



A Network of Paralogous Stress Response Transcription Factors in the Human Pathogen *Candida glabrata*

Jawad Merhej¹, Antonin Thiebaut¹, Corinne Blugeon², Juliette Pouch², Mohammed El Amine Ali Chaouche², Jean-Michel Camadro³, Stéphane Le Crom⁴, Gaëlle Lelandais^{3*} and Frédéric Devaux^{1*}

¹ Laboratoire de Biologie Computationnelle et Quantitative, Centre National de la Recherche Scientifique, Institut de Biologie Paris-Seine, UMR 7238, Sorbonne Universités, Université Pierre et Marie Curie, Paris, France, ² École Normale Supérieure, Paris Sciences et Lettres Research University, Centre National de la Recherche Scientifique, Institut National de la Santé et de la Recherche Médicale, Institut de Biologie de l'École Normale Supérieure, Plateforme Génomique, Paris, France, ³ Centre National de la Recherche Scientifique, UMR 7592, Institut Jacques Monod, Université Paris Diderot, Sorbonne Paris Cité, Paris, France, ⁴ Évolution, Centre National de la Recherche Scientifique, Institut de Biologie Paris-Seine, UMR 7138, Sorbonne Universités, Université Pierre et Marie Curie, Paris, France

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*Correspondence:

Gaëlle Lelandais
gaelle.lelandais@univ-paris-diderot.fr;
Frédéric Devaux
Frederic.devaux@upmc.fr

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The yeast *Candida glabrata* has become the second cause of systemic candidemia in humans. However, relatively few genome-wide studies have been conducted in this organism and our knowledge of its transcriptional regulatory network is quite limited. In the present work, we combined genome-wide chromatin immunoprecipitation (ChIP-seq), transcriptome analyses, and DNA binding motif predictions to describe the regulatory interactions of the seven Yap (Yeast AP1) transcription factors of *C. glabrata*. We described a transcriptional network containing 255 regulatory interactions and 309 potential target genes. We predicted with high confidence the preferred DNA binding sites for 5 of the 7 CgYaps and showed a strong conservation of the Yap DNA binding properties between *S. cerevisiae* and *C. glabrata*. We provided reliable functional annotation for 3 of the 7 Yaps and identified for Yap1 and Yap5 a core regulon which is conserved in *S. cerevisiae*, *C. glabrata*, and *C. albicans*. We uncovered new roles for CgYap7 in the regulation of iron-sulfur cluster biogenesis, for CgYap1 in the regulation of heme biosynthesis and for CgYap5 in the repression of *GRX4* in response to iron starvation. These transcription factors define an interconnected transcriptional network at the cross-roads between redox homeostasis, oxygen consumption, and iron metabolism.

Keywords: yeast, Yap, ChIP-seq, transcriptome, regulatory networks, evolution

INTRODUCTION

Candida glabrata is a unicellular yeast from the Hemiascomycetes phylogenetic group and a commensal host of the human mucosal microbiota. However, in patients with severe immunodeficiency, it can cause invasive systemic infections, with high mortality rates (about 40–60%). While *Candida albicans* remains the main cause of systemic candidiasis (50–70%), *C. glabrata* ranks second (20–25%), and its prevalence has increased in the last decades (Perlroth et al., 2007). An important prerequisite to the acquisition of virulence traits in *C. glabrata* is its

ability to adapt and be resistant to environmental variations, which allows the pathogen to colonize many different niches and organs in the human body, with very different features in terms of pH, temperature, redox potential, iron, zinc, or oxygen availability, etc. (Domergue et al., 2005; Nevitt and Thiele, 2011). Moreover, *C. glabrata* is able to survive and grow in the harsh environment of the phagolysosomes of macrophages (Seider et al., 2011, 2014).

The mechanisms that allow these adaptations rely partially on transcriptional regulatory networks. Systematic combination of transcriptome analyses of loss of function mutants for different transcription factors and of genome-wide chromatin immunoprecipitation has led to the comprehensive description of genome-wide transcriptional regulatory networks in the model yeast *Saccharomyces cerevisiae* (Babu et al., 2004; Harbison et al., 2004; Teixeira et al., 2014). These global approaches have also been extensively used in *C. albicans*. In this species, more than 75 specific transcription factors (over a total of about 150) have been analyzed by genome-wide approaches and comprehensive network-based models are available for ribosome biogenesis, iron homeostasis, or biofilm formation (Lavoie et al., 2010; Chen et al., 2011; Nobile et al., 2012; Fox et al., 2015). In contrast, the global transcriptional regulation in *C. glabrata* has been poorly studied. As of 2016, only 10 transcription factors have been analyzed on a genome-wide scale in this species (Vermitsky et al., 2006; Lelandais et al., 2008; Roetzer et al., 2008, 2011; Kuo et al., 2010a; Caudle et al., 2011; Ferrari et al., 2011; Miyazaki et al., 2013; Noble et al., 2013; Paul et al., 2014; Merhej et al., 2015; Wu et al., 2015). While *C. glabrata* and *C. albicans* share a common ecological niche and are from the same genus, they are very distant species according to their genomic sequence, and their ability to successfully infect humans involves quite different strategies (Brunke and Hube, 2013). For instance, in contrast to *C. albicans*, the ancestor of *C. glabrata* and *S. cerevisiae* experienced a whole-genome duplication event. In addition, *C. albicans* is usually diploid, switching frequently from yeast to hyphal growth under stress conditions, while *C. glabrata* is strictly haploid and grows mostly in the yeast form. Therefore, the *C. glabrata* transcriptional networks cannot be simply inferred from the knowledge acquired in *C. albicans* (Gabaldon and Carrete, 2015).

In the present work, we conducted a network-based analysis of the seven transcription factors belonging to the Yap (Yeast AP1) family in *C. glabrata*. The Yap proteins belong to the pap subfamily of bZIP transcription factors and are homologous to the CREB, ATF2, and Fos/Jun transcription factors of vertebrates (Fujii et al., 2000; Reinke et al., 2013). The model yeast *S. cerevisiae* has 8 Yap members (named ScYap thereafter; Supplementary File S1), most of which are involved in adaptation to environmental changes (Rodrigues-Pousada et al., 2010). ScYap1 is the major regulator of oxidative stress responses caused by reactive oxygen species (ROS), metals and drugs (reviewed in Rodrigues-Pousada et al., 2010). ScYap2 (also named Cad1), the ohnolog of Yap1, is involved in cadmium resistance (Hirata et al., 1994; Azevedo et al., 2007; Mazzola et al., 2015). The role of ScYap3 is unknown but it has been shown to contribute to the resistance to benzenic compounds and to 6-Nonadecynoic acid

(North et al., 2011; Xu et al., 2012; Adeboye et al., 2014). The ScYap4 (also named Cin5) and ScYap6 ohnologs are involved in salt stress response (Mendizabal et al., 1998; Nevitt et al., 2004; Ni et al., 2009). ScYap5 is responsible for the activation of the high iron stress response (Li et al., 2008; Pimentel et al., 2012). ScYap7, the ohnolog of ScYap5, was recently found to be a repressor of the nitric oxide oxidase encoding gene *YHB1* (Merhej et al., 2015) and ScYap8 is involved in the response to arsenic (Bobrowicz et al., 1997; Amaral et al., 2013; Kumar et al., 2015).

C. glabrata has 7 Yap members, which slightly differ from the *S. cerevisiae* family (Supplementary File S1). Orthologues of Yap1 (*CAGL0H04631g*), Yap2 (*CAGL0F03069g*, named CgYap1 thereafter), Yap5 (*CAGL0K08756g*, named CgYap5 thereafter), and Yap7 (*CAGL0F01265g*, named CgYap7 thereafter) are present, but Yap8 is absent. Two versions of Yap3 (*CAGL0K02585g* and *CAGL0M10087g*, named CgYap3a and CgYap3b thereafter) and only one ortholog for the Yap4 and Yap6 pair (*CAGL0M08800g*, named CgYap4/6 thereafter) are found. Only three of these Yaps have been studied previously in *C. glabrata*. CgYap1 has been shown to be involved in oxidative stress response, with a set of targets which is significantly conserved compared to ScYap1, but with different DNA binding preferences (Chen et al., 2007; Cuellar-Cruz et al., 2008; Lelandais et al., 2008; Kuo et al., 2010a; Goudot et al., 2011; Roetzer et al., 2011). CgYap7 has been shown to have a conserved role in nitric oxide oxidase repression (Merhej et al., 2015). Finally, as in *S. cerevisiae*, CgYap5 is involved in the activation of the *CCC1* and *GRX4* genes under high iron conditions (Merhej et al., 2015). In the present work, we conducted chromatin immunoprecipitation experiments followed by high-throughput sequencing (ChIP-seq) and transcriptome analyses to determine the targets for the seven Yap transcription factors of *C. glabrata*. The CgYap network included 309 genes and 255 regulatory interactions. From these results, we could predict with high confidence the preferred DNA binding sites for 5 of the 7 CgYaps and show a strong conservation of the Yap DNA binding properties between *S. cerevisiae* and *C. glabrata*. We provided functional annotation for 3 of the 7 CgYaps and identified for Yap1 and Yap5 a core regulon which is conserved in *S. cerevisiae*, *C. glabrata*, and *C. albicans*. Our data pointed out new roles for CgYap7 in the regulation of iron-sulfur cluster biogenesis, for CgYap1 in the regulation of heme biosynthesis and for CgYap5 in the repression of *GRX4* in response to iron starvation.

MATERIALS AND METHODS

Strains

The list of the strains used in this study is available in Supplementary File S2. All the strains were derived from the Δ HTU parental strain (Kitada et al., 1995). The genomic myc-tagging and deletion of the different CgYAP was performed as described previously (Merhej et al., 2014, 2015). Briefly, deletion or myc-tagging cassettes were PCR amplified from the *M. Longtine's* plasmids (Longtine et al., 1998) with oligonucleotides containing in 5' homology sequences flanking the desired genomic insertion points. At least 10 micrograms of purified PCR product was used to transform Δ HTU cells using a standard yeast

transformation protocol (Merhej et al., 2015). Genotyping of the clones growing on selective media was done by PCR. The PCR-verified clones for the knock-out were then verified by southern blot (Merhej et al., 2015). The correct myc-tagging of the *CgYAP* was verified by sequencing of the gene and western blot (Merhej et al., 2015). All the oligonucleotides used for cassette preparation and genotyping are listed in Supplementary File S2.

Yeast Cultures and Growth Conditions

All cultures were conducted in a rotative shaker at 30°C in YPD (Glucose 2%, yeast extract 1%, Bactopeptone 1%). Stress conditions used were: 1 mM sodium selenite, 1 M NaCl, 2 mM cadmium chloride, 5 mM iron sulfate, or 0.5 mM bathophenanthroline disulfonate (BPS). These doses were chosen, based on preliminary microarray and growth assay experiments, to induce a transcriptional response in the wild type without causing significant differences of growth rates between the wild type and the *CgYAP* knock-out mutants (data not shown).

Chromatin Immunoprecipitation and High-Throughput Sequencing

For ChIP, myc-tagged *CgYAP* strains were grown in YPD until exponential phase (OD = 0.8) and then stressing agents were added for 30 min. Cross-linking of the cells and ChIP were performed as described previously (Lelandais et al., 2016). The parental Δ HTU (untagged strain) was grown and processed the same way to provide the mock-IP samples. Sequencing of the IPs, Input DNAs and mock IPs samples and primary data analyses (quality controls and mapping of the reads) were performed as described previously (Lelandais et al., 2016). All experiments were performed twice and the reads of the replicate averaged before the peak calling step, except for *CgYap5* for which one of the two replicates had poor read coverage and was not used for further analyses. Peak calling was performed with the bpeaks software (Merhej et al., 2014), using both the Input DNA and the mock IP as references. For peak calling using the Input DNA as reference, the bpeaks parameters were T1 = 2, T2 = 2, T3 = 1.5, T4 = 0.7. For peak calling using the Mock IP as reference, the bpeaks parameters were T1 = 2, T2 = 2, T3 = 1.5, and T4 = 0. Only the peaks which were found by the two analyses were kept for further processing. These peaks were then manually checked on a genome browser (Thorvaldsdottir et al., 2013) to discard artifactual peaks (e.g., peaks centered on a tRNA locus or perfectly overlapping a highly expressed ORF) which would have escaped the bpeaks filter (Supplementary Files S3, S4). The ChIP seq data can be downloaded from the GEO database (accession number: GSE77904).

Transcriptome Analyses

Knock-out and wild type strains were grown in 50 mL of YPD until exponential phase (OD = 0.8) and then stressing agents were added. After 30 min, 20 mL of each cell cultures were flash-frozen in two volumes of cold ethanol and collected by centrifugation. The OD of the cultures were monitored before the stress treatment and every 30 min for 2 h after stress treatment. Only samples from wild type and knock-out cultures which showed

similar growth rates ($\pm 10\%$) (Thompson et al., 2013) were used for transcriptome comparisons. Total RNA was extracted, quality controlled and quantified as described previously (Merhej et al., 2015). One microgram of total RNA was used for fluorescent cDNA synthesis according to the amino-allyl protocol (Merhej et al., 2015). The cDNA were labeled with Cy3 and Cy5 and hybridization was performed as previously described (Merhej et al., 2015). Two biologically independent experiments were performed for each condition, using dye switch. We used custom *C. glabrata* Agilent arrays in an 8 × 60 k format (array express accession number: A-MEXP-2402). After overnight hybridization and washing, the slides were scanned using a 2-micron Agilent microarray scanner. The images were analyzed using the feature extraction software (Agilent technologies) and normalized using global LOESS (Lemoine et al., 2006). The mean of the biological replicates was calculated. A gene was considered as being differentially expressed if its mean absolute Log₂(fold change) value was more than 0.75 and if its expression variation was considered as being statistically significant using the LIMMA package with a cut-off *p*-value of 0.02 (Ritchie et al., 2015). The complete microarray data are available at Array express database under the accession number: E-MTAB-4457.

TFBS Predictions

DNA sequences of ChIP peaks were retrieved from their genomic locations (BED file) using the “getfasta” function from the BEDTOOLS suite (Quinlan and Hall, 2010). These genomic sequences were used as inputs for the “peak-motif” tool to search for regulatory motifs (Thomas-Chollier et al., 2012). An additional filtering step was added to the standard peak motif procedure to discard low complexity motifs (e.g., CCCCCC) or motifs which were found in <20% of the peaks (Supplementary File S5).

Network Building

The ChIP peaks were assigned to genes as described previously (Merhej et al., 2014). When a peak was located in a divergent promoter (i.e., an intergenic region in between two divergent genes) the two genes were fused in one target in the network named “gene 1/gene 2,” unless we had transcriptome evidence supporting the regulation of one of the two genes. In this case, only the name of the regulated gene was kept. The network was represented using the igraph library (igraph.org, R programming language; Csardi and Nepusz, 2006), combining three different types of information (Supplementary Table 1). The ChIP parameter was used to define interactions (arrows) between the different Yap factors and their target promoters. The transcriptome parameter was used to color arrows depending on the directionality of the regulation (activation, repression, or no detected change). The TFBS parameter was used to color target promoters depending of the presence of the identified TFBS in the corresponding ChIP peaks.

Gene Ontology and Gene Set Enrichment Analyses

GO analyses were performed using the “GO term finder” tool at the CGD database, with default parameters (Ingliš et al., 2012).

GSEA were performed using the GSEA module of the Jexpress software suite with a cut-off FDR of 1% (Subramanian et al., 2005; Stavrum et al., 2008). We used as input files transcriptome analyses of the responses of *C. glabrata* wild type cells to fluconazole (Kuo et al., 2010b), sodium salt (Roetzer et al., 2008; Wapinski et al., 2010), heat shock (Roetzer et al., 2008; Wapinski et al., 2010), hydrogen peroxide (Wapinski et al., 2010; Roetzer et al., 2011), menadione (Roetzer et al., 2011), glucose starvation (Roetzer et al., 2008), sorbic acid (Jandric et al., 2013), iron excess and iron starvation (this work, E-MTAB-4457), cadmium chloride and sodium selenite (Thiebaut et al., unpublished data).

RESULTS

The Yap Network in *C. glabrata*

To identify the gene targets and characterize the regulatory interactions of the seven Yap transcription factors of *C. glabrata*, we used three different approaches. First, we performed ChIP-seq experiments using myc-tagged versions of each factor. For CgYap1, CgYap2, CgYap4/6, and CgYap5, we submitted the corresponding tagged strains to stress conditions known to induce full activity of their *S. cerevisiae* orthologues, i.e., oxidative stress caused by a metalloid (namely selenium) for Yap1 (Haugen et al., 2004; Salin et al., 2008), cadmium for Yap2 (Azevedo et al., 2007; Mazzola et al., 2015), salt excess for Yap4/6 (Nevitt et al., 2004; Ni et al., 2009), and iron excess for Yap5 (Li et al., 2008; Pimentel et al., 2012). For CgYap7, which was shown to have a constitutive activity (Merhej et al., 2015), the experiments were performed in standard growth conditions. For the two orthologs of Yap3, whose role remains unknown in *S. cerevisiae*, the experiments were performed using cells grown in YPD and cells exposed to a pleiotropic stress inducer (selenite; Salin et al., 2008). The ChIP-seq data were analyzed taking as a reference both the input control and the mock IP control, to sort out a maximum of the false positive peaks due to highly expressed loci (Park et al., 2013; Teytelman et al., 2013). Second, we used the ChIP-seq data to predict the preferred Transcription Factor DNA binding sites (TFBS) for each CgYap. Reciprocally, we identified all the ChIP-peaks which contain the predicted consensus in their promoter sequence. Third, we compared the transcriptome of wild type and null mutants for each CgYap, using the same growth conditions as those used for ChIP-seq experiments. Hence, we identified the genes for which expression was altered, directly or indirectly, in the absence of the corresponding transcription factor.

To build a network from these three sources of information, we used a scoring system based on simple but meaningful logical rules (Supplementary Table 1). Briefly, a regulatory interaction was included in the network if it was detected by ChIP-seq. Then, the interactions and the edges were differently labeled depending on the transcriptome and TFBS data (Figure 1). As a consequence, the different interactions in the final network do not have the same value, depending on whether they were supported by one, two, or three experimental evidences (Supplementary Table 1). The final CgYap network contained 6 transcription factors and 255 regulatory interactions involving 214 promoters and 309 potential target genes (Figure 1). We could not identify any target gene for CgYap3a, neither by

ChIP-seq or by transcriptome analyses. Notably, 62% of the interactions in the network were supported by at least two evidences (Figure 1). The majority of the ChIP targets that we identified had only one peak in their promoter. However, in few cases, several binding sites could be unambiguously detected for CgYap1 and CgYap7 (Supplementary File S6). The number of interactions for each CgYap was highly variable, from 3 for CgYap2, up to 118 for CgYap7 (Figure 1). Relatively few redundancies were observed between the different Yaps, only 38 genes (18% of the edges) are targeted by more than one CgYap.

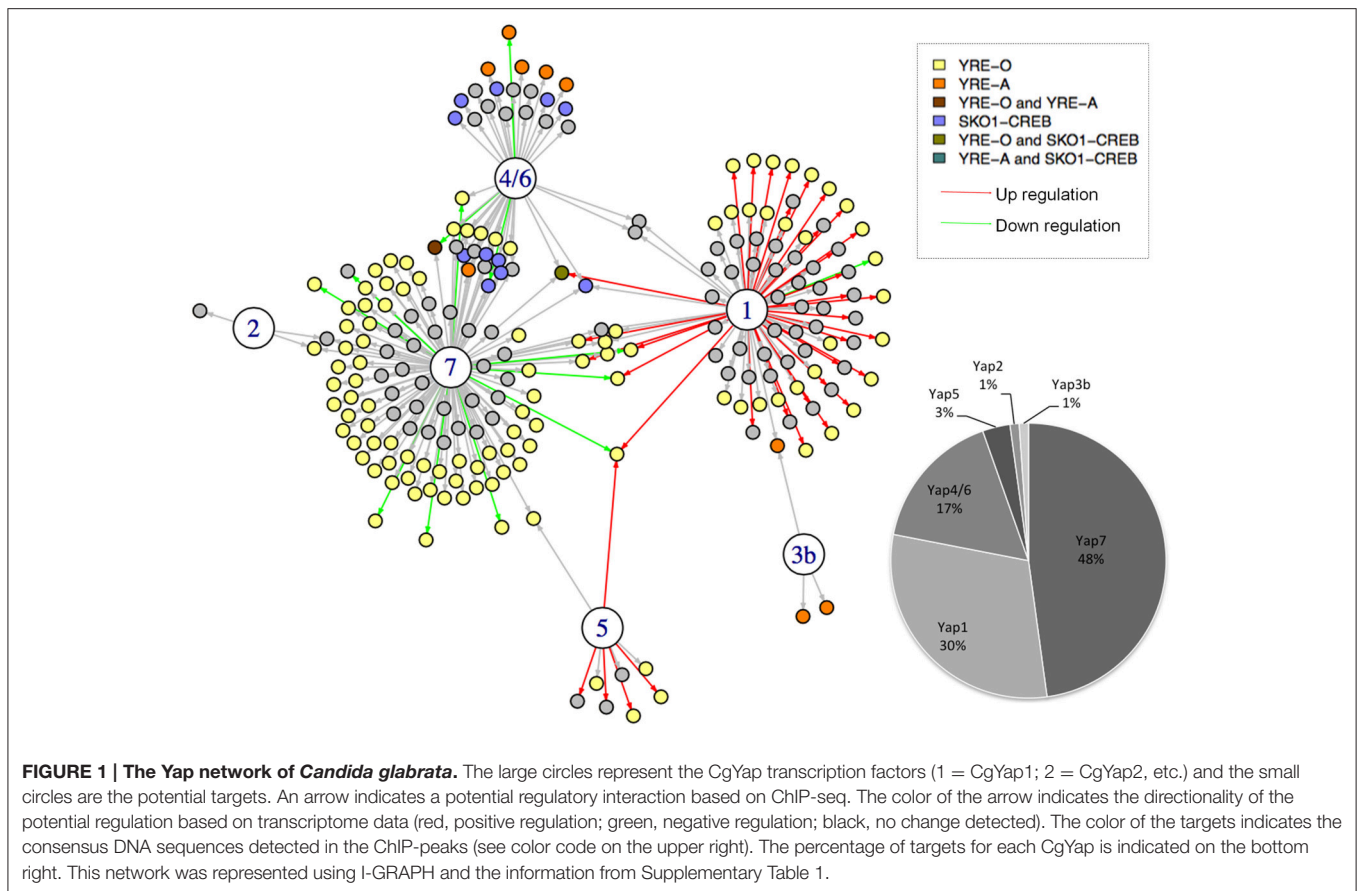
C. glabrata Yap Transcription Factor Binding Sites

As mentioned above, the peaks identified from ChIP-seq data were used to predict the TFBS for 5 of the 7 studied CgYap (Figure 2). In *S. cerevisiae*, the Yap proteins were classified in two categories based on their preferred TFBS (named YRE for Yap Response Elements): ScYap1, ScYap2, ScYap5, and ScYap7 bind to YRE-O (TTACTAA) motifs while ScYap3, ScYap4, and ScYap6 rather recognize YRE-A (TTACGTAA) motifs (Tan et al., 2008; Kuo et al., 2010a). We found that this dichotomy was remarkably conserved in *C. glabrata* (Figure 2). The predicted binding sites for CgYap1, CgYap5 and CgYap7 were very close to the perfect YRE-O consensus. In contrast, the YRE-A motif was enriched in the ChIP peaks of CgYap3b and CgYap4/6. These YRE motifs were identified as the best predicted motifs for all Yaps, except for CgYap4/6 (Figure 2, Supplementary File S5). For this transcription factor, the best identified motif was ATGACGTCAT, which differs from the canonical YRE-A motif by its higher GC content and which actually corresponds to the consensus motif published for another bZIP subfamily, the CREB/ATF2 factors (Fujii et al., 2000). Interestingly enough, in *S. cerevisiae* this motif was associated to the Sko1 transcription factor, which is a yeast homolog of the ATF2/CREB factors (Pascual-Ahuir et al., 2001; Gordan et al., 2011). Sko1 has been shown to contribute to the salt stress response of *S. cerevisiae*, together with ScYap4 and ScYap6, and to share a large number of targets with these factors (Ni et al., 2009).

The proportion of ChIP peaks containing the best predicted TFBS was remarkably high (from 41% for Yap1 up to 100% for Yap3b) (Figures 1, 2). This provided a posteriori confirmation that our ChIP-seq analyses procedure efficiently filtered out the false positive peaks.

Directionality of the CgYap Activities

Because we included transcriptome data in our network, we could determine the directionality (i.e., activation or repression) of some of the regulatory interactions set up by the different Yap proteins (Figure 1). This allowed us to predict the activator or repressor nature of these CgYaps. We observed that, in the conditions that were studied (selenite, excess of iron, salt stress, or YPD, respectively), CgYap1 and CgYap5 were strict activators, while CgYap4/6 and CgYap7 were strict repressors (Figure 1). The deletion of CgYap2 and CgYap3b had no effect on their targets in the conditions that we studied and therefore we had no information on their activity. The overlaps between transcriptome results and ChIP-seq results were remarkably



high in the sense that, taking into account the directionality mentioned above, between 40 and 90% of the expression changes observed in the transcriptome analyses involved genes to which promoters were bound by the TF according to ChIP-seq data (Supplementary File S7). The reciprocal was not true for CgYap7 and CgYap4/6, for which most of the ChIP targets showed unchanged expression in the mutant. This may be due either to functional redundancies for the regulation of these genes or because our transcriptome experiments were not conducted in conditions in which these regulations were active.

Functional Annotation of the Yap Network

We performed Gene Ontology (GO) term enrichment analyses on the whole set of genes in the network and on the individual sets of target of each CgYap (Figure 3, Supplementary Table 1). The whole network was enriched in GO categories related to oxido-reduction processes and iron homeostasis (Figure 3). CgYap1, CgYap5, and CgYap7 were the main contributors to these categories, while CgYap2, CgYap3b, and CgYap4/6 did not show any significant enrichment. More specifically, CgYap1 was the main contributor of targets related to oxidative stress response, oxido-reduction and chemical stress response (e.g., *TRR1*, *TRX2*, *OYE2*, *GPX2*, *TSA1*, *CTA1*, *SRX1*, ...). Its target set was also enriched in genes involved in heme biosynthesis (*HEM1*, *HEM3*, *HEM15*, and *HEM2*). CgYap5 target list was

clearly associated to iron sulfur cluster binding (*ISA1*, *TYW1*, *ACO1*, *RLI1*, *SDH2*, *GLT1*) and iron homeostasis (*GRX4*, *CCC1*). The only GO category to be enriched in CgYap7 targets was iron sulfur cluster metabolism. This includes genes involved in the cytosolic and mitochondrial iron sulfur assembly pathway (*CIA1*, *CIA2*, *DRE2*, *NAR1*, *CFD1*, *IBA57*, *JAC1*) and in iron sulfur cluster binding (*LYS4*, *SDH2*). CgYap7 also targets 10 genes encoding oxidoreductases (*CCP1*, *ERG11*, *YHB1*, *OYE2*, etc...) and genes encoding heme containing proteins (*YHB1*, *CCP1*) or related to heme metabolism (*HEM3*, *CYC3*). The targets of CgYap1 and CgYap7 included many genes encoding transcription factors. For instance, several transcription factors involved in oxygen homeostasis and oxidative stress responses are targeted by CgYap1 (*ROX1*, *MSN4*, *RPN4*, *SKN7*, *IXR1*). Remarkably, CgYap1 and CgYap7 both bound their own promoter, suggesting auto-regulation (Supplementary Table 1).

Conservation of the CgYap1/5/7 Subnetwork

The functional annotation presented above pointed out a CgYap1/CgYap5/CgYap7 network centered on iron and redox homeostasis. To assess the conservation of this sub-network we compared the targets of CgYap1, CgYap5, and CgYap7 with the targets of their orthologues in *S. cerevisiae* and *C. albicans* (Li et al., 2008, 2011; Salin et al., 2008; Znaidi et al., 2009; Chen

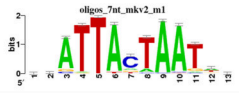
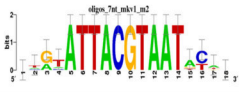


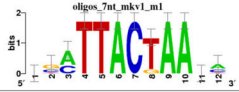

TF	Motifs in <i>C. glabrata</i> (this work)	Motif type	% of ChIP peaks with motif	Motifs in <i>C. glabrata</i> (previous studies)	Motifs in <i>S. cerevisiae</i> (previous studies)
Yap1		YRE-O	41%	YRE-A and/or YRE-O	YRE-O
Yap2	ND	NA	NA	ND	YRE-O
Yap3a	ND	NA	NA	ND	YRE-A
Yap3b		YRE-A	100%	ND	
Yap4/6		Sko1/CREB	46%	ND	YRE-A
		YRE-A	26%		
Yap5		YRE-O	67%	ND	YRE-O
Yap7		YRE-O	66%	ND	YRE-O

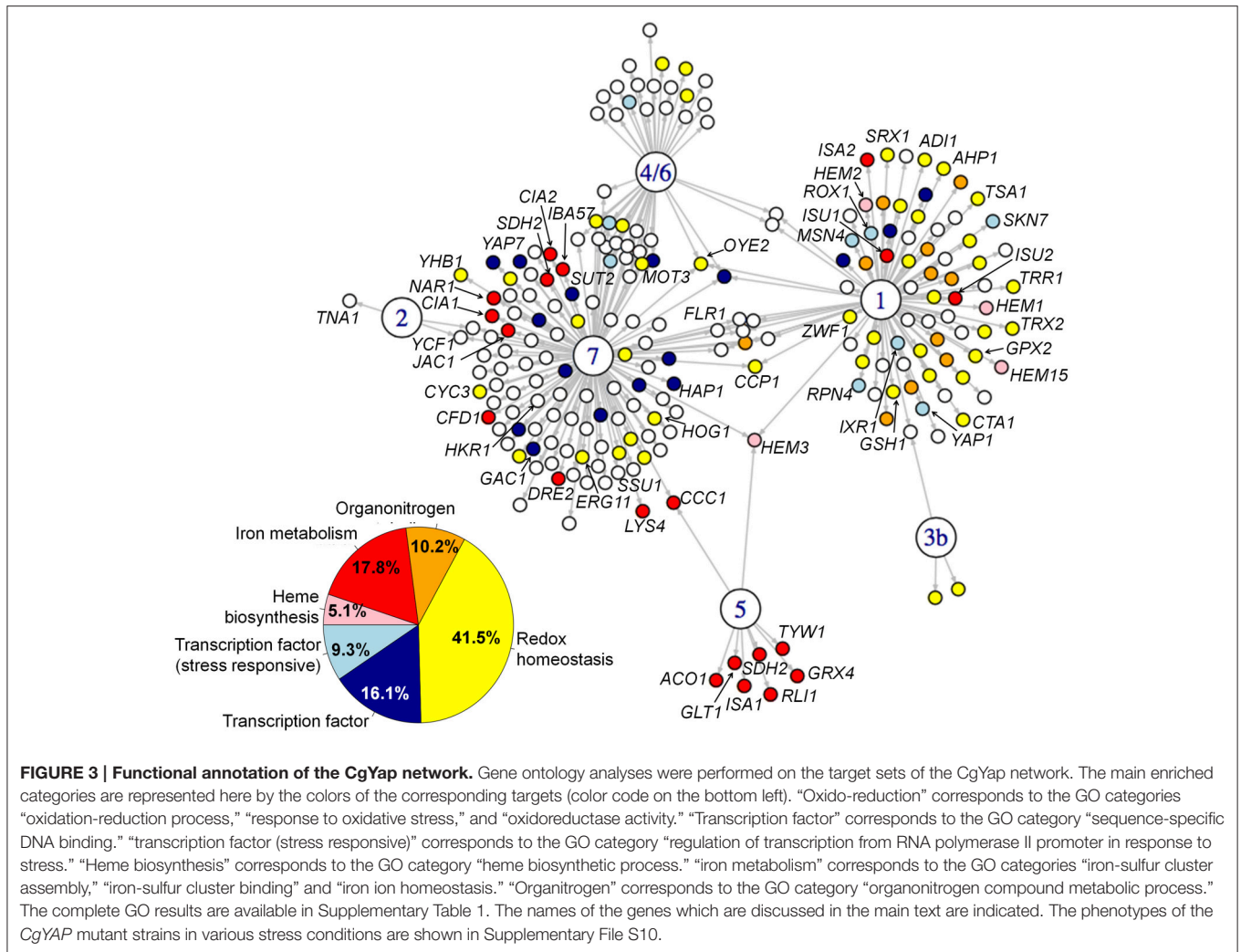
FIGURE 2 | CgYaps Transcription Factor Binding Sites predictions based on ChIP peaks. The motifs were predicted from the ChIP-peaks sequences using Peak Motif (Thomas-Chollier et al., 2012). The motifs previously found for the Yap transcription factors in *S. cerevisiae* and for CgYap1 in *C. glabrata* are indicated (Tan et al., 2008; Kuo et al., 2010a; Goudot et al., 2011). The complete Peak Motif predictions are available in Supplementary File S5.

et al., 2011; Hsu et al., 2011; Singh et al., 2011; Pimentel et al., 2012; Supplementary Table 2). The functional categories in which Yap1 and Yap5 are involved were remarkably conserved: ScYap1 and Cap1 are also involved in oxido-reduction processes (Salin et al., 2008; Znaidi et al., 2009), ScYap5 and Hap43 are involved in iron homeostasis, but ScYap5 is an activator of iron stress response while Hap43 has a role in repressing iron consuming genes in iron limiting conditions (Li et al., 2008; Hsu et al., 2011; Singh et al., 2011). No clear GO category could be attributed to ScYap7 besides its role in *YHB1* repression but all previous genome-wide studies (for instance Harbison et al., 2004) have been conducted in a strain background in which the *ScYAP7* gene is interrupted by a frame-shift mutation (Merhej et al., 2015). In terms of gene targets, the three Yap5 targets which had been validated in *S. cerevisiae* (*CCC1*, *GRX4*, and *TYW1*; Li et al., 2008, 2011; Pimentel et al., 2012) were conserved in *C. glabrata* (Figure 4A). Remarkably, despite the large evolutionary distance between *C. glabrata* and *C. albicans*, all CgYap5 targets, except *GRX4*, are targets of its orthologue Hap43 (Figure 4B). Hap43 also shares 19 targets with CgYap7, many of which are involved in iron sulfur cluster metabolism, redox homeostasis or heme metabolism (e.g., *NAR1*, *DRE2*, *LYS4*, *SDH2*, *SSU1*, *OYE2*, *YHB1*, *CCP1*, *HEM3*, ...).

Among the 82 targets that we identified for CgYap1, 28 are orthologous to targets of ScYap1 and 20 to Cap1 targets (Figures 4C,D). Of these, 15 were common to the three orthologs. This set of highly conserved Yap1 targets includes several enzymes known to play important roles in redox balance (*OYE2*, *TRX2*, *TRR1*, *CTA1*, *GPX2*, *GSH1*, *ZWF1*, *CCP1*), the MFS permease *FLR1* (named *MDR1* in *C. albicans*) and, remarkably, Yap1 itself, suggesting that the auto-regulation of Yap1 may actually play a significant role in its function. Besides this relatively high conservation of the Yap1 regulon in yeasts, a remarkable specificity of CgYap1 is its role in the direct regulation of several genes encoding enzymes of the heme biosynthetic pathway, a feature which was documented neither in *S. cerevisiae* nor in *C. albicans*.

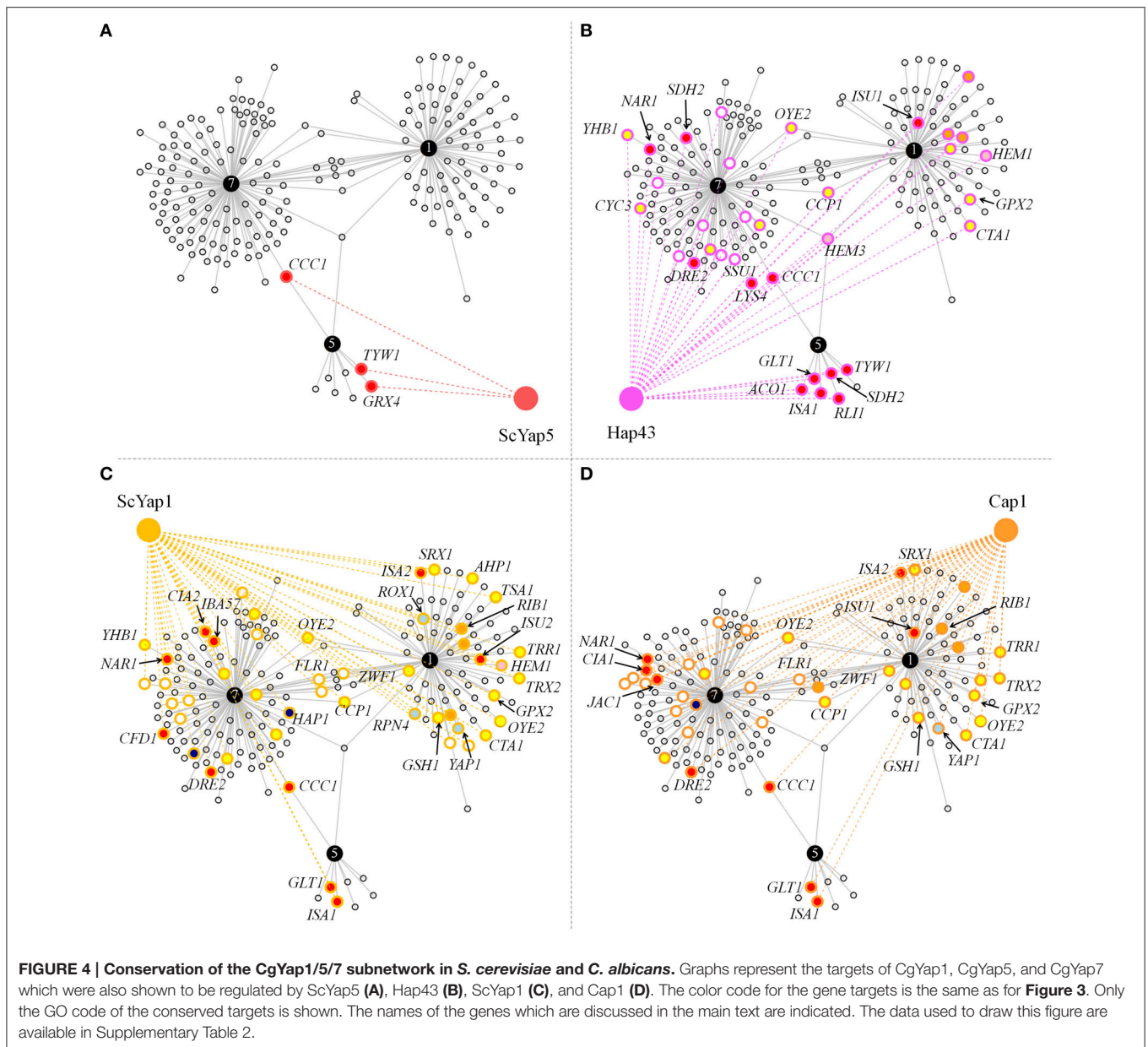
Overlap between the CgYap Network and Stress Responses in *C. glabrata*

We next used Gene Set Enrichment Analyses (GSEA; Subramanian et al., 2005) to look for enrichments of our CgYap target sets in transcriptome analyses of *C. glabrata* wild type cells to various environmental stresses. The list and origin of the transcriptome data which were used can be found in the methods. As could have been expected from the results presented



in the previous chapters, the CgYap1 targets were significantly enriched among the genes induced by oxidative stress causing agents (H₂O₂, selenite, iron, with the exception of menadione; Supplementary File S8), supporting the general role of this transcription factor in the oxidative stress response of *C. glabrata* (Lelandais et al., 2008; Roetzer et al., 2011). More surprisingly, the targets of CgYap1 were enriched in heat shock and sorbic acid stress responses. Interestingly enough, the HSE (Heat Shock Element) was found to be present in 25% of the CgYap1 ChIP peaks (Supplementary File S5). In *S. cerevisiae*, previous works have identified connections between Yap1 activity on one hand and Hsf1 and some gene regulatory modules induced by heat shock on the other (Hahn et al., 2006; Wu and Li, 2008; Nussbaum et al., 2014). Our results suggest that these connections might be conserved in *C. glabrata*. In contrast, the targets of CgYap4/6 were not particularly enriched in osmotic stress responses caused by NaCl excess (Supplementary File S8), which questions the conservation in *C. glabrata* of the role of the ScYap4 and ScYap6 in this process. As expected, the targets of CgYap5 were globally induced by iron excess in *C. glabrata* but we also observed that the CgYap5 set of targets

was significantly repressed in iron depleted conditions caused by BPS treatment or selenite exposure (Supplementary File S8, **Figure 5A**). Moreover, our transcriptome analyses of iron starvation and iron excess responses in wild type *C. glabrata* cells showed that CgYAP5 itself had expression levels which were inversely correlated to the iron concentration (**Figure 5B**), suggesting an active role for this transcription factor in iron starvation. Obviously, the fact that CgYap5 targets were repressed by iron starvation did not necessarily mean that CgYap5 was directly involved in this regulation. For instance, in *S. cerevisiae*, the repression of many iron consuming genes such as *ACO1*, *SDH2*, *ISA1*, and *CCC1* in iron starved conditions occurs post-transcriptionally and is mediated by the RNA binding proteins Cth1 and Cth2 (Puig et al., 2005). To assess the actual role of CgYap5 in the iron starvation response of *C. glabrata*, we analyzed the transcriptome response of Δ Cgyp5 cells to BPS treatment and compared this response with the one of wild type cells. We observed that most of the genes, which were dependent on Yap5 for their high iron induction, were similarly repressed by BPS in the wild type and in the Δ Cgyp5 mutant (exemplified by *SDH2* on **Figure 5C**), indicating that this



repression was not CgYap5 dependent. The only exception was *GRX4*, for which repression was totally abolished when CgYap5 was absent (Figure 5D). Consistent with these results, ChIP-seq experiments conducted in iron-replete conditions showed that the binding of CgYap5 to *GRX4* promoter was constitutive, while binding to its other targets was detected only at high iron concentrations (Figures 5E,F). Notably, the *GRX4* expression level was previously shown to be independent of CgYap5 in standard growth conditions (Merhej et al., 2015), indicating that the effect detected here was indeed specific of iron starvation. These results strongly suggest that, in addition to its role in the iron stress response, CgYap5 plays an active role in the iron starvation response by directly repressing the expression of *GRX4*.

DISCUSSION

A Methodology to Build Highly Consistent Regulatory Networks

Global ChIP and transcriptome analyses are powerful tools to achieve comprehensive descriptions of large transcriptional regulatory networks (Babu et al., 2004; Harbison et al., 2004). However, the interpretation of these networks is dampened by the tendency of these techniques to generate large numbers of false positives. For instance, highly expressed genomic regions (tRNA genes, glycolytic enzymes encoding genes, etc.) have been shown to be nonspecifically enriched in ChIP-seq experiments, leading to tens to hundreds of misidentified “targets” (Park et al., 2013; Teytelman et al., 2013). This bias is

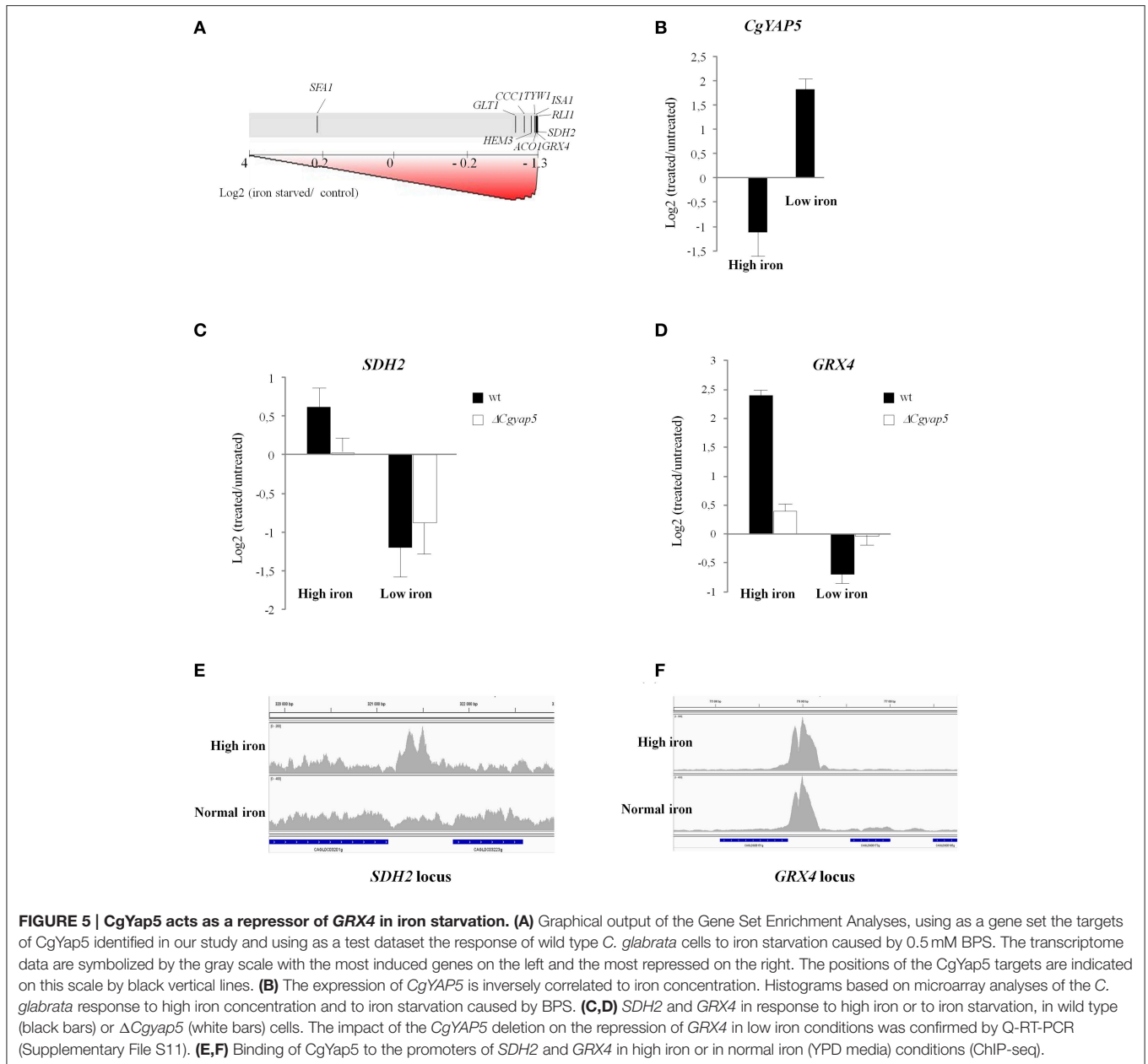


FIGURE 5 | CgYap5 acts as a repressor of GRX4 in iron starvation. (A) Graphical output of the Gene Set Enrichment Analyses, using as a gene set the targets of CgYap5 identified in our study and using as a test dataset the response of wild type *C. glabrata* cells to iron starvation caused by 0.5 mM BPS. The transcriptome data are symbolized by the gray scale with the most induced genes on the left and the most repressed on the right. The positions of the CgYap5 targets are indicated on this scale by black vertical lines. **(B)** The expression of *CgYAP5* is inversely correlated to iron concentration. Histograms based on microarray analyses of the *C. glabrata* response to high iron concentration and to iron starvation caused by BPS. **(C,D)** *SDH2* and *GRX4* in response to high iron or to iron starvation, in wild type (black bars) or $\Delta Cgyap5$ (white bars) cells. The impact of the *CgYAP5* deletion on the repression of *GRX4* in low iron conditions was confirmed by Q-RT-PCR (Supplementary File S11). **(E,F)** Binding of CgYap5 to the promoters of *SDH2* and *GRX4* in high iron or in normal iron (YPD media) conditions (ChIP-seq).

better captured, but only partially corrected, using mock IP as reference for peak calling, rather than input DNA (Park et al., 2013; Krebs et al., 2014). For transcriptome analyses, relatively minor differences between wild type and mutant growth rates or stress response dynamics can eventually produce tens of differentially expressed genes which have no real relationship with the mutation being tested (Thompson et al., 2013). We took into account these previous observations and designed experimental and bioinformatics procedures in which, 1- peak calling was performed using both the input DNA and the mock IP as references to efficiently sort out peaks corresponding to tRNA or highly expressed ORFs loci; and 2- only wild type and mutant cell cultures having very similar growth rates before and after the stress treatment were compared in transcriptome

analyses. These simple precautions led to a final network showing unusually high consistency between ChIP results and TFBS predictions on the one hand, and ChIP and transcriptome results on the other hand. Indeed, between 40 and 100% of the ChIP peaks identified contained one or several YRE (Yap Response Element). For comparison, the rate of YRE containing peaks in previous ChIP-chip studies conducted on the Yaps of *S. cerevisiae* (Tan et al., 2008; Ni et al., 2009) or on Yap1 in *C. glabrata* (Kuo et al., 2010a) ranged from 15 to 30%. Similarly, between 40 and 90% of the genes for which the transcriptome data showed an expression change had a ChIP peak in their promoter. This percentage ranged from 0 to 25% in a previous study of the *S. cerevisiae* Yap family (Tan et al., 2008).

These data allowed us to propose targets with a reasonably high level of confidence for 6 of the 7 Yaps in *C. glabrata*, to predict the preferred binding motifs of 5 of them and identify enriched functional categories for CgYap1, CgYap5, and CgYap7.

Conserved DNA Binding Properties of the Yap Transcription Factors

TFBS predictions based on the ChIP peaks pointed out a perfect conservation in the DNA binding properties of Yap1, Yap3b, Yap4/6, Yap5, and Yap7. As in *S. cerevisiae* (Tan et al., 2008), CgYap1, CgYap5, and CgYap7 were predicted to recognize preferentially the YRE-O site (TTACTAA) while CgYap3b and CgYap4/6 rather binds YRE-A (TTACGTAA). This result is consistent with the high conservation of the DNA binding domains of these proteins between *C. glabrata* and *S. cerevisiae* (Kuo et al., 2010a). Yet, this was unexpected in the case of CgYap1. Indeed, it was previously proposed, based on bioinformatic predictions from ChIP-chip data, that CgYap1 shifted its binding preferences from YRE-O to YRE-A due to a single mutation in the DNA binding domain compared to ScYap1 (Kuo et al., 2010a). This model was toned down by further analyses of the same dataset, which showed that YRE-O were as frequent as YRE-A in the promoters of CgYap1 targets (Goudot et al., 2011). Our ChIP-seq data unambiguously suggest that YRE-O is the preferred DNA binding site of CgYap1 and that the co-evolution scenario previously published has to be reconsidered. The discrepancy between these different studies may rely on the size of the genomic sequences which were used for TFBS predictions. Indeed previous ChIP-chip data provided peaks which were as wide as intergenic regions (800 base pairs in Goudot et al., 2011) while the binding regions identified by our ChIP-seq analyses for CgYap1 were 300 base pairs in average, leading to a much more precise identification of the actual binding location of the transcription factor (Supplementary File S4).

The Yap1 Core Regulon

Hence, the DNA binding properties of Yap1 were remarkably conserved between *S. cerevisiae*, *C. glabrata*, and *C. albicans*, since the YRE-O was also shown to be the preferred binding motif of Cap1 (Znaidi et al., 2009; Goudot et al., 2011). What was true for DNA binding was also true at the level of gene targets and functional annotations. Indeed, our data confirmed the role of CgYap1 in redox homeostasis, as previously demonstrated (Chen et al., 2007; Lelandais et al., 2008; Kuo et al., 2010a; Roetzer et al., 2011). We identified a core Yap1 regulon of 28 targets between *S. cerevisiae* and *C. glabrata* and 15 conserved targets between *S. cerevisiae*, *C. glabrata*, and *C. albicans*. Notably, this enlarged by more than two-fold the list of conserved Yap1 targets which were previously identified (Kuo et al., 2010a; Goudot et al., 2011). This core Yap1 regulon is composed mostly of well-known and highly conserved actors of oxidative stress response such as glutathione peroxidase and glutathione synthetase, catalase, mitochondrial peroxidase, enzymes of the thioredoxin pathway (thioredoxins, thioredoxin reductase, thioredoxin peroxidase, and thioredoxin peroxidase reductase), the FLR1/MDR1 permease, enzymes involved in NADPH metabolism (NADPH oxydoreductase of the Old Yellow

Enzyme family and glucose phosphate dehydrogenase). This confirms that, as suggested by previous work, oxidative stress response in general and the Yap1 control of this response in particular, do not fundamentally differ in *C. glabrata* compared with *S. cerevisiae* and *C. albicans* (Cuellar-Cruz et al., 2008; Lelandais et al., 2008; Kuo et al., 2010a; Gulshan et al., 2011; Roetzer et al., 2011; Briones-Martin-Del-Campo et al., 2014). Also, Yap1 binding to its own promoter, which had been demonstrated in *C. albicans* and *S. cerevisiae* (Salin et al., 2008; Znaidi et al., 2009), was conserved in *C. glabrata*. Although the primary activation of Yap1 has been shown to be at the post-translational level (Kuge et al., 1997; Zhang et al., 2000), this high conservation suggests that Yap1 autoregulation could play a role in conditions of acute oxidative stress, such as the ones used in the aforementioned studies. In support to this hypothesis, ScYAP1, CgYAP1 and CAP1 were all shown to be induced by oxidative stress at the mRNA level (Salin et al., 2008; Znaidi et al., 2009).

Insights in the Roles of Yap2 and Yap4/6 in *C. glabrata*

Only three ChIP targets could be detected for CgYap2 and CgYap3b, which did not allow us to propose functional annotations for these two factors. Still, it is interesting to notice that CgYap2 targets *YCF1*, which encodes a vacuolar transporter playing a key role in cadmium detoxification in *S. cerevisiae* (Wemmie et al., 1994; Li et al., 1997) and *TNA1*, the inactivation of which leads to cadmium sensitivity (Ruotolo et al., 2008). This suggests that the role of ScYap2 in cadmium resistance is conserved in *C. glabrata*. In *S. cerevisiae*, the activity of Yap2 is hidden by its partial functional redundancy with Yap1 (Azevedo et al., 2007; Iwai et al., 2010; Mazzola et al., 2015). Hence, it would be interesting to conduct Δ Cgyap2 transcriptome analyses in a context in which CgYAP1 has been knocked out.

We identified about 40 potential targets for CgYap4/6. The enrichment of the Sko1 binding motifs in the ChIP peaks indicated that the role of ScYap4 and ScYap6 in osmotic stress response may be conserved in *C. glabrata*. However, our GSEA and GO analyses showed that the CgYap4/6 targets were enriched neither in NaCl responsive genes nor in any particular functional category and the actual role of this factor remains to be elucidated. Our transcriptome analyses suggested that CgYap4/6 acts as a transcriptional repressor in the conditions that we tested. In *S. cerevisiae*, this point is controversial. ScYap4 and ScYap6 have been shown to recruit the general transcriptional repressor Tup1 (Hanlon et al., 2011) and bioinformatic analyses based on transcriptome data have suggested that they could be both repressors and activators (Tan et al., 2008). However, previous studies based on northern blots had shown that ScYap4 positively impacts the expression of three genes in response to osmotic shock (Nevitt et al., 2004).

CgYap5 Can Act Both as an Activator and a Repressor of Glutaredoxin Expression, Depending on Iron Availability

Similarly to CgYap1, CgYap5 has a conserved role in the iron stress response. In *S. cerevisiae*, Yap5 acts at three levels against iron excess by 1- the induction of the glutaredoxin Grx4

which senses iron-sulfur clusters in the cytoplasm (Muhlenhoff et al., 2010) and negatively controls the activity of the Aft1/2 transcriptional activators of iron uptake (Ojeda et al., 2006; Ueta et al., 2012); 2- the induction of Ccc1 which transports cytoplasmic iron into the vacuole (Li et al., 2008); and 3- the overexpression of Tyw1 which is an iron-sulfur cluster containing protein and therefore contribute to iron sequestration (Li et al., 2011). We showed here that, in iron excess conditions, CgYap5 similarly controls the expression of *GRX4*, *CCC1*, and genes encoding proteins involved in iron sequestration either through iron-sulfur cluster binding and biogenesis (*TYW1*, *GLT1*, *ACO1*, *RLI1*, *SDH2*, *ISAI*) or through heme biosynthesis (*HEM3*).

In addition to this conserved role in the detoxification of iron excess, we showed that CgYap5 is overexpressed in response to iron starvation and that it represses the expression of *GRX4* in these conditions. As mentioned above, Grx4 inhibits the iron starvation response by promoting the nuclear export of Aft1/2 when iron-sulfur clusters are abundant (reviewed in Lill et al., 2014). The regulation of glutaredoxin activity is mostly post-translational (Lill et al., 2014), but transcriptional repression of *GRX4* by CgYap5 may provide a supplementary layer of regulation to ensure full Aft1/2 activity in iron starvation conditions (Figure 6). This new role of Yap5 may be conserved in *S. cerevisiae*, since ScYap5 has been shown to bind the *GRX4* promoter independently of iron concentration and since the deletion of ScYAP5 negatively impacts the nuclear localization of Aft1/2 in iron limiting conditions (Pimentel et al., 2012). However, the inactivation of ScYAP5 does not seem to impact *GRX4* expression in iron-replete cells (Pimentel et al., 2012).

This dual role of CgYap5 is reminiscent of its HapX orthologs. HapX was initially identified as a key regulator of iron starvation response in filamentous ascomycetes (Hortschansky et al., 2007; Schrettl et al., 2010; Lopez-Berges et al., 2012), basidiomycetes (Jung et al., 2010), and hemiascomycetes of the *C. albicans* clade (where it is called Hap43; Hsu et al., 2011; Singh et al., 2011). HapX acts by repressing iron consuming genes when iron is limiting and HapX proteins are more expressed in iron starvation than in iron excess growth conditions (Singh et al.,

2011; Gsaller et al., 2014). However, HapX was shown recently to play an additional role in activating the expression of some of its targets in response to iron excess in *Aspergillus* and *Fusarium sp.* (Gsaller et al., 2014). Interestingly, Yap5 and HapX both sense iron by directly binding iron-sulfur clusters through a conserved cysteine rich domain (CRD; Gsaller et al., 2014; Rietzschel et al., 2015). Our data indicate that Yap5 and HapX may have more in common than just this CRD. As indicated above, CgYap5 is also overexpressed and able to repress transcription in iron starved cells (Figure 6). Moreover, all the CgYap5 targets that we identified, except *GRX4*, are targets of Hap43. This conservation of targets is remarkable considering that Hap43 and Yap5 have different DNA binding properties: Hap43, like HapX in other fungi, mainly binds CCAAT boxes through its Hap4L like domain (Hortschansky et al., 2007, 2015; Chen et al., 2011), while Yap5 binds YRE-O using exclusively its bZIP region (this work, Li et al., 2008; Pimentel et al., 2012). Still, the role of CgYap5 in iron starvation is limited to a modest repression of *GRX4* expression, while HapX strongly represses the expression of tens of iron consuming genes in these conditions (Supplementary File S9).

This new role of Yap5 opens the question of the molecular mechanisms which would allow CgYap5 to be a transcriptional repressor when iron is limiting. Several hypotheses can be mentioned, based on the literature. The transcriptional repression by Hap43 requires its Hap4L domain and involves the CCAAT binding complex (Singh et al., 2011). Yap5 only has a vestigial Hap4L domain (Merhej et al., 2015), which was supposed to be non-functional although its activity has actually never been tested so far. Second, Yap7, the ortholog of Yap5, has been shown to repress transcription by recruiting the general repressor Tup1 (Merhej et al., 2015). Moreover, functional connections between Hap43 and Tup1 have been reported in *C. albicans* (Hsu et al., 2011). Additionally, the shift of Yap5 from an activator to a repressor could be controlled by iron availability, since the binding of iron-sulfur clusters to ScYap5 was shown to significantly change its conformation (Rietzschel et al., 2015).

Interconnections between CgYap1, CgYap5, CgYap7, and the *C. glabrata* Stress Response Network

Our data led us to propose new roles for Yap1 and Yap7 in the regulation of genes involved, respectively, in heme biosynthesis and in the biogenesis of iron-sulfur cluster proteins. Previously, Yap7 was shown to be a repressor of the heme-containing nitric oxide oxidase Yhb1 in *C. glabrata* and *S. cerevisiae* (Merhej et al., 2015). Our ChIP-seq data indicated that CgYap7 binds many genes encoding enzymes of the cytoplasmic and mitochondrial iron sulfur cluster biogenesis pathway. This result is particularly interesting considering the important role played by the mitochondrial iron sulfur clusters in the sensing of iron availability and in regulating the activity of Yap5, the ortholog of Yap7 (Lill et al., 2014; Rietzschel et al., 2015).

In *S. cerevisiae*, there is little evidence of a connection between Yap1 and heme biosynthesis, besides the binding of ScYap1 to *HEM1* (Salin et al., 2008) and the regulation by

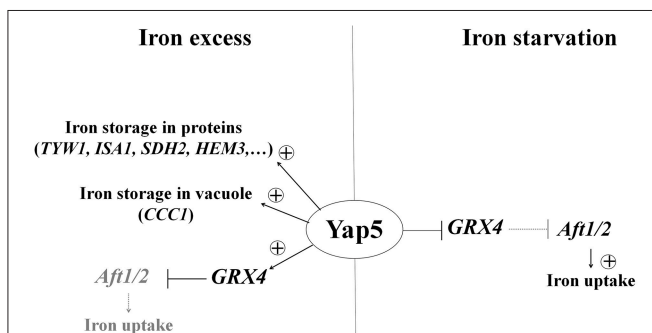
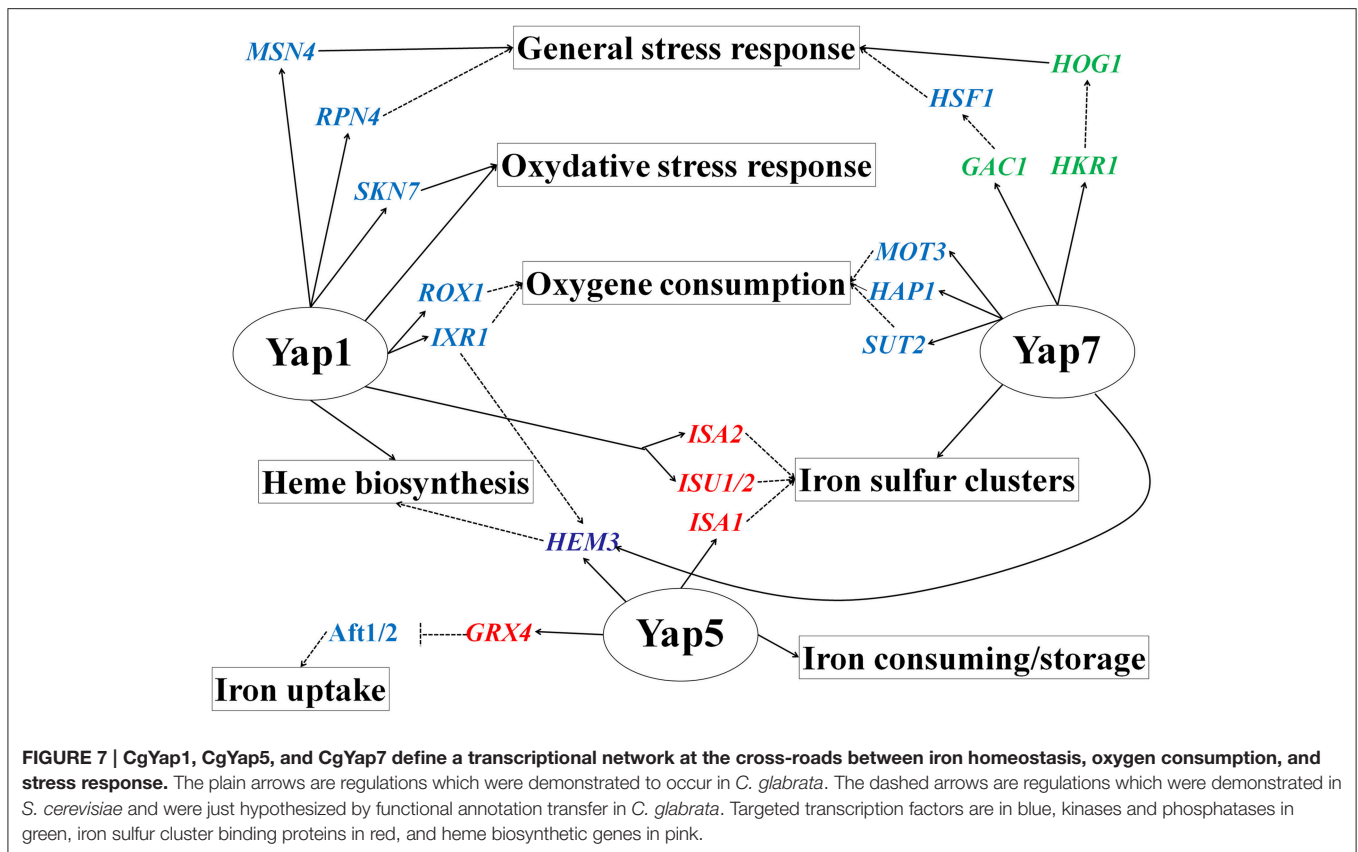


FIGURE 6 | A dual role for CgYap5 in iron excess and iron starvation. In iron excess (Left Panel), CgYap5 induces an iron stress response which is very similar to what was described in *S. cerevisiae*. In iron starvation (Right Panel), CgYap5 represses *GRX4* expression and may indirectly contribute to the induction of iron uptake genes by the Aft1/2 transcription factors.



ScYap1 of the heme-dependent repressors *IXR1* and *ROX1* (Castro-Prego et al., 2010; Caetano et al., 2015). We show here that CgYap1 directly targets four of the eight enzymes involved in this pathway (*HEM1*, *HEM2*, *HEM3*, *HEM15*) and CgYAP1 deletion clearly impacted the expression of three of them (*HEM1*, *HEM3*, and *HEM15*). Hence, we defined here a CgYap1/CgYap5/CgYap7 network deeply involved in oxygen and iron sensing by tuning redox homeostasis, heme biosynthesis, iron storage, and iron sulfur cluster metabolism (Figure 7). Each factor has its own specificity (CgYap1 in redox balance and heme biosynthesis, CgYap5 in iron storage and CgYap7 in iron-sulfur clusters biogenesis) but there are some interconnections between them. For instance, CgYap5 and CgYap1 positively control the mitochondrial iron-sulfur cluster proteins maturation factors *Isa1* and *Isu1/2/Isa2*, respectively. Also, the *HEM3* gene, encoding the prophobilinogen deaminase (third step of heme biosynthesis) came out as a “hub” in this network, being positively regulated by CgYap1 and CgYap5 in response to oxidative stress or iron excess, respectively, and repressed by CgYap7 in standard growth conditions (Figure 3). Similarly, *CCP1*, encoding the cytochrome c peroxidase which acts as a heme-based sensor of the mitochondrial oxidative stress (Martins et al., 2013), is positively regulated by CgYap1 and repressed by CgYap7 (Figure 3).

Another interesting feature of CgYap1 and CgYap7 is the strong enrichment for transcription factors among their targets. For instance, they potentially control the expression of several

transcription factors known in *S. cerevisiae* for being involved in the regulation of hypoxic genes and oxygen consumption (*Rox1* and *Ixr1* for CgYap1 and *Mot3*, *Hap1* and *Sut2* for CgYap7; Figure 7; Castro-Prego et al., 2010; Gonzalez Siso et al., 2012). They also target some regulators involved in the general Environmental Stress response. This is for instance the case of the *Hog1*, *Hkr1*, and *Gac1* stress signaling proteins for CgYap7. Additionally, CgYap1 seems to positively control the expression of *SKN7*, which encodes a regulator of oxidative stress response known to cooperate with Yap1 both in *S. cerevisiae* and in *C. glabrata* (Morgan et al., 1997; Lee et al., 1999; Saijo et al., 2010; Roetzer et al., 2011). This suggests that the network described here is tightly connected to other transcriptional responses and that CgYap1 and CgYap7 are master regulators in the *C. glabrata* hierarchy of transcription factors (Jothi et al., 2009; Bhardwaj et al., 2010).

AUTHOR CONTRIBUTIONS

JM constructed the strains and performed the ChIP-seq and transcriptome analyses. AT performed transcriptome analyses. CB, JP, MA, and SL performed the high-throughput sequencing. GL and JC performed the bioinformatics analyses and the network building. FD designed the experiments and wrote the manuscript.

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REFERENCES

- Adeboye, P. T., Bettiga, M., and Olsson, L. (2014). The chemical nature of phenolic compounds determines their toxicity and induces distinct physiological responses in *Saccharomyces cerevisiae* in lignocellulose hydrolysates. *AMB Express* 4, 46. doi: 10.1186/s13568-014-0046-7
- Amaral, C., Pimentel, C., Matos, R. G., Arraiano, C. M., Matzapetakis, M., and Rodrigues-Pousada, C. (2013). Two residues in the basic region of the yeast transcription factor Yap8 are crucial for its DNA-binding specificity. *PLoS ONE* 8:e83328. doi: 10.1371/journal.pone.0083328
- Azevedo, D., Nascimento, L., Labarre, J., Toledano, M. B., and Rodrigues-Pousada, C. (2007). The *S. cerevisiae* Yap1 and Yap2 transcription factors share a common cadmium-sensing domain. *FEBS Lett.* 581, 187–195. doi: 10.1016/j.febslet.2006.11.083
- Babu, M. M., Luscombe, N. M., Aravind, L., Gerstein, M., and Teichmann, S. A. (2004). Structure and evolution of transcriptional regulatory networks. *Curr. Opin. Struct. Biol.* 14, 283–291. doi: 10.1016/j.sbi.2004.05.004
- Bhardwaj, N., Kim, P. M., and Gerstein, M. B. (2010). Rewiring of transcriptional regulatory networks: hierarchy, rather than connectivity, better reflects the importance of regulators. *Sci. Signal.* 3, ra79. doi: 10.1126/scisignal.2001014
- Bobrowicz, P., Wysocki, R., Owsianik, G., Goffeau, A., and Ulaszewski, S. (1997). Isolation of three contiguous genes, ACR1, ACR2 and ACR3, involved in resistance to arsenic compounds in the yeast *Saccharomyces cerevisiae*. *Yeast* 13, 819–828.
- Briones-Martin-Del-Campo, M., Orta-Zavalza, E., Juarez-Cepeda, J., Gutierrez-Escobedo, G., Canas-Villamar, I., Castano, I., et al. (2014). The oxidative stress response of the opportunistic fungal pathogen *Candida glabrata*. *Rev. Iberoam. Micol.* 31, 67–71. doi: 10.1016/j.riam.2013.09.012
- Brunke, S., and Hube, B. (2013). Two unlike cousins: *Candida albicans* and *C. glabrata* infection strategies. *Cell. Microbiol.* 15, 701–708. doi: 10.1111/cmi.12091
- Caetano, S. M., Menezes, R., Amaral, C., Rodrigues-Pousada, C., and Pimentel, C. (2015). Repression of the low affinity iron transporter gene FET4: a novel mechanism against cadmium toxicity orchestrated by YAP1 via ROX1. *J. Biol. Chem.* 290, 18584–18595. doi: 10.1074/jbc.M114.600742
- Castro-Prego, R., Lamas-Maceiras, M., Soengas, P., Carneiro, I., Gonzalez-Siso, I., and Cerdan, M. E. (2010). Regulatory factors controlling transcription of *Saccharomyces cerevisiae* IXR1 by oxygen levels: a model of transcriptional adaptation from aerobiosis to hypoxia implicating ROX1 and IXR1 cross-regulation. *Biochem. J.* 425, 235–243. doi: 10.1042/BJ20091500
- Caudle, K. E., Barker, K. S., Wiederhold, N. P., Xu, L., Homayouni, R., and Rogers, P. D. (2011). Genomewide expression profile analysis of the *Candida glabrata* Pdr1 regulon. *Eukaryot. Cell* 10, 373–383. doi: 10.1128/EC.00073-10
- Chen, C., Pande, K., French, S. D., Tuch, B. B., and Noble, S. M. (2011). An iron homeostasis regulatory circuit with reciprocal roles in *Candida albicans* commensalism and pathogenesis. *Cell Host Microbe* 10, 118–135. doi: 10.1016/j.chom.2011.07.005
- Chen, K. H., Miyazaki, T., Tsai, H. F., and Bennett, J. E. (2007). The bZip transcription factor Cgap1p is involved in multidrug resistance and required for activation of multidrug transporter gene CgFLR1 in *Candida glabrata*. *Gene* 386, 63–72. doi: 10.1016/j.gene.2006.08.010

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2016.00645>

- Csardi, G., and Nepusz, T. (2006). The igraph software package for complex network research. *InterJournal Complex Syst.* 1695. Available online at: www.interjournal.org
- Cuellar-Cruz, M., Briones-Martin-del-Campo, M., Canas-Villamar, I., Montalvo-Arredondo, J., Riego-Ruiz, L., Castano, I., et al. (2008). High resistance to oxidative stress in the fungal pathogen *Candida glabrata* is mediated by a single catalase, Cta1p, and is controlled by the transcription factors Yap1p, Skn7p, Msn2p, and Msn4p. *Eukaryot. Cell* 7, 814–825. doi: 10.1128/EC.00011-08
- Domergue, R., Castano, I., De Las Penas, A., Zupancic, M., Lockatell, V., Hebel, J. R., et al. (2005). Nicotinic acid limitation regulates silencing of *Candida adhesins* during UTI. *Science* 308, 866–870. doi: 10.1126/science.1108640
- Ferrari, S., Sanguinetti, M., Torelli, R., Posteraro, B., and Sanglard, D. (2011). Contribution of CgPDR1-regulated genes in enhanced virulence of azole-resistant *Candida glabrata*. *PLoS ONE* 6:e17589. doi: 10.1371/journal.pone.0017589
- Fox, E. P., Bui, C. K., Nett, J. E., Hartooni, N., Mui, M. C., Andes, D. R., et al. (2015). An expanded regulatory network temporally controls *Candida albicans* biofilm formation. *Mol. Microbiol.* 96, 1226–1239. doi: 10.1111/mmi.13002
- Fujii, Y., Shimizu, T., Toda, T., Yanagida, M., and Hakoshima, T. (2000). Structural basis for the diversity of DNA recognition by bZIP transcription factors. *Nat. Struct. Biol.* 7, 889–893. doi: 10.1038/82822
- Gabaldon, T., and Carrete, L. (2015). The birth of a deadly yeast: tracing the evolutionary emergence of virulence traits in *Candida glabrata*. *FEMS Yeast Res.* 16:fov110. doi: 10.1093/femsyr/fov110
- Gonzalez Siso, M. I., Becerra, M., Lamas Maceiras, M., Vizoso Vazquez, A., and Cerdan, M. E. (2012). The yeast hypoxic responses, resources for new biotechnological opportunities. *Biotechnol. Lett.* 34, 2161–2173. doi: 10.1007/s10529-012-1039-8
- Gordan, R., Murphy, K. F., McCord, R. P., Zhu, C., Vedenko, A., and Bulyk, M. L. (2011). Curated collection of yeast transcription factor DNA binding specificity data reveals novel structural and gene regulatory insights. *Genome Biol.* 12:R125. doi: 10.1186/gb-2011-12-12-r125
- Goudot, C., Etchebest, C., Devaux, F., and Lelandais, G. (2011). The reconstruction of condition-specific transcriptional modules provides new insights in the evolution of yeast AP-1 proteins. *PLoS ONE* 6:e20924. doi: 10.1371/journal.pone.0020924
- Gsaller, F., Hortschansky, P., Beattie, S. R., Klammer, V., Tuppatsch, K., Lechner, B. E., et al. (2014). The Janus transcription factor HapX controls fungal adaptation to both iron starvation and iron excess. *EMBO J.* 33, 2261–2276. doi: 10.15252/embj.201489468
- Gulshan, K., Lee, S. S., and Moye-Rowley, W. S. (2011). Differential oxidant tolerance determined by the key transcription factor Yap1 is controlled by levels of the Yap1-binding protein, Ybp1. *J. Biol. Chem.* 286, 34071–34081. doi: 10.1074/jbc.M111.251298
- Hahn, J. S., Neef, D. W., and Thiele, D. J. (2006). A stress regulatory network for co-ordinated activation of proteasome expression mediated by yeast heat shock transcription factor. *Mol. Microbiol.* 60, 240–251. doi: 10.1111/j.1365-2958.2006.05097.x
- Hanlon, S. E., Rizzo, J. M., Tatomer, D. C., Lieb, J. D., and Buck, M. J. (2011). The stress response factors Yap6, Cin5, Phd1, and Skn7 direct targeting of the conserved co-repressor Tup1-Ssn6 in *S. cerevisiae*. *PLoS ONE* 6:e19060. doi: 10.1371/journal.pone.0019060

- Harbison, C. T., Gordon, D. B., Lee, T. I., Rinaldi, N. J., MacIsaac, K. D., Danford, T. W., et al. (2004). Transcriptional regulatory code of a eukaryotic genome. *Nature* 431, 99–104. doi: 10.1038/nature02800
- Haugen, A. C., Kelley, R., Collins, J. B., Tucker, C. J., Deng, C., Afshari, C. A., et al. (2004). Integrating phenotypic and expression profiles to map arsenic-response networks. *Genome Biol.* 5:R95. doi: 10.1186/gb-2004-5-12-r95
- Hirata, D., Yano, K., and Miyakawa, T. (1994). Stress-induced transcriptional activation mediated by YAP1 and YAP2 genes that encode the Jun family of transcriptional activators in *Saccharomyces cerevisiae*. *Mol. Gen. Genet.* 242, 250–256. doi: 10.1007/BF00280413
- Hortschansky, P., Ando, E., Tuppatsch, K., Arikawa, H., Kobayashi, T., Kato, M., et al. (2015). Deciphering the combinatorial DNA-binding code of the CCAAT-binding complex and the iron-regulatory basic region leucine zipper (bZIP) transcription factor HapX. *J. Biol. Chem.* 290, 6058–6070. doi: 10.1074/jbc.M114.628677
- Hortschansky, P., Eisendle, M., Al-Abdallah, Q., Schmidt, A. D., Bergmann, S., Thon, M., et al. (2007). Interaction of HapX with the CCAAT-binding complex—a novel mechanism of gene regulation by iron. *EMBO J.* 26, 3157–3168. doi: 10.1038/sj.emboj.7601752
- Hsu, P. C., Yang, C. Y., and Lan, C. Y. (2011). *Candida albicans* Hap43 is a repressor induced under low-iron conditions and is essential for iron-responsive transcriptional regulation and virulence. *Eukaryot. Cell* 10, 207–225. doi: 10.1128/EC.00158-10
- Inglis, D. O., Arnaud, M. B., Binkley, J., Shah, P., Skrzypek, M. S., Wymore, F., et al. (2012). The *Candida* genome database incorporates multiple *Candida* species: multispecies search and analysis tools with curated gene and protein information for *Candida albicans* and *Candida glabrata*. *Nucleic Acids Res.* 40, D667–D674. doi: 10.1093/nar/gkr945
- Iwai, K., Naganuma, A., and Kuge, S. (2010). Peroxiredoxin Ahp1 acts as a receptor for alkylhydroperoxides to induce disulfide bond formation in the Cad1 transcription factor. *J. Biol. Chem.* 285, 10597–10604. doi: 10.1074/jbc.M109.090142
- Jandric, Z., Gregori, C., Klopff, E., Radolf, M., and Schuller, C. (2013). Sorbic acid stress activates the *Candida glabrata* high osmolarity glycerol MAP kinase pathway. *Front. Microbiol.* 4:350. doi: 10.3389/fmicb.2013.00350
- Jothi, R., Balaji, S., Wuster, A., Grochow, J. A., Gsponer, J., Przytycka, T. M., et al. (2009). Genomic analysis reveals a tight link between transcription factor dynamics and regulatory network architecture. *Mol. Syst. Biol.* 5:294. doi: 10.1038/msb.2009.52
- Jung, W. H., Saikia, S., Hu, G., Wang, J., Fung, C. K., D'Souza, C., et al. (2010). HapX positively and negatively regulates the transcriptional response to iron deprivation in *Cryptococcus neoformans*. *PLoS Pathog.* 6:e1001209. doi: 10.1371/journal.ppat.1001209
- Kitada, K., Yamaguchi, E., and Arisawa, M. (1995). Cloning of the *Candida glabrata* TRP1 and HIS3 genes, and construction of their disruptant strains by sequential integrative transformation. *Gene* 165, 203–206. doi: 10.1016/0378-1119(95)00552-H
- Krebs, W., Schmidt, S. V., Goren, A., De Nardo, D., Labzin, L., Bovier, A., et al. (2014). Optimization of transcription factor binding map accuracy utilizing knockout-mouse models. *Nucleic Acids Res.* 42, 13051–13060. doi: 10.1093/nar/gku1078
- Kuge, S., Jones, N., and Nomoto, A. (1997). Regulation of yAP-1 nuclear localization in response to oxidative stress. *EMBO J.* 16, 1710–1720. doi: 10.1093/emboj/16.7.1710
- Kumar, N. V., Yang, J., Pillai, J. K., Rawat, S., Solano, C., Kumar, A., et al. (2015). Arsenic directly binds to and activates the yeast AP-1-like transcription factor Yap8. *Mol. Cell Biol.* 36, 913–922. doi: 10.1128/MCB.00842-15
- Kuo, D., Licon, K., Bandyopadhyay, S., Chuang, R., Luo, C., Catalana, J., et al. (2010a). Coevolution within a transcriptional network by compensatory trans and cis mutations. *Genome Res.* 20, 1672–1678. doi: 10.1101/gr.111765.110
- Kuo, D., Tan, K., Zinman, G., Ravasi, T., Bar-Joseph, Z., and Ideker, T. (2010b). Evolutionary divergence in the fungal response to fluconazole revealed by soft clustering. *Genome Biol.* 11:R77. doi: 10.1186/gb-2010-11-7-r77
- Lavoie, H., Hogues, H., Mallick, J., Sellam, A., Nantel, A., and Whiteway, M. (2010). Evolutionary tinkering with conserved components of a transcriptional regulatory network. *PLoS Biol.* 8:e1000329. doi: 10.1371/journal.pbio.100329
- Lee, J., Godon, C., Lagniel, G., Spector, D., Garin, J., Labarre, J., et al. (1999). Yap1 and Skn7 control two specialized oxidative stress response regulons in yeast. *J. Biol. Chem.* 274, 16040–16046. doi: 10.1074/jbc.274.23.16040
- Lelandais, G., Blugeon, C., and Merhej, J. (2016). ChIPseq in yeast species: from chromatin immunoprecipitation to high-throughput sequencing and bioinformatics data analyses. *Methods Mol. Biol.* 1361, 185–202. doi: 10.1007/978-1-4939-3079-1_11
- Lelandais, G., Tanty, V., Geneix, C., Etchebest, C., Jacq, C., and Devaux, F. (2008). Genome adaptation to chemical stress: clues from comparative transcriptomics in *Saccharomyces cerevisiae* and *Candida glabrata*. *Genome Biol.* 9:R164. doi: 10.1186/gb-2008-9-11-r164
- Lemoine, S., Combes, F., Servant, N., and Le Crom, S. (2006). Goulphar: rapid access and expertise for standard two-color microarray normalization methods. *BMC Bioinformatics* 7:467. doi: 10.1186/1471-2105-7-467
- Li, L., Bagley, D., Ward, D. M., and Kaplan, J. (2008). Yap5 is an iron-responsive transcriptional activator that regulates vacuolar iron storage in yeast. *Mol. Cell Biol.* 28, 1326–1337. doi: 10.1128/MCB.01219-07
- Li, L., Jia, X., Ward, D. M., and Kaplan, J. (2011). Yap5 protein-regulated transcription of the TYW1 gene protects yeast from high iron toxicity. *J. Biol. Chem.* 286, 38488–38497. doi: 10.1074/jbc.M111.286666
- Li, Z. S., Lu, Y. P., Zhen, R. G., Szczycka, M., Thiele, D. J., and Rea, P. A. (1997). A new pathway for vacuolar cadmium sequestration in *Saccharomyces cerevisiae*: YCF1-catalyzed transport of bis(glutathionato)cadmium. *Proc. Natl. Acad. Sci. U.S.A.* 94, 42–47. doi: 10.1073/pnas.94.1.42
- Lill, R., Srinivasan, V., and Muhlenhoff, U. (2014). The role of mitochondria in cytosolic-nuclear iron-sulfur protein biogenesis and in cellular iron regulation. *Curr. Opin. Microbiol.* 22, 111–119. doi: 10.1016/j.mib.2014.09.015
- Longtine, M. S., McKenzie, A. III., Demarini, D. J., Shah, N. G., Wach, A., Brachat, A., et al. (1998). Additional modules for versatile and economical PCR-based gene deletion and modification in *Saccharomyces cerevisiae*. *Yeast* 14, 953–961.
- Lopez-Berges, M. S., Capilla, J., Turra, D., Schafferer, L., Matthijs, S., Jochl, C., et al. (2012). HapX-mediated iron homeostasis is essential for rhizosphere competence and virulence of the soilborne pathogen *Fusarium oxysporum*. *Plant Cell* 24, 3805–3822. doi: 10.1105/tpc.112.098624
- Martins, D., Kathiresan, M., and English, A. M. (2013). Cytochrome c peroxidase is a mitochondrial heme-based H₂O₂ sensor that modulates antioxidant defense. *Free Radic. Biol. Med.* 65, 541–551. doi: 10.1016/j.freeradbiomed.2013.06.037
- Mazzola, D., Pimentel, C., Caetano, S., Amaral, C., Menezes, R., Santos, C. N., et al. (2015). Inhibition of Yap2 activity by MAPKAP kinase Rck1 affects yeast tolerance to cadmium. *FEBS Lett.* 589, 2841–2849. doi: 10.1016/j.febslet.2015.07.049
- Mendizabal, I., Rios, G., Mulet, J. M., Serrano, R., and de Larrinoa, I. F. (1998). Yeast putative transcription factors involved in salt tolerance. *FEBS Lett.* 425, 323–328. doi: 10.1016/S0014-5793(98)00249-X
- Merhej, J., Delaveau, T., Guitard, J., Palancade, B., Hennequin, C., Garcia, M., et al. (2015). Yap7 is a transcriptional repressor of nitric oxide oxidase in yeasts, which arose from neofunctionalization after whole genome duplication. *Mol. Microbiol.* 96, 951–972. doi: 10.1111/mmi.12983
- Merhej, J., Frigo, A., Le Crom, S., Camadro, J. M., Devaux, F., and Lelandais, G. (2014). bPeaks: a bioinformatics tool to detect transcription factor binding sites from ChIPseq data in yeasts and other organisms with small genomes. *Yeast* 31, 375–391. doi: 10.1002/yea.3031
- Miyazaki, T., Nakayama, H., Nagayoshi, Y., Kakeya, H., and Kohno, S. (2013). Dissection of Ire1 functions reveals stress response mechanisms uniquely evolved in *Candida glabrata*. *PLoS Pathog.* 9:e1003160. doi: 10.1371/journal.ppat.1003160
- Morgan, B. A., Banks, G. R., Toone, W. M., Raitt, D., Kuge, S., and Johnston, L. H. (1997). The Skn7 response regulator controls gene expression in the oxidative stress response of the budding yeast *Saccharomyces cerevisiae*. *EMBO J.* 16, 1035–1044. doi: 10.1093/emboj/16.5.1035
- Muhlenhoff, U., Molik, S., Godoy, J. R., Uzarska, M. A., Richter, N., Seubert, A., et al. (2010). Cytosolic monothiol glutaredoxins function in intracellular iron sensing and trafficking via their bound iron-sulfur cluster. *Cell Metab.* 12, 373–385. doi: 10.1016/j.cmet.2010.08.001
- Nevitt, T., Pereira, J., Azevedo, D., Guerreiro, P., and Rodrigues-Pousada, C. (2004). Expression of YAP4 in *Saccharomyces cerevisiae* under osmotic stress. *Biochem. J.* 379, 367–374. doi: 10.1042/bj20031127

- Nevitt, T., and Thiele, D. J. (2011). Host iron withholding demands siderophore utilization for *Candida glabrata* to survive macrophage killing. *PLoS Pathog.* 7:e1001322. doi: 10.1371/journal.ppat.1001322
- Ni, L., Bruce, C., Hart, C., Leigh-Bell, J., Gelperin, D., Umansky, L., et al. (2009). Dynamic and complex transcription factor binding during an inducible response in yeast. *Genes Dev.* 23, 1351–1363. doi: 10.1101/gad.1781909
- Nobile, C. J., Fox, E. P., Nett, J. E., Sorrells, T. R., Mitrovich, Q. M., Hernday, A. D., et al. (2012). A recently evolved transcriptional network controls biofilm development in *Candida albicans*. *Cell* 148, 126–138. doi: 10.1016/j.cell.2011.10.048
- Noble, J. A., Tsai, H. F., Suffis, S. D., Su, Q., Myers, T. G., and Bennett, J. E. (2013). STB5 is a negative regulator of azole resistance in *Candida glabrata*. *Antimicrob. Agents Chemother.* 57, 959–967. doi: 10.1128/AAC.01278-12
- North, M., Tandon, V. J., Thomas, R., Loguinov, A., Gerlovina, I., Hubbard, A. E., et al. (2011). Genome-wide functional profiling reveals genes required for tolerance to benzene metabolites in yeast. *PLoS ONE* 6:e24205. doi: 10.1371/journal.pone.0024205
- Nussbaum, I., Weindling, E., Jubran, R., Cohen, A., and Bar-Nun, S. (2014). Deteriorated stress response in stationary-phase yeast: Sir2 and Yap1 are essential for Hsf1 activation by heat shock and oxidative stress, respectively. *PLoS ONE* 9:e111505. doi: 10.1371/journal.pone.0111505
- Ojeda, L., Keller, G., Muhlenhoff, U., Rutherford, J. C., Lill, R., and Winge, D. R. (2006). Role of glutaredoxin-3 and glutaredoxin-4 in the iron regulation of the Aft1 transcriptional activator in *Saccharomyces cerevisiae*. *J. Biol. Chem.* 281, 17661–17669. doi: 10.1074/jbc.M602165200
- Park, D., Lee, Y., Bhupindersingh, G., and Iyer, V. R. (2013). Widespread misinterpretable ChIP-seq bias in yeast. *PLoS ONE* 8:e83506. doi: 10.1371/journal.pone.0083506
- Pascual-Ahuir, A., Serrano, R., and Proft, M. (2001). The Sko1p repressor and Gcn4p activator antagonistically modulate stress-regulated transcription in *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 21, 16–25. doi: 10.1128/MCB.21.1.16-25.2001
- Paul, S., Bair, T. B., and Moye-Rowley, W. S. (2014). Identification of genomic binding sites for *Candida glabrata* Pdr1 transcription factor in wild-type and rho0 cells. *Antimicrob. Agents Chemother.* 58, 6904–6912. doi: 10.1128/AAC.03921-14
- Perlroth, J., Choi, B., and Spellberg, B. (2007). Nosocomial fungal infections: epidemiology, diagnosis, and treatment. *Med. Mycol.* 45, 321–346. doi: 10.1080/13693780701218689
- Pimentel, C., Vicente, C., Menezes, R. A., Caetano, S., Carreto, L., and Rodrigues-Pousada, C. (2012). The role of the Yap5 transcription factor in remodeling gene expression in response to Fe bioavailability. *PLoS ONE* 7:e37434. doi: 10.1371/journal.pone.0037434
- Puig, S., Askeland, E., and Thiele, D. J. (2005). Coordinated remodeling of cellular metabolism during iron deficiency through targeted mRNA degradation. *Cell* 120, 99–110. doi: 10.1016/j.cell.2004.11.032
- Quinlan, A. R., and Hall, I. M. (2010). BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* 26, 841–842. doi: 10.1093/bioinformatics/btq033
- Reinke, A. W., Baek, J., Ashenberg, O., and Keating, A. E. (2013). Networks of bZIP protein-protein interactions diversified over a billion years of evolution. *Science* 340, 730–734. doi: 10.1126/science.1233465
- Rietzschel, N., Pierik, A. J., Bill, E., Lill, R., and Muhlenhoff, U. (2015). The basic leucine zipper stress response regulator Yap5 senses high-iron conditions by coordination of [2Fe-2S] clusters. *Mol. Cell. Biol.* 35, 370–378. doi: 10.1128/MCB.01033-14
- Ritchie, M. E., Phipson, B., Wu, D., Hu, Y., Law, C. W., Shi, W., et al. (2015). limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res.* 43, e47. doi: 10.1093/nar/gkv007
- Rodrigues-Pousada, C., Menezes, R. A., and Pimentel, C. (2010). The Yap family and its role in stress response. *Yeast* 27, 245–258. doi: 10.1002/yea.1752
- Roetzer, A., Gregori, C., Jennings, A. M., Quintin, J., Ferrandon, D., Butler, G., et al. (2008). *Candida glabrata* environmental stress response involves *Saccharomyces cerevisiae* Msn2/4 orthologous transcription factors. *Mol. Microbiol.* 69, 603–620. doi: 10.1111/j.1365-2958.2008.06301.x
- Roetzer, A., Klopff, E., Gratz, N., Marcet-Houben, M., Hiller, E., Rupp, S., et al. (2011). Regulation of *Candida glabrata* oxidative stress resistance is adapted to host environment. *FEBS Lett.* 585, 319–327. doi: 10.1016/j.febslet.2010.12.006
- Ruotolo, R., Marchini, G., and Ottonello, S. (2008). Membrane transporters and protein traffic networks differentially affecting metal tolerance: a genomic phenotyping study in yeast. *Genome Biol.* 9:R67. doi: 10.1186/gb-2008-9-4-r67
- Saijo, T., Miyazaki, T., Izumikawa, K., Mihara, T., Takazono, T., Kosai, K., et al. (2010). Skn7p is involved in oxidative stress response and virulence of *Candida glabrata*. *Mycopathologia* 169, 81–90. doi: 10.1007/s11046-009-9233-5
- Salin, H., Fardeau, V., Piccini, E., Lelandaïs, G., Tanty, V., Lemoine, S., et al. (2008). Structure and properties of transcriptional networks driving selenite stress response in yeasts. *BMC Genomics* 9:333. doi: 10.1186/1471-2164-9-333
- Schrettl, M., Beckmann, N., Varga, J., Heinekamp, T., Jacobsen, I. D., Jochl, C., et al. (2010). HapX-mediated adaptation to iron starvation is crucial for virulence of *Aspergillus fumigatus*. *PLoS Pathog.* 6:e1001124. doi: 10.1371/journal.ppat.1001124
- Seider, K., Brunke, S., Schild, L., Jablonowski, N., Wilson, D., Majer, O., et al. (2011). The facultative intracellular pathogen *Candida glabrata* subverts macrophage cytokine production and phagolysosome maturation. *J. Immunol.* 187, 3072–3086. doi: 10.4049/jimmunol.1003730
- Seider, K., Gerwien, F., Kasper, L., Allert, S., Brunke, S., Jablonowski, N., et al. (2014). Immune evasion, stress resistance, and efficient nutrient acquisition are crucial for intracellular survival of *Candida glabrata* within macrophages. *Eukaryot. Cell* 13, 170–183. doi: 10.1128/EC.00262-13
- Singh, R. P., Prasad, H. K., Sinha, I., Agarwal, N., and Natarajan, K. (2011). Cap2-HAP complex is a critical transcriptional regulator that has dual but contrasting roles in regulation of iron homeostasis in *Candida albicans*. *J. Biol. Chem.* 286, 25154–25170. doi: 10.1074/jbc.M111.233569
- Stavrum, A. K., Petersen, K., Jonassen, I., and Dysvik, B. (2008). Analysis of gene-expression data using J-Express. *Curr. Protoc. Bioinformatics* Chapter 7, Unit 7.3. doi: 10.1002/0471250953.bi0703s21
- Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., et al. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc. Natl. Acad. Sci. U.S.A.* 102, 15545–15550. doi: 10.1073/pnas.0506580102
- Tan, K., Feizi, H., Luo, C., Fan, S. H., Ravasi, T., and Ideker, T. G. (2008). A systems approach to delineate functions of paralogous transcription factors: role of the Yap family in the DNA damage response. *Proc. Natl. Acad. Sci. U.S.A.* 105, 2934–2939. doi: 10.1073/pnas.0708670105
- Teixeira, M. C., Monteiro, P. T., Guerreiro, J. F., Goncalves, J. P., Mira, N. P., dos Santos, S. C., et al. (2014). The YEASTRACT database: an upgraded information system for the analysis of gene and genomic transcription regulation in *Saccharomyces cerevisiae*. *Nucleic Acids Res.* 42, D161–D166. doi: 10.1093/nar/gkt1015
- Teytelman, L., Thurtle, D. M., Rine, J., and van Oudenaarden, A. (2013). Highly expressed loci are vulnerable to misleading ChIP localization of multiple unrelated proteins. *Proc. Natl. Acad. Sci. U.S.A.* 110, 18602–18607. doi: 10.1073/pnas.1316064110
- Thomas-Chollier, M., Herrmann, C., Defrance, M., Sand, O., Thieffry, D., and van Helden, J. (2012). RSAT peak-motifs: motif analysis in full-size ChIP-seq datasets. *Nucleic Acids Res.* 40, e31. doi: 10.1093/nar/gkr1104
- Thompson, D. A., Roy, S., Chan, M., Styczynsky, M. P., Pfiffner, J., French, C., et al. (2013). Evolutionary principles of modular gene regulation in yeasts. *Elife* 2:e00603. doi: 10.7554/eLife.00603
- Thorvaldsdottir, H., Robinson, J. T., and Mesirov, J. P. (2013). Integrative Genomics Viewer (IGV): high-performance genomics data visualization and exploration. *Brief. Bioinformatics* 14, 178–192. doi: 10.1093/bib/bbs017
- Ueta, R., Fujiwara, N., Iwai, K., and Yamaguchi-Iwai, Y. (2012). Iron-induced dissociation of the Aft1p transcriptional regulator from target gene promoters is an initial event in iron-dependent gene suppression. *Mol. Cell. Biol.* 32, 4998–5008. doi: 10.1128/MCB.00726-12
- Vermitsky, J. P., Earhart, K. D., Smith, W. L., Homayouni, R., Edlind, T. D., and Rogers, P. D. (2006). Pdr1 regulates multidrug resistance in *Candida glabrata*: gene disruption and genome-wide expression studies. *Mol. Microbiol.* 61, 704–722. doi: 10.1111/j.1365-2958.2006.05235.x
- Wapinski, I., Pfiffner, J., French, C., Socha, A., Thompson, D. A., and Regev, A. (2010). Gene duplication and the evolution of ribosomal protein gene regulation in yeast. *Proc. Natl. Acad. Sci. U.S.A.* 107, 5505–5510. doi: 10.1073/pnas.0911905107
- Wemmie, J. A., Szczypka, M. S., Thiele, D. J., and Moye-Rowley, W. S. (1994). Cadmium tolerance mediated by the yeast AP-1 protein requires the presence of

- an ATP-binding cassette transporter-encoding gene, YCF1. *J. Biol. Chem.* 269, 32592–32597.
- Wu, J., Chen, X., Cai, L., Tang, L., and Liu, L. (2015). Transcription factors Asg1p and Hal9p regulate pH homeostasis in *Candida glabrata*. *Front. Microbiol.* 6:843. doi: 10.3389/fmicb.2015.00843
- Wu, W. S., and Li, W. H. (2008). Identifying gene regulatory modules of heat shock response in yeast. *BMC Genomics* 9:439. doi: 10.1186/1471-2164-9-439
- Xu, T., Tripathi, S. K., Feng, Q., Lorenz, M. C., Wright, M. A., Jacob, M. R., et al. (2012). A potent plant-derived antifungal acetylenic acid mediates its activity by interfering with fatty acid homeostasis. *Antimicrob. Agents Chemother.* 56, 2894–2907. doi: 10.1128/AAC.05663-11
- Zhang, X., De Micheli, M., Coleman, S. T., Sanglard, D., and Moye-Rowley, W. S. (2000). Analysis of the oxidative stress regulation of the *Candida albicans* transcription factor, Cap1p. *Mol. Microbiol.* 36, 618–629. doi: 10.1046/j.1365-2958.2000.01877.x
- Znaidi, S., Barker, K. S., Weber, S., Alarco, A. M., Liu, T. T., Boucher, G., et al. (2009). Identification of the *Candida albicans* Cap1p regulon. *Eukaryot. Cell* 8, 806–820. doi: 10.1128/EC.00002-09

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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