



# Can Synovial Pathobiology Integrate with Current Clinical and Imaging Prediction Models to Achieve Personalized Health Care in Rheumatoid Arthritis?

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Although great progress has been made in the past decade toward understanding the pathogenesis of rheumatoid arthritis (RA), clinicians remain some distance from a goal of personalized health care. The capacity to diagnose RA early, predict prognosis, and moreover predict response to biologic therapies has been a research focus for many years. How currently available clinical prediction models can facilitate such goals is reviewed in this article. In addition, the role of current imaging techniques in this regard is also discussed. Finally, the authors review the current literature regarding synovial biomarkers and consider whether integration of synovial pathobiology into clinical prediction algorithms may enhance their predictive value.

**Keywords:** rheumatoid arthritis, synovial membrane, pathology, imaging, personalized medicine

## INTRODUCTION

Rheumatoid arthritis (RA) is characterized by a chronic symmetrical synovitis and underlying erosion of the subchondral bone and cartilage. It is associated with significant morbidity and mortality, with erosive damage directly related to disability and an economic burden for the UK economy alone of around £4 billion. It is now well recognized that the early diagnosis and treatment of RA equates to better long-term outcomes (1). The use of biologic drugs has transformed the treatment of RA, and since infliximab first came into routine clinical practice in the late 1990s, the biologics market for RA has expanded to include more than nine licensed preparations, with significantly more in late stage clinical development. Generally such agents are well tolerated, but adverse events can be severe and fatal and added to this is a high financial cost, with an annual cost of £200 million in the UK alone. Although variable, clinical trial data suggest response rates of around 60% (2) with first biologic treatment, as assessed by the composite clinical assessment tool of the American College of Rheumatology (ACR) 20 response or a fall in disease activity score of 28 joint count (DAS28) of >1.2. However, such level of clinical response is of modest significance to patient functionality, while an ACR 70 response that would enable patients to work and look after their family is reached in only 20–25%. Thus, a large proportion of patients (approximately 80%) are left with no response or significant ongoing disease activity. Additionally, a further proportion of patients will develop

secondary failure following an initially promising response, leaving altogether a huge unmet medical need.

In the era of personalized health care, a major focus of rheumatological research has been on predicting prognosis of RA, focusing on three main areas: (i) the early diagnosis and treatment of RA, (ii) predicting prognosis following diagnosis of RA, and (iii) predicting response to biologic therapies. On this basis, a number of clinical prediction models have been developed with variable reports of specificity and sensitivity for both early diagnosis and predicting prognosis of RA.

Synovial tissue is a crucial mediator of cartilage and subchondral bone erosion in RA and as such remains at the epicenter of joint pathology, although its exact place in the hierarchy of disease initiation and/or perpetuation is not entirely clear. In particular, the transition from the pre-clinical phase [breach of tolerance in secondary lymphoid organs (SLOs), mucosal associated lymphoid tissues, e.g., gut (GALT) and bronchial (BALT)] to localizing the disease to the joint is not fully understood. However, once the disease gets hold of the joint, examination of pathobiological specimens from the target organ, similarly to many other branches of medicine, including nephrology, dermatology, gastroenterology, and oncology, may become part of standard clinical care, with management decisions directed by the integration of pathobiology into clinical prediction models. With the advent of new techniques, such as ultrasound-guided synovial biopsy (3), the acquisition of synovial tissue has become a simple and well-tolerated procedure and emulating such a diagnostic/prognostic oncological model a potential goal. It is in this context that this review will discuss the currently available clinical prediction models and address whether the future integration of synovial pathobiology may make personalized health care in RA a more tangible objective.

## CURRENT CLINICAL PREDICTION MODELS

### Predictive Models for the Diagnosis of RA in Early Inflammatory Arthritis

The first clinical prediction model for use in early inflammatory arthritis was that developed by Henk Visser et al. (4). In a study of 524 early arthritis patients, seven variables: (i) symptom duration at first visit, (ii) early morning stiffness (EMS), (iii) arthritis in three or more joints, (iv) bilateral compression pain in the metatarsophalangeal (MTP) joints, (v) rheumatoid factor (RF) positivity, (vi) anti-citrullinated protein antibodies (ACPA) positivity, and (vii) the presence of erosions (hands/feet) were modeled for predictivity and shown to reliably discriminate between self-limiting, persistent non-erosive, and persistent erosive arthritis at 2-year follow-up. A subsequent model, again originating from the Leiden Early Arthritis Clinic (5), was developed for use in patients with early undifferentiated inflammatory arthritis to assess progression to erosive arthritis RA. The authors examined nine clinical variables: (i) sex, (ii) age, (iii) localization of symptoms, (iv) severity of EMS, (v) the tender joint count, (vi) the swollen joint count (SJC), (vii) C-reactive protein (CRP) level, and the presence of (viii) RF, or (ix) ACPA. These were

modeled (6) into a 14-point scale in a cohort of 570 early arthritis patients. They demonstrated that the positive predictive value for the development of RA with a score of 8 or greater was 91% and that the negative predictive value for patients with a score of 6 or lower was 84% and, further, that virtually none of the patients with a score of 3 or less were ultimately diagnosed with RA. This prediction rule has been validated in a number of other patient cohorts (7, 8) with similarly encouraging results. It should be noted, however, that 25% of patients in the study had a score between 6 and 8 and for these patients the chance of developing RA or not was equal, indicating the need for improvement of available predictive models in a quarter of patients.

The recognition that the 1987 ACR classification criteria for RA were unable to reliably diagnose patients with early disease led to the development of the ACR/EULAR 2010 classification criteria for RA (9), a process that incorporated a data-driven approach with consensus opinion. The 2010 classification criteria weigh the acute phase response [erythrocyte sedimentation rate (ESR)/CRP], joint involvement (number and size affected), duration of symptoms (>6 weeks), and seropositivity for RF/ACPA antibodies. These criteria (along with the van der Helm and Visser criteria) have most recently been explored in the REACH study cohort (10) with outcome measures defined as methotrexate use and persistent disease at 12 months. When comparing the ACR/EULAR 2010 criteria with the van der Helm and Visser scores, a superior sensitivity (0.74 vs 0.1 vs 0.59), although lower specificity (0.66 vs 1.0 vs 0.93) was demonstrated for the ACR/EULAR criteria (10). Currently, the individual criteria of the ACR/EULAR 2010 criteria are being analyzed within other early arthritis cohorts with the aim of further improving its diagnostic value (11).

### Predictive Models for the Prognosis of RA

Once RA is diagnosed, predicting the subsequent clinical course remains a challenge and in particular those patients at risk of rapid radiographic progression. Although a number of factors including seropositivity for RF/ACPA, baseline radiographic erosions, and high inflammatory markers associate with a higher erosive load (12–16), none are sufficient independent predictors of outcome (15). A number of clinical prediction models have, therefore, been developed in an attempt to overcome this difficulty (14, 17, 18) with data in general demonstrating that combining factors improves predictive power. For example, a model combining ACPA, sex, ESR, and IgM RF, was reported to have an accuracy of 73.6% to predict radiographic progression (19). A more recent report has suggested that integrating easily accessible clinical and laboratory variables (28 SJC, RF, and CRP or ESR) into a visual matrix model enables identification of those patients at risk of rapid radiological progression (6). The same visual matrix model was subsequently validated in an early arthritis cohort by Durnez et al. (20) to examine whether its application would improve therapeutic decision-making, but although it appeared to prevent overtreatment in the population studied, the predictive capacity for rapid radiological progression was low. Finally, this model (6) was also examined alongside two other prediction models, the BEST (21) and the SWEFOT model (19) in an observational RA cohort (22), and all three were found to have limited power to predict rapid radiological progression. More recently, a simplified

model incorporating three clinical parameters has been reported although sensitivity/specificity of prediction of radiographic damage was less with an AUC of 0.75 (23).

Finally, although not specifically a clinical model the widely validated multi-biomarker disease activity score, incorporating 12 serum biomarkers has recently been demonstrated to reliably predict radiographic progression at 12-month follow-up in a cohort of early RA patients although requires validation in further cohorts (24).

## INCORPORATING IMAGING INTO CLINICAL PREDICTION MODELS FOR THE DIAGNOSIS AND PROGNOSIS OF RA

Magnetic resonance imaging (MRI) and ultrasonography (US) have shown utility both in the early diagnosis of inflammatory arthritis and in monitoring its progression, and as such, a number of studies have examined their use as diagnostic/prognostic markers. MRI has been demonstrated to identify bone erosions early (25) and, furthermore, as a sensitive means to detect synovitis and bone marrow edema (26, 27). What is currently unclear, however, is how MRI functions as a tool to diagnose RA or further to predict its prognosis. Indeed a recent systematic review by Suter et al. (28) found no consensus on early MRI diagnostic criteria for RA and further concluded that bone marrow edema was the only significant predictor of radiographic progression (29). This is line with results from Duer-Jensen et al. (30) who examined predictive MRI features in an early undifferentiated arthritis cohort and identified BME as an independent predictor of subsequent RA development. The authors went on to integrate BME into a clinical matrix prediction model and directly compared the performance with the previously described van der Helm-van Mijl model (vdHvM model) (5) and reported an enhanced performance in this cohort with the former model correctly identifying the development of RA in 82 vs 60.2% of patients. The importance of BME was also highlighted by Tamai et al., who demonstrated in an early arthritis cohort that the combination of BME and ACPA positivity was equivalent to the application of the vdHvM model (31). Furthermore, the demonstration in a RF-negative cohort of 40 early arthritis patients (32) that MRI features of synovitis, erosions, and BME were significantly more sensitive than ACPA in correctly identifying RA patients suggest a potentially valuable role for MRI in this particular cohort.

Magnetic resonance imaging continues to be an expensive imaging modality, however, and is also time consuming and cumbersome for the patient. Conversely US can be performed swiftly and relatively cheaply within outpatient clinics, a considerable advantage, particularly as immediate treatment decisions can then be made following the scan mostly carried out and interpreted by rheumatologists. Again, it has been shown to be a sensitive tool to identify early erosions and synovitis (33). An initial report by Freeston et al. demonstrated a diagnostic benefit to adding ultrasonographic examination of the metacarpal-phalangeal (MCP) joints, wrists, and flexor tendons to conventional clinical tools (RF/ACPA status, ESR/CRP, radiographic damage) for the diagnosis of very early inflammatory arthritis (34). The predictive

algorithm appeared particularly helpful in seronegative patients, with a probability of certain diagnosis increasing from 30 to 94% once ultrasonographic features were incorporated. Interestingly, a recent study performed in very early arthritis incorporated US (of the MCP, wrist, and MTP joints) into both the van der Helm clinical prediction model and the ACR/EULAR 2010 classification criteria and demonstrated that incorporating ultrasound increased sensitivity for both clinical prediction models (35). Conversely, however, data from a real-life early arthritis clinic (36) cohort of 379 patients did not demonstrate a benefit in the addition of US into a risk metric derived from 12 clinical and serological parameters for the prediction of the development of either persistent inflammatory arthritis or RA.

## Clinical Predictive Models for Response to Biologic Treatment in RA

As already mentioned predicting the response to biologic agents is critical to the management of patients in order to optimize response and limit serious side effects. Although there are no clearly validated models for any of the biologic agents, a number of biomarkers have been explored. First, seropositivity for RF and ACPA and the primary response to anti-TNF agents has been examined in a number of cohorts (37–39) with mixed results, and thus at present, neither can be recommended as reliable biomarkers. Interestingly, however, it is becoming increasingly clear that seropositivity for RF and/or ACPA does enhance response to rituximab (40–42). Similarly, enhanced retention to abatacept in patients seropositive for RF and/or ACPA has also been observed in two independent cohorts (43, 44). Despite these observations, significant clinical challenges remain in predicting response to rituximab or abatacept with wide variability seen in response rates within the seropositive groups. Finally, preliminary data from an observational cohort of 530 RA patients have suggested that high levels of CRP retention with tocilizumab (45).

## WHAT CAN SYNOVIAL PATHOBIOLOGY ADD TO CURRENT CLINICAL PREDICTION MODELS?

Thus, while clinical prediction models offer potential (summarized in **Table 1**), they have a number of limitations, and at present, none are used in routine clinical practice, primarily because of lack of validation in large routine patient cohorts outside specialized centers and levels of sensitivity/specificity. These are important considerations and with a long-term goal of personalized health care much effort has gone into the search for additional biomarkers.

Rheumatoid arthritis is characterized by a thickened synovial membrane infiltrated by a diverse array of inflammatory cells, including macrophages, T and B lymphocytes, and NK cells associated with the proliferation of resident synovial fibroblasts. One of the characteristics of rheumatoid synovial tissue is the frequent capacity of the inflammatory cell infiltrate to organize into aggregates, a process termed ectopic lymphopoiesis (**Figure 1**). These aggregates are primarily composed of B and T lymphocytes, macrophages, and frequently follicular dendritic

**TABLE 1 | Summary of diagnostic clinical prediction models in rheumatoid arthritis.**

Clinical prediction model	Variables	Target population	Outcome	Sensitivity/specificity	Validated on external cohorts yes/no
Visser et al. (4)	i. Symptom duration, ii. early morning stiffness (EMS), iii. arthritis in >3 joints, iv. +ve metatarsophalangeal compression test, v. RF+ve, vi. ACPA+ve, vii. presence of erosions	Inflammatory arthritis	Self limiting vs persistent non-erosive (PNE), persistent erosive (PE) at 2-year follow-up	AUC 0.84 SI vs PE AUC 0.91 PNE vs PE (4)	Yes, e.g., Ref. (10)
Van der Helm-van Mil et al. (5)	i. Sex, ii. age, iii. localization of symptoms, iv. severity of EMS, v. tender joint count, vi. swollen joint count, vii. C-reactive protein, viii. RF+/- or ix. ACPA+/-	Undifferentiated arthritis	Erosive disease	AUC 0.87 (5)	Yes, e.g., Ref. (7, 10)
ACR/EULAR 2010 criteria, Aletaha et al. (9)	i. ESR/CRP, ii. joint involvement (number and size affected), iii. duration of symptoms (>6 weeks), iv. RF/ACPA+ve	Inflammatory arthritis	Persistent arthritis	Sensitivity 0.71 Specificity 0.65 (11)	Yes, e.g., Ref. (10, 46)

Variables, target population, outcome assessed, and sensitivity and specificity of each model are described.

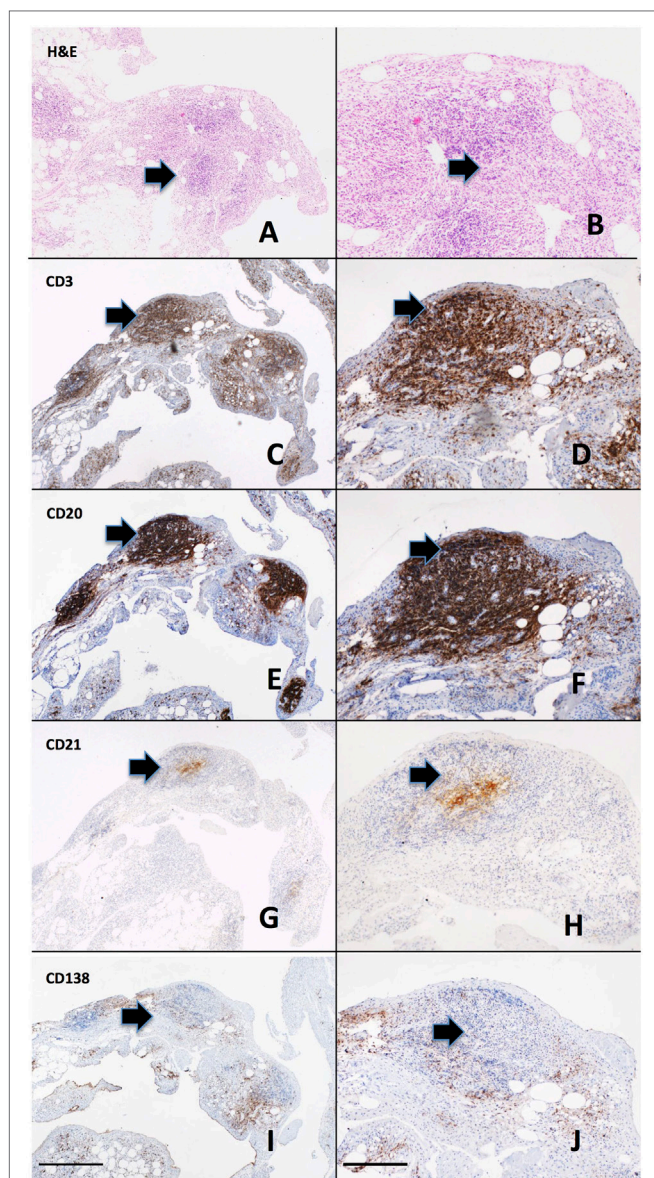
cells (FDCs). Such organized ectopic lymphoid structures (ELS) within the target organ of an autoimmune disease are not restricted to RA and indeed are seen within the salivary glands of patients with Sjogren's syndrome (47), the meninges of patients with multiple sclerosis (48) and in the thyroid of patients with autoimmune thyroid disease (49). Within rheumatoid synovial tissue, ELS can appear as tightly organized clusters closely resembling SLOs, expressing high endothelial venules and CD21+ follicular dendritic cells (50) (**Figure 1**). Following these observations, much debate has focused on their pathophysiological significance. Indeed, a direct functional role in inflammation and autoantibody production for synovial ELS in RA pathogenesis has come from a number of studies including our own work using the Human RA SCID mouse model (51). We demonstrated that synovial graft ELS support the proliferation and differentiation of B cells as well as the expression of activation-induced cytidine deaminase, an enzyme critical for the processes of class switch recombination and affinity maturation of antibodies (52), which correlated with human ACPA titers (IgG) within mouse serum. In addition, grafts characterized by the presence of ELS (vs ELS-ve grafts) showed higher level of production of B cell growth factors, TNF alpha, and RANK-ligand, supporting a direct functional role for autoantibody and pro-inflammatory mediators production by such structures. Of considerable importance, it should be noted that these events occurred in the absence of new immune cells infiltrating the grafts (SCID are immune deficient), indicating that ELS contribute to disease pathogenesis *via* protracted self-sustained immune activation. Further evidence includes the expression of RANK-ligand by activated T cells within synovial aggregates (53).

Direct evidence for a role in pathogenesis has also been suggested from clinical studies: a significant association was reported in established RA between synovial aggregates and erosive burden (54), and in a prospective cohort of early arthritis, increased synovial B cells were associated with the subsequent development of RA (55). However, the same authors (56) and others in larger cross sectional cohorts have found no association between synovial aggregates and radiological damage (57) and a recent prospective study of early arthritis patients could not confirm

either a diagnostic or prognostic role for aggregate synovial pathotypes (58). This has led a number of authors to conclude that lymphoid aggregates are a byproduct of inflammation, rather than a key component of the pathological process (59). However, interpretation of these data is complicated by a number of issues. First, no specific criteria are consistently used between studies to define aggregate synovial tissue histologically and as such the definition of ELS has been variously used to refer to aggregates of differing size and/or expressing different specific immunological cells (e.g., FDC, T cells, B cells). Further, comparing data from cross sectional studies including patients with varying (a) disease duration, (b) radiographic damage, and (c) treatment exposure (different therapeutic agents may influence synovial pathobiology differentially) has inherent methodological discrepancies. Finally, excessive prevalence of synovial tissue from the knee, as is the case with a number of studies, may introduce systematic bias, by including those patients with the most aggressive disease (60). These are crucial considerations, since a significant amount of data suggest that the induction of specific pathways related to lymphoid organization within the synovium (51, 53, 61, 62) may have short- and/or long-term effects on disease evolution. Thus, this concept requires further examination in a large-scale prospective study of early arthritis patients naïve to all therapy.

## SYNOVIAL PATHOTYPE AS A PREDICTOR OF RESPONSE TO THERAPY

Clearly, given the relative ease of access, the identification of biomarkers within peripheral blood would be preferable to those from synovial tissue. However, the search for peripheral biomarkers for response to TNF inhibitors (TNFi) has so far been challenging (63–67), and overall, it can be concluded that at present the currently examined peripheral blood biomarkers lack either sufficient sensitivity or specificity for predicting response to TNFis. Notably, however, the type 1 interferon signature has been identified as a potential marker of resistance to rituximab therapy in RA (68), but this still requires further validation in independent cohorts. The search for synovial biomarkers has, therefore, been driven by the ongoing challenge of selecting the



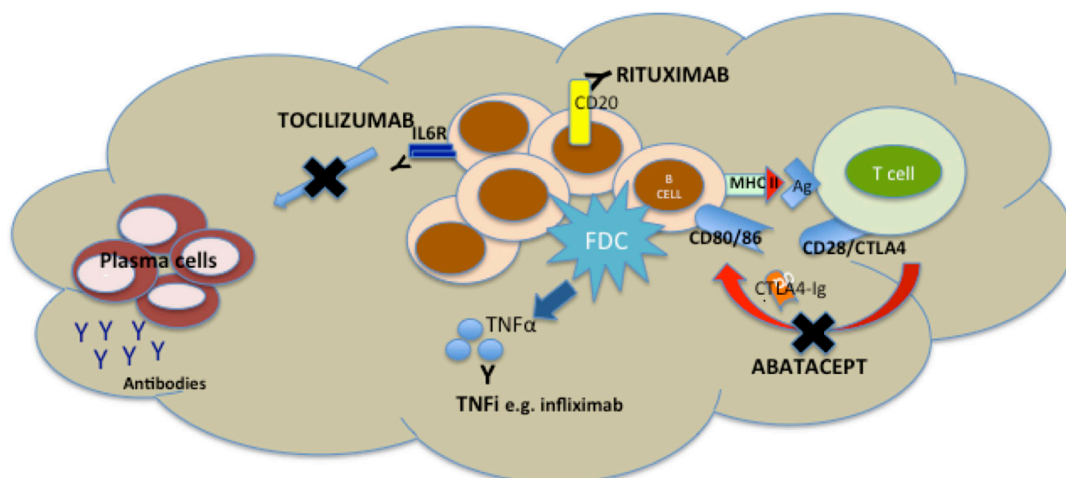
**FIGURE 1 | Ectopic lymphoneogenesis within the rheumatoid synovial membrane.** The synovial inflammatory cell infiltrate has been demonstrated to organize into discrete organized clusters clearly visible following routine hematoxylin and eosin staining (A,B). The aggregate is primarily composed of T cells [CD3, (C,D)] and B cells [CD20, (E,F)] with a central follicular dendritic cell network [CD21, (G,H)]. Plasma cells [CD138, (I,J)] are shown surrounding the aggregate. Sequentially cut 3  $\mu$ m sections of paraffin embedded rheumatoid arthritis (RA) synovial tissue from a patient with early RA are shown. Arrows indicate regions of positively stained cells. Left hand panel 4x magnification, right hand panel 20x, scale bar 200  $\mu$ m.

correct biologic in routine clinical practice, on the background of a burgeoning market for biologics usage in RA and by the limited utility so far demonstrated by peripheral blood biomarkers or clinical parameters.

TNF inhibitors are the most widely used biologic agents and have been in routine use for the longest and as such synovial pathobiological changes are probably the most exhaustively studied.

TNF $\alpha$ +/- LT $\alpha$  are inhibited by TNFi drugs, and both cytokines are key mediators of both the induction and maintenance of SLOs (69), so the capacity for modulation of synovial aggregates by TNFi has the potential to dissect the pathophysiology of the disease (Figure 2) as well as identification of biomarkers. Results from two groups have produced somewhat opposite results. An initial report from Cañete et al. (57) investigated the relationship between response to TNFi and the identification of large B cell aggregates with a pretreatment arthroscopic synovial biopsy in 86 patients. Further, in a subgroup of 24 patients, the capacity of TNFi to modulate B cell aggregates and the relationship with clinical response was explored. The data suggested that synovial lymphoneogenesis was an independent negative predictor of response to therapy, and response to treatment was associated with regression of ELS. However, a subsequent report from Klaasen et al. (70) in which synovial biopsies were performed on 97 patients prior to commencing treatment with infliximab identified the presence of lymphocyte aggregates as positive predictors of response at 16 weeks. There are a number of reasons for such discrepancies between the two groups including patient cohorts differing in terms of past TNFi exposure, disease duration, and variable follow-up times. In addition, a different definition of aggregate positive synovial tissue was used within each study, and the uniform use of a single TNFi was not consistent either within (57) or between the studies, a crucial consideration as increasing data suggest that efficacy of TNFis may involve more complex mechanisms than the simple division of drugs into either TNF antibody or TNF receptor (71). Interestingly, however, when the presence of synovial lymphocytic aggregates was added into a clinical prediction model (70) with TNF expression within the synovium, baseline DAS 28 score, and ACPA positivity, the power to predict response increased from 19 to 29%. A further potential synovial biomarker was identified when a preliminary study identified pretreatment levels of synovial TNF as a positive predictor of response to treatment (72) and a subsequent synovial-based study of 143 patients (73) supported these results with TNF expression within the synovial sublining explaining about 10% of the variance in response to therapy when multivariate linear regression was used to investigate synovial markers. Further, when a clinical prediction model incorporating both disease activity and TNF expression with the synovial sublining was formulated, 17% of variance in response to therapy could be explained (73). Thus, although at present, we remain some distance from personalized health care, these studies demonstrate proof of concept that in the long term the integration of synovial pathobiology into clinical prediction models may prove to be a useful clinical tool.

Whole tissue analysis using microarray technology has also been investigated to identify markers of response to therapy with such technology having the advantage of providing multiple simultaneous biomarkers. Its use was supported by two studies: the first where synovial tissue available from 10 patients pre- and post-TNF therapy (74) was examined and suggested a differential expression of genes between responders and non-responders and modulation of transcription profiling following treatment. The second study including 18 patients again suggested a differential expression of predominantly inflammatory genes in TNFi responders (75). Conversely, another group reported in a biopsy-based study of



**FIGURE 2 | Mechanisms of disruption of ectopic lymphoneogenesis within rheumatoid synovial tissue via currently licensed biologic agents for rheumatoid arthritis (RA).**

Disruption of ectopic lymphoneogenesis within the rheumatoid synovial membrane may be mediated by a number of biologic agents licensed for the treatment of RA and, therefore, has a putative role as a biomarker of response/resistance to biologic therapies. Rituximab induces death of CD20+ B cells via Ab or cell-mediated cytotoxicity, phagocytosis, or cell lysis. Abatacept, a CTLA-Ig fusion protein, prevents endogenous CTLA from binding to CD80/86 on B cells (functioning as antigen presenting cells) and so prevents co-stimulatory signals to the B cell. TNF inhibitors (TNFi) (inhibitors) such as infliximab bind to soluble and membrane-bound TNF $\alpha$  released by follicular dendritic cells (FDCs), and so inhibiting FDC-mediated B cell attraction. Tocilizumab inhibits binding of IL6 to its receptor, preventing IL6-mediated B cell proliferation and differentiation.

25 patients that a gene signature characterized predominantly by markers of cell division and immune activation predicted response to therapy (76). A large study of synovial tissue from 62 RA patients was established to attempt to identify further biomarkers of response using transcriptional analysis (77). This study did identify 38 genes associated with good response, but only within aggregate positive patients. Interestingly, more recently published data from Dennis et al. (78) have utilized whole tissue microarray as a tool to characterize the molecular profile of three previously reported (79) pathobiological synovial subgroups: myeloid, fibroid, and lymphoid subtypes. The same authors (78) also went on to identify two soluble serum markers associated with synovial pathotypes: ICAM1 (myeloid) and CXCL13 (lymphoid), and demonstrated that each was found to predict response to adalimumab (ICAM1) and tocilizumab (CXCL13). Such data require validation in larger prospective cohorts but highlight that disease heterogeneity may be more complex than that dissected by simple histopathological assessment of synovial tissue and, furthermore, suggest the potential role for synovial tissue in rationalizing choice of peripheral biomarkers. Therefore, it seems likely that any robust prediction model will have to incorporate multiple pathobiological as well as clinical biomarkers.

Thus, at present, despite these observations, the capacity to predict response to TNFi remains remote to practicing clinicians. What seems crucial to stress, particularly with an ever-increasing number of TNFi available, each with unique pharmacological characteristics and capable of modulating unique pathobiological processes, is that extrapolating biomarkers from individual TNFi to the whole group will likely prove a futile exercise. What is needed are large-scale pathobiological studies conducted in patients naive to all biologic therapy and focusing on individual TNFi, this approach promises a unique opportunity to dissect disease mechanism in parallel with identifying novel biomarkers.

Rituximab, a monoclonal antibody against CD20, has demonstrated efficacy in RA, with response rates of around 60% (80). Although seropositivity for RF and/or ACPA has emerged as a positive predictor of response to treatment following a subgroup analysis of a number of studies (81–83), outcome to treatment remains unpredictable. Clinical studies pre and post rituximab therapy have demonstrated a number of potential pathways within synovial tissue that may explain the heterogenous clinical response seen despite apparent complete depletion in the peripheral blood. First, in a small study of 13 patients with synovial biopsies taken prior and at 8 weeks following standard rituximab treatment a trend toward clinical response and depletion of both synovial B cells and immunoglobulin synthesis was demonstrated (84). Second, a study of 24 patients identified reduction in the number of plasma cells, potentially originating locally within the synovial tissue, 16 weeks post therapy as a predictor of response to therapy (85). Finally, the identification of circulating pre plasma cells, by high sensitivity FACs analysis following the first cycle of rituximab associated with a poor response to treatment (86). Such cells are thought to originate from solid tissue, e.g., bone marrow, though the contribution of autoreactive clones escaping from the synovium may also play a role. It may well be concluded from these studies that modulating the functional B cell activity within synovial tissue has an important contribution on response to treatment, a hypothesis that is currently being explored by the authors of this paper through two randomized biopsy driven clinical trials (a) the National Institute for Health Research: Response, Relapse, Resistance to Rituximab (R<sub>4</sub>RA) study and (b) Stratification of Biologic Therapies for Rheumatoid Arthritis by Pathobiology (STRAP) study jointly funded by the Medical Research Council and Arthritis Research UK.

Early data on the synovial response to abatacept, a specific modulator of T-cell activation via inhibition of the CD80/86-CD28

interaction, have suggested a characteristic response in patients responding to treatment. In the study involving 16 RA patients with synovial tissue obtained prior and 16 weeks following the initiation of therapy, a significant downregulation of a number of pro inflammatory genes and a specific reduction in synovial CD20+ B cells was seen in patients responding to treatment (87). Further studies are awaited.

## CONCLUSION

A number of major advances in the management of RA have been made in the past decade. Importantly, overwhelming evidence now supports early intervention (1). The introduction of biologic drugs into the therapeutic armamentarium has been associated with a paradigm shift in treatment goals, from an often previously ill-defined response to therapy to disease remission or even cure. Such aims require early diagnosis and to target biologic treatments appropriately, to patients with the worst prognosis and to those most likely to respond to individual agents. The currently available clinical prediction models are insufficiently sensitive or specific to allow reliable early diagnosis or indeed predict those with the worst prognosis and so far biomarkers for response to biologics are widely lacking. Although the simple pathological characterization of synovial tissue into subtypes seems unlikely to aid prognosis *per se*, an enormous number of questions remain before fully understanding how the heterogeneous synovial pathotypes integrate with disease pathogenesis and ultimately the

heterogeneity of clinical phenotypes of disease. The complexity and diversity of RA pathogenesis is perhaps as such that would require more integrated and modular approaches as needed for cancer medicine (79, 88, 89). What is clear from the current conflicting data is that future synovial biopsy studies need to be conducted on large cohorts, including early arthritis patients naïve to DMARD therapeutic intervention. Significant progress in minimally invasive ultrasound-guided techniques which are easily carried out by rheumatologists, safe and well tolerated by patients, and allow for the acquisition of synovial tissue both from small and large joint in most patients will facilitate the conduction of such studies. These may not only predict prognosis but have the potential to predict response to future therapeutic intervention at first presentation. It seems realistic to suggest that in future synovial pathobiology will be integrated into current clinical prediction models with positive benefits for patient care and health economics through accurate patient stratification.

## AUTHOR CONTRIBUTIONS

All authors substantially contributed to the conception/design, acquisition, and analysis of the work. All authors contributed to drafting of the work and gave final approval of the version to be published and are all in agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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