



OPEN ACCESS

EDITED BY

Arun Malhotra,
University of Miami, United States

REVIEWED BY

Nahid Arghiani,
Stockholm University, Sweden
Yu-Ping Yang,
University of Miami, United States

*CORRESPONDENCE

Zhe-Jia Zhang,
✉ zhangzhejia@csu.edu.cn
Jun-Pu Wang,
✉ wang-jp2013@csu.edu.cn

RECEIVED 10 May 2023

ACCEPTED 27 July 2023

PUBLISHED 04 August 2023

CITATION

Duan S-L, Fu W-J, Jiang Y-K, Peng L-S,
Ousmane D, Zhang Z-J and Wang J-P
(2023), Emerging role of exosome-
derived non-coding RNAs in tumor-
associated angiogenesis of
tumor microenvironment.
Front. Mol. Biosci. 10:1220193.
doi: 10.3389/fmolb.2023.1220193

COPYRIGHT

© 2023 Duan, Fu, Jiang, Peng, Ousmane,
Zhang and Wang. This is an open-access
article distributed under the terms of the
[Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or
reproduction in other forums is
permitted, provided the original author(s)
and the copyright owner(s) are credited
and that the original publication in this
journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Emerging role of exosome-derived non-coding RNAs in tumor-associated angiogenesis of tumor microenvironment

Sai-Li Duan^{1,2}, Wei-Jie Fu², Ying-Ke Jiang¹, Lu-Shan Peng³,
Diabate Ousmane^{2,3}, Zhe-Jia Zhang^{1,2*} and Jun-Pu Wang^{2,3,4,5*}

¹Department of General Surgery, Xiangya Hospital Central South University, Changsha, China, ²Xiangya School of Medicine, Central South University, Changsha, China, ³Department of Pathology, Xiang-ya Hospital, Central South University, Changsha, China, ⁴Key Laboratory of Hunan Province in Neurodegenerative Disorders, Xiangya Hospital, Central South University, Changsha, China, ⁵National Clinical Research Center for Geriatric Disorders, Xiangya Hospital, Central South University, Changsha, China

The tumor microenvironment (TME) is an intricate ecosystem that is actively involved in various stages of cancer occurrence and development. Some characteristics of tumor biological behavior, such as proliferation, migration, invasion, inhibition of apoptosis, immune escape, angiogenesis, and metabolic reprogramming, are affected by TME. Studies have shown that non-coding RNAs, especially long-chain non-coding RNAs and microRNAs in cancer-derived exosomes, facilitate intercellular communication as a mechanism for regulating angiogenesis. They stimulate tumor growth, as well as angiogenesis, metastasis, and reprogramming of the TME. Exploring the relationship between exogenous non-coding RNAs and tumor-associated endothelial cells, as well as their role in angiogenesis, clinicians will gain new insights into treatment as a result.

KEYWORDS

exosomes, endothelial cells, exosomes-derived non-coding RNAs, tumor-associated angiogenesis, tumor microenvironment, lncRNA, miRNA, cancer

1 Introduction

The tumor microenvironment (TME), an intricate ecosystem, actively participates in every stage of cancer development (Hanahan and Weinberg, 2011; Yang et al., 2020). As a dynamic ecosystem containing a variety of cell types and non-cellular components, TME plays a major role in tumor growth, metastasis, and drug resistance. Cancers exhibit some

Abbreviations: TME, Tumor microenvironment; ECs, endothelial cells; VEGF, vascular endothelial growth factor; ncRNAs, non-coding RNAs; lncRNAs, long-chain non-coding RNAs; miRNAs, microRNAs; TAM, Tumor-associated macrophage; TEXs, Tumor-derived exosomes; PDAC, pancreatic ductal adenocarcinoma; UTR, untranslated region; VASH2, Vasohibin 2; HCC, hepatocellular carcinoma; VE-Cad, VE-Cadherin; CRC, colorectal cancer; HUVEC, human umbilical vein endothelial cells; PHD1 and 2, prolyl hydroxylases 1 and 2; HIF-1 α , hypoxia-inducible factor 1 alpha; LUAD, lung adenocarcinoma; SCLC, small cell lung cancer; NSCLC, non-small cell lung cancer; HBMEC, human brain microvascular endothelial cells; ERK1/2, extracellular signal-regulated kinase 1/2; EOC, epithelial ovarian cancer; APC, Adenomatous Polyp in Colon; EOC, epithelial ovarian cancer.

biological behaviors, such as proliferation, migration, invasion, immune escape, angiogenesis, and metabolic reprogramming, all of which are affected by TME. Biological functions, including autocrine and paracrine functions, are regulated by the complex communication network within TMEs. Exocrine-mediated communication is an important emerging pathway of paracrine signal transduction (Giraldo et al., 2019). Exosomes can carry molecules such as DNA, RNA, and proteins to adjacent cells, where they act as effective signaling molecules between cancer cells and surrounding cells constituting TME. Nontumor cells in TME, such as fibroblasts, endothelial cells (ECs), and immune cells, are affected by tumor-associated active substances and their original cell functions undergo tumor-like changes, constantly adapting to the new environment and promoting tumor growth. The TME is composed of different cell types with various functions, which regulates excessive cell-cell interactions. These interactions orchestrate reprogramming to the environment allowed by each cancer and may have a significant impact on cancer development, progression, and treatment resistance.

ECs are involved in tumor growth, tumor-induced angiogenesis, and vascular secretory functions for self-renewal and differentiation after trauma and thrombosis (Barachini et al., 2023). Angiogenesis plays an important role in all stages of cancer development (Aguilar-Cazares et al., 2019). Angiogenesis is a complex process of growing new capillaries from preexisting blood vessels, typically involving the following steps: stimulation of ECs with vascular endothelial growth factor (VEGF), proliferation, migration, and differentiation of vascular ECs, vessel branches and vessel formation (Ahir et al., 2020; Yang et al., 2022). Tumor vascular growth is a key factor in cancer progression, which is closely related to metastasis and a poor prognosis. Tumor angiogenesis is a recognized target for anticancer therapy by targeting growth factors, their cell surface receptors, and associated signaling pathways. Tissue hypoxia induces an overproduction of VEGF, leading to an imbalance between pro-angiogenic factors and anti-angiogenic factors, causing excessive abnormal angiogenesis that plays a central role in tumor progression (Jászai and Schmidt, 2019). The supply of energy and the removal of waste products are key factors in the development of cancer cells

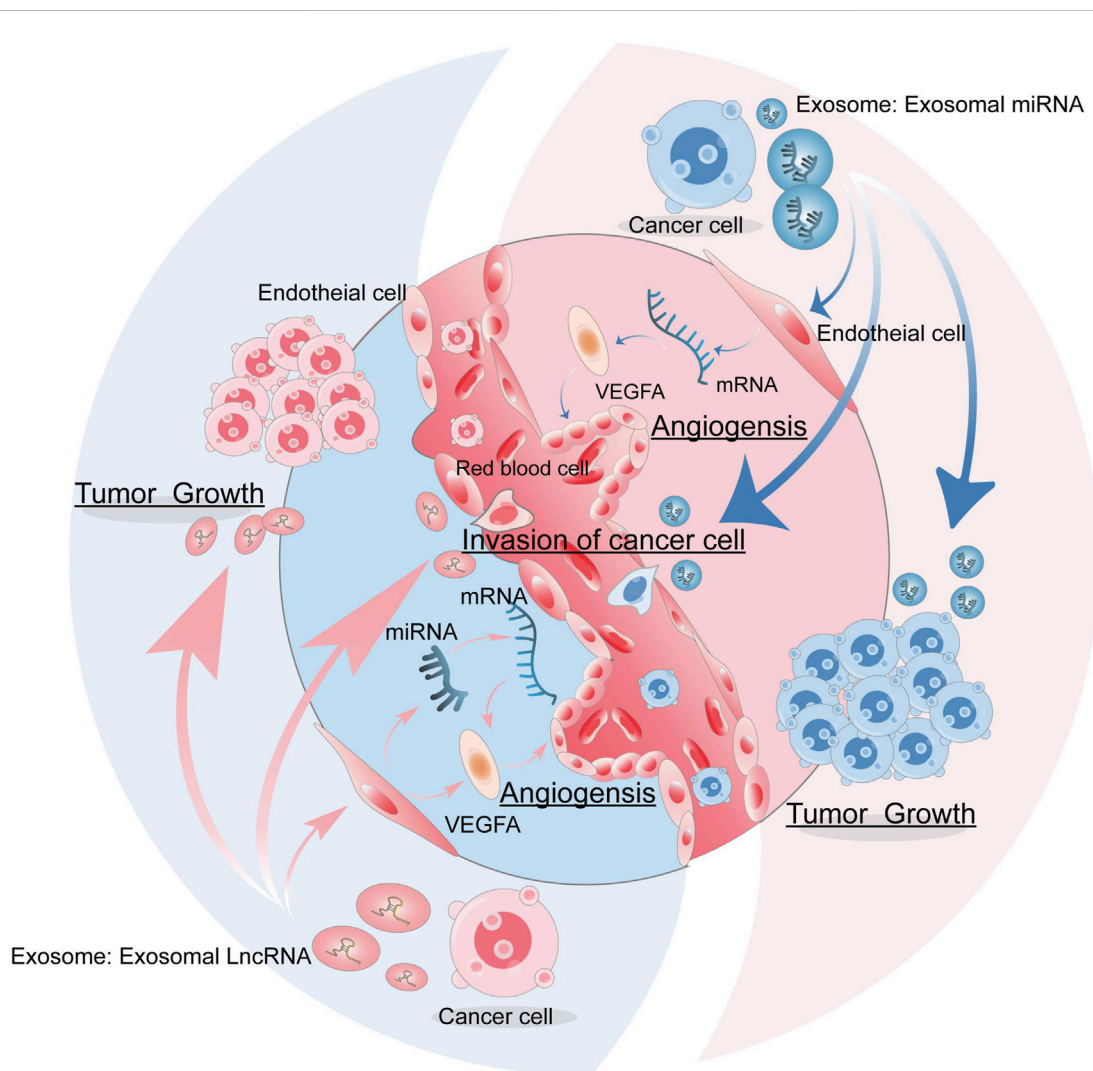


FIGURE 1

The role of exosomes in tumor angiogenesis. Tumor cells can release exosomes, which carry miRNA and lncRNAs that can act on endothelial cells to promote tumor angiogenesis, as well as stimulate tumor growth, invasion, and metastasis.

(Anderson and Simon, 2020). Tumor cells can communicate with adjacent tissues through the release of exosomes (Stec et al., 2015; Dominiak et al., 2020). Exosomes contain a variety of substances that promote angiogenesis and thus accelerate cancer invasion and metastasis (Głuszko et al., 2019), and the release of some exosomes also affects immune function (Aslan et al., 2019). Evidence suggests that non-coding RNAs (*ncRNAs*), especially long-chain non-coding RNAs (*lncRNAs*) and microRNAs (*miRNAs*) in cancer-derived exosomes, play an important role in regulating angiogenesis by facilitating intercellular communication, which in turn stimulates tumor growth, as well as angiogenesis, metastasis, and reprogramming of TME (shown in Figure 1) (Zhao et al., 2020).

In hypoxic environments, hypoxia can induce overexpression of *ncRNA*, which is released by exosomes and participates in tumor angiogenesis by reacting with ECs and other angiogenic cells, thus affecting tumor progression (He et al., 2022; Jia et al., 2022; Yang et al., 2022). In addition to ECs, there are many other remaining cell-derived exosomes in TME that can also promote angiogenesis and thus help tumor cell metastasis. For example, exosomal *ncRNAs* released by tumor cells regulate ECs and promote or inhibit angiogenesis (Ahmadi and Rezaie, 2020). Tumor-associated macrophage (TAM)-derived exosomal *ncRNAs* regulate tumor cells and promote angiogenesis (Xu et al., 2022). Stem cell-derived exosomal *ncRNAs* regulate tumor cells and inhibit tumor angiogenesis (Yang and Teng, 2023). Tumor-derived exosomes (TEXs) of lung cancer cells can transfer *miR-21* to ECs *in vitro* and stimulate ECs angiogenesis to increase VEGF expression and secretion, thus helping to invade and metastasize lung cancer cells (Forder et al., 2021). The overexpression of exosome-derived *miR-16* and *miR-100* from mesenchymal stem cells downregulates VEGF expression in breast cancer cells, thus inhibiting angiogenesis and tumor growth *in vivo* and *in vitro* (Soheilifar et al., 2022). In hepatoma cells, cancer stem cells upregulate VEGF by delivering overexpressed *lncRNAH19* to ECs to promote angiogenesis and tumor growth (Yao et al., 2023). TAM-derived exosomes are enriched with *miR-501-3p*, which enhances the metastatic capacity of pancreatic ductal adenocarcinoma (PDAC) cells (Yin et al., 2019; Cocks et al., 2022). Blood vessel formation is inseparable from the role of ECs, and the relationship between exosomal *miRNAs* and *lncRNAs* and endothelial cells in TME is the focus of this review. By summarizing their relationship to explore the role of exogenous *ncRNAs* in tumor-associated endothelial cells and also their specific role in angiogenesis, clinicians will be able to gain new insights in cancer treatment.

2 Important position of exosomes

Previous studies have shown that a series of growth factors, cell surface receptors, and a large number of signaling molecules drive remodeling of the blood and lymphatic system in cancer (Stacker et al., 2014; Fares et al., 2020; Arcucci et al., 2021a). Recent studies have identified important roles for *ncRNAs* in the regulation of key aspects of cancer biology, including tumor angiogenesis and lymphangiogenesis. *NcRNAs* are a class of RNA molecules that do not encode proteins (Zampetaki et al., 2018), of which *miRNA* is the most studied, which along with *lncRNA* is the main focus of this review. *miRNAs* are small RNA molecules that

mediate post-transcriptional regulation by targeting *mRNAs*, thereby resulting in the reduction of gene expression through *mRNA* degradation and/or translational repression. Nuclear *miRNAs* have been shown to play a role in transcriptional regulation through the recruitment of transcriptional activators and chromatin remodeling proteins of repressors (Bartel, 2009; Liu et al., 2018). It should be noted that different *miRNAs* can work together to focus on the expression of the same or multiple genes in related molecular pathways (Uhlmann et al., 2012). *LncRNA* exhibits a series of different regulatory functions in different cell compartments (Zampetaki et al., 2018). *LncRNA* plays a role in transcriptional regulation by binding chromatin remodeling proteins and recruiting transcription factors, activators, and inhibitors (Man et al., 2018).

Nuclear *miRNAs* can affect transcription by active ting or silencing of transcribed genes (Liu et al., 2018), and *miRNAs* participate in post-transcriptional processes by regulating *mRNA*. For example, *miR-29-b* regulates the expression of VEGFA and Akt3 by negatively inhibiting angiogenesis (Chen et al., 2017; Li et al., 2017). *LncRNA Hotair* can promote angiogenesis by directly activating the transcription of VEGFA genes (Fu et al., 2016). *LncRNA* can influence the cell cycle by regulating *mRNAs*. For example, *lncRNA MALAT1* can regulate the variable splicing of the carcinogenic transcription factor B-MYB in endothelial cells (Tripathi et al., 2013), *WTAPP1 lncRNA* promotes migration by increasing the expression of matrix metalloproteinase MMP1 (Li et al., 2018), and *tie-1As lncRNA* selectively binds and degrades *tie-1 mRNA*, leading to specific defects in cell connection and tube formation (Li et al., 2010). Furthermore, *lncRNA H19* regulates the biological behaviors of endothelial cells by suppressing *miR-29a*, thus inhibiting angiogenesis (Jia et al., 2016). *LncRNAs* facilitate epigenetic control of gene expression by recruiting transcription activators or inhibitors (Lam et al., 2013; Melo et al., 2013) or chromatin remodeling proteins as transcription regulators (Creamer and Lawrence, 2017). After gene transcription, *LncRNAs* can also be regulated, mainly by regulating mRNA splicing (Gong and Maquat, 2011), or by eliciting proteins that degrade *mRNAs* (Hutchinson et al., 2007) or acting as bait for proteins involved in *mRNA* degradation (Lee et al., 2016). *LncRNAs* can regulate various cancer-associated *mRNAs* by competitively sponging various *miRNAs*, and thus participate in relevant signaling pathways (Zhong et al., 2019). It is worth emphasizing that both *miRNAs* and *lncRNAs* can regulate the gene expression in complex biological responses: *miRNAs* regulate gene expression of proteins associated with their related molecular pathways by targeting *mRNAs*, and in addition, *miRNAs* can collaborate with other molecules to precisely mediate gene silencing. *LncRNAs* regulate gene expression by controlling chromatin remodeling, or by targeting *miRNAs* regulate gene expression by controlling chromatin remodeling or by targeting *miRNAs* (Guo et al., 2020; Mao et al., 2020).

2.1 The relationship between exosomal miRNAs and endothelial cells

Endothelial cells can form vascular systems to transport nutrients and metabolites, which can help tumor proliferation, invasion, and metastasis. Crosstalk stimulation between tumor

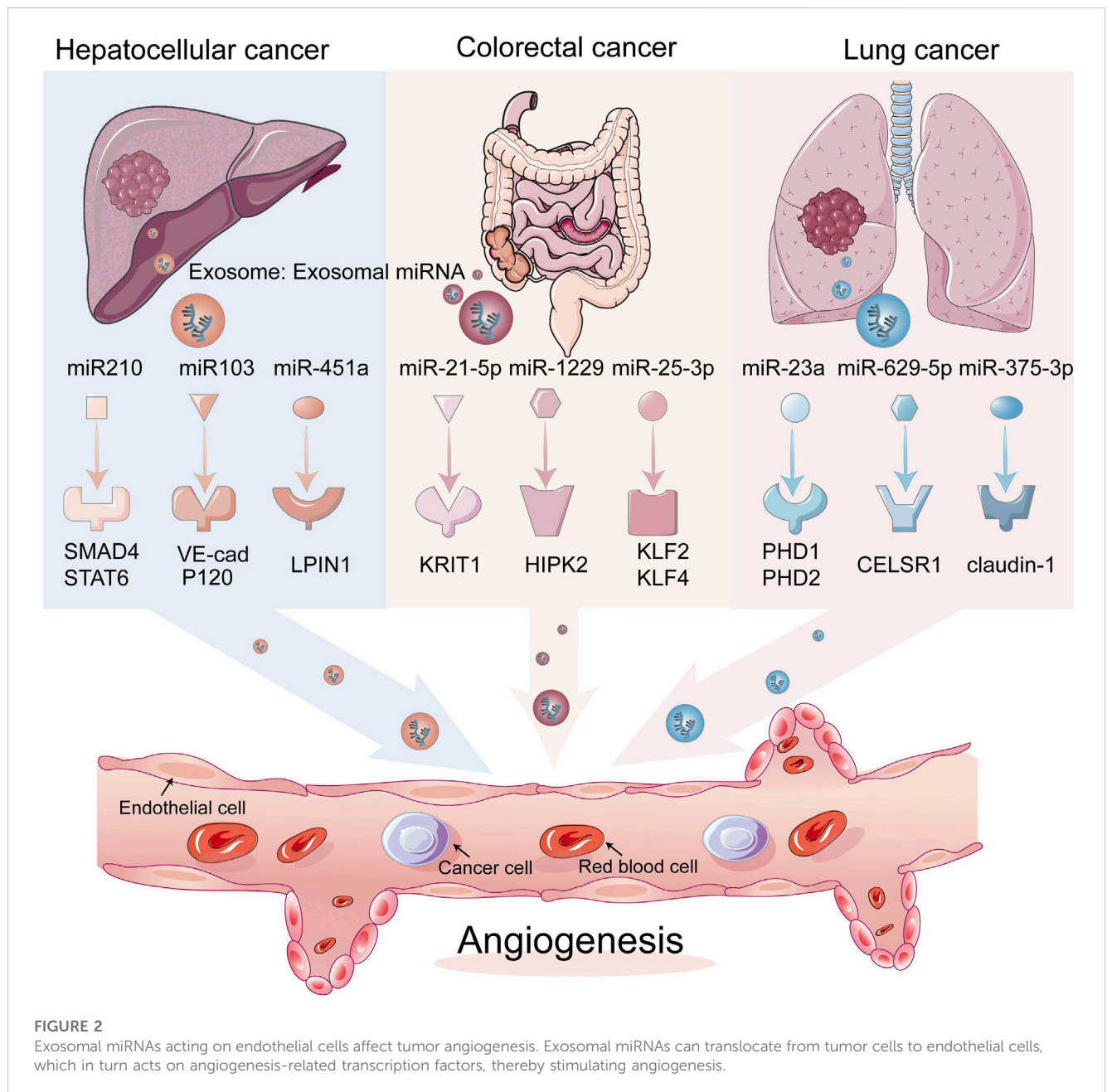
TABLE 1 Relationship between exosomal miRNAs and angiogenesis in different types of cancer.

Types of cancers	MiRNAs in exosome	Roles of miRNAs in angiogenesis	Receptor cells	References
Hepatocellular carcinoma (HCC)	<i>miR-103</i>	Inhibiting the expression of VE cadherin	Endothelial cells	Fang et al. (2018)
	<i>miR-210</i>	Targeting Smad4 and STAT6	Endothelial cells	Lin et al. (2018a)
	<i>miR-1290</i>	Targeting SMEK1	Endothelial cells	Wang et al. (2021a)
	<i>miR-451a</i>	Targeting LPIN1	Endothelial cells	Zhao et al. (2019)
	<i>miR-638</i>	Down-regulating the expression of VE cadherin and ZO-1	Endothelial cells	Yokota et al. (2021)
	<i>miR-200b-3p</i>	Enhancing the expression of endothelial ERG	Endothelial cells	Moh-Moh-Aung et al. (2020)
	<i>miR-378b</i>	Directly promoting	Endothelial cells	Shi et al. (2021)
Lung cancer (LC)	<i>miR-296</i>	Being responsible for lymphangiogenesis	Endothelial cells	Shi et al. (2019)
	<i>miR-23a</i>	Inhibiting its targets PHD1 and 2	Endothelial cells	Hsu et al. (2017)
	<i>miR-629-5p</i>	Inhibiting CELSR1	Endothelial cells	Li et al. (2020b)
	<i>miR-30a-5p</i>	Inhibiting cell proliferation, migration and invasion abilities	Lung adenocarcinoma (LUAD) cells	Tao et al. (2021)
	<i>miR-141</i>	Targeting KLF12	Endothelial cells	Mao et al. (2020)
	<i>miR-375-3p</i>	Binding the 3'UTR of the tight junction protein claudin-1	Endothelial cells	Mao et al. (2021)
	<i>miR-486-5p</i>	Targeting the CADM1/tight junction axis	Endothelial cells	Sun et al. (2021)
Glioma (GBMLGG)	<i>miR-148a-3p</i>	Inhibiting ERRF1 and activating EGFR/MAPK signaling pathway	Endothelial cells	Wang et al. (2020a)
	<i>miR-26a</i>	Targeting PTEN	Endothelial cells	Wang et al. (2019)
	<i>miR-21</i>	Via miR-21/VEGF signaling pathway	Endothelial cells	Mezzadra et al. (2017)
	<i>miR-944</i>	Inhibiting AKT/ERK signaling	Endothelial cells	Jiang et al. (2021)
	<i>miR-182-5p</i>	Inhibiting Kruppel-like factors 2 and 4	Endothelial cells	Li et al. (2020a)
Colorectal Cancer (CRC)	<i>miR-27b-3p</i>	Transferring to human umbilical vein endothelial cells	Endothelial cells	Dou et al. (2021)
	<i>miR-21-5p</i>	Transferring to human umbilical vein endothelial cells	Endothelial cells	He et al. (2021a)
	<i>miR-25-3p</i>	Targeting KLF2 and KLF4	Endothelial cells	Zeng et al. (2018)
	<i>miR-1229</i>	Targeting HIPK2	Endothelial cells	Hu et al. (2019)
Oral Squamous Cell Carcinoma (OSCC)	<i>miR-210-3p</i>	Targeting EFNA3 via PI3K/AKT pathway regulation	Endothelial cells	Wang et al. (2020b)
	<i>miR-130b-3p</i>	Inhibiting human umbilical vein endothelial cells	Endothelial cells	Yan et al. (2021)
	<i>miR-221-3p</i>	Targeting PIK3R1	Endothelial cells	He et al. (2021b)
cervical squamous cell carcinoma (CESC)	<i>miR-221-3p</i>	Targeting THBS2	Endothelial cells	Zhou et al. (2019)
	<i>miR-663b</i>	Inhibiting vinculin	Endothelial cells	You et al. (2021)
	<i>miR-142-5p</i>	Inducing IDO expression via ARID2-DNMT1-IFN- γ signaling	Lymphatic endothelial cells (LECs)	Zhou et al. (2021)
	<i>miR-221-3p</i>	Targeting VASH1	Endothelial cells	Wu et al. (2019)
	<i>miR-9</i>	Targeting MDK and modulating the PDK/AKT pathway	Endothelial cells	Lu et al. (2018)

(Continued on following page)

TABLE 1 (Continued) Relationship between exosomal miRNAs and angiogenesis in different types of cancer.

Types of cancers	MiRNAs in exosome	Roles of miRNAs in angiogenesis	Receptor cells	References
	<i>miR-23a</i>	Inhibiting TSGA10	Endothelial cells	Bao et al. (2018)
Ovarian Cancer (OV)	<i>miR-205</i>	Via PTEN-AKT pathway	Endothelial cells	He et al. (2019)
	<i>miR-141-3p</i>	Activating JAK/STAT3 and NF-κB signaling pathways	Endothelial cells	Masoumi-Dehghi et al. (2020)
Pancreatic Cancer (PAAD)	<i>miR-27a</i>	Via BTG2	Endothelial cells	Shang et al. (2020)
Renal clear cell carcinoma (RCCC)	<i>miR-185-5p</i>	Binding to the promoter region of HIF2A mRNA	Endothelial cells	Braga et al. (2019)



cells and endothelial cells can promote the growth of both, improve tumor malignancy, and even develop resistance to treatment (Shweiki et al., 1992; Carmeliet and Jain, 2011). Tumor cells and certain immune cell subsets can promote angiogenesis by expressing and secreting growth factors or inducing hypoxia (Ding et al., 2014; Zhou et al., 2014), resulting in leakage of vascular structures that promote angiogenesis and metastatic spread of tumor cells. *MiRNAs* are endogenous *ncRNAs* consisting of 21–25 nucleotides that promote post-transcriptional regulation of target genes mainly by binding to the 3′ untranslated region (UTR) of *mRNAs*. Meanwhile, *miRNAs* regulate more than 30% of gene expression in the body, and their functions are closely related to cell proliferation, differentiation, apoptosis, embryonic development, tissue and organ formation, as well as the occurrence and development of various diseases (Bartel, 2004). Recent studies have shown that exosome-mediated *miRNAs* transfer from cancer cells to endothelial cells, contributing to the breakdown of the endothelial cell barrier and allowing cancer cells to spread and metastasize to distant locations, such as cell-derived exosomal *miR-27b-3p* in colorectal cancer (Zhou et al., 2014; Dou et al., 2021). Furthermore, *miRNA*-containing exosomes from leukemia cells, such as *miR-17-92*, play an important role in communication between tumor and endothelial cells, thus regulating the process of tumor angiogenesis (Umezumi et al., 2013).

Exosomal *miRNAs* can regulate the migration of tumor endothelial cells and the formation of lymphatic and blood vessels (Table 1; Figure 2). Within tumors, most exosomal *miRNAs* are thought to be produced by tumor cells (Huang et al., 2022). When internalized by endothelial cells, some of these *miRNAs* can stimulate angiogenesis or lymphangiogenesis by inhibiting the expression of proteins that inhibit the main pathways driving these processes (Duan et al., 2019; Kim et al., 2020; Masoumi-Dehghi et al., 2020). Exosomal *miRNAs* have been shown to downregulate several anti-angiogenic transcription factors in endothelial cells or inhibit the expression of VEGFA, a key inducer of angiogenesis, thus turning on the angiogenic switch (Li J. et al., 2020). For example, in gastric cancer, exosomal *miR-130a* and *miR-155* secreted by gastric cancer cells can inhibit the expression of the transcription factor c-MYB, indirectly promoting the expression of VEGFA (Arcucci et al., 2021b), which promotes angiogenesis and further assists in invasion and metastasis of gastric cancer cells.

2.1.1 Hepatocellular carcinoma cells

Current studies have shown that in hepatocellular carcinoma (HCC), exosomal *miR-210* secreted by HCC cells can be transferred to endothelial cells, thus promoting tumor angiogenesis by targeting SMAD4 and STAT6 (Lin X. J. et al., 2018). *miR-1290* targeting SMEK1 promotes angiogenesis of hepatocellular carcinoma, and *miR-451a* targeting LPIN1 suppresses hepatocellular tumorigenesis by regulating tumor cell apoptosis and angiogenesis (Zhao et al., 2019; Wang Q. et al., 2021). HANR is responsible for lymphangiogenesis in HCC cells via the exosomal *miR-296* and the EAG1/VEGF axis (Shi et al., 2019). *miR-103* was delivered to ECs through exosomes and then attenuated the integrity of the endothelial junction by directly inhibiting the expression of VE-Cadherin (VE-Cad) (Fang et al., 2018). *miR-638* can promote vascular permeability by downregulating endothelial expression

of VE-Cad and ZO-1 (Yokota et al., 2021). Exosomal *miR-200b-3p* from hepatocytes inhibited endothelial ERG expression, while reduction of *miR-200b-3p* in cancer cells promoted angiogenesis in HCC tissues by improving endothelial ERG expression (Moh-Moh-Aung et al., 2020).

2.1.2 Colorectal cancer cells

The exosome *miR-21-5p* can be delivered from colon cancer cells to endothelial cells, targeting KRIT1 and thus inducing angiogenesis and vascular permeability, as can the exosome *miR-25-3p*, which also transfers to ECs and promotes CRC metastasis by targeting KLF2 and KLF4 to regulate growth factors in endothelial cells. Furthermore, there is *miR-1229* that promotes angiogenesis by targeting HIPK2 (Zeng et al., 2018; Hu et al., 2019; He Q. et al., 2021). *miR-27b-3p* is transferred by EMT-CRC cells into the exosomes of human umbilical vein endothelial cells (HUVEC), weakening the vascular barrier (Dou et al., 2021).

2.1.3 Lung cancer cells

miR-23a directly inhibits its targets, prolyl hydroxylases 1 and 2 (PHD1 and 2), in exosomes from lung cancer cells, resulting in the accumulation of hypoxia-inducible factor 1 alpha (HIF-1α) in endothelial cells. Finally, hypoxic lung cancer cells enhanced angiogenesis through hypoxic cancer-derived exosomes under normoxic and hypoxic conditions (Hsu et al., 2017). For lung adenocarcinoma (LUAD), *miRNAs* affect cancer cells and ECs bidirectionally; for example, *miR-629-5p* in lung adenocarcinoma transfers to endothelial cells, and by inhibiting CELSR1, which is lower in endothelial cells in invasive LUAD (a *miR-30a-5p*, a non-canonical cadherin, increases endothelial monolayer permeability, while overexpression of *miR-30a-5p* in endothelial cells inhibited tumor development (Li et al., 2020b; Tao et al., 2021). The exosome *miR-141* is transported into HUVEC cells and targets KLF12 to promote angiogenesis in small cell lung cancer (SCLC), and *miR-375-3p* destroys vascular endothelial cells by directly binding to the 3′UTR of the tight junction protein CLDN1 and negatively regulating its expression tight junctions (Mao et al., 2020; Mao et al., 2021). *miR-486-5p* in non-small cell lung cancer (NSCLC) targets the CADM1/tight junction axis in vascular endothelial cells to promote metastasis of non-small cell lung cancer cells (Sun et al., 2021).

2.2 The relationship between exosomal lncRNAs and endothelial cells

LncRNAs are a diverse class of transcribed RNA molecules that are more than 200 nucleotides long and have limited protein coding potential (Nagano and Fraser, 2011; Spizzo et al., 2012). Current estimates from the GENCODE database (www.genecodegenes.org) suggest that the human genome contains approximately 16,000 *lncRNA* genes encoding over 28,000 distinct *lncRNAs*. Many *lncRNAs* have emerged as key players in the regulation of numerous biological processes in cancer, such as differentiation, cell cycle regulation, and immune responses (Guttman et al., 2009; Qiu et al., 2015; Bach and Lee, 2018). They can act directly as tumor suppressors or oncogenes, or be regulated by well-known tumor suppressors or oncogenes at the transcriptional or post-

TABLE 2 Ways of exosomal lncRNAs to promote angiogenesis in different types of cancer.

Types of cancers	lncRNAs in exosome	Ways of lncRNAs to promote angiogenesis	Receptor cells	References
Hepatocellular carcinoma (HCC)	<i>lncRNA UBE2CP3</i>	Activating the ERK/HIF-1 α /p70S6K signaling cascade	Endothelial cells	Lin et al. (2018b)
	<i>lncRNA H19</i>	Affecting its tumor microenvironment	Endothelial cells	Conigliaro et al. (2015)
	<i>MALAT1</i>	Activating ERK1/2 signaling	Endothelial cells	Malakoti et al. (2021)
	<i>SNHG16</i>	Sponging miR-4500	Endothelial cells	Li et al. (2021)
	<i>lncRNA HULC</i>	Via VEGF and ESM-1	Endothelial cells	Zhu et al. (2016)
	<i>lncRNA-OR3A4</i>	Via AGGF1/akt/mTOR	Endothelial cells	Li et al. (2019)
Hepatoblastomas (HBs)	<i>lncRNA CRNDE</i>	Modulating mTOR signaling	Endothelial cells	Dong et al. (2017)
Gastric cancer (GC)	<i>lncRNA PVT1</i>	Inducing the STAT3/VEGFA axis	Endothelial cells	Zhao et al. (2018)
	<i>X26nt</i>	Binding to the 3'UTR of VE-cadherin mRNA	Endothelial cells	Chen et al. (2021)
	<i>LINC01410</i>	Depleting miR-532-5p	Endothelial cells	Zhang et al. (2018)
Non-Small Cell Lung Cancer (NSCLC)	<i>TNK2-AS1</i>	Enhancing STAT3 signaling through increasing VEGFA expression	Endothelial cells	Wang et al. (2018a)
	<i>lncRNA-p21</i>	Promoting tube formation and enhancing adhesion of tumor cells	Endothelial cells	Castellano et al. (2020)
	<i>lncRNA LINC01356</i>	Remodeling the blood-brain barrier	Endothelial cells	Geng et al. (2022)
	<i>lnc-MMP2-2</i>	Targeting the miRNA-1207-5p/EPB41L5 axis	Endothelial cells	Wu et al. (2021)
Pancreatic Cancer (PAAD)	<i>lncRNA UCA1</i>	Through the miR-96-5p/AMOTL2/ERK1/2 axis	Endothelial cells	Guo et al. (2020)
	<i>CCAT1</i>	Binding to miR-138-5p to increase HMGA1 expression	Endothelial cells	Han et al. (2021)
Glioma (GBMLGG)	<i>lnc-POU3F3</i>	Secreting linc-POU3F3-enriched exosomes	Endothelial cells	Lang et al. (2017a)
	<i>lnc-CCAT2</i>	Activating VEGFA and TGF β	Endothelial cells	Lang et al. (2017b)
Osteosarcoma (OS)	<i>EWSAT1</i>	Increasing secretion of angiogenic factors	Endothelial cells	Qiu et al. (2018)
	<i>lncRNA RAMP2-AS1</i>	Acting as a ceRNA of miR-2355-5p and regulating the expression of VEGFR2	Endothelial cells	Cheng et al. (2020)
	<i>MALAT1</i>	Blocking the pro-angiogenic effects potentially	Endothelial cells	Zhang et al. (2017b)
Breast cancer (BC)	<i>lncRNA AC073352.1</i>	Binding to YBX1	Endothelial cells	Kong et al. (2021)
	<i>MEG3</i>	Inactivating AKT signaling	Endothelial cells	Zhang et al. (2017c)
	<i>lncRNA GSI-600G8.5</i>	Reducing TEER and increasing BBB permeability	Endothelial cells	Lu et al. (2020)
Colorectal Cancer (CRC)	<i>lncRNA-APC1</i>	Activating the MAPK pathway	Endothelial cells	Wang et al. (2021b)
	<i>lncRNA PCAT1</i>	Regulating the activity of the miR-329-3p/Netrin-1-CD146 complex	Endothelial cells	Fang et al. (2022)
salivary adenoid cystic carcinoma (SACC)	<i>MRPL23-AS1</i>	Forming an RNA-protein complex with EZH2	Endothelial cells	Chen et al. (2020)
Nasopharyngeal carcinoma (NPC)	<i>CCAT2</i>	Via nasopharyngeal carcinoma-derived exosomal lncRNA CCAT2	Endothelial cells	Zhou et al. (2020)
Lung Adenocarcinoma (LAD)	<i>lncRNA LOC100132354</i>	Activating the VEGFA/VEGFR2/RAF/MEK/ERK signaling pathway	Endothelial cells	Wang et al. (2018b)
Epithelial ovarian cancer (EOC)	<i>MALAT1</i>	Transferring to recipient HUVECs and affecting HUVECs	Endothelial cells	Qiu et al. (2018)
Bladder cancer (BCa)	<i>lncRNA BCYRN1</i>	Enhancing VEGF-C/VEGFR3 signaling-induced BCa lymphatic metastasis	Endothelial cells	Lei and Mou (2020)
Cervical Cancer (CC)	<i>TUG1</i>	Being transferred to the recipient HUVEC	Endothelial cells	Tao et al. (2020)
Thyroid Cancer (TC)	<i>FGD5-AS1</i>	Targeting the miR-6838-5p/VAV2 axis	Endothelial cells	Liu et al. (2022)

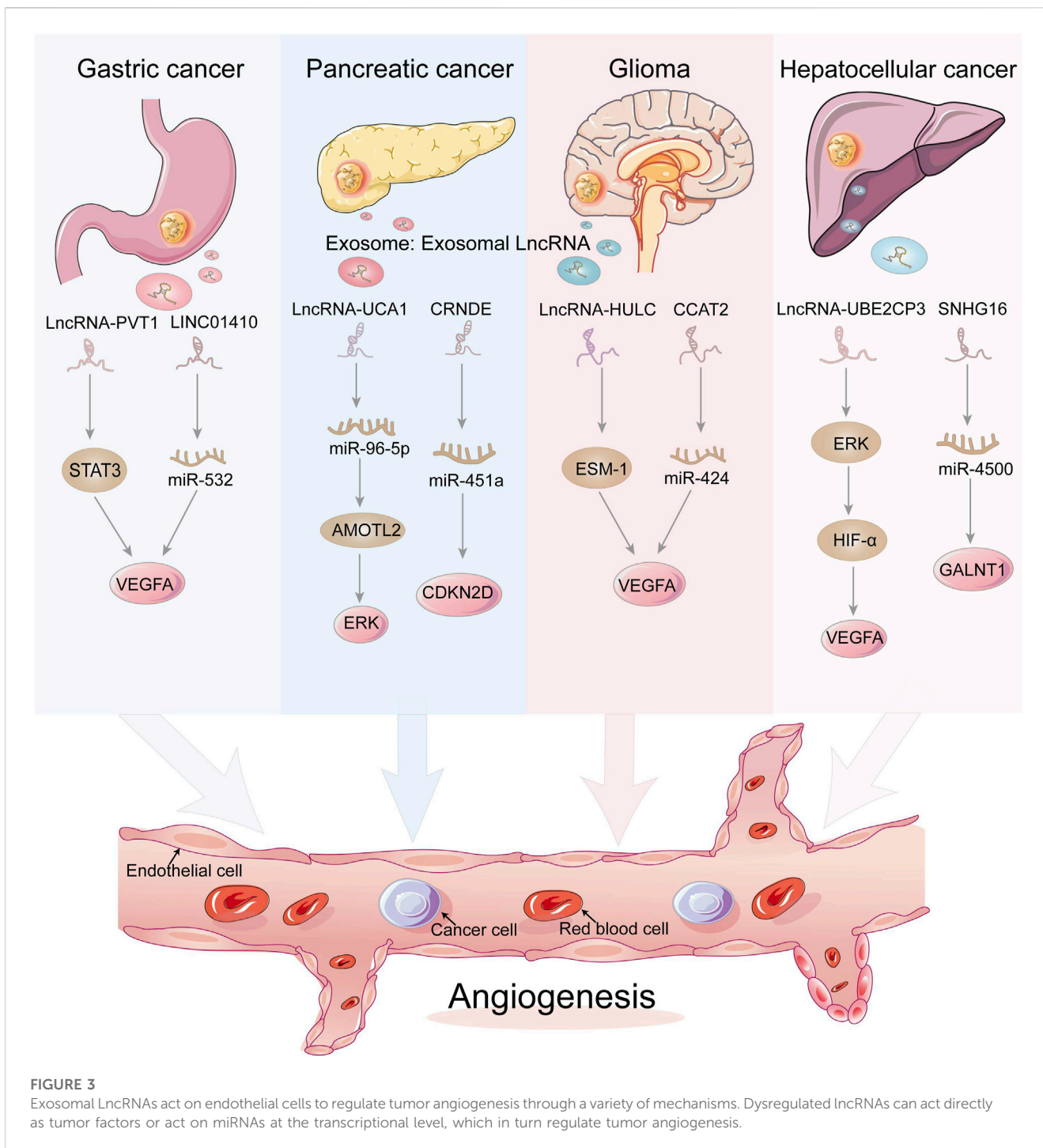


FIGURE 3 Exosomal lncRNAs act on endothelial cells to regulate tumor angiogenesis through a variety of mechanisms. Dysregulated lncRNAs can act directly as tumor factors or act on miRNAs at the transcriptional level, which in turn regulate tumor angiogenesis.

transcriptional level (Barsyte-Lovejoy et al., 2006; Huarte et al., 2010). ECs that line the inner surface of the blood vessels are an important part of the matrix in the TME (Junttila and de Sauvage, 2013; Kohlhapp et al., 2015). They are believed to be critical for angiogenesis and tumor metastasis, and lncRNAs may affect tumor progression by regulating endothelial cell biological behavior (Table 2; Figure 3). For example, lncRNA H19 has been reported to be significantly upregulated in glioma-associated endothelial cells cultured in glioma-conditioned medium. Knockdown of lncRNA H19 inhibited glioma-induced endothelial cell proliferation, migration, and tube formation *in vitro*. Mechanistic evidence

suggests that lncRNA H19 regulates the biological behavior of glioma-associated endothelial cells by inhibiting miR-29a (Jia et al., 2016). Furthermore, lncRNA-APC1 plays an important tumor suppressor role in the pathogenesis of colorectal cancer. The following mechanistic studies show that lncRNA-APC1 reduces exosome production in colorectal cancer cells by reducing Rab5b mRNA stability, and this effect inhibits tumor angiogenesis by inhibiting the over-activation of the MAPK pathway in endothelial cells (Wang F. W. et al., 2021). Dysregulated lncRNAs affect endothelial cell biological behavior through multiple mechanisms, so regulation of specific lncRNA

expression in tumor cells or/and endothelial cells may have a significant impact on cancer progression.

2.2.1 Gastric cancer cells

PVT1 is an oncogenic *lncRNA* that is significantly expressed in gastric cancer, especially in patients with low differentiation and progressive stages. *PVT1* can bind to different proteins to exert oncogenic effects, and in gastric cancer, *PVT1* can bind to the signal transduction activator STAT3 to ensure that it is not degraded, thus activating the STAT3 signaling pathway and thus increasing VEGFA in gastric cancer, thus activating the STAT3 signaling pathway and increasing the expression of VEGFA to promote gastric cancer angiogenesis. At the same time, activated STAT3 can also occupy the promoter of *PVT1* and promote *PVT1* expression, forming a positive feedback regulation (Zhao et al., 2018). Similarly, in NSCLC, the *lncRNA TNK2-AS1* can also bind to STAT3 to inhibit its degradation, thus activating the STAT3 signaling pathway and promoting tumor progression and angiogenesis. In addition, STAT3 can also bind to the *lncRNA TNK2-AS1* promoter to promote its transcription in positive feedback (Wang et al., 2018a). *LINC01410* is also one of the molecules that promote angiogenesis in gastric cancer. *LINC01410* can inhibit *miR-532-5p* expression, while silencing *miR-532-5p* reduces inhibition of NCF2, thus upregulating NCF2 expression and activating the NF- κ B signaling pathway, exacerbating malignant progression and angiogenesis of gastric cancer. Interestingly, NCF2 can bind to the *LINC01410* promoter, thereby promoting its transcription, forming a positive feedback loop that exacerbates the development of gastric carcinogenesis (Zhang et al., 2018).

2.2.2 Pancreatic cancer cells

An important feature of the tumor microenvironment is hypoxia caused by inadequate oxygen flow and abnormal tumor vasculature, and exposure of cancer cells to conditions of oxygen deficiency increases the release of exosomes, which in turn promotes angiogenesis and tumor metastasis. In hypoxic PC cells, the expression of *lncRNA UCA1* increases and can be transferred to human microvascular endothelial cells HUVECs, promoting angiogenesis and tumor growth via the miR-96-5p/AMOTL2/ERK1/2 axis (Guo et al., 2020). In addition to this, PC cell-derived exosomal *CRNDE* enhanced angiogenesis by binding to *miR-451a* to increase *CDKN2D* expression (Zhu et al., 2021).

2.2.3 Glioma cells

One of the keys to glioma development is abnormal generation of tumor blood vessels, and high-grade gliomas clearly have a higher density of tumor blood vessels that contribute more to tumor development than low-grade gliomas. It has been shown that glioma cells can regulate the tumor microenvironment by secreting exosomes, for example, glioma exosomes can promote angiogenesis by transferring *LINC-POU3F3* to human brain microvascular endothelial cells (HBMEC) (Lang et al., 2017a). Additionally, *LINC-CCAT2* was found to be highly expressed in glioma cells U87-MG and could be transferred to HUVECs to activate the production of the angiogenic factors VEGFA and TGF β , while inhibiting the expression of the apoptotic molecules Bax and caspase-3, thus promoting angiogenesis and inhibiting apoptosis in glioma cells (Lang et al., 2017b). *lncRNA HULC* is

one of the most common oncogenes with the potential to promote invasion and angiogenesis. In glioma, Zhu Yu et al. showed that *HULC* can activate the PI3K/AKT/mTOR signaling pathway, which in turn regulates downstream angiogenic factors VEGF and ESM-1. Furthermore, in a hypoxic environment, *HULC* can upregulate HIF-1 α , which is also one of the key molecules that promote the secretion of angiogenic factors (Zhu et al., 2016).

2.2.4 Hepatocellular carcinoma cells

As tumor growth requires more and more nutrients, this requires the secretion of angiogenic substances to promote tumor angiogenesis. *lncRNA* has been shown to regulate ECs function and promote the expression of angiogenic factors to regulate angiogenesis. Lin et al. demonstrated that the *lncRNA UBE2CP3* can activate the ERK/HIF-1 α /p70S6K signaling pathway, increase VEGFA expression and regulate ECs function, thus promoting angiogenesis in hepatocellular carcinoma (Lin J. et al., 2018). Cancer stem-like cells, also known as CD90⁺ hepatocellular carcinoma cells, are enriched in *lncRNA H19*, which can be released by encapsulating in exosomes and then transported to endothelial cells, promoting the expression of the angiogenic factor VEGF in endothelial cells and thus regulating hepatocellular carcinoma angiogenesis (Conigliaro et al., 2015). Direct exosomal transfer of *MALAT1* to hepatocytes leads to increased invasion and migration of hepatocytes through activation of extracellular signal-regulated kinase 1/2 (ERK1/2) signaling (Li et al., 2020c). Exosomal *SNHG16* increases *GALNT1* expression by sponging *miR-4500* to promote angiogenesis. The *SNHG16/miR-4500/GALNT1* axis plays an important role in exosome-mediated angiogenesis and tumor growth *in vitro* and *in vivo* (Li et al., 2021). Furthermore, elevated expression of *lncRNA-OR3A4* in hepatocellular carcinoma is associated with angiogenesis and promotes the tube formation capacity of HUVEC, mainly through activation of the AGGF1/AKT/mTOR pathway (Li et al., 2019). *CRNDE* is upregulated in many tumors, promotes cell growth and migration, and is a recognized oncogene, also in hepatoblastoma. *CRNDE* knockdown inhibits tumor angiogenesis and reduces cell viability in hepatoblastoma, primarily through regulation of mTOR signaling (Dong et al., 2017).

2.2.5 Other cancer cells

Some other cancer exosomal *lncRNAs* are still associated with endothelial cells (Table 2). Osteosarcoma originates from bone and is the most common of primary malignancies. Zhang et al. showed that *lncRNA MALAT1* is associated with osteosarcoma angiogenesis and hypoxic response and that *MALAT1* activates the mTOR/HIF-1 α pathway, thereby promoting the production of angiogenic factors (Zhang Z. C. et al., 2017). In lung adenocarcinoma, the *lncRNA LOC100132354* can affect the downstream target gene VEGFA to promote tumor angiogenesis (Wang et al., 2018b). Some non-angiogenic *lncRNAs* have the ability to inhibit angiogenesis. For example, *GAS5* can inhibit the activation of the Wnt/ β -catenin pathway to suppress angiogenesis in CRC (Song et al., 2019). Regarding MEG3, a recognized tumor suppressor, it inhibits tumor progression in breast cancer mainly by suppressing AKT signaling and also inhibits capillary angiogenesis in endothelial cells by reducing the expression of tumor angiogenic factors (Lu et al., 2020). The *lncRNA MALAT1* can be transported through exosomes

to endothelial cells in epithelial ovarian cancer (EOC) and then regulates the vasculature of endothelial cells by generating related genes that stimulate pro-angiogenic behavior. In addition, serum exosomal *MALAT1* levels were strongly associated with advanced and metastatic outcomes, which were independent predictors of overall survival in EOC (Qiu et al., 2018). Interestingly, *lncRNAs* can affect exosome production in addition to being transported by exosomes. In colorectal cancer, activation of the Adenomatous Polyp in Colon (APC) gene of *lncRNA* (*lncRNA APC1*) can directly affect the stability of *Rab5b mRNA*, thereby inhibiting exosome production by CRC cells and ultimately tumor angiogenesis (Wang F. W. et al., 2021). Moreover, exosomal *lncRNA PDAT1* regulates the activity of the *miR-329-3p/Netrin-1-CD146* complex to promote tumor metastasis (Fang et al., 2022). In lung cancer, the exosomal *lncRNA LINC01356* and the exosomal *lnc-MMP2-2* derived from NSCLC cells play a key role in the remodeling of the blood-brain barrier, thereby participating in brain metastasis (Geng et al., 2022). Exosomal *lnc-MMP2-2* promotes brain metastasis via the *miRNA-1207-5p/EPB41L5* axis (Wu et al., 2021). In thyroid cancer, exosome *FGD5-AS1* targets the *miR-6838-5p/VAV2* axis to promote angiogenesis and metastasis (Liu et al., 2022).

3 Conclusion and prospect on endothelial cells and exosomes

Exosomes are important carriers of cell-to-cell communication signals and genetic material in the tumor microenvironment. In this review, we divide them into different types of cancer and summarize the relationship between *miRNAs* and *lncRNAs* with endothelial cells, promoting tumor angiogenesis and tumor angiogenesis. Mechanisms of lymphangiogenesis, demonstrating the complexity of their mediated angiogenesis in cancer development. Although *ncRNAs* do not encode proteins, they do play critical roles in regulating the levels of many cellular and extracellular proteins, particularly in the early stages of certain tumors, by mediating gene silencing at the transcriptional level to regulate the expression of cancer-related proteins, which in turn affects aspects of angiogenesis, apoptosis, and tumor metastasis. *NcRNAs* can be used as a new class of markers for early clinical diagnosis and prognosis, and exosomes can be used as carriers to deliver them to various parts of the body, helping them participate more actively in intercellular communication and function. Cancer-derived exosomal *ncRNAs* can promote tumor angiogenesis and lymphangiogenesis by altering gene expression in a variety of cell types, including endothelial cells. Therefore, the regulatory functions of *ncRNAs* in tumor angiogenesis and lymphangiogenesis can be considered multidimensional.

The mechanistic summary in this paper can help develop effective and precise cancer therapies and, based on current research related to the regulation of tumor angiogenesis by *ncRNAs*, can be used to develop new cancer biomarkers and therapies depending on the type of cancer. Identifying the different mechanisms involved in identifying therapeutic

approaches has seminal implications for new cancer treatments, and more research is needed to achieve this. In addition, certain specific *ncRNAs* can be used as a new class of markers for early clinical diagnosis and prognosis, also providing a new idea for tumor treatment. *lncRNAs* and *miRNAs* may be a feasible strategy to monitor the efficacy of anti-angiogenic therapy and predict prognosis. In addition, regulation of angiogenesis-related signaling pathways may also serve as a new therapeutic direction, and the molecular mechanisms of *miRNAs* and *lncRNAs* in tumor development and development need to be investigated in more depth, thus contributing to the improvement of tumor diagnosis and treatment.

Author contributions

S-LD and W-JF contributed to the direction and guidance of this review; S-LD and Y-KJ collected formal resources, wrote the original draft and prepared the figures; L-SP, DO, Z-JZ and J-PW provided critical revisions and contributed to the editing of the paper. All authors contributed to the article and approved the submitted version.

Funding

This work was partially supported by the National Natural Science Foundation of China (project NO. 81602167), the Hunan Provincial Natural Science Foundation of China (project NO. 2017JJ3494 and 2021JJ31100), and the Science and Technology Program Foundation of Changsha City (project NO. kq2004085).

Acknowledgments

We thank all authors to collect data and make improvement of this manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Aguilar-Cazares, D., Chavez-Dominguez, R., Carlos-Reyes, A., Lopez-Camarillo, C., Hernandez de la Cruz, O. N., and Lopez-Gonzalez, J. S. (2019). Contribution of angiogenesis to inflammation and cancer. *Front. Oncol.* 9, 1399. doi:10.3389/fonc.2019.01399
- Ahir, B. K., Engelhard, H. H., and Lakka, S. S. (2020). Tumor development and angiogenesis in adult brain tumor: Glioblastoma. *Mol. Neurobiol.* 57, 2461–2478. doi:10.1007/s12035-020-01892-8
- Ahmadi, M., and Rezaie, J. (2020). Tumor cells derived-exosomes as angiogenic agents: Possible therapeutic implications. *J. Transl. Med.* 18, 249. doi:10.1186/s12967-020-02426-5
- Anderson, N. M., and Simon, M. C. J. C. B. (2020). The tumor microenvironment. *tumor Microenviron.* 30, R921–R925. doi:10.1016/j.cub.2020.06.081
- Arcucci, V., Stacker, S. A., and Achen, M. G. (2021b). Control of gene expression by exosome-derived non-coding RNAs in cancer angiogenesis and lymphangiogenesis. *Biomolecules* 11, 249. doi:10.3390/biom11020249
- Arcucci, V., Stacker, S. A., and Achen, M. G. J. B. (2021a). Control of gene expression by exosome-derived non-coding RNAs in cancer angiogenesis and lymphangiogenesis. *Biomolecules* 11, 249. doi:10.3390/biom11020249
- Aslan, C., Maralbashi, S., Salari, F., Kahroba, H., Sigaroodi, F., Kazemi, T., et al. (2019). Tumor-derived exosomes: Implication in angiogenesis and antiangiogenesis cancer therapy. *J. Cell. physiology* 234, 16885–16903. doi:10.1002/jcp.28374
- Bach, D. H., and Lee, S. K. (2018). Long noncoding RNAs in cancer cells. *Cancer Lett.* 419, 152–166. doi:10.1016/j.canlet.2018.01.053
- Bao, L., You, B., Shi, S., Shan, Y., Zhang, Q., Yue, H., et al. (2018). Metastasis-associated miR-23a from nasopharyngeal carcinoma-derived exosomes mediates angiogenesis by repressing a novel target gene TSGA10. *Oncogene* 37, 2873–2889. doi:10.1038/s41388-018-0183-6
- Barachini, S., Ghelardoni, S., and Madonna, R. J. J. o. C. M. (2023). Vascular progenitor cells: From cancer to tissue repair. *J. Clin. Med.* 12, 2399. doi:10.3390/jcm12062399
- Barsyte-Lovejoy, D., Lau, S. K., Boutros, P. C., Khosravi, F., Jurisica, I., Andrusis, I. L., et al. (2006). The c-Myc oncogene directly induces the H19 noncoding RNA by allele-specific binding to potentiate tumorigenesis. *Cancer Res.* 66, 5330–5337. doi:10.1158/0008-5472.CAN-06-0037
- Bartel, D. P. (2004). MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* 116, 281–297. doi:10.1016/s0092-8674(04)00045-5
- Bartel, D. P. (2009). MicroRNAs: Target recognition and regulatory functions. *Cell* 136, 215–233. doi:10.1016/j.cell.2009.01.002
- Braga, E. A., Fridman, M. V., Loginov, V. I., Dmitriev, A. A., and Morozov, S. G. (2019). Molecular mechanisms in clear cell renal cell carcinoma: Role of miRNAs and hypermethylated miRNA genes in crucial oncogenic pathways and processes. *Front. Genet.* 10, 320. doi:10.3389/fgene.2019.00320
- Carmeliet, P., and Jain, R. K. (2011). Molecular mechanisms and clinical applications of angiogenesis. *Nature* 473, 298–307. doi:10.1038/nature10144
- Castellano, J. J., Marrades, R. M., Molins, L., Viñolas, N., Moises, J., Canals, J., et al. (2020). Extracellular vesicle lincRNA-p21 expression in tumor-draining pulmonary vein defines prognosis in NSCLC and modulates endothelial cell behavior. *Cancers (Basel)* 12, 734. doi:10.3390/cancers12030734
- Chen, C. W., Fu, M., Du, Z. H., Zhao, F., Yang, W. W., Xu, L. H., et al. (2020). Long noncoding RNA MRPL23-AS1 promoteoid cystic carcinoma lung metastasis. *Cancer Res.* 80, 2273–2285. doi:10.1158/0008-5472.CAN-19-0819
- Chen, H. X., Xu, X. X., Tan, B. Z., Zhang, Z., and Zhou, X. D. (2017). MicroRNA-29b inhibits angiogenesis by targeting VEGFA through the MAPK/ERK and PI3K/akt signaling pathways in endometrial carcinoma. *Cell. physiology Biochem. Int. J. Exp. Cell. physiology, Biochem. Pharmacol.* 41, 933–946. doi:10.1159/000460510
- Chen, X., Zhang, S., Du, K., Zheng, N., Liu, Y., Chen, H., et al. (2021). Gastric cancer-secreted exosomal X26nt increases angiogenesis and vascular permeability by targeting VE-cadherin. *Cancer Sci.* 112, 1839–1852. doi:10.1111/cas.14740
- Cheng, C., Zhang, Z., Cheng, F., and Shao, Z. (2020). Exosomal lincRNA RAMP2-AS1 derived from chondrosarcoma cells promotes angiogenesis through miR-2355-5p/VEGFR2 Axis. *Onco Targets Ther.* 13, 3291–3301. doi:10.2147/OTT.S244652
- Cocks, A., Del Vecchio, F., Martinez-Rodriguez, V., Schukking, M., and Fabbri, M. (2022). Pro-tumoral functions of tumor-associated macrophage EV-miRNA. *Seminars cancer Biol.* 86, 58–63. doi:10.1016/j.semcancer.2021.08.001
- Conigliaro, A., Costa, V., Lo Dico, A., Saieva, L., Buccheri, S., Dieli, F., et al. (2015). CD90+ liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lincRNA. *Mol. Cancer* 14, 155. doi:10.1186/s12943-015-0426-x
- Creamer, K. M., and Lawrence, J. B. (2017). Xist RNA: A window into the broader role of RNA in nuclear chromosome architecture. *Philosophical Trans. R. Soc. Lond. Ser. B, Biol. Sci.* 372, 20160360. doi:10.1098/rstb.2016.0360
- Ding, B. S., Cao, Z., Lis, R., Nolan, D. J., Guo, P., Simons, M., et al. (2014). Divergent angiocrine signals from vascular niche balance liver regeneration and fibrosis. *Nature* 505, 97–102. doi:10.1038/nature12681
- Dominiak, A., Chelstowska, B., Olejars, W., and Nowicka, G. (2020). Communication in the cancer microenvironment as a target for therapeutic interventions. *Cancers* 12, 1232. doi:10.3390/cancers12051232
- Dong, R., Liu, X. Q., Zhang, B. B., Liu, B. H., Zheng, S., and Dong, K. R. (2017). Long non-coding RNA-CRND: A novel regulator of tumor growth and angiogenesis in hepatoblastoma. *Oncotarget* 8, 42087–42097. doi:10.18632/oncotarget.14992
- Dou, R., Liu, K., Yang, C., Zheng, J., Shi, D., Lin, X., et al. (2021). EMT-cancer cells-derived exosomal miR-27b-3p promotes circulating tumour cells-mediated metastasis by modulating vascular permeability in colorectal cancer. *Clin. Transl. Med.* 11, e595. doi:10.1002/ctm2.595
- Duan, B., Shi, S., Yue, H., You, B., Shan, Y., Zhu, Z., et al. (2019). Exosomal miR-17-5p promotes angiogenesis in nasopharyngeal carcinoma via targeting BAMB1. *J. Cancer* 10, 6681–6692. doi:10.7150/jca.30757
- Fang, J. H., Zhang, Z. J., Shang, L. R., Luo, Y. W., Lin, Y. F., Yuan, Y., et al. (2018). Hepatoma cell-secreted exosomal microRNA-103 increases vascular permeability and promotes metastasis by targeting junction proteins. *Hepatology* 68, 1459–1475. doi:10.1002/hep.29920
- Fang, X., Xu, Y., Li, K., Liu, P., Zhang, H., Jiang, Y., et al. (2022). Exosomal lincRNA PCAT1 promotes tumor circulating cell-mediated colorectal cancer liver metastasis by regulating the activity of the miR-329-3p/netrin-1-cd146 complex. *J. Immunol. Res.* 2022, 9916228. doi:10.1155/2022/9916228
- Fares, J., Fares, M. Y., Khachfe, H. H., Salhab, H. A., Fares, Y. J. S. t., and therapy, t. (2020). Molecular principles of metastasis: A hallmark of cancer revisited. *Signal Transduct. Target. Ther.* 5, 28. doi:10.1038/s41392-020-0134-x
- Forder, A., Hsing, C. Y., Trejo Vazquez, J., and Garnis, C. (2021). Emerging role of extracellular vesicles and cellular communication in metastasis. *Cells* 10, 3429. doi:10.3390/cells10123429
- Fu, W. M., Lu, Y. F., Hu, B. G., Liang, W. C., Zhu, X., Yang, H. D., et al. (2016). Long noncoding RNA Hotair mediated angiogenesis in nasopharyngeal carcinoma by direct and indirect signaling pathways. *Oncotarget* 7, 4712–4723. doi:10.18632/oncotarget.6731
- Geng, S., Tu, S., Bai, Z., and Geng, Y. (2022). Exosomal lincRNA LINC01356 derived from brain metastatic non-small-cell lung cancer cells remodels the blood-brain barrier. *Front. Oncol.* 12, 825899. doi:10.3389/fonc.2022.825899
- Giraldo, N. A., Sanchez-Salas, R., Peske, J. D., Vano, Y., Becht, E., Petitprez, F., et al. (2019). The clinical role of the TME in solid cancer. *Br. J. cancer* 120, 45–53. doi:10.1038/s41416-018-0327-z
- Gluzsko, A., Szczepański, M. J., Ludwig, N., Mirza, S. M., and Olejars, W. (2019). Exosomes in cancer: Circulating immune-related biomarkers. *BioMed Res. Int.* 2019, 1628029. doi:10.1155/2019/1628029
- Gong, C., and Maquat, L. E. (2011). lncRNAs transactivate STAU1-mediated mRNA decay by duplexing with 3' UTRs via Alu elements. *Nature* 470, 284–288. doi:10.1038/nature09701
- Guo, Z., Wang, X., Yang, Y., Chen, W., Zhang, K., Teng, B., et al. (2020). Hypoxic tumor-derived exosomal long noncoding RNA UCA1 promotes angiogenesis via miR-96-5p/AMOTL2 in pancreatic cancer. *Nucleic acids.* 22, 179–195. doi:10.1016/j.omtn.2020.08.021
- Guttman, M., Amit, I., Garber, M., French, C., Lin, M. F., Feldser, D., et al. (2009). Chromatin signature reveals over a thousand highly conserved large non-coding RNAs in mammals. *Nature* 458, 223–227. doi:10.1038/nature07672
- Han, W., Sulidankazha, Q., Nie, X., Yildan, R., and Len, K. (2021). Pancreatic cancer cells-derived exosomal long non-coding RNA CCAT1/microRNA-138-5p/HMGA1 axis promotes tumor angiogenesis. *Life Sci.* 278, 119495. doi:10.1016/j.lfs.2021.119495
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. *Cell* 144, 646–674. doi:10.1016/j.cell.2011.02.013
- He, G., Peng, X., Wei, S., Yang, S., Li, X., Huang, M., et al. (2022). Exosomes in the hypoxic TME: From release, uptake and biofunctions to clinical applications. *Mol. Cancer* 21, 19. doi:10.1186/s12943-021-01440-5
- He, L., Zhu, W., Chen, Q., Yuan, Y., Wang, Y., Wang, J., et al. (2019). Ovarian cancer cell-secreted exosomal miR-205 promotes metastasis by inducing angiogenesis. *Theranostics* 9, 8206–8220. doi:10.7150/thno.37455
- He, Q., Ye, A., Ye, W., Liao, X., Qin, G., Xu, Y., et al. (2021a). Cancer-secreted exosomal miR-21-5p induces angiogenesis and vascular permeability by targeting KRIT1. *Cell Death Dis.* 12, 576. doi:10.1038/s41419-021-03803-8
- He, S., Zhang, W., Li, X., Wang, J., Chen, X., Chen, Y., et al. (2021b). Oral squamous cell carcinoma (OSCC)-derived exosomal MiR-221 targets and regulates phosphoinositide-3-kinase regulatory subunit 1 (PIK3R1) to promote human umbilical vein endothelial cells migration and tube formation. *Bioengineered* 12, 2164–2174. doi:10.1080/21655979.2021.1932222

- Hsu, Y. L., Hung, J. Y., Chang, W. A., Lin, Y. S., Pan, Y. C., Tsai, P. H., et al. (2017). Hypoxic lung cancer-secreted exosomal miR-23a increased angiogenesis and vascular permeability by targeting prolyl hydroxylase and tight junction protein ZO-1. *Oncogene* 36, 4929–4942. doi:10.1038/ncr.2017.105
- Hu, H. Y., Yu, C. H., Zhang, H. H., Zhang, S. Z., Yu, W. Y., Yang, Y., et al. (2019). Exosomal miR-1229 derived from colorectal cancer cells promotes angiogenesis by targeting HIPK2. *Int. J. Biol. Macromol.* 132, 470–477. doi:10.1016/j.ijbiomac.2019.03.221
- Huang, Y., Kanada, M., Ye, J., Deng, Y., He, Q., Lei, Z., et al. (2022). Exosome-mediated remodeling of the tumor microenvironment: From local to distant intercellular communication. *Cancer Lett.* 543, 215796. doi:10.1016/j.canlet.2022.215796
- Huarte, M., Guttman, M., Feldser, D., Garber, M., Koziol, M. J., Kenzelmann-Broz, D., et al. (2010). A large intergenic noncoding RNA induced by p53 mediates global gene repression in the p53 response. *Cell* 142, 409–419. doi:10.1016/j.cell.2010.06.040
- Hutchinson, J. N., Ensminger, A. W., Clemson, C. M., Lynch, C. R., Lawrence, J. B., and Chess, A. (2007). A screen for nuclear transcripts identifies two linked noncoding RNAs associated with SC35 splicing domains. *BMC genomics* 8, 39. doi:10.1186/1471-2164-8-39
- Jászai, J., and Schmidt, M. H. H. (2019). Trends and challenges in tumor anti-angiogenic therapies. *Cells* 8, 1102. doi:10.3390/cells8091102
- Jia, P., Cai, H., Liu, X., Chen, J., Ma, J., Wang, P., et al. (2016). Long non-coding RNA H19 regulates glioma angiogenesis and the biological behavior of glioma-associated endothelial cells by inhibiting microRNA-29a. *Cancer Lett.* 381, 359–369. doi:10.1016/j.canlet.2016.08.009
- Jia, Z., Jia, J., Yao, L., and Li, Z. (2022). Crosstalk of exosomal non-coding RNAs in the tumor microenvironment: Novel Frontiers. *Front. Immunol.* 13, 900155. doi:10.3389/fimmu.2022.900155
- Jiang, J., Lu, J., Wang, X., Sun, B., Liu, X., Ding, Y., et al. (2021). Glioma stem cell-derived exosomal miR-944 reduces glioma growth and angiogenesis by inhibiting AKT/ERK signaling. *Aging (Albany NY)* 13, 19243–19259. doi:10.18632/aging.203243
- Junttila, M. R., and de Sauvage, F. J. (2013). Influence of tumour micro-environment heterogeneity on therapeutic response. *Nature* 501, 346–354. doi:10.1038/nature12626
- Kim, D. H., Park, S., Kim, H., Choi, Y. J., Kim, S. Y., Sung, K. J., et al. (2020). Tumor-derived exosomal miR-619-5p promotes tumor angiogenesis and metastasis through the inhibition of RCAN1.4. *Cancer Lett.* 475, 2–13. doi:10.1016/j.canlet.2020.01.023
- Kohlhapp, F. J., Mitra, A. K., Lengyel, E., and Peter, M. E. (2015). MicroRNAs as mediators and communicators between cancer cells and the tumor microenvironment. *Oncogene* 34, 5857–5868. doi:10.1038/ncr.2015.89
- Kong, X., Li, J., Li, Y., Duan, W., Qi, Q., Wang, T., et al. (2021). A novel long non-coding RNA AC073352.1 promotes metastasis and angiogenesis via interacting with YBX1 in breast cancer. *Cell Death Dis.* 12, 670. doi:10.1038/s41419-021-03943-x
- Lam, M. T., Cho, H., Lesch, H. P., Gosselin, D., Heinz, S., Tanaka-Oishi, Y., et al. (2013). Rev-Erbs repress macrophage gene expression by inhibiting enhancer-directed transcription. *Nature* 498, 511–515. doi:10.1038/nature12209
- Lang, H. L., Hu, G. W., Chen, Y., Liu, Y., Tu, W., Lu, Y. M., et al. (2017a). Glioma cells promote angiogenesis through the release of exosomes containing long non-coding RNA POU3F3. *Eur. Rev. Med. Pharmacol. Sci.* 21, 959–972.
- Lang, H. L., Hu, G. W., Zhang, B., Kuang, W., Chen, Y., Wu, L., et al. (2017b). Glioma cells enhance angiogenesis and inhibit endothelial cell apoptosis through the release of exosomes that contain long non-coding RNA CCAT2. *Oncol. Rep.* 38, 785–798. doi:10.3892/or.2017.5742
- Lee, S., Kopp, F., Chang, T. C., Sataluri, A., Chen, B., Sivakumar, S., et al. (2016). Noncoding RNA NORAD regulates genomic stability by sequestering PUMILIO proteins. *Cell* 164, 69–80. doi:10.1016/j.cell.2015.12.017
- Lei, L., and Mou, Q. (2020). Exosomal taurine up-regulated 1 promotes angiogenesis and endothelial cell proliferation in cervical cancer. *Cancer Biol. Ther.* 21, 717–725. doi:10.1080/15384047.2020.1764318
- Li, J., Yuan, H., Xu, H., Zhao, H., and Xiong, N. (2020a). Hypoxic cancer-secreted exosomal miR-182-5p promotes glioblastoma angiogenesis by targeting kruppel-like factor 2 and 4. *Mol. Cancer Res.* 18, 1218–1231. doi:10.1158/1541-7786.MCR-19-0725
- Li, K., Blum, Y., Verma, A., Liu, Z., Pramanik, K., Leigh, N. R., et al. (2010). A noncoding antisense RNA in tie-1 locus regulates tie-1 function *in vivo*. *Blood* 115, 133–139. doi:10.1182/blood-2009-09-242180
- Li, S., Qi, Y., Huang, Y., Guo, Y., Huang, T., and Jia, L. (2021). Exosome-derived SNHG16 sponging miR-4500 activates HUVEC angiogenesis by targeting GALNT1 via PI3K/Akt/mTOR pathway in hepatocellular carcinoma. *J. Physiol. Biochem.* 77, 667–682. doi:10.1007/s13105-021-00833-w
- Li, W. D., Zhou, D. M., Sun, L. L., Xiao, L., Liu, Z., Zhou, M., et al. (2018). LncRNA WTAPP1 promotes migration and angiogenesis of endothelial progenitor cells via MMP1 through MicroRNA 3120 and akt/PI3K/autophagy pathways. *Stem cells Dayt. Ohio* 36, 1863–1874. doi:10.1002/stem.2904
- Li, W., Fu, Q., Man, W., Guo, H., and Yang, P. (2019). LncRNA OR3A4 participates in the angiogenesis of hepatocellular carcinoma through modulating ACGF1/akt/mTOR pathway. *Eur. J. Pharmacol.* 849, 106–114. doi:10.1016/j.ejphar.2019.01.049
- Li, Y., Cai, B., Shen, L., Dong, Y., Lu, Q., Sun, S., et al. (2017). MiRNA-29b suppresses tumor growth through simultaneously inhibiting angiogenesis and tumorigenesis by targeting Akt3. *Cancer Lett.* 397, 111–119. doi:10.1016/j.canlet.2017.03.032
- Li, Y., Zhang, H., Fan, L., Mou, J., Yin, Y., Peng, C., et al. (2020b). MiR-629-5p promotes the invasion of lung adenocarcinoma via increasing both tumor cell invasion and endothelial cell permeability. *Oncogene* 39, 3473–3488. doi:10.1038/s41388-020-1228-1
- Li, Y., Zhang, X., Zheng, Q., Zhang, Y., Ma, Y., Zhu, C., et al. (2020c). YAP1 inhibition in HUVECs is associated with released exosomes and increased hepatocarcinoma invasion and metastasis. *Mol. Ther. Nucleic Acids* 21, 86–97. doi:10.1016/j.omtn.2020.05.021
- Lin, J., Cao, S., Wang, Y., Hu, Y., Liu, H., Li, J., et al. (2018b). Long non-coding RNA UBE2CP3 enhances HCC cell secretion of VEGFA and promotes angiogenesis by activating ERK1/2/HIF-1 α /VEGFA signalling in hepatocellular carcinoma. *J. Exp. Clin. Cancer Res.* 37, 113. doi:10.1186/s13046-018-0727-1
- Lin, X. J., Fang, J. H., Yang, X. J., Zhang, C., Yuan, Y., Zheng, L., et al. (2018a). Hepatocellular carcinoma cell-secreted exosomal MicroRNA-210 promotes angiogenesis *in vitro* and *in vivo*. *Mol. Ther. Nucleic Acids* 11, 243–252. doi:10.1016/j.omtn.2018.02.014
- Li, B., Chen, J., Shang, F., Lian, M., Shen, X., and Fang, J. (2022). Tumor-derived exosome FGD5-AS1 promotes angiogenesis, vascular permeability, and metastasis in thyroid cancer by targeting the miR-6838-5p/VAV2 Axis. *J. Oncol.* 2022, 4702855. doi:10.1155/2022/4702855
- Liu, H., Lei, C., He, Q., Pan, Z., Xiao, D., and Tao, Y. (2018). Nuclear functions of mammalian MicroRNAs in gene regulation, immunity and cancer. *Mol. Cancer* 17, 64. doi:10.1186/s12943-018-0765-5
- Lu, J., Liu, Q. H., Wang, F., Tan, J. J., Deng, Y. Q., Peng, X. H., et al. (2018). Exosomal miR-9 inhibits angiogenesis by targeting MDK and regulating PDK/AKT pathway in nasopharyngeal carcinoma. *J. Exp. Clin. Cancer Res.* 37, 147. doi:10.1186/s13046-018-0814-3
- Lu, Y., Chen, L., Li, L., and Cao, Y. (2020). Exosomes derived from brain metastatic breast cancer cells destroy the blood-brain barrier by carrying lncRNA GS1-600g8.5. *Biomed. Res. Int.* 2020, 7461727. doi:10.1155/2020/7461727
- Malakoti, F., Targhazeh, N., Karimzadeh, H., Mohammadi, E., Asadi, M., Asemi, Z., et al. (2021). Multiple function of lncRNA MALAT1 in cancer occurrence and progression. *Chem. Biol. Drug Des.* 101, 1113–1137. doi:10.1111/cbdd.14006
- Man, H. S. J., Sukumar, A. N., Lam, G. C., Turgeon, P. J., Yan, M. S., Ku, K. H., et al. (2018). Angiogenic patterning by STEEL, an endothelial-enriched long noncoding RNA. *Proc. Natl. Acad. Sci. U. S. A.* 115, 2401–2406. doi:10.1073/pnas.1715182115
- Mao, S., Lu, Z., Zheng, S., Zhang, H., Zhang, G., Wang, F., et al. (2020). Exosomal miR-141 promotes tumor angiogenesis via KLF12 in small cell lung cancer. *J. Exp. Clin. Cancer Res.* 39, 193. doi:10.1186/s13046-020-01680-1
- Mao, S., Zheng, S., Lu, Z., Wang, X., Wang, Y., Zhang, G., et al. (2021). Exosomal miR-375-3p breaks vascular barrier and promotes small cell lung cancer metastasis by targeting claudin-1. *Transl. Lung Cancer Res.* 10, 3155–3172. doi:10.21037/tlcr-21-356
- Masoumi-Dehghi, S., Babashah, S., and Sadeghizadeh, M. (2020). microRNA-141-3p-containing small extracellular vesicles derived from epithelial ovarian cancer cells promote endothelial cell angiogenesis through activating the JAK/STAT3 and NF- κ B signaling pathways. *J. Cell Commun. Signal* 14, 233–244. doi:10.1007/s12079-020-00548-5
- Melo, C. A., Drost, J., Wijchers, P. J., van de Werken, H., de Wit, E., Oude Vrielink, J. A., et al. (2013). eRNAs are required for p53-dependent enhancer activity and gene transcription. *Mol. Cell* 49, 524–535. doi:10.1016/j.molcel.2012.11.021
- Mezzadra, R., Sun, C., Jae, L. T., Gomez-Eerland, R., de Vries, E., Wu, W., et al. (2017). Identification of CMTM6 and CMTM4 as PD-L1 protein regulators. *Nature* 549, 106–110. doi:10.1038/nature23669
- Moh-Moh-Aung, A., Fujisawa, M., Ito, S., Katayama, H., Ohara, T., Ota, Y., et al. (2020). Decreased miR-200b-3p in cancer cells leads to angiogenesis in HCC by enhancing endothelial ERG expression. *Sci. Rep.* 10, 10418. doi:10.1038/s41598-020-67425-4
- Nagano, T., and Fraser, P. (2011). Non-nonsense functions for long noncoding RNAs. *Cell* 145, 178–181. doi:10.1016/j.cell.2011.03.014
- Qiu, J. J., Lin, X. J., Tang, X. Y., Zheng, T. T., Lin, Y. Y., and Hua, K. Q. (2018). Exosomal metastasis-associated lung adenocarcinoma transcript 1 promotes angiogenesis and predicts poor prognosis in epithelial ovarian cancer. *Int. J. Biol. Sci.* 14, 1960–1973. doi:10.7150/ijbs.28048
- Qiu, M., Xu, Y., Wang, J., Zhang, E., Sun, M., Zheng, Y., et al. (2015). A novel lncRNA, LUADT1, promotes lung adenocarcinoma proliferation via the epigenetic suppression of p27. *Cell Death Dis.* 6, e1858. doi:10.1038/cddis.2015.203
- Shang, D., Xie, C., Hu, J., Tan, J., Yuan, Y., Liu, Z., et al. (2020). Pancreatic cancer cell-derived exosomal microRNA-27a promotes angiogenesis of human microvascular endothelial cells in pancreatic cancer via BTG2. *J. Cell Mol. Med.* 24, 588–604. doi:10.1111/jcmm.14766
- Shi, W., Zhang, C., Ning, Z., Hua, Y., Li, Y., Chen, L., et al. (2021). CMTM8 as an LPA1-associated partner mediates lysophosphatidic acid-induced pancreatic cancer metastasis. *Ann. Transl. Med.* 9, 42. doi:10.21037/atm-20-1013
- Shi, Y., Yang, X., Xue, X., Sun, D., Cai, P., Song, Q., et al. (2019). HANR promotes lymphangiogenesis of hepatocellular carcinoma via secreting miR-296 exosome and regulating EAG1/VEGFA signaling in HDLEC cells. *J. Cell Biochem.* 120, 17699–17708. doi:10.1002/jcb.29036

- Shweiki, D., Itin, A., Soffer, D., and Keshet, E. (1992). Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature* 359, 843–845. doi:10.1038/359843a0
- Soheilifar, M. H., Masoudi-Khoram, N., Madadi, S., Nobari, S., Maadi, H., Keshmiri Neghab, H., et al. (2022). Angioregulatory microRNAs in breast cancer: Molecular mechanistic basis and implications for therapeutic strategies. *J. Adv. Res.* 37, 235–253. doi:10.1016/j.jare.2021.06.019
- Song, J., Shu, H., Zhang, L., and Xiong, J. (2019). Long noncoding RNA GAS5 inhibits angiogenesis and metastasis of colorectal cancer through the Wnt/ β -catenin signaling pathway. *J. Cell Biochem.* 120, 6937–6951. doi:10.1002/jcb.27743
- Spizzo, R., Almeida, M. I., Colombatti, A., and Calin, G. A. (2012). Long non-coding RNAs and cancer: A new frontier of translational research? *Oncogene* 31, 4577–4587. doi:10.1038/onc.2011.621
- Stacker, S. A., Williams, S. P., Karnezis, T., Shayan, R., Fox, S. B., and Achen, M. G. (2014). Lymphangiogenesis and lymphatic vessel remodelling in cancer. *Nat. Rev. Cancer* 14, 159–172. doi:10.1038/nrc3677
- Stec, M., Baj-Krzyworzeka, M., Baran, J., Węglarczyk, K., Zembala, M., Barbasz, J., et al. (2015). Isolation and characterization of circulating micro(nano)vesicles in the plasma of colorectal cancer patients and their interactions with tumor cells. *Oncol. Rep.* 34, 2768–2775. doi:10.3892/or.2015.4228
- Sun, B., Han, Y., and Shi, M. (2021). Stromal-derived miR-486-5p promotes metastasis of non-small-cell lung cancer cells by targeting the CADM1/tight junctions axis in vascular endothelial cells. *Cell Biol. Int.* 45, 849–857. doi:10.1002/cbin.11531
- Tao, K., Liu, J., Liang, J., Xu, X., Xu, L., and Mao, W. (2021). Vascular endothelial cell-derived exosomal miR-30a-5p inhibits lung adenocarcinoma malignant progression by targeting CCNE2. *Carcinogenesis* 42, 1056–1067. doi:10.1093/carcin/bgab051
- Tao, S. C., Huang, J. Y., Wei, Z. Y., Li, Z. X., and Guo, S. C. (2020). EWSAT1 acts in concert with exosomes in osteosarcoma progression and tumor-induced angiogenesis: The "double stacking effect. *Adv. Biosyst.* 4, e2000152. doi:10.1002/adbi.202000152
- Tripathi, V., Shen, Z., Chakraborty, A., Giri, S., Freier, S. M., Wu, X., et al. (2013). Long noncoding RNA MALAT1 controls cell cycle progression by regulating the expression of oncogenic transcription factor B-MYB. *PLoS Genet.* 9, e1003368. doi:10.1371/journal.pgen.1003368
- Uhlmann, S., Mannsperger, H., Zhang, J. D., Horvat, E., Schmidt, C., Küblbeck, M., et al. (2012). Global microRNA level regulation of EGFR-driven cell-cycle protein network in breast cancer. *Mol. Syst. Biol.* 8, 570. doi:10.1038/msb.2011.100
- Umez, T., Ohyashiki, K., Kuroda, M., and Ohyashiki, J. H. (2013). Leukemia cell to endothelial cell communication via exosomal miRNAs. *Oncogene* 32, 2747–2755. doi:10.1038/onc.2012.295
- Wang, F. W., Cao, C. H., Han, K., Zhao, Y. X., Cai, M. Y., Xiang, Z. C., et al. (2021b). APC-activated long noncoding RNA inhibits colorectal carcinoma pathogenesis through reduction of exosome production. *J. Clin. Invest.* 131, e149666. doi:10.1172/JCI149666
- Wang, H., Wang, L., Zhou, X., Luo, X., Liu, K., Jiang, E., et al. (2020b). OSCC exosomes regulate miR-210-3p targeting EFNA3 to promote oral cancer angiogenesis through the PI3K/AKT pathway. *Biomed. Res. Int.* 2020, 2125656. doi:10.1155/2020/2125656
- Wang, M., Zhao, Y., Yu, Z. Y., Zhang, R. D., Li, S. A., Zhang, P., et al. (2020a). Glioma exosomal microRNA-148a-3p promotes tumor angiogenesis through activating the EGFR/MAPK signaling pathway via inhibiting ERRF1. *Cancer Cell Int.* 20, 518. doi:10.1186/s12935-020-01566-4
- Wang, Q., Wang, G., Niu, L., Zhao, S., Li, J., Zhang, Z., et al. (2021a). Exosomal MiR-1290 promotes angiogenesis of hepatocellular carcinoma via targeting SMEK1. *J. Oncol.* 2021, 6617700. doi:10.1155/2021/6617700
- Wang, Y., Han, D., Pan, L., and Sun, J. (2018a). The positive feedback between lncRNA TNK2-AS1 and STAT3 enhances angiogenesis in non-small cell lung cancer. *Biochem. Biophys. Res. Commun.* 507, 185–192. doi:10.1016/j.bbrc.2018.11.004
- Wang, Y., Zhang, F., Wang, J., Hu, L., Jiang, F., Chen, J., et al. (2018b). lncRNA LOC100132354 promotes angiogenesis through VEGFA/VEGFR2 signaling pathway in lung adenocarcinoma. *Cancer Manag. Res.* 10, 4257–4266. doi:10.2147/CMAR.S177327
- Wang, Z. F., Liao, F., Wu, H., and Dai, J. (2019). Glioma stem cells-derived exosomal miR-26a promotes angiogenesis of microvessel endothelial cells in glioma. *J. Exp. Clin. Cancer Res.* 38, 201. doi:10.1186/s13046-019-1181-4
- Wu, D., Deng, S., Li, L., Liu, T., Zhang, T., Li, J., et al. (2021). TGF- β 1-mediated exosomal lnc-MMP2-2 increases blood-brain barrier permeability via the miRNA-1207-5p/EPB41L5 axis to promote non-small cell lung cancer brain metastasis. *Cell Death Dis.* 12, 721. doi:10.1038/s41419-021-04004-z
- Wu, X. G., Zhou, C. F., Zhang, Y. M., Yan, R. M., Wei, W. F., Chen, X. J., et al. (2019). Cancer-derived exosomal miR-221-3p promotes angiogenesis by targeting THBS2 in cervical squamous cell carcinoma. *Angiogenesis* 22, 397–410. doi:10.1007/s10456-019-09665-1
- Xu, Z., Chen, Y., Ma, L., Chen, Y., Liu, J., Guo, Y., et al. (2022). Role of exosomal non-coding RNAs from tumor cells and tumor-associated macrophages in the tumor microenvironment. *Mol. Ther. J. Am. Soc. Gene Ther.* 30, 3133–3154. doi:10.1016/j.ythet.2022.01.046
- Yan, W., Wang, Y., Chen, Y., Guo, Y., Li, Q., and Wei, X. (2021). Exosomal miR-130b-3p promotes progression and tubular formation through targeting PTEN in oral squamous cell carcinoma. *Front. Cell Dev. Biol.* 9, 616306. doi:10.3389/fcell.2021.616306
- Yang, E., Wang, X., Gong, Z., Yu, M., Wu, H., Zhang, D. J. S. t., et al. (2020). Exosome-mediated metabolic reprogramming: The emerging role in tumor microenvironment remodeling and its influence on cancer progression. *Signal Transduct. Target. Ther.* 5, 242. doi:10.1038/s41392-020-00359-5
- Yang, J., and Teng, Y. (2023). Harnessing cancer stem cell-derived exosomes to improve cancer therapy. *J. Exp. Clin. Cancer Res.* CR 42, 131. doi:10.1186/s13046-023-02717-x
- Yang, K., Zhou, Q., Qiao, B., Shao, B., Hu, S., Wang, G., et al. (2022). Exosome-derived noncoding RNAs: Function, mechanism, and application in tumor angiogenesis. *Nucleic Acids.* 27, 983–997. doi:10.1016/j.omtn.2022.01.009
- Yao, C., Wu, S., Kong, J., Sun, Y., Bai, Y., Zhu, R., et al. (2023). Angiogenesis in hepatocellular carcinoma: Mechanisms and anti-angiogenic therapies. *Cancer Biol. Med.* 20, 25–43. doi:10.20892/j.issn.2095-3941.2022.0449
- Yin, Z., Ma, T., Huang, B., Lin, L., Zhou, Y., Yan, J., et al. (2019). Macrophage-derived exosomal microRNA-501-3p promotes progression of pancreatic ductal adenocarcinoma through the TGFBR3-mediated TGF- β signaling pathway. *J. Exp. Clin. Cancer Res.* 38, 310–320. doi:10.1186/s13046-019-1313-x
- Yokota, Y., Noda, T., Okumura, Y., Kobayashi, S., Iwagami, Y., Yamada, D., et al. (2021). Serum exosomal miR-638 is a prognostic marker of HCC via downregulation of VE-cadherin and ZO-1 of endothelial cells. *Cancer Sci.* 112, 1275–1288. doi:10.1111/cas.14807
- You, X., Sun, W., Wang, Y., Liu, X., Wang, A., Liu, L., et al. (2021). Cervical cancer-derived exosomal miR-663b promotes angiogenesis by inhibiting vinculin expression in vascular endothelial cells. *Cancer Cell Int.* 21, 684. doi:10.1186/s12935-021-02379-9
- Zampetaki, A., Albrecht, A., and Steinhilber, K. (2018). Long non-coding RNA structure and function: Is there a link? *Front. Physiology* 9, 1201. doi:10.3389/fphys.2018.01201
- Zeng, Z., Li, Y., Pan, Y., Lan, X., Song, F., Sun, J., et al. (2018). Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. *Nat. Commun.* 9, 5395. doi:10.1038/s41467-018-07810-w
- Zhang, C. Y., Yu, M. S., Li, X., Zhang, Z., Han, C. R., and Yan, B. (2017c). Overexpression of long non-coding RNA MEG3 suppresses breast cancer cell proliferation, invasion, and angiogenesis through AKT pathway. *Tumour Biol.* 39, 1010428317701311. doi:10.1177/1010428317701311
- Zhang, J. W., Liu, T. F., Chen, X. H., Liang, W. Y., Feng, X. R., Wang, L., et al. (2017b). Validation of aspirin response-related transcripts in patients with coronary artery disease and preliminary investigation on CMTM5 function. *Gene* 624, 56–65. doi:10.1016/j.gene.2017.04.041
- Zhang, J. X., Chen, Z. H., Chen, D. L., Tian, X. P., Wang, C. Y., Zhou, Z. W., et al. (2018). LINC01410-miR-532-NCF2-NF- κ B feedback loop promotes gastric cancer angiogenesis and metastasis. *Oncogene* 37, 2660–2675. doi:10.1038/s41388-018-0162-y
- Zhang, Z. C., Tang, C., Dong, Y., Zhang, J., Yuan, T., Tao, S. C., et al. (2017a). Targeting the long noncoding RNA MALAT1 blocks the pro-angiogenic effects of osteosarcoma and suppresses tumour growth. *Int. J. Biol. Sci.* 13, 1398–1408. doi:10.7150/ijbs.22249
- Zhao, J., Du, P., Cui, P., Qin, Y., Hu, C., Wu, J., et al. (2018). lncRNA PVT1 promotes angiogenesis via activating the STAT3/VEGFA axis in gastric cancer. *Oncogene* 37, 4094–4109. doi:10.1038/s41388-018-0250-5
- Zhao, S., Li, J., Zhang, G., Wang, Q., Wu, C., Zhang, Q., et al. (2019). Exosomal miR-451a functions as a tumor suppressor in hepatocellular carcinoma by targeting LPIN1. *Cell Physiol. Biochem.* 53, 19–35. doi:10.33594/0000001118
- Zhao, Z., Sun, W., Guo, Z., Zhang, J., Yu, H., and Liu, B. (2020). Mechanisms of lncRNA/microRNA interactions in angiogenesis. *Life Sci.* 254, 116900. doi:10.1016/j.lfs.2019.116900
- Zhong, M. E., Chen, Y., Zhang, G., Xu, L., Ge, W., and Wu, B. (2019). lncRNA H19 regulates PI3K-akt signal pathway by functioning as a ceRNA and predicts poor prognosis in colorectal cancer: Integrative analysis of dysregulated ncRNA-associated ceRNA network. *Cancer Cell Int.* 19, 148. doi:10.1186/s12935-019-0866-2
- Zhou, C. F., Ma, J., Huang, L., Yi, H. Y., Zhang, Y. M., Wu, X. G., et al. (2019). Cervical squamous cell carcinoma-secreted exosomal miR-221-3p promotes lymphangiogenesis and lymphatic metastasis by targeting VASH1. *Oncogene* 38, 1256–1268. doi:10.1038/s41388-018-0511-x
- Zhou, C., Zhang, Y., Yan, R., Huang, L., Mellor, A. L., Yang, Y., et al. (2021). Exosome-derived miR-142-5p remodels lymphatic vessels and induces Ido to promote immune privilege in the tumour microenvironment. *Cell Death Differ.* 28, 715–729. doi:10.1038/s41418-020-00618-6
- Zhou, S. K., Gao, F., Zhong, Z. S., and Yao, H. (2020). Long non-coding RNA colon cancer associated transcript-2 from nasopharyngeal carcinoma-derived exosomes promotes angiogenesis. *Zhonghua Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 55, 944–951. doi:10.3760/cma.j.cn115330-20200423-00322
- Zhou, W., Fong, M. Y., Min, Y., Somlo, G., Liu, L., Palomares, M. R., et al. (2014). Cancer-secreted miR-105 destroys vascular endothelial barriers to promote metastasis. *Cancer Cell* 25, 501–515. doi:10.1016/j.ccr.2014.03.007
- Zhu, H. Y., Gao, Y. J., Wang, Y., Liang, C., Zhang, Z. X., and Chen, Y. (2021). lncRNA CRNDE promotes the progression and angiogenesis of pancreatic cancer via miR-451a/CDKN2D axis. *Transl. Oncol.* 14, 101088. doi:10.1016/j.tranon.2021.101088
- Zhu, Y., Zhang, X., Qi, L., Cai, Y., Yang, P., Xuan, G., et al. (2016). HULC long noncoding RNA silencing suppresses angiogenesis by regulating ESM-1 via the PI3K/Akt/mTOR signaling pathway in human gliomas. *Oncotarget* 7, 14429–14440. doi:10.18632/oncotarget.7418