Check for updates

OPEN ACCESS

EDITED BY Shengxi Chen, Arizona State University, United States

REVIEWED BY

Daniela Miladinova, Saints Cyril and Methodius University of Skopje, North Macedonia Yuqin Shang, Arizona State University, United States Ping Tang, Loyola University Medical Center, United States

*CORRESPONDENCE Nicola Fusco, ⊠ nicola.fusco@ieo.it

SPECIALTY SECTION

This article was submitted to Molecular Diagnostics and Therapeutics, a section of the journal Frontiers in Molecular Biosciences

RECEIVED 28 February 2023 ACCEPTED 23 March 2023 PUBLISHED 03 April 2023

CITATION

Sajjadi E, Guerini-Rocco E, De Camilli E, Pala O, Mazzarol G, Venetis K, Ivanova M and Fusco N (2023), Pathological identification of HER2-low breast cancer: Tips, tricks, and troubleshooting for the optimal test. *Front. Mol. Biosci.* 10:1176309. doi: 10.3389/fmolb.2023.1176309

COPYRIGHT

© 2023 Sajjadi, Guerini-Rocco, De Camilli, Pala, Mazzarol, Venetis, Ivanova and Fusco. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Pathological identification of HER2-low breast cancer: Tips, tricks, and troubleshooting for the optimal test

Elham Sajjadi^{1,2}, Elena Guerini-Rocco^{1,2}, Elisa De Camilli², Oriana Pala², Giovanni Mazzarol², Konstantinos Venetis², Mariia Ivanova² and Nicola Fusco^{1,2}*

¹Department of Oncology and Hemato-Oncology, University of Milan, Milan, Italy, ²Division of Pathology, IEO, European Institute of Oncology IRCCS, Milan, Italy

The introduction of novel anti-HER2 antibody-drug conjugates (ADC) for the treatment of HER2-low breast cancers has transformed the traditional dichotomy of HER2 status to an expanded spectrum. However, the identification of HER2-low (i.e., immunohistochemistry (IHC) score 1 + or IHC score 2+, without gene amplification) tumors is challenged by methodological and analytical variables that might influence the sensitivity and reproducibility of HER2 testing. To open all possible therapeutic opportunities for HER2-low breast cancer patients the implementation of more accurate and reproducible testing strategies is mandatory. Here, we provide an overview of the existing barriers that may trouble HER2-low identification in breast cancer and discuss practical solutions that could enhance HER-low assessment.

KEYWORDS

breast cancer, HER2, HER2-low, pathology, precision medicine, immunohistochemistry, *in situ* hybridization

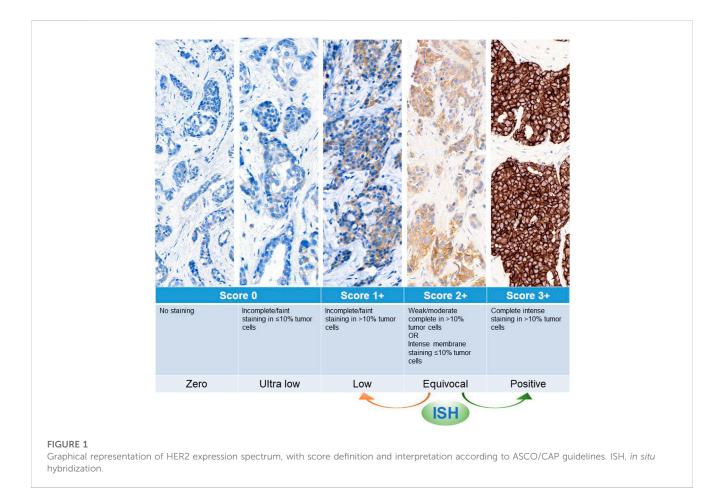
Introduction

HER2 testing is a standard procedure for all new breast cancer diagnoses, as well as in case of tumor progression and/or residual tumor after neoadjuvant treatment (Venetis et al., 2022; Pondé et al., 2018; Viale and Fusco, 2022; Fusco et al., 2022). This analysis relies on a combination of immunohistochemistry (IHC) and in situ hybridization (ISH). In particular, IHC detects the expression and intensity of HER2 protein on the cell membrane by a three-tier scoring system (from score 0 to score 3+) Table 1; Figure 1, while ISH detects the presence of gene amplification using HER2 and CEP17 probes (Sajjadi et al., 2022). To date, HER2+ breast cancer patients are defined as IHC score 3 + or score 2+ with a positive ISH, and subsequently qualified for anti-HER2 targeted therapy (Pondé et al., 2018; Wolff et al., 2018). Based on the DESTINY-Breast04 trial, metastatic breast cancer (mBC) with low levels of HER2 expression (i.e., IHC score 1 + or score 2+/ISH-negative) could benefit of the new anti-HER2 antibody-drug conjugate (ADC) Trastuzumab Deruxtecan (Modi et al., 2022). This drug contains a strong chemotherapeutic payload that is guided against tumor cells even with the presence of low traces of HER2 positivity thanks to the so-called "bystander effect" (Zhang et al., 2020; Venetis et al., 2022). The precise identification of HER2-low, and in particular the discrimination of score 1 + vs. score 0, is impacted by methodological and analytical

Membrane staining pattern	Tumor cells	Score	Classical category	Expanded spectrum
Intense, complete	>10%	3+	HER2+	HER2+
Weak-to-moderate, complete	>10%	2+	HER2+ (if ISH+)	HER2+ (if ISH+)
			HER2- (if ISH-)	HER2-low (if ISH-)
Faint/barely perceptible, incomplete	>10%	1+	HER2-	HER2-low
Faint/barely perceptible, incomplete	≤10%	0	HER2-	HER2 Ultra low
No staining			HER2-	HER2-zero

TABLE 1 HER2 scoring by immunohistochemistry according to the expanded spectrum of positivity (ASCO/CAP 2021).

Abbreviations: IHC, immunohistochemistry; ISH, in situ hybridization; HER2, human epidermal growth factor receptor 2.



variables (Sajjadi et al., 2022). In this respect, several studies are ongoing for the development of new technologies and implementation of dedicated guidelines that could provide more comprehensive and accurate assessments of HER2 status in view of HER2-low mBC. Among novel methods, artificial intelligence (AI)-based digital pathology has the potential to complement the traditional pathological analysis and to improve the accuracy of HER2 testing. In this mini review we will discuss all these challenges and opportunities, providing practical suggestions for HER2 testing, and "old" but at the same time "new" task in pathology.

Current challenges: how far from the perfect test?

The identification of low HER2 expression levels is not a trivial task because it relies on multiple methodological and analytical variables (Table 2). These variables might trouble the testing sensitivity and reproducibility, particularly for the discrimination between HER2-low score 1+ and "HER2-zero" (i.e., IHC score 0), which comprises also the subset of "HER2 ultra-low" (i.e., score 0 with incomplete and faint staining in $\leq 10\%$ of tumor cells) (Sajjadi et al., 2022). The

Phase	Current challenges	Possible solutions	
Pre- analytical	Variables influencing the IHC staining intensity: fixation, antigen retrieval, reaction time, temperature, and substrate concentration	Implementation and follow-up of strict SOPs, describing precisely the workflow	
		Repeating the test for equivocal results may help to exclude technical problems	
	The availability of different antibody clones with varying specificity	Updated guidelines that assess the reliability of commonly used HER2 IHC testing methods	
Analytical	The subjective mode of HER2 assessment, observer variability	Rigorous quality control procedures and well-defined guidelines based on amplified clinical trials and patient recruitment to further educate pathologists for higher concordance in scoring	
		Implementation of methodologies to improve HER2 assessment including machine learning approaches	
	The heterogeneity of HER2 expression and/or amplification	Recruitment of further diagnostic approaches with a more precise cut-off	
		Further investigation on whether the presence of intratumor heterogeneity may affect the efficacy of ADCs	

TABLE 2 Summary table listing the current challenges and future directions to improve HER2-low scoring.

Abbreviations: IHC, immunohistochemistry; SOPs, standard operating procedures; HER2, human epidermal growth factor receptor 2; ADC, antibody drug conjugate.

tissue sample handling and processing remains a crucial task, mainly subjected to the "garbage-in-garbage-out" paradigm (Perez et al., 2014). Variables such as fixation, antigen retrieval, antibody clones, reaction time, temperature, and substrate concentration can all influence the IHC staining intensity (Sapino et al., 2013; Zhang et al., 2021). Additionally, the choice of staining methodology, particularly antigen retrieval, and the availability of different antibody clones with varying specificity can also impact the accuracy and reproducibility of results (Schrohl et al., 2011; Zhang et al., 2021). The VENTANA PATHWAY anti-HER2/ neu (4B5) rabbit monoclonal primary antibody has been approved by the FDA as the first companion diagnostic test to identify HER2-low mBC patients for trastuzumab deruxtecan (T-DXd) targeted therapy (Zhang and Peng, 2022). However, assessing the reliability of other commonly used HER2 IHC testing methods is still pending (Zhang and Peng, 2022). Repeating the test for equivocal results may help to exclude technical problems, however, it does not always lead to a definitive result. It should be noted that the HER2 IHC testing was initially designed to distinguish high levels of HER2 expression (i.e., almost 2 million molecules/cell which corresponds to HER2 IHC 3+) from lower levels of HER2 expression (i.e., 20,000 to 500,000 molecules/cell for HER2 IHC 0 to 2+). Therefore, this method has not been developed for detecting HER2-low tumors (Zhang and Peng, 2022). On the other hand, in the post-analytical phase, interobserver variability can occur due to the lack of consistent epithelial internal positive control for HER2 in non-neoplastic breast tissue (Sajjadi et al., 2022; Zhang et al., 2022). The subjective mode of HER2 assessment and HER2 intratumoral heterogeneity in HER2-low breast cancer are other barriers with great influence on HER2-low assessment (Bianchi et al., 2015; Jensen et al., 2020; Zhang et al., 2020). To determine the suitability of the current standard HER2 IHC assays for identifying patients with HER2-low breast cancers, a concordance survey of 18 pathologists reading 170 breast cancer biopsies was carried out (Fernandez et al., 2022). Not

surprisingly, the concordance for IHC scores between score 0 and score 1+ was lower compared to score 2 + vs. score 3+ (26% and 58% concordance, respectively). This difference is likely related to the fact that before the T-DXd approval the distinction between score 0 and score 1+, unlike score 2+ and score 3+, was not related to true clinical implications (Fernandez et al., 2022). It should be noted that almost 50% of patients with breast cancer have HER2-low disease (Banerji et al., 2019; Tarantino et al., 2020). This inaccuracy in the real-world clinical practice could lead to misassignment of a great number of potential candidates for ADC therapies (Yue et al., 2021).

The heterogeneity of HER2 expression and/or amplification is another important challenge that should be considered for improving HER2-low identification. HER2 heterogeneity could be seen in both HER2 gene copy number and/or protein expression (Brunelli et al., 2009; Seol et al., 2012). The American Society of Clinical Oncology (ASCO)/College of American Pathologists (CAP) guidelines have clearly defined intra-tumor heterogeneity of HER2 amplification (i.e., HER2/CEP17 signal ratios >2.2 in 5%-50% of the neoplastic cells). However, there is a lack of consensus to define HER2 heterogeneity for IHC (Ahn et al., 2020; Sajjadi et al., 2022). It should be noted that intratumoral HER2 IHC heterogeneity (i.e., uneven distribution of HER2 expression or different intensities of HER2 staining in tumor cells) is more frequent in the HER2-low (2 + or 1 + IHC score) samples (Seol et al., 2012; Wu et al., 2023a). This condition may contribute to the poor consistency of HER2-low interpretation (Wu et al., 2023a). Moreover, whether the presence of intratumor heterogeneity may affect the efficacy of ADCs is yet to be further investigated (Wu et al., 2023b). The misdiagnosis of HER2low cases influences directly the prognostic and therapeutic implications of ADCs. Hence, urge the need for rigorous quality control procedures and well-defined guidelines to further educate pathologists for higher concordance in scoring (Sajjadi et al., 2022). Implementation and follow-up of strict standard operating procedures (SOPs), describing the diagnostic workflow from the specimen excision to HER2 report is pivotal (Cree et al., 2014). The invention and application of improved diagnostic methods that are

more precise and reproducible may lead to improved responses for a larger number of patients (Ivkovic-Kapic et al., 2019; Angerilli et al., 2021).

Novel complementary methodologies and future directions

Updates in current guidelines, and advances in methodologies for a more precise measurement could pave the way to ensure consistent and accurate HER2-low identification (Wu et al., 2023b). Clinical trials and patient recruitment are fundamental to improving diagnostic sensitivity for HER2-low breast cancer. In this respect, DESTINY-Breast06 (NCT04494425) is an ongoing phase 3, randomized, multicenter, open-label study on T-DXd vs. chemotherapy in HER2-low, hormone receptor-positive breast cancer patients with disease progression on endocrine therapy in the metastatic setting. In this trial HER2-low tumors (IHC2+/ISH-, IHC 1+, or IHC >0 < 1+) are determined by central laboratory testing results (Wu et al., 2023b). The findings of such trials could contribute to the understanding of clear-cut limits of HER2 expression required for ADC treatment strategy using the available diagnostic techniques (Wu et al., 2023b; Fan and Xu, 2023). Yet, to develop a robust definition of HER2-low, the number of recruited patients in such randomized clinical trials needs to be amplified (Atallah et al., 2022). Recruitment of further diagnostic approaches with a more precise cut-off could facilitate the detection of patients suitable for targeted therapies. For instance, Muotafi et al. developed an assay that is complementary to the conventional methods for the identification of HER2-low unamplified tumors through a combination of quantitative immunofluorescence and mass spectrometry (Moutafi et al., 2022). This method measured absolute amounts of HER2 protein on conventional histology sections where HER2-low expression ranged between 2 and 20 attomol/mm2. By applying this assay to 364 breast cancers, they found that 67% of the tumors had HER2 expression above the limit of quantification and below the level seen in HER2amplified breast cancer. However, this assay can distinguish between HER2 gene amplified from unamplified. To fully span the dynamic range of HER2 expression more than one assay is needed (Moutafi et al., 2022). Other studies have shown strong between multiple-reaction monitoring agreement mass spectrometry (MRM-MS) results and IHC/ISH, and also demonstrated a correlation between MRM-based measurements and clinical response to HER2-targeted therapy (Steiner et al., 2015; Nuciforo et al., 2016). Kennedy et al. used immunoaffinityenrichment coupled immuno-MRM-MS to quantify HER2 protein expression in 96 frozen and 119 FFPE breast cancer samples. By examining the agreement of HER2 normalized by glyceraldehyde-3phosphate dehydrogenase (GAPDH) (as a surrogate for tumor cellularity) to HER2 status, they found an excellent agreement between the GAPDH-normalized immuno-MRM-MS measurements of HER2 and HER2 status (IHC ± ISH) in both the FFPE (*p*-value = 1.09 e-16 by Mann-Whitney test) and frozen (p-value = 1.14 e-7 by Mann-Whitney test) tissues. In addition, the immuno-MRM-MS assay showed good sensitivity and specificity in both FFPE (AUC = 0.971) and frozen (AUC = 0.962) samples. Hence, the findings showed this method enables precise, relative quantification of HER2 in HER2-low and HER2-negative tumors (Kennedy et al., 2021). Other methodologies to improve HER2 assessment include machine learning approaches. Increasingly, pathological diagnoses are aided by artificial intelligence (AI), particularly in image processing and quantitative assessments (Koopman et al., 2019; Zakrzewski et al., 2019). Through AI, it is possible to create a computer algorithm that can analyze images, count the amount of HER2 membrane staining, and generate reliable and reproducible results (Koopman et al., 2019; Zakrzewski et al., 2019). Despite great achievements in assessing HER2-positive and -negative tumors, only a few studies have utilized AI for differentiating HER2 0 from 1 + tumors. Palm et al. evaluated an AI-assisted workflow that involved both IHC and ISH for determining the HER2 status of primary and metastatic breast cancer as well as assessing its performance on HER2-low breast tumors. For this, a preliminary cohort of primary breast carcinomas (n = 495), and a study cohort of 67 primary breast carcinomas and 30 metastatic deposits were evaluated for HER2 status by IHC and ISH. After having built the ground truth by practicing breast pathologists, the results of the AI digital image analysis were provided to them and the slides were reassessed. The findings showed that HER2 IHC and its interpretation were troubled by low inter-rater agreement, particularly in the 0 and 1 + range. The IHC/ISH combination was suitable for the detection of amplified and non-amplified HER2 tumors (Cohen's $\kappa = 0.94$) however limitations in the classification of HER2 low tumors were observed. While a combined IHC/ISH digitalized AI-assisted workflow for HER2 status determination in primary and metastatic breast cancer including the HER2 low tumors was feasible (Palm et al., 2023). Another recent research has reported of digital image analysis (DIA) efficacy compared to assisted reading (AR) in 761 tumors from 727 patients, IHC stained for HER2 (Sode et al., 2023). The authors reported moderate agreement of 73% (κ : 0. 55) between DIA and AR, being mostly discordant in cases with heterogeneous and aberrant staining, representing a major setback in the evaluation of HER2. Of note, pathologists evaluated fewer tumors as HER2-1+ and more in the HER2-0 and HER2-2+ categories compared with DIA when looking at the HER2-low category, mostly downgrading the HER2 score to HER2-0 compared to DIA HER2-1+. Thus, authors suggest that pathologist will remain the most significant factor in HER2 evaluation in the near future (Sode et al., 2023). A multiinstitutional study showed improved accuracy of HER2 0 and 1 + tumor interpretation when pathologists were assisted by AI compared to not being assisted (0.93 vs. 0.8). Importantly, the accuracy was significantly improved even with the presence of heterogeneity (0.68-0.89) (Wu et al., 2023a). Given the complications of HER2 evaluation in the presence of heterogeneity and the higher prevalence of HER2 heterogeneity in HER21 + and 2 + than in HER23 + tumors (Seol et al., 2012), this methodology could assist pathologists in a more precise HER2-low evaluation (Wu et al., 2023a). Gustavson et al. used deep learningbased image analysis to generate a novel HER2 Quantitative Continuous Score (QCS). The results showed a high correlation between QCS obtained from automatically detected membranes and those annotated by pathologists (R = 0.993). Moreover, HER2 QCS results had a broad quantitative overlap between IHC and ISH (Gustavson et al., 2021). The use of the artificial neural network

(ANN) models that can learn and shape non-linear and complex relationships has been implemented to assist in refining the HER2low definition (Atallah et al., 2022). Atallah et al., developed a model by implementing as input the HER2 scoring parameters (i.e., intensity and its distribution, completeness, and percentage of positive cells and cytoplasmic staining) and as an output the HER2 mRNA-based clusters variable. The ANN model configured HER2 IHC score 1 + as membranous staining in invasive cells either as faint intensity in \geq 20% of cells regardless of the circumferential completeness, weak complete staining in ≤10%, weak incomplete staining in >10% and moderate incomplete staining in \leq 10%. This model showed statistical significance with Oncotype DX scores and a perfect intra- and inter-observer agreement (kappa value = 0.8 and 0.9, respectively) (Atallah et al., 2022). Such methodologies have shown promising outcomes for a more accurate and reproducible HER2-low assessment compared to the currently available methods. Yet, further investigation to tailor these novel approaches as cost effective and easily accessible assets, as well as performing extensive analytic and clinical validation are necessary prior to introducing them in the real-life clinical practice. Consecutively, the significance of continuous education, updates, quality controls, and multidisciplinary discussions should not be underestimated for optimal patient management.

Discussion

To date, IHC-ISH combined test is the gold standard approach for assessing HER2 in breast cancer. However, this method suffers from high inter- and intra-observer variability, with the results being conditioned by subjective factors. Moreover, the heterogeneity of HER2 expression and/or amplification adds a layer of complexity to a precise and reproducible HER2-low assessment. To this end, the implementation of clear guidelines, careful supervision of preanalytical and analytical issues, and specialized training for accurate HER2 testing play a crucial role in this process. Moreover, the recruitment of novel diagnostic approaches with a more precise cut-off is essential to define patients who are eligible for targeted therapies specifically ADCs in HER2-low tumors. Among alternative methods, AI and machine learning-based predictors have shown promising results in terms of speed, accuracy, and costeffectiveness and are likely to play an increasingly important role in this field in the future. It is important to note that AI algorithms are not yet widely used in the clinical setting for the identification of HER2-low breast cancer, and further research is required to validate their accuracy and reliability. However, the potential benefits of AI in this area are significant, and there is ongoing work to develop and refine these algorithms to improve patient

References

Ahn, S., Woo, J. W., Lee, K., and Park, S. Y. (2020). HER2 status in breast cancer: Changes in guidelines and complicating factors for interpretation. *J. pathology Transl. Med.* 54 (1), 34–44. doi:10.4132/jptm.2019.11.03

Angerilli, V., Galuppini, F., Pagni, F., Fusco, N., Malapelle, U., and Fassan, M. (2021). The role of the pathologist in the next-generation era of tumor molecular characterization. *Diagn. (Basel)* 11 (2), 339. doi:10.3390/diagnostics11020339

outcomes. Introducing such novel approaches in the real-life clinical practice requires extensive analytic and clinical validation. Further investigation including multidisciplinary clinical studies on identifying this fairly large group of patients through novel methodologies with the implementation of strict guidelines, is warranted.

Author contributions

ES wrote the initial version of the manuscript; NF supervised and revised the paper; EG-R, EC, OP, GM, KV, and MI provided a critical revision of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding

This work was partially supported by the Italian Ministry of Health with Ricerca Corrente 5 x 1,000 funds. MI was supported by Fondazione Umberto Veronesi; KV was supported by Fondazione IEO—MONZINO.

Conflict of interest

NF has received honoraria for consulting, advisory role, speaker bureau, travel, and/or research grants from MSD, Boehringer Ingelheim, Novartis, Roche, AstraZeneca, Daiichi Sankyo, GlaxoSmithKline (GSK), Gilead, Diaceutics, Adicet Bio, and Sermonix. EGR has received honoraria for consulting, advisory role, speaker bureau, travel, and/or research grants from Thermo Fisher Scientific, Novartis, AstraZeneca, Roche, Biocartis, Exact Science, GSK, and Illumina.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Atallah, N. M., Toss, M. S., Green, A. R., Mongan, N. P., Ball, G., and Rakha, E. A. (2022). Refining the definition of HER2-low class in invasive breast cancer. *Histopathology* 81 (6), 770–785. doi:10.1111/his.14780

Banerji, U., van Herpen, C. M. L., Saura, C., Thistlethwaite, F., Lord, S., Moreno, V., et al. (2019). Trastuzumab duocarmazine in locally advanced and metastatic solid tumours and HER2-expressing breast cancer: A phase 1 dose-escalation and dose-expansion study. *Lancet Oncol.* 20 (8), 1124–1135. doi:10.1016/S1470-2045(19)30328-6

Bianchi, S., Caini, S., Paglierani, M., Saieva, C., Vezzosi, V., Baroni, G., et al. (2015). Accuracy and reproducibility of HER2 status in breast cancer using immunohistochemistry: A quality control study in tuscany evaluating the impact of updated 2013 ASCO/CAP recommendations. *Pathol. Oncol. Res.* 21 (2), 477–485. doi:10.1007/s12253-014-9852-0

Brunelli, M., Manfrin, E., Martignoni, G., Miller, K., Remo, A., Reghellin, D., et al. (2009). Genotypic intratumoral heterogeneity in breast carcinoma with HER2/neu amplification: Evaluation according to ASCO/CAP criteria. *Am. J. Clin. Pathol.* 131 (5), 678–682. doi:10.1309/AJCP09VUTZWZXBMJ

Cree, I. A., Deans, Z., Ligtenberg, M. J., Normanno, N., Edsjö, A., Rouleau, E., et al. (2014). Guidance for laboratories performing molecular pathology for cancer patients. *J. Clin. Pathol.* 67 (11), 923–931. doi:10.1136/jclinpath-2014-202404

Fan, P., and Xu, K. (2023). Antibody-drug conjugates in breast cancer: Marching from HER2-overexpression into HER2-low. *Biochimica Biophysica Acta (BBA) - Rev. Cancer* 1878 (1), 188849. doi:10.1016/j.bbcan.2022.188849

Fernandez, A. I., Liu, M., Bellizzi, A., Brock, J., Fadare, O., Hanley, K., et al. (2022). Examination of low ERBB2 protein expression in breast cancer tissue. *JAMA Oncol.* 8 (4), 1–4. doi:10.1001/jamaoncol.2021.7239

Fusco, N., Rizzo, A., Costarelli, L., Santinelli, A., Cerbelli, B., Scatena, C., et al. (2022). Pathological examination of breast cancer samples before and after neoadjuvant therapy: Recommendations from the Italian group for the study of breast pathology Italian society of pathology (GIPaM-SIAPeC). *Pathologica* 114 (2), 104–110. doi:10. 32074/1591-951X-747

Gustavson, M., Haneder, S., Spitzmueller, A., Kapil, A., Schneider, K., Cecchi, F., et al. (2021). Abstract PD6-01: Novel approach to HER2 quantification: Digital pathology coupled with AI-based image and data analysis delivers objective and quantitative HER2 expression analysis for enrichment of responders to trastuzumab deruxtecan (T-DX4; DS-8201), specifically in HER2-low patients. *Cancer Res.* 81, PD6–01. doi:10. 1158/1538-7445.sabcs20-pd6-01

Ivkovic-Kapic, T., Knezevic Usaj, S., Moldvaji, E., Jovanic, I., Milovanovic, Z., Milentijevic, M., et al. (2019). Interlaboratory concordance in HER2 testing: Results of a Serbian ring-study. *J. buon* 24 (3), 1045–1053.

Jensen, S. G., Thomas, P. E., Christensen, I. J., Balslev, E., Hansen, A., and Høgdall, E. (2020). Evaluation of analytical accuracy of HER2 status in patients with breast cancer: Comparison of HER2 GPA with HER2 IHC and HER2 FISH. *Apmis* 128 (11), 573–582. doi:10.1111/apm.13076

Kennedy, J. J., Whiteaker, J. R., Kennedy, L. C., Bosch, D. E., Lerch, M. L., Schoenherr, R. M., et al. (2021). Quantification of human epidermal growth factor receptor 2 by immunopeptide enrichment and targeted mass spectrometry in formalin-fixed paraffinembedded and frozen breast cancer tissues. *Clin. Chem.* 67 (7), 1008–1018. doi:10.1093/ clinchem/hvab047

Koopman, T., Buikema, H. J., Hollema, H., de Bock, G. H., and van der Vegt, B. (2019). What is the added value of digital image analysis of HER2 immunohistochemistry in breast cancer in clinical practice? A study with multiple platforms. *Histopathology* 74 (6), 917–924. doi:10.1111/his.13812

Modi, S., Jacot, W., Yamashita, T., Sohn, J., Vidal, M., Tokunaga, E., et al. (2022). Trastuzumab deruxtecan in previously treated HER2-low advanced breast cancer. *N. Engl. J. Med.* 387 (1), 9–20. doi:10.1056/ NEJMoa2203690

Moutafi, M., Robbins, C. J., Yaghoobi, V., Fernandez, A. I., Martinez-Morilla, S., Xirou, V., et al. (2022). Quantitative measurement of HER2 expression to subclassify ERBB2 unamplified breast cancer. *Lab. Investig.* 102 (10), 1101–1108. doi:10.1038/ s41374-022-00804-9

Nuciforo, P., Thyparambil, S., Aura, C., Garrido-Castro, A., Vilaro, M., Peg, V., et al. (2016). High HER2 protein levels correlate with increased survival in breast cancer patients treated with anti-HER2 therapy. *Mol. Oncol.* 10 (1), 138–147. doi:10.1016/j. molonc.2015.09.002

Palm, C., Connolly, C. E., Masser, R., Padberg Sgier, B., Karamitopoulou, E., Simon, Q., et al. (2023). Determining HER2 status by artificial intelligence: An investigation of primary, metastatic, and HER2 low breast tumors. *Diagn. [Internet]* 13 (1), 168. doi:10. 3390/diagnostics13010168

Perez, E. A., Cortés, J., Gonzalez-Angulo, A. M., and Bartlett, J. M. S. (2014). HER2 testing: Current status and future directions. *Cancer Treat. Rev.* 40 (2), 276–284. doi:10.1016/j.ctrv.2013.09.001 Pondé, N., Brandão, M., El-Hachem, G., Werbrouck, E., and Piccart, M. (2018). Treatment of advanced HER2-positive breast cancer: 2018 and beyond. *Cancer Treat. Rev.* 67, 10–20. doi:10.1016/j.ctrv.2018.04.016

Sajjadi, E., Venetis, K., Ivanova, M., and Fusco, N. (2022). Improving HER2 testing reproducibility in HER2-low breast cancer. *Cancer Drug Resist* 5 (4), 882–888. doi:10. 20517/cdr.2022.29

Sapino, A., Goia, M., Recupero, D., and Marchiò, C. (2013). Current challenges for HER2 testing in diagnostic pathology: State of the art and controversial issues. *Front. Oncol.* 3, 129. doi:10.3389/fonc.2013.00129

Schrohl, A. S., Pedersen, H. C., Jensen, S. S., Nielsen, S. L., and Brünner, N. (2011). Human epidermal growth factor receptor 2 (HER2) immunoreactivity: Specificity of three pharmacodiagnostic antibodies. *Histopathology* 59 (5), 975–983. doi:10.1111/j. 1365-2559.2011.04034.x

Seol, H., Lee, H. J., Choi, Y., Lee, H. E., Kim, Y. J., Kim, J. H., et al. (2012). Intratumoral heterogeneity of HER2 gene amplification in breast cancer: Its clinicopathological significance. *Mod. Pathol.* 25 (7), 938–948. doi:10.1038/modpathol.2012.36

Sode, M., Thagaard, J., Eriksen, J. O., and Laenkholm, A-V. (2023). Digital image analysis and assisted reading of the HER2 score display reduced concordance: Pitfalls in the categorisation of HER2-low breast cancer. *Histopathology* 2023. doi:10.1111/his. 14877

Steiner, C., Tille, J. C., Lamerz, J., Kux van Geijtenbeek, S., McKee, T. A., Venturi, M., et al. (2015). Quantification of HER2 by targeted mass spectrometry in formalin-fixed paraffin-embedded (FFPE) breast cancer tissues. *Mol. Cell Proteomics* 14 (10), 2786–2799. doi:10.1074/mcp.O115.049049

Tarantino, P., Hamilton, E., Tolaney, S. M., Cortes, J., Morganti, S., Ferraro, E., et al. (2020). HER2-Low breast cancer: Pathological and clinical landscape. *J. Clin. Oncol.* 38 (17), 1951–1962. doi:10.1200/JCO.19.02488

Venetis, K., Crimini, E., Sajjadi, E., Corti, C., Guerini-Rocco, E., Viale, G., et al. (2022). HER2 low, ultra-low, and novel complementary biomarkers: Expanding the spectrum of HER2 positivity in breast cancer. *Front. Mol. Biosci.* 9, 834651. doi:10.3389/fmolb.2022. 834651

Viale, G., and Fusco, N. (2022). Pathology after neoadjuvant treatment - how to assess residual disease. *Breast* 62, S25–S28. doi:10.1016/j.breast.2021.11.009

Wolff, A. C., Hammond, M. E. H., Allison, K. H., Harvey, B. E., Mangu, P. B., Bartlett, J. M. S., et al. (2018). Human epidermal growth factor receptor 2 testing in breast cancer: American society of clinical oncology/college of American pathologists clinical practice guideline focused update. J. Clin. Oncol. 36 (20), 2105–2122. doi:10.1200/JCO.2018.77.8738

Wu, S., Yue, M., Zhang, J., Li, X., Li, Z., Zhang, H., et al. (2023). The role of artificial intelligence in accurate interpretation of HER2 immunohistochemical scores 0 and 1+ in breast cancer. *Mod. Pathol.* 36 (3), 100054. doi:10.1016/j.modpat.2022.100054

Wu, Y., Zhong, R., and Ma, F. (2023). HER2-low breast cancer: Novel detections and treatment advances. *Crit. Rev. Oncol. Hematol.* 181, 103883. doi:10.1016/j.critrevonc. 2022.103883

Yue, M., Zhang, J., Wang, X., Yan, K., Cai, L., Tian, K., et al. (2021). Can AI-assisted microscope facilitate breast HER2 interpretation? A multi-institutional ring study. *Virchows Arch.* 479 (3), 443–449. doi:10.1007/s00428-021-03154-x

Zakrzewski, F., de Back, W., Weigert, M., Wenke, T., Zeugner, S., Mantey, R., et al. (2019). Automated detection of the HER2 gene amplification status in Fluorescence *in situ* hybridization images for the diagnostics of cancer tissues. *Sci. Rep.* 9 (1), 8231. doi:10.1038/s41598-019-44643-z

Zhang, H., Karakas, C., Tyburski, H., Turner, B. M., Peng, Y., Wang, X., et al. (2022). HER2-low breast cancers: Current insights and future directions. *Seminars Diagnostic Pathology* 39 (5), 305–312. doi:10.1053/j.semdp.2022.07.003

Zhang, H., Katerji, H., Turner, B. M., and Hicks, D. G. (2021). HER2-Low breast cancers: New opportunities and challenges. *Am. J. Clin. Pathology* 157, 328. doi:10.1093/ajcp/aqab117

Zhang, H., Moisini, I., Ajabnoor, R. M., Turner, B. M., and Hicks, D. G. (2020). Applying the new guidelines of HER2 testing in breast cancer. *Curr. Oncol. Rep.* 22 (5), 51. doi:10.1007/s11912-020-0901-4

Zhang, H., and Peng, Y. (2022). Current biological, pathological and clinical landscape of HER2-low breast cancer. *Cancers (Basel)* 15 (1), 126. doi:10.3390/ cancers15010126