci. Res.* 95, 336–344. doi: 10.1002/jnr.23897
- Whitmer, R. A., Quesenberry, C. P., Zhou, J., and Yaffe, K. (2011). Timing of hormone therapy and dementia: the critical window theory revisited. *Ann. Neurol.* 69, 163–169. doi: 10.1002/ana.22239
- Wiesmann, M., Roelofs, M., van der Lugt, R., Heerschap, A., Kiliaan, A. J., and Claassen, J. A. (2017). Angiotensin II, hypertension and angiotensin II receptor antagonism: Roles in the behavioural and brain pathology of a mouse model of Alzheimer's disease. *J. Cereb. Blood Flow Metab.* 37, 2396–2413. doi: 10.1177/0271678x16667364
- Williams, J. K. (2005). A mouse model of the perimenopausal transition: importance for cardiovascular research. *Arterioscler. Thromb. Vasc. Biol.* 25, 1765–1766. doi: 10.1161/01.atv.0000175757.28698.c2
- Woolley, C. S., and McEwen, B. S. (1992). Estradiol mediates fluctuation in hippocampal synapse density during the estrous cycle in the adult rat. *J. Neurosci.* 12, 2549–2554. doi: 10.1523/JNEUROSCI.12-10-j0001.1992
- Wright, L. E., Christian, P. J., Rivera, Z., Van Alstine, W. G., Funk, J. L., Boussein, M. L., et al. (2008). Comparison of skeletal effects of ovariectomy versus chemically induced ovarian failure in mice. *J. Bone Miner. Res.* 23, 1296–1303. doi: 10.1359/jbmr.080309
- Xu, H., Gouras, G. K., Greenfield, J. P., Vincent, B., Naslund, J., Mazzarelli, L., et al. (1998). Estrogen reduces neuronal generation of Alzheimer  $\beta$ -amyloid peptides. *Nat. Med.* 4, 447–451. doi: 10.1038/nm0498-447
- Xu, H., Wang, R., Zhang, Y. W., and Zhang, X. (2006). Estrogen,  $\beta$ -amyloid metabolism/trafficking, and Alzheimer's disease. *Ann. N Y Acad. Sci.* 1089, 324–342. doi: 10.1196/annals.1386.036
- Xue, B., Badaue-Passos, D. Jr., Guo, F., Gomez-Sanchez, C. E., Hay, M., and Johnson, A. K. (2009). Sex differences and central protective effect of 17 $\beta$ -estradiol in the development of aldosterone/NaCl-induced hypertension. *Am. J. Physiol. Heart Circ. Physiol.* 296, H1577–H1585. doi: 10.1152/ajpheart.01255.2008
- Xue, B., Johnson, A. K., and Hay, M. (2013). Sex differences in angiotensin II- and aldosterone-induced hypertension: the central protective effects of estrogen. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 305, R459–R463. doi: 10.1152/ajpregu.00222.2013
- Xue, B., Zhang, Z., Beltz, T. G., Guo, F., Hay, M., and Johnson, A. K. (2014). Estrogen regulation of the brain renin-angiotensin system in protection against angiotensin II-induced sensitization of hypertension. *Am. J. Physiol. Heart Circ. Physiol.* 307, H191–H198. doi: 10.1152/ajpheart.01012.2013
- Yanes, L. L., and Reckelhoff, J. F. (2011). Postmenopausal hypertension. *Am. J. Hypertens.* 24, 740–749. doi: 10.1038/ajh.2011.71
- Yao, J., Hamilton, R. T., Cadenas, E., and Brinton, R. D. (2010). Decline in mitochondrial bioenergetics and shift to ketogenic profile in brain during reproductive senescence. *Biochim. Biophys. Acta* 1800, 1121–1126. doi: 10.1016/j.bbagen.2010.06.002
- Yao, J., Irwin, R., Chen, S., Hamilton, R., Cadenas, E., and Brinton, R. D. (2012). Ovarian hormone loss induces bioenergetic deficits and mitochondrial  $\beta$ -amyloid. *Neurobiol. Aging* 33, 1507–1521. doi: 10.1016/j.neurobiolaging.2011.03.001
- Yao, J., Irwin, R. W., Zhao, L., Nilsen, J., Hamilton, R. T., and Brinton, R. D. (2009). Mitochondrial bioenergetic deficit precedes Alzheimer's pathology in female mouse model of Alzheimer's disease. *Proc. Natl. Acad. Sci. U S A* 106, 14670–14675. doi: 10.1073/pnas.0903563106
- Yao, M., Nguyen, T. V., and Pike, C. J. (2007). Estrogen regulates Bcl-w and Bim expression: role in protection against  $\beta$ -amyloid peptide-induced neuronal death. *J. Neurosci.* 27, 1422–1433. doi: 10.1523/JNEUROSCI.2382-06.2007
- Yue, M., Hanna, A., Wilson, J., Roder, H., and Janus, C. (2011). Sex difference in pathology and memory decline in rTg4510 mouse model of tauopathy. *Neurobiol. Aging* 32, 590–603. doi: 10.1016/j.neurobiolaging.2009.04.006
- Yue, X., Lu, M., Lancaster, T., Cao, P., Honda, S., Staufienbiel, M., et al. (2005). Brain estrogen deficiency accelerates A $\beta$  plaque formation in an Alzheimer's disease animal model. *Proc. Natl. Acad. Sci. U S A* 102, 19198–19203. doi: 10.1073/pnas.0505203102
- Zanchetti, A., Facchetti, R., Cesana, G. C., Modena, M. G., Pirrelli, A., Sega, R., et al. (2005). Menopause-related blood pressure increase and its relationship to age and body mass index: the SIMONA epidemiological study. *J. Hypertens* 23, 2269–2276. doi: 10.1097/01.hjh.0000194118.35098.43
- Zandi, P. P., Carlson, M. C., Plassman, B. L., Welsh-Bohmer, K. A., Mayer, L. S., Steffens, D. C., et al. (2002). Hormone replacement therapy and incidence of Alzheimer disease in older women: the Cache County Study. *JAMA* 288, 2123–2129. doi: 10.1001/jama.288.17.2123
- Zhao, L., Mao, Z., Woody, S. K., and Brinton, R. D. (2016). Sex differences in metabolic aging of the brain: insights into female susceptibility to Alzheimer's disease. *Neurobiol. Aging* 42, 69–79. doi: 10.1016/j.neurobiolaging.2016.02.011
- Zhao, L., Yao, J., Mao, Z., Chen, S., Wang, Y., and Brinton, R. D. (2011). 17 $\beta$ -Estradiol regulates insulin-degrading enzyme expression via an ER $\beta$ /PI3-K pathway in hippocampus: relevance to Alzheimer's prevention. *Neurobiol. Aging* 32, 1949–1963. doi: 10.1016/j.neurobiolaging.2009.12.010
- Zheng, H., Xu, H., Uljon, S. N., Gross, R., Hardy, K., Gaynor, J., et al. (2002). Modulation of A( $\beta$ ) peptides by estrogen in mouse models. *J. Neurochem.* 80, 191–196. doi: 10.1046/j.0022-3042.2001.00690.x
- Zlokovic, B. V. (2011). Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat. Rev. Neurosci.* 12, 723–738. doi: 10.1038/nrn3114

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