Check for updates

OPEN ACCESS

EDITED BY Xiangwei Xiao, Children's Hospital of Pittsburgh, University of Pittsburgh, United States

REVIEWED BY Yuling Han, Cornell University, United States Elisabeth M. Lodder, University of Amsterdam, Netherlands

*CORRESPONDENCE Jian Yang 1313027@zju.edu.cn Shenjiang Hu hzhzl85519933@zju.edu.cn

[†]These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION This article was submitted to Inflammation, a section of the journal Frontiers in Immunology

RECEIVED 11 September 2022 ACCEPTED 24 March 2023 PUBLISHED 05 April 2023

CITATION

Chen Y, Lou L, Zhang X, Jin L, Chen Y, Chen L, Li Z, Zhang F, Fu T, Hu S and Yang J (2023) Association between circulating leukocytes and arrhythmias: Mendelian randomization analysis in immuno-cardiac electrophysiology. *Front. Immunol.* 14:1041591. doi: 10.3389/fimmu.2023.1041591

COPYRIGHT

© 2023 Chen, Lou, Zhang, Jin, Chen, Chen, Li, Zhang, Fu, Hu and Yang. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Association between circulating leukocytes and arrhythmias: Mendelian randomization analysis in immunocardiac electrophysiology

Yuxiao Chen^{1†}, Lian Lou^{1†}, Xuan Zhang^{1†}, Luyang Jin², Yao Chen¹, Lele Chen¹, Zhihang Li¹, Fen Zhang³, Ting Fu⁴, Shenjiang Hu^{1*} and Jian Yang^{1*}

¹Department of Cardiology, The First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, China, ²Department of Gynecology, The First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, China, ³Department of Cardiology, Jinhua People's Hospital, Jinhua, China, ⁴Department of Cardiology, Yiwu Central Hospital, Jinhua, China

Background: Cardiac arrhythmia is a common disease associated with high mortality and morbidity. Circulating leukocyte counts, which serve as a biomarker for assessing systemic immune status, have been linked to arrhythmias in observational studies. However, observational studies are plagued by confounding factors and reverse causality, whether alterations in circulating leukocyte components are causally associated with arrhythmias remains uncertain. The present study explored this question based on genetic evidence.

Methods and findings: We performed Mendelian randomization (MR) analysis to evaluate whether alterations in leukocyte counts affect aggregated risk of all types of arrhythmia or risk of five specific types of arrhythmia. Single-nucleotide polymorphisms serving as proxies for leukocyte differential counts were retrieved from the Blood Cell Consortium, and statistical data on arrhythmias were obtained from the UK Biobank), FinnGenand a meta-analysis of genomewide association studies for atrial fibrillation. We applied inverse varianceweighted method as the primary analysis, complemented by a series of sensitivity analyses. Bidirectional analyses were conducted to assess reverse causality. Finally, multivariable MR was performed to study the joint effects of multiple risk factors. We found that genetically predicted differential leukocyte counts were not significantly associated with aggregated occurrence of all types of arrhythmia. In contrast, each 1-standard deviation increase in lymphocyte count was associated with 46% higher risk of atrioventricular block (OR 1.46, 95% CI 1.11–1.93, p=0.0065). A similar effect size was observed across all MR sensitivity analyses, with no evidence of horizontal pleiotropy. Reverse MR analysis suggested that atrioventricular block was unlikely to cause changes in lymphocyte count. Primary MR analysis based on the inverse-variance weighted method suggested that changes in neutrophil count alter risk of right bundle branch block, and changes in basophil count alter risk of atrial fibrillation. However, these causal relationships were not robust in sensitivity analyses. We

found no compelling evidence that neutrophil or lymphocyte counts cause atrial fibrillation.

Conclusion: Our data support higher lymphocyte count as a causal risk factor for atrioventricular block. These results highlight the importance of immune cells in the pathogenesis of specific cardiac conduction disorders.

KEYWORDS

leukocyte, lymphocyte, arrhythmia, atrioventricular block, Mendelian randomization

1 Introduction

Cardiac arrhythmia, a condition in which the heartbeat is irregular, too fast or too slow, is a relatively common heart disease. Some arrhythmias are brief and asymptomatic, while others are persistent and can lead to hemodynamic instability, thromboembolic events, and even cardiac sudden death, imposing a significant burden on healthcare systems. However, evidence on how to effectively prevent and treat arrhythmias, thus far, has been limited.

Although how exactly leukocytes participate in arrhythmogenesis is not fully understood, it is generally accepted that leukocytes might contribute to arrhythmia either directly through coupling to cardiomyocytes, or indirectly by producing cytokines and antibodies (1). Neutrophils, which are rarely found in the healthy myocardium, are rapidly recruited to the heart in response to stress signals, and they exert the arrhythmogenic effect by releasing myeloperoxidase (2) and lipocalin (3), promoting oxidative stress and interstitial fibrosis. Monocytes/macrophages are the most numerous leukocytes in the heart, and can effectively clear dysfunctional mitochondria, apoptotic cells and debris, thus preventing ventricular tachycardia and fibrillation after myocardial infarction (3). However, macrophages can also secrete cytokines like IL-1 β , which then prolong the action potential duration of cardiomyocytes and induces arrhythmias (4). Recent research has uncovered a non-canonical leukocyte function for macrophages in cardiac electrical conduction, demonstrating that they can directly couple to conducting cardiomyocytes via gap junctions containing Cx43, altering their electrical properties (5). T lymphocytes elicit cellmediated immunity, and subsets of T lymphocytes may produce cytokines like IFN-y, IL-2 or IL-17, exacerbating the neutrophilic inflammation, then promote micro-scar formation among myocardial tissue, leading to insulating fibrosis (6). B lymphocytes can promote cardiac arrhythmias by means of autoantibodies targeting specific calcium, potassium, or sodium channels on the surface of cardiomyocytes (7). Last, less is known about the function of basophils and eosinophils in arrhythmias, but recent experimental data highlight that basophil-derived IL-14 plays an essential role in the heart, by balancing macrophage polarization. While eosinophils may play an anti-inflammatory and cardioprotective role after myocardial infarction, reducing cardiomyocyte death and inflammatory cell accumulation (8, 9). Due to these pioneering

works, researchers have attempted to integrate electrophysiology and immunology, and a new terminology "immuno-cardiac electrophysiology" was introduced to highlight the emerging essential role of immune cells in arrhythmias (1, 10, 11).

Circulating leukocytes are crude markers of the systemic immunological status of individuals, and they can modulate local inflammatory responses. Cellular numbers are the most critical parameter for homeostasis of circulating immune cells. So far, some cross-sectional clinical surveys have linked circulating leukocyte counts to incidence of cardiac arrhythmias. In the CALIBER study of 775,231 individuals, high neutrophil count (12), low eosinophil count, and low lymphocyte count (13) were associated with ventricular arrhythmia. In the Framingham Heart Study, white blood cell counts correlated with risk of atrial fibrillation (14). Other studies have linked risk of atrial fibrillation to eosinophil count (15) and proportion of monocyte subsets (16). However, that literature does not definitively establish a role for leukocyte counts in the pathogenesis of arrhythmias because observational studies are prone to residual unmeasured confounding and reverse causation. Of particular concern is the potential for reverse causation. Atrial fibrillation itself might promote systemic inflammation during atrial remodeling and induce a spurious inverse association. In addition, observational studies have come to conflicting conclusions about the association of leukocyte counts with supraventricular tachycardia (17, 18). Therefore, evidence from observational studies alone is insufficient. The causal effect of leukocyte counts on the risk of arrhythmias remains unknown. And additional studies are needed to characterize the role of each immune cell subtype in different types of arrhythmias. Addressing these causal questions can accelerate the discovery of mechanisms underlying disease and open new prevention and treatment avenues.

Mendelian randomization (MR) is an epidemiologic approach that strives to address some key limitations of observational studies, such as confounding and reverse causation (19). It uses genetic variants, usually single-nucleotide polymorphisms (SNPs), as proxies for clinical interventions (as a result of exposure) in order to assess whether the genetic variants are associated with the outcome. In this way, MR supports inferences about causality (20), placing it at the interface between traditional observational epidemiology and interventional trials (21). MR should be robust to confounders, given that alleles are randomly distributed at conception, and it should be robust to reverse causation, since an individual's genetic code is fixed at birth, before the outcome of interest. In the present study, two-sample MR was used to estimate whether leukocyte counts cause changes in arrhythmia risk, based on summary data in genome-wide association studies (GWAS).

2 Methods

2.1 Study design

For the current study, we conducted two-sample MR analysis of circulating leukocyte counts on arrhythmias using data from publicly available GWAS. The five subtypes of leukocytes were considered: neutrophils, eosinophils, basophils, monocytes, and lymphocytes. Arrythmia was defined as all types in aggregate or as one of the following five specific types: atrial fibrillation, atrioventricular block, left bundle branch block (LBBB), right bundle branch block (RBBB), and paroxysmal tachycardia.

All study procedures were performed in accordance with the World Medical Association Declaration of Helsinki ethical principles for medical research. Ethics approval was considered unnecessary for the present study because the included GWAS reported appropriate ethical approval from their respective institutions, and the present analyses were performed only on summary-level data.

2.2 Selection of genetic instruments for circulating leukocyte counts

We extracted summary statistics from the largest meta-analyzed GWAS data provided by the Blood Cell Consortium (22). The Blood Cell Consortium Phase 2 includes 563,946 European participants from 26 GWAS cohorts, after excluding patients with blood cancer, acute medical/surgical illness, myelodysplastic syndrome, bone marrow transplant, congenital/hereditary anemia, HIV, end-stage kidney disease, splenectomy, cirrhosis or extreme blood cell counts. An overview of the data sources is provided in Supplementary Table S1, and more detail is available in the original article (23).

SNPs associated with the counts of the five leukocyte counts were selected at the genome-wide significance level ($P < 5 \times 10^{-8}$) and defined as genetic instruments. To ensure that SNPs were independent, a clumping procedure was performed, and the SNPs were pruned at a stringent linkage disequilibrium (LD) of $R^2 < 0.001$ within a 10,000-kb window.

The proportions of variance in respective leukocyte counts explained by the selected SNPs were estimated (Supplementary Table S2), and F-statistics were calculated as measures of instrument strength (24). The F value for all genetic instruments was > 10, ensuring that weak bias would be <10% at least 95% of the time (Supplementary Table S2).

2.3 Data sources for arrhythmia

To more thoroughly evaluate the association of leukocyte counts and the risk of arrhythmias, we aimed to include all

eligible GWAS of arrhythmias by extensively searching the public Integrative Epidemiology Unit (IEU) GWAS database (https:// gwas.mrcieu.ac.uk/). We selected GWAS with the largest samples, leading to seven GWAS whose summary statistics for different types of arrhythmias were used in the present study.

Genetic association estimates for the outcome of all types of arrhythmia were obtained from the UK Biobank (UK Biobank field ID 20002, value 1077), based on the UKB GWAS pipeline set up for the MRC IEU. We restricted the analytical cohort to individuals of European descent. Individuals with cardiac arrhythmia were identified via self-report during a face-to-face interview with a trained nurse. The GWAS dataset on atrial fibrillation was obtained from a meta-analysis comprising 1,030,836 participants of European ancestry (25). Cases of atrial fibrillation were defined as those patients with paroxysmal atrial fibrillation, permanent atrial fibrillation, or atrial flutter. Summary data for the other four types of arrhythmias (atrioventricular block, LBBB, RBBB, and paroxysmal tachycardia) were retrieved from the FinnGen project (release 2), where cases were defined as those assigned the corresponding ICD-10 diagnosis codes. Specifically, cases of atrioventricular block were defined as patients with first degree (ICD10:I440), second degree (I441), third degree atrioventricular block (I442) or other unspecified atrioventricular block (I443). LBBB included left anterior fascicular block (I444), left posterior fascicular block(I445), other fascicular block (I446) and unspecified LBBB (I447). While RBBB inlcuded right fascicular block (I450) and other RBBB (I451). And the term paroxysmal tachycardia referred to re-entry ventricular, supraventricular, ventricular tachycardia and unspecified paroxysmal tachycardia (I47). The FinnGen project included 102,739 Finnish participants and combined genetic data from Finnish biobanks and health records from Finnish health registries. Further details on data sources are included in Supplementary Table S1.

Prior to the MR analyses, we harmonized the SNPs identified from exposure GWAS with SNPs in outcome GWAS in order to align alleles on the same strand.

2.4 Statistical analyses

We used the inverse-variance weighted (IVW) method as the primary analysis. Then we applied a range of sensitivity analyses to assess the robustness of the IVW findings against potential violations, including MR-Egger, weighted median, MR-PRESSO and multivariable MR (MVMR) analyses. Although these methods have relatively low statistical efficiency on their own, they have different theoretical properties to control for different types of biases, and they are robust to certain assumption violations.

The IVW method (random effects model) can provide the greatest statistical power (26), assuming all genetic instruments are valid. This method is equivalent to a weighted linear regression of the SNP-exposure effects against the SNP-outcome effects, with the intercept constrained to zero. Owing to this constraint, it can lead to a relatively high rate of false positives in the presence of horizontal pleiotropy. Cochran's Q statistic (27) from IVW analysis was used for global heterogeneity testing. Based on the notion that

pleiotropy is one of the main sources of heterogeneity, low heterogeneity (Cochran's Q p > 0.05) implies the minor possibility of pleiotropy.

MR-Egger regression is performed similarly as IVW, except the intercept is not fixed to zero (28). Therefore, the slope coefficient of MR-Egger regression gives an adjusted causal estimate, even when pleiotropy is present. The intercept of MR-Egger regression is an indicator of average pleiotropic effect across the genetic variants. An intercept of zero associated with P > 0.05 was considered evidence for absence of pleiotropic bias.

The weighted median method (29) is a consensus approach that takes the median of the ratio estimate distribution as the overall causal estimate. It has the advantage that it provides unbiased estimates when more than 50% of the weight comes from valid variants. It is less affected when a few genetic variants have pleiotropic effects, and it can be viewed as an implicit outlier removal approach.

The MR-PRESSO (30), a newly proposed MR method, is a variation on the IVW method. MR-PRESSO global test is used to assess the presence of overall horizontal pleiotropy. If pleiotropy is detected, the MR-PRESSO outlier test allows the detection of individual pleiotropic outliers through calculation of the residual sum of squares. Finally, the causal estimate is obtained by applying the IVW method to the genetic variants remaining after exclusion of outliers.

Steiger filtering (31), which computes the amount of variance each SNP explains in the exposure and in the outcome variable, identifies variant instruments that are likely to reflect reverse causation.

When significant horizontal pleiotropy was detected, we also used Cook's distance to identify outliers. Cook's distance identifies SNPs that exert disproportionate influences on the overall estimates as outliers.

MVMR, an extension of the standard MR approach, considers multiple correlated exposures within a single model, allowing the disentanglement of independent associations of each exposure with the outcome. This method was performed while considering associations of SNPs with diabetes mellitus (DM), hypertension, and coronary artery disease (CAD) as covariates in order to estimate the direct effects of leukocyte counts independently of risk factors known to influence risk of arrhythmia. Given the strong correlations between leukocyte subtypes, we also performed MVMR to determine the effect of each of the five leukocyte subtypes separately on arrhythmia, after adjusting for the effects of the other four subtypes.

We performed reverse-direction MR analysis to evaluate whether there is genetic evidence for the possibility that arrhythmia alters circulating leukocyte counts. Because we detected few genome-wide significant SNPs for arrhythmias (defined as $p < 5 \times 10^{-8}$), we used a less stringent statistical threshold ($p < 1 \times 10^{-5}$) to select genetic instruments (32). In fact, we were unable to detect eligible SNPs associated with the aggregated occurrence of all types of arrhythmia, even at the suggestive level of $p < 1 \times 10^{-5}$, so this outcome was not included in the analysis. In this reverse-direction analysis, IVW, MR-Egger and weighted median analyses were performed as described above.

All statistical analyses were conducted using the TwoSampleMR, MendelianRandomization, and MR-PRESSO

packages in R (version 4.0.3). Effect estimates for dichotomous outcomes were reported as odds ratios (ORs) with corresponding 95% confidence intervals (CIs).

2.5 Interpretation of results

Normally, Bonferroni-corrected p values are used to adjust for multiple testing. However, given the large number of arrhythmia outcomes and leukocyte counts in the study, we judged this correction procedure to be unnecessarily conservative (33). Therefore, we applied the conventional p value threshold of 0.05, and we interpreted p values near 0.05 with caution.

We considered casual associations to be strongly supported if the following four criteria were satisfied. (1) Primary IVW analysis gave a statistically significant causal estimate (p < 0.05). (2) All sensitivity analyses yielded concordant estimates, despite making different assumptions. (3) No evidence of unbalanced horizontal pleiotropy was observed, defined as p > 0.05 for Cochran's Q statistic, MR–Egger intercept test and MR-PRESSO global pleiotropy test. (4) No evidence of reverse causation from arrhythmias to leukocyte differential counts was observed, defined as p > 0.05 in the IVW, MR-Egger, and weighted median analyses in reverse MR analysis.

3 Results

3.1 Circulating leukocyte counts and heart arrhythmias: Primary results

First, we investigated the causal effect of each leukocyte subtype count on arrhythmias using IVW methods with multiplicative random effects. The IVW approach is recommended as the primary method in MR analysis because it is optimally efficient when all genetic variants are valid (34). The results of IVW analysis are presented in Figure 1.

We did not find clear evidence supporting causal effects of any leukocyte subtype counts on the overall occurrence of all-type arrhythmia (Figure 1). Nevertheless, there was evidence that different leukocyte subtype counts causally affected three specific types of arrhythmias. A genetically estimated 1-standard deviation increase in lymphocyte count was associated with 46% higher risk of atrioventricular block (OR 1.46, 95% CI 1.11–1.93, p=0.0065). We also found moderate evidence for causal effects of basophil count on atrial fibrillation (OR 1.08, 95% CI 1.01–1.58, p=0.0237), and neutrophil count on RBBB (OR 2.32, 95% CI 1.11–4.86, p=0.0259). No significant associations were observed for the other outcomes.

3.2 Sensitivity analyses of positive results

We assessed the robustness of the significant causal estimates from the above IVW analysis using sensitivity analyses. These sensitivity analyses are generally considered less powerful than



bundle-branch block; OR, odds ratio; CI, confidence interval.

the conventional IVW approach, but robust to different forms of biases (see Methods). Therefore we conducted MR-Egger, MR-PRESSO, weighted median, Steiger filtering and multivariable MR analyses on the following three combinations of exposure and outcome: (1) lymphocyte count and atrioventricular block, (2) neutrophil count and RBBB, and (3) basophil count and atrial fibrillation.

3.2.1 Lymphocyte count and atrioventricular block

Sensitivity analyses supported the causal link between lymphocyte count and atrioventricular block (Figure 2): the MR-Egger approach indicated an OR 1.95 (95% CI 1.12-3.39; p=0.019), and the weighted median approach indicated an OR 1.76 (95% CI 1.20-2.78; p=0.015). With respect to pleiotropy detection, Cochran's



Q test gave a p value of 0.586, suggesting no evidence of heterogeneity between genetic instruments and therefore no pleiotropy. Similarly, bias due to pleiotropy was not detectable in the IVW analyses, based on a p value of 0.249 for the MR-Egger intercept test and p value of 0.566 for MR-PRESSO global pleiotropy test. Additionally, the absence of outliers detected through the Steiger filtering reinforced this conclusion (Supplementary Table S3).

Using MVMR analysis, we confirmed this causal relationship after adjusting for risk factors of arrhythmia (CAD, DM and hypertension) and for effects from the other four subtypes of leukocytes (Figure 2).

3.2.2 Neutrophil count and RBBB

The weighted median method (OR 3.40, 95% CI 1.02-11.28; p=0.049) and MR-Egger method (OR 3.13, 95% CI 0.64-15.32; p=0.158) produced results similar with those of the primary IVW analysis (Figure 3). But these CIs were wide and the p values near 0.05 or above 0.05, likely due to lack of statistical power. There was no indication of heterogeneity or pleiotropy in the corresponding Cochran's Q test (p =0.200), MR-Egger intercept test (p=0. 674) or MR-PRESSO global pleiotropy test (p=0.209). And Steiger filtering did not detect any outliers (Supplementary Table S3). In MVMR analysis, accounting for counts of lymphocytes and eosinophils abolished the direct effect of neutrophil count on RBBB (Figure 2). Together, these analyses suggest no direct, independent effect of neutrophil counts on the risk of RBBB.

3.2.3 Basophil count and atrial fibrillation

For basophil count and atrial fibrillation, the issue of horizontal pleiotropy is a particular concern (Figure 4). Although the intercept estimated from the MR-Egger regression was centered around zero (-0.0004, p=0.801), and Steiger filtering did not identify any outliers (Supplementary Table S3), we determined the presence of overall horizontal pleiotropy among all genetic instruments using MR-PRESSO (global pleiotropy p<0.001). After removing five outlier SNPs, the causal estimate of basophil count on atrial fibrillation no longer achieved statistical significance (MR-PRESSO outlier correction p=0.106). Similarly, effect estimates from MR-Egger and weighted median analyses were not significant. In conclusion, these analyses suggest that the estimate from IVW analysis may be strongly affected by pleiotropy, and that no compelling evidence exists in support of a causal association between basophil count and atrial fibrillation.

3.3 Sensitivity analyses of negative results

To reduce the incidence of false negative findings, sensitivity analyses (MR-Egger, weighted median, Steiger filtering, MR-PRESSO) were also performed to assess the validity of negative results. Empirically, we focused on the causal relationships of lymphocyte count or neutrophil count with atrial fibrillation.

For lymphocyte count and atrial fibrillation, both MR-Egger and MR-PRESSO methods gave negative, non-significant estimates

	Outcome: RBBB	:	OR (95% CI)	p-value	Pleiotropy Test
	MR-Egger			0.158	Intercept p value=0.674
	Weighted median		3.40 (1.02-11.28)	0.049	
	MR-PRESSO(Raw)*		2.32 (1.11-4.86)	0.027	Global pleiotropy p value=0.209
	MVMR adjusted for DM		2.62 (1.23-5.54)	0.012	
	MVMR adjusted for hypertension		2.52 (1.19-5.34)	0.016	
	MVMR adjusted for CAD		2.06 (1.05-4.05)	0.036	
	MVMR adjusted for eosinophils		1.52 (0.69-3.36)	0.302	
	MVMR adjusted for basophils		3.52 (1.52-8.17)	0.003	
	MVMR adjusted for monocytes		2.87 (1.07-7.69)	0.037	
	MVMR adjusted for lymphocytes		1.76 (0.73-4.21)	0.208	
	-	01 4 7 10 13 OR (95% CI)	16		
GURE 3					

FIGU

Outcome: AF		OR (95% CI)	p-value	Pleiotropy Test
MR-Egger	·	1.10 (0.96-1.26)	0.172	Intercept p value=0.801
Weighted median		1.07 (0.98-1.17)	0.122	
MR-PRESSO(Raw)		1.08 (1.01-1.16)	0.025	Global pleiotropy p value<0.001
MR-PRESSO(5 outliers Corrected)		1.05 (0.99-1.12)	0.106	
MVMR adjusted for DM		1.10 (1.02-1.19)	0.012	
MVMR adjusted for hypertension		1.08 (0.99-1.16)	0.051	
MVMR adjusted for CAD		1.10 (1.03-1.18)	0.007	
MVMR adjusted for neutrophils		1.11 (1.02-1.21)	0.021	
MVMR adjusted for eosinophils		1.15 (1.05-1.26)	0.002	
MVMR adjusted for monocytes	÷	1.07 (0.98-1.16)	0.122	
MVMR adjusted for lymphocytes		1.14 (1.04-1.24)	0.003	
	1 1.5 OR (95% CI)	-		

Sensitivity MR analyses of the causal association between basophil count and risk of atrial fibrillation using MR-Egger, weighted median, MR-PRESSO and MVMR analyses. Statistical significance was defined as p<0.05.

similar to those of the IVW analysis (Table 1). Only weighted median analysis showed a significant, albeit small, effect. Steiger filtering identified one outlier SNP, but the results of above analysis remained essentially unchanged after removing the outlier (Supplementary Table S3). The finding from weighted median analysis alone is insufficient evidence. Overall, we conclude the absence of strong evidence for a causal association between lymphocyte count and risk of atrial fibrillation.

For neutrophil count and atrial fibrillation, the MR-Egger and weighted median analyses showed a statistically significant causal estimate, which was inconsistent with the IVW analysis (Table 1). These two methods are perceived as methods that have natural robustness to pleiotropy. Meanwhile, we found evidence of pleiotropy based on the p values for the MR-Egger intercept test (p=0.013) and MR-PRESSO global pleiotropy test(p<0.001), as well as evidence of substantial heterogeneity based on the p value for Cochran's Q statistic (p<0.001). We suspect that pleiotropy biased the effect estimate towards null in the IVW analysis, even if pleiotropy more often biases estimates away from null. To remove potential pleiotropy as much as possible, we applied two additional different methods(MR-PRESSO outlier test and Cook's distance) to further determine and exclude potential outliers. Using MR-PRESSO and Cook's distance, we identified 9 and 19 outliers, respectively. After removing the outlier SNPs, the causal estimates still did not reach statistical significance (Table 1). In fact, the estimates were even smaller than before. Taken together, our analyses indicate no compelling evidence for a causal effect of neutrophil count on atrial fibrillation.

Sensitivity analyses of the other 29 exposure-outcome combinations yielded negative findings similar to those of the IVW analyses (Supplementary Table S4).

3.4 Reverse MR analysis to assess the effect of arrhythmias on leukocyte counts

To examine the possibility that reverse causation could be driving our findings, we performed extensive reverse MR analysis in which the risk of arrhythmia was the exposure and counts of the five leukocyte subtypes were the outcome. Although the IVW analysis showed that atrial fibrillation, paroxysmal tachycardia, LBBB and RBBB all had effects on the differential leukocyte counts, the effect sizes were so small that their practical significance is highly questionable (Supplementary Table S5). Moreover, these causal effects did not achieve statistical significance in either MR-Egger or weighted median analysis

TABLE 1 Sensitivity MR analyses evaluating the causal effects of neutrophil and lymphocyte counts on atrial fibrillation.

Exposure	Outcome	Method	OR (95% CI)	p value	Pleiotropy test p value
Lymphocyte count	Atrial fibrillation	MR-Egger	0.99 (0.90-1.08)	0.762	0.66
		Weighted median	0.93 (0.88-0.99)	0.014	
		MR-PRESSO (Raw)	0.97 (0.92-1.01)	0.147	<0.001
		MR-PRESSO (remove 9 outliers)	0.97 (0.93-1.01)	0.129	
		IVW (remove 25 outliers identified by cook's distance)	0.96 (0.92-1.00)	0.051	
Neutrophil count	Atrial fibrillation	MR-Egger	1.17 (1.05-1.29)	0.003	0.013
		Weighted median	1.09 (1.03-1.16)	0.006	
		MR-PRESSO (Raw)	1.04 (0.99-1.10)	0.095	<0.001
		MR-PRESSO (remove 9 outliers)	1.04 (0.99-1.09)	0.131	
		IVW (remove 19 outliers identified by cook's distance)	1.04 (0.99-1.09)	0.17	

(Supplementary Table S5). Therefore, we did not found any robust evidence of reverse associations. In particular, we did not observe causal effects of atrioventricular block on lymphocyte count in IVW method (OR 1.001, 95% CI 0.998–1.004; *P*p=0.44) (Supplementary Table S5). Similar results were observed in MR-Egger and weighted median analyses (Supplementary Table S5).

4 Discussion

In this study, using large publicly available genomic datasets, we conducted MR analyses to investigate the causal effects of leukocyte counts on different types of arrhythmias. Our principal findings are that genetically determined high lymphocyte count increases risk of atrioventricular block. In contrast, we did not detect a significant causal effect of either neutrophil or lymphocyte count on risk of atrial fibrillation. Although sparse observational studies have reported relationships between leukocyte counts and some types of arrhythmias, the unique contribution of the present study is that we precisely investigated the association of each differential leukocyte count with five specific types of arrhythmias. In addition, we used MR methods, which help to minimize bias due to confounding factors and reverse causation, allowing us to draw conclusions about causal relationships, not merely associations.

Diversity is an intrinsic characteristic of the immune system, which exerts an important influence on an individual's risk of developing immune mediated diseases. Although the abundance of circulating immune cells is particularly prone to change in the context of infection or injury, it has been demonstrated to be highly variable even among "healthy" individuals (35). Moreover, evidence has suggested that immune cell composition is associated with risks of cancer (36) and cardiovascular disease (12) among healthy people without prior corresponding diseases, although the exact causal relationship between immune cell composition changes and disease remains unclear. The analyses in the present study were carried out on data in the Blood Cell Consortium, for which mean leukocyte counts were within the normal range (22). Thus, our results may support the potential of leukocyte counts for predicting assessing arrhythmia risk in disease-free individuals.

High-degree atrioventricular block is the leading reason for pacemaker implantation. First-degree atrioventricular block, previously thought to be associated with a favorable prognosis, may actually be linked to adverse cardiovascular outcomes and increased mortality (37). However, due to the unknown mechanism of atrioventricular block, prevention and non-invasive treatment strategies are largely lacking in clinical practice. In particular, whether changes in circulating leukocyte components affect the risk of developing atrioventricular block remains unclear, as is the question of which types of leukocyte exert greater influence on atrioventricular block. Macrophages have been implicated in the disorder: they are abundant at the atrioventricular node and affect its physiological function though electrical coupling with cardiomyocytes (5). However, the current study did not find evidence supporting a causal effect of circulating monocyte count on atrioventricular block. We assume that this discrepancy stems from the fact that most cardiac macrophages, especially those resident in the atrioventricular node, populate the heart during embryogenesis and self-maintain locally with minimal exchange with the population of circulating monocytes (5, 38). On the other hand, our results revealed that genetically determined high lymphocyte count increases the risk of atrioventricular block. To the best of our knowledge, data on the impact of lymphocyte on atrioventricular block are scarce. The etiology of atrioventricular block is related to fibrosis of the conduction system (39), electrical remodeling of atrioventricular node myocytes (40, 41), and elevated vagal tone (42). Depending on the types of cells involved, it is speculated that lymphocytes may affect atrioventricular conduction in various ways. For instance, by secreting cytokines, lymphocytes can regulate monocyte/macrophage recruitment and differentiation (43). As previously mentioned, macrophages can directly affect the action potential of cardiomyocytes through gap junctions (5). Additionally, lymphocytes can promote fibroblast activation by secreting inflammatory mediators (44), leading to fibrosis in the atrioventricular node area and subsequent electrical isolation. Moreover, it may be possible that during cardiac injury, endogenous antigens in the conduction system are exposed, triggering the proliferation of autoreactive T and B cells and subsequent damage to atrioventricular node myocytes. Finally, it is worth investigating whether lymphocytes can directly couple to cardiomyocytes or produce autoantibodies that cross-react with ion channels in cardiomyocytes and ultimately affect their action potential. In conclusion, our results justify detailed studies into the role of lymphocytes in the pathogenesis of atrioventricular block, as well as their utility as a biomarker in disease risk assessment. Atrial fibrillation is the most common arrhythmia, and it increases the risk of stroke, heart failure and mortality (45). Previous observational studies have reported links between the disorder and high ratios of circulating neutrophils to lymphocytes (46, 47). Animal studies further support that atrial fibrillation involves atrial infiltration by neutrophils (48). However, we did not find any significant association between genetically predicted neutrophil or lymphocyte counts and atrial fibrillation. In particular, although our effect estimates for neutrophil counts were directionally concordant with the results from observational studies, the effect sizes were small and the CIs wide. These findings, coupled with inconsistent estimates from our various sensitivity analyses, lead us to conclude that genetically determined neutrophil counts do not substantially influence risk of atrial fibrillation. One potential reason for the differences between our work and previous epidemiological studies is that our MR analysis evaluated how lifelong exposure to increased leukocyte counts affected risk of atrial fibrillation (21). In contrast, observational studies typically have limited follow-up and may focus on short-term effects of leukocyte counts on the risk of postoperative atrial fibrillation (49). This study has limitations worth considering. First, it was restricted to a population of European descent for the sake of genetic homogeneity, so its generalizability to other ethnic groups is unclear. Second, lymphocytes are a diverse population of cells that have distinct phenotypic and functional properties. The aggregated count of all lymphocytes is far from fully representing the heterogeneous changes of lymphocyte subpopulations. Future studies should examine specific subsets of circulating lymphocytes,

such as through fluorescence-activated cell sorting. Third, we were unable to distinguish different subtype of each kind of arrhythmias in our analysis, due to the lack of detailed original GWAS data. Fourth, no MR analysis can entirely exclude the influence of pleiotropic effects. Nevertheless, the observed consistency of effect estimates across multiple sensitivity analyses implies minimal confounding and bias. Fifth, this study did not encompass ventricular tachycardia or ventricular fibrillation, as large-scale population-based GWAS summary statistics on ventricular arrhythmias are currently unavailable. Recruiting patients with ventricular fibrillation in the setting of acute myocardial infarction is challenging when compared to the ease of recruitment of atrial fibrillation patients (50). Existing GWAS primarily focus on electrophysiological parameters that highly correlated with ventricular tachyarrhythmia, such as PR interval (51), QT interval (50, 52), or specific diseases like Brugada syndrome (53) or long QT syndrome (54), which are predominantly characterized by ventricular arrhythmias. Sixth, the "all types of arrhythmias" analyzed in the study represent a collection of phenotypes. It may introduce composition bias. This is because the proportion of each arrhythmia in the dataset is unknown, and changes in the proportion can significantly impact the causal effects, thereby reducing reproducibility. Furthermore, if the causal effects of leukocytes are opposite on different types of arrhythmias, they may mutually cancel out, resulting in inaccurate findings.

5 Conclusion

In conclusion, our study provides strong evidence of a causal effect of genetically high lymphocyte count on the risk of atrioventricular block. We failed to find evidence supporting a causal effect of lymphocyte or neutrophil count on atrial fibrillation. Our results provide insights into the role of systemic immune changes in the pathogenesis of arrhythmias.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Ethics statement

Ethical review and approval was not required for the study on human participants in accordance with the local legislation and institutional requirements. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

Author contributions

YuC, JY and SH contributed to conception and design of the study. YuC, LJ, XZ, YaC, LC, FZ, ZL and TF performed the statistical analysis. YuC and LL wrote the first draft of the manuscript. YuC, LL, XZ and YaC wrote sections of the manuscript All authors contributed to the article and approved the submitted version.

Funding

JY was supported by National Natural Science Foundation of China (Grant Number:82070409) and Natural Science Foundation of Zhejiang province (Grant Number:8LY19H020008); FZ was supported by Jinhua Science and Technology Bureau (Grant Number:2018-4-051); TF was supported by Jinhua Science and Technology Bureau (Grant Number: 2020-4-149).

Acknowledgments

We would also like to thank Professor Jianzhong Sheng and Professor Hefeng Huang for providing laboratory facilities.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2023.1041591/ full#supplementary-material

References

1. Grune J, Yamazoe M, Nahrendorf M. Electroimmunology and cardiac arrhythmia. *Nat Rev Cardiol* (2021) 18(8):547-64. doi: 10.1038/s41569-021-00520-9

2. Rudolph V, Andrie RP, Rudolph TK, Friedrichs K, Klinke A, Hirsch-Hoffmann B, et al. Myeloperoxidase acts as a profibrotic mediator of atrial fibrillation. *Nat Med* (2010) 16(4):470-4. doi: 10.1038/nm.2124

3. Grune J, Lewis A, Yamazoe M, Hulsmans M, Rohde D, Xiao L, et al. Neutrophils incite and macrophages avert electrical storm after myocardial infarction. *Nat Cardiovasc Res* (2022) 1(7):649–64. doi: 10.1038/s44161-022-00094-w

4. Monnerat G, Alarcon ML, Vasconcellos LR, Hochman-Mendez C, Brasil G, Bassani RA, et al. Macrophage-dependent IL-1beta production induces cardiac arrhythmias in diabetic mice. *Nat Commun* (2016) 7:13344. doi: 10.1038/ncomms13344

5. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages facilitate electrical conduction in the heart. *Cell* (2017) 169(3):510–22. doi: 10.1016/j.cell.2017.03.050

6. Liu Y, Shi Q, Ma Y, Liu Q. The role of immune cells in atrial fibrillation. J Mol Cell Cardiol (2018) 123:198–208. doi: 10.1016/j.yjmcc.2018.09.007

7. Lazzerini PE, Capecchi PL, Laghi-Pasini F, Boutjdir M. Autoimmune channelopathies as a novel mechanism in cardiac arrhythmias. *Nat Rev Cardiol* (2017) 14(9):521-35. doi: 10.1038/nrcardio.2017.61

 Toor IS, Ruckerl D, Mair I, Ainsworth R, Meloni M, Spiroski AM, et al. Eosinophil deficiency promotes aberrant repair and adverse remodeling following acute myocardial infarction. *JACC Basic Transl Sci* (2020) 5(7):665–81. doi: 10.1016/ jiacbts.2020.05.005

9. Liu J, Yang C, Liu T, Deng Z, Fang W, Zhang X, et al. Eosinophils improve cardiac function after myocardial infarction. *Nat Commun* (2020) 11(1):6396. doi: 10.1038/s41467-020-19297-5

10. Swirski FK, Nahrendorf M. Cardioimmunology: the immune system in cardiac homeostasis and disease. *Nat Rev Immunol* (2018) 18(12):733-44. doi: 10.1038/s41577-018-0065-8

11. Lazzerini PE, Laghi-Pasini F, Boutjdir M, Capecchi PL. Cardioimmunology of arrhythmias: the role of autoimmune and inflammatory cardiac channelopathies. *Nat Rev Immunol* (2019) 19(1):63–4. doi: 10.1038/s41577-018-0098-z

12. Shah AD, Denaxas S, Nicholas O, Hingorani AD, Hemingway H. Neutrophil counts and initial presentation of 12 cardiovascular diseases: A CALIBER cohort study. J Am Coll Cardiol (2017) 69(9):1160–9. doi: 10.1016/j.jacc.2016.12.022

13. Shah AD, Denaxas S, Nicholas O, Hingorani AD, Hemingway H. Low eosinophil and low lymphocyte counts and the incidence of 12 cardiovascular diseases: a CALIBER cohort study. *Open Heart* (2016) 3(2):e477. doi: 10.1136/ openhrt-2016-000477

14. Rienstra M, Sun JX, Magnani JW, Sinner MF, Lubitz SA, Sullivan LM, et al. White blood cell count and risk of incident atrial fibrillation (from the framingham heart study). *Am J Cardiol* (2012) 109(4):533–7. doi: 10.1016/j.amjcard.2011.09.049

15. Chen P, Chen J, Xie X, Zhu J, Xia L. Eosinophils in patients with lone atrial fibrillation. *Pacing Clin Electrophysiol* (2017) 40(8):955–8. doi: 10.1111/pace.13146

16. Suzuki A, Fukuzawa K, Yamashita T, Yoshida A, Sasaki N, Emoto T, et al. Circulating intermediate CD14++CD16+monocytes are increased in patients with atrial fibrillation and reflect the functional remodelling of the left atrium. *Europace* (2017) 19(1):40–7. doi: 10.1093/europace/euv422

17. Cosgun M, Gunes Y, Sincer I, Mansiroglu AK. Assessment of the hemogram parameters in patients with paroxysmal supraventricular tachycardia: a retrospective study. *Rev Assoc Med Bras (1992)* (2020) 66(10):1371–5. doi: 10.1590/1806-9282.66.10.1371

18. Aydin M, Yildiz A, Yuksel M, Polat N, Aktan A, Islamoglu Y. Assessment of the neutrophil/lymphocyte ratio in patients with supraventricular tachycardia. *Anatol J Cardiol* (2016) 16(1):29–33. doi: 10.5152/akd.2015.5927

19. Holmes MV, Ala-Korpela M, Smith GD. Mendelian randomization in cardiometabolic disease: challenges in evaluating causality. *Nat Rev Cardiol* (2017) 14(10):577–90. doi: 10.1038/nrcardio.2017.78

20. O'Donnell CJ, Sabatine MS. Opportunities and challenges in mendelian randomization studies to guide trial design. *JAMA Cardiol* (2018) 3(10):967. doi: 10.1001/iamacardio.2018.2863

21. Davies NM, Holmes MV, Davey SG. Reading mendelian randomisation studies: a guide, glossary, and checklist for clinicians. *BMJ* (2018) 362:k601. doi: 10.1136/bmj.k601

22. Chen MH, Raffield LM, Mousas A, Sakaue S, Huffman JE, Moscati A, et al. Trans-ethnic and ancestry-specific blood-cell genetics in 746, 667 individuals from 5 global populations. *Cell* (2020) 182(5):1198–213. doi: 10.1016/j.cell.2020.06.045

23. Roselli C, Chaffin MD, Weng LC, Aeschbacher S, Ahlberg G, Albert CM, et al. Multi-ethnic genome-wide association study for atrial fibrillation. *Nat Genet* (2018) 50 (9):1225–33. doi: 10.1038/s41588-018-0133-9

24. Palmer TM, Lawlor DA, Harbord RM, Sheehan NA, Tobias JH, Timpson NJ, et al. Using multiple genetic variants as instrumental variables for modifiable risk factors. *Stat Methods Med Res* (2012) 21(3):223–42. doi: 10.1177/0962280210394459

25. Nielsen JB, Thorolfsdottir RB, Fritsche LG, Zhou W, Skov MW, Graham SE, et al. Biobank-driven genomic discovery yields new insight into atrial fibrillation biology. *Nat Genet* (2018) 50(9):1234–9. doi: 10.1038/s41588-018-0171-3

26. Larsson SC, Mason AM, Back M, Klarin D, Damrauer SM, Michaelsson K, et al. Genetic predisposition to smoking in relation to 14 cardiovascular diseases. *Eur Heart J* (2020) 41(35):3304–10. doi: 10.1093/eurheartj/ehaa193

27. Bowden J, Del GMF, Minelli C, Davey SG, Sheehan N, Thompson J. A framework for the investigation of pleiotropy in two-sample summary data mendelian randomization. *Stat Med* (2017) 36(11):1783-802. doi: 10.1002/sim.7221

28. Bowden J, Davey SG, Burgess S. Mendelian randomization with invalid instruments: effect estimation and bias detection through egger regression. *Int J Epidemiol* (2015) 44(2):512–25. doi: 10.1093/ije/dyv080

29. Bowden J, Davey SG, Haycock PC, Burgess S. Consistent estimation in mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol* (2016) 40(4):304–14. doi: 10.1002/gepi.21965

30. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from mendelian randomization between complex traits and diseases. *Nat Genet* (2018) 50(5):693–8. doi: 10.1038/s41588-018-0099-7

31. Hemani G, Tilling K, Davey SG. Correction: Orienting the causal relationship between imprecisely measured traits using GWAS summary data. *PloS Genet* (2017) 13 (12):e1007149. doi: 10.1371/journal.pgen.1007149

32. Savage JE, Jansen PR, Stringer S, Watanabe K, Bryois J, de Leeuw CA, et al. Genome-wide association meta-analysis in 269, 867 individuals identifies new genetic and functional links to intelligence. *Nat Genet* (2018) 50(7):912–9. doi: 10.1038/s41588-018-0152-6

33. Lee S, Lee DK. What is the proper way to apply the multiple comparison test? *Korean J Anesthesiol* (2018) 71(5):353-60. doi: 10.4097/kja.d.18.00242

34. Burgess S, Davey SG, Davies NM, Dudbridge F, Gill D, Glymour MM, et al. Guidelines for performing mendelian randomization investigations. *Wellcome Open Res* (2019) 4:186. doi: 10.12688/wellcomeopenres.15555.1

35. Patin E, Hasan M, Bergstedt J, Rouilly V, Libri V, Urrutia A, et al. Natural variation in the parameters of innate immune cells is preferentially driven by genetic factors. *Nat Immunol* (2018) 19(3):302–14. doi: 10.1038/s41590-018-0049-7

36. Le Cornet C, Schildknecht K, Rossello CA, Fortner RT, Gonzalez MS, Katzke VA, et al. Circulating immune cell composition and cancer risk: A prospective study using epigenetic cell count measures. *Cancer Res* (2020) 80(9):1885–92. doi: 10.1158/0008-5472.CAN-19-3178

37. Kwok CS, Rashid M, Beynon R, Barker D, Patwala A, Morley-Davies A, et al. Prolonged PR interval, first-degree heart block and adverse cardiovascular outcomes: a systematic review and meta-analysis. *Heart* (2016) 102(9):672–80. doi: 10.1136/ heartjnl-2015-308956

38. Hashimoto D, Chow A, Noizat C, Teo P, Beasley MB, Leboeuf M, et al. Tissueresident macrophages self-maintain locally throughout adult life with minimal contribution from circulating monocytes. *Immunity* (2013) 38(4):792–804. doi: 10.1016/j.immuni.2013.04.004

39. Barra SN, Providencia R, Paiva L, Nascimento J, Marques AL. A review on advanced atrioventricular block in young or middle-aged adults. *Pacing Clin Electrophysiol* (2012) 35(11):1395–405. doi: 10.1111/j.1540-8159.2012.03489.x

40. Mesirca P, Nakao S, Nissen SD, Forte G, Anderson C, Trussell T, et al. Intrinsic electrical remodeling underlies atrioventricular block in athletes. *Circ Res* (2021) 129 (1):e1–e20. doi: 10.1161/CIRCRESAHA.119.316386

41. Wang DW, Viswanathan PC, Balser JR, George AJ, Benson DW. Clinical, genetic, and biophysical characterization of SCN5A mutations associated with atrioventricular conduction block. *Circulation* (2002) 105(3):341–6. doi: 10.1161/hc0302.102592

42. Alboni P, Holz A, Brignole M. Vagally mediated atrioventricular block: pathophysiology and diagnosis. *Heart* (2013) 99(13):904-8. doi: 10.1136/heartjnl-2012-303220

43. Zouggari Y, Ait-Oufella H, Bonnin P, Simon T, Sage AP, Guerin C, et al. B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. *Nat Med* (2013) 19(10):1273–80. doi: 10.1038/nm.3284

44. Nevers T, Salvador AM, Velazquez F, Ngwenyama N, Carrillo-Salinas FJ, Aronovitz M, et al. Th1 effector T cells selectively orchestrate cardiac fibrosis in nonischemic heart failure. *J Exp Med* (2017) 214(11):3311–29. doi: 10.1084/jem.20161791

45. Camm AJ, Lip GY, De Caterina R, Savelieva I, Atar D , Hohnloser SH, et al. 2012 focused update of the ESC guidelines for the management of atrial fibrillation: an update of the 2010 ESC guidelines for the management of atrial fibrillation–developed with the special contribution of the European heart rhythm association. *Europace* (2012) 14(10):1385–413. doi: 10.1093/europace/eus305

46. Hu YF, Chen YJ, Lin YJ, Chen SA. Inflammation and the pathogenesis of atrial fibrillation. *Nat Rev Cardiol* (2015) 12(4):230–43. doi: 10.1038/nrcardio.2015.2

47. Shao Q, Chen K, Rha SW, Lim HE, Li G, Liu T. Usefulness of Neutrophil/ Lymphocyte ratio as a predictor of atrial fibrillation: A meta-analysis. *Arch Med Res* (2015) 46(3):199–206. doi: 10.1016/j.arcmed.2015.03.011

48. Ishii Y, Schuessler RB, Gaynor SL, Yamada K, Fu AS, Boineau JP, et al. Inflammation of atrium after cardiac surgery is associated with inhomogeneity of atrial conduction and atrial fibrillation. *Circulation* (2005) 111(22):2881–8. doi: 10.1161/CIRCULATIONAHA.104.475194

49. Im SI, Shin SY, Na JO, Kim YH, Choi CU, Kim SH, et al. Usefulness of neutrophil/lymphocyte ratio in predicting early recurrence after radiofrequency catheter ablation in patients with atrial fibrillation. *Int J Cardiol* (2013) 168(4):4398–400. doi: 10.1016/j.ijcard.2013.05.042

50. Glinge C, Lahrouchi N, Jabbari R, Tfelt-Hansen J, Bezzina CR. Genome-wide association studies of cardiac electrical phenotypes. *Cardiovasc Res* (2020) 116(9):1620–34. doi: 10.1093/cvr/cvaa144

51. Lin H, van Setten J, Smith AV, Bihlmeyer NA, Warren HR, Brody JA, et al. Common and rare coding genetic variation underlying the electrocardiographic PR interval. *Circ Genom Precis Med* (2018) 11(5):e2037. doi: 10.1161/CIRCGEN.117.002037

52. Arking DE, Pulit SL, Crotti L, van der Harst P, Munroe PB, Koopmann TT, et al. Genetic association study of QT interval highlights role for calcium signaling pathways in myocardial repolarization. *Nat Genet* (2014) 46(8):826–36. doi: 10.1038/ng.3014

53. Barc J, Tadros R, Glinge C, Chiang DY, Jouni M, Simonet F, et al. Genome-wide association analyses identify new brugada syndrome risk loci and highlight a new mechanism of sodium channel regulation in disease susceptibility. *Nat Genet* (2022) 54 (3):232–9. doi: 10.1038/s41588-021-01007-6

54. Lahrouchi N, Tadros R, Crotti L, Mizusawa Y, Postema PG, Beekman L, et al. Transethnic genome-wide association study provides insights in the genetic architecture and heritability of long QT syndrome. *Circulation* (2020) 142(4):324–38. doi: 10.1161/CIRCULATIONAHA.120.045956