Check for updates

OPEN ACCESS

EDITED BY George Tsiamis, University of Patras, Greece

REVIEWED BY Liang Wang, Guangdong Provincial People's Hospital, China Oumaima Zidi, University of Manouba, Tunisia

*CORRESPONDENCE Heng Yang ⊠ johnnelyang@hotmail.com

¹These authors have contributed equally to this work

SPECIALTY SECTION This article was submitted to Systems Microbiology, a section of the journal Frontiers in Microbiology

RECEIVED 03 December 2022 ACCEPTED 09 February 2023 PUBLISHED 27 February 2023

CITATION

Zeng Y, Cao S and Yang H (2023) Roles of gut microbiome in epilepsy risk: A Mendelian randomization study. *Front. Microbiol.* 14:1115014. doi: 10.3389/fmicb.2023.1115014

COPYRIGHT

© 2023 Zeng, Cao and Yang. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Roles of gut microbiome in epilepsy risk: A Mendelian randomization study

Youjie Zeng^{1†}, Si Cao^{1†} and Heng Yang^{2*}

¹Department of Anesthesiology, Third Xiangya Hospital, Central South University, Changsha, Hunan, China, ²Department of Neurology, Third Xiangya Hospital, Central South University, Changsha, Hunan, China

Background: Recent studies have suggested an association between gut microbiomes (GMs) and epilepsy. However, the GM taxa identified in different studies are variable. In addition, observational studies cannot indicate causality. Therefore, our study aimed to explore the causal association of GMs with epilepsy and identify the most influential GM taxa.

Methods: We conducted a Mendelian randomization (MR) study using summary statistics from genome-wide association studies (GWAS) of 211 GM taxa and epilepsy. The GWAS summary statistics for 211 GM taxa (from phylum to genus level) were generated by the MiBioGen consortium, while the FinnGen consortium provided the GWAS summary statistics for epilepsy. The primary analytical method to assess causality was the inverse-variance weighted (IVW) approach. To complement the IVW method, we also applied four additional MR methods: MR-Egger, weighted median, simple mode, and weighted. In addition, we conducted sensitivity analyses using Cochrane's *Q*-test, MR-Egger intercept test, MR-PRESSO global test, and leave-one-out analysis.

Results: We evaluated the causal effect of 211 GM taxa (from phylum to genus level) on epilepsy, generalized epilepsy, and focal epilepsy. After using the Bonferroni method for multiple testing correction, *Class Betaproteobacteria* [odds ratio (OR)=1.357, 95% confidence interval (CI): 1.126-1.635, p=0.001] and *Order Burkholderiales* (OR=1.336, 95% CI: 1.112-1.606, p=0.002). In addition, 21 nominally significant causal relationships were also identified. Further, the MR-Egger intercept test and MR-PRESSO global test suggested that our MR analysis was unaffected by horizontal pleiotropy (p>0.05). Finally, the leave-one-out analysis suggested the robustness of the results.

Conclusion: Through the MR study, we analyzed the causal relationship of 211 GM taxa with epilepsy and determined the specific intestinal flora associated with increased epilepsy risk. Our findings may provide helpful biomarkers for disease progression and potential candidate therapeutic targets for epilepsy. In addition, in-depth analysis of large-scale microbiome GWAS datasets based on metagenomics sequencing is necessary for future studies.

KEYWORDS

gut microbiome, epilepsy, causal relationship, incidence risk, Mendelian randomization, MiBioGen, FinnGen, microbiota-gut-brain axis

1. Introduction

Epilepsy is a common, chronic neurological disorder characterized by sudden abnormal excessive ultra-synchronized neuron discharges that result in temporary involuntary brain dysfunction (Fisher et al., 2005). Globally, there are 70 million people with epilepsy, with the highest incidence in infants and the elderly, posing a tremendous social burden throughout the world (Collaborators, 2019; Thijs et al., 2019). Despite advances and innovations in antiepileptic medications, approximately one-third of the patients suffer from drug-resistant epilepsy (de Biase et al., 2019). Therefore, further insights into the pathogenesis and the exploration of novel therapeutic targets for epilepsy are required.

There is growing evidence that the gut microbiome (GM) can regulate host homeostasis, including cardiovascular function, metabolism, and immune/inflammatory response (Le Chatelier et al., 2013). Recent research has shown that GMs play a role in neuropsychiatric disorders (Iannone et al., 2019), as they regulate brain function and behavior via the microbiota-gut-brain axis (Johnson and Foster, 2018). Differences in GM taxa have been identified in epilepsy patients compared to controls (Dong et al., 2022). The ketogenic diet (KD) is a treatment approach for intractable epilepsy (D'Andrea Meira et al., 2019). During the KD treatment of drug-resistant epilepsy, the GM pattern was altered simultaneously (Lindefeldt et al., 2019). Consequently, GMs may be involved in the crosstalk between KD and epilepsy (Fan et al., 2019). Furthermore, researchers are investigating the possibility of using the change in GM composition as a surrogate marker for the efficacy of the KD treatment in patients with drug-resistant epilepsy (Thambi et al., 2020). However, the effect of various GM taxa on epilepsy has not yet been determined. The 16S rRNA and metagenomic sequencing are the most widely used methods for identifying GM taxonomic characteristics (Durazzi et al., 2021), providing the basis for identifying the potential role of GM taxa. Recent research has increasingly focused on the causal effects of GMs on epilepsy, particularly refractory epilepsy (Lum et al., 2020). In addition, perturbations for certain GM taxa levels have been reported to affect the activity of epileptic neurons (Darch and McCafferty, 2022).

Nevertheless, the specific contribution of various GM taxa to epilepsy warrants further exploration. Similar to randomized controlled trials (RCT), the Mendelian randomization (MR) study is a novel research method for exploring the causal association between exposure and outcome (Swanson et al., 2017). In MR studies, single nucleotide polymorphisms (SNPs) are considered instrumental variables (IVs) to estimate the causal association between exposures and the outcomes of interest (Burgess et al., 2017). SNPs conform to the principle of random assignment of genetic variants at meiosis, which avoids the effect of confounding factors and the potential impact of reverse causation since genetic variants precede the onset of disease (Lawlor et al., 2008). Therefore, the causal associations of exposure factors of interest to outcomes can be identified more rapidly by MR analysis compared to RCT. For example, a recent MR study by Cai et al. has identified several blood metabolites with potential causal associations with epilepsy (Cai et al., 2022). Here, we conducted an MR study using large-scale GWAS summary statistics of GMs and epilepsy to identify potentially influential GM taxa, which could provide confidence to some existing evidence and may yield new insights into the prevention and treatment of epilepsy.

2. Materials and methods

2.1. Study design

The overall flow chart of this study is shown in Figure 1. MR studies are required to satisfy the following three assumptions: (i) IVs are strongly associated with exposure factors, (ii) IVs are independent of confounding factors, and (iii) IVs are solely associated with outcomes through exposure factors (Burgess et al., 2017). Specifically, we identified GM taxa that have a causal effect on epilepsy, generalized epilepsy, and focal epilepsy by performing a two-sample MR analysis. Our results were reported in accordance with the STROBE-MR guidelines (Skrivankova et al., 2021).

2.2. Data sources for the exposure

A study from the MiBioGen consortium analyzed the host genotypes and 16S fecal microbiomes rRNA gene sequencing profiles of 18,340 participants (Kurilshikov et al., 2021). This GWAS study examined 211 GM taxa (from genus to phylum level) and ultimately identified genetic variants associated with nine phyla, 16 classes, 20 orders, 35 families, and 131 genera. The GWAS summary statistics of GMs are available for download at¹ (Swertz and Jansen, 2007; Swertz et al., 2010; van der Velde et al., 2019).

2.3. Data sources for the outcome

We obtained GWAS summary statistics for epilepsy from the FinnGen consortium R7 release² (Kurki et al., 2022). In addition, we downloaded GWAS summary data for generalized epilepsy and focal epilepsy. Epilepsy diagnosis in FinnGen was based on G40 in the 10th version of the International Classification of Diseases (ICD). Cases of generalized and focal epilepsy were narrower endpoints of epilepsy under the strict definition. Table 1 shows the details of the exposure and outcome analyzed in this MR study.

2.4. Identification of IVs

SNPs strongly associated with each GM taxon were used as IVs in this MR study. Since the number of IVs obtained under the strict threshold ($p < 5 \times 10^{-8}$) was extremely minimal, we adopted a more comprehensive threshold ($p < 1 \times 10^{-5}$) to obtain relatively more IVs to achieve relatively robust results. In addition, to ensure each IV's independence, SNPs within a window size of 10,000 kb at a threshold of $r^2 < 0.001$ were pruned to mitigate linkage disequilibrium (LD). Then, palindromic SNPs and SNPs not present in the outcome were removed from the IVs. Finally, we calculated the F-statistic of IVs to assess the degree of weak instrumental bias. If the F-statistic >10, it was considered that no bias was caused by weak IVs (Pierce et al., 2011).

¹ https://mibiogen.gcc.rug.nl/

² https://r7.finngen.fi/



TABLE 1 Details of the exposure and outcome.

Trait	Consortium	Samples	Case	Control			
Exposure							
211 GM taxa	MiBioGen	18,340	1	/			
Outcome							
Epilepsy	FinnGen (R7)	252,026	8,523	243,503			
Generalized epilepsy	FinnGen (R7)	302,828	2,197	300,631			
Focal epilepsy	FinnGen (R7)	301,493	862	300,631			

GM, gut microbiome.



(A) Forest plot of GM taxa associated with epilepsy identified by IVW_FE method. (B) Forest plot of GM taxa associated with generalized epilepsy identified by IVW_FE method. (C) Forest plot of GM taxa associated with focal epilepsy identified by IVW_FE method.

2.5. Statistical methods

The inverse variance weighted fixed-effect (IVW-FE) method or the IVW random effect (IVW-RE) method was used as the primary MR method for inferring causality. The choice of IVW-FE or IVW-RE was determined based on Cochrane's Q heterogeneity test. The IVW method is an extension of the Wald ratio estimator based on the principles of Meta-analysis (Pagoni et al., 2019).

For each GM taxon, if the IVW method identified a causal association (p < 0.05), four additional MR methods, MR-Egger, weighted median, simple mode, and weighted mode, would be performed to supplement the IVW result (Bowden et al., 2016; Burgess and Thompson, 2017). The criterion for using the weighted median method is that at least 50% of the SNPs must satisfy the premise that they are valid IVs (Bowden et al., 2016). The MR-Egger method provides unbiased estimates even when all selected IVs are

Exposure	Method		OR (95% CI)	P-value
Class				
Betaproteobacteria	IVW (FE)	+I	1.357 (1.126, 1.635)	0.001
ID: 2867)	IVW (RE)		1.357 (1.126, 1.635)	0.001
	MR Egger		1.378 (0.763, 2.486)	0.319
	Simple mode	·····	1.319 (0.852, 2.042)	0.245
	Weighted median	······	1.332 (1.025, 1.730)	0.032
	Weighted mode	k	1.352 (0.899, 2.031)	0.181
verrucomicrobiae	IVW (FE)	· · · · · · · · · · · · · · · · · · ·	1.184 (1.021, 1.372)	0.025
ID: 4029)	IVW (BE)	·····	1 184 (1 003 1 397)	0.046
	MB Egger		1 833 (1 102 3 048)	0.044
	Simple mode	••••••••	1 325 (0.030 1 888)	0.150
	Weighted modian	· · · · · · · · · · · · · · · · · · ·	1 160 (0.053, 1.434)	0.135
	Weighted median		1.225 (0.055, 1.434)	0.133
.	weighted mode	· · ·	1.525 (0.955, 1.859)	0.125
Jraer	NAV (PP)		1226 (1112-1700)	0.002
Surkholderlaies	IVW (FE)		1.336 (1.112, 1.606)	0.002
ID: 2874)	IVW (RE)		1.336 (1.112, 1.606)	0.002
	MR Egger		1.362 (0.761, 2.438)	0.329
	Simple mode		1.263 (0.845, 1.886)	0.284
	Weighted median	••••••	1.283 (0.998, 1.650)	0.052
	Weighted mode	I	1.277 (0.863, 1.889)	0.253
rrucomicrobiales	IVW (FE)	[!·····]	1.184 (1.021, 1.372)	0.025
ID: 4030)	IVW (RE)	••••••	1.184 (1.003, 1.397)	0.046
	MR Egger	H	1.833 (1.102, 3.048)	0.044
	Simple mode	F	1.325 (0.924, 1.900)	0.157
	Weighted median	······•	1.169 (0.945, 1.446)	0.151
	Weighted mode		1.325 (0.926 1.895)	0 154
amily	ineighted hidde		Tible (dibled, 1050)	01101
Verrugomiarobiaceae	IVW (FE)	······	1 184 (1 021 1 272)	0.025
ID: 4026)	IVW (PE)		1.184 (1.021, 1.372)	0.025
1D. 4030)	IV W (RE)	· · · · · · · · · · · · · · · · · · ·	1.164 (1.005, 1.597)	0.040
	MR Egger		1.833 (1.102, 3.048)	0.044
	Simple mode		1.325 (0.924, 1.900)	0.157
	Weighted median		1.169 (0.951, 1.436)	0.137
	Weighted mode	· · · · · · · · · · · · · · · · · · ·	1.325 (0.944, 1.859)	0.135
Genus				
Subacterium Xylanophilus Group	IVW (FE)		0.816 (0.694, 0.959)	0.014
ID: 14375)	IVW (RE)		0.816 (0.688, 0.967)	0.019
	MR Egger		0.851 (0.495, 1.465)	0.579
	Simple mode	••••••••••••••••••••••••••••••••••••••	0.747 (0.517, 1.078)	0.157
	Weighted median	••••••	0.786 (0.629, 0.982)	0.034
	Weighted mode	·····•	0.734 (0.508, 1.059)	0.136
Akkermansia	IVW (FE)		1.184 (1.021, 1.372)	0.025
ID: 4037)	IVW (RE)	·····•	1.184 (1.003, 1.397)	0.046
	MR Egger	+•	1.834 (1.104, 3.047)	0.044
	Simple mode	·······	1.324 (0.947, 1.850)	0.132
	Weighted median	······•	1.169 (0.944, 1.447)	0.153
	Weighted mode		1.324 (0.932, 1.879)	0.148
Anaerotruncus	IVW (FF)		1.238 (1.050, 1.459)	0.011
ID: 2054)	IVW (RF)	 	1,238 (1,050, 1,459)	0.011
	MR Egger		1 162 (0 710 1 877)	0.553
	Simple mode		1.02 (0.717, 1.077)	0.555
	Waighted mode	· · · · · · · · · · · · · · · · · · ·	1.092 (0.773, 1.340)	0.023
	weighted median		1.157 (0.929, 1.440)	0.193
	Weighted mode	······	1.109 (0.791, 1.555)	0.560
cuminococcaceae UCG014	IVW (FE)		1.178 (1.016, 1.367)	0.030
ID: 11371)	IVW (RE)		1.178 (1.016, 1.367)	0.030
	MR Egger	H	1.316 (0.920, 1.881)	0.167
	Simple mode	······	1.119 (0.806, 1.553)	0.518
	Weighted median	I	1.124 (0.910, 1.388)	0.280
	Weighted mode	ŀ····	1.115 (0.876, 1.420)	0.396
Jnknown genus	IVW (FE)	I+++••+++	0.819 (0.715, 0.939)	0.004
ID: 826)	IVW (RE)	h	0.819 (0.690, 0.973)	0.023
	MR Egger		0.882 (0.527, 1.476)	0.641
	Simple mode	J	0.988 (0.734 1.332)	0.940
	Weighted median		0.935 (0.751, 1.140)	0.520
	Weighted median	· · · · · · · · · · · · · · · · · · ·	0.733 (0.701, 1.149)	0.020
	weighted mode		0.979 (0.742, 1.292)	0.000
	0.0	0.2 0.4 0.6 0.8 1.0 1.2 1.4 1.6 1.8 2.0 2.2 2.4 2. Odds Ratio(95%CI)	o 2.8 3.0	
IDE 7				

multivariate (Burgess and Thompson, 2017). Finally, the results of causal associations were presented as odds ratios (OR) and 95% confidence intervals (95% CI). The significance threshold was set at p < 0.05. In addition, the Bonferroni method was used for multiple testing corrections. The threshold for various levels was p < 0.05/n, where n represents the number of taxa at a particular level.

Only exposure-outcome pairs with the same direction identified by all MR methods were considered to have a causal association. To test the stability of the causal association, we further performed several sensitivity analyses. First, the MR-Egger intercept test and MR-PRESSO global test were utilized to detect horizontal pleiotropy (Rees et al., 2017; Verbanck et al., 2018). In addition, the leave-one-out analysis was performed to assess the robustness of the results. Furthermore, we performed replicated MR analyses after excluding potential confounders from the IVs. Specifically, the confoundersrelated SNPs were retrieved from the PhenoScanner V2 database³ (Staley et al., 2016; Kamat et al., 2019), including education level (Wang et al., 2021), diabetes (Marcovecchio et al., 2015), obesity (Hafizi et al., 2017), and smoking (Yuan et al., 2021).

All analyses in this study were performed based on R software(version 4.2.1). The "TwoSampleMR" R package⁴ and the "MRPRESSO" R package⁵ were used in our MR study.

³ http://www.phenoscanner.medschl.cam.ac.uk/

⁴ https://mrcieu.github.io/TwoSampleMR/

⁵ https://github.com/rondolab/MR-PRESSO



IGURE 4

Diverse MR results for 5 GM taxa causally associated with generalized epilepsy.

Exposure	Method			OR (95% CI)	P-value
hylum					
errucomicrobia	IVW (FE)		II	1.697 (1.101, 2.616)	0.017
D: 3982)	IVW (RE)			1.697 (1.094, 2.633)	0.018
	MR Egger	+	••••••	1.332 (0.401, 4.424)	0.650
	Simple mode	+-	•••••••••••••••••••••••••••••••••••••••	2.421 (0.849, 6.909)	0.127
	Weighted median		kk	2.045 (1.116, 3.749)	0.021
	Weighted mode		ļI	2.371 (0.988, 5.692)	0.079
ass					
errucomicrobiae	IVW (FE)			1.740 (1.103, 2.745)	0.017
D: 4029)	IVW (RE)			1.740 (1.071, 2.827)	0.025
	MR Egger	ŀ	•	1.081 (0.196, 5.947)	0.931
	Simple mode		•	3.109 (0.973, 9.928)	0.085
	Weighted median			2.169 (1.149, 4.092)	0.017
	Weighted mode			3.011 (1.072, 8.456)	0.063
rder					
rrucomicrobiales	IVW (FE)		······	1.740 (1.103, 2.745)	0.017
D: 4030)	IVW (RE)		••	1.740 (1.071, 2.827)	0.025
	MR Egger	ŀ	•	1.081 (0.196, 5.947)	0.931
	Simple mode		ç	3.109 (0.977, 9.895)	0.084
	Weighted median			2.169 (1.148, 4.098)	0.017
	Weighted mode)I	3.011 (1.019, 8.899)	0.074
amily	-				
errucomicrobiaceae	IVW (FE)		·····•	1.739 (1.102, 2.745)	0.017
D: 4036)	IVW (RE)			1.739 (1.070, 2.827)	0.025
	MR Egger	ŀ	•	1.078 (0.196, 5.930)	0.933
	Simple mode			3.115 (1.011, 9.601)	0.076
	Weighted median		······	2.165 (1.117, 4.195)	0.022
	Weighted mode		•	3.018 (0.999, 9.111)	0.079
nus				,	
kkermansia	IVW (FE)			1.739 (1.102, 2.744)	0.017
0: 4037)	IVW (RE)			1.739 (1.070, 2.826)	0.025
	MR Egger	ŀ	•	1.079 (0.196, 5.928)	0.933
	Simple mode			3 116 (0.938, 10.351)	0.093
	Weighted median			2 174 (1 136 4 158)	0.019
	Weighted mode		· · · · · · · · · · · · · · · · · · ·	2.987 (0.954, 9.355)	0.090
loprevotella	IVW (FF)		•••••••	1 499 (1 052 2 137)	0.025
D: 961)	IVW (RF)			1.499 (1.052, 2.137)	0.025
,	MR Egger		•	2 478 (0 114 53 708)	0.604
	Simple mode		· · · · · · · · · · · · · · · · · · ·	1 707 (0.941 3 096)	0.153
	Weighted median			1 652 (1 051 2 595)	0.029
	Weighted mode		P	1 707 (0 957, 3 045)	0.144
itterella	IVW (FF)	···•		0 534 (0 310 0 919)	0.024
D: 2896)	IVW (RE)	···•	· 	0 534 (0 267, 1 065)	0.024
	MR Egger	J		0.150 (0.033, 0.692)	0.075
	Simple mode		M	0.307 (0.039, 0.052)	0.110
	Weighted median	. <u>.</u>	1	0.440 (0.205, 0.084)	0.046
	Weighted mode	. <u>-</u>	, ,	0.337 (0.116, 0.981)	0.040
petridium Saneu Stricto I	IVW (FE)		۲ ۱۰۰۰۰۰۰۰۰ <mark>۵</mark> ۰۰۰۰۰۰۰۰	1.662 (1.038, 2.661)	0.095
OSUTURIUM SENSU SUFICIO I	IVW (FE)			1.002 (1.038, 2.001)	0.035
J: 1873)	IVW (KE)	1.	, ,	1.002 (1.038, 2.001)	0.035
	MK Egger	1		0.949 (0.908, 53.195)	0.091
	Simple mode	1		2.703 (0.839, 8.719)	0.124
	Weighted median	I-		1.718 (0.893, 3.305)	0.105

FIGURE 5

Diverse MR results for 8 GM taxa causally associated with focal epilepsy.

Epilepsy	Generalized epilepsy	Focal epilepsy		
			Actinobacteria (ID: 400)	Phylum
			Verrucomicrobia (ID: 3982)	Class
			Bacilli (ID: 1673)	Order
			Betaproteobacteria (ID: 2867)	Famliy
			Verrucomicrobiae (ID: 4029)	Genus
			Burkholderiales (ID: 2874)	
			Verrucomicrobiales (ID: 4030)	
			Verrucomicrobiaceae (ID: 4036)	P<0.01
			Akkermansia (ID: 4037)	P<0.05
			Alloprevotella (ID: 961)	NS
			Anaerotruncus (ID: 2054)	P<0.05
			Bifidobacterium (ID: 436)	P<0.01
			Clostridium Sensu Stricto 1 (ID: 1873)	
			Coprobacter (ID: 949)	
			Eubacterium Xylanophilus Group (ID: 14	1375)
			Ruminococcaceae UCG014 (ID: 11371)	
			Sutterella (ID: 2896)	
			Unknown genus (ID: 826)	

FIGURE 6

Heat map of GM taxa causally associated with epilepsy, generalized epilepsy, and focal epilepsy identified by IVW method. Red represents risk factors, while blue represents protective factors.

FABLE 2 Horizontal pleiot	opy analysis for IVs	of 10 GM taxa	associated with e	pilepsy.
---------------------------	----------------------	---------------	-------------------	----------

Exposure	MR-Egger intercept test			MR-PRESSO global test	
	Egger_ intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value
Class					
Betaproteobacteria (ID: 2867)	-0.001	0.021	0.959	9.543	0.578
Verrucomicrobiae (ID: 4029)	-0.036	0.020	0.111	15.497	0.266
Order					
Burkholderiales (ID: 2874)	-0.001	0.021	0.948	9.840	0.551
Verrucomicrobiales (ID: 4030)	-0.036	0.020	0.111	15.497	0.271
Family					
Verrucomicrobiaceae (ID: 4036)	-0.036	0.020	0.112	15.499	0.274
Genus					
Eubacterium Xylanophilus Group(ID: 14375)	-0.004	0.023	0.876	11.147	0.433
Akkermansia (ID: 4037)	-0.036	0.020	0.111	15.484	0.280
Anaerotruncus (ID: 2054)	0.005	0.017	0.788	8.363	0.865
Ruminococcaceae UCG014(ID: 11371)	-0.010	0.016	0.522	11.928	0.479
Unknown genus (ID: 826)	-0.007	0.024	0.771	23.163	0.105

SE, standard error; RSS, residual sum of squares.

3. Results

3.1. Details of IVs

Overall, 2,252 SNPs were identified as final IVs. These SNPs were classified according to five levels: phylum, class, order, family, and

genus. Specifically, there were 102 IVs in 9 phyla, 179 IVs in 16 classes, 216 IVs in 20 orders, 383 IVs in 35 families, and 1,372 IVs in 131 genera. In addition, all IVs were more strongly associated with exposure than with outcome ($p_{exposure} < p_{outcome}$), and all F-statistics were greater than 10. Details of the IVs are presented in Supplementary Table S1.

TABLE 3 Horizontal pleiotropy analysis for IVs of 5 GM taxa associated with generalized epilepsy.

Exposure	MR-Egger intercept test	MR-PRESSO global test				
	Egger_intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value	
Phylum						
Actinobacteria (ID: 400)	-0.007	0.040	0.858	7.596	0.920	
Class						
Bacilli (ID: 1673)	0.015	0.028	0.605	18.510	0.532	
Genus						
Bifidobacterium (ID: 436)	-0.049	0.027	0.102	12.796	0.483	
Coprobacter (ID: 949)	-0.012	0.053	0.830	12.348	0.384	
Unknown genus (ID: 826)	-0.023	0.046	0.630	21.511	0.128	

SE, standard error; RSS, residual sum of squares.

TABLE 4 Horizontal pleiotropy analysis for IVs of 8 GM taxa associated with focal epilepsy.

Exposure	MR-Egger intercept test			MR-PRESSO global test	
	Egger_intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value
Phylum					
Verrucomicrobia (ID: 3982)	0.023	0.054	0.678	13.273	0.450
Class					
Verrucomicrobiae (ID: 4029)	0.039	0.068	0.581	13.605	0.379
Order					
Verrucomicrobiales (ID: 4030)	0.039	0.068	0.581	13.605	0.364
Family					
Verrucomicrobiaceae(ID: 4036)	0.039	0.068	0.579	13.610	0.341
Genus					
Akkermansia (ID: 4037)	0.039	0.068	0.579	13.608	0.364
Alloprevotella (ID: 961)	-0.070	0.217	0.768	1.914	0.895
Clostridium Sensu Stricto 1(ID: 1873)	0.137	0.078	0.138	12.978	0.187
Sutterella (ID: 2896)	-0.098	0.069	0.187	12.428	0.525

SE, standard error; RSS, residual sum of squares.

3.2. MR analysis

First, we performed MR analysis to assess the causal association of 211 GM taxa at five levels with epilepsy. The results assessed by the IVW-FE showed that class *Betaproteobacteria* (ID: 2867), class *Verrucomicrobiae* (ID: 4029), order *Burkholderiales* (ID: 2874), order *Verrucomicrobiales* (ID: 4030), family *Verrucomicrobiaeeae* (ID: 4036), genus *Akkermansia* (ID: 4037), genus *Anaerotruncus* (ID: 2054) and genus *Ruminococcaeae UCG 014* (ID: 11371) were associated with an increased risk for epilepsy, while genus *Eubacterium Xylanophilus Group* (ID: 14375) and genus *Unknown genus* (ID: 826) were associated with a decreased risk for epilepsy (Figure 2A). Furthermore, the results of Cochran's Q test indicated the absence of heterogeneity. After applying the Bonferroni correction, class *Betaproteobacteria* (ID: 2867) [OR=1.357 (1.126, 1.635), p = 0.001] and order *Burkholderiales* (ID: 2874) [OR=1.336 (1.112, 1.606), p = 0.002] remained risk factors for epilepsy.

Subsequently, we further evaluated the causal association of 211 GM taxa with generalized epilepsy using the IVW-FE method. The results showed that phylum *Actinobacteria* (ID: 400) and genus

Bifidobacterium (ID: 436) were associated with an increased risk for generalized epilepsy, while class *Bacilli* (ID: 1673), genus *Coprobacter* (ID: 949), genus *Unknown genus* (ID: 826) and genus *Unknown genus* (ID: 1868) were associated with a decreased risk for generalized epilepsy (Figure 2B). However, after Bonferroni correction, the causal effect of these GM taxa on generalized epilepsy was insignificant. Furthermore, the results of Cochran's Q-test suggested heterogeneity in the MR analysis of Genus *Unknown genus* (ID: 1868); thus, the IVW random effect (RE) was applied to explain the causal association of this GM taxon with generalized epilepsy, with results indicating no causal association.

Finally, we assessed the causal association of 211 GM taxa with focal epilepsy using the IVW-FE method. The results showed that phylum *Verrucomicrobia* (ID: 3982), class *Verrucomicrobiae* (ID: 4029), order *Verrucomicrobiales* (ID: 4030), family *Verrucomicrobiaceae* (ID: 4036), genus *Akkermansia* (ID: 4037), genus *Alloprevotella* (ID: 961), and genus *Sutterella* (ID: 2896) were associated with an increased risk for focal epilepsy, while genus *Clostridium Sensu Stricto 1* (ID: 1873) was associated with a decreased risk for focal epilepsy (Figure 2C). The results of Cochran's Q test

suggested no heterogeneity in the MR analysis. However, after Bonferroni correction, the causal effect of these GM taxa on generalized epilepsy was insignificant.

In addition, four additional methods, MR-Egger, weighted median, simple mode, and weighted mode, were performed to assess the causal effect of these GM taxa on epilepsy (Figure 3), generalized epilepsy (Figure 4), and focal epilepsy (Figure 5). Similarly, the results were parallel to the IVW results (Supplementary Figure S1). The heat map visualized the causal association of GM taxa identified in our study with epilepsy, generalized epilepsy, and focal epilepsy (Figure 6).

3.3. Sensitivity analysis

The results of the MR-Egger intercept test and MR-PRESSO global test showed that there was no horizontal pleiotropy ($p_{\text{MR-Egger}}$) and global $p_{\text{MR-PRESSO}}$ >0.05) in (i) IVs of 10 GM taxa associated with epilepsy (Table 2), (ii) IVs of 5 GM taxa associated

with generalized epilepsy (Table 3), and (iii) IVs of 8 GM taxa associated with focal epilepsy (Table 4). In addition, the leave-one-out analysis indicated the robustness of the MR results since excluding any one IV did not shift the overall results (Supplementary Figure S2).

3.4. Replicated analysis after removing confounders-related IVs

Among the IVs of 10 GM taxa associated with epilepsy, rs4936098 was associated with obesity and rs2321387 with education level. In addition, among the IVs of 5 GM taxa associated with generalized epilepsy, rs12634544, rs182549, rs1397793, rs7570971, rs35344081, and rs35344081 were associated with obesity; rs182549 with diabetes mellitus; and rs2952251 with smoking. Furthermore, among the IVs of 8 GM taxa associated with focal epilepsy, rs4936098 was associated with obesity, and rs2321387 with education level. After removing these SNPs from the IVs, the causal associations of these GM taxa were re-evaluated by the IVW-FE method. The results showed that,

TABLE 5 Replicated MR analysis by IVW method after removing confounders-related IVs.

Exposure	Outcome	<i>p</i> -value	OR (95% CI)
Phylum			
Actinobacteria (ID: 400)	Generalized epilepsy	0.132	1.318 (0.920, 1.889)
Verrucomicrobia (ID: 3982)	Focal epilepsy	0.017	1.697 (1.101, 2.616)
Class			
Betaproteobacteria(ID: 2867)	Epilepsy	0.001	1.381 (1.136, 1.679)
Verrucomicrobiae (ID: 4029)	Epilepsy	0.030	1.186 (1.016, 1.384)
Bacilli (ID: 1673)	Generalized epilepsy	0.025	0.718 (0.537, 0.960)
Verrucomicrobiae (ID: 4029)	Focal epilepsy	0.008	1.900 (1.178, 3.063)
Order			
Burkholderiales (ID: 2874)	Epilepsy	0.002	1.359 (1.121, 1.648)
Verrucomicrobiales (ID: 4030)	Epilepsy	0.030	1.186 (1.016, 1.384)
Verrucomicrobiales (ID: 4030)	Focal epilepsy	0.008	1.900 (1.178, 3.063)
Family			
Verrucomicrobiaceae (ID: 4036)	Epilepsy	0.030	1.186 (1.016, 1.385)
Verrucomicrobiaceae (ID: 4036)	Focal epilepsy	0.008	1.900 (1.178, 3.063)
Genus			
Eubacterium Xylanophilus Group (ID: 14375)	Epilepsy	0.014	0.816 (0.694, 0.959)
Akkermansia (ID: 4037)	Epilepsy	0.030	1.186 (1.017, 1.385)
Anaerotruncus (ID: 2054)	Epilepsy	0.011	1.238 (1.050, 1.459)
Ruminococcaceae UCG014(ID: 11371)	Epilepsy	0.030	1.178 (1.016, 1.367)
Unknown genus (ID: 826)	Epilepsy	0.004	0.819 (0.715, 0.939)
Bifidobacterium (ID: 436)	Generalized epilepsy	0.036	1.389 (1.021, 1.890)
Coprobacter (ID: 949)	Generalized epilepsy	0.044	0.787 (0.624, 0.993)
Unknown genus (ID: 826)	Generalized epilepsy	0.005	0.682 (0.524, 0.889)
Akkermansia (ID: 4037)	Focal epilepsy	0.008	1.899 (1.178, 3.062)
Alloprevotella (ID: 961)	Focal epilepsy	0.025	1.499 (1.052, 2.137)
Clostridium Sensu Stricto 1(ID: 1873)	Focal epilepsy	0.024	0.534 (0.310, 0.919)
Sutterella (ID: 2896)	Focal epilepsy	0.032	1.715 (1.048, 2.806)

OR, odds ratio; CI, confidence interval.

except for phylum *Actinobacteria* (ID: 400), the causal effects of the above GM taxa remained significant (Table 5).

4. Discussion

Our study comprehensively assessed the causal effect of 211 GM taxa (from phylum to genus level) on epilepsy and its sub-types. Finally, we identified a total of 23 causal relationships, of which 21 were nominal causal relationships, and two were strong causal relationships, thus highlighting the importance of GMs in epilepsy.

Accumulating evidence has suggested crosstalk between GMs and the central nervous system (CNS) (Cryan and Dinan, 2012). Investigations have shown that GMs play a vital role in the development of the enteric nervous system, blood-brain barrier, and glial cells, which are all important for cognitive development and behavior regulation (Braniste et al., 2014; Collins et al., 2014). Various neurological disorders, including multiple sclerosis (Jangi et al., 2016), autism (Mulle et al., 2013), Alzheimer's disease (Jiang et al., 2017), and Parkinson's disease (Parashar and Udayabanu, 2017), have been linked to intestinal dysbiosis. Recent findings also suggest that GMs may also play a role in epilepsy (Russo, 2022). Several studies have examined the effect of the KD, a treatment for refractory epilepsy, on GMs to explore the potential mechanisms of GMs in KD treatment (Lum et al., 2020). However, it is inconclusive which GM taxa have the most significant impact on epilepsy. As one-third of patients with epilepsy are diagnosed with refractory epilepsy (Dahlin and Prast-Nielsen, 2019), exploring biomarkers of epilepsy on the GMs level could offer promising alternative treatment options and potentially prevent the need for invasive treatments such as vagus nerve stimulation (VNS) or epilepsy surgery (Braakman and van Ingen, 2018).

Our study identified two strong causal relationships. Class *Betaproteobacteria* (OR = 1.357, 95% CI: 1.126–1.635, *p* = 0.001) and *Order Burkholderiales* (OR = 1.336, 95% CI: 1.112–1.606, *p*=0.002) significantly elevated the epilepsy risk after Bonferroni correction. Burkholderiales, an order of Betaproteobacteria, was found to have a potential impact on epilepsy from our MR study, which was consistent with the findings of some previous investigations. For instance, Safak et al. identified the genus Delftia and genus Lautropia, which are members of Burkholderiales, to be significantly higher in the intestine of epilepsy patients versus healthy individuals (Safak et al., 2020). In addition, another genus of Burkholderiales, Sutterella, which was reported with increased intestinal abundance in adult patients with epilepsy (Dong et al., 2022), was also identified in our study to be nominally associated with an increased risk of focal epilepsy. The present MR study could provide evidence and confidence for the increased level of genera belonging to Order Burkholderiales in the intestines of epilepsy patients.

It's important to note that the Bonferroni correction can result in false negatives. Our findings showed 21 GM taxa with nominal causal connections, but these correlations vanished after applying the Bonferroni correction. This may be due to the crosstalk between the gut-brain axis being usually coordinated by multiple factors and that the role of a single microbiota in the genus level in causing disease may not be as important as previously thought. In fact, several GM

taxa with nominal causal relationships identified in this study corroborate the findings of previous research. For instance, Huang et al. revealed that patients with cerebral palsy and epilepsy contained a higher proportion, in comparison to healthy controls, of Bifidobacterium and Akkermansia (Huang et al., 2019). In addition, Gong and colleagues identified Bifidobacterium, Ruminococcaceae UCG 014, and Akkermansia at the genus level were increased in patients with epilepsy compared to healthy controls (Gong et al., 2020). Further, Lee et al. identified Enterococcus faecium (species of class Bacilli), Bifidobacterium longum (species of genus Bifidobacterium), and Eggerthella lenta (species of phylum Actinobacteria) as biomarkers for drug-resistant epilepsy (Lee et al., 2021). Although only nominal causal associations were identified at the genus level for these GM taxa, the coordination and crosstalk between various GM taxa remain worthy of in-depth study in the future.

The mechanisms involved in the relationship between GMs and epilepsy have not been fully determined. However, some evidence suggests potential mechanisms. (i) Studies have reported that GMs can alter neurotransmitter levels such as glutamate, gammaaminobutyric acid (GABA), 5-hydroxytryptamine (5-HT) (Mittal et al., 2017), as well as increase levels of cytokines, chemokines, such as TNFa and MCP-1, lipopolysaccharides (LPS) which led to generalized immune activation or inflammation (Blander et al., 2017), contributing to the risk of seizures. (ii) GMs have been demonstrated to interact with gut-derived metabolites, resulting in both beneficial and detrimental mechanisms for the central nervous system (Tran and Mohajeri, 2021). (iii) In addition, GMs can affect the hypothalamicpituitary-adrenal (HPA) axis (Sudo et al., 2004) and the levels of brain-derived neurotrophic factor (BDNF) (Maqsood and Stone, 2016), which promote seizure propensity. (iv) GMs also regulate peripheral metabolites and central neurotransmitter metabolism, which affect seizure susceptibility (Lum et al., 2020). Nevertheless, the specific mechanism and crosstalk between different GM taxa remain to be verified by future studies.

The limitations of the present study should be noted: (i) Since the number of IVs fulfilling the strict threshold $(p < 5 \times 10^{-8})$ was extremely small, a relatively lenient threshold $(p < 1 \times 10^{-5})$ was adopted for screening IVs. (ii) This study included individuals of essentially European ancestry, so extrapolating the findings to other populations is limiting. (iii) The number of cases of the two subtypes of epilepsy under strict definition (generalized epilepsy and focal epilepsy) is relatively small, so future analysis based on a larger sample size of GWAS summary data is necessary to increase the confidence of the results. (iv) The GM-related GWAS summary-level dataset included in this study was based on 16S rRNA sequencing, and thus further analysis based on large-scale studies with more advanced methods, such as metagenomics sequencing, is required in the future in order to evaluate the species-level. (v) Current studies of GMs have focused only on bacteria; however, other types of GMs may also have potential functions.

5. Conclusion

Overall, by performing MR analysis of the causal effects of 211 GM taxa on epilepsy and its sub-types, we finally identified 21

nominal causal relationships and two strong causal relationships. Among them, *Class Betaproteobacteria* and *Order Burkholderiales* are significantly associated with increased epilepsy risk. However, it is essential to recognize that since the present study was conducted based on the GWAS summary-level dataset generated from 16S rRNA sequencing, further in-depth analyses based on more advanced large-scale studies generated from metagenomics sequencing are necessary. Nevertheless, our findings may provide helpful biomarkers for disease progression and potential candidate therapeutic targets for epilepsy.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found at: https://mibiogen.gcc.rug.nl/, https://r7.finngen.fi/.

Ethics statement

Publicly available de-identified data from participant studies approved by an ethical standards committee were used in this study. Therefore, no additional separate ethical approval was required for this study.

Author contributions

YZ designed the study, analyzed the data, and wrote the manuscript. SC assisted in analyzing the data and revising the manuscript. HY critically read and edited the manuscript. All authors contributed to the article and approved the submitted version.

References

Blander, J. M., Longman, R. S., Iliev, I. D., Sonnenberg, G. F., and Artis, D. (2017). Regulation of inflammation by microbiota interactions with the host. *Nat. Immunol.* 18, 851–860. doi: 10.1038/ni.3780

Bowden, J., Davey Smith, G., Haycock, P. C., and Burgess, S. (2016). Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genet. Epidemiol.* 40, 304–314. doi: 10.1002/gepi. 21965

Braakman, H. M. H., and van Ingen, J. (2018). Can epilepsy be treated by antibiotics? J. Neurol. 265, 1934–1936. doi: 10.1007/s00415-018-8943-3

Braniste, V., Al-Asmakh, M., Kowal, C., Anuar, F., Abbaspour, A., Toth, M., et al. (2014). The gut microbiota influences blood-brain barrier permeability in mice. *Sci. Transl. Med.* 6:263ra158. doi: 10.1126/scitranslmed.3009759

Burgess, S., Small, D. S., and Thompson, S. G. (2017). A review of instrumental variable estimators for Mendelian randomization. *Stat. Methods Med. Res.* 26, 2333–2355. doi: 10.1177/0962280215597579

Burgess, S., and Thompson, S. G. (2017). Interpreting findings from Mendelian randomization using the Mr-Egger method. *Eur. J. Epidemiol.* 32, 377–389. doi: 10.1007/s10654-017-0255-x

Cai, J., Li, X., Wu, S., Tian, Y., Zhang, Y., Wei, Z., et al. (2022). Assessing the causal association between human blood metabolites and the risk of epilepsy. *J. Transl. Med.* 20:437. doi: 10.1186/s12967-022-03648-5

Collaborators, G. B. D. E. (2019). Global, regional, and National Burden of epilepsy, 1990-2016: a systematic analysis for the global burden of disease study 2016. *Lancet Neurol.* 18, 357–375. doi: 10.1016/S1474-4422(18)30454-X

Collins, J., Borojevic, R., Verdu, E. F., Huizinga, J. D., and Ratcliffe, E. M. (2014). Intestinal microbiota influence the early postnatal development of the enteric nervous system. *Neurogastroenterol. Motil.* 26, 98–107. doi: 10.1111/nmo.12236

Funding

This research was funded by the Natural Science Foundation of Hunan Province (2022JJ70069).

Acknowledgments

We want to acknowledge the MiBioGen consortium for providing GM-related GWAS summary data. We want to acknowledge the participants and investigators of the FinnGen study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmicb.2023.1115014/ full#supplementary-material

Cryan, J. F., and Dinan, T. G. (2012). Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat. Rev. Neurosci.* 13, 701–712. doi: 10.1038/nrn3346

D'Andrea Meira, I., Romão, T. T., Pires do Prado, H. J., Krüger, L. T., Pires, M. E. P., and da Conceição, P. O. (2019). Ketogenic diet and epilepsy: what we know so far. *Front. Neurosci.* 13:5. doi: 10.3389/fnins.2019.00005

Dahlin, M., and Prast-Nielsen, S. (2019). The gut microbiome and epilepsy. *EBioMedicine* 44, 741–746. doi: 10.1016/j.ebiom.2019.05.024

Darch, H., and McCafferty, C. P. (2022). Gut microbiome effects on Neuronal Excitability & Activity: implications for epilepsy. *Neurobiol. Dis.* 165:105629:105629. doi: 10.1016/j.nbd.2022.105629

de Biase, S., Nilo, A., Bernardini, A., Gigli, G. L., Valente, M., and Merlino, G. (2019). Timing use of novel anti-epileptic drugs: is earlier better? *Expert. Rev. Neurother.* 19, 945–954. doi: 10.1080/14737175.2019.1636649

Dong, L., Zheng, Q., Cheng, Y., Zhou, M., Wang, M., Xu, J., et al. (2022). Gut microbial characteristics of adult patients with epilepsy. *Front. Neurosci.* 16:803538. doi: 10.3389/fnins.2022.803538

Durazzi, F., Sala, C., Castellani, G., Manfreda, G., Remondini, D., and De Cesare, A. (2021). Comparison between 16s Rrna and shotgun sequencing data for the taxonomic characterization of the gut microbiota. *Sci. Rep.* 11:3030. doi: 10.1038/s41598-021-82726-y

Fan, Y., Wang, H., Liu, X., Zhang, J., and Liu, G. (2019). Crosstalk between the ketogenic diet and epilepsy: from the perspective of gut microbiota. *Mediat. Inflamm.* 2019:8373060, 1–9. doi: 10.1155/2019/8373060

Fisher, R. S., van Emde, B. W., Blume, W., Elger, C., Genton, P., Lee, P., et al. (2005). Epileptic seizures and epilepsy: definitions proposed by the international league against epilepsy (Ilae) and the International Bureau for Epilepsy (Ibe). *Epilepsia* 46, 470–472. doi: 10.1111/j.0013-9580.2005.66104.x

Gong, X., Liu, X., Chen, C., Lin, J., Li, A., Guo, K., et al. (2020). Alteration of gut microbiota in patients with epilepsy and the potential index as a biomarker. *Front. Microbiol.* 11:517797. doi: 10.3389/fmicb.2020.517797

Hafizi, N., Alipoor, E., and Hosseinzadeh-Attar, M. J. (2017). Obesity and epilepsy: the first step of a long road. *Epilepsy Behav.* 67, 135–136. doi: 10.1016/j.yebeh.2016. 12.004

Huang, C., Li, Y., Feng, X., Li, D., Li, X., Ouyang, Q., et al. (2019). Distinct gut microbiota composition and functional category in children with cerebral palsy and epilepsy. *Front. Pediatr.* 7:394. doi: 10.3389/fped.2019.00394

Iannone, L. F., Preda, A., Blottiere, H. M., Clarke, G., Albani, D., Belcastro, V., et al. (2019). Microbiota-gut brain Axis involvement in neuropsychiatric disorders. *Expert. Rev. Neurother.* 19, 1037–1050. doi: 10.1080/14737175.2019.1638763

Jangi, S., Gandhi, R., Cox, L. M., Li, N., von Glehn, F., Yan, R., et al. (2016). Alterations of the human gut microbiome in multiple sclerosis. *Nat. Commun.* 7:12015. doi: 10.1038/ncomms12015

Jiang, C., Li, G., Huang, P., Liu, Z., and Zhao, B. (2017). The gut microbiota and Alzheimer's disease. *J. Alzheimers Dis.* 58, 1–15. doi: 10.3233/JAD-161141

Johnson, K. V., and Foster, K. R. (2018). Why does the microbiome affect behaviour? *Nat. Rev. Microbiol.* 16, 647–655. doi: 10.1038/s41579-018-0014-3

Kamat, M. A., Blackshaw, J. A., Young, R., Surendran, P., Burgess, S., Danesh, J., et al. (2019). Phenoscanner V2: An expanded tool for searching human genotype-phenotype associations. *Bioinformatics* 35, 4851–4853. doi: 10.1093/bioinformatics/btz469

Kurilshikov, A., Medina-Gomez, C., Bacigalupe, R., Radjabzadeh, D., Wang, J., Demirkan, A., et al. (2021). Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat. Genet.* 53, 156–165. doi: 10.1038/ s41588-020-00763-1

Kurki, M., Karjalainen, J., Palta, P., Sipilä, T., Kristiansson, K., Donner, K., et al. (2022). Finngen: unique genetic insights from combining isolated population and national health register data. *medRxiv*. doi: 10.1101/2022.03.03.22271360

Lawlor, D. A., Harbord, R. M., Sterne, J. A., Timpson, N., and Davey, S. G. (2008). Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat. Med.* 27, 1133–1163. doi: 10.1002/sim.3034

Le Chatelier, E., Nielsen, T., Qin, J., Prifti, E., Hildebrand, F., Falony, G., et al. (2013). Richness of human gut microbiome correlates with metabolic markers. *Nature* 500, 541–546. doi: 10.1038/nature12506

Lee, K., Kim, N., Shim, J. O., and Kim, G. H. (2021). Gut bacterial Dysbiosis in children with intractable epilepsy. *J. Clin. Med.* 10:5. doi: 10.3390/jcm10010005

Lindefeldt, M., Eng, A., Darban, H., Bjerkner, A., Zetterstrom, C. K., Allander, T., et al. (2019). The ketogenic diet influences taxonomic and functional composition of the gut microbiota in children with severe epilepsy. *NPJ Biofilms Microbiomes* 5:5:5. doi: 10.1038/s41522-018-0073-2

Lum, G. R., Olson, C. A., and Hsiao, E. Y. (2020). Emerging roles for the intestinal microbiome in epilepsy. *Neurobiol. Dis.* 135:104576. doi: 10.1016/j.nbd.2019. 104576

Maqsood, R., and Stone, T. W. (2016). The gut-brain Axis, BDNF, NMDA and CNS disorders. *Neurochem. Res.* 41, 2819–2835. doi: 10.1007/s11064-016-2039-1

Marcovecchio, M. L., Petrosino, M. I., and Chiarelli, F. (2015). Diabetes and epilepsy in children and adolescents. *Curr. Diab. Rep.* 15:21. doi: 10.1007/s11892-015-0588-3

Mittal, R., Debs, L. H., Patel, A. P., Nguyen, D., Patel, K., O'Connor, G., et al. (2017). Neurotransmitters: the critical modulators regulating gut-brain Axis. *J. Cell. Physiol.* 232, 2359–2372. doi: 10.1002/jcp.25518

Mulle, J. G., Sharp, W. G., and Cubells, J. F. (2013). The gut microbiome: a new frontier in autism research. *Curr. Psychiatry Rep.* 15:337. doi: 10.1007/s11920-012-0337-0 Pagoni, P., Dimou, N. L., Murphy, N., and Stergiakouli, E. (2019). Using Mendelian randomisation to assess causality in observational studies. *Evid. Based Ment. Health* 22, 67–71. doi: 10.1136/ebmental-2019-300085

Parashar, A., and Udayabanu, M. (2017). Gut microbiota: implications in Parkinson's disease. *Parkinsonism Relat. Disord.* 38, 1–7. doi: 10.1016/j.parkreldis.2017.02.002

Pierce, B. L., Ahsan, H., and Vanderweele, T. J. (2011). Power and instrument strength requirements for Mendelian randomization studies using multiple genetic variants. *Int. J. Epidemiol.* 40, 740–752. doi: 10.1093/ije/dyq151

Rees, J. M. B., Wood, A. M., and Burgess, S. (2017). Extending the Mr-Egger method for multivariable Mendelian randomization to correct for both measured and unmeasured pleiotropy. *Stat. Med.* 36, 4705–4718. doi: 10.1002/sim.7492

Russo, E. (2022). The gut microbiota as a biomarker in epilepsy. *Neurobiol. Dis.* 163:105598. doi: 10.1016/j.nbd.2021.105598

Safak, B., Altunan, B., Topcu, B., and Eren, T. A. (2020). The gut microbiome in epilepsy. *Microb. Pathog.* 139:103853:103853. doi: 10.1016/j.micpath.2019.103853

Skrivankova, V. W., Richmond, R. C., Woolf, B. A. R., Yarmolinsky, J., Davies, N. M., Swanson, S. A., et al. (2021). Strengthening the reporting of observational studies in epidemiology using Mendelian randomization: the Strobe-Mr statement. *JAMA* 326, 1614–1621. doi: 10.1001/jama.2021.18236

Staley, J. R., Blackshaw, J., Kamat, M. A., Ellis, S., Surendran, P., Sun, B. B., et al. (2016). Phenoscanner: a database of human genotype-phenotype associations. *Bioinformatics* 32, 3207–3209. doi: 10.1093/bioinformatics/btw373

Sudo, N., Chida, Y., Aiba, Y., Sonoda, J., Oyama, N., Yu, X. N., et al. (2004). Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J. Physiol.* 558, 263–275. doi: 10.1113/jphysiol.2004.063388

Swanson, S. A., Tiemeier, H., Ikram, M. A., and Hernan, M. A. (2017). Nature as a Trialist?: deconstructing the analogy between Mendelian randomization and randomized trials. *Epidemiology* 28, 653–659. doi: 10.1097/EDE.000000000000699

Swertz, M. A., Dijkstra, M., Adamusiak, T., van der Velde, J. K., Kanterakis, A., Roos, E. T., et al. (2010). The Molgenis toolkit: rapid prototyping of biosoftware at the push of a button. *BMC Bioinformatics* 11:S12. doi: 10.1186/1471-2105-11-S12-S12

Swertz, M. A., and Jansen, R. C. (2007). Beyond standardization: dynamic software infrastructures for systems biology. *Nat. Rev. Genet.* 8, 235–243. doi: 10.1038/nrg2048

Thambi, M., Nathan, J., and Radhakrishnan, K. (2020). Can change in gut microbiota composition be used as a surrogate marker of treatment efficacy of ketogenic diet in patients with drug-resistant epilepsy? *Epilepsy Behav.* 113:107444. doi: 10.1016/j. yebeh.2020.107444

Thijs, R. D., Surges, R., O'Brien, T. J., and Sander, J. W. (2019). Epilepsy in adults. Lancet 393, 689-701. doi: 10.1016/S0140-6736(18)32596-0

Tran, S. M., and Mohajeri, M. H. (2021). The role of gut bacterial metabolites in brain development, aging and disease. *Nutrients* 13:732. doi: 10.3390/nu13030732

van der Velde, K. J., Imhann, F., Charbon, B., Pang, C., van Enckevort, D., Slofstra, M., et al. (2019). Molgenis research: advanced bioinformatics data software for non-Bioinformaticans. *Bioinformatics* 35, 1076–1078. doi: 10.1093/bioinformatics/bty742

Verbanck, M., Chen, C. Y., Neale, B., and Do, R. (2018). Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat. Genet.* 50, 693–698. doi: 10.1038/s41588-018-0099-7

Wang, M., Zhang, Z., Liu, D., Xie, W., Ma, Y., Yao, J., et al. (2021). Educational attainment protects against epilepsy independent of cognitive function: a Mendelian randomization study. *Epilepsia* 62, 1362–1368. doi: 10.1111/epi.16894

Yuan, S., Tomson, T., and Larsson, S. C. (2021). Modifiable risk factors for epilepsy: a two-sample Mendelian randomization study. *Brain Behav.* 11:e02098. doi: 10.1002/brb3.2098