



A Systematic Review of *In vitro* and *In vivo* Activities of Anti-*Toxoplasma* Drugs and Compounds (2006–2016)

Mahbobeh Montazeri^{1,2}, Mehdi Sharif^{1,3}, Shahabeddin Sarvi^{1,3}, Saeed Mehrzadi⁴, Ehsan Ahmadpour⁵ and Ahmad Daryani^{1,3*}

¹ Toxoplasmosis Research Center, Mazandaran University of Medical Sciences, Sari, Iran, ² Student Research Committee, Mazandaran University of Medical Sciences, Sari, Iran, ³ Department of Parasitology and Mycology, Sari Medical School, Mazandaran University of Medical Sciences, Sari, Iran, ⁴ Department of Pharmacology, School of Medicine, Iran University of Medical Sciences Tehran, Iran, ⁵ Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

OPEN ACCESS

Edited by:

Octavio Luiz Franco,
Universidade Católica de Brasília,
Brazil

Reviewed by:

Osmar Nascimento Silva,
Universidade Católica Dom Bosco,
Brazil

Nasib Singh,
Eternal University, India

*Correspondence:

Ahmad Daryani
daryanii@yahoo.com

Specialty section:

This article was submitted to
Antimicrobials, Resistance and
Chemotherapy,
a section of the journal
Frontiers in Microbiology

Received: 01 September 2016

Accepted: 05 January 2017

Published: 20 January 2017

Citation:

Montazeri M, Sharif M, Sarvi S,
Mehrzadi S, Ahmadpour E and
Daryani A (2017) A Systematic Review
of *In vitro* and *In vivo* Activities of
Anti-*Toxoplasma* Drugs and
Compounds (2006–2016).
Front. Microbiol. 8:25.
doi: 10.3389/fmicb.2017.00025

The currently available anti-*Toxoplasma* agents have serious limitations. This systematic review was performed to evaluate drugs and new compounds used for the treatment of toxoplasmosis. Data was systematically collected from published papers on the efficacy of drugs/compounds used against *Toxoplasma gondii* (*T. gondii*) globally during 2006–2016. The searched databases were PubMed, Google Scholar, Science Direct, ISI Web of Science, EBSCO, and Scopus. One hundred and eighteen papers were eligible for inclusion in this systematic review, which were both *in vitro* and *in vivo* studies. Within this review, 80 clinically available drugs and a large number of new compounds with more than 39 mechanisms of action were evaluated. Interestingly, many of the drugs/compounds evaluated against *T. gondii* act on the apicoplast. Therefore, the apicoplast represents as a potential drug target for new chemotherapy. Based on the current findings, 49 drugs/compounds demonstrated *in vitro* half-maximal inhibitory concentration (IC₅₀) values of below 1 μM, but most of them were not evaluated further for *in vivo* effectiveness. However, the derivatives of the ciprofloxacin, endochin-like quinolones and 1-[4-(4-nitrophenoxy) phenyl] propane-1-one (NPPP) were significantly active against *T. gondii* tachyzoites both *in vitro* and *in vivo*. Thus, these compounds are promising candidates for future studies. Also, compound 32 (*T. gondii* calcium-dependent protein kinase 1 inhibitor), endochin-like quinolones, miltefosine, rolipram abolish, and guanabenz can be repurposed into an effective anti-parasitic with a unique ability to reduce brain tissue cysts (88.7, 88, 78, 74, and 69%, respectively). Additionally, no promising drugs are available for congenital toxoplasmosis. In conclusion, as current chemotherapy against toxoplasmosis is still not satisfactory, development of well-tolerated and safe specific immunoprophylaxis in relaxing the need of dependence on chemotherapeutics is a highly valuable goal for global disease control. However, with the increasing number of high-risk individuals, and absence of a proper vaccine, continued efforts are necessary for the development of novel treatment options against *T. gondii*. Some of the novel compounds reviewed here may represent good starting points for the discovery of effective new drugs. In further, bioinformatic and *in silico* studies are needed in order to identify new potential toxoplasmodicidal drugs.

Keywords: *Toxoplasma gondii*, toxoplasmosis, drugs, compounds, *in vitro*, *in vivo*

INTRODUCTION

Toxoplasma gondii (*T. gondii*), an obligate intracellular, parasitic protozoan, is the etiologic agent of toxoplasmosis. About 30–50% of the world population is infected with the parasite, and it is the most prevalent infection among humans (Tenter et al., 2000; Flegr et al., 2014). Worldwide, over 1 billion people are estimated to be infected with *T. gondii* (Hoffmann et al., 2012). Its prevalence in some countries is high (e.g., Brazil, 77.5%; Sao Tome and Principe, 75.2%; Iran, 63.9%; Colombia, 63.5%; and Cuba, 61.8%) (Pappas et al., 2009). The annual incidence of congenital toxoplasmosis was estimated to be 190,100 cases globally (Torgerson and Mastroiacovo, 2013).

In the United States, the Centers for Disease Control and Prevention (CDC) reported that 22.5% of the population 12 years and older have been infected with *Toxoplasma* with 1.1 million new infections each year, making it the second most common cause of deaths due to foodborne diseases (an estimated 327 deaths) and the fourth leading cause of hospitalizations attributable to foodborne illness (an estimated 4428 hospitalizations). Also, an estimated 400–4000 infants are born with congenital toxoplasmosis in the United States each year (Jones et al., 2014).

T. gondii has three infectious stages of sporozoites (in oocysts), tachyzoites (rapidly multiplying form), and bradyzoites (tissue cyst form). Among them, tachyzoites are responsible for clinical manifestations and the acute phase of the disease. They are susceptible to the immune response of the host and to drug action. The resistant cyst form of the parasite is resistant to both the immune system and drugs (Hill and Dubey, 2002).

Acute toxoplasmosis in healthy individuals is usually subclinical and asymptomatic, but may lead to chronic infection. However, toxoplasmosis can lead to great morbidity and mortality rates in immunocompromised or congenitally infected individuals (Dubey and Jones, 2008; Ahmadpour et al., 2014). In AIDS patients, presence of the parasite causes necrotizing encephalitis and focal cerebral lesions in the central nervous system (CNS) from primary or recrudescent infection. In immunocompetent patients, latent toxoplasmosis occurs with the formation of cysts principally in the CNS (Martins-Duarte et al., 2006).

In the recent years, the development of well-tolerated and safe specific immunoprophylaxis against toxoplasmosis is a highly valuable goal for global disease control (Lim and Othman, 2014). Immunotherapeutic strategies for improving toxoplasmosis control could either be a vaccine which would induce strong protective immunity against toxoplasmosis, or passive immunization in cases of disease recrudescence. In the last few years, much progress has been made in vaccine research on DNA vaccination, protein vaccination, live attenuated vaccinations, and heterologous vaccination; while there were few candidates on passive immunization. New vaccine candidates have been tested, including in particular proteins from *T. gondii* ROP, MIC, and GRA organelles, multi-antigen vaccines, novel adjuvants but until now the researches could not access to a proper vaccine for prevention of toxoplasmosis in human (Zhang et al., 2013, 2015).

The recommended drugs for treatment or prophylaxis of toxoplasmosis are pyrimethamine and sulfadiazine. Unfortunately, these drugs have side effects such as neutropenia, severe drop of platelet count, thrombocytopenia, leucopenia, elevation in serum creatinine and serum liver enzymes, hematological abnormalities, and hypersensitivity reactions (Bosch-Driessen et al., 2002; Silveira et al., 2002; Schmidt et al., 2006). In addition, other drugs, such as azithromycin, clarithromycin, spiramycin, atovaquone, dapsone, and cotrimoxazole (trimethoprim-sulfamethoxazole), have been used for clinical toxoplasmosis. However, these drugs are poorly tolerated and have no effect on the bradyzoite form (Araujo and Remington, 1992; Petersen and Schmidt, 2003; Serranti et al., 2011).

In a clinical trial, 24% of sera positive women treated with spiramycin and pyrimethamine plus sulfadoxine combination delivered *Toxoplasma* infected infants in France (Bessières et al., 2009). Spiramycin monotherapy can be effective during the early stage of pregnancy to prevent prenatal transmission (Julliac et al., 2010). More than 50% of patients treated with spiramycin retained *T. gondii* DNA in blood and remained infected (Habib, 2008).

In recent years, studies have focused on finding safe drugs with novel mechanisms of action against *T. gondii*. Accordingly, there is an urgent need to evaluate new drugs based on novel and innovative therapeutic strategies against *T. gondii* that are both efficacious and nontoxic for humans (Rodriguez and Szajnman, 2012; Vanagas et al., 2012; Angel et al., 2013). Therefore, the goal of the present systematic review was to retrieve published studies related to *in vitro* and *in vivo* evaluation of drugs and compounds for the treatment of toxoplasmosis (2006–2016) in order to prepare comprehensive data for designing more accurate investigations in future.

METHODOLOGY

This review followed the preferred reporting items for systematic reviews (PRISMA) guidelines (Moher et al., 2009).

Literature Search, Study Selection, and Data Extraction

English databases, including PubMed, Science Direct, Scopus, Google Scholar, ISI Web of Science, and EBSCO, were systematically searched for papers on *in vitro* and *in vivo* evaluation of anti-*Toxoplasma* activities of drugs and compounds, published worldwide, from 2006 to 2016. The keywords included were: “Toxoplasmosis,” “*T. gondii*,” “Anti-*Toxoplasma*,” “Drug,” “Anticoccidial,” “Treatment,” “*In vitro*,” “*In vivo*,” and “Compound.”

Papers written in English were selected. Gray literature and abstracts of articles which were published in congresses were not explored. In addition, in order to avoid missing any articles, whole references of the papers were meticulously hand-searched. Among English articles found with the mentioned strategies, full text papers that used laboratory method both *in vitro* and *in vivo* were included.

Also, studies with at least one of the following criteria were excluded: (1) studies that were not relevant; (2) articles not available in English; (3) studies on treatments for ocular infection; (4) articles that were of review or descriptive study type; (5) articles which contained no eligible data; (6) case series reports; (7) the data were duplicated from other studies or we were unable to obtain them; (8) those that were on efficacy of anti-*T. gondii* medicines in humans; and (9) any drug with an IC_{50} value $> 10 \mu M$.

Data Collection

All the experimental studies that were carried out to evaluate the efficacy of either drugs or compounds against *T. gondii* both *in vitro* and *in vivo* were included, and replicates were excluded. The inclusion criteria for selection of *in vitro* studies were important information about medication used for the experiments, type of cells used for culture, identification of the *Toxoplasma* strain, laboratory methods used for assessing drug activities, and main results comprising of the 50% inhibitory concentration (IC_{50}). We reported *in vivo* studies used animal models, *Toxoplasma* strain, route of infection, the

treatment schedule (dosage, route of administration, duration of treatment), the criteria for assessing drug activity (mainly survival for acute toxoplasmosis, histology, and brain cyst burdens for chronic infection), and the main results.

RESULTS

Analysis of the Included Literature

A total of 118 papers (83 studies *in vitro*, 59 *in vivo*, 27 both *in vitro* and *in vivo*) published from 2006 to 2016, were included in the systematic review. **Figure 1** briefly shows the search process in this systematic review article.

Mechanisms of Action

In the current systematic review, 80 clinically available drugs (**Table 1**) and several new compounds with more than 39 pathways/ mechanisms of action were evaluated against *T. gondii* in both *in vitro* and *in vivo* studies. Several target based drug screens were also identified against *T. gondii* include mitochondrial electron transport chain, calcium-dependent protein kinase 1, type II fatty acid synthesis, DNA synthesis, DNA

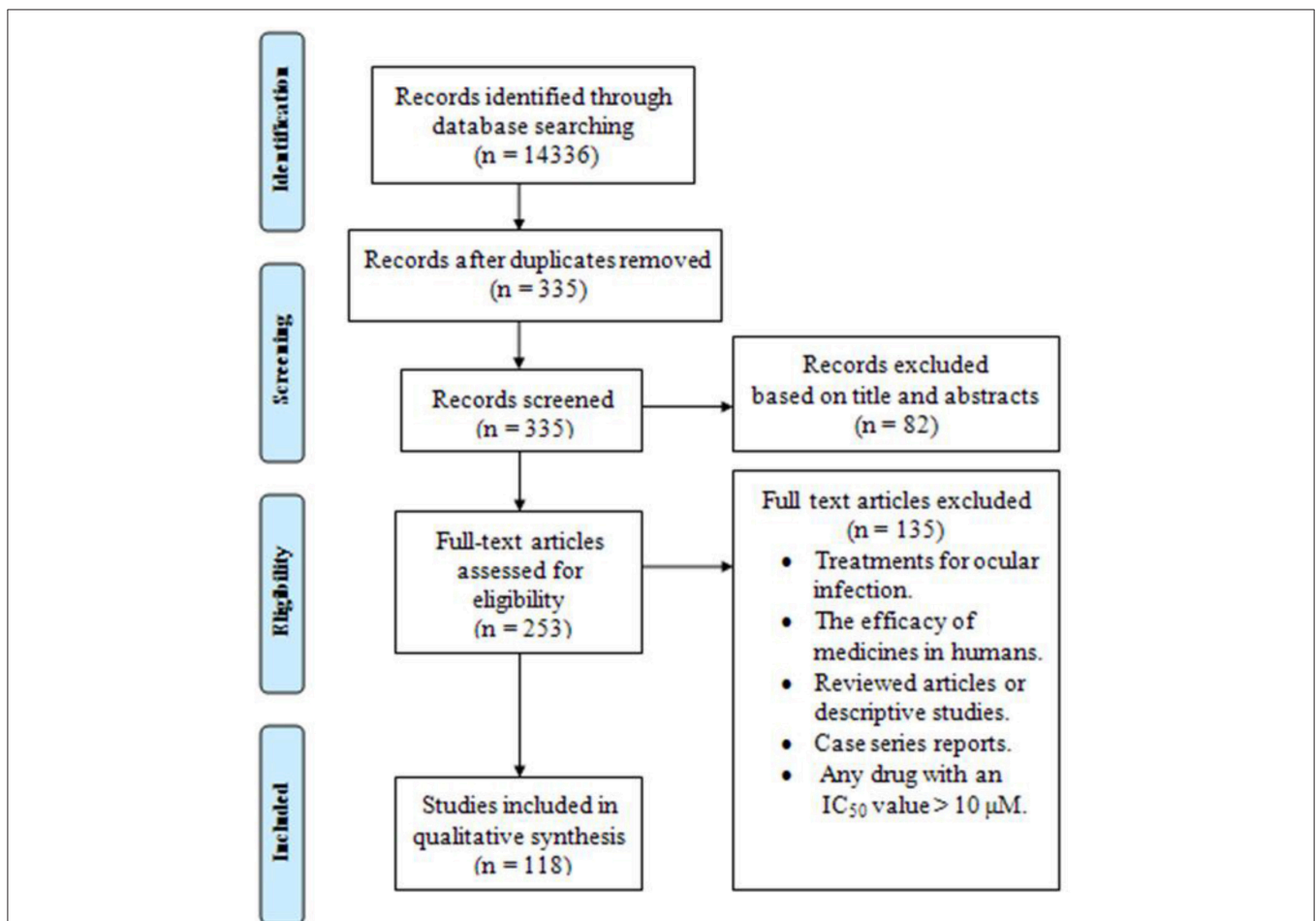


FIGURE 1 | The PRISMA flow diagram of the search strategy, study selection, and data management procedure of *in vitro* and *in vivo* activities of anti-*Toxoplasma* drugs and compounds (2006–2016).

TABLE 1 | Clinically available drugs/compounds evaluated against *T. gondii* in vitro and in vivo studies.

Common clinical uses	Drugs/compounds	References
Antiprotozoal agents	Bisphosphonates Diamidine analogs Spiramycin (Rovamycin) Thiosemicarbazides 4-thiazolidinones 1,3,4-thiadiazoles Naphthalene-sulfonyl-indole Thiosemicarbazone Phenylsemicarbazone Ivermectin Silver nanoparticles Novel ferrocenic atovaquone derivatives Triclosan Triclosan liposomal nanoparticles Metronidazole 1,25(OH) ₂ D ₃ Naphthoquinone PHNQ6 ^a Novel azasterols Apicidin	Baramée et al., 2006; Ferreira et al., 2006; Rajapakse et al., 2007; Strobl et al., 2007; Leepin et al., 2008; Shubar et al., 2008; Liesen et al., 2010; Aquino et al., 2011; Franco et al., 2011; Martins-Duarte et al., 2011; Chew et al., 2012; Asgari et al., 2013; Bilgin et al., 2013; Gomes et al., 2013; Gaafar et al., 2014; da Silva et al., 2015; El-Zawawy et al., 2015a,b
Antimalarial agents	Pyrimethamine Atovaquone Triazine JPC-2067-B Spiroindolone Endochin-like quinolones Halofuginone	Meneceur et al., 2008; Mui et al., 2008; Doggett et al., 2012; Zhou et al., 2014; Jain et al., 2015
Antibacterial agents	Sulfadiazine Azithromycin Enrofloxacin Fusidic acid Ciprofloxacin Chitosan	Meneceur et al., 2008; Costa et al., 2009; Barbosa et al., 2012; Payne et al., 2013; Castro-Filice et al., 2014; Gaafar et al., 2014; Martins-Duarte et al., 2015
Antiretroviral agents	Atazanavir Fosamprenavir Indinavir Nelfinavir Ritonavir Saquinavir	Monzote et al., 2013
Anticoccidial agents	NPPP ^b Diclazuril Toltrazuril	Kul et al., 2013; Choi et al., 2014; Oz, 2014a,b
Anthelmintic agents	Nicosamide Nitazoxanide	Fomovska et al., 2012; Galván-Ramírez et al., 2013
Antifungal agents	Itraconazole Fluconazole Chitosan	Martins-Duarte Edos et al., 2008; Martins-Duarte et al., 2010, 2013; Gaafar et al., 2014

(Continued)

TABLE 1 | Continued

Common clinical uses	Drugs/compounds	References
Anticancer agents	SAHA ^c Pterocarpanquinone Ruthenium complexes Quinoline derivatives 4-aminoquinoline 4-piperazinyloquinoline analogs Miltefosine Tetraoxanes Gefitinib 3-bromopyruvate Tamoxifen	Strobl et al., 2007; Portes Jde et al., 2012; Leyke et al., 2012; Barna et al., 2013; Kadri et al., 2014; de Lima et al., 2015; Eissa et al., 2015; Opsenica et al., 2015; Dittmar et al., 2016
Immunosuppressants agents	Auranofin Am80 Betamethasone Pyridinylimidazole Imidazopyrimidine	Ghaffarifar et al., 2006; Wei et al., 2007; Andrade et al., 2014; Ihara and Nishikawa, 2014
Immunomodulators agents	Rolipram	Affi and Al-Rabia, 2015
Immunoregulatory agents	Levamisole	Köksal et al., 2016
Antipsychotic agents	Aripiprazole	Saraei et al., 2015
Antioxidant agents	Resveratrol	Bottari et al., 2015
Antischizophrenic agents	Haloperidol Clozapine Fluphenazine Trifluoperazine Thioridazine Amisulpride Cyamemazine Levomepromazine Loxapine Olanzapine Risperidone Tiapride	Goodwin et al., 2011; Fond et al., 2014; Saraei et al., 2016
Moodstabilizing agents	Valproate	Fond et al., 2014
Anti hypertensive agents	Guanabenz	Benmerzoug et al., 2015
Anti hypertensive and irregular heart rate agents	Propranolol	Montazeri et al., 2015, 2016

^a2-hydroxy-3-(1'-propen-3-phenyl)-1,4-naphthoquinone.

^b(4-nitrophenoxy) phenyl] propane one.

^cSuberoylanilide hydroxamic acid.

replication, etc. (Table 2). Also, drugs/compounds with known mechanisms of action on life stages of *T. gondii* are shown in Figure 2. Our collective data indicated that many of the drugs/compounds evaluated against *T. gondii* act on the apicoplast. Therefore, the apicoplast represents as a potential drug target for new chemotherapy.

The Investigated Strains

T. gondii has three main clonal lineages in population structure; type I (including a highly virulent RH strain), Type II (including ME49 and PRU, avirulent strains), and Type III (including avirulent strains like NED), which is correlated with virulence expression in mice (Howe and Sibley, 1995).

TABLE 2 | Drugs/compounds with pathways/ mechanisms of action against *T. gondii*.

Pathway/mechanism of action	Drugs/compounds	References
Electron transport chain	PHNQ6 ^a HDQ ^b Atovaquone* Endochin-like quinolones* Ferrocenic atovaquone derivatives Naphthoquinones Toltrazuril 3-Bromopyruvate	Baramée et al., 2006; Ferreira et al., 2006, 2012; Saleh et al., 2007; Meneceur et al., 2008; Bajohr et al., 2010; Doggett et al., 2012; Kul et al., 2013; de Lima et al., 2015
Sterol biosynthesis	Novel quinclidine (ER119884, E5700)	Martins-Duarte et al., 2006
Synthesis of cholesterol	Am80*	Ihara and Nishikawa, 2014
Antifolate	Pyrimethamine* Sulfadiazine* Dihydrotriazine* (JPC-2067-B, JPC-2056)	Meneceur et al., 2008; Mui et al., 2008; Martins-Duarte et al., 2013
Calcium-dependent protein kinase 1	1 NM-PP1* Bumped Kinase Inhibitor 1294* Imidazo [1,2-b] pyridazines* Compound 32*	Sugi et al., 2011; Doggett et al., 2014; Moine et al., 2015b; Vidadala et al., 2016
Human mitogen-activated protein kinase	Pyridinylimidazole* Imidazopyrimidine*	Wei et al., 2007
Nucleoside triphosphate hydrolase (NTPase)	2-(Naphthalene-2-ylthio)-1H indole*	Asgari et al., 2013, 2015
Isoprenoid pathway	2- alkylaminoethyl- 1,1- bisphosphonic acids* Newly synthesized bisphosphonates* Atorvastatin*	Shubar et al., 2008; Szajman et al., 2008; Li et al., 2013
Type II fatty acid synthesis	Thiolactomycin* 53 novel compounds* Inhibitors of enoyl reductase Triclosan and triclosan liposomal*	Martins-Duarte et al., 2009; Tipparaju et al., 2010; El-Zawawy et al., 2015a,b
Protein synthesis	Azithromycin* Spiramycin* Spiroindolone 3-aminomethyl benzoxaborole (AN6426)	Costa et al., 2009; Franco et al., 2011; Chew et al., 2012; Zhou et al., 2014; Palencia et al., 2016
Disappearance of the Apicoplast	Quinoline derivatives* (MC1626, quinoline, 8-hydroquinoline and B23)	Smith et al., 2007; Kadri et al., 2014
Histone deacetylase enzyme	SAHA ^c SBHA ^d Scriptaid* Trichostatin A* Di-cationic pentamidine-analog* FR235222, FR235222 derivative*	Strobl et al., 2007; Maubon et al., 2010; Kropf et al., 2012
DNA synthesis	Metronidazole* Phenylsemeicarbazone*	Liesen et al., 2010; Chew et al., 2012; Gomes et al., 2013

(Continued)

TABLE 2 | Continued

Pathway/mechanism of action	Drugs/compounds	References
	Phenylthiosemicarbazones* Thiosemicarbazides* 4-Thiazolidinones* 1,3,4-thiadiazoles*	
Cyclic AMP signaling pathways	Rolipram*	Afifi et al., 2014; Afifi and Al-Rabia, 2015
Post-translational modification by N-linked glycosylation of proteins	Tunicamycin*	Luk et al., 2008
Membrane permeability	Novel diamidine analog*	Leepin et al., 2008
Microfilament functional	Cromolyn sodium Ketotifen Propranolol Oryzalin analogs	Endeshaw et al., 2010; Rezaei et al., 2014; Montazeri et al., 2015, 2016
Micronemal secretion pathway, cysteine protease	Peptidyl vinyl sulfone compounds* (LHVS and ZL3VS)	Teo et al., 2007
Immuno-regulatory	Levamisole*	Köksal et al., 2016
Translational control	Guanabenz* Fusidic acid Halofuginone*	Payne et al., 2013; Benmerzouga et al., 2015; Jain et al., 2015
DNA gyrase activity, transcription	Enrofloxacin Ciprofloxacin derivatives*	Barbosa et al., 2012; Martins-Duarte et al., 2015
Thioredoxin reductase	Auranofin	Andrade et al., 2014
Topoisomerases I and II HSP90 protein	Harmene, norharmene, and harmine	Alomar et al., 2013
Metabolism of neurotransmitters in the brain	Resveratrol	Bottari et al., 2015
Effect on the liver biochemical parameters	ATT-5126 and KH-0562	Choi et al., 2014
Vascular ATP synthase subunit C and/or methyltransferase	NPPP	Choi et al., 2015
Sterol biosynthesis enzyme-sterol methyl transferase.	22, 26-azasterol and 24, 25-(R, S)-epiminolanosterol	Martins-Duarte et al., 2011
Downregulates expression of serine/threonine protein phosphatase	Diclazuril	Oz, 2014a,b
Ergosterol synthesis	Fluconazole Itraconazole	Martins-Duarte Edos et al., 2008; Martins-Duarte et al., 2013
Interruption of mitosis	Trifluralin	Wiengcharoen et al., 2007
Oxidative phosphorylation	Niclosamide	Fomovska et al., 2012
Apocynin-dependent pathway	NSC3852	Strobl et al., 2009
Phospholipid metabolism	Miltefosine	Eissa et al., 2015

(Continued)

TABLE 2 | Continued

Pathway/mechanism of action	Drugs/compounds	References
Quinone oxidoreductase expression	Nitaxozanide	Galván-Ramírez et al., 2013
Kinase inhibitors	Small-molecules	Kamau et al., 2012
Tyrosine kinase	Gefitinib Crizotinib	Yang et al., 2014
Adenosine kinase in the purine salvage pathways	N6-benzyladenosine analog*	Kim et al., 2007; Szajman et al., 2008
Purine nucleoside phosphorylase	3-(thiophen-2-yl)-1,2,4-triazole-5-thione	Dzitko et al., 2014b
Damage on the microneme proteins	7-nitroquinoxalin-2-ones (VAM2-2)	Fernández et al., 2016

*Drugs/compounds with known pathway/mechanisms of action against *T. gondii*.

^a2-hydroxy-3-(1'-propen-3-phenyl)-1,4-naphthoquinone.

^b1-hydroxy-2-dodecyl-4 (1H) quinolone.

^cSuberoylanilide hydroxamic acid.

^dSuberic bishydroxamic acid.

In vitro and *in vivo* screening methods were used of type I *T. gondii* (mostly RH strain; 76 studies *in vitro*, and 36 *in vivo*). Because type I RH strain is highly virulent in mice, causing 100% mortality, but types II and III are relatively less virulent. Although in some studies, ME49 (7 studies *in vitro*, and 17 *in vivo*), Prugniaud, EGS, and VEG strains were used, which showed that the outcome of infections depends on the challenge dose and on the genotype of the host (Szabo and Finney, 2016). Details about the investigated strains *in vitro* and *in vivo* are shown in Tables 3, 4, respectively.

Cell Culture

The cell cultures used in *in vitro* studies were mostly human foreskin fibroblast (HFF; 39 studies), LLCMK2 (12 studies), Vero (11 studies), HeLa (6 studies), mouse macrophage cell line (J774A.1) (5 studies), and MRC-5 (2 studies; Table 3).

Laboratory Animals

T. gondii can infect most warm-blooded animals, and is studied in different animal models depending on the nature of the investigation (Szabo and Finney, 2016). The animal model used in studies was mostly mice (16 studies BALB/c and 19 studies Swiss-Webster). In murine models of acute toxoplasmosis, some medicines were protective even when administered at low dosages. But some drugs despite of their excellent *in vitro* activity were poorly protective in murine models with acute toxoplasmosis (Payne et al., 2013).

Diagnostic Tests and Evaluation Methods

The present review outlines the results of *in vitro* screening methods including morphological assay, incorporation of [3H] uracil assay, plaque assays, enzyme-linked immunosorbent assay (ELISA), colorimetric micro titer assay (b-galactosidase assay), flow cytometric quantification assay, and cell viability assay. Numerous versions of fluorescent proteins have been expressed in *T. gondii* (Kim et al., 2001). The reporter genes used *in vitro*

and *in vivo* studies were the green fluorescent protein (GFP) and yellow fluorescent protein (YFP). Parasites expressing fluorescent proteins can also be analyzed and sorted by flow cytometry. This technology used for drugs screening in 10 studies.

Details about the diagnostic methods and drug dosage under *in vivo* conditions are shown in Table 4. Also, a comprehensive list of drugs/compounds evaluated against *T. gondii* with regard to IC₅₀ is illustrated in Table 5.

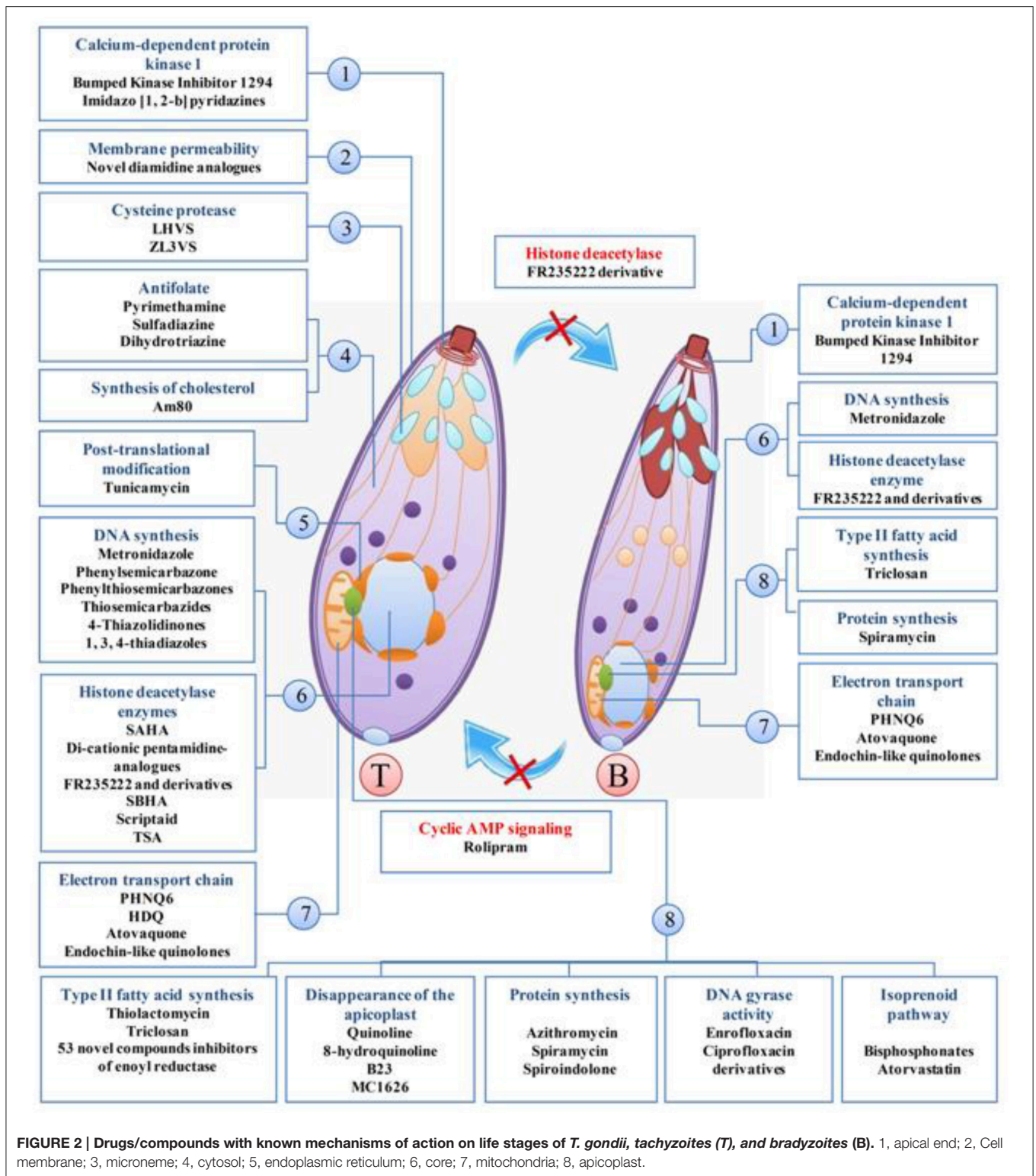
DISCUSSION

The aim of this systematic review was to investigate the *in vitro* and *in vivo* effects of anti-Toxoplasma drugs and synthetic compounds, from 2006 to 2016. The current anti-*T. gondii* chemotherapy is deficient; as it is not well-tolerated by immunocompromised patients and cannot completely eradicate tissue cysts produced by the parasite (Rodriguez and Szajman, 2012). Therefore, developing new, safe, effective, and well-tolerated drugs with novel mechanisms of action could be a global priority (Lai et al., 2012). An ideal drug for prophylaxis and/or treatment of toxoplasmosis would show effective penetration and concentration in the placenta, transplacental passage, parasitocidal properties vs. the different parasitic stages, penetration into cysts, and distribution in the main sites. No available drug fulfills these criteria (Derouin et al., 2000; Montoya and Liesenfeld, 2004).

Thus, the findings of the present systematic review article encourage and support more accurate investigations for future to select new anti-Toxoplasma drugs and strategies in designing new targets with specific activity against the parasite.

Activities of Anti-Toxoplasma Clinically Available Drugs

With growing parasite resistance to therapeutic drugs and in the absence of a vaccine, to increase the



effectiveness of drugs, various changes have been made in construction of the clinically available medicines. Thus, the activity of new formulations of clinically available drugs against *T. gondii* should be evaluated to find

alternative treatments for toxoplasmosis (da Cunha et al., 2010).

Interestingly, encapsulation of pyrimethamine improved the efficacy and tolerability of this drug against acute toxoplasmosis

TABLE 3 | Summary of in vitro studies evaluated the anti-Toxoplasma activity of drugs/compounds.

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
1	Two novel quinucidine (ER119884, E5700)	RH	LLCMK2	24, 48 h	IC ₅₀ values ^a	IC ₅₀ ER119884, E5700 = 0.66, 0.23 μM	Effective	Sulfadiazine, pyrimethamine	Martins-Duarte et al., 2006
2	Fourteen novel ferrocenic atovaquone derivatives	76K, PLK, A to R	HFF	48 h	IC ₅₀ values	IC ₅₀ 2d, 2e, 2f = 5.0, 2.5, 6.25 μM	Effective 2d, 2e, 2f	-	Barames et al., 2006
3	Betamethasone and IFN-γ ^b	RH	Hela	24, 48, 72 h	Counting the number of tachyzoites	High number of plaques was seen in group with 40 μg/ml of betamethasone.	Betamethasone not effective, IFN-γ effective	-	Ghafarifar et al., 2006
4	Suberonylanilide hydroxamic, suberic bishydroxamic acid, scriptaid, trichostatin A	RH	HS68 HFF	48, 72 h	IC ₅₀ values	IC ₅₀ scriptaid = 0.039 μM	Scriptaid was the most effective	-	Strobl et al., 2007
5	RWJ67657, RWJ64809 ^c , RWJ68198 ^d	RH, ME49	HFF	48 h	IC ₅₀ values	RWJ67657 was at least as potent as RWJ68198, SB203580, or SB202190 in reducing of <i>T. gondii</i> replication	RWJ67657, SB203580 effective	-	Wei et al., 2007
6	Novel drug compounds (A-I) (B, F, G, H) (trifluralin analogs)	RH	Vero	72 h	MTT assay ^e , crystal violet assay	IC ₅₀ drug F = 10 μM	Drugs F was the most effective	-	Wiengcharoen et al., 2007
7	1-hydroxy-2-dodecyl-4(1H) quinolone (HDQ)	RH	HFF	24 h	Replication rate determined	IC ₅₀ HDQ = 0.0024 ± 0.0003 μM	Effective	-	Saleh et al., 2007
8	Quinoline derivative MC1626	RH	HFF	24 h	Standard [³ H]uracil uptake and plaque assays	100 μM reducing growth	Effective	-	Smith et al., 2007
9	N6-benzyladenosine analogs	RH	HFF	24 h	MTT assay	IC ₅₀ N6-(2,4-dimethoxybenzyl) Adenosine = 8.7 ± 0.6 μM, exhibited the most favorable activity	Effective	Sulfadiazine, pyrimethamine	Kim et al., 2007
10	Fluorine-containing aryloxyethyl thiocyanate derivatives	RH	HFF	24 h	IC ₅₀ values	IC ₅₀ compounds 1 and 3 = 2.80 and 3.99 μM	Effective	Atovaquone	Liñares et al., 2007
11	LHVS, ZL3VS ^f	RH or 2F1	HFF	45 min	B gal ^g , Red/green invasion assay, SDS-PAGE, immunoblotting, gliding motility assay	IC ₅₀ LHVS and ZL3 VS = 10 and 12.5 μM	Effective	3,4-dichloroisocoumarin	Teo et al., 2007
12	1,25(OH) 2D3	RH	MICc12	72 h	Trypan blue assay	Ruled out any toxic effects of 1,25(OH) 2D 3 for <i>T. gondii</i>	Effective	-	Rajapakse et al., 2007
13	Tunicamycin	RH	HFF	2, 24, or 48 h	Fluorescence and electron microscopy	N-Glycosylation is completely inhibited by treatment of parasites with tunicamycin	Effective	Pyrimethamine	Luk et al., 2008
14	Novel diamidine analogs	RH	Vero HFF	2 or 3 days	IC ₅₀ values, Q-PCR ^h	IC ₅₀ DB750, DB786 = 0.16, 0.22 μM	Effective	-	Leepin et al., 2008
15	Pyrimethamine, sulfadiazine, and atovaquone	17 strains <i>T. gondii</i>	THP-1 MRC-5	7 days	IC ₅₀ , real-time PCR	IC ₅₀ pyrimethamine = 0.0002, 0.01 μM IC ₅₀ atovaquone = 0.0001, 0.00005 μM IC ₅₀ sulfadiazine = 0.01, 0.07 μM for	Effective	-	Meneceur et al., 2008

(Continued)

TABLE 3 | Continued

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
16	Novel triazine JPC-2067-B	RH	HFF	3 days	Liquid scintillation counting	13 strains and were > 0.1 µM for three strains IC ₅₀ JPC-2067-B = 0.02 µM, IC ₉₀ JPC-2067-B = 0.05 µM	Effective	-	Mui et al., 2008
17	Newly synthesized bisphosphonates (15 new compounds)	RH	Mouse macrophages (J 744A, 1)	24, 48 h	MTT assay, flow cytometry	91A and 282A showed moderate and low toxicity (cell viability between 70% and 100%) IC ₅₀ compound 19 = 2.6 µM	Effective	-	Shubar et al., 2008
18	2-alkylaminoethyl-1,1-bisphosphonic acids	RH	HFF	Daily	IC ₅₀ values, radiometric assay		Compound 19 was very effective		Szajman et al., 2008
19	Itraconazole	RH	LLCMK2	24 or 48 h	IC ₅₀ values, TEM ¹ analysis	IC ₅₀ = 0.11, 0.05 µM for 24, 48h	Effective	-	Martins-Duarte Edos et al., 2008
20	Thiolactomycin analogs (8 new compounds)	RH	LLCMK2	24, 48 h	IC ₅₀ values, Lipid extraction, chromatographic analysis	IC ₅₀ compounds = 1.6-29.4 µM	Compound 5 was very effective	Sulfadiazine, pyrimethamine	Martins-Duarte et al., 2009
21	NSC3852	RH	HS 68 HFF	2 h	SYBR green assay, MTS assay, ROS assay, NO assays	EC ₅₀ NSC3852 = 0.08 µM,	NSC3852, NSC74949 were the most effective	-	Strobl et al., 2009
22	FR235222, FR235222 derivative compounds (W363, W371, W399, W406, W425)	RH, PRU (type I)	HFF	24 h	EC50 determination, Western blot analysis, immunofluorescence microscopy	EC ₅₀ NSC74949 = 0.6 µM 100% altered cysts 24 h after treatment with the lowest concentration of FR235222	Effective	-	Maubon et al., 2010
23	Thiosemicarbazides, 4-thiazolidinones and 1,3,4-thiadiazoles	RH	Vero	24 h	Mean number of intracellular parasites, LD ₅₀ ^k	A significant decrease in the percentage of infected cells and in the mean number of tachyzoites per cell from the concentrations of 0.1, 1, 10mM	Effective	Hydroxyurea, sulfadiazine	Liesen et al., 2010
24	FLZ ¹ and ITZ ^m	RH	LLCMK2	24, 48 h	IC ₅₀ values	IC ₅₀ FLZ = 8.9, 3.1 µM after 24, 48h IC ₅₀ ITZ = 0.1, 0.05 µM for 24, 48h	Effective	Sulfadiazine, pyrimethamine	Martins-Duarte et al., 2010
25	1-Hydroxy-2-Alkyl-4(1H) Quinolone Derivatives	RH (type I)	HFF	24 h	IC ₅₀ values	IC ₅₀ compound A, B = 0.0004, 0.0008 µM	Effective	Atovaquone	Bajohr et al., 2010
26	Oryzalin Analogs	RH	HFF	8 day 26 h	Plaque assay, immunofluorescence assay, IC ₅₀ values	IC ₅₀ 18b = 0.03 µM	Effective	-	Endeshaw et al., 2010
27	53 novel compounds (Inhibitors of Enoyl reductase)	RH	HFF	3 days	IC ₅₀ values	IC ₅₀ compounds 2, 19 = 0.04, 0.02 µM, IC ₅₀ compounds 39 less active	Compounds 2, 19, 39 greatest effect	-	Tipparaju et al., 2010
28	Haloperidol, clozapine, fluphenazine, trifluoperazine, thioridazine	RH	HFF	48 h	IC ₅₀ values	IC ₅₀ fluphenazine, thioridazine, trifluoperazine = 1, 1.2, and 3.8 µM	Fluphenazine, thioridazine, trifluoperazine were effective	-	Goodwin et al., 2011

(Continued)

TABLE 3 | Continued

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
29	Azithromycin, spiramycin	RH	Bewo cell line	24 h	MTT assay, measurement of Th1/Th2	Increase TNF- α , IL-10, IL-4 production, but decreased IFN- γ	Effective	-	Franco et al., 2011
30	Novel azasterols	RH ME49	LLCMK2	24 or 48 h	IC ₅₀ values, immunofluorescence assays	IC ₅₀ compounds 1, 2, 3 = 0.8–4.7 μ M	Compound 3 was the most effective	-	Martins-Duarte et al., 2011
31	Ciprofloxacin derivatives	RH	LLC-MK2	24 or 48 h	IC ₅₀ -MTS assay	IC ₅₀ compounds 2, 4, 5 = 0.42, 1.24, and 0.46 μ M	Effective	-	Dubar et al., 2011
32	2-hydrazolyl-3-phenyl-5-(4-nitrobenzylidene)-4-thiazolidinone substituted	RH	Vero	24 h	LD ₅₀ values	LD ₅₀ = 0.5, 10 mM	Effective	Hydroxyurea, Sulfadiazine	Aquino et al., 2011
33	Nanoparticles	RH (CAT-GFP)	Macrophages J774-A1	3 day	HPLC ^o : flow cytometry	Ca ²⁺ .85% observed maximum in Toxoplasmosis therapy efficiency	Effective	-	Leyke et al., 2012
34	Enrofloxacin	RH	HFF	72 h	MTT assays	Enrofloxacin resulted in a significant inhibition of the percentage of infected cells by the parasite (68.72%)	Effective	Sulfadiazine, pyrimethamine	Barbosa et al., 2012
35	ELQ-271 and ELQ-316 ⁹	2F	HFF	4 days	Host-cell toxicity	IC ₅₀ ELQ-271, ELQ-316 = 0.0001, and 0.000007 μ M	Effective	Atovaquone	Doggett et al., 2012
36	Pterocarpanquinone	RH	LLCMK2	24 or 48 h	Direct counts, viability, immunofluorescence assays	IC ₅₀ = 2.5 μ M	Effective	-	Portes Jde et al., 2012
37	New naphthoquinones and an alkaloid	RH, EGS	HFF	48 h	MTT assays	IC ₅₀ QUI-5, and QUI-6 ⁷ = 69.35, and 172.81 μ M	Effective	Atovaquone, Sulfadiazine	Ferreira et al., 2012
38	Spiramycin coadministered with metronidazole	ME49	Vero E6	1 week	Numbers of cysts and tachyzoites	Spiramycin reduced <i>in vitro</i> reactivation, metronidazole alone did not have significant effect	Effective	-	Chew et al., 2012
39	Di-cationic pentamidine-analogs	RH ME49	HFF	72 h	Cytotoxicity assays	IC ₅₀ arylimidamide DB745 = 0.11, 0.13 μ M (tachyzoites of Rh, Me49)	Effective	Atovaquone	Kropf et al., 2012
40	Small-Molecule (n=527)	Strains 5A10 (type III strain)	HFF	72 h	Luciferasebased assay, Host cell viability, electron microscopy, invasion, motility assays	EC ₅₀ s for the 14 compounds = 0.14–8.7 μ M	14 compounds effect	-	Kamau et al., 2012
41	Salicylic acids (39 compounds)	RH, RH-YFP, and ME49	HFF	1 h	[³ H]-Uracil incorporation and YFP Fluorescence assay	3i, 3j, 7a, 14a, and 14b were active at low nanomolar concentrations	Effective	Pyrimethamine, Sulfadiazine	Fomovska et al., 2012
42	FLZ combined with sulfadiazine and pyrimethamine	RH	LLCMK2	24 h	IO50 values and MTS assay	IC ₅₀ FLZ = 8.4 \pm 1.2, IC ₅₀ sulfadiazine/pyrimethamine, pyrimethamine = 8.7 \pm 0.8 μ M	Effective	-	Martins-Duarte et al., 2013

(Continued)

TABLE 3 | Continued

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
43	Harmaline, Norharmaline (β -carboline alkaloids)	RH	Vero HFF	1, 24h	Parasite invasion and replication rate	harmaline and harmine showed 2.5- to 3.5-fold decrease in the invasion rates at doses of 40 μ M, norharmaline 2.5 μ M	Effective	Sulfadiazine	Alomar et al., 2013
44	Fusidic acid	Prugniaud	HFF	7 days	Lytic plaques counted	IC ₅₀ = 7.7 μ M, decreased the number of <i>T. gondii</i> plaques in a dose-dependent manner	Effective	-	Payne et al., 2013
45	Two naphthalene-sulfonyl-indole compounds	RH	-	1.5h	Stained by PI, analyzed by FACS	LD ₅₀ compound A, B = 62, 800 μ mol	Effective	Saponin	Asgari et al., 2013
46	(Benzaldehyde)-4-phenyl-3-thiosemicarbazone, (benzaldehyde)-(4 or 1)-phenylsemicarbazone (9 compounds)	RH	Vero	24h	Cytotoxicity, number of intracellular parasites	LD ₅₀ compound 8 = 0.3 mM, reduced the number of intracellular parasites by 82 % in a concentration of 0.01 mM	Effective	Sulfadiazine	Gomes et al., 2013
47	Ivermectin and sulphadiazine	RH	Hep-2	24, 48, 72h	IC ₅₀ , invert microscopy, ELISA assay	IC ₅₀ ivermectin and sulphadiazine = 0.2, and 29.1 μ M	Effective	-	Bligh et al., 2013
48	Novel ruthenium complexes	RH	HFF	72h	cytotoxicity assessment, TEM	EC ₅₀ compounds 16, 18 = 18.7, 41.1 nM	Compounds 16, and 18 effective	-	Barna et al., 2013
49	Atazanavir, fosamprenavir, indinavir, nelfinavir, ritonavir, and saquinavir	RH	Macrophages Swiss Webster	48h	IC ₅₀ determination, MTT assay	IC ₅₀ atazanavir ritonavir, and saquinavir = > 1 μ M	Effective	Pyrimethamine	Monzote et al., 2013
50	Atorvastatin	RH	HFF	8 days	IC ₅₀ values	IC ₅₀ fosamprenavir, and nelfinavir = > 5 μ M	Effective	-	Li et al., 2013
51	Nitazoxanide	RH	Astrocyte	24, 48h	Immunocytochemical method, microscopic analysis, viability	IC ₅₀ = 50 μ M	Effective	Pyrimethamine	Galván-Ramirez et al., 2013
52	Amisulpride, cyamemazine, fluphenazine, haloperidol, levomepromazine, loxapine, olanzapine, risperidone, tiapride, and valproate	RH	HFF	4h	Growth inhibition assay	Nitazoxanide produced 97% <i>T. gondii</i> death in a concentration of 10 mg/mL in 48 h infected astrocytes	Zuclopenthixol, high effective	-	Fond et al., 2014
53	Spiroindolone	RH	HFF	72h	Fluorescence assays, cytotoxicity assessment	Amisulpride, tiapride and valproate did not have inhibitory activity	Effective	Pyrimethamine, sulfadiazine	Zhou et al., 2014
54	Auranofin	RH	HFF	5 days	Invasion and replication assays and plaque assays	IC ₅₀ = 1 μ M	Effective	Pyrimethamine, Sulfadiazine	Andrade et al., 2014
55	Azithromycin	2 F1	Placental tissues	48h	Production of cytokines and hormones	TD ₅₀ = 8.21 μ M, IC ₅₀ = 0.28 μ M	Effective	Pyrimethamine, Sulfadiazine, folic acid	Castro-Filice et al., 2014
56	6-Trifluoromethyl-2-thiouracil (ATT-5126), (KH-0562)	RH	Hela	24h	MTS assay, IC ₅₀	Increases IL-6 production, reduced secretion of estradiol, progesterone, and HCG + β IC ₅₀ ATT-5126, KH-0562 = 19.7, 32.2 μ M	Effective	Pyrimethamine	Choi et al., 2014
						CC ₅₀ ATT-5126, KH-0562 = 35.4, 56.3 μ M			

(Continued)

TABLE 3 | Continued

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
57	Cromolyn sodium and ketotifen	RH	Macrophage monolayer	24 h	Inhibition rate	After 60 min the best efficacy was observed at 15 µg/ml (78.9 ± 1.70, 91.97 ± 0.37%)	Effective	-	Rezaei et al., 2014
58	200 drug-like and 200 probe-like compounds of Malaria Box	TS-4 (mutant of the RH)	HFF	24 h	Cytotoxicity assays	Seven compounds with IC ₅₀ < 5 µM, SI > 6	7 compounds effected	Pyrimethamine, sulfadiazine	Boyom et al., 2014
59	Am80	RH, PLK, its recombinants	J774A.1	20 h	Uracil incorporation assay, RT-PCR ^s , flow cytometry	Am80 inhibited parasite growth by decreasing intracellular accumulation of cholesterol	Effective	-	Inara and Nishikawa, 2014
60	Pyrimethamine-loaded lipid-core nanocapsules	RH	LLC-MK2	72 h	MTS assay	TC ₅₀ PYR loaded lipid-core nanocapsules = 6.0 µM	Effective	-	Pissinate et al., 2014
61	Quinoline derivatives (58 compounds)	2F	HFF	4 days	Cytotoxicity assays	IC ₅₀ B23 = 0.4 ± 0.03 µM, the most effective compound	32 compounds effected	-	Kadri et al., 2014
62	74 novel thiazolidin-4-one derivatives	-	HFF	5 days	Cytotoxicity assays	IC ₅₀ derivatives 12 A, 27 A = 0.9, 2.9 µM	Effective	Timethoprim	D'Ascenzo et al., 2014
63	Gefitinib and Crizotinib	RH	HeLa	24, 48, 72 h	Counting the number of <i>T. gondii</i> per parasitophorous vacuolar membrane	Gefitinib inhibited the growth of <i>T. gondii</i> over 5 µM whereas Sunitinib did not	Gefitinib effected	Pyrimethamine	Yang et al., 2014
64	1,4-disubstituted thiosemicarbazides	RH	Mouse L929 fibroblasts	24 h	MTT assay and q-pcr	19, 2b, 3d, 3l showed significant anti-parasitic effects	1 g was very effective	Sulfadiazine	Dzitko et al., 2014a
65	3-(thiophen-2-yl)-1,2,4-triazole-5-thione	RH	Mouse L929 fibroblasts	24 h	IC ₅₀ values and q-pcr	IC ₅₀ at least 30 times better than that of sulfadiazine	Effective	Sulfadiazine	Dzitko et al., 2014b
66	1-[4-(4-nitrophenoxy)phenyl]propane-1-one (NPPP)	RH	HeLa	24 h	CC ₅₀ , EC ₅₀ values	EC ₅₀ , CC ₅₀ = 36.2 ± 0.2, 67.0 ± 0.2 µM	Effective	-	Choi et al., 2015
67	C-type lectin from <i>Bothrops pauloensis</i> venom	RH	HeLa	24 h	MTT assay, cytokine measurements	MTT assay between 0.195, 12.5 µg/mL MIF, IL-6 productions were increased	Effective	-	Castanheira et al., 2015
68	Ciprofloxacin derivatives Compounds (2, 4, 5)	RH	LLCMK2 HFF	24, 48, 72 h	Immunofluorescence, TEM	Inhibited parasite replication early in the first cycle of infection	Effective	-	Martins-Duarte et al., 2015
69	New chiral N-cysulfonamide bis-oxazolidin-2-ones	RH	MRC-5	6 h	IC ₅₀ values	IC ₅₀ of Mol 1 was less than Mol 2	Effective	Sulfadiazine	Meriem et al., 2015
70	Guanabenz	ME49 Prugnialud	HFF	32 h	EC ₅₀ values	EC ₅₀ = 6 µM	Effective	-	Benmeizouga et al., 2015
71	3-Bromopyruvate, Atovaquone	RH	LLC-MK2	24, 48 h, or 6 days	Light-microscopic analysis, indirect immunofluorescent assays	73 and 71% reduction in intracellular parasites after 24, 48 h	Effective	-	de Lima et al., 2015
72	Biphenylimidazoazines	RH	HFF	96 h	EC ₅₀ values and fluorescence microscopy assay	EC ₅₀ < 1 µM	Effective	Pyrimethamine	Moine et al., 2015a
73	Halofuginone	RH	HFF	24 h	EC ₅₀ values	EC ₅₀ = 0.94 nM	Effective	Pyrimethamine	Jain et al., 2015
74	Naphthoquinone derivative	RH	LLC-MK2	24, 48 h	IC ₅₀ and MTT assay	IC ₅₀ LQB 151 = < 1 µM	Effective	-	da Silva et al., 2015

(Continued)

TABLE 3 | Continued

No	Drug	Strain	Cells	Culture	Evaluation	Main results	Effectivity	Positive control	References
75	Aryloxyethyl thiocyanates	RH	Vero	24 h	Determination of ED ₅₀	ED ₅₀ derivatives 15 and 16 = 1.6 μM and 1.9 μM	Effective	-	Chao et al., 2015
76	Imidazo [1,2-b] pyridazines derivatives	RH-GFP	HFF	24 h	Cytotoxicity assay	EC ₅₀ 16a, 16f = 100, 70 nM	Effective	-	Moine et al., 2015b
77	Nitrofurantoin	RH	HeLa	24 h	MTS assay	Selectivity = 2.3 EC ₅₀ = 14.7 μM	Effective	Pyrimethamine	Yeo et al., 2016
78	Quinoxaline derivatives	RH	HEP-2 ^t	24 h	IC ₅₀ values, viability, invasion, and intracellular growth	MIC ₅₀ VAM2-2 = 3.3 ± 1.8 μM	VAM2-2 was very effective	-	Fernández et al., 2016
79	1120 compounds	RH-GFP	HFF	72 h	Parasite invasion, Microneme secretion, Luciferase, and LC3-GFP assays	94 compounds with IC ₅₀ < 5 μM	Tamoxifen effective	-	Dittmar et al., 2016
80	3-aminomethyl benzoxaborole (AN6426)	RH	HFF	24 h	Determination of EC ₅₀	EC ₅₀ = 76.9 μM	Effective	Pyrimethamine	Palencia et al., 2016
81	Sulfur-containing linear bisphosphonates	RH, Prugnard	Human fibroblasts (hTert cells)	5 days	Determination of EC ₅₀	EC ₅₀ = 0.11 ± 0.02 μM	Compound 22 was very effective	-	Szajman et al., 2016
82	Fluorine-containing Analogs of WC-9 (4-phenoxyphenoxyethyl thiocyanate)	RH	Vero	24 h	Determination of EC ₅₀	EC ₅₀ 3-(3-fluorophenoxy), 3-(4-fluorophenoxy) phenoxyethyl thiocyanates, and 2-[3-(phenoxy)phenoxyethylthio]ethyl-1,1-bisphosphonat = 1.6 4.9 and 0.7 μM	Effective	-	Chao et al., 2016
83	6-(1,2,6,7-tetraoxaspiro[7.11] nonadec-4-yl)hexan-1-ol (N-251)	RH	Human hepatocyte, Huh-7	72 h	IC ₅₀ values, q-por, ultrastructural Change by TEM	LC ₅₀ = 1.11 μg/ml	Effective	Sulfadiazine	Xin et al., 2016

^aHalf maximal inhibitory concentration.

^bInterferon gamma.

^cPyridinylimidazole.

^dImidazopyrimidine.

^e3-(4, 5-Dimethyl-2-Thiazyl) - 2, 5-Diphenyl-2H-Tetrazolu Bromide.

^fMorpholinourea-leucyl-homophenolalaninyl-phenyl-vinylsulfone, N-benzoylcarbonyl-(leucyl) 3-phenyl-vinyl-sulfone.

^gB galactosidase.

^hQuantitative polymerase chain reaction.

ⁱTransmission electron microscopy.

^j5-nitroso-8-quinolind.

^kLethal Dose, 50%.

^lFluconazole.

^mItraconazole.

ⁿTumor necrosis factor

^oHigh Performance Liquid Chromatography.

^pCarriers achieved.

^qEndochin-like quinolones.

^r7-(4-methyl-3-pentenyl)-2-pyrrolidone-[1, 4]-naphthoquinone (QUI-5), 6-(4-methyl-3-pentenyl)-2-pyrrolidone-[1, 4]-naphthoquinone (QUI-6).

^sReverse transcription polymerase chain reaction.

^tHuman larynx epidermoid carcinoma epithelial cells.

TABLE 4 | Summary of in vivo studies evaluated the anti-Toxoplasma activity of drugs/ compounds.

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
1	PHN06 ^a alone or combined with sulfadiazine	Female Swiss mice	RH EGS P	Acute, chronic	1000 tachyzoites (ip) 10 brain cysts (orally)	PHN06 50 mg/kg/day Sulfadiazine, 40 mg/L	Survival rates, IFAT ^b , and liver histology	Treatment protected at least 70, 90% of mice infected with RH and EGS strains	Effective	Sulfadiazine	Ferreira et al., 2006
2	1, 25(OH) 2D3	BALB/c	ME49	Acute	20 cysts	0.5 µg/kg/2 days ip	Histopathology, RT-PCR ^c	Low parasitic burdens were found	Effective	-	Rejapakse et al., 2007
3	Pyridinylimidazole (RWJ67667, RWJ64809), imidazopyrimidine (RWJ68198)	Female CBAXJ, CD81/-	RH ME49	Acute	1000, 100, and 20 tachyzoites	3.8, 7.5, 15, 30, or 60 mg/kg ip	Survival rates	The highest dose (60 mg/kg) significantly improved survival	RWJ67667 effective	-	Wei et al., 2007
4	Novel triazine JPC-2067-B	Outbred Swiss Webster	RH	Acute	10000 tachyzoites ip	1.25 mg/kg/day orally	Peritoneal <i>T. gondii</i> burden	Intraperitoneal parasite numbers were reduced	Effective	-	Mui et al., 2008
5	Newly synthesized bisphosphonates	NMIRI	RH	Acute	100 000 tachyzoites ip	490, 1000, 512, 44.05, and 47.6 µM	Flow cytometry	Therapeutic efficacy was 100% for bisphosphonates 2F 3B, 18A, 22A, and 30B	Effective	-	Shubar et al., 2008
6	Azithromycin, Artemisia annua, spiramycin, SPFA	Females C. callosus	ME49	Chronic	20 cysts	Azithromycin (9 mg/24 h), A. annua (1.0 mg/8 h), spiramycin (0.15 mg/8 h)	Morphological, immunohistochemical analyses, mouse bioassay, and PCR ^d	No morphological changes were seen in the placenta and embryonic tissues from females treated with azithromycin, spiramycin, and SPFA	Azithromycin more effective	-	Costa et al., 2009
7	Dihydroartemisinin and azithromycin	Kunming mice	-	Acute	2 × 10 ³ tachyzoites	Dihydroartemisinin and azithromycin 75 and 200 mg/kg	The ultrastructure of tachyzoites	The ultrastructure of tachyzoites was observed in the treatment groups such as edema, enlarged, broken or damaged	Effective	-	Yin et al., 2009
8	FLZ ^e and ITZ ^f	Outbred female Swiss	CF1 ME49	Chronic	20 cysts of the ME49 orally or ip	10, 20 mg/kg/day orally	Survival rates and brain cyst burden	ITZ survival of 90, 87% FLZ survival rate of 71, 85%	Effective	Sulfadiazine, pyrimethamine	Marins-Duarte et al., 2010
9	HDQ ^h derivatives	Female NMRI, IRF-8/-	RH ME49	Acute, chronic	10 ⁵ green fluorescent protein, i.p 10 cysts	32 mg/kg body weight/day	Parasite loads in lungs, livers by qPCR ^e , and flow cytometry analyses	Derivatives of HDQ had lower parasite concentrations than mice treated with HDQ	Effective	Atovaquone	Bejohir et al., 2010
10	FR235222 FR235222 derivative, (W363, W371, W399, W406, W425)	Outbred female Swiss	PRU	Chronic	Living cysts i.p	200 nM	Presence or absence of cysts in brain was assessed by staining	No cysts were detected in mice inoculated with FR235222-treated	Effective	Pyrimethamine	Maubon et al., 2010
11	Azithromycin combined with metronidazole	BALB/c	-	Acute	50 tissue cysts orally or ip	250, 200 mg/kg/day	Microscopical examination, bioassay were done for brain, and survival rates	Cure rate 100%	Effective	-	H-A-Jader and Al-Mukhtar, 2010
12	Novel compounds 2,19 (Inhibitors of Enoyl Reductase)	CD1	RH	Acute	2000	10 mg/kg ip	Parasite burdens in the peritoneal cavity and survival rates	Reduction of parasite burden	Effective	-	Tipparaju et al., 2010

(Continued)

TABLE 4 | Continued

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
13	SDS-coated atovaquone	C57BL/6	ME49	Acute, chronic	10 cysts orally	100 mg/kg	Histology, PCR	Parasite loads and inflammatory changes in brains were significantly reduced	Effective	-	Shubar et al., 2011
14	1NM-PP1	Old female ICR strain	RH	Acute	1.0 × 10 ⁵ tachyzoites i.p	5 μM orally	Survival rates, parasite load by qPCR	Reduced the parasite load in the brains, livers, lungs	Effective	-	Sugi et al., 2011
15	Enrofloxacin	Calomys callosus, C57BL/6	RH ME49	Acute, chronic	100 tachyzoites RH strain 20 cysts per 100/ l (orally)	Subcutaneously for 3 days, 3 mg/kg twice a week for the duration of 25-day	Histological analysis, immunohistochemical assay, survival, cyst counts	diminished significantly the tissue parasitism as well as the inflammatory alterations in the brain	Effective	Sulfadiazine, pyrimethamine	Barbosa et al., 2012
16	Small-Molecule (C1, C2, C3, C5)	BALB/c	5A10, PB3-10	Acute, chronic	10,000 tachyzoites i.p	4.4 mg/kg/day	Survival rates, recording the total number of photons per second from each mouse	C2 showed a significant reduction in parasite load in acute and reduced levels of parasite proliferation and increased survival in chronic phase	C2 effective	-	Kamau et al., 2012
17	Endochin-like quinolones (ELO-271, ELO-316)	Female CF-1, CBA/J	RH ME49	Acute, chronic	20000 tachyzoites (express YFP) i.p 18 cysts of ME49	50, 20, 5, 1 mg/kg for 5 day	Counted by flow cytometry	ED50 values of 0.14, 0.08 mg/kg reducing cyst burden by 76-88%	Effective	Atovaquone	Doggett et al., 2012
18	Spiramycin coadministered with metronidazole	Male BALB/c	ME49	Chronic	1000 tachyzoites orally	5 or 25 mg/kg for 16 day 400 mg/kg daily for 7 days	Brain cysts counted	Metronidazole increased spiramycin brain penetration, causing a significant reduction of <i>T. gondii</i> brain cysts	Metronidazole alone showed no effect	-	Ohew et al., 2012
19	New naphthoquinones, an alkaloid	Female Swiss-Webster	EGS	Chronic	10 tissue cysts orally	500 mg/kg daily 50 μg/mL of QUI-11, 100 μg/mL of either QUI-6 or QUI-11	Presence of tachyzoites in the peritoneal cavities and survival rates	The survival rates increased	Effective	Atovaquone	Ferreira et al., 2012
20	Prednisolone	Swiss albino	RH ME49	Acute, chronic	1 × 10 ⁴ tachyzoites, iP	235, 470, 705 mg/kg	Number of tachyzoites present	Greatly improved the number of tachyzoite, cyst forms in mice	No effective	-	Puvanesusarian et al., 2012
21	Salicylic acids compounds 14a, 14b	Swiss Webster	RH, RH-YFP, ME49	Acute	Oocysts oral gavage	100 or 25 mg/kg orally	Survival rates	Increased survival by 1 day	Effective	Pyrimethamine, sulfadiazine	Fomovska et al., 2012
22	Atorvastatin	Female Swiss Webster, BALB/c	RH	Acute	5-20 tachyzoites i.p 10,000-100,000 tachyzoites	20 mg/kg/day ip	Plaque assays and containing tachyzoites in peritoneal fluid	Atorvastatin protect mice against death, cures a lethal infection	Effective	-	Li et al., 2013
23	Fusidic acid	Female BALB/c	Prughiaud	Acute	5 × 10 ³ or 5 × 10 ⁴ tachyzoites i.p	20 mg/kg	Parasite burdens, analyses of host cytokine, and survival rates	There was no statistically significant difference between mice treated with fusidic acid versus saline	No effective	Trimethoprim, sulfadiazine	Payne et al., 2013

(Continued)

TABLE 4 | Continued

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
24	FLZ combined with sulfadiazine, and pyrimethamine	CF1	RH	Acute	10 ³ tachyzoites	10 mg/kg/day of fluconazole with 40/1 mg/kg/day sulfadiazine, pyrimethamine	Survival rates	93% survival	Effective	Sulfadiazine, pyrimethamine	Martins-Duarte et al., 2013
25	Two naphthalene-sulfonyl-imide compounds	BALB/c	RH	Acute	2 × 10 ⁶ tachyzoites	25-800 μmol.i.p	Survival rates, liver touch smears with giemsa stained	Both of the compounds was preserved	Effective		Asgari et al., 2013
26	Toltrazuril	lambs	ME49	Chronic	1 × 10 ⁵ oocysts	20, 40 mg/kg orally 2 times, once every week	Presence of tissue cysts by histopathology, immunohistochemistry, and nested-PCR	Cyst presence was determined as 44.4%	Effective		Kul et al., 2013
27	Auranofin	Chicken embryos	RH	Acute	1 × 10 ⁴ tachyzoites chorioallantoic vein	1 mg/kg	Histopathology, immunohistochemistry, and qPCR	Significantly reduced parasite load	Effective	Pyrimethamine, sulfadiazine	Andrade et al., 2014
28	Spiroindolone	Mice	RH	Acute	2000 tachyzoites	100 mg/kg/day	Parasite burdens, measuring the fluorescence intensity	Reduced the parasite burden in mice by 90%	Effective	-	Zhou et al., 2014
29	6-Trifluoromethyl-2-thiouraci KH-0562 ¹ and ATT-5126 ¹	ICR female	RH	Acute	1 × 10 ⁵ tachyzoites	100 mg/kg KH-0562 ¹ or ATT-5126 ¹ orally	Measuring amount of the tachyzoites in mice ascites, LPO ^k , GSH, ALT ^m , AST ^m in mouse liver	LPO level-KH-0562 and ATT-5126 = 87.4 and 105.2 nmol/g	KH-0562 more effective	Pyrimethamine	Choi et al., 2014
30	Pyrazolopyrimidine-1,2,94	BALB/c	RH Pru	Acute, chronic	10 ⁵ tachyzoites	100, 30 mg/kg/day for 5 days	Survival rates and number of <i>T. gondii</i> per ml	Decreasing the numbers of <i>T. gondii</i> tachyzoites at both 100, 30 mg/kg	Effective	-	Doggett et al., 2014
31	6-Trifluoromethyl-2-thiouraci KH-0562 ¹ , ATT-5126 ¹	Female ICR	RH	Acute	1 × 10 ⁵ tachyzoites	100 mg/kg	Proteomic profiles of <i>T. gondii</i> tachyzoites	Decreased the amount of tachyzoites, mean numbers of tachyzoites = (66.8 ± 0.8) × 10 ⁶	Effective	Pyrimethamine	Choi et al., 2014
32	Cromolyn sodium, ketotifen	Balb/c	RH	Acute	4 × 10 ⁵ tachyzoites	Ketotifen 1, 2 mg/kg, cromolyn sodium 5, 10 mg/kg, ip	Inhibition evaluated under a light microscope with giemsa staining	After 60 min ketotifen at 2 mg/kg (69.83 ± 2.25 %), cromolyn sodium, at 10 mg/kg in (80.47 ± 2/49 %) had the best effect	Effective	-	Rezaei et al., 2014
33	Diclazuril plus atovaquone	CD1 mice	PTG Strain	Chronic	600 tachyzoites-i.p	65, 120 mg/kg diclazuril	Hematoxylin eosin, Giemsa, immunohistochemical staining	Combination diclazuril plus atovaquone was safe	Effective	-	Oz, 2014b
34	Diclazuril plus atovaquone	CD1 mice	PTG strain	Chronic	300, or 600 tachyzoites.i.p	65, 120 mg/kg diclazuril	Hematoxylin and eosin, slides evaluated of colonic tissues	Combined therapy synergistically normalized pathology and to a lesser degree monotherapy	Effective	-	Oz, 2014a
35	Am80	BALB/c mice	RH, PLK	Acute	1 × 10 ³ tachyzoites.i.p	1 mg/kg	Survival rates	Percent survival of mice increased statistically	Effective	-	Ihara and Nishikawa, 2014
36	Chitosan and silver nanoparticles	Swiss albino	RH	Acute	3.5 × 10 ³ tachyzoites.i.p	100, 200 μg/ml	Parasite density and ultrastructural parasite changes	Statistically significant decrease in the mean number of the parasite count in the liver and the spleen	Effective	Pyrimethamine	Gaafar et al., 2014

(Continued)

TABLE 4 | Continued

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
37	Pyrimethamine/sulfadiazine	Female C57BL/6 mice	ME49	Chronic	20 cysts i.p	Pyrimethamine, sulfadiazine 4, 100 mg/kg daily for one month	Histology, qPCR, measured KP metabolites	Significant increases in these kynurenine pathway metabolites were observed in the brain at 28 days post-infection	Effective	-	Notarangelo et al., 2014
38	Pyrimethamine-loaded lipid-core nanocapsules	Female CF1 mice	RH	Acute	10 ³ tachyzoites	5.0–10 mg/kg/day	Surviving mice, cyst brain evaluation, bioassay urea, AST and ALP _o	Survival rate higher than the animals treated with the same doses of non-encapsulated pyrimethamine	Effective	-	Pissinatte et al., 2014
39	Atovaquone and astragalus combination	BALB/c	RH	Acute	2 × 10 ⁴ /ml trophozoites	Atovaquone, astragalus 100, 0.075 mg/kg/day oral gavage	Peritoneal trophozoite numbers, IL-2, IL-12, IFN- γ levels were determined by ELISA	The number of trophozoites in the combination groups were found significantly lower than the number of trophozoites in the control group	Effective	-	Söhmezz et al., 2014
40	Roilbram	Female Swiss albino mice	KSU strain	Chronic	20 tissue cysts	10 mg/kg daily for three weeks	Life expectancy, serum ALT, histopathology of liver and brain	Roilbram exerts a significant lowering effect on ALT levels, pathology	Partially effective	-	Alfi et al., 2014
41	Roilbram	Female Swiss albino mice	Low pathogenic strain	Chronic	20 tissue cysts	-	Tissue injury scoring, brain cyst count, specific Ig G titers, TNF- α , IFN- γ and IL-12 assays	Significant reduction of TNF α (84.6%), IFN- γ (76.7%), IL-12 (71%)	Partially effective	-	Alfi and Al-Rablia, 2015
42	Triclosan (TS) and triclosan-loaded liposomal nanoparticles	Swiss strain Albino mice	RH HXGPRT (-)	Acute	10 ⁴ tachyzoites	150 mg/kg TS or 100 mg/kg TS liposomes	Mice mortality, peritoneal, liver parasite burdens	Reduction in mice mortality, parasite burden	Effective	-	Ei-Zawawy et al., 2015b
43	Sulfamethoxazole-trimethoprim (ST) associated with resveratrol	Male Swiss albino mice	VEG strain	Chronic	50 cysts containing bradyzoites	ST (groups B, F), free resveratrol (groups C,G) 0.5, 100 mg kg ⁻¹	Cyst counts in the brain, and histopathology analyses	Combination was able to reduce the number of cysts in the brain, inflammatory infiltrates in the liver, prevented the occurrence of hepatocytes lesions	Effective	Sulfamethoxazole, trimethoprim	Botian et al., 2015
44	Ciprofloxacin derivatives (compounds 2, 4, 5)	Female Swiss mice	RH	Acute	5 × 10 ³ tachyzoites i.p	25, 50, 100, or 200 mg/kg/day a single oral dose	Survival rate, determine the serum levels of urea and creatinine kinase	Increased mouse survival significantly, with 13–25% of mice surviving for up to 60 days post infection	Effective	-	Martins-Duarte et al., 2015
45	Triclosan (TS), TS liposomal	Swiss albino mice	ME49	Chronic	10 cysts	200, 120 mg/kg	Mortality, brain parasite burden	TS significant diminution in the parasite burden, great reduction in the infectivity power of <i>T.gondii</i> cysts	Effective	-	Ei-Zawawy et al., 2015a

(Continued)

TABLE 4 | Continued

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
46	2-(Naphthalene-2-ylthio)-1H-Indole 2-(naphthalene-2-ylthio)-1H-Indole	BALB/c	RH	Acute	2×10^6 tachyzoites exposed to the concentrations of the compound i.p.	25-800 μ M for 1.5h	Surviving mice, stained by PI and analyzed by fluorescence-activated cell sorting (FACS)	The longevity of mice was dose dependent. Five mice out of group 400 μ M and 3 out of group 800 μ M showed immunization to the parasite	Effective	-	Asgari et al., 2015
47	Propranolol	BALB/c	RH	Acute	1×10^3 tachyzoites i.p	2, 3 mg/kg/day	Parasite load determined	In the pre-treatment group, propranolol combined with pyrimethamine was more effective	Effective	Pyrimethamine	Montazeri et al., 2015
48	Aripiprazole	BALB/c	Tehran strain	Chronic	50 tissue cysts, i.p	10, 20 mg/kg	Cysts counted in smears prepared from brain homogenate by optical microscope	No significant difference between mean logarithms of brain cyst numbers of aripiprazole groups compared with control	No effective	-	Saraei et al., 2015
49	Pyrimethamine (PYR) and sulphadiazine (SDZ) combined with levamisole and echinacea	BALB/c	RH	30 days after treatment	10^5 tachyzoite i.p	PYR: 6.25, 12.5 SDZ; 100, 200 PYR, SDZ, levamisole; 2.5, echinacea; 130, 260 mg/kg/day oral treatment 24 h later for 10 days	Survival rates	Survival rate PYR+SDZ, and levamisole = 33.3% to 88.9%	Effective	Pyrimethamine, sulfadiazine	Köksal et al., 2016
50	Miltefosine	Swiss albino mice	RH ME49	Acute, chronic	2500 tachyzoites i.p 10 cysts orally	20 mg/kg for 5 days	Survival rates, tachyzoites count in the liver, spleen, cyst count and size in the brain ultra structural study, and histopathological study	Survival rate in acute = 30% Survival rate in chronic = 5%	No effective in acute, Partially effective in chronic	Sulphadiazine	Eissa et al., 2015
51	Tetraoxanes	Female Swiss Webster	RH	Acute	10^2 and 10^6 tachyzoite i.p	20 mg/kg/day 60 days post infection for 15 days 10 mg/kg/day, subcutaneously for 8 days	Survival rates and pathohistological analysis	Survival rate = 20 %	Effective	-	Opšenica et al., 2015
52	Guanabenz	BALB/c	ME49 Prughaud	Acute, chronic	10^4 ME49 or 10^6 Pru tachyzoites, i.p	5 or 10 mg/kg repeated every 2 days	Survival of mice, qPCR	Enhanced survival, reduces cyst burdens in chronically infected mice	Effective	-	Benmerzouga et al., 2015
53	Fluphenazine and Thioridazine	BALB/c	Tehran strain	chronic	20 tissue cysts i.p	Thioridazine 10, 20, fluphenazine 0.06 mg/kg/ three days after inoculation for 3 weeks	The number of brain cysts	Drugs reduced the percent of cysts at higher dose compared to lower doses	Effective, not significant	Pyrimethamine	Saraei et al., 2016
54	Nitrofurantoin	Female ICR mice	RH	Acute	1×10^5 tachyzoites	20, 50, and 100 mg/kg, orally once/day for 4 days	The numbers of tachyzoites in the peritoneal cavity, Hematology and biochemical parameters	The inhibition rate = 44.7% hematology indicators and biochemical parameters reduced by nitrofurantoin significantly	Effective	Pyrimethamine	Yeo et al., 2016

(Continued)

TABLE 4 | Continued

No	Drug	Animal	Strain	Type of infection	Inoculum	Treatment	Assessment of efficacy	Main results	Effectivity	Positive control	References
55	Dextran sulfate	Pigs	RH	Acute	1 × 10 ⁶ tachyzoites, intravenously	50–500 µg per head	host clinical, pathological, and immunological analyses	High-dose caused reversible hepatocellular degeneration of the liver	Effective	.	Kato et al., 2016
56	Propranolol	BALB/c	RH	Acute, chronic	1 × 10 ³ tachyzoites i.p	2, 3 mg/kg/day	Parasite load determined by qPCR, and survival rate	Decreased the parasite load in brain, eye, and spleen tissues	Effective	Pyrimethamine	Montazeri et al., 2016
57	Resveratrol and sulfamethoxazole-trimetoprim	Male Swiss Webster	VEG	Chronic	50 cysts orally	Oral doses of 0.5 and 100 mg/kg/day	Counting brain cysts, tissue oxidant and antioxidant levels, and histopathology	A reduction on the number of cysts in the brain was observed	Co-administration more effective	Sulfamethoxazole-trimetoprim	Bottari et al., 2016
58	Compound 22 of sulfur-containing linear bisphosphonates	Webster mice	RH	Acute	20 or 100 or 5000 tachyzoites i.p	0.05, 0.1, 0.5, and 1 mg/kg of 22/ i.p. for 10 days	Survival rate	ED ₅₀ = 0.02 mg/kg	Effective	.	Szajman et al., 2016
59	Compound 32 (TgCDPK1 inhibitor)	Female CF-1 CBA/J	RH ME49	Acute, chronic	less than 100 tachyzoites/mL	20 mg/kg for 5 days/ oral gavage 30 mg/kg for 14 days	The numbers of tachyzoites in spleen, brain/ and the number of brain cysts	Reducing infection in spleen and brain (99%, 95%) 88.7% reduction of brain cyst	Effective	.	Vidadala et al., 2016

^a2-hydroxy-3-(1-propen-3-phenyl)-1,4-naphthoquinone.

^bIndirect immunofluorescence antibody test.

^cReverse transcription polymerase chain reaction.

^dPolymerase chain reaction.

^eQuantitative Polymerase chain reaction.

^fFluconazole.

^gItraconazole.

^h1-Hydroxy-2-Alkyl-4(1H) Quinolone.

ⁱ6-trifluoromethyl-2-thiouacil.

^j3-[(2-((E)-furan-2-ylmethylene) hydrazinyl) methylenyl] methylene-1, 3-dihydroindol-2-one.

^kLipid peroxidation.

^lGlutathione-S-transferase.

^mAlanine amino transferase.

ⁿAspartate amino transferase.

^oAlkaline phosphatase.

^pInterferon gamma.

^qTumor necrosis factor.

TABLE 5 | A comprehensive list of drugs/compounds evaluated against *T. gondii* with regard to IC₅₀.

Drug	IC ₅₀ (μM)			References
	<1	1–5	5–10	
Novel quinclidine	+			Martins-Duarte et al., 2006
Novel ferrocenic atovaquone derivatives	Atovaquone (PLK strain)	2d, 2e, 2f		Baramée et al., 2006
SAHA ^a , SBHA ^b , Scriptaid, Trichostatin A	Scriptaid Trichostatin A SAHA SBHA			Strobl et al., 2007
Pyridinylimidazoles SB203580 and SB202190	RWJ67657, (ME49 strain)	SB202190 SB203580 RWJ68198, (ME49 strain) RWJ67657, (RH strain)	SB203580 RWJ68198, (RH strain)	Wei et al., 2007
1-hydroxy-2-dodecyl-4(1H) quinolone	+			Saleh et al., 2007
Fluorine-containing aryloxyethyl thiocyanate derivatives		Compound 1, 3, 9	Compound 10	Liñares et al., 2007
Novel diamidine analog	+			Leepin et al., 2008
Pyrimethamine, sulfadiazine, atovaquone	+			Meneceur et al., 2008
Novel triazine JPC-2067-B	+			Mui et al., 2008
2-alkylaminoethyl-1,1-bisphosphonic acids		Compound 19	Compound 14, 17	Szajman et al., 2008
Itraconazole	+			Martins-Duarte Edos et al., 2008
Thiolactomycin analog		Compound 5, 6	Compound 2	Martins-Duarte et al., 2009
Fluconazole (FLZ)		FLZ (48 h)	FLZ (24 h)	Martins-Duarte et al., 2010
1-Hydroxy-2-Alkyl-4(1H) Quinolone derivatives	+			Bajohr et al., 2010
Haloperidol, clozapine, fluphenazine, trifluoperazine, thioridazine		+		Goodwin et al., 2011
Novel azasterols	Compound 3 (48 h)	Compound 1 (48 h), 2, 3 (24 h)	Compound 1 (24 h)	Martins-Duarte et al., 2011
Endochin-like quinolones	+			Doggett et al., 2012
Pterocarpanquinone		+		Portes Jde et al., 2012
New naphthoquinones (QUI), an alkaloid	QUI-11 Liriodenine			Ferreira et al., 2012
Di-cationic, pentamidine-analog	+			Kropf et al., 2012
Fuconazole combined with sulfadiazine and pyrimethamine	Pyrimethamine		+	Martins-Duarte et al., 2013

(Continued)

TABLE 5 | Continued

Drug	IC ₅₀ (μM)			References
	<1	1–5	5–10	
Antipsychotic drugs and valproate		Fluphenazine Thioridazine	Zuclopenthixol	Fond et al., 2014
Fusidic acid			+	Payne et al., 2013
Ivermectin and sulphadiazine	Ivermectin		Sulphadiazine	Bilgin et al., 2013
Novel ruthenium complexes,(compounds 16 and 18)	+			Barna et al., 2013
Auranofin	+			Andrade et al., 2014
6-Trifluoromethyl-2-thiouracil		+		Choi et al., 2014
200 drug-like, 200 probe-like compounds of Malaria Box	MMV007791	MMV007881 MMV007363 MMV006704 MMV666095 MMV020548 MMV085203		Boyom et al., 2014
Quinoline derivatives	8-Hydroxyquinoline, A 11, A14, A18, B11, B12, B15, B23, B24	A2-6, A12, A15–17, A23, B16, B22, B26, B27, B29, Chloroquine	Quinoline 2-chloroquinoline 5-Nitroqu Inoline Quinoline N-oxide hydrate A7, B18	Kadri et al., 2014
Bumped Kinase Inhibitor 1294	+			Doggett et al., 2014
Salicylanilides	3i, 3j, 7a, 14a, 14b			Fomovska et al., 2012
Antiretroviral compounds		Atazanavir Ritonavir Saquinavir	Fosamprenavir Nelfinavir	Monzote et al., 2013
Spiroindolone		+		Zhou et al., 2014
Ciprofloxacin derivatives	Compound 2, 5	Compound 4		Dubar et al., 2011
Thiazolidin-4-one derivatives	12A	27, 34 A	36 A	D'Ascenzio et al., 2014
N6-benzyladenosine analog			Compound 11 e, g, j, n, o, q, u, v	Kim et al., 2007
Naphthoquinone derivative	LQB151 (48 h)	LQB94 LQB151 (24 h) LQB150 (24, 48 h)		da Silva et al., 2015
Oryzalin analogs	Compound 6a, h, i, 14a, 18a, b, c	Compound 6b, g, j, l, n, 12	Compound 6m, 14b	Endeshaw et al., 2010
94 compounds		+		Dittmar et al., 2016
6-(1,2,6,7-tetraoxaspiro[7.11]nonadec-4-yl)hexan-1-ol (N-251)		+		Xin et al., 2016

^aSuberoylanilide hydroxamic acid.

^bSuberic bishydroxamic acid.

in mice and can be considered as an alternative for reducing the dose and side effects of pyrimethamine (Pissinate et al., 2014). Recently, researchers reported that computational analysis of biochemical differences between human and *T. gondii* dihydrofolate reductase enabled the design of inhibitors with both improved potency and selectivity against *T. gondii* (Welsch et al., 2016). El-Zawawy et al. reported that incorporating triclosan into the lipid bilayer of liposomes allowed its use in lower doses, which in turn, reduced its biochemical adverse effects (El-Zawawy et al., 2015b). In another study, sodium dodecyl sulfate (SDS)-coated atovaquone nanosuspensions (ANSs) considerably increased the therapeutic efficacy against experimentally reactivated and acquired toxoplasmosis by improving passage of gastrointestinal and blood-brain barriers. Accordingly, coating of ANSs with SDS may improve the treatment of toxoplasmic encephalitis and other cerebral diseases (Shubar et al., 2011).

Also, various studies showed that a number of drugs were investigated for the mechanisms of action summarized in **Table 2** and **Figure 2**. One study discussing the metabolic differences between the host and the parasite noted that dihydrofolate reductase, isoprenoid pathway, and *T. gondii* histone deacetylase are promising molecular targets (Rodriguez and Szajnman, 2012).

Novel triazine JPC-2067-B (4, 6-diamino-1, 2-dihydro-2, 2-dimethyl-1-(3'(2-chloro-, 4-trifluoromethoxyphenoxy)propyloxy)-1, 3, 5-triazine), the anti-folate medicines, is highly effective against *T. gondii* with an IC_{50} of $0.02 \mu\text{M}$, which is more efficacious than pyrimethamine and has *in vitro* cidal activity. Additionally, pro-drug JPC-2056 (1-(3'-(2-chloro-4-trifluoromethoxyphenoxy)propyl oxy)-5-isopropylbiguanide) is effective *in vivo* when administered orally (Mui et al., 2008). Moreover, histone deacetylase is potentially a very important drug target in *T. gondii*, since scriptaid and trichostatin A had the highest effect against *T. gondii* tachyzoite proliferation with the IC_{50} of 0.039 and $0.041 \mu\text{M}$, respectively (Strobl et al., 2007). For promising anti-*T. gondii* drugs/compounds, assessment of their ability to control parasite growth is a key step in drug development (McFarland et al., 2016).

A large number of research papers suggested that the apicoplast represents a potential drug target for new chemotherapy, as it is essential to the parasite and it is absent in host cells. Functions of the apicoplast include fatty acid synthesis, protein synthesis, DNA replication, electron transport, and heme biosynthesis (Yung and Lang-Unnasch, 2004). Some of the drugs evaluated against *T. gondii* are shown to act in the apicoplast such as thiolactomycin, triclosan (TS), azithromycin, fusidic acid, ciprofloxacin, and quinoline derivatives (Costa et al., 2009; Martins-Duarte et al., 2009, 2015; Payne et al., 2013; Kadri et al., 2014; El-Zawawy et al., 2015b).

In *T. gondii*, FAS-II enzymes are present in the apicoplast and are essential for its survival. The key enzyme in this process is the ENR enzyme, which is not found in mammals (Surolia and Surolia, 2001). This enzyme catalyzes the last reductive step of the type II FAS pathway. The TS, which inhibits type II FAS, significantly reduced mice mortality, parasite burden, as well as viability and infectivity of tachyzoites

and cysts harvested from infected treated mice and their brains. Accordingly, TS is proved as an effective, promising, and safe preventive drug against acute and chronic murine toxoplasmosis. Liposomal formulation of TS enhanced its efficacy and allowed its use at a lower dose (Surolia and Surolia, 2001; El-Zawawy et al., 2015a,b). Among apicoplast pathways, DNA replication is an important potential chemotherapeutic target. Fluoroquinolones are the known DNA replication inhibitors that target prokaryotic type II topoisomerases (Collin et al., 2011). In two studies, researchers showed that derivatives of the antibiotic ciprofloxacin, a fluoroquinolone, are active against *T. gondii* tachyzoites both *in vitro* and *in vivo* (Neville et al., 2015). While all mice treated with ciprofloxacin died by day 10 post-infection, some mice treated with ciprofloxacin derivatives remained alive for at least 60 days, suggesting that ciprofloxacin derivatives cured *T. gondii* infection in treated mice (Dubar et al., 2011; Martins-Duarte et al., 2015).

Anti-Toxoplasma Activities of New Synthetic Compounds

There are numerous reports on efficacy of many new synthetic compounds with a focus on identifying drug candidates with innovative and acceptable profiles against *T. gondii*. The anti-coccidial effect of 1-[4-(4-nitrophenoxy) phenyl] propane-1-one (NPPP), a synthetic compound, was studied both *in vitro* and *in vivo*. Treatment with NPPP showed anti-Toxoplasma activity *in vitro* with a lower EC_{50} value than pyrimethamine. In ICR mice infected with *T. gondii*, oral administration of NPPP for 4 days showed statistically significant anti-Toxoplasma activity with lower number of tachyzoites than those of the negative control (Choi et al., 2015).

In a study by Kadri et al. anti-Toxoplasma properties of 58 newly synthesized quinoline compounds were evaluated. A significant improvement in anti-Toxoplasma effect among quinoline derivatives was detected in B11, B12, B23, and B24. Among these compounds, B23 was the most effective compound with the IC_{50} value of $<1 \mu\text{M}$, displaying its anti-Toxoplasma effects and ability to cause the disappearance of the apicoplast (40–45% of the parasites lost their apicoplasts; Kadri et al., 2014).

In a study by Boyom et al. the strategy adopted was to repurpose the open access Malaria Box to identify chemical series active against *T. gondii*. The results showed that the most interesting compound was MMV007791, a piperazine acetamide, which has an IC_{50} of $0.19 \mu\text{M}$. This compound is novel for its anti-Toxoplasma activity, and of course, further studies on the rates and mechanisms of compound action will elucidate these considerations (Boyom et al., 2014).

Tetraoxanes, anti-cancer molecules, were tested *in vivo* against *T. gondii*. Subcutaneous, administration of a 10 mg/kg/day dose of derivative 21, for 8 days allowed the survival of 20% of infected mice, demonstrating the high potential of tetraoxanes for the treatment of *T. gondii* (Opsenica et al., 2015).

In another study by Moine et al. researchers evaluated *in vitro* anti-*T. gondii* activity of 51 compounds with a biphenylimidazoazine scaffold. Eight of these compounds displayed highly potent activity against *T. gondii* growth *in vitro*,

with 50% effective concentration (EC₅₀) below 1 mM, without demonstrating cytotoxic effects on human fibroblastic cell at equivalent concentrations. However, these compounds have to be evaluated in animal models so as to confirm their *in vivo* activity (Moine et al., 2015a).

Several pathways were characterized and shown to differ significantly from those of the mammalian host cells, thus, revealing an attractive area for therapeutic intervention. 1-Hydroxy-2-Alkyl-4 (1H) quinolone derivatives inhibit the fourth step of the essential *de novo* synthesis of pyrimidine, which uses ubiquinol reduction as an electron sink for dihydroorotate oxidation (Saleh et al., 2007). Also, newly synthesized bisphosphonates interfere with the mevanolate pathway, which leads to the synthesis of sterols and polyisoprenoid compounds that are important for parasite survival (Shubar et al., 2008).

Interestingly, Kamau et al. identified novel kinases that are integral to essential pathways, elucidating their mechanism of action and ultimately, identifying new drug targets (Kamau et al., 2012). In that study, 527 compounds were evaluated *in vitro*; also, they assessed the impact of the inhibitory compounds C1, C2, C3, and C5 in mouse models of toxoplasmosis. C2 was found quite effective in decreasing the parasite burden and increasing mice survival. These results should be considered with caution, since there are a number of factors are at play in whether a compound will be *in vivo* effective, such as solubility *in vivo*, access to different tissues, and host metabolic processes (Kamau et al., 2012). In a recent study, Dittmar et al. screened a collection of 1,120 compounds, 94 of which were blocked parasite replications with IC₅₀ of <5 μM. These data suggest that tamoxifen restricts *Toxoplasma* growth by inducing xenophagy or autophagic destruction of this parasite (Dittmar et al., 2016). According to a new study, *in silico* screening is useful, particularly in the identification of molecular targets in the laboratory. Fernandez et al. synthesized VAM2 compounds (7-nitroquinoxalin-2-ones), based on the design obtained from an *in silico* prediction with the software TOMOCOMD-CARDD. From the group of VAM2 compounds, Fernandez et al. chose VAM2-2 with an IC₅₀ of 3.3 μM against *T. gondii*. However, more studies are required to evaluate its effect on the cysts formed by of the parasite and in animal models of toxoplasmosis (Fernández et al., 2016).

Activity of Drugs, Compounds, and Combined Therapy against Cysts

An ideal drug against toxoplasmosis should not only be effective against the proliferative stage of the parasite but also exert dual activity against the tissue cyst stage and penetration into cysts (Benmerzouga et al., 2015). Currently, there is no approved therapy that eliminates the tissue cysts responsible for chronic infection (Innes, 2010). Derouin reported that among the drugs commonly used in humans, only atovaquone and azithromycin were found effective after long-term incubation. Besides, arpinocid-N-oxyde, an anticoccidial for veterinary use, was efficient at a high dosage (Derouin, 2005).

Recently, investigators have focused on guanabenz for *in vivo* studies, as guanabenz inhibitor of eIF2α dephosphorylation, is already an food and drug administration (FDA) approved drug and has excellent solubility with good penetration into the CNS. The results of that study show that guanabenz (5 mg/kg/day) not only protects mice against acute toxoplasmosis, but also reduces 69% of the number of brain cysts in chronically infected animals. This finding suggested that guanabenz can be repurposed into an effective antiparasitic with a unique ability to diminish tissue cysts in the brain (Benmerzouga et al., 2015).

Another study showed that miltefosine had no efficacy in controlling acute toxoplasmosis after 5 days of treatment; however, a 15-day treatment against the established chronic stage led to a 78% reduction of cysts in the brain. Additionally, the remaining cysts were noticeably smaller upon microscopic examination, suggesting that the drug effectively penetrates the blood-brain barrier, and that extension of treatment time may produce greater effects (Eissa et al., 2015).

In another study by Maubon et al. FR235222 and its derivatives were identified as new lead compounds for use against acute and chronic toxoplasmosis both *in vitro* and *in vivo*. *In vivo* experiments indicated that FR235222, as a histone deacetylase inhibitor, is able to access the bradyzoites within the cyst. The ability of FR235222 to permeate the membrane wall is a major advantage for crossing the blood-brain barrier and CNS tissue, where *Toxoplasma* cysts are located. This opens a promising way to develop drugs that are selective against *Toxoplasma* and those that have sterilizing activity, especially in patients with cysts, who are at risk for reactivating acute toxoplasmosis (patients with HIV infection, hematological malignancies, or transplantation). Still, effectiveness of FR235222 against chronically infected mice remains to be directly demonstrated *in vivo* (Maubon et al., 2010).

In a new study Vidadala et al. identified compounds 32 (*T. gondii* calcium-dependent protein kinase 1 inhibitor) a promising lead for the development of a new antitoxoplasmosis therapy. Compounds 32 is CNS-penetrant and highly effective in acute and latent mouse models of *T. gondii* infection, significantly reducing brain cysts by 88.7% (Vidadala et al., 2016).

Many studies reported anti-*Toxoplasma* effects of different drugs combination with novel compounds. The compound 2-hydroxy-3-(1'-propen-3-phenyl)-1, 4-naphthoquinone (PHNQ6), (50 mg/kg/day) combined with sulfadiazine showed reduction or elimination of brain cysts *in vivo* (Ferreira et al., 2006). In another study that coadministered spiramycin and metronidazole, spiramycin, did not reach effective concentrations in the brain due to the presence of the efflux transporters multidrug-resistant protein 2, and P-glycoprotein. Metronidazole increased brain penetration of spiramycin, causing a significant reduction of *T. gondii* brain cysts, with potential clinical translatability for chronic toxoplasmosis treatment. According to the reports, combination therapy leads to faster recovery, less relapse, lower doses of drugs, and fewer side effects of the disease. Furthermore, such combinations are highly promising to develop a drug that is able to eliminate the cyst stage of the parasite, and thus, efficiently impairs relapse of the disease (Chew et al., 2012; Martins-Duarte et al., 2013).

Activity of Drugs against Congenital Toxoplasmosis

In pregnant women, current toxoplasmosis treatment is based on the administration of spiramycin or a drug combination such as sulphadiazine-pyrimethamine-folinic acid (SPFA) in cases of confirmed fetal infection. However, these drugs are not well-tolerated and present many adverse effects due to their toxic effects to the host (Degerli et al., 2003).

Degerli et al. evaluated the effectiveness of azithromycin, artemisia annua infusion, spiramycin, and SPFA in *Calomys callosus*, such as model of congenital toxoplasmosis. The results demonstrated that the treatment of pregnant *C. callosus* with azithromycin was effective for inhibiting the vertical transmission of *T. gondii* ME49 strain, suggesting that it may be an alternative drug of choice for the treatment of congenital infection, since it is able to inhibit fetal infection and offers new perspectives for the treatment of congenital toxoplasmosis. Azithromycin is one of the new generation macrolides with numerous advantages. Mechanism of action of azithromycin is based on the inhibition of protein synthesis in both *T. gondii* tachyzoite and bradyzoite stages (Degerli et al., 2003), but it may present limited effectiveness against *T. gondii*, requiring high drug concentrations (Costa et al., 2009). In another study, Oz et al. reported that combined atovaquone and diclazuril therapy is a novel synergistic prophylactic and therapeutic approach to fetal maternal toxoplasmosis (Oz, 2014a). Atovaquone, an inhibitor of mitochondrial electron-transport processes, is an FDA-approved toxoplasmosis therapy but not for use in congenital toxoplasmosis treatment (Oz, 2014a). Another compound, diclazuril, and its related benzeneacetonitriles have long been used in the treatment and prevention of poultry and livestock coccidiosis. In addition, it is known to be a safe compound at therapeutic dose levels (Assis et al., 2010).

Adverse Effects of Drugs

However, anti-*Toxoplasma* effects of drugs/compounds were reported in many trials, but prednisolone increased the number of tachyzoites and bradyzoites in immunosuppressed infected mice (Puvanuesuaran et al., 2012). In addition, betamethasone can

escalate the invasion of tachyzoites, in cell culture. It could be suggested that patients under prolonged use of betamethasone and prednisolone should be protected against *T. gondii* infection. Also, if individuals receiving betamethasone are infected with *T. gondii*, interferon-gamma may be used to reduce the invasion of tachyzoites (Ghaffarifar et al., 2006).

CONCLUSIONS

As current chemotherapy against toxoplasmosis is still not satisfactory, the development of well-tolerated and safe specific immunoprophylaxis in relaxing the need of dependence on chemotherapeutics is a highly valuable goal for global disease control. Immunotherapeutics strategies for improving toxoplasmosis control could either be a vaccine which would induce strong protective immunity against toxoplasmosis, or passive immunization in cases of disease recrudescence. However, with the increasing number of high-risk individuals, such as immunocompromised patients, and absence of a proper vaccine, continued efforts are necessary for the development of novel treatment options against *T. gondii*. Some of the novel compounds reviewed here may represent good starting points for the discovery of effective new drugs. In further bioinformatic and *in silico* studies are needed in order to identify new potential toxoplasmodicidal drugs.

AUTHOR CONTRIBUTIONS

AD and MS conceived the idea for this review. MM and SS searched the databases for potentially eligible articles based on their titles and abstracts. AD and MM participated in the study design and wrote the manuscript. SM and EA critically reviewed the manuscript. All authors read and approved the final manuscript for publication.

ACKNOWLEDGMENTS

We also would like to thank of financial support by Vice Chancellors for Research of Mazandaran University of Medical Sciences, Sari, Iran (Grant number: 2085).

REFERENCES

- Al-jader, Z. H., and Al-Mukhtar, M. A. (2010). Synergistic activity of azithromycin combined with metronidazole against toxoplasmosis in experimental mice. *Iraq. J. Pharm.* 10, 1–4.
- Affi, M. A., and Al-Rabia, M. W. (2015). The immunomodulatory effects of rolipram abolish drug-resistant latent phase of *Toxoplasma gondii* infection in a murine model. *J. Microsc. Ultrastruct.* 3, 86–91. doi: 10.1016/j.jmau.2014.12.001
- Affi, M. A., Jiman-Fatani, A. A., Al-Rabia, M. W., and Al-Hussainy, N. H. (2014). Application of a phosphodiesterase-4 (PDE4) inhibitor to abort chronic toxoplasmosis and to mitigate consequential pathological changes. *J. Microsc. Ultrastruct.* 2, 94–99. doi: 10.1016/j.jmau.2014.05.002
- Ahmadpour, E., Daryani, A., Sharif, M., Sarvi, S., Aarabi, M., Mizani, A., et al. (2014). Toxoplasmosis in immunocompromised patients in Iran: a systematic review and meta-analysis. *J. Infect. Dev. Ctries.* 8, 1503–1510. doi: 10.3855/jidc.4796
- Alomar, M. L., Rasse-Suriani, F. A., Ganuza, A., Cóceres, V. M., Cabrerizo, F. M., and Angel, S. O. (2013). *In vitro* evaluation of β -carboline alkaloids as potential anti-*Toxoplasma* agents. *BMC Res. Notes* 6:193. doi: 10.1186/1756-0500-6-193
- Andrade, R. M., Chaparro, J. D., Capparelli, E., and Reed, S. L. (2014). Auranofin is highly efficacious against *Toxoplasma gondii* *in vitro* and in an *in vivo* experimental model of acute toxoplasmosis. *PLoS Negl. Trop. Dis.* 8:e2973. doi: 10.1371/journal.pntd.0002973
- Angel, S. O., Matrajt, M., and Echeverria, P. C. (2013). A review of recent patents on the protozoan parasite HSP90 as a drug target. *Recent Pat. Biotechnol.* 7, 2–8. doi: 10.2174/1872208311307010002
- Aquino, T. M., Nascimento, A. A., Spacov, I. C., Carvalho, C. S., Lima, V. T., Alves, A. Q., et al. (2011). Synthesis, anti-*Toxoplasma gondii* and antimicrobial activities of 2-hydrazolyl-3-phenyl-5-(4-nitrobenzylidene)-4-thiazolidinone substituted derivatives. *Lat. Am. J. Pharm.* 30, 1567–1573. doi: 10.1016/j.bmc.2007.09.025

- Araujo, F. G., and Remington, J. S. (1992). Recent advances in the search for new drugs for treatment of toxoplasmosis. *Int. J. Antimicrob. Agents* 1, 153–164. doi: 10.1016/0924-8579(92)90002-9
- Asgari, Q., Keshavarz, H., Rezaeian, M., Motazedian, M. H., Shojaee, S., Mohebbi, M., et al. (2013). Direct effect of two naphthalene-sulfonyl-indole compounds on *Toxoplasma gondii* tachyzoite. *J. Parasitol. Res.* 2013:716976. doi: 10.1155/2013/716976
- Asgari, Q., Keshavarz, H., Rezaeian, M., Sadeghpour, H., Miri, R., and Motazedian, M. H. (2015). Anti-toxoplasma activity of 2-(naphthalene-2-ylthiol)-1H indole. *Iran. J. Parasitol.* 10, 171–180. Available online at: <http://ijpa.tums.ac.ir>
- Assis, R. C., Luns, F. D., Beletti, M. E., Assis, R. L., Nasser, N. M., Faria, E. S., et al. (2010). Histomorphometry and macroscopic intestinal lesions in broilers infected with *Eimeria acervulina*. *Vet. Parasitol.* 168, 185–189. doi: 10.1016/j.vetpar.2009.11.017
- Bajohr, L. L., Ma, L., Platte, C., Liesenfeld, O., Tietze, L. F., Groß, U., et al. (2010). *In vitro* and *in vivo* activities of 1-hydroxy-2-alkyl-4 (1H) quinolone derivatives against *Toxoplasma gondii*. *Antimicrob. Agents Chemother.* 54, 517–521. doi: 10.1128/AAC.01001-09
- Baramée, A., Coppin, A., Mortuaire, M., Pelinski, L., Tomavo, S., and Brocard, J. (2006). Synthesis and *in vitro* activities of ferrocenic aminohydroxynaphthoquinones against *Toxoplasma gondii* and *Plasmodium falciparum*. *Bioorg. Med. Chem.* 14, 1294–1302. doi: 10.1016/j.bmc.2005.09.054
- Barbosa, B. F., Gomes, A. O., Ferro, E. A., Napolitano, D. R., Mineo, J. R., and Silva, N. M. (2012). Enrofloxacin is able to control *Toxoplasma gondii* infection in both *in vitro* and *in vivo* experimental models. *Vet. Parasitol.* 187, 44–52. doi: 10.1016/j.vetpar.2011.12.039
- Barna, F., Debache, K., Vock, C. A., Küster, T., and Hemphill, A. (2013). *In vitro* effects of novel ruthenium complexes in *Neospora caninum* and *Toxoplasma gondii* tachyzoites. *Antimicrob. Agents Chemother.* 57, 5747–5754. doi: 10.1128/AAC.02446-12
- Benmerzouga, I., Checkley, L. A., Ferdig, M. T., Arrizabalaga, G., Wek, R. C., and Sullivan, W. J. (2015). Guanabenz repurposed as an antiparasitic with activity against acute and latent toxoplasmosis. *Antimicrob. Agents Chemother.* 59, 6939–6945. doi: 10.1128/AAC.01683-15
- Bessières, M., Berrebi, A., Cassaing, S., Fillaux, J., Cambus, J., Berry, A., et al. (2009). Diagnosis of congenital toxoplasmosis: prenatal and neonatal evaluation of methods used in Toulouse University Hospital and incidence of congenital toxoplasmosis. *Mem. Inst. Oswaldo Cruz* 104, 389–392. doi: 10.1590/S0074-02762009000200038
- Bilgin, M., Yıldırım, T., and Hökelek, M. (2013). *In vitro* effects of ivermectin and sulphadiazine on *Toxoplasma gondii*. *Balkan Med. J.* 30, 19. doi: 10.5152/balkanmedj.2012.098
- Bosch-Driessen, L. H., Verbraak, F. D., Suttorp-Schulten, M. S., van Ruyven, R. L., Klok, A. M., Hoyng, C. B., et al. (2002). A prospective, randomized trial of pyrimethamine and azithromycin vs pyrimethamine and sulfadiazine for the treatment of ocular toxoplasmosis. *Am. J. Ophthalmol.* 134, 34–40. doi: 10.1016/S0002-9394(02)01537-4
- Bottari, N. B., Baldissera, M. D., Tonin, A. A., Rech, V. C., Alves, C. B., D'Avila, F., et al. (2016). Synergistic effects of resveratrol (free and inclusion complex) and sulfamethoxazole-trimethoprim treatment on pathology, oxidant/antioxidant status and behavior of mice infected with *Toxoplasma gondii*. *Microb. Pathog.* 95, 166–174. doi: 10.1016/j.micpath.2016.04.002
- Bottari, N. B., Baldissera, M. D., Tonin, A. A., Rech, V. C., Nishihira, V. S., Thomé, G. R., et al. (2015). Sulfamethoxazole-trimethoprim associated with resveratrol for the treatment of toxoplasmosis in mice: influence on the activity of enzymes involved in brain neurotransmission. *Microb. Pathog.* 79, 17–23. doi: 10.1016/j.micpath.2015.01.001
- Boyom, F. F., Fokou, P. V., Tchokouaha, L. R., Spangenberg, T., Mfopa, A. N., Kouipou, R. M., et al. (2014). Repurposing the open access malaria box to discover potent inhibitors of *Toxoplasma gondii* and *Entamoeba histolytica*. *Antimicrob. Agents Chemother.* 58, 5848–5854. doi: 10.1128/AAC.02541-14
- Castanheira, L., Naves de Souza, D. L., Silva, R. J., Barbosa, B., Mineo, J. R., Tudini, K. A., et al. (2015). Insights into anti-parasitism induced by a C-type lectin from *Bothrops pauloensis* venom on *Toxoplasma gondii*. *Int. J. Biol. Macromol.* 74, 568–574. doi: 10.1016/j.ijbiomac.2014.11.035
- Castro-Filice, L. S., Barbosa, B. F., Angeloni, M. B., Silva, N. M., Gomes, A. O., Alves, C. M., et al. (2014). Azithromycin is able to control *Toxoplasma gondii* infection in human villous explants. *J. Transl. Med.* 12:132. doi: 10.1186/1479-5876-12-132
- Chao, M. N., Li, C., Storey, M., Falcone, B. N., Szajnman, S. H., Bonesi, S. M., et al. (2016). Activity of fluorine-containing analogues of WC-9 and structurally related analogues against two intracellular parasites: *Trypanosoma cruzi* and *Toxoplasma gondii*. *ChemMedChem* 11, 2690–2702. doi: 10.1002/cmdc.201600505
- Chao, M. N., Matiuzzi, C. E., Storey, M., Li, C., Szajnman, S. H., Docampo, R., et al. (2015). Aryloxyethyl thiocyanates are potent growth inhibitors of *Trypanosoma cruzi* and *Toxoplasma gondii*. *ChemMedChem* 10, 1094–1108. doi: 10.1002/cmdc.201500100
- Chew, W. K., Segarra, I., Ambu, S., and Mak, J. W. (2012). Significant reduction of brain cysts caused by *Toxoplasma gondii* after treatment with spiramycin coadministered with metronidazole in a mouse model of chronic toxoplasmosis. *Antimicrob. Agents Chemother.* 56, 1762–1768. doi: 10.1128/AAC.05183-11
- Choi, H. J., Lee, J. H., Yeo, S. J., Kaewintajuk, K., Yi, K. Y., Kim, S., et al. (2015). Evaluation of anti-coccidial effects of 1-[4-(4-nitrophenoxy)phenyl] propane-1-one and identification of its potential target proteins in *Toxoplasma gondii*. *Arch. Pharm. Res.* 38, 752–760. doi: 10.1007/s12272-014-0400-y
- Choi, H. J., Yu, S. T., Lee, K. I., Choi, J. K., Chang, B. Y., Kim, S. Y., et al. (2014). 6-trifluoromethyl-2-thiouracil possesses anti-*Toxoplasma gondii* effect *in vitro* and *in vivo* with low hepatotoxicity. *Exp. Parasitol.* 143, 24–29. doi: 10.1016/j.exppara.2014.05.002
- Collin, F., Karkare, S., and Maxwell, A. (2011). Exploiting bacterial DNA gyrase as a drug target: current state and perspectives. *Appl. Microbiol. Biotechnol.* 92, 479–497. doi: 10.1007/s00253-011-3557-z
- Costa, I. N., Angeloni, M. B., Santana, L. A., Barbosa, B. F., Silva, M. C., Rodrigues, A. A., et al. (2009). Azithromycin inhibits vertical transmission of *Toxoplasma gondii* in *Calomys callosus* (Rodentia: Cricetidae). *Placenta* 30, 884–890. doi: 10.1016/j.placenta.2009.08.002
- da Cunha, E. F., Ramalho, T. C., Mancini, D. T., Fonseca, E., and Oliveira, A. A. (2010). New approaches to the development of anti-protozoan drug candidates: a review of patents. *J. Braz. Chem. Soc.* 21, 1787–1806. doi: 10.1590/S0103-50532010001000002
- da Silva, L. L. R., de Araujo Portes, J., de Araújo, M. H., Silva, J. L., Rennó, M. N., Netto, C. D., et al. (2015). Further evidence that naphthoquinone inhibits *Toxoplasma gondii* growth *in vitro*. *Parasitol. Int.* 64, 622–631. doi: 10.1016/j.parint.2015.08.010
- D'Ascenzio, M., Bizzarri, B., De Monte, C., Carradori, S., Bolasco, A., Secci, D., et al. (2014). Design, synthesis and biological characterization of thiazolidin-4-one derivatives as promising inhibitors of *Toxoplasma gondii*. *Eur. J. Med. Chem.* 86, 17–30. doi: 10.1016/j.ejmech.2014.08.046
- de Lima, L. P., Seabra, S. H., Carneiro, H., and Barbosa, H. S. (2015). Effect of 3-bromopyruvate and atovaquone on infection during *in vitro* interaction of *Toxoplasma gondii* and LLC-MK2 cells. *Antimicrob. Agents Chemother.* 59, 5239–5249. doi: 10.1128/AAC.00337-15
- Degerli, K., Kilimcioglu, A. A., Kurt, Ö., Tamay, A. T., and Özbilgin, A. (2003). Efficacy of azithromycin in a murine toxoplasmosis model, employing a *Toxoplasma gondii* strain from Turkey. *Acta Trop.* 88, 45–50. doi: 10.1016/S0001-706X(03)00194-3
- Derouin, F. (2005). *Systematic Search and Analysis of Preclinical Published Data on In vitro and In vivo Activities of Antitoxoplasma Drugs (Panel 2: Treatment Issues)*. Bordeaux: The Eurotox Group.
- Derouin, F., Jacqz-Aigrain, E., Thulliez, P., Couvreur, J., and Lepout, C. (2000). Cotrimoxazole for prenatal treatment of congenital toxoplasmosis? *Parasitol. Today* 16, 254–256. doi: 10.1016/S0169-4758(00)01667-7
- Dittmar, A. J., Drozda, A. A., and Blader, I. J. (2016). Drug repurposing screening identifies novel compounds that effectively inhibit *Toxoplasma gondii* growth. *mSphere* 1, e00042-15. doi: 10.1128/mSphere.00042-15
- Doggett, J. S., Nilsen, A., Forquer, I., Wegmann, K. W., Jones-Brando, L., Yolken, R. H., et al. (2012). Endochin-like quinolones are highly efficacious against acute and latent experimental toxoplasmosis. *Proc. Natl. Acad. Sci. U.S.A.* 109, 15936–15941. doi: 10.1073/pnas.1208069109

- Doggett, J. S., Ojo, K. K., Fan, E., Maly, D. J., and Van Voorhis, W. C. (2014). Bumped kinase inhibitor 1294 treats established *Toxoplasma gondii* infection. *Antimicrob. Agents Chemother.* 58, 3547–3549. doi: 10.1128/AAC.01823-13
- Dubar, F., Wintjens, R., Martins-Duarte, É. S., Vommaro, R. C., de Souza, W., Dive, D., et al. (2011). Ester prodrugs of ciprofloxacin as DNA-gyrase inhibitors: synthesis, antiparasitic evaluation and docking studies. *Medchemcomm* 2, 430–435. doi: 10.1039/c1md00022e
- Dubey, J. P., and Jones, J. L. (2008). *Toxoplasma gondii* infection in humans and animals in the United States. *Int. J. Parasitol.* 38, 1257–1278. doi: 10.1016/j.ijpara.2008.03.007
- Dzitko, K., Paneth, A., Plech, T., Pawelczyk, J., Śtaczek, P., Stefańska, J., et al. (2014a). 1, 4-disubstituted thiosemicarbazide derivatives are potent inhibitors of *Toxoplasma gondii* proliferation. *Molecules* 19, 9926–9943. doi: 10.3390/molecules19079926
- Dzitko, K., Paneth, A., Plech, T., Pawelczyk, J., Węglińska, L., and Paneth, P. (2014b). Triazole-based compound as a candidate to develop novel medicines to treat toxoplasmosis. *Antimicrob. Agents Chemother.* 58, 7583–7585. doi: 10.1128/AAC.03832-14
- Eissa, M. M., Barakat, A. M., Amer, E. I., and Younis, L. K. (2015). Could miltefosine be used as a therapy for toxoplasmosis? *Exp. Parasitol.* 157, 12–22. doi: 10.1016/j.exppara.2015.06.005
- El-Zawawy, L. A., El-Said, D., Mossallam, S. F., Ramadan, H. S., and Younis, S. S. (2015a). Preventive prospective of triclosan and triclosan-liposomal nanoparticles against experimental infection with a cystogenic ME49 strain of *Toxoplasma gondii*. *Acta Trop.* 141, 103–111. doi: 10.1016/j.actatropica.2014.09.020
- El-Zawawy, L. A., El-Said, D., Mossallam, S. F., Ramadan, H. S., and Younis, S. S. (2015b). Triclosan and triclosan-loaded liposomal nanoparticles in the treatment of acute experimental toxoplasmosis. *Exp. Parasitol.* 149, 54–64. doi: 10.1016/j.exppara.2014.12.007
- Endeshaw, M. M., Li, C., de Leon, J., Yao, N., Latibeaudiere, K., Premalatha, K., et al. (2010). Synthesis and evaluation of oryzalin analogs against *Toxoplasma gondii*. *Bioorg. Med. Chem. Lett.* 20, 5179–5183. doi: 10.1016/j.bmcl.2010.07.003
- Ferreira, R. A., de Oliveira, A. B., Gualberto, S. A., Miguel Del Corral, J. M., Fujiwara, R. T., Gazzinelli Guimarães, P. H., et al. (2012). New naphthoquinones and an alkaloid with *in vitro* activity against *Toxoplasma gondii* RH and EGS strains. *Exp. Parasitol.* 132, 450–457. doi: 10.1016/j.exppara.2012.09.003
- Ferreira, R. A., Oliveira, A. B., Ribeiro, M. F., Tafuri, W. L., and Vitor, R. W. (2006). *Toxoplasma gondii*: *in vitro* and *in vivo* activities of the hydroxynaphthoquinone 2-hydroxy-3-(1'-propen-3-phenyl)-1, 4-naphthoquinone alone or combined with sulfadiazine. *Exp. Parasitol.* 113, 125–129. doi: 10.1016/j.exppara.2005.12.006
- Flegel, J., Prandota, J., Soviěková, M., and Israili, Z. H. (2014). Toxoplasmosis—a global threat. Correlation of latent toxoplasmosis with specific disease burden in a set of 88 countries. *PLoS ONE* 9:e90203. doi: 10.1371/journal.pone.0090203
- Fomovska, A., Wood, R. D., Mui, E., Dubey, J. P., Ferreira, L. R., Hickman, M. R., et al. (2012). Salicylanilide inhibitors of *Toxoplasma gondii*. *J. Med. Chem.* 55, 8375–8391. doi: 10.1021/jm3007596
- Fond, G., MacGregor, A., Tamouza, R., Hamdani, N., Meary, A., Leboyer, M., et al. (2014). Comparative analysis of anti-toxoplasma activity of antipsychotic drugs and valproate. *Eur. Arch. Psychiatry Clin. Neurosci.* 264, 179–183. doi: 10.1007/s00406-013-0413-4
- Franco, P. S., Gomes, A. O., Barbosa, B. F., Angeloni, M. B., Silva, N. M., Teixeira-Carvalho, A., et al. (2011). Azithromycin and spiramycin induce anti-inflammatory response in human trophoblastic (BeWo) cells infected by *Toxoplasma gondii* but are able to control infection. *Placenta* 32, 838–844. doi: 10.1016/j.placenta.2011.08.012
- Gaafar, M. R., Mady, R. F., Diab, R. G., and Shalaby, T. I. (2014). Chitosan and silver nanoparticles: promising anti-toxoplasma agents. *Exp. Parasitol.* 143, 30–38. doi: 10.1016/j.exppara.2014.05.005
- Galván-Ramírez Mde, L., Jiménez, J. M. D., Pérez, L. R. R., Troyo-Sanroman, R., Ramírez-Herrera, M., and García-Iglesias, T. (2013). Effect of nitaxozanide and pyrimethamine on astrocytes infected by *Toxoplasma gondii* *in vitro*. *Arch. Med. Res.* 44, 415–421. doi: 10.1016/j.arcmed.2013.07.002
- Ghaffarifar, F., Asl, A. D., Sharifi, Z., Ghasemi, S., Solhjoo, K., and Mohammadi, S. R. (2006). The effect of betamethasone and IFN- γ on replication of *Toxoplasma gondii* (RH Strain) and nitric oxide production in HeLa cell culture. *Iran. J. Allergy Asthma Immunol.* 5, 75–78.
- Gomes, M. A. G., Carvalho, L. P., Rocha, B. S., Oliveira, R. R., de Melo, E. J., and Maria, E. J. (2013). Evaluating anti-*Toxoplasma gondii* activity of new serie of phenylsemicarbazone and phenylthiosemicarbazones *in vitro*. *Med. Chem. Res.* 22, 3574–3580. doi: 10.1007/s00044-012-0347-9
- Goodwin, D. G., Strobl, J. S., and Lindsay, D. S. (2011). Evaluation of five antischizophrenic agents against *Toxoplasma gondii* in human cell cultures. *J. Parasitol.* 97, 148–151. doi: 10.1645/G.E.2536.1
- Habib, F. (2008). Post-treatment assessment of acute *Toxoplasma* infection during pregnancy. *J. Obstet. Gynaecol. (Lahore)*. 28, 593–595. doi: 10.1080/01443610802344332
- Hill, D., and Dubey, J. (2002). *Toxoplasma gondii*: transmission, diagnosis and prevention. *Clin. Microbiol. Infect.* 8, 634–640. doi: 10.1046/j.1469-0691.2002.00485.x
- Hoffmann, S., Batz, M. B., and Morris, J. G. Jr. (2012). Annual cost of illness and quality-adjusted life year losses in the United States due to 14 foodborne pathogens. *J. Food Protect.* 75, 1292–1302. doi: 10.4315/0362-028X.JFP-11-417
- Howe, D. K., and Sibley, L. D. (1995). *Toxoplasma gondii* comprises three clonal lineages: correlation of parasite genotype with human disease. *J. Infect. Dis.* 172, 1561–1566. doi: 10.1093/infdis/172.6.1561
- Ihara, F., and Nishikawa, Y. (2014). Synthetic retinoid Am80 controls growth of intracellular *Toxoplasma* by inhibiting acquisition and synthesis of cholesterol in macrophages. *J. Protozool. Res.* 24, 1–10.
- Innes, E. A. (2010). Vaccination against *Toxoplasma gondii*: an increasing priority for collaborative research? *Expert Rev. Vaccines* 9, 1117–1119. doi: 10.1586/erv.10.113
- Jain, V., Yogavel, M., Oshima, Y., Kikuchi, H., Touquet, B., Hakimi, M. A., et al. (2015). Structure of prolyl-tRNA synthetase-halfofuginone complex provides basis for development of drugs against malaria and toxoplasmosis. *Structure* 23, 819–829. doi: 10.1016/j.str.2015.02.011
- Jones, J. L., Parise, M. E., and Fiore, A. E. (2014). Neglected parasitic infections in the United States: toxoplasmosis. *Am. J. Trop. Med. Hyg.* 90, 794–799. doi: 10.4269/ajtmh.13-0722
- Julliac, B., Théophile, H., Begorre, M., Richez, B., and Haramburu, F. (2010). Side effects of spiramycin masquerading as local anesthetic toxicity during labor epidural analgesia. *Int. J. Obstet. Anesth.* 19, 331–332. doi: 10.1016/j.ijoa.2010.03.002
- Kadri, D., Crater, A. K., Lee, H., Solomon, V. R., and Ananvoranich, S. (2014). The potential of quinoline derivatives for the treatment of *Toxoplasma gondii* infection. *Exp. Parasitol.* 145, 135–144. doi: 10.1016/j.exppara.2014.08.008
- Kamau, E. T., Srinivasan, A. R., Brown, M. J., Fair, M. G., Caraher, E. J., and Boyle, J. P. (2012). A focused small-molecule screen identifies 14 compounds with distinct effects on *Toxoplasma gondii*. *Antimicrob. Agents Chemother.* 56, 5581–5590. doi: 10.1128/AAC.00868-12
- Kato, K., Murata, Y., Horiuchi, N., Inomata, A., Terkawi, M. A., Ishiwa, A., et al. (2016). Dextran sulfate inhibits acute *Toxoplasma gondii* infection in pigs. *Parasit. Vectors* 9, 134. doi: 10.1186/s13071-016-1421-9
- Kim, K., Eaton, M. S., Schubert, W., Wu, S., and Tang, J. (2001). Optimized expression of green fluorescent protein in *Toxoplasma gondii* using thermostable green fluorescent protein mutants. *Mol. Biochem. Parasitol.* 113, 309–313. doi: 10.1016/S0166-6851(01)00212-2
- Kim, Y. A., Sharon, A., Chu, C. K., Rais, R. H., Al Safarjalani, O. N., Naguib, F. N., et al. (2007). Synthesis, biological evaluation and molecular modeling studies of N 6-benzyladenosine analogues as potential anti-toxoplasma agents. *Biochem. Pharmacol.* 73, 1558–1572. doi: 10.1016/j.bcp.2007.01.026
- Köksal, Z. Ş., Yanik, K., Bilgin, K., Yılmaz, E. M., and Hokelek, M. (2016). *In vivo* efficacy of drugs acting on *Toxoplasma gondii* combined with immunomodulators. 69, 113–117. *Jpn. J. Infect. Dis.* doi: 10.7883/yoken.JJID.2015.023
- Kropf, C., Debache, K., Rampa, C., Barna, F., Schorer, M., Stephens, C. E., et al. (2012). The adaptive potential of a survival artist: characterization of the *in vitro* interactions of *Toxoplasma gondii* tachyzoites with di-cationic compounds in human fibroblast cell cultures. *Parasitology* 139, 208–220. doi: 10.1017/S0031182011001776

- Kul, O., Yildiz, K., Ocal, N., Freyre, A., Deniz, A., Karahan, S., et al. (2013). *In-vivo* efficacy of toltrazuril on experimentally induced *Toxoplasma gondii* tissue cysts in lambs: a novel strategy for prevention of human exposure to meat-borne toxoplasmosis. *Res. Vet. Sci.* 94, 269–276. doi: 10.1016/j.rvsc.2012.08.001
- Lai, B. S., Witola, W. H., El Bissati, K., Zhou, Y., Mui, E., Fomovska, A., et al. (2012). Molecular target validation, antimicrobial delivery, and potential treatment of *Toxoplasma gondii* infections. *Proc. Natl. Acad. Sci. U.S.A.* 109, 14182–14187. doi: 10.1073/pnas.1208775109
- Leepin, A., Stüdl, A., Brun, R., Stephens, C. E., Boykin, D. W., and Hemphill, A. (2008). Host cells participate in the *in vitro* effects of novel diamidine analogues against tachyzoites of the intracellular apicomplexan parasites *Neospora caninum* and *Toxoplasma gondii*. *Antimicrob. Agents Chemother.* 52, 1999–2008. doi: 10.1128/AAC.01236-07
- Leyke, S., Köhler-Sokolowska, W., Paulke, B. R., and Presber, W. (2012). Effects of nanoparticles in cells infected by *Toxoplasma gondii*. *e-Polymers* 12, 647–663. doi: 10.1515/epoly.2012.12.1.647
- Li, Z. H., Ramakrishnan, S., Striepen, B., and Moreno, S. N. (2013). *Toxoplasma gondii* relies on both host and parasite isoprenoids and can be rendered sensitive to atorvastatin. *PLoS Pathog.* 9:e1003665. doi: 10.1371/journal.ppat.1003665
- Liñares, G. G., Gismondi, S., Codesido, N. O., Moreno, S. N., Docampo, R., and Rodriguez, J. B. (2007). Fluorine-containing aryloxyethyl thiocyanate derivatives are potent inhibitors of *Trypanosoma cruzi* and *Toxoplasma gondii* proliferation. *Bioorg. Med. Chem. Lett.* 17, 5068–5071. doi: 10.1016/j.bmcl.2007.07.012
- Liesen, A. P., de Aquino, T. M., Carvalho, C. S., Lima, V. T., de Araújo, J. M., de Lima, J. G., et al. (2010). Synthesis and evaluation of anti-*Toxoplasma gondii* and antimicrobial activities of thiosemicarbazides, 4-thiazolidinones and 1, 3, 4-thiadiazoles. *Eur. J. Med. Chem.* 45, 3685–3691. doi: 10.1016/j.ejmech.2010.05.017
- Lim, S. S., and Othman, R. Y. (2014). Recent advances in *Toxoplasma gondii* immunotherapeutics. *Korean J. Parasitol.* 52, 581–593. doi: 10.3347/kjp.2014.52.6.581
- Luk, F. C., Johnson, T. M., and Beckers, C. J. (2008). N-linked glycosylation of proteins in the protozoan parasite *Toxoplasma gondii*. *Mol. Biochem. Parasitol.* 157, 169–178. doi: 10.1016/j.molbiopara.2007.10.012
- Martins-Duarte Edos, S., De Souza, W., and Vommaro, R. C. (2008). Itraconazole affects *Toxoplasma gondii* endodyogeny. *FEMS Microbiol. Lett.* 282, 290–298. doi: 10.1111/j.1574-6968.2008.01130.x
- Martins-Duarte, É. S., de Souza, W., and Vommaro, R. C. (2013). *Toxoplasma gondii*: the effect of fluconazole combined with sulfadiazine and pyrimethamine against acute toxoplasmosis in murine model. *Exp. Parasitol.* 133, 294–299. doi: 10.1016/j.exppara.2012.12.011
- Martins-Duarte, E. S., Dubar, F., Lawton, P., da Silva, C. F., Soeiro Mde, N., de Souza, W., et al. (2015). Ciprofloxacin derivatives affect parasite cell division and increase the survival of mice infected with *Toxoplasma gondii*. *PLoS ONE* 10:e0125705. doi: 10.1371/journal.pone.0125705
- Martins-Duarte, E. S., Jones, S. M., Gilbert, I. H., Atella, G. C., de Souza, W., and Vommaro, R. C. (2009). Thiolactomycin analogues as potential anti-*Toxoplasma gondii* agents. *Parasitol. Int.* 58, 411–415. doi: 10.1016/j.parint.2009.08.004
- Martins-Duarte, É. S., Lemgruber, L., de Souza, W., and Vommaro, R. C. (2010). *Toxoplasma gondii*: fluconazole and itraconazole activity against toxoplasmosis in a murine model. *Exp. Parasitol.* 124, 466–469. doi: 10.1016/j.exppara.2009.12.011
- Martins-Duarte, É. S., Lemgruber, L., Lorente, S. O., Gros, L., Magaraci, F., Gilbert, I. H., et al. (2011). Evaluation of three novel azasterols against *Toxoplasma gondii*. *Vet. Parasitol.* 177, 157–161. doi: 10.1016/j.vetpar.2010.11.034
- Martins-Duarte, É. S., Urbina, J. A., de Souza, W., and Vommaro, R. C. (2006). Antiproliferative activities of two novel quinuclidine inhibitors against *Toxoplasma gondii* tachyzoites *in vitro*. *J. Antimicrob. Chemother.* 58, 59–65. doi: 10.1093/jac/dkl180
- Maubon, D., Bougdour, A., Wong, Y. S., Brenier-Pinchart, M. P., Curt, A., Hakimi, M. A., et al. (2010). Activity of the histone deacetylase inhibitor FR235222 on *Toxoplasma gondii*: inhibition of stage conversion of the parasite cyst form and study of new derivative compounds. *Antimicrob. Agents Chemother.* 54, 4843–4850. doi: 10.1128/AAC.00462-10
- McFarland, M. M., Zach, S. J., Wang, X., Potluri, L.-P., Neville, A. J., Vennerstrom, J. L., et al. (2016). Review of experimental compounds demonstrating anti-toxoplasma activity. *Antimicrob. Agents Chemother.* 60, 7017–7034. doi: 10.1128/AAC.01176-16
- Meneceur, P., Bouldouyre, M. A., Aubert, D., Villena, I., Menotti, J., Sauvage, V., et al. (2008). *In vitro* susceptibility of various genotypic strains of *Toxoplasma gondii* to pyrimethamine, sulfadiazine, and atovaquone. *Antimicrob. Agents Chemother.* 52, 1269–1277. doi: 10.1128/AAC.01203-07
- Meriem, B., Hajira, B., Radia, B., Malika, B., and Med Reda, D. (2015). *In vitro* and *in vivo* potential effect of the N-acetylsulfonamide Bis-oxazolodi-2-ones on *Toxoplasma gondii*. *Biomed. Biol. Eng.* 2. Available online at: scholar.waset.org/1999.16/24904
- Moher, D., Liberati, A., Tetzlaff, J., Altman, D.G., and The PRISMA Group (2009). Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *BMJ* 339:b2535. doi: 10.1136/bmj.b2535
- Moine, E., Denevault-Sabourin, C., Debierre-Grockiego, F., Silpa, L., Gorgette, O., Barale, J. C., et al. (2015a). A small-molecule cell-based screen led to the identification of biphenylimidazoazines with highly potent and broad-spectrum anti-apicomplexan activity. *Eur. J. Med. Chem.* 89, 386–400. doi: 10.1016/j.ejmech.2014.10.057
- Moine, E., Dimier-Poisson, I., Enguehard-Gueffier, C., Logé, C., Pénichon, M., Moiré, N., et al. (2015b). Development of new highly potent imidazo [1, 2-b] pyridazines targeting *Toxoplasma gondii* calcium-dependent protein kinase 1. *Eur. J. Med. Chem.* 105, 80–105. doi: 10.1016/j.ejmech.2015.10.004
- Montazeri, M., Daryani, A., Ebrahimzadeh, M., Ahmadpour, E., Sharif, M., and Sarvi, S. (2015). Effect of propranolol alone and in combination with pyrimethamine on acute murine toxoplasmosis. *Jundishapur J. Microbiol.* 8:e22572. doi: 10.5812/jjm.22572
- Montazeri, M., Ebrahimzadeh, M. A., Ahmadpour, E., Sharif, M., Sarvi, S., and Daryani, A. (2016). Evaluation of propranolol effect on experimental acute and chronic toxoplasmosis using quantitative PCR. *Antimicrob. Agents Chemother.* 60, 7128–7133. doi: 10.1128/AAC.01323-16
- Montoya, J. G., and Liesenfeld, O. (2004). Toxoplasmosis. *Lancet* 363, 1965–1976. doi: 10.1016/S0140-6736(04)16412-X
- Monzote, L., Rodriguez, M., Alfonso, Y., and Cox, R. (2013). Antiretroviral activity of protease inhibitors against *Toxoplasma gondii*. *Rev. Inst. Med. Trop. S. Paulo* 55, 65–67. doi: 10.1590/S0036-46652013000100012
- Mui, E. J., Schiehs, G. A., Milhous, W. K., Hsu, H., Roberts, C. W., Kirisits, M., et al. (2008). Novel triazine JPC-2067-B inhibits *Toxoplasma gondii* *in vitro* and *in vivo*. *PLoS Negl. Trop. Dis.* 2:e190. doi: 10.1371/journal.pntd.0000190
- Neville, A. J., Zach, S. J., Wang, X., Larson, J. J., Judge, A. K., Davis, L. A., et al. (2015). Clinically available medicines demonstrating anti-toxoplasma activity. *Antimicrob. Agents Chemother.* 59, 7161–7169. doi: 10.1128/AAC.02009-15
- Notarangelo, F. M., Wilson, E. H., Horning, K., Thomas, M., Harris, T., Fang, Q., et al. (2014). Evaluation of kynurenine pathway metabolism in *Toxoplasma gondii*-infected mice: implications for schizizophrenia. *Schizophr. Res.* 152, 261–267. doi: 10.1016/j.schres.2013.11.011
- Ospenica, D., Radivojević, J., Matić, I., Štajner, T., Knežević-Ušaj, S., Djurković-Djaković, O., et al. (2015). Tetraoxanes as inhibitors of Apicomplexan parasites *Plasmodium falciparum* and *Toxoplasma gondii* and anti-cancer molecules. *J. Serb. Chem. Soc.* 80, 1339–1359. doi: 10.2298/JSC150430063O
- Oz, H. S. (2014a). Novel synergistic protective efficacy of atovaquone and diclazuril on fetal-maternal toxoplasmosis. *Int. J. Clin. Med.* 5, 921. doi: 10.4236/ijcm.2014.515124
- Oz, H. S. (2014b). Toxoplasmosis complications and novel therapeutic synergism combination of diclazuril plus atovaquone. *Front. Microbiol.* 5:484. doi: 10.3389/fmicb.2014.00484
- Palencia, A., Liu, R. J., Lukarska, M., Gut, J., Bougdour, A., Touquet, B., et al. (2016). Cryptosporidium and Toxoplasma parasites are inhibited by a benzoxaborole targeting leucyl-tRNA synthetase. *Antimicrob. Agents Chemother.* 60, 5817–5827. doi: 10.1128/AAC.00873-16
- Pappas, G., Rousos, N., and Falagas, M. E. (2009). Toxoplasmosis snapshots: global status of *Toxoplasma gondii* seroprevalence and implications for pregnancy and congenital toxoplasmosis. *Int. J. Parasitol.* 39, 1385–1394. doi: 10.1016/j.ijpara.2009.04.003
- Payne, A. J., Neal, L. M., and Knoll, L. J. (2013). Fusidic acid is an effective treatment against *Toxoplasma gondii* and *Listeria monocytogenes* *in vitro*, but not in mice. *Parasitol. Res.* 112, 3859–3863. doi: 10.1007/s00436-013-3574-1

- Petersen, E., and Schmidt, D. R. (2003). Sulfadiazine and pyrimethamine in the postnatal treatment of congenital toxoplasmosis: what are the options? *Exp. Rev. Anti-Infect. Ther.* 1, 175–182. doi: 10.1586/14787210.1.1.175
- Pissinate, K., dos Santos Martins-Duarte, É., Schaffazick, S. R., de Oliveira, C. P., Vommaro, R. C., Guterres, S. S., et al. (2014). Pyrimethamine-loaded lipid-core nanocapsules to improve drug efficacy for the treatment of toxoplasmosis. *Parasitol. Res.* 113, 555–564. doi: 10.1007/s00436-013-3715-6
- Portes Jde, A., Netto, C. D., da Silva, A. J. M., Costa, P. R. R., DaMatta, R. A., dos Santos, T. A. T., et al. (2012). A new type of pterocarpanquinone that affects *Toxoplasma gondii* tachyzoites *in vitro*. *Vet. Parasitol.* 186, 261–269. doi: 10.1016/j.vetpar.2011.11.008
- Puvanesuaran, V. R., Nowroji, K., Sreenivasan, S., Noordin, R., and Balakrishnan, V. (2012). Use of prednisolone to aid propagation of *Toxoplasma gondii* in mice. *Eur. Rev. Med. Pharmacol. Sci.* 16, 1028–1032.
- Rajapakse, R., Uring-Lambert, B., Andarawewa, K. L., Rajapakse, R. P., Aboubacar, A., Marcellin, L., et al. (2007). 1, 25 (OH) 2D3 inhibits *in vitro* and *in vivo* intracellular growth of apicomplexan parasite *Toxoplasma gondii*. *J. Steroid Biochem. Mol. Biol.* 103, 811–814. doi: 10.1016/j.jsbmb.2006.12.058
- Rezaei, F., Ebrahimzadeh, M. A., Daryani, A., Sharif, M., Ahmadpour, E., and Sarvi, S. (2014). The inhibitory effect of cromolyn sodium and ketotifen on *Toxoplasma gondii* entrance into host cells *in vitro* and *in vivo*. *J. Parasit. Dis.* 40, 1001–1005. doi: 10.1007/s12639-014-0623-3
- Rivera Fernández, N., Castelán, M. M., Pozos, S. G., Flores, C. J. R., González, R. M., de León, C. T. G., et al. (2016). A new type of quinoxalinone derivatives affects viability, invasion, and intracellular growth of *Toxoplasma gondii* tachyzoites *in vitro*. *Parasitol. Res.* 115, 2081–2096. doi: 10.1007/s00436-016-4953-1
- Rodriguez, J. B., and Szajnman, S. H. (2012). New antibacterials for the treatment of toxoplasmosis; a patent review. *Expert Opin. Ther. Pat.* 22, 311–333. doi: 10.1517/13543776.2012.668886
- Saleh, A., Friesen, J., Baumeister, S., Gross, U., and Bohne, W. (2007). Growth inhibition of *Toxoplasma gondii* and *Plasmodium falciparum* by nanomolar concentrations of 1-hydroxy-2-dodecyl-4 (1H) quinolone, a high-affinity inhibitor of alternative (type II) NADH dehydrogenases. *Antimicrob. Agents Chemother.* 51, 1217–1222. doi: 10.1128/AAC.00895-06
- Saraei, M., Ghaderi, Y., Mosavi, T., Shahnazi, M., Nassiri-Asl, M., and Jahanihashemi, H. (2016). The effect of fluphenazine and thioridazine on *Toxoplasma gondii* *in vivo*. *Iran. J. Parasitol.* 11, 226–231. Available online at: <http://tums.ac.ir>
- Saraei, M., Samadzadeh, N., Khoeni, J., Shahnazi, M., Nassiri-Asl, M., and Jahanihashemi, H. (2015). *In vivo* anti-Toxoplasma activity of aripiprazole. *Iran. J. Basic Med. Sci.* 18, 938. doi: 10.22038/ijbms.2015.5219
- Schmidt, D. R., Hogh, B., Andersen, O., Hansen, S. H., Dalhoff, K., and Petersen, E. (2006). Treatment of infants with congenital toxoplasmosis: tolerability and plasma concentrations of sulfadiazine and pyrimethamine. *Eur. J. Pediatr.* 165, 19–25. doi: 10.1007/s00431-005-1665-4
- Serranti, D., Buonsenso, D., and Valentini, P. (2011). Congenital toxoplasmosis treatment. *Eur. Rev. Med. Pharmacol. Sci.* 15, 193–198.
- Shubar, H. M., Lachenmaier, S., Heimesaat, M. M., Lohman, U., Mauludin, R., Mueller, R. H., et al. (2011). SDS-coated atovaquone nanosuspensions show improved therapeutic efficacy against experimental acquired and reactivated toxoplasmosis by improving passage of gastrointestinal and blood-brain barriers. *J. Drug Target.* 19, 114–124. doi: 10.3109/10611861003733995
- Shubar, H. M., Mayer, J. P., Hopfenmüller, W., and Liesenfeld, O. (2008). A new combined flow-cytometry-based assay reveals excellent activity against *Toxoplasma gondii* and low toxicity of new bisphosphonates *in vitro* and *in vivo*. *J. Antimicrob. Chemother.* 61, 1110–1119. doi: 10.1093/jac/dkn047
- Silveira, C., Belfort, R. Jr., Muccioli, C., Holland, G. N., Victora, C. G., Horta, B. L., et al. (2002). The effect of long-term intermittent trimethoprim/sulfamethoxazole treatment on recurrences of toxoplasmic retinochoroiditis. *Am. J. Ophthalmol.* 134, 41–46. doi: 10.1016/S0002-9394(02)01527-1
- Smith, A. T., Livingston, M. R., Mai, A., Filetici, P., Queener, S. F., and Sullivan, W. J. (2007). Quinoline derivative MC1626, a putative GCN5 histone acetyltransferase (HAT) inhibitor, exhibits HAT-independent activity against *Toxoplasma gondii*. *Antimicrob. Agents Chemother.* 51, 1109–1111. doi: 10.1128/AAC.01256-06
- Sönmez, N., Büyükbaba Boral, O., Kaşali, K., and Tekeli, F. (2014). [Effects of atovaquone and astragalus combination on the treatment and IL-2, IL-12, IFN- γ levels on mouse models of acute toxoplasmosis]. *Mikrobiyol. Bul.* 48, 639–651. doi: 10.5578/mb.8025
- Strobl, J. S., Cassell, M., Mitchell, S. M., Reilly, C. M., and Lindsay, D. S. (2007). Scriptaid and suberoylanilide hydroxamic acid are histone deacetylase inhibitors with potent anti-*Toxoplasma gondii* activity *in vitro*. *J. Parasitol.* 93, 694–700. doi: 10.1645/GE-1043R.1
- Strobl, J. S., Seibert, C. W., Li, Y., Nagarkatti, R., Mitchell, S. M., Rosypal, A. C., et al. (2009). Inhibition of *Toxoplasma gondii* and *Plasmodium falciparum* infections *in vitro* by NSC3852, a redox active antiproliferative and tumor cell differentiation agent. *J. Parasitol.* 95, 215–223. doi: 10.1645/GE-1608.1
- Sugi, T., Kato, K., Kobayashi, K., Kurokawa, H., Takemae, H., Gong, H., et al. (2011). 1NM-PP1 treatment of mice infected with *Toxoplasma gondii*. *J. Vet. Med. Sci.* 73, 1377–1379. doi: 10.1292/jvms.11-0085
- Surolia, N., and Surolia, A. (2001). Triclosan offers protection against blood stages of malaria by inhibiting enoyl-ACP reductase of *Plasmodium falciparum*. *Nat. Med.* 7, 167–173. doi: 10.1038/84612
- Szabo, E. K., and Finney, C. A. (2016). *Toxoplasma gondii*: one organism, multiple models. *Trends Parasitol.* doi: 10.1016/j.pt.2016.11.007. [Epub ahead of print].
- Szajnman, S. H., Galaka, T., Li, Z.-H., Li, C., Howell, N. M., Chao, M. N., et al. (2016). 1. *In vitro* and *in vivo* activity of sulfur-containing linear bisphosphonates against apicomplexan parasites. *Antimicrob. Agents Chemother.* doi: 10.1128/AAC.01590-16. [Epub ahead of print].
- Szajnman, S. H., García Liñares, G. E., Li, Z.-H., Jiang, C., Galizzi, M., Bontempi, E. J., et al. (2008). Synthesis and biological evaluation of 2-alkylaminoethyl-1, 1-bisphosphonic acids against *Trypanosoma cruzi* and *Toxoplasma gondii* targeting farnesyl diphosphate synthase. *Bioorg. Med. Chem.* 16, 3283–3290. doi: 10.1016/j.bmc.2007.12.010
- Tenter, A. M., Heckerroth, A. R., and Weiss, L. M. (2000). *Toxoplasma gondii*: from animals to humans. *Int. J. Parasitol.* 30, 1217–1258. doi: 10.1016/S0020-7519(00)00124-7
- Teo, C. F., Zhou, X. W., Bogoyo, M., and Carruthers, V. B. (2007). Cysteine protease inhibitors block *Toxoplasma gondii* microneme secretion and cell invasion. *Antimicrob. Agents Chemother.* 51, 679–688. doi: 10.1128/AAC.01059-06
- Tipparaju, S. K., Muench, S. P., Mui, E. J., Ruzheinikov, S. N., Lu, J. Z., Hutson, S. L., et al. (2010). Identification and development of novel inhibitors of *Toxoplasma gondii* enoyl reductase. *J. Med. Chem.* 53, 6287–6300. doi: 10.1021/jm9017724
- Torgerson, P. R., and Mastroiacovo, P. (2013). The global burden of congenital toxoplasmosis: a systematic review. *Bull. World Health Organ.* 91, 501–508. doi: 10.2471/BLT.12.111732
- Vanagas, L., Jeffers, V., Bogado, S. S., Dalmaso, M. C., Sullivan, W. J. Jr., and Angel, S. O. (2012). Toxoplasma histone acetylation remodelers as novel drug targets. *Expert Rev. Anti Infect. Ther.* 10, 1189–1201. doi: 10.1586/eri.12.100
- Vidadala, R. S. R., Rivas, K. L., Ojo, K. K., Hulverson, M. A., Zambriski, J. A., Bruzual, I., et al. (2016). Development of an orally available and Central Nervous System (CNS) penetrant *Toxoplasma gondii* calcium-dependent protein Kinase 1 (Tg CDPK1) inhibitor with minimal human Ether-a-go-go-Related Gene (hERG) activity for the treatment of toxoplasmosis. *J. Med. Chem.* 59, 6531–6546. doi: 10.1021/acs.jmedchem.6b00760
- Wei, S., Daniel, B. J., Brumlik, M. J., Burrow, M. E., Zou, W., Khan, I. A., et al. (2007). Drugs designed to inhibit human p38 mitogen-activated protein kinase activation treat *Toxoplasma gondii* and *Encephalitozoon cuniculi* infection. *Antimicrob. Agents Chemother.* 51, 4324–4328. doi: 10.1128/AAC.00680-07
- Welsch, M. E., Zhou, J., Gao, Y., Yan, Y., Porter, G., Agnihotri, G., et al. (2016). Discovery of potent and selective leads against *Toxoplasma gondii* dihydrofolate reductase via structure-based design. *ACS Med. Chem. Lett.* 7, 1124–1129. doi: 10.1021/acsmedchemlett.6b00328
- Wiengcharoen, J., O., Hanley, R., Armstrong, T., Best, W., Sukthana, Y., and Thompson, R. A. (2007). Novel drug compounds against *Neospora caninum* and *Toxoplasma gondii* *in vitro*. *Southeast Asian J. Trop. Med. Public Health* 38, 15.
- Xin, C. F., Kim, H. S., Sato, A., Lee, H. J., Lee, Y. W., Pyo, K. H., et al. (2016). *In vitro* inhibition of *Toxoplasma gondii* by the anti-malarial candidate, 6-(1, 2, 6, 7-tetraoxaspiro [7.11] nonadec-4-yl) hexan-1-ol. *Parasitol. Int.* 65, 494–499. doi: 10.1016/j.papint.2016.06.013

- Yang, Z., Ahn, H. J., and Nam, H. W. (2014). Gefitinib inhibits the growth of *Toxoplasma gondii* in HeLa cells. *Korean J. Parasitol.* 52:439. doi: 10.3347/kjp.2014.52.4.439
- Yeo, S. J., Jin, C., Kim, S., and Park, H. (2016). *In vitro* and *in vivo* effects of nitrofurantoin on experimental toxoplasmosis. *Korean J. Parasitol.* 54:155. doi: 10.3347/kjp.2016.54.2.155
- Yin, W. D., Gao, Q. C., Liu, X. D., and Tang, H. W. (2009). [In vivo effect of dihydroartemisinin and azithromycin on the ultrastructure of *Toxoplasma gondii* tachyzoites]. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 27, 4325–4327.
- Yung, S. C., and Lang-Unnasch, N. (2004). “Targeting the *Toxoplasma gondii* apicoplast for chemotherapy,” in *Opportunistic Infections: Toxoplasma, Sarcocystis, and Microsporidia*, eds D. S. Lindsay and L. M. Weiss (Boston, MA: Springer), 39–49.
- Zhang, N. Z., Chen, J., Wang, M., Petersen, E., and Zhu, X. Q. (2013). Vaccines against *Toxoplasma gondii*: new developments and perspectives. *Expert Rev. Vaccines* 12, 1287–1299. doi: 10.1586/14760584.2013.844652
- Zhang, N. Z., Wang, M., Xu, Y., Petersen, E., and Zhu, X. Q. (2015). Recent advances in developing vaccines against *Toxoplasma gondii*: an update. *Expert Rev. Vaccines* 14, 1609–1621. doi: 10.1586/14760584.2015.1098539
- Zhou, Y., Fomovska, A., Muench, S., Lai, B. S., Mui, E., and McLeod, R. (2014). Spiroindolone that inhibits PfATPase4 is a potent, cidal inhibitor of *Toxoplasma gondii* tachyzoites *in vitro* and *in vivo*. *Antimicrob. Agents Chemother.* 58, 1789–1792. doi: 10.1128/AAC.02225-13

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Montazeri, Sharif, Sarvi, Mehrzadi, Ahmadpour and Daryani. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.