### Check for updates

### OPEN ACCESS

EDITED BY Liping Feng, Duke University, United States

#### REVIEWED BY Gil Mor, Wayne State University, United States Carla Guenther, Osaka University, Japan

\*CORRESPONDENCE Meirong Du mrdu@fudan.edu.cn Lu Liu Iulu11n@163.com

RECEIVED 05 November 2023 ACCEPTED 18 December 2023 PUBLISHED 11 January 2024

#### CITATION

Yin T, Li X, Li Y, Zang X, Liu L and Du M (2024) Macrophage plasticity and function in cancer and pregnancy. *Front. Immunol.* 14:1333549. doi: 10.3389/fimmu.2023.1333549

#### COPYRIGHT

© 2024 Yin, Li, Li, Zang, Liu and Du. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Macrophage plasticity and function in cancer and pregnancy

Tingxuan Yin<sup>1</sup>, Xinyi Li<sup>1</sup>, Yanhong Li<sup>1</sup>, Xingxing Zang<sup>2</sup>, Lu Liu<sup>1\*</sup> and Meirong Du<sup>1\*</sup>

<sup>1</sup>Lab of Reproduction Immunology, Shanghai Key Laboratory of Female Reproductive Endocrine-Related Diseases, Hospital of Obstetrics and Gynecology, Fudan University Shanghai Medical College, Shanghai, China, <sup>2</sup>Department of Microbiology and Immunology, Albert Einstein College of Medicine, Bronx, NY, United States

As the soil of life, the composition and shaping process of the immune microenvironment of the uterus is worth exploring. Macrophages, indispensable constituents of the innate immune system, are essential mediators of inflammation and tissue remodeling as well. Recent insights into the heterogeneity of macrophage subpopulations have renewed interest in their functional diversity in both physiological and pathological settings. Macrophages display remarkable plasticity and switch from one phenotype to another. Intrinsic plasticity enables tissue macrophages to perform a variety of functions in response to changing tissue contexts, such as cancer and pregnancy. The remarkable diversity and plasticity make macrophages particularly intriguing cells given their dichotomous role in either attacking or protecting tumors and semi-allogeneic fetuses, which of both are characterized functionally by immunomodulation and neovascularization. Here, we reviewed and compared novel perspectives on macrophage biology of these two settings, including origin, phenotype, differentiation, and essential roles in corresponding microenvironments, as informed by recent studies on the heterogeneity of macrophage identity and function, as well as their mechanisms that might offer opportunities for new therapeutic strategies on malignancy and pregnancy complications.

#### KEYWORDS

macrophages, plasticity, maternal-fetal interface, tumor-associated macrophages (TAMs), immunosuppression, immune microenvironment

Abbreviations: ARG1, arginase 1; ANG-2, angiopoietin 2; CTL, cytotoxic T cells; CAFs, cancer-associated fibroblasts; CSF-1, macrophage-stimulating factor 1; CCL, chemokine (C-C motif) ligand; CXCL, chemokine (C-X-C) ligand; DC, dendritic cell; dMφ, decidual macrophages; EVT, extravillous trophoblasts; FBP, fructose-1,6-bisphosphate; FGR, fetal growth restriction; iNOS, nitric oxide synthase; IDO, Indoleamine 2,3-dioxygenase; IFN- $\gamma$ , interferon-gamma; ICAM1, intercellular adhesion molecule 1; IL, interleukin; Kyn, Kynurenine; LPA, lysophosphatidic acid; MDSCs, myeloid-derived suppressor cells; mIHC, multiplex immunochemistry staining; PE, preeclampsia; RPL, recurrent pregnancy loss; scRNA-seq, single-cell RNA sequencing; TGF- $\beta$ , transforming growth factor-beta; TAM, tumor-associated macrophages; TME, tumor microenvironment; TRMs, tissue-resident macrophages; TNF- $\alpha$ , tumor necrosis factor-alpha; VEGFA, vascular endothelial growth factor-A.

### 1 Introduction

Macrophages are a subset of mononuclear phagocytes of the innate immune system. As a multifunctional immune population, they not only play a central role in inflammation and host defense but also exert homeostatic functions in a wide range of peripheral tissues. Remarkably, macrophages are one of the most plastic and diverse cells among human hematopoietic cells, involved in nearly all diseases in humans. In local tissues, microenvironmental clues mold macrophages morphologically and functionally according to the homeostatic needs of their surroundings, leaving a profound imprint on the tissue-specific functions of macrophages.

As a place where the embryo implants and develops, the maternalfetal interface is highly dynamic and multi-functional to maintain a homeostatic environment that requires both immunotolerance toward the semi-allogeneic fetus, defense against pathogens, and tissue remodeling. At the maternal-fetal interface, macrophages are present in the whole stage of gestation, where they participate in various biological processes dynamically, including decidualization, trophoblast invasion, angiogenesis, parturition, and postpartum uterine involution (1). Similarly, in the setting of cancer, phenotype shift and function skewing of macrophages also occur along with tumor initiation, progression, and metastasis (2). Originally, activated macrophages central to the chronic inflammatory process mediate the elimination of senescent epithelial cells and tumor tissue disruption, which drives subsequent carcinogenesis (3). Later upon tumor establishment, cancer cell products transform neighboring macrophages from a classically activated state to a trophic, immunoregulatory, and pro-angiogenic phenotype that favors tumor escape (4).

Dates back 30 years ago, some researchers noticed that the penetrative property of hemochorial placentation mimics the phenomena observed in highly invasive tumors, and both are accompanied by a hypoxic environment and abundant immune cells (5, 6). Indeed, as a kind of versatile immune component, macrophages contribute a lot to the formation of immune tolerance and new blood vessels through their extreme plasticity and diversity (7). However, investigations that draw parallels between the two are lacking (5, 7, 8). Recent studies combining lineage tracing, single-cell multi-omics, and morphologic and molecular analysis have illustrated the essential role and mechanisms of diverse macrophage populations in mediating various aspects of both pathological processes like tumor development and physiological status as pregnancy (9-11). In this review, we described the phenotypical heterogeneity and factors regulating macrophage differentiation in the context of both cancer and pregnancy and compared the concrete mechanisms of how macrophages exert immune-modulatory and angiogenic capacities.

# 2 The microenvironment of tumor and decidua

The maternal-fetal interface is composed of the fetal-derived placenta and the maternal-derived decidua, whose proper establishment guarantees a successful pregnancy. For better adaption to embryo implantation, on the maternal side, endometrial stromal cells (ESCs) begin to proliferate and differentiate into large, round, cytoplasm-rich, multi-nucleated decidual cells, known as decidual stromal cells (DSCs), and this process is defined as "decidualization" in a narrow sense (12, 13). Upon embryo implantation, on the fetal side, the outermost layer of the blastocysts transforms into cytotrophoblasts (CTBs), which then generate multinucleated synctiotrophoblast (SCTs) and invasive extravillous cytotrophoblasts (EVTs) after series of chorionic villi differentiations. SCT is the outer lining of the chorionic villi that is in direct contact with maternal glandular secretions and maternal blood flowing into the intervillous space full of maternal peripheral blood, whereas EVTs get in touch with decidua and infiltrate into local lymphatics, veins, and glands, participating in placenta construction and fetal development (14, 15). Derived from the anchoring villi (the distal tips of chorionic villi in contact with the decidua), proliferative EVTs migrate into the decidua and experience two differentiation pathways toward opposite directions: The interstitial EVT invades across the decidual stroma toward the maternal spiral arteries as far as the muscular layer of the uterine wall, whereas the endovascular EVT moves in a retrograde manner down the artery to form a plug that prevents blood from entering the intervillous space and replaces endothelial cells (16). At the 10th week of gestation, when the placenta switches from histiotrophic to haemotrophic nutrition, EVT plugs dissolve, licensing abundant blood support to the fetus (17).

In the first trimester of pregnancy, decidual immune cells concede the invasion of fetal-derived cells and cooperate with the process of initial placental development and angiogenesis by forming an antiinflammatory network and releasing pro-angiogenetic cytokines (18). In this period, up to 40% of decidual cells are leukocytes, where natural killer (NK) cells account for the most at around 70%, followed by decidual macrophages (dM $\phi$ , 20%–25%) and T cells (3%–10%), dendritic cells (DC), and B cells (18). Among these, dM $\phi$  performs crucial functions, including spiral arteries remodeling, facilitating trophoblast invasion, scavenging apoptotic cells, canonical antimicrobial function, and aiding in the construction of the immune tolerant microenvironment (19).

The tumor microenvironment (TME) is a unique milieu featuring a shortage of nutrients and scarce oxygenation, which contains tumor cells, extracellular matrix, stromal cells (such as fibroblasts, mesenchymal stromal cells, pericytes, adipocytes, blood, and lymphatic vasculatures), and immune cells (including T and B lymphocytes, DC, NK cells, monocytes, and macrophages) (20). Like the decidua, a dynamic network of tumor-related cytokines and chemokines, growth factors, and metabolic products together achieve the balance of tumor inhibition and promotion (5). As the most abundant immune population within the TME, tumorassociated macrophages (TAMs) constitute a plastic and heterogeneous cell population that accounts for up to 50% of some solid neoplasms and are associated with adverse prognoses (21). They have been demonstrated to exert multiple protumor activities that range from tumor initiation and promotion, metastasis and colonization, immune evasion, angiogenesis, and extracellular matrix remodeling (22). Importantly, because of their intrinsic plasticity, TAMs can rapidly respond to surrounding perturbations occurring in the process of tumor progression and therapeutic challenges. Moreover, TAMs prefer to enrich in a

hypoxic TME milieu with poor blood supply and contribute to tumor vasculature formation by producing molecules and enzymes with proangiogenic effects (23)

It can be perceived that the early maternal-fetal interface bears a notable resemblance with TME in terms of structural composition, in which tumor cells and trophoblasts act as the center surrounded by extracellular matrix (ECMs), stromal cells, and immune components, and they both have unusual vascularity. However, the proportion of various immune cells is inconsistent between the two settings, as TAMs dominate the immune environment of tumors, whereas decidual NK cells lead at the maternal-fetal interface. Different from the close relationship between tumor and tumor-draining lymph nodes (TdLNs), ESC decidualization results in a loss of lymphatics (24, 25), which restricts the DC migration and homing to uterine-draining lymph nodes (UdLNs) and avoids the susceptibility to mounting an overwhelming immune response to attack the fetus. Figure 1 characterizes these microenvironments in both settings.

# 3 Diversity of origins and phenotypes of macrophages

Macrophages are the preliminary immune system to appear during embryonic development, and their exact ontogeny has been widely explored. In adults, macrophages are ubiquitous and diffuse across various bodily tissues and organs, and, for a long time, they are assumed to originate exclusively from bone marrow-derived progenitors and blood monocyte intermediates (27, 28). So far, mounting evidence has demonstrated that macrophages have additional origins in a developmental stage- and tissue-specific way (11). Fate-mapping studies and single-cell analyses in murine indicate that tissue-resident macrophages (TRMs) develop either from the macrophage's embryonic yolk sac progenitors or fetal liver-derived erythro-myeloid progenitors (EMPs) (11). In humans, the embryo has two waves of TRMs from either yolk sac-derived primitive macrophages or yolk sac-derived myeloid-biased progenitor (YSMP)-derived embryonic liver monocytes, which corresponds to EMPs in mice (29). The diverse macrophage ontogeny is deciphered in detail in Figure 2.

In tumors, TAMs are essentially comprised of two origins: existing TRMs and circulating bone marrow-derived macrophages (BMDMs) (35). Monocyte-related MDSCs (M-MDSCs) are currently known as the main circulating precursor of TAMs, which are induced by a broad range of inflammatory signals to infiltrate into the tumor entity and are influenced by extreme survival conditions, like hypoxia (36). Ligand/receptors axes mediating TAMs recruitment and accumulation mainly include macrophage colony-stimulating factor/colony-stimulating factor 1 receptor (M-CSF/CSF-1R), CCL2/CCR2, CX3CL1/CX3CR1, CCL3/ CCR1, CCL3/CCR5, CCL5/CCR5, and vascular endothelial growth factor-A/vascular endothelial growth factor receptor 1 (VEGF-A/ VEGFR1) (37). Compared to physiological conditions, tumors are featured by a significantly higher expansion of TRMs, such as lung carcinoma, glioblastoma, and hepatocellular carcinoma (35).



#### FIGURE 1

The architecture and surroundings of the immune microenvironment in tumor and decidua. The diagonal frame shows that macrophages constitute a major component of both immune microenvironments. The two microenvironments are composed of multiple cell types including tumor cells/ trophoblasts, fibroblasts/stromal cells, endothelial cells, epithelial cells, and various immune cells, where macrophages constitute a major immune component (5). At the decidual face, growing invasive fetal cells come in direct contact with decidua, as well as the abundant maternal blood after the EVT plug is eroded at the beginning of the second trimester (16). Unlike the primary role of NK cells in the decidua, TME of cancer is dominantly infiltrated with macrophages (26). In particular, dendritic cells efferent from tumors can present cancer-related antigens to T cells after they enter lymph nodes, whereas decidual dendritic cells cannot migrate to draining lymph nodes due to the degeneration of lymphatic vessels (25). CTB, cytotrophoblast; CAF, cancer-associated fibroblast; DSC, decidual stromal cells; EVT, extravillous trophoblasts; SCT, synctiotrophoblast; TdLN, tumor-draining lymph nodes.



#### FIGURE 2

The ontogeny and phenotypic differentiation of macrophages. The top half of the graph depicts the murine fetal macrophage development. The first wave, known as primitive hematopoiesis, directly occurs in the blood islands of the extra-embryonic yolk sac at E7.5 and the yolk sac differentiates into microglia (11); the second wave is initially generated from proliferative EMPs in the yolk sac around E8.25 and subsequently arrive fetal liver, where progenitors produce fetal liver monocyte-derived macrophages, seeding diverse tissues, and differentiate into tissue-resident macrophages with renewing ability (11). The final wave relies on HSCs and begins to produce fetal hematopoietic stem and progenitor cells from approximately E9.5 in the intra-embryonic mesoderm and then transfers to the fetal liver for further maturation (30). Later, HSCs migrate to and colonize the developing fetal bone marrow by E16.5, where they remain and deliver monocytes to peripheral blood throughout adulthood (31). The different colors represent each origin of macrophages. The lower half shows the traditional M1 and M2 nomenclature, involving distinct transcription factors, phenotypes, and secretory profiles, resulting in different functions according to the complex external contexts (32). M1 macrophages are acutely stimulated by IFN- $\gamma$  or LPS and secrete inflammatory factors and killing molecules against foreign invaders (26). Depending on the activating stimulus, M2 macrophages can be further divided into four different subsets consisting of M2a, M2b, M2c, and M2d (33). In particular, M2a (induced by IL-4 and IL-13) and M2b (induced by TLR or IL-1R agonists) exert immunoregulatory functions and drive Th2 responses, whereas M2c macrophages (induced by IL-10) are more related to anti-inflammatory activities against apoptotic cells and M2d macrophages (induced by TLR antagonists) are inclined to induce tumor progression (33, 34). ACs, apoptotic cells; BM, bone-marrow; CXCL, chemokine (C-X-C) ligand; CCL, chemokine (C-C motif) ligand; EMPs, erythro-myeloid progenitors; GC, glucocorticoids; GM-CSF, granulocyte-macrophage colony-stimulating factor; HSC, hematopoietic stem cells; HIF, hypoxia-inducible transcription factors; IFNg, interferon-gamma; iNOS, inducible nitric oxide synthase; MHC, major histocompatibility complex; MPs, myeloid progenitors; M-CSF, macrophages-stimulating factor; PPARy, peroxisome proliferator activated receptor  $\gamma_{c}$  TLR, Toll-like receptor; TNF- $\alpha_{c}$  tumor necrosis factor-alpha; TGF- $\beta_{c}$  transforming growth factor-beta; Ym1/2, Ym1 and Ym2.

Evidence suggests that TAMs of different origins may coexist in tumors, and TAMs of different origins display different functions within a certain TME (35). However, recent studies suggest that different niches can direct TAMs activation profiling in a cancer type-, tissue-, and stage-specific way, highlighting a determinate role of specific TME rather than origin in the education of TAM populations (38).

Compared to TAMs, evidence covering the origin of  $dM\phi$  is scarcer (30). During menstruation, an influx of macrophages derived from classical monocytes flow into secretory endometrium emerges, concomitant with the increase of macrophage-derived cytokines and proteases (31). At 18 days after fertilization, a group macrophage of fetal origin emerges in the chorionic plate, later migrating to villi at the placenta, which is referred to as Hofbauer cells (human, CD14<sup>+</sup>HLA-DR<sup>-</sup>FOLR2<sup>+</sup>; and mouse, CD11b<sup>10</sup>F4/80<sup>hi</sup>CX3CR1<sup>hi</sup>) (39). Decidual macrophages of maternal origin (human, CD14<sup>+</sup>HLA-DR<sup>hi</sup>; mouse, CD11b<sup>hi</sup>F4/ 80<sup>lo</sup>), replenished from the maternal circulation by HSC-derived cells, are present in decidua basalis throughout gestation (39). These two populations account for 70% and 30% of macrophages at the maternal-fetal interface, respectively (40). CSF-1 and CX3CR1 are reported to regulate monocyte extravasation from blood vessels to the uterus (31, 41). The specific cluster of  $dM\phi$  with self-renewing capacity existed since the embryonic stage of the mother and is more likely dominant to exert a homeostatic role at decidua during pregnancy than subsequent circulating monocytes (41).

Dynamic TAM phenotype drives different functions, which could influence cancer outcome. During a long time, it is commonly accepted that macrophages are classified into two distinct subsets M1 and M2 that represent two extreme activation conditions, where the former excels at fighting against pathogens to protect the host, whereas the latter is considered to be associated with immunoregulatory and pro-tumoral activities (26, 32). Figure 2 shows the current M1/M2 nomenclature, involving distinct transcription factors, phenotypes, and secretory profiles, resulting in different functions according to the complex external contexts. However, it should be noted that M2-like macrophages should not be regarded as cancer-specific, even less TAMs (26) but

rather present an alternative activated status displaying high phenotypic heterogeneity (currently thought to be four subsets) and governing functions at the interface of immunity, tissue homeostasis, and metabolism (33, 34, 42). Indeed, M2-like macrophages can occur after any immune response in healthy organisms to initiate tissue remodeling and dissolve inflammation. In cancer, macrophages crewing toward M2 status are hijacked to benefit tumor progression (43). Therefore, this binary-polarization model neglects that the adaption driven by environmental signals in the TME is flexible rather than static and could not decipher the complex activation and heterogeneity of TAMs (44). Backed by some large-scale transcriptomic studies, the model "multi-function knife" began to be widely accepted indicating that TAMs are present in a transitional intermediate state, consistent with extraordinary plasticity and a broad spectrum of activation and functional states (10), and some recently identified TAMs subsets (45-48) with specific functions have been listed in Table 1.

 $dM\phi$  possesses functional and quantitative plasticity with remarkable adaptation to changing environments (55). Initially, seminal plasma triggers an essential inflammatory milieu that

TABLE 1 The summary of the recent classification of TAMs and  $dM\phi.$ 

contains an increase in M1-skewed macrophages in the periimplantation uterus (56). Upon EVT attachment onto the endometrial lining and invasion into decidua, it transits to a mixed profile of M1/M2 macrophages (14). A predominantly M2 phenotype gradually forms along with uterine vascular remodeling, culminating with completed placental development at the beginning of the second trimester (57). These M2-like  $dM\phi$ promote immune tolerance to semi-allogenic fetuses and protect fetal growth till parturition, which is a major source of interleukin-10 (IL-10) at decidua. Finally, the surge of M1 dMo at parturition, together with an outflux of inflammatory cytokines, suggests their important role in triggering the term delivery (58). DMqs exhibit excellent plasticity and are sensitive to small changes in the microenvironment, including nutritional status, smoking habits, inflammatory conditions, and physiological stress. Inappropriate macrophage plasticity, whenever it occurs, could impart adverse effects on pregnancy outcomes, leading to complications like infertility, recurrent pregnancy loss (RPL; defined as loss of two or more consecutive pregnancies), preeclampsia [PE; defined as a pregnancy-related syndrome with high blood pressure (≥140/90 mmHg) and/or proteinuria ( $\geq 0.3$  g/L)], fetal growth restriction

	Time	Method	Source	Subset (phenotype and function)	Marker	Reference
TAMs	2019	scRNA-seq	CRC, HCC, and OSCC	FCN1 <sup>+</sup> TAMs	FCN1, VCAN, FCGR2A, FCGR3A, S100A6, S100A8, S100A9, and S100A12	(45-47)
	2017	scRNA-seq	CRC, HCC, and NSCLC	C1Q <sup>+</sup> TAMs	C1QA/B/C, APOE, TREM2, and SLC40A1	(46-48)
	2020	scRNA-seq	CRC	SPP1 <sup>+</sup> TAMs	SPP1, IL1RN, OLR1, VEGFA, EREG, FN1, C15ORF48, PHLDA1, AQP9, TNS3, and NDRG1	(46)
dΜφ	2011	Flow cytometry	NP decidua of 6– 12 weeks	CD14 <sup>+</sup> CD11c <sup>hi</sup> : lipid metabolism and inflammation, process protein Ag CD14 <sup>+</sup> CD11c <sup>lo</sup> : extracellular matrix formation, muscle regulation, and tissue growth	CD14, CD45, and CD11c	(49)
	2011	Flow cytometry	NP decidua of 7– 11 weeks	CD163 <sup>high</sup> CD206 <sup>high</sup> CD209 <sup>high</sup> NRP-1 <sup>high</sup> ICAM-3 <sup>low</sup>	CD14, CD163, CD206, CD209, and NRP-1	(50)
	2018	scRNA-seq	NP decidua and placenta of 6–14 weeks	dM1: remodeling dM2: self-renewal and remodeling	ITGAX and IL-10	(51)
	2018	Flow cytometry	NP decidua of 6-8weeks	CCR2 <sup>+</sup> CD11c <sup>hi</sup> : pro-inflammatory characteristics CCR2 <sup>-</sup> CD11c <sup>hi</sup> : anti-oxidative and anti- inflammatory CCR2 <sup>-</sup> CD11c <sup>lo</sup> : antigen presentation	CCR2, HMOX1, MARCO, MHCII, and IL1B	(49, 52)
	2021	scRNA-seq and ATAC-seq	Decidua of 15 NP (7-24 weeks) and nine RPL (8- 50 weeks) women	CD11b <sup>+</sup> : M1 polarization characteristics CD11c <sup>+</sup> : M2 macrophage homeostasis in the decidua	CD11b <sup>+</sup> : S100A10, S100A6, S100B, and S100A4; CD11c <sup>+</sup> : PLAU, DAB2, EGR1, SEPP1, and MAF	(9)
	2021	scRNA-seq	Decidua of six NP (7 weeks) and six RPL (5 weeks) women	Ten clusters according to transcriptional states, suggesting that dMs do not fit the conventional M1/M2 classification	IL1BIL6, IL10, MS4A7, CCL2, CCL3, and CCL4	(53)
	2021	scRNA-seq	Decidua of 10 NP and 14 RPL women (6–8 weeks)	CD14-expressing dM cells	CD14, ITGAX, CXCL8, CXCL3, C1QA, C1QB, and C1QC	(54)

CRC, colorectal cancer; dM, decidual macrophages; HCC, hepatocellular cancer; NSCLC, non-small-cell lung cancer; NP, normal pregnancy; OSCC, oral squamous cell carcinoma; RPL, recurrent pregnancy loss; scRNA-seq, single-cell RNA sequencing. ATAC-seq, Assay for Transposase Accessible Chromatin with high-throughput sequencing.

(FGR; defined as a failure of fetus to reach its expected biological growth), and preterm delivery (59). In the past decades, scientists have made efforts to identify  $dM\phi$  subgroups and confirm phenotypes of  $dM\phi$ s related to the maintenance of healthy pregnancy (49–54, 60), which is summarized in Table 1.

# 4 Factors regulating macrophage phenotype and function

### 4.1 Metabolic reprogramming

### 4.1.1 Hypoxia

With restricted oxygen availability, tumor cells undergo metabolic adaptations and bear the capacity to initiate high rates of aerobic glycolysis, which generates a nutrient-deficient TME (61). In this context, TAM states and phenotypes can be adjusted by disparities in tumor tissue oxygenation, which benefits tumor cells to overcome nutritive deprivation and convert the TME into more hospitable sites (2). More specifically, M2 cells tend to be in hypoxic, necrotic, or perivascular tumor sites where they dampen anti-tumor inflammation, whereas M1 cells enrich normoxic regions (62). Although hypoxia generates both pro-inflammatory markers, like tumor necrosis factor-alpha (TNF- $\alpha$ ), IL-1 $\beta$ , and inducible nitric oxide synthase (iNOS), as well as anti-inflammatory markers such as IL-10 and arginase 1 (ARG1), TAMs shift mainly toward the M2-like phenotype (63). Beyond enhancing TAMs homing to tumor sites via hypoxia-induced release of chemoattractant, hypoxia also fine-tunes the expression level of major histocompatibility complex II (MHC-II) on TAMs to induce "immune invisibility" of tumor-specific antigen, as MHC-II<sup>lo</sup> TAMs always populate hypoxic areas, whereas MHC-II<sup>hi</sup> TAMs prefer normoxic (64).

Compared to TME located in hypoxic niches (0.1%-5% O<sub>2</sub>) of solid tumors (64), the hypoxic level of the whole placenta is more moderate. Although the dramatic changes of O<sub>2</sub> levels in villous regions during the first (2%-3%) and second trimester (8%-10%) as the perfusion of the placenta is thoroughly established, O2 levels at decidua (5%-6%) maintain relatively constant during early gestation, which is comparable to the unpregnant uterus (39, 65). The sites located near the uterus arteries even bathe within a rich oxygen supply (up to 10%-14% O<sub>2</sub>) (39). Thus, hypoxia seems to make less impact on dMo phenotypic transformation compared to TAMs. Still and all, the proper extent of hypoxia is indispensable to a thriving placenta (5). In the early stage, hypoxia aids peripheral NK cells in homing to the uterus and promotes normal expansion and development of Hofbauer cell progenitors (66). In pregnancy complications where misfunctioned  $dM\phi$  can be seen, like PE and RPL, insufficient uterine spiral arteries remodeling and uteroplacental ischemia cause extreme hypoxia (67), suggesting the importance of a fine balance of oxygen concentration on  $dM\phi$ differentiation and function.

### 4.1.2 Lactic acid

Lactic acid (LA) is a metabolic product produced from glucose through glycolysis and the conversion of pyruvate by lactate dehydrogenase (LDH) under oxygen-deprivation conditions (68). In extremely hypoxia TME, the glucose restriction and low pH mediated by tumor-derived LA induce a poorly glycolytic M2-like profile along with the high levels of VEGF and ARG1, fatty acid oxidation (FAO) upregulation, and weak capacity for antigen presentation (69). It has also been reported that modulation of LA levels can redistribute M2-TAM subsets and upregulate PD-L1 to assist tumor immune escape via inducing T-cell apoptosis (70).

At the maternal-fetal interface, trophoblasts and DSCs are a major source of LA, especially during peri-implantation and decidualization (68). Unlike tumors, decidua-derived LA exerts a dual role in regulating macrophage polarization according to the concentration of oxygen and LA (71). Under normoxia, light trophoblast-derived LA promotes macrophage polarization to the M2 phenotype by upregulating mitochondrial oxidative phosphorylation (OXPHOS) and VEGF expression independent of hypoxia-inducible transcription factor- $1\alpha$  (HIF- $1\alpha$ ), whereas, upon hypoxia, vast trophoblast-derived LA promotes M1 phenotype polarization by upregulating glycolysis and iNOS mediated by HIF-1α/SRC/LDHA pathway (71). Clinically, the maternal-fetal microenvironment in patients with RPL is characterized by enhanced LA accumulation and exaggerated hypoxia, suggesting that glucose metabolism may be a therapeutic target by reining  $dM\phi$  phenotype and function (72).

### 4.1.3 Amino acid metabolism

Macrophages from both human and mouse TME are desperate for non-essential amino acids, thus depleting the local environment of amino acids necessary for optimal effector function of immune cells and thus subverting immunosurveillance (73). Among these, arginine metabolism is a key regulator (73). The hypoxic state of the tumor prefers the catabolism of Arg1 over iNOS, resulting in more M2-like polarization and elevated tumor-supporting factors into TME (74).

In addition, the concentration of tryptophan (Trp) and its byproduct Kynurenine (Kyn) in the local microenvironment also contribute to the polarization of macrophages (75). Trpconsuming enzyme Indoleamine 2,3-dioxygenase (IDO) is highly expressed on TAMs and  $dM\phi$  and causes a functional loss on NK and T cells (43, 44). In addition to the deprival of Trp, immunosuppressive Kyn interacts with the ligand-activated transcription factor aryl hydrocarbon receptor (AHR) and drives the generation of Tregs and tolerogenic myeloid cells and the upregulation of Anti-programmed cell death 1 (PD-1) in CD8<sup>+</sup> T cells, modulating M2-like macrophages activity (76). The blockade of IDO could reduce the recruitment and phagocytosis of macrophages, switching them to M1-like phenotype (76). Clinically, a remarkable decrease of IDO<sup>+</sup> dM $\phi$  is observed in patients with RPL; meanwhile, the treatment with IDO inhibitor, 1-methyl-D-tryptophan, during early pregnancy leads to obvious uterus inflammation and fetus loss (77).

### 4.1.4 Other metabolites

As recently reported, lysophosphatidic acid (LPA), an important intermediate in lipid metabolism produced by uterine epithelium, binds nuclear receptor peroxisome proliferatoractivated receptor gamma (PPAR $\gamma$ ) within dM $\phi$  and thus induces the transcription of M2-differentiation genes and facilitates the accumulation of decidua-resident macrophages, which favors trophoblast invasion and placental development (65). In line with, clinical observation found that a low expression of LPA receptor in villous decidua is associated with the risk of early RPL. In addition to this, the effect of LPA on TAM formation and polarization also has been reported in ovarian carcinoma (78).

In addition, because of the unique maternal–fetal interface components, like trophoblasts and DSCs, and pregnancy-related hormones, there may be some distinct mechanisms driving  $dM\phi$ toward tolerant phenotypes that TMEs do not involve. For example, fructose-1,6-bisphosphate (FBP), a crucial glucose metabolite during decidualization, is found to increase the secretion of IL-27 from DSCs, which induces M2-like  $dM\phi$  differentiation and suppresses their production of pro-inflammation molecules (79). Given the low level of FBP observed in decidua and plasma of patients with RPL (79), FBP supplementation could be a promising strategy to prevent fetal loss by targeting IL-27-triggered M2-like macrophages.

### 4.2 Epigenetic regulation

Via modifying the differentiation and functional programming of macrophages, epigenetic control provides a highly flexible mechanism for the activation and deactivation of transcription in macrophages in response to diverse stimuli in the TME, acting as a basis of plasticity. Firstly, DNA methylation. There are pieces of evidence that DNA methyltransferases (DNMTs) have a specific effect on the formation of macrophage phenotypes. Ubiquitin-like protein containing plant homeodomain (PHD) and RING finger domain 1 (UHRF1) maintains the DNA methylation status by recruiting DNMT1 (80). A recent study found that decreased UHRF1 in trophoblasts promotes the expression of proinflammatory IL-1β, which recruits monocytes to decidua and shifts them into the M1 phenotype, further exacerbating RPL (80). In cancer, DNA methylation is critical for the suppression of the expression of tumor suppressor genes, whereas the loss of DNA methylation mediated by removal of methyl groups leads to the overexpression of oncogenes. For example, the expression of demethylase ten-eleven translocation 2 (TET2) is increased in TAMs in the setting of melanoma in an IL-1R-MyD88 pathwaydependent way (81).

Secondly, histone methylation and acetylation also participate in the epigenetic regulation of macrophages. In tumors, histone deacetylase (HDAC) class I (HDAC3) is central in M2 activation, whereas HDAC class II (HDAC 4, 5, 6, and 7) mainly targets the M1 subgroup (44). Enhancer of zeste homolog 2 (EZH2) is the enzymatic subunit of polycomb repressive complex 2 (PRC2), which can catalyze trimethylates lysine 27 of histone H3 (H3K27me3) to mediate gene transcriptional silencing and chromatin compaction (82). It has been reported that the suppression of EZH2 in various malignant tumors like colorectal cancer (CRC) shifts TAMs toward the M1 phenotype via decreasing the H3K27me3 levels on the promoters of STAT3 (83), which is beneficial for tumor therapy, whereas a significantly decreased expression of EZH2 was found to be associated with adverse pregnancy outcomes such as RPL (84). Expectedly, inhibition of EZH2 in trophoblasts disturbed the phenotypic differentiation of macrophages, transforming M2-like into M1-like dM $\phi$ , via reducing the production of transforming growth factor–beta (TGF- $\beta$ ), IL-10, IL-6, IL-4, CXCL-16, and PD-L1 (84). Moreover, decreased HDAC8 expression downregulates the level of M2 marker genes like CD163 in dM $\phi$ s, which may disturb immune tolerance and induce pro-inflammatory response at the maternal–fetal interface and be relevant to RPL (85).

Moreover, noncoding RNAs are another modification to macrophages. Abundant evidence shows more than 100 micro-RNAs (miRNAs) impinge on macrophage polarization and function, forming a complex post-transcriptional regulating network (86). In tumors, for example, miRNA regulates TAM polarization to the M1 phenotype, via miR-155, miR-181, and miR-451, or to the M2 phenotype, via miR-99a, miR-146a, miR-125a, and miR-145-5p (73). Regarding dM $\varphi$ , miR-19a-3p and miR-130b-3p, and miR-10a-5p and miR-150–5p have been reported to inhibit the M1-marker gene expression and increase the production of M2-phenotype markers. Like TAMs, miR-155 derived from dM $\varphi$ also promotes M1 polarization while impairing M2 status (4), the alteration of which is associated with gestational complications such as miscarriage and PE.

# 4.3 Environmental growth factors, cytokines, and chemokines

The role of secreted or surface molecules in cancer progression and TAM phenotypic transformations remains inadequate, which can be explained by TAMs' plasticity shaped by the heterogeneity of source of certain molecules, tumor types, and stages, and systemic immunity of hosts. A certain molecule could exert diverse functions in regulating macrophage polarization according to specific microenvironments or subgroups of macrophages, such as Dectin 1 (87, 88) and CCL2/MCP-1 axis (89). Given various essential modulators involved in macrophage polarization in tumors and pregnancy, targeting can help switch or regulate macrophage polarization, thus probably facilitating potential immunotherapy. Table 2 summarizes the effect of current reported environmental factors on the polarization of macrophages from tumors and decidua (90–94, 96–106).

# 5 Macrophages function in immune tolerance

### 5.1 TAMs in tumor immune escape

### 5.1.1 TAMs interact with stromal cells

As the most abundant stromal cells in the TME, cancerassociated fibroblasts (CAFs) promote the recruitment of

Interference factor	Source	Influence on macrophage phenotype	Clinical relation	Reference
GM-CSF	Trophoblast and DSCs	M1 polarization	Tumor; pregnancy	(35, 90)
M-CSF	Trophoblast and DSCs	M2 polarization	Tumor; pregnancy	(35, 90)
VEGF	Trophoblast and DSCs	M2 polarization	Pregnancy	(91)
CCL2/MCP-1	DSCs; tumor cells	Balanced M1/M2 dMφ	LLC; pregnancy	(89, 92)
TGF-β	CA-MSCs	Immunosuppressive M2-like phenotype	Cancer	(93)
CX3CL1	CA-MSCs	Immunosuppressive M2-like phenotype	Cancer	(89, 93)
IL-10, IL-4, and IL-13	Trophoblast	M2-like subtype dMφ	Pregnancy	(50)
GAS6	DSCs; apoptotic cells	Immunosuppressive phenotype	RSA; cancer	(94, 95)
GdA	DSCs	dMφ-like phenotypes	Pregnancy	(96)
IL-33	Dying cell; macrophages	M2 polarization	RSA	(97)
IL-34	Trophoblast; DSC	Immunoregulatory phenotype	Pregnancy	(98)
CXCL16	Trophoblast	M2 polarization; lower IL-15 level	Pregnancy	(99)
IL-6	Tumor cells; trophoblast	M2 phenotype	TNBC; pregnancy	(100, 101)
Hyaluronan	Trophoblast	M2 phenotype	Pregnancy	(102)
RANKL/RANK	DSCs	M2 phenotype; higher adhesion molecules	Pregnancy	(103)
Progesterone	Ovary, placenta, and fetus	M2-like phenotype	Pregnancy	(104)
HCG	Placenta	M2 polarization	Pregnancy	(105)
PGE2	DSCs	M1 polarization	RSA	(106)
Dectin 1	Trophoblast	M2 polarization	GBM; PDAC	(87, 88)

TABLE 2 The various environmental factors that affect macrophage polarization and phenotype.

CA-MSC, cancer-associated mesenchymal stem cells; DSC, decidual stromal cell; GM-CSF, granulocyte-macrophage colony-stimulating factor; GdA, glycodelin-A; GBM, glioblastoma; HCG, human chorionic gonadotropin; LLC, Lewis lung cancer; PGE2, prostaglandin E2; PDAC, pancreatic cancer; RSA, recurrent spontaneous abortion; TNBC, triple-negative breast cancer.

monocytes and their transformation into M2 phenotype via secreting multiple regulatory molecules, such as IL-6, M-CSF, monocyte chemoattractant protein-1 (MCP-1, or CCL2), and stromal cell-derived factor 1 (SDF-1, or CXCL12) (107). More importantly, CAFs can motivate the immunosuppressive properties of TAMs, including higher PD-1 expression (45). Reciprocally, M2-like TAMs regulate CAF activation and progression and promote the EMT process, which consequently creates a positive loop between CAFs and TAMs, together promoting tumor evasion and survival (107).

# 5.1.2 TAMs regulate the function of other immune cells

TAMs can directly inhibit cytotoxic immune cell responses. Firstly, TAMs affect the phenotypic and functional differentiation of T cells via reversing the anti-tumor cytotoxic T lymphocyte (CTL) and CD4<sup>+</sup> Th1 cells to tolerant Th2 cells, Th17 cells, and Tregs cells in a manner dependent on inhibitory cytokines, such as TGF- $\beta$  and IL-10 (36). TAMs are also involved in tumor immune microenvironment (TIME) formation through their expression of immune checkpoints such as PD-L1/L2 and CD80/86 that bind to their receptors (PD-1/cytotoxic T- lymphocyte-associated protein 4 (CTLA-4)) that constitutively express on T cells, directly restraining the function of activated anti-tumor T cells (36). Subsequently, Th cells, in turn, remodel tumor TME partly by altering the polarization direction of macrophage, as Th2 cells secrete IL-4, IL-5, and IL-10 to induce more M2 macrophages (44). In addition, TAMs play a crucial intermediary role through both soluble mediator crosstalk and cell-to-cell contact to weaken tumorassociated NK cells. TAMs harboring inhibitory molecules like human leukocyte antigen-G (HLA-G) and human leukocyte antigen-E (HLA-E) interact with NK/T cells via coinhibitory signal molecules ILT2 and NKG2, respectively (108), which have been proved to neighbor TAMs physically in vivo and appear "disarmed." In addition to the direct contact, the secretion of IL-10 and TGF- $\beta$  by TAMs also educates NK cells toward an immunosuppressive state (109). As previously mentioned, TAMmediated metabolic activities, including the depletion of metabolites (such as L-arginine and Trp) also promote Treg activity and impair both CTL proliferation and interferon-gamma (IFN-y) production, thus engaging in the immune-suppressive TME (61). Moreover, IL-10 released by TAMs suppresses IL-12 expression by conventional DCs in tumors, resulting in a diminished CTL response (36).

TAMs also recruit immunosuppressive populations and build positive feedback loops to facilitate the expansion of each other and reinforce the suppressive TIME. TAMs have been shown to promote Treg cell recruitment to human carcinomas via CCL22 and CCL1, which are the respective ligands of CCR8 and CCR4 on Treg cells (23). In the stage of tumor progression, TAMs also secrete high levels of IDO1 and CXCL10 to attract surrounding Treg cells

10.3389/fimmu.2023.1333549

(36). In turn, Treg cells from TME provoke the production of IL-10, TGF- $\beta$ , and CD73 by TAMs and increase expression of negative coinhibitory molecules such as PD-L1 in an autocrine way, which later binds to effector T cells and restrains their anti-tumor activity (110).

TAMs also interact with unconventional T cells in tumor immunity, although not all lead to immune suppression. Invariant natural killer T cells (iNKT) were found to directly suppress M2-like TAMs and promote the survival of M1-like macrophages in a CD40/Fas-dependent way, thereby weakening prostate cancers in a transgenic mouse model (111). Moreover, TCR $\alpha\beta^+$ CD4<sup>-</sup>CD8<sup>-</sup>NK1.1<sup>-</sup> innate  $\alpha\beta$  T cells (i $\alpha\beta$ T) promote M1like proinflammatory reprogramming of macrophages in a CCR5dependent manner with enhanced antigen presentation and T-cell chemoattraction, this way improving CD4<sup>+</sup> and CD8<sup>+</sup> cytotoxic Tcell toward the tumor (112). As mentioned above,  $\gamma\delta$ -T cells can be stimulated by CCR2<sup>+</sup>TAMs to secrete IL-17, thus orchestrating a pro-metastatic response (113).

### 5.1.3 TAMs interact with tumor cell

It prevails that malignant tumor cells are encouraged to evade immune surveillance and "cool" the immune response (114). In this case, it is another key underlying mechanism for tumors avoiding immune surveillance that TAMs being hijacked and manipulated by tumor cells prefer to establish a tolerant TME. Initially, tumor cells highly express self-labels like CD47, B2M, and CD24, which can phosphorylate ITIMs and thus downregulate the phagocytic function of macrophages (63). Furthermore, tumor cells produce a plethora of cytokines and chemokines to bind to receptors on the surface of TAMs, handling their expression of relevant immunosuppressive genes. Manipulated TAMs thus gain secretion of various tumor-promoting factors including CCL18, IL-10, and TGF- $\beta$  while inhibiting the release of cytotoxic factors like IL-2, IL-12, TNF- $\alpha$ , and IFN- $\gamma$ , impeding the anti-tumor capacity of other immune components and prolong tumor survival (115).

# 5.1.4 Immune checkpoint molecules involved in TAMs

TAM is a major source of immune checkpoint molecules in the TME. In addition to the PD-1/PD-L1 and CTLA-4/CD86, TAMs can also induce immune escape along with other immune checkpoints, such as IDO/AHR, T-cell immunoglobulin and mucin domain protein 3 (Tim-3)/Galectin-9, and Fas/Fas-L (44). A recent study has found that Phosphatase and tensin homolog (PTEN)-deficient glioblastoma (GBM) cells secrete high levels of Galectin-9 via the AKT-GSK3β-IRF1 pathway, which drives M2 polarization by activating Tim-3 (116). Su et al. have revealed an unexpected immunosuppressive effect of macrophages that undergo antibody-dependent cellular phagocytosis by overexpressing PD-L1 and IDO, via inhibiting both NK-cell- and T-cell-mediated cytotoxicity (117). Clinically, the PD-1 expression on TAM of human cancers elevates over time with the stage of the disease, which downregulates TAM proliferation and activation, accelerating tumor cells' escape (118).

# 5.2 DM $\phi$ in immune tolerance at the maternal-fetal interface

### 5.2.1 Crosstalk between $dM\phi$ and decidua stroma

DSCs, the largest population at the decidua, are critical in maintaining a successful pregnancy not only by the production of hormones such as prolactin but also by shaping the local immunological microenvironment via cytokines and chemokines (30). Initially, DSCs promote the survival of CD14<sup>+</sup> blood-derived monocytes and induce them into the CD14<sup>bright</sup>CD163<sup>+</sup>CD209<sup>+</sup>CD86<sup>dim</sup> phenotype and trigger the expansion and phenotypic shift of dMp mainly via M-CSF and TGF- $\beta$  signaling (119). Subsequently, chemokines such as CCL2, secreted by DSCs, are a key regulator to maintain the fine M1/M2 mixed immune status (120). Growth arrest-specific factor 6 (GAS6) produced by DSCs also favors polarizing dMo toward an M2-like phenotype and induces cell proliferation (94). Compared with normal pregnancy, less GAS6 secretion by DSCs in early miscarriage suggests a critical role of dialog between DSCs and dMq in maintaining a healthy pregnancy (94). Similarly, TAM receptors (Tyro3, Axl, and MerTK), the cognate receptors of GAS6 on apoptotic cells, are also expressed on TAMs and skew polarization toward a pro-tumor M2-like phenotype, participating in immune homeostasis and tumor progression (95).

### 5.2.2 Crosstalk between $dM\phi$ and trophoblast

Analogous to tumor cells, trophoblasts secrete high levels of multiple signaling molecules to recruit and educate immune cells and establish a tolerant environment at the maternal-fetal interface (121). In general, trophoblast-derived factors, including IDO, TGFβ, IL-10, M-CSF, IL-34, IL-6, CXCL16, and PD-L1, play critical roles in shaping  $dM\phi$  into a fetus-friendly immune population (122, 123). The PD-1/PD-L1 axis modulates dMp polarization to a phenotype subset that modulates the maternal-fetal tolerance, which inhibition has been proven to associate with increased fetal loss (124). Intrinsically, trophoblasts tend to upregulate nonclassical HLA class Ib molecules HLA-E, HLA-F, and HLA-G, to provide inhibitory signals to a broad of immune cells, including NK cells, T cells, and myeloid cells via surface inhibitory receptors CD94/ NKG2A, KIR3DL1, immunoglobulin-like transcript 2 and 4 (ILT-2/4), and KIR2DL4 (125), among which, ILT-2/4 is especially enriched on macrophages. Lee et al. found that sHLA-G5 polarized macrophages toward the M2 phenotype, with higher phagocytic activity and increased expression of IDO, IL-6, and CXCL1, which impaired the proliferation and cytotoxic function of T cells and promoting trophoblast invasion (126). During normal pregnancy, the plasma level of sHLA-G5 increases in the first trimester and gradually declines as the pregnancy advances, and the low sHLA-G level in early gestation often relates to pregnancy complications, such as RPL, PE, and FGR (127).

In addition, trophoblast debris has been reported to modulate the expression of cytokines in macrophages, upregulating the expression of inhibitory molecules, like PD-L1, IDO, IL-1Ra, IL-6, and IL-10, while reducing the expression of costimulatory molecules, such as MHC-II molecules and intercellular adhesion molecule 1(ICAM-1) and pro-inflammatory cytokines including IL-8 and IL-1 $\beta$  (14). In turn, M $\phi$ -derived cytokine IL-33 acts as a critical factor for placental growth in early pregnancy, inducing the proliferation of trophoblasts (59).

# 5.2.3 Crosstalk between $dM\phi$ and other immune cells

In the first trimester of pregnancy,  $dM\phi$  exercises a strong influence on the regulation of immune tolerance in the maternalfetal interface (18). As the dominant APC,  $dM\phi$  plays a role in regulating T-cell activation (127). Two decades ago, the suppressive effect of  $dM\phi$  on the proliferation of effective T cells was first described in early pregnancy (128). DM $\phi$  also attracts and promotes the proliferation of Treg cells, which are in the control of maternal immune effector functions and in preventing fetal rejection (30).

Various immune checkpoints prevent an overreacted maternal inflammatory response to fetal antigens during pregnancy. Compared with peripheral blood monocytes, decidual macrophages suppress the production of Th1-type cytokine IFN-y by decidual T cells via PD-L1/PD-1 interactions (59). Tim-3+ dMp subset induces the exhaustion or apoptosis of cytotoxic T lymphocytes and favors Th2 and Treg bias in decidual CD4+T cells (129), and Tim-3 blockade has been proven to cause more fetal loss (90). Given pre-clinical cancer studies that found co-blockade of the PD-1/Tim-3 pathway endowed successful control over tumor growth, whether the joint effect of Tim-3 and PD-1 also make optimal protection to the fetus is attractive. Indeed, compared with the single intervention, the combination of PD-L1and Gal-9 fusion proteins are proven to exhibit obvious advantages in lowering the risk of miscarriage, trophoblast invasion, and fetal vascular development, indicating a synergistic effect of co-inhibitory pathways to better pregnancy outcomes (130).

As the most abundant leukocytes during pregnancy, dNK cells reside in close contact with decidual CD14<sup>+</sup> myelomonocytic cells, indicating a potential crosstalk between them. Indeed, TGF- $\beta$ secreted from dM $\phi$  restrains dNK killing toward fetal cells (131). Moreover, CXCL16 from trophoblasts decreases IL-15 secretion in dM $\phi$  (99). As an important cytokine regulating differentiation, maturation, and survival of NK cells, the reduction of IL-15 impairs NK cytotoxicity to establish immune tolerance (99). Following interaction with dNK cells, dM $\phi$  expresses IDO, which, in turn, induces the expansion of Tregs through TGF- $\beta$  production or CTLA-4-mediated interactions (132).

# 5.2.4 Intrinsic phagocytosis of $dM\phi$ helps inflammation prevention

Anyway, it cannot be ignored that as a type of professional APC the intrinsic capacity of macrophages is phagocytosis. Foreign pathogens and shedding trophoblast debris accumulated at the maternal-fetal interface become an important source of provoking inflammation and aggregating cytotoxic immune cells, which is likely to disturb the fetus-protecting homeostasis (127). In this case, either pathogen-associated molecular patterns of pathogens or damage-associated molecular patterns from apoptotic cells could help recruit decidual macrophages to the site of inflammation to prevent harmful inflammatory stress (133). Meanwhile, the pattern recognition receptors on the surface reverse these macrophages to an M1-like phenotype temporally, suggesting that they retain the role of canonical antimicrobial (127). Interestingly, how the manipulated dM $\phi$  by foreign agents make an effect on pregnancy outcomes, pro-M1 or pro-M2, is specifically related to distinct virulent strains of intrauterine pathogens, suggesting the complicated plasticity of dM $\phi$  answering external stimuli (134).

Above all, in the settings of tumor and pregnancy, macrophages, at the core of immunosuppressive cells and cytokines networks, are educated by tumor cells or trophoblasts to assist tumor immune evasion or fetus protection through their capacity of recruiting multiple immune components containing cells, cytokines, and inhibitory immune checkpoints. Meanwhile, dMqs also contribute to the defense against potential intrauterine infections, acting to suppress inflammatory stress that may disturb normal pregnancy. The similarities and distinctions of how TAMs and dMqs create an immunosuppressive environment are described in Figure 3.

### 6 Macrophages in angiogenesis

# 6.1 TAMs exert a crucial effect on tumor neo-angiogenesis

Like the placenta, a tumor as a developing organ requires blood vessels to transport oxygen and nutrients, which can exponentially accelerate the tumor's growth. The onset of neovascularization is orchestrated by various mechanisms, among which TAMs play a leading role in angiogenesis by promoting endothelial cell proliferation, migration, and survival, as well as ECM remodeling (135). During this process, hypoxia and LA regulate the effects of TAMs on the tumor-associated blood vessels via finely tuning the activation of TAMs and stimulating pro-angiogenic gene transcription in a HIF- $\alpha$ -dependent manner, such as VEGFA, TIE2, and neuropilin-1 (NRP1) (62, 136, 137). Endothelial cells can also exert an effect on TAMs to maintain their proangiogenic phenotype metabolically. Among these, PPAR-y is suggested to take part in the interaction between macrophage polarization and tumor angiogenesis (138), on which endothelial cells rely to promote the alternative activation of TAMs (139), and induces the transcriptional accumulation of HIF-2 $\alpha$  (140).

Moreover, M2-like TAMs induced by tolerant TIME promote tumor angiogenesis through the secretion of pro-angiogenic factors, including VEGFA, FGFs, CXCL8, CXCL12, WNT7B, and BV8, proinflammatory cytokines, namely, IL-1 $\beta$ , IL-6, and TNF, as well as pro-angiogenic proteases, including matrix metalloproteinases (MMPs) and cathepsins (141). Among those, VEGFA guarantees both angiogenic switch and abnormal vasculature featured by vascular permeability, thereby favoring tumor intravasation and distant metastasis (142). MMPs mobilize ECM-bound VEGFA and enable their binding to VEGF receptor 2, which is expressed on endothelial cells, triggering angiogenesis (143).



#### FIGURE 3

Crosstalk between macrophages and other immune components in the tumor immune microenvironment or at the maternal-fetal interface. Whether in the tumor or decidual immune microenvironment, macrophages establish close connections with other cell components to jointly create an environment of immune tolerance, which is conducive to the survival of rapidly expanding tumor cells and trophoblasts, respectively (5, 8). These macrophages either express high level of co-inhibitory molecules that suppress cytotoxicity of CTL, promote the generation of Treqs, and promote the conversion of Th1 to Th2 type, either produces various immune regulators, to inhibit the killing capacity of NK and effector T cells (36 67). In the tumor microenvironment, macrophages maintain closer hormonal communication with tumor cells and CAFs, forming positive feedback, which also exists between DSCs and decidual macrophage crosstalk (107, 119). During the rapid development of the fetus and its belongings, the rapid clearance of trophoblast debris by decidual macrophages also prevents potential inflammatory response. CAFs, cancer-associated fibroblast; DSCs, decidual stromal cells; tros, trophoblast; CTL, cytotoxic T cells; GAS6, growth arrest-specific factor 6.

The angiopoietins (Ang-1 to Ang-4) are a family of angiogenic growth factors primarily involved in vessel stabilization in the latter stages of angiogenesis, which signal through membrane-bound tyrosine kinase receptors (Tie-1 and Tie-2). Ang-2 as one of the best-characterized Ang family numbers has also been shown to play a role in the recruitment and education of TAMs (144). Hypoxic TME induces the expression of CXCL12 and Ang-2, which stimulates the recruitment and perivascular accumulation of CXCR4<sup>+</sup>TIE2<sup>+</sup> TAMs (145). Subsequently, the paracrine interaction between TAMs and endothelial cells that secrete Ang-2 facilitates tumor angiogenesis (136), offering close pro-angiogenic and tissue-remodeling support to sprouting and anastomotic blood vessels.

### 6.2 DM $\phi$ is essential in spiral artery remodeling

In a successful pregnancy, angiogenesis and spiral artery remodeling of the decidua are necessary to guarantee necessary blood flow and nutrition to the placenta and fetus, which requires the joint coordination of dNK cells, EVT,  $dM\phi$ , and vascular smooth muscle cells (VSMCs). Previous studies indicate that spiral arteriole remodeling in human pregnancy occurs in two phases (146). The first phase is trophoblast-independent and occurs within the spiral artery in the endometrium. In this phase, the perivascular VSMC is disrupted, the endothelium swells and loses its continuity, and VSMCs and endothelial cells are partially induced into apoptosis, which proves to be associated with the localized accumulation of dMq (145). Here, dMq infiltrating remodeling vessels assist in clearing apoptotic cells and producing MMP-7 and MMP-9, which facilitate perivascular ECM degradation and the vascular remodeling process (146). The second phase, namely, trophoblast-dependent, refers to the EVT migrating into the spiral arteries and forming a pseudo-endothelium with the loss of most VSMCs and endothelial cells. Here, vasoactive intestinal peptide and pregnancy-specific glycoproteins, produced by trophoblasts, disrupt endothelial cell networks and promote the growth of nascent blood vessels via an effect on macrophages (147). In both stages, the decidual spiral arteriole endothelial cells express and secrete CCL14 to induce chemotaxis of dNK and dMq (148). Similar to tumor angiogenesis, Ang-1, Ang-2, and Tie-2, are identified to present in the developing placenta as well as EVT and endothelial cells and VSMCs of the uterine spiral arteries, resulting in the organized breakdown and remodeling of the vasculature (144). However, the exact role of Angs in dMp recruitment or phenotypes induction has not been explored.

Clinically, abnormal angiogenesis is an important factor leading to PE, in which dysfunctional  $dM\phi$  could accelerate disease progression (149). Similar to TAMs, dMq can regulate vascular remodeling by secreting abundant angiogenic regulators, such as VEGFA (57), placental growth factor (PlGF), and their receptors FMS-like tyrosine kinase (Flt-1) (150), among others. In pregnant

mice, uterine macrophages express elevated levels of iNOS and VEGF during the implantation window compared to that of non-pregnant women (151).

It can be perceived that both TAM and  $dM\phi$ 's regulation of angiogenesis is largely based on VEGF. Compared with studies on TAMs, studies on  $dM\phi$  are limited to the mice models (152, 153), which cannot represent authentic  $dM\phi$ 's dynamic change along with the human pregnancy, and, due to ethical issues and technical difficulty, it is often infeasible to acquire abundant clinical samples. However, emerging and promising technologies including organoids, microfluidics, and organ-on-a-chip are paving the way for a deeper investigation of vasculogenesis involved in placental development and tumorigenesis (154, 155). The effect of TAM and  $dM\phi$  on vasculature remodeling is illustrated in Figure 4.

### 7 Conclusion and perspectives

The build of solid tumors is a complex multistep process requiring the interplay between different cell types and a surrounding TME to promote survival. In addition, the establishment of pregnancy adopted analogous strategies to embrace the implantation and growth of a semi-allogeneic fetus as follows (5, 8). Firstly, in terms of immunomodulation, immune tolerance is facilitated via the lack of classical MHC-I expression, presenting co-inhibitory signals and producing immunosuppressive cytokines and regulatory metabolites in both settings (156). Secondly, regarding metabolism and energy taking, both tumor and trophoblast proliferate at low oxygen levels and keep invasion toward higher oxygen levels, meanwhile releasing lactate as an energy source and signaling molecule (5). Thirdly, as mentioned before, several correlates have been drawn between tumor and pregnancy establishment in terms of angiogenesis and cell invasion, such as the upregulated level of Ang-2 and VEGFA in both settings. To maintain the growing demand for oxygen and nutrients, both tumor and trophoblast strive for the host's blood supply via the releasing of pro-angiogenic factors and attracting various leukocytes including macrophages and NK cells (5). The above explains the similarities of macrophage phenotypes and functions in both contexts as this review has illustrated earlier.

However, there is ample evidence indicating that metabolic, proliferative, migrative, and invasive states of trophoblasts are supervised tightly and finely, in contrast to tumor cells (6, 157). For example, ordered spiral artery remodeling and confused vasculogenic mimicry as well as the well-organized invasion endpoint and distant metastases are present in the decidua and the tumor site, respectively (158). Different from the solitary tumor, ahead of embryo implantation and trophoblast presenting, the human uterus has braced itself for the fetus and finished decidualization orchestrated by various sex hormones, generating an essential nourishing and supporting population as DSCs (159). They stand opposite the trophoblast at the decidua and play an active role in directing a moderate trophoblast invasion via the secretion of a variety of different cytokines and growth factors rather than passively receiving the intrusion without any restrains, including leukemia inhibitory factor, IL-6, IL-11, IL-15, CXCL-10, and granulocyte-macrophage colony-stimulating factor (GM-CSF) (110). Also, fluctuated hormone levels at the decidua during pregnancy, especially estrogen and progesterone, affect macrophages' phenotype and response (160). In addition, as



#### FIGURE 4

The tumor neo-angiogenesis and spiral artery remodeling during early pregnancy mediated by TAMs and dMφ. In response to distinct signals, such as hypoxia, acidosis, and high lactate, TAMs tend to accumulate in hypoxic areas of tumors and increase the secretion of a series of pro-angiogenic factors and angiogenesis-modulating factors, via upregulating HIFs (61, 145). Similarly, decidual macrophages support spiral artery remodeling by producing angiogenic factors, especially VEGF and MMPs; moreover, they maintain a homeostatic tissue environment by for instance phagocytosing apoptotic cells and debris (39). ANGPT2, angiopoietin 2; HIF, hypoxia-inducible transcription factors; MMPs, matrix metalloproteinase; TME, tumor microenvironment; VSMC, vascular smooth muscle cells; VEGFA, vascular endothelial growth factor–A.

mentioned earlier, owing to some fine-tuned structures like EVT plugs and disappearing lymphatics, placenta establishment and immune response can be modulated by the mother in a timely way, although this initiative may be taken away during tumor growth (25). Moreover, what is easily ignored is the distinction between fetus/tumor-specific antigen recognition in pregnancy and tumors. In pregnancy, it has been reported that maternal T cells become aware of the fetal allograft exclusively through "indirect" antigen presentation as the trophoblasts also express paternal MHC molecules while infiltrating T cells in tumor sites that recognize tumor-specific antigen via both "indirect" antigen presentation (161, 162). In other words, decidual macrophages

presentation (eer, roc), in only works, accutate interopring present different MHC molecules from the fetus, whereas the TAMs and tumors present the same, suggesting the slightly different functions of macrophages in terms of antigen-specific presentation and response in these two settings. In this review, we illustrate the heterogeneity and plasticity of

TAMs and dMq and compare the maternal-fetal immune tolerance with tumor immune escape and the placental vascular remodeling with tumor neovascularization. The remarkably high plasticity and diversity allow for performing a plethora of functions with phenotype governed by the local microenvironmental milieu, which can be corroborated by both TAMs and  $dM\phi$ . In general, macrophages at both the tumor sites and maternal-fetal interfaces are inclined to adopt a homeostatic M2like phenotype, resulting in negative outcomes (tumor progression and spreading) and positive outcomes (fetus development and pregnancy maintenance), respectively. To suppress cancer, we strive to reverse the state of exhausted immune status and enhance attack by reducing antiinflammatory TAMs and enabling pro-inflammatory TAMs. During pregnancy, dMq provides an immunotolerant environment for the semi-allogeneic fetus to be carried in the uterus. Therefore, the switching of macrophage polarity could be a potential therapeutic target to treat pregnancy complications as is predicted for cancer treatment and vice versa. The signaling pathways and chemokine/ cytokines mentioned earlier make it tantalizing to speculate that the same regulator could impact macrophage biology to aid both physiological and pathological processes. It urges researchers to make a more precise understanding of molecular mechanisms involved in macrophage function, based on animal models and clinical data, which will be crucial for developing macrophage-targeted therapeutics.

Furthermore, in addition to simple tumors or pregnancy, some particular cases should be taken into consideration as well. Decidual macrophages secrete both pro-inflammatory and anti-inflammatory cytokines, which serves as an attempt to maintain fetal tolerance while responding to microbial challenge. Take *Listeria monocytogenes*: a Gram-positive bacterium initially infiltrates placental tissues and leads to intrauterine infection, which can cause miscarriage or preterm birth (163). Upon *Listeria* infection during pregnancy, the signatures for pro-inflammatory pathways, as well as cytokine production, are strongly upregulated in dMqs (163), indicating that dangerous stimuli may disturb the balance of M1/M2 mix, reverse the dominated homeostatic dMqs, and lead to adverse outcomes. Moreover, regarding patients with cancer with pregnancy, most studies focus on the epidemiology and individual management and little discuss the dynamic phenotype and function of immune cells at decidua and concurrent tumor sites (164). In the future, whether the plasticity and polarization of local macrophages can be mutually affected by two abnormal immune statuses requires further investigations, which is quite meaningful for patients with cancer who wish to become or are pregnant.

### Author contributions

TY: Visualization, Writing – original draft, Writing – review & editing. XL: Investigation, Writing – original draft. YL: Conceptualization, Supervision, Writing – review & editing. XZ: Conceptualization, Supervision, Writing – review & editing. LL: Conceptualization, Supervision, Writing – review & editing. MD: Conceptualization, Supervision, Validation, Writing – review & editing.

### Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the National Natural Science Foundation of China [82271713, 31970859, 81630036], the National Key R&D Program of China [2021YFE0206500], the international cooperation project between Macau and Shanghai (20410760300), the funding of Innovative research team of high-level local universities in Shanghai and a key laboratory program of the Education Commission of Shanghai Municipality (ZDSYS14005), Shanghai Municipal Health and Family Planning Commission 20204Y0403, Shanghai Sailing Program(1YF1403700).

## Acknowledgments

Figures 1, 3, 4 in this review were created with BioRender.com.

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.Abbreviation

## References

1. Houser BL. Decidual macrophages and their roles at the maternal-fetal interface. *Yale J Biol Med* (2012) 85(1):105–18.

2. Lin Y, Xu J, Lan H. Tumor-associated macrophages in tumor metastasis: Biological roles and clinical therapeutic applications. J Hematol Oncol (2019) 12 (1):1-16. doi: 10.1186/s13045-019-0760-3

3. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* (2010) 141(1):39–51. doi: 10.1016/j.cell.2010.03.014

4. Yao Y, Hao F, Tang LC, Xu XH, Jin L. Downregulation of HDAC8 expression decreases CD163 levels and promotes the apoptosis of macrophages by activating the ERK signaling pathway in recurrent spontaneous miscarriage. *Mol Hum Reprod* (2020) 26(7):521–31. doi: 10.1093/molehr/gaaa035

5. Krstic J, Deutsch A, Fuchs J, Gauster M, Sparovec TG, Hiden U, et al. (Dis) similarities between the decidual and tumor microenvironment. *Biomedicines* (2022) 10(5):1–35. doi: 10.3390/biomedicines10051065

6. Muralidhara P, Sood V, Vinayak Ashok V, Bansal K. Pregnancy and tumour: the parallels and differences in regulatory T cells. *Front Immunol* (2022) 13:866937. doi: 10.3389/fimmu.2022.866937

7. Soeters PB, Grimble RF. The conditional role of inflammation in pregnancy and cancer. *Clin Nutr* (2013) 32(3):460–5. doi: 10.1016/j.clnu.2012.07.010

8. Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. *Nat Rev Immunol [Internet]* (2017) 17(8):469–82. doi: 10.1038/nri.2017.64

9. Guo C, Cai P, Jin L, Sha Q, Yu Q, Zhang W, et al. Single-cell profiling of the human decidual immune microenvironment in patients with recurrent pregnancy loss. *Cell Discov* (2021) 7(1):1–15. doi: 10.1038/s41421-020-00236-z

10. Wu K, Lin K, Li X, Yuan X, Xu P, Ni P, et al. Redefining tumor-associated macrophage subpopulations and functions in the tumor microenvironment. *Front Immunol* (2020) 11:1731. doi: 10.3389/fimmu.2020.01731

11. Hoeffel G, Chen J, Lavin Y, Low D, Almeida FF, See P, et al. C-myb+ Erythromyeloid progenitor-derived fetal monocytes give rise to adult tissue-resident macrophages. *Immunity* (2015) 42(4):665–78. doi: 10.1016/j.immuni.2015.03.011

12. Io S, Kabata M, Iemura Y, Semi K, Morone N, Minagawa A, et al. Capturing human trophoblast development with naive pluripotent stem cells in vitro. *Cell Stem Cell* (2021) 28(6):1023–39.e13. doi: 10.1016/j.stem.2021.03.013

13. Haider S, Meinhardt G, Saleh L, Kunihs V, Gamperl M, Kaindl U, et al. Self-renewing trophoblast organoids recapitulate the developmental program. *Stem Cell Rep* (2018) 11:1–15. doi: 10.1016/j.stemcr.2018.07.004

14. Ding J, Zhang Y, Cai X, Diao L, Yang C, Yang J. Crosstalk between trophoblast and macrophage at the maternal-fetal interface: current status and future perspectives. *Front Immunol* (2021) 12:1–10. doi: 10.3389/fimmu.2021.758281

15. Abbas Y, Turco MY, Burton GJ, Moffett A. Investigation of human trophoblast invasion in vitro. *Hum Reprod Update* (2020) 26(4):501–13. doi: 10.1093/humupd/dmaa017

16. Turco MY, Moffett A. Development of the human placenta. (2019) 146(22): dev163428. doi: 10.1242/dev.163428

17. Pollheimer J, Vondra S, Baltayeva J, Beristain AG, Knöfler M. Regulation of placental extravillous trophoblasts by the maternal uterine environment. *Front Immunol* (2018) 9:1–18s. doi: 10.3389/fimmu.2018.02597

18. Li X, Zhou J, Fang M, Yu B. Pregnancy immune tolerance at the maternal-fetal interface. Int Rev Immunol (2020) 39(6):247-63. doi: 10.1080/08830185.2020.1777292

19. Murata H, Tanaka S, Okada H. Immune tolerance of the human decidua. *J Clin Med* (2021) 10(2):1–16. doi: 10.3390/jcm10020351

20. Rodríguez-Cerdeira C, Gregorio MC, López-Barcenas A, Sánchez-Blanco E, Sánchez-Blanco B, Fabbrocini G, et al. Advances in immunotherapy for melanoma: A comprehensive review. *Mediators Inflamm* (2017) 3264217. doi: 10.1155/2017/3264217

21. Wagner J, Rapsomaniki MA, Chevrier S, Anzeneder T, Langwieder C, Dykgers A, et al. A single-cell atlas of the tumor and immune ecosystem of human breast cancer. *Cell* (2019) 177(5):1330–45.e18. doi: 10.1016/j.cell.2019.03.005

22. Ma RY, Black A, Qian BZ. Macrophage diversity in cancer revisited in the era of single-cell omics. *Trends Immunol* (2022) 43(7):546–63. doi: 10.1016/j.it.2022.04.008

23. Pan Y, Yu Y, Wang X, Zhang T. Tumor-associated macrophages in tumor immunity. Front Immunol (2020) 11. doi: 10.3389/fimmu.2020.583084

24. Collins MK, Tay CS, Erlebacher A. Dendritic cell entrapment within the pregnant uterus inhibits immune surveillance of the maternal/fetal interface in mice. *J Clin Invest* (2009) 119(7):2062–73. doi: 10.1172/JCI38714

25. Volchek M, Girling JE, Lash GE, Cann L, Kumar B, Robson SC, et al. Lymphatics in the human endometrium disappear during decidualization. *Hum Reprod* (2010) 25 (10):2455–64. doi: 10.1093/humrep/deq224

26. Chávez-Galán L, Olleros ML, Vesin D, Garcia I. Much more than M1 and M2 macrophages, there are also CD169+ and TCR+ macrophages. *Front Immunol* (2015) 6:1–15. doi: 10.3389/fimmu.2015.00263

27. Wu Y, Hirschi KK. Tissue-resident macrophage development and function. Front Cell Dev Biol (2021) 8:1-7. doi: 10.3389/fcell.2020.617879

28. Locati M, Curtale G, Mantovani A. Diversity, mechanisms, and significance of macrophage plasticity. *Annu Rev Pathol Mech Dis* (2020) 15:123–47. doi: 10.1146/annurev-pathmechdis-012418-012718

29. Bian Z, Gong Y, Huang T, Lee CZW, Bian L, Bai Z, et al. Deciphering human macrophage development at single-cell resolution. *Nature* (2020) 582(7813):571–6. doi: 10.1038/s41586-020-2316-7

30. Sun F, Wang S, Du M. Functional regulation of decidual macrophages during pregnancy. J Reprod Immunol (2021) 143:103264. doi: 10.1016/j.jri.2020.103264

31. Hogg C, Horne AW, Greaves E. Endometriosis-associated macrophages: origin, phenotype, and function. *Front Endocrinol (Lausanne)* (2020) 11:1–15. doi: 10.3389/ fendo.2020.00007

32. Tugal D, Liao X, Jain MK. Transcriptional control of macrophage polarization. *Arterioscler Thromb Vasc Biol* (2013) 33(6):1135-44. doi: 10.1161/ATVBAHA.113.301453

33. Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaeili SA, Mardani F, et al. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol* (2018) 233(9):6425–40. doi: 10.1002/jcp.26429

34. Boutilier AJ, Elsawa SF. Macrophage Polarization States in the Tumor Microenvironment. Int J Mol Sci (2021) 22(13):6995. doi: 10.3390/ijms22136995

35. Hourani T, Holden JA, Li W, Lenzo JC, Hadjigol S, O'Brien-Simpson NM. Tumor associated macrophages: origin, recruitment, phenotypic diversity, and targeting. *Front Oncol* (2021) 11:1–20. doi: 10.3389/fonc.2021.788365

36. Qiu Y, Chen T, Hu R, Zhu R, Li C, Ruan Y, et al. Next frontier in tumor immunotherapy: macrophage-mediated immune evasion. *biomark Res* (2021) 9(1):1–19. doi: 10.1186/s40364-021-00327-3

37. Kielbassa K, Vegna S, Ramirez C, Akkari L. Understanding the origin and diversity of macrophages to tailor their targeting in solid cancers. *Front Immunol* (2019) 10:2215. doi: 10.3389/fimmu.2019.02215

38. Laviron M, Boissonnas A. Ontogeny of tumor-associated macrophages. Front Immunol (2019) 10:1799. doi: 10.3389/fimmu.2019.01799

39. Zhao H, Wong RJ, Stevenson DK. The impact of hypoxia in early pregnancy on placental cells. *Int J Mol Sci* (2021) 22(18):9675. doi: 10.3390/ijms22189675

40. Mezouar S, Katsogiannou M, Ben Amara A, Bretelle F, Mege JL. Placental macrophages: Origin, heterogeneity, function and role in pregnancy-associated. (2021) 103:94–103. doi: 10.1016/j.placenta.2020.10.017

41. Tagliani E, Shi C, Nancy P, Tay CS, Pamer EG, Erlebacher A. Coordinate regulation of tissue macrophage and dendritic cell population dynamics by CSF-1. *J Exp Med* (2011) 208(9):1901–16. doi: 10.1084/jem.20110866

42. Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* (2004) 25(12):677–86. doi: 10.1016/j.it.2004.09.015

43. Wang S, Liu G, Li Y, Pan Y. Metabolic reprogramming induces macrophage polarization in the tumor microenvironment. *Front Immunol* (2022) 13:1–14. doi: 10.3389/fimmu.2022.840029

44. Chamseddine AN, Assi T, Mir O, Chouaib S. Modulating tumor-associated macrophages to enhance the efficacy of immune checkpoint inhibitors: A TAM-pting approach. *Pharmacol Ther* (2022) 231:107986. doi: 10.1016/j.pharmthera.2021.107986

45. Song H, Lou C, Ma J, Gong Q, Tian Z, You Y, et al. Single-cell transcriptome analysis reveals changes of tumor immune microenvironment in oral squamous cell carcinoma after chemotherapy. *Front Cell Dev Biol* (2022) 10(June):1–14. doi: 10.3389/ fcell.2022.914120

46. Zhang L, Li Z, Skrzypczynska KM, Fang Q, Zhang W, O'Brien SA, et al. Singlecell analyses inform mechanisms of myeloid-targeted therapies in colon cancer. *Cell* (2020) 181(2):442–59.e29. doi: 10.1016/j.cell.2020.03.048

47. Zhang Q, He Y, Luo N, Patel SJ, Han Y, Gao R, et al. Landscape and dynamics of single immune cells in hepatocellular carcinoma. *Cell [Internet]* (2019) 179(4):829–845.e20. doi: 10.1016/j.cell.2019.10.003

48. Lavin Y, Kobayashi S, Leader A, Amir E ad D, Elefant N, Bigenwald C, et al. Innate immune landscape in early lung adenocarcinoma by paired single-cell analyses. Cell (2017) 169(4):750–65.e17. doi: 10.1016/j.cell.2017.04.014

49. Houser BL, Tilburgs T, Hill J, Nicotra ML, Strominger JL. Two unique human decidual macrophage populations. *J Immunol* (2011) 186(4):2633-42. doi: 10.4049/jimmunol.1003153

50. Svensson J, Jenmalm MC, Matussek A, Geffers R, Berg G, Ernerudh J. Macrophages at the fetal-maternal interface express markers of alternative activation and are induced by M-CSF and IL-10. *J Immunol* (2011) 187(7):3671–82. doi: 10.4049/jimmunol.1100130

51. Vento-tormo R, Efremova M, Botting RA, Turco MY, Vento-tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal – fetal interface in humans. *Nature* (2018) 563(7731):347–53. doi: 10.1038/s41586-018-0698-6

52. Jiang X, Du MR, Li M, Wang H. Three macrophage subsets are identified in the uterus during early human pregnancy. *Cell Mol Immunol* (2018) 15(12):1027–37. doi: 10.1038/s41423-018-0008-0

53. Chen P, Zhou L, Chen J, Lu Y, Cao C, Lv S, et al. The immune atlas of human deciduas with unexplained recurrent pregnancy loss. *Front Immunol* (2021) 12 (June):1–16. doi: 10.3389/fimmu.2021.689019

54. Wang F, Jia W, Fan M, Shao X, Li Z, Liu Y, et al. Single-cell immune landscape of human recurrent miscarriage. *Genomics Proteomics Bioinforma* (2021) 19(2):208–22. doi: 10.1016/j.gpb.2020.11.002

55. Mor G, Cardenas I. The immune system in pregnancy: A unique complexity. *Am J Reprod Immunol* (2010) 63(6):425–33. doi: 10.1111/j.1600-0897.2010.00836.x

56. Jaiswal MK, Mallers TM, Larsen B, Kwak-Kim J, Chaouat G, Gilman-Sachs A, et al. V-ATPase upregulation during early pregnancy: A possible link to establishment of an inflammatory response during preimplantation period of pregnancy. *Reproduction* (2012) 143(5):713–25. doi: 10.1530/REP-12-0036

57. Yao Y, Xu XH, Jin L. Macrophage polarization in physiological and pathological pregnancy. *Front Immunol* (2019) 10:1–13. doi: 10.3389/fimmu.2019.00792

58. Zha Y, Liu H, Lin X, Yu L, Gao P, Li Y, et al. Immune deviation in the decidua during term and preterm labor. *Front Immunol* (2022) 13:1–15. doi: 10.3389/fmmu.2022.877314

59. Lemon SM, Walker CM. Role of macrophages in pregnancy and related complications. *Arch Immunol Ther Exp* (2019) 67(5):295–309. doi: 10.1007/s00005-019-00552-7

60. Jiang X, Wang H. Macrophage subsets at the maternal-fetal interface. Cell Mol Immunol (2020) 17(8):889–91. doi: 10.1038/s41423-020-0435-6

61. Vitale I, Manic G, Coussens LM, Kroemer G, Galluzzi L. Macrophages and metabolism in the tumor microenvironment. *Cell Metab* (2019) 30(1):36–50. doi: 10.1016/j.cmet.2019.06.001

62. Casazza A, Laoui D, Wenes M, Rizzolio S, Bassani N, Mambretti M, et al. Impeding macrophage entry into hypoxic tumor areas by sema3A/nrp1 signaling blockade inhibits angiogenesis and restores antitumor immunity. *Cancer Cell* (2013) 24 (6):695–709. doi: 10.1016/j.ccr.2013.11.007

63. Chen D, Zhang X, Li Z, Zhu B. Metabolic regulatory crosstalk between tumor microenvironment and tumor-associated macrophages. *Theranostics* (2020) 11 (3):1016–30. doi: 10.7150/thno.51777

64. Henze AT, Mazzone M. The impact of hypoxia on tumor-associated macrophages. J Clin Invest (2016) 126(10):3672-9. doi: 10.1172/JCI84427

65. Yang HL, Lai ZZ, Shi JW, Zhou WJ, Mei J, Ye JF, et al. A defective lysophosphatidic acid-autophagy axis increases miscarriage risk by restricting decidual macrophage residence. *Autophagy* (2022) 00(00):1–22. doi: 10.1080/15548627.2022.2039000

66. Garcés-Lázaro I, Kotzur R, Cerwenka A, Mandelboim O. NK cells under hypoxia: the two faces of vascularization in tumor and pregnancy. *Front Immunol* (2022) 13:1–13. doi: 10.3389/fimmu.2022.924775

67. Jena MK, Nayak N, Chen K, Nayak NR. Role of macrophages in pregnancy and related complications. *Arch Immunol Ther Exp (Warsz)* (2019) 67(5):295–309. doi: 10.1007/s00005-019-00552-7

68. Ma LN, Huang XB, Muyayalo KP, Mor G, Liao AH. Lactic acid: A novel signaling molecule in early pregnancy? *Front Immunol* (2020) 11(February):1–10. doi: 10.3389/fimmu.2020.00279

69. Colegio OR, Chu NQ, Szabo AL, Chu T, Rhebergen AM, Jairam V, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature* (2014) 513(7519):559–63. doi: 10.1038/nature13490

70. Shan T, Chen S, Chen X, Wu T, Yang Y, Li S, et al. M2-TAM subsets altered by lactic acid promote T-cell apoptosis through the PD-L1/PD-1 pathway. *Oncol Rep* (2020) 44(5):1885–94. doi: 10.3892/or.2020.7767

71. Gao L, Xu QH, Ma LN, Luo J, Muyayalo KP, Wang LL, et al. Trophoblastderived lactic acid orchestrates decidual macrophage differentiation *via* SRC/LDHA signaling in early pregnancy. *Int J Biol Sci* (2022) 18(2):599–616. doi: 10.7150/ ijbs.67816

72. Zhang T, Shen HH, Qin XY, Li MQ. The metabolic characteristic of decidual immune cells and their unique properties in pregnancy loss\*. *Immunol Rev* (2022) 308 (1):168–86. doi: 10.1111/imr.13085

73. Larionova I, Kazakova E, Patysheva M, Kzhyshkowska J. Transcriptional, epigenetic and metabolic programming of tumor-associated macrophages irina. *Cancers (Basel)* (2020) 12(1411):1-41. doi: 10.3390/cancers12061411

74. Geeraerts X, Fernández-Garcia J, Hartmann FJ, de Goede KE, Martens L, Elkrim Y, et al. Macrophages are metabolically heterogeneous within the tumor microenvironment. *Cell Rep* (2021) 37(13):110171. doi: 10.1016/j.celrep.2021.110171

75. Zhai L, Bell A, Ladomersky E, Lauing KL, Bollu L, Sosman JA, et al. Immunosuppressive IDO in cancer: mechanisms of action, animal models, and targeting strategies. *Front Immunol* (2020) 11:1–15. doi: 10.3389/fimmu.2020.01185

76. Campesato LF, Budhu S, Tchaicha J, Weng CH, Gigoux M, Cohen IJ, et al. Blockade of the AHR restricts a Treg-macrophage suppressive axis induced by L-Kynurenine. *Nat Commun* (2020) 11(1):1–11. doi: 10.1038/s41467-020-17750-z

77. Huang HL, Yang HL, Lai ZZ, Yang SL, Li MQ, Li DJ. Decidual IDO+ macrophage promotes the proliferation and restricts the apoptosis of trophoblasts. *J Reprod Immunol* (2021) 148:103364. doi: 10.1016/j.jri.2021.103364

78. Feng Y, Xiao M, Zhang Z, Cui R, Jiang X, Wang S, et al. Potential interaction between lysophosphatidic acid and tumor-associated macrophages in ovarian

carcinoma. J Inflammation (United Kingdom) (2020) 17(1):1–5. doi: 10.1186/s12950-020-00254-4

79. Zhou W, Yang H, Mei J, Chang K, Lu H, Lai Z, et al. Fructose-1, 6-bisphosphate prevents pregnancy loss by inducing decidual COX-2 + macrophage differentiation. *Sci Adv* (2022) 8(8):eabj2488. doi: 10.1126/sciadv.abj2488

80. Liu H, Wang LL, Xu QH, Wang J, Zhang YJ, Luo J, et al. UHRF1 shapes both the trophoblast invasion and decidual macrophage differentiation in early pregnancy. FASEB J (2022) 36(4):1–17. doi: 10.1096/fj.202101647RR

81. Larionova I, Kazakova E, Patysheva M, Kzhyshkowska J. Transcriptional, epigenetic and metabolic programming of tumor-associated macrophages. *Cancers* (*Basel*) (2020) 12(6):1411. doi: 10.3390/cancers12061411

82. Nancy P, Siewiera J, Rizzuto G, Tagliani E, Osokine I, Manandhar P, et al. H3K27me3 dynamics dictate evolving uterine states in pregnancy and parturition. *J Clin Invest* (2018) 128(1):233–47. doi: 10.1172/JCI95937

83. Li C, Song J, Guo Z, Gong Y, Zhang T, Huang J, et al. EZH2 inhibitors suppress colorectal cancer by regulating macrophage polarization in the tumor microenvironment. *Front Immunol* (2022) 13:1–15. doi: 10.3389/fimmu.2022.857808

84. Shang Y, Wu S, Li SJ, Qin X, Chen J, Ding J, et al. Downregulation of EZH2 in trophoblasts induces decidual M1 macrophage polarization: a potential cause of recurrent spontaneous abortion. *Reprod Sci* (2022) (10):2820–8. doi: 10.1007/s43032-021-00790-1

85. Wang LL, Li ZH, Wang H, Kwak-Kim J, Liao AH. Cutting edge: the regulatory mechanisms of macrophage polarization and function during pregnancy. *J Reprod Immunol* (2022) 151:103627. doi: 10.1016/j.jri.2022.103627

86. Wang L, Deng Z, Yang J, Zhao Y, Zhou L, Diao L, et al. Epigenetic and transcriptomic characterization of maternal-fetal interface in patients with recurrent miscarriage via an integrated multi-omics approach. *J Reprod Immunol* (2022) 154:103754. doi: 10.1016/j.jri.2022.103754

87. Daley D, Mani VR, Mohan N, Akkad N, Ochi A, Heindel DW, et al. Dectin 1 activation on macrophages by galectin 9 promotes pancreatic carcinoma and peritumoral immune tolerance. *Nat Med* (2017) 23(5):556–67. doi: 10.1038/nm.4314

88. Tiwari RK, Singh S, Gupta CL, Pandey P, Singh VK, Sayyed U, et al. Repolarization of glioblastoma macrophage cells using non-agonistic Dectin-1 ligand encapsulating TLR-9 agonist: plausible role in regenerative medicine against brain tumor. *Int J Neurosci* (2021) 131(6):591–8. doi: 10.1080/00207454.2020.1750393

89. Schmall A, Al-Tamari HM, Herold S, Kampschulte M, Weigert A, Wietelmann A, et al. Macrophage and cancer cell cross-talk *via* CCR2 and CX3CR1 is a fundamental mechanism driving lung cancer. *Am J Respir Crit Care Med* (2014) 191(4):437–47. doi: 10.1164/rccm.201406-1137OC

90. Zhang YH, He M, Wang Y, Liao AH. Modulators of the balance between M1 and M2 macrophages during pregnancy. *Front Immunol* (2017) 8:1–12. doi: 10.3389/fmmu.2017.00120

91. Wheeler KC, Jena MK, Pradhan BS, Nayak N, Das S, Hsu C, et al. VEGF may contribute to macrophage recruitment and M2 polarization in the decidua. *PLoS One* (2018) 13(1):e0191040. doi: 10.1371/journal.pone.0191040

92. Lin Z, Shi JL, Chen M, Zheng ZM, Li MQ, Shao J. CCL2: An important cytokine in normal and pathological pregnancies: A review. *Front Immunol* (2023) 13:40–5. doi: 10.3389/fimmu.2022.1053457

93. Cascio S, Chandler C, Zhang L, Sinno S, Gao B, Onkar S, et al. Cancer-associated MSC drive tumor immune exclusion and resistance to immunotherapy, which can be overcome by Hedgehog inhibition. *Sci Adv* (2021) 7(46):eabi5790. doi: 10.1126/sciadv.abi5790

94. Dai JC, Yang JY, Chang RQ, Liang Y, Hu XY, Li H, et al. GAS6-mediated dialogue between decidual stromal cells and macrophages is essential for early pregnancy maintenance by inducing M2-like polarization and cell proliferation of decidual macrophages. *Mol Hum Reprod* (2022) 28(3):1–14. doi: 10.1093/molehr/gaac006

95. Myers KV, Amend SR, Pienta KJ. Targeting Tyro3, Axl and MerTK (TAM receptors): Implications for macrophages in the tumor microenvironment. *Mol Cancer* (2019) 18(1):1–14. doi: 10.1186/s12943-019-1022-2

96. Vijayan M, Lee CL, Wong VHH, Wang X, Bai K, Wu J, et al. Decidual glycodelin-A polarizes human monocytes towards a decidual macrophage-like phenotype via siglec-7. *J Cell Sci* (2020) 133(14):jcs244400. doi: 10.1242/jcs.244400

97. Sheng YR, Hu WT, Wei CY, Tang LL, Liu YK, Liu YY, et al. IL-33/ST2 axis affects the polarization and efferocytosis of decidual macrophages in early pregnancy. *Am J Reprod Immunol* (2018) 79(6):1–10. doi: 10.1111/aji.12836

98. Lindau R, Mehta RB, Lash GE, Papapavlou G, Boij R, Berg G, et al. Interleukin-34 is present at the fetal-maternal interface and induces immunoregulatory macrophages of a decidual phenotype in vitro. *Hum Reprod* (2018) 33(4):588–99. doi: 10.1093/humrep/dey037

99. Wang XQ, Zhou WJ, Hou XX, Fu Q, Li DJ. Trophoblast-derived CXCL16 induces M2 macrophage polarization that in turn inactivates NK cells at the maternal-fetal interface. *Cell Mol Immunol* (2018) 15(12):1038–46. doi: 10.1038/s41423-018-0019-x

100. Ding J, Yang C, Cheng Y, Wang J, Zhang S, Yan S, et al. Trophoblast-derived IL-6 serves as an important factor for normal pregnancy by activating Stat3-mediated M2 macrophages polarization. *Int Immunopharmacol* (2021) 90:106788. doi: 10.1016/j.intimp.2020.106788

101. Weng YS, Tseng HY, Chen YA, Shen PC, Al Haq AT, Chen LM, et al. MCT-1/ miR-34a/IL-6/IL-6R signaling axis promotes EMT progression, cancer stemness and M2 macrophage polarization in triple-negative breast cancer. *Mol Cancer* (2019) 18 (1):1–15. doi: 10.1186/s12943-019-0988-0

102. Wang S, Sun F, Han M, Liu Y, Zou Q, Wang F, et al. Trophoblast-derived hyaluronan promotes the regulatory phenotype of decidual macrophages. *Reproduction* (2019) 157(2):189–98. doi: 10.1530/REP-18-0450

103. Meng YH, Zhou WJ, Jin LP, Liu LB, Chang KK, Mei J, et al. RANKL-mediated harmonious dialogue between fetus & mother guarantees smooth gestation by inducing decidual M2 macrophage polarization. *Cell Death Dis* [Internet] (2017) 8(10):1–13. doi: 10.1038/cddis.2017.505

104. Tsai YC, Tseng JT, Wang CY, Su MT, Huang JY, Kuo PL. Medroxyprogesterone acetate drives M2 macrophage differentiation toward a phenotype of decidual macrophage. *Mol Cell Endocrinol* (2017) 452:74–83. doi: 10.1016/j.mce.2017.05.015

105. Furcron AE, Romero R, Mial TN, Balancio A, Panaitescu B, Hassan SS, et al. Human chorionic gonadotropin has anti-inflammatory effects at the maternal-fetal interface and prevents endotoxin-induced preterm birth, but causes dystocia and fetal compromise in mice. *Biol Reprod* (2016) 94(6):25–8. doi: 10.1095/biolreprod.116.139345

106. Ye Y, Peng L, Chelariu-Raicu A, Kuhn C, Dong X, Jeschke U, et al. Prostaglandin E2 receptor 3 promotes M1 macrophages polarization in unexplained recurrent pregnancy loss. *Biol Reprod* (2022) 106(5):910–8. doi: 10.1093/biolre/ioac030

107. Mao X, Xu J, Wang W, Liang C, Hua J, Liu J, et al. Crosstalk between cancerassociated fibroblasts and immune cells in the tumor microenvironment: new findings and future perspectives. *Mol Cancer* (2021) 20(1):1–30. doi: 10.1186/s12943-021-01428-1

108. Marchesi M, Andersson E, Villabona L, Seliger B, Lundqvist A, Kiessling R, et al. HLA-dependent tumour development: A role for tumour associate macrophages? J Transl Med (2013) 11(1):1–15. doi: 10.1186/1479-5876-11-247

109. Zhou J, Zhang S, Guo C. Crosstalk between macrophages and natural killer cells in the tumor microenvironment. *Int Immunopharmacol* (2021) 101(PB):108374. doi: 10.1016/j.intimp.2021.108374

110. Li C, Jiang P, Wei S, Xu X, Wang J. Regulatory T cells in tumor microenvironment: New mechanisms, potential therapeutic strategies and future prospects. *Mol Cancer* (2020) 19(1):1–23. doi: 10.1186/s12943-020-01234-1

111. Cortesi F, Delfanti G, Grilli A, Calcinotto A, Gorini F, Pucci F, et al. Bimodal CD40/fas-dependent crosstalk between iNKT cells and tumor-associated macrophages impairs prostate cancer progression. *Cell Rep* (2018) 22(11):3006–20. doi: 10.1016/ j.celrep.2018.02.058

112. Banerjee S, Dudeja V, Saluja A. Unconventional T cells in the pancreatic tumor microenvironment: Thinking outside the box. *Cancer Discov* (2019) 9(9):1164–6. doi: 10.1158/2159-8290.CD-19-0722

113. Kersten K, Coffelt SB, Hoogstraat M, Verstegen NJM, Vrijland K, Ciampricotti M, et al. Mammary tumor-derived CCL2 enhances pro-metastatic systemic inflammation through upregulation of IL1 $\beta$  in tumor-associated macrophages. *Oncoimmunology* (2017) 6(8):1–14. doi: 10.1080/2162402X.2017.1334744

114. Fabian KP, Padget MR, Fujii R, Schlom J, Hodge JW. Differential combination immunotherapy requirements for inflamed (warm) tumors versus T cell excluded (cool) tumors: Engage, expand, enable, and evolve. *J Immunother Cancer* (2021) 9(2): e001691. doi: 10.1136/jitc-2020-001691

115. Bai R, Li Y, Jian L, Yang Y, Zhao L, Wei M. The hypoxia-driven crosstalk between tumor and tumor-associated macrophages: mechanisms and clinical treatment strategies. *Mol Cancer* (2022) 21(1):1–22. doi: 10.1186/s12943-022-01645-2

116. Ni X, Wu W, Sun X, Ma J, Yu Z, He X, et al. Interrogating glioma-M2 macrophage interactions identifies Gal-9/Tim-3 as a viable target against PTEN-null glioblastoma. *Sci Adv* (2022) 8(27):eabl5165. doi: 10.1126/sciadv.abl5165

117. Su S, Zhao J, Xing Y, Zhang X, Liu J, Ouyang Q, et al. Immune checkpoint inhibition overcomes ADCP-induced immunosuppression by macrophages. *Cell* (2018) 175(2):442–57.e23. doi: 10.1016/j.cell.2018.09.007

118. Gordon SR, Maute RL, Dulken BW, Hutter G, George BM, McCracken MN, et al. PD-1 expression by tumour-associated macrophages inhibits phagocytosis and tumour immunity. *Nature* (2017) 545(7655):495–9. doi: 10.1038/nature22396

119. Lindau R, Vondra S, Spreckels J, Solders M, Svensson-Arvelund J, Berg G, et al. Decidual stromal cells support tolerance at the human foetal-maternal interface by inducing regulatory M2 macrophages and regulatory T-cells. *J Reprod Immunol* (2021) 146:103330. doi: 10.1016/j.jri.2021.103330

120. Wei CY, Li MQ, Zhu XY, Li DJ. Immune status of decidual macrophages is dependent on the CCL2/CCR2/JAK2 pathway during early pregnancy. *Am J Reprod Immunol* (2021) 86(5):1–11. doi: 10.1111/aji.13480

121. Schmall A, Al-Tamari HM, Herold S, Kampschulte M, Weigert A, Wietelmann A, et al. Macrophage and cancer cell cross-talk via CCR2 and CX3CR1 is a fundamental mechanism driving lung cancer. *Am J Respir Crit Care Med* (2015) 191(4):437–47. doi: 10.1164/rccm.201406-1137OC

122. Fang XH, Li ZJ, Liu CY, Mor G, Liao AH. Macrophage memory: Types, mechanisms, and its role in health and disease. *Immunology* (2023) 171(1):18–30. doi: 10.1111/imm.13697

123. Liao HQ, Han MT, Cheng W, Zhang C, Li H, Li MQ, et al. Decidual-derived RANKL facilitates macrophages accumulation and residence at the maternal-fetal interface in human early pregnancy. *Am J Reprod Immunol* (2021) 86(2):1–9. doi: 10.1111/aji.13406

124. Zhang Y, Ma L, Hu X, Ji J, Mor G, Liao A. The role of the PD-1/PD-L1 axis in macrophage differentiation and function during pregnancy. *Hum Reprod* (2019) 34 (1):25–36. doi: 10.1093/humrep/dey347

125. Drukker M, Katz G, Urbach A, Schuldiner M, Markel G, Itskovitz-Eldor J, et al. Characterization of the expression of MHC proteins in human embryonic stem cells. *Proc Natl Acad Sci USA* (2002) 99(15):9864–9. doi: 10.1073/pnas.142298299

126. Lee CL, Guo Y, So KH, Vijayan M, Guo Y, Wong VHH, et al. Soluble human leukocyte antigen G5 polarizes differentiation of macrophages toward a decidual macrophage-like phenotype. *Hum Reprod* (2015) 30(10):2263–74. doi: 10.1093/ humrep/dev196

127. Parasar P, Guru N, Nayak NR. Contribution of macrophages to fetomaternal immunological tolerance. *Hum Immunol* (2021) 82(5):325-31. doi: 10.1016/j.humimm.2021.02.013

128. Kämmerer U, Eggert AO, Kapp M, McLellan AD, Geijtenbeek TBH, Dietl J, et al. Unique appearance of proliferating antigen-presenting cells expressing DC-SIGN (CD209) in the decidua of early human pregnancy. *Am J Pathol* (2003) 162(3):887–96. doi: 10.1016/S0002-9440(10)63884-9

129. Li M, Sun F, Xu Y, Chen L, Chen C, Cui L, et al. Tim-3+ decidual Møs induced Th2 and Treg bias in decidual CD4+T cells and promoted pregnancy maintenance *via* CD132. *Cell Death Dis* (2022) 13(5):1–12. doi: 10.1038/s41419-022-04899-2

130. Hu XH, Li ZH, Muyayalo KP, Wang LL, Liu CY, Mor G, et al. A newly intervention strategy in precclampsia: Targeting PD-1/Tim-3 signaling pathways to modulate the polarization of decidual macrophages. *FASEB J* (2022) 36(1):1–18. doi: 10.1096/fj.202101306R

131. Co EC, Gormley M, Kapidzic M, Rosen DB, Scott MA, Stolp HAR, et al. Maternal decidual macrophages inhibit NK cell killing of invasive cytotrophoblasts during human pregnancy. *Biol Reprod* (2013) 88(6):1–9. doi: 10.1095/biolreprod.112.099465

132. Vacca P, Cantoni C, Vitale M, Prato C, Canegallo F, Fenoglio D, et al. Crosstalk between decidual NK and CD14+ myelomonocytic cells results in induction of Tregs and immunosuppression. *Proc Natl Acad Sci USA* (2010) 107(26):11918–23. doi: 10.1073/pnas.1001749107

133. Kapellos TS, Iqbal AJ. Epigenetic control of macrophage polarisation and soluble mediator gene expression during inflammation. *Mediators Inflamm* (2016) 2016:6591703. doi: 10.1155/2016/6591703

134. Brown MB, von Chamier M, Allam AB, Reyes L. M1/M2 macrophage polarity in normal and complicated pregnancy. *Front Immunol* (2014) 5:1–11. doi: 10.3389/fimmu.2014.00606

135. Murdoch C, Muthana M, Coffelt SB, Lewis CE. The role of myeloid cells in the promotion of tumour angiogenesis. *Nat Rev Cancer* (2008) 8(8):618–31. doi: 10.1038/nrc2444

136. Mazzieri R, Pucci F, Moi D, Zonari E, Ranghetti A, Berti A, et al. Targeting the ANG2/TIE2 axis inhibits tumor growth and metastasis by impairing angiogenesis and disabling rebounds of proangiogenic myeloid cells. *Cancer Cell* (2011) 19(4):512–26. doi: 10.1016/j.ccr.2011.02.005

137. Duran CL, Borriello L, Karagiannis GS, Entenberg D, Oktay MH, Condeelis JS. Targeting tie2 in the tumor microenvironment: From angiogenesis to dissemination. *Cancers (Basel)* (2021) 13(22):5730. doi: 10.3390/cancers13225730

138. Du S, Wagner N, Wagner KD. The emerging role of PPAR beta/delta in tumor angiogenesis. PPAR Res (2020) 2020:3608315. doi: 10.1155/2020/3608315

139. Wang Q, He Z, Huang M, Liu T, Wang Y, Xu H, et al. Vascular niche IL-6 induces alternative macrophage activation in glioblastoma through HIF-2 $\alpha$ . Nat Commun (2018) 9(1):559. doi: 10.1038/s41467-018-03050-0

140. Takeda N, O'Dea EL, Doedens A, Kim JW, Weidemann A, Stockmann C, et al. Differential activation and antagonistic function of HIF- $\alpha$  isoforms in macrophages are essential for NO homeostasis. *Genes Dev* (2010) 24(5):491–501. doi: 10.1101/gad.1881410

141. De Palma M, Biziato D, Petrova TV. Microenvironmental regulation of tumour angiogenesis. *Nat Rev Cancer [Internet]* (2017) 17(8):457–74. doi: 10.1038/nrc.2017.51

142. Claesson-Welsh L, Welsh M. VEGFA and tumour angiogenesis. J Intern Med (2013) 273(2):114–27. doi: 10.1111/joim.12019

143. Schatz F, Guzeloglu-Kayisli O, Arlier S, Kayisli UA, Lockwood CJ. The role of decidual cells in uterine hemostasis, menstruation, inflammation, adverse pregnancy outcomes and abnormal uterine bleeding. *Hum Reprod Update* (2016) 22(4):497–515. doi: 10.1093/humupd/dmw004

144. Wang Q, Lash GE. Angiopoietin 2 in placentation and tumor biology: The yin and yang of vascular biology. *Placenta* (2017) 56:73–8. doi: 10.1016/j.placenta.2017.03.021

145. Hughes R, Qian BZ, Rowan C, Muthana M, Keklikoglou I, Olson OC, et al. Perivascular M2 macrophages stimulate tumor relapse after chemotherapy. *Cancer Res* (2015) 75(17):3479–91. doi: 10.1158/0008-5472.CAN-14-3587

146. Smith SD, Dunk CE, Aplin JD, Harris LK, Jones RL. Evidence for immune cell involvement in decidual spiral arteriole remodeling in early human pregnancy. *Am J Pathol* (2009) 174(5):1959–71. doi: 10.2353/ajpath.2009.080995

147. Paparini DE, Choudhury RH, Vota DM, Karolczak-Bayatti M, Finn-Sell S, Grasso EN, et al. Vasoactive intestinal peptide shapes first-trimester placenta trophoblast, vascular, and immune cell cooperation. *Br J Pharmacol* (2019) 176 (7):964–80. doi: 10.1111/bph.14609

148. Choudhury RH, Dunk CE, Lye SJ, Aplin JD, Harris LK, Jones RL. Extravillous trophoblast and endothelial cell crosstalk mediates leukocyte infiltration to the early remodeling decidual spiral arteriole wall. *J Immunol* (2017) 198(10):4115–28. doi: 10.4049/jimmunol.1601175

149. Rong M, Yan X, Zhang H, Zhou C, Zhang C. Dysfunction of decidual macrophages is a potential risk factor in the occurrence of preeclampsia. *Front Immunol* (2021) 12:1–14. doi: 10.3389/fimmu.2021.655655

150. Kumazaki K, Nakayama M, Suehara N, Wada Y. Expression of vascular endothelial growth factor, placental growth factor, and their receptors Flt-1 and KDR in human placenta under pathologic conditions. *Hum Pathol* (2002) 33 (11):1069–77. doi: 10.1053/hupa.2002.129420

151. Tan W, Chen L, Guo L, Ou X, Xie D, Quan S. Relationship between macrophages in mouse uteri and angiogenesis in endometrium during the periimplantation period. *Theriogenology* (2014) 82(7):1021-7. doi: 10.1016/ j.theriogenology.2014.07.025

152. Hemberger M, Hanna CW, Dean W. Mechanisms of early placental development in mouse and humans. *Nat Rev Genet* (2020) 21(1):27-43. doi: 10.1038/s41576-019-0169-4

153. Brown EAB, To drive or be driven SA. the path of a mouse model of recurrent pregnancy loss. *Reproduction* (2014) 2147(5):R153–67. doi: 10.1530/REP-13-0583

154. Chen H, Williams KE, Kwan EY, Kapidzic M, Puckett KA, Aburajab RK, et al. Global proteomic analyses of human cytotrophoblast differentiation/invasion. *Dev* (2021) 148(13):1–11. doi: 10.1242/dev.199561

155. Sun W, Luo Z, Lee J, Kim HJ, Lee KJ, Tebon P, et al. Organ-on-a-chip for cancer and immune organs modeling. *Adv Healthc Mater* (2019) 8(4):1–12. doi: 10.1002/adhm.201801363

156. Reeves E, James E. Tumour and placenta establishment: The importance of antigen processing and presentation. *Placenta* (2017) 56:34–9. doi: 10.1016/ j.placenta.2017.02.025

157. Costanzo V, Bardelli A, Siena S, Abrignani S. Exploring the links between cancer and placenta development. *Open Biol* (2018) 8(6):180081. doi: 10.1098/ rsob.180081

158. Tutar R, Çelebi-Saltik B. Modeling of artificial 3D human placenta. Cells Tissues Organs (2022) 211(4):527-36. doi: 10.1159/000511571

159. Shao Q, Liu X, Huang Y, Chen X, Wang H. Human decidual stromal cells in early pregnancy induce functional re-programming of monocyte-derived dendritic cells *via* crosstalk between G-CSF and IL-1 $\beta$ . *Front Immunol* (2020) 11:1–11. doi: 10.3389/fimmu.2020.574270

160. Hoffmann JP, Liu JA, Seddu K, Klein SL. Sex hormone signaling and regulation of immune function. *Immunity* (2023) 56(11):2472-91. doi: 10.1016/j.immuni.2023.10.008

161. Erlebacher A, Vencato D, Price KA, Zhang D, Glimcher LH. Constraints in antigen presentation severely restrict T cell recognition of the allogeneic fetus. *J Clin Invest.* (2007) 117(5):1399–411. doi: 10.1172/JCI28214

162. DhatChinamoorthy K, Colbert JD, Rock KL. Cancer immune evasion through loss of MHC class I antigen presentation. *Front Immunol* (2021) 12. doi: 10.3389/fimmu.2021.636568

163. True H, Blanton M, Sureshchandra S, Messaoudi I. Monocytes and macrophages in pregnancy: The good, the bad, and the ugly\*. *Immunol Rev* (2022) 308(1):77–92. doi: 10.1111/imr.13080

164. Albright CM, Wenstrom KD. Malignancies in pregnancy. Best Pract Res Clin Obstet Gynaecol (2016) 33:2-18. doi: 10.1016/j.bpobgyn.2015.10.004