



Transient Receptor Potential Channels and Inflammatory Bowel Disease

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The transient receptor potential (TRP) cation channels are present in abundance across the gastrointestinal (GI) tract, serving as detectors for a variety of stimuli and secondary transducers for G-protein coupled receptors. The activation of TRP channels triggers neurogenic inflammation with related neuropeptides and initiates immune reactions by extra-neuronally regulating immune cells, contributing to the GI homeostasis. However, under pathological conditions, such as inflammatory bowel disease (IBD), TRP channels are involved in intestinal inflammation. An increasing number of human and animal studies have indicated that TRP channels are correlated to the visceral hypersensitivity (VHS) and immune pathogenesis in IBD, leading to an exacerbation or amelioration of the VHS or intestinal inflammation. Thus, TRP channels are a promising target for novel therapeutic methods for IBD. In this review, we comprehensively summarize the functions of TRP channels, especially their potential roles in immunity and IBD. Additionally, we discuss the contradictory findings of prior studies and offer new insights with regard to future research.

Keywords: transient receptor potential channels, gastrointestinal tract, neurogenic inflammation, immune cells, inflammatory bowel disease

INTRODUCTION

Inflammatory bowel disease (IBD) is a chronic relapsing GI inflammatory disorder, comprising of Crohn's disease (CD) and ulcerative colitis (UC). It is acknowledged that IBD is related to inappropriate immunity, commensal bacteria, genetics, and environmental factors. The exact pathogenesis of IBD, however, remains unknown (1). Nowadays, various receptors in the gastrointestinal (GI) tract are proposed to play a role in the pathophysiology of IBD, amongst which transient receptor potential (TRP) ion channels have been identified and are considered to be potentially effective. TRP channels are polymodal ion channels that serve as sensors for chemical noxious and physical stimuli. These channels are widely distributed in the GI tract and exert various effects, contributing to the somatic and visceral nociception and the maintenance of physiological function of the GI tract (2). The activation of TRP channels can evoke neurogenic inflammation, namely the inflammation initiated by the local release of immunomodulatory neuropeptides, including calcitonin-gene-related peptide (CGRP) and substance P (SP) released by unmyelinated afferent neurons (3, 4). Some TRP channels are also expressed in multiple immune cells, and are primarily responsible for modulating actions, such as cytokine release, cell migration, and phagocytic activity (2). Therefore, numerous studies have indicated that TRP channels are mainly

involved in the visceral hypersensitivity (VHS) and immune pathogenesis of IBD due to their comprehensive functions of sensors and immunomodulators. Different subtypes of TRP channels seem to have distinct effects. Here, we briefly review the correlation between TRP channels and IBD with a focus on TRPV1, TRPA1, TRPV4, TRPM2, and TRPM8, which have been documented to be the most relevant TRP channels in IBD.

TRP CHANNELS IN THE GI TRACT AND RELATED NEUROPEPTIDES

In the GI tract, TRP channels are mainly expressed on the extrinsic primary afferent nerves with some on epithelial, endocrine cells, and intrinsic enteric neurons (5, 6). Intriguingly, 97% of TRPA1-positive (TRPA1⁺) afferents co-express TRPV1, and 30% of the TRPV1-positive (TRPV1⁺) neurons co-express TRPA1, hinting at the potential interaction between the two channels (5, 7). Capsaicin is a significant agonist for TRPV1 with an exquisite selectivity and allyl isothiocyanate (AITC), the pungent ingredient in garlic, is the prototypical agonist for TRPA1 (8, 9). TRPV4 colocalizes with TRPV1, TRPA1, and protease-activated receptors 2 (PAR-2) in the GI tract, in response to strong acidosis, hypotonicity, warmth, and mechanical stimuli (10, 11). TRPM2 is sensitive to heat stimulus while TRPM8 is essential to cold-induced pain (12). Most of TRP channels are non-selective cation channels and show the permeability to calcium ion (Ca²⁺). Upon stimulation, TRP channels in afferents can lead to autonomic reflex responses by transmitting signals to the central nervous system. Meanwhile, TRP channels can transduce sensory signal of G-protein coupled receptors (GPCRs) based on the phosphorylation sites in N-terminus for serine and threonine protein kinases, such as protein kinase A (PKA) and protein kinase C (PKC) (11, 13).

Additionally, TRP channels in the GI tract can mediate the crosstalk between the nervous and immune systems by modulating the release of neuropeptides. TRPV1, TRPA1, and TRPV4 are especially often found to colocalize with CGRP and SP (6, 14).

CGRP, which is generated from the alternative RNA processing of the gene for calcitonin, serves as a potent peptide vasodilator and is involved in the transmission of nociception (15). CGRP plays a protective role in the inflammation and inhibits the capacity of immune cells. For dendritic cells (DCs) and macrophages, CGRP could restrain their ability in the presentation of antigens and the secretion of pro-inflammatory cytokines, such as tumor necrosis factor alpha (TNF- α) (16, 17). CGRP also downregulated DCs' responses to Toll-like receptor 4 (TLR4), a receptor for lipopolysaccharides (LPS) which is an abundant outer wall glycolipid of Gram-negative bacteria (18). CGRP can exert an inhibitory effect on the activation and chemotaxis of neutrophils (19), and inhibit neutrophil-mediated killing of bacteria mainly through suppressing the activity of the bactericidal enzyme myeloperoxidase (MPO) in a dose-dependent manner (20). CGRP was found to restrain group 2 innate lymphoid cells proliferation and type 2 innate immune responses (21), and be required for the induction

of protective innate type 17 immunity after the activation of cutaneous TRPV1⁺ neurons (22). Furthermore, CGRP could induce the upregulation of interleukin (IL)-10 and was beneficial in preserving mucosal integrity and limiting tissue damage (16). These observations demonstrate the negative regulatory role of CGRP in innate immunity and the benefits of CGRP in the GI tract. Conversely, GRCP was reported to be capable of stimulating T-cell migration and promoting the release of interferon gamma (IFN- γ) and IL-2 from T-helper cells (23, 24). In IBD patients, the expression of CGRP in the colonic mucosa was significantly increased and was closely associated with the severity of disease (25, 26). Therefore, CGRP might also play a part in the pro-inflammatory process.

SP belongs to tachykinin family. The receptor for SP is the neurokinin 1 receptor (NK-1R) (15). Similar to CGRP, SP serves as a potent vasodilator and the SP-induced vasodilatation is based on nitric oxide (NO) release (27). Of interest, high-dose CGRP was reported to restrain the SP-evoked vasodilation but facilitate SP-evoked plasma protein extravasation (28), suggesting a crosstalk between CGRP and SP. The expression of SP and NK-1R has been well-documented in DCs, monocytes, eosinophils, neutrophils, mast cells, natural killer cells, and T cells, enabling SP to regulate functions of different types of immune cell (29). SP also modulated the immune response to microbial infection (29). It was recently demonstrated that SP could promote the migration and activation of mast cells, inducing the release of multiple pro-inflammatory cytokines and chemokines (30). Noteworthy, SP directly caused the secretion of IL-8 in human colonic epithelial cell lines (31), hinting at the potential pro-inflammatory role of SP in intestine. SP was detected to be elevated in tissue extracts from the colon and rectum of IBD patients, and the level of SP was correlated with disease activity (32). However, in animal studies, SP ameliorated dextran sulfate sodium (DSS)-induced colitis by promoting the enrichment of M2 macrophages and regulatory T cells, or maintaining barrier structure and regulating immune response (33, 34). Such results remind us of the possible protective effect of SP in colitis. Taking these contradictory observations with regard to the properties of CGRP and SP into consideration, we can conclude that the neurogenic inflammation triggered by TRP channels has bidirectional functions on immunity and colitis. However, the exact function of neuropeptides on a certain physical or pathological condition has not yet been discovered and further studies are required.

TRP CHANNELS IN IMMUNE CELLS

Besides the roles in nervous system of the GI tract, TRP channels are also expressed in immune cells and directly contribute to immune responses. In bone marrow-derived macrophages, the TRPV1 expression was increased and intracellular Ca²⁺-transients were triggered after oxidized low-density lipoprotein (ox-LDL)-stimulation (35). TRPV1 could dose-dependently modulate the level of inducible NO synthase in stimulated peritoneal macrophages through the inhibition of nuclear factor kappa B (NF- κ B), thus influencing the secretion of

pro-inflammatory cytokines involved in this pathway (2). In a sepsis model, the LPS-stimulated peritoneal macrophages showed an impaired phagocytosis when TRPV1 was knocked-out (36), suggesting the putative role of TRPV1 to potentiate macrophages. In CD4⁺ T cells, TRPV1 was associated with T cell receptor (TCR) and facilitated TCR-induced Ca²⁺ inflow (37), and the activity of CD4⁺ T cells was impaired via the inhibition of TRPV1 (38). The activation of TRPV1 was also reported to enhance leukocyte rolling and adhesion (39). These data indicated the possible pro-inflammatory properties of TRPV1 in immune cells. Intriguingly, TRPV1 activation can trigger the production of the endocannabinoid anandamide, which increases the level of regulatory CX3CR1 (hi) macrophages in the gut and enhances their immunosuppressive activity (40). In DCs, TRPV1 mediates the downregulation of TLR4/NF- κ B signaling pathway that leads to the maturation of DCs (41). A recent research concerning lethal *Staphylococcus aureus* pneumonia stated that the activation of TRPV1⁺ nociceptors by capsaicin could suppress cytokine release, inhibit the recruitment and surveillance of neutrophils, and alter lung $\gamma\delta$ T cell numbers; thus impairing lung bacterial clearance (42). As discussed above, TRPV1 is an important immunomodulator that regulates the activation and function of immune cells.

As for TRPA1, its expression was increased in stimulated T cells, and TRPA1 was vital for the T cell activation and release of cytokines like TNF- α , IFN- γ and IL-2 (43). TRPA1 also expresses in mast cells and DCs (44). In TRPA1-knockout (*Trpa1*^{-/-}) mice, mast cells, leukocytes, and T cells, together with the expression of IL-1 β , IL-6, IL-17, IL-22, and IL-23 were decreased in the lesions of skin (45), indicating the ability of TRPA1 to induce inflammation through these immune cells. In addition to its pro-inflammatory function, the activation of TRPA1 could suppress the pro-inflammatory effect of LPS-stimulated peritoneal macrophages by decreasing the level of NO, which is an abundant pro-inflammatory mediator (46). Taken together, TRPA1 has the ability to regulate immune cells in diverse manner. The crosstalk between TRP channels and bacteria is noteworthy. It was discovered that LPS interacted with TLR4 on the TRPV1⁺ afferent neurons. This then activated or sensitized TRPV1 via its phosphorylation binding sites through PKC, thus resulting in an increased release of CGRP (17, 47). Antagonists for TRPV1 and CGRP could reverse LPS-induced motility disturbance of the intestine (48). Another study showed that a probiotic bacterium named *Lactobacillus reuteri* and its condition medium dose-dependently reduced the capsaicin- and distension- evoked firing of jejunal spinal afferents in mice (49), revealing the engagement of afferents in bacteria-induced GI sensory-motor dysfunction. TRPA1 in nociceptive neurons could be sensitized by LPS in a TLR4-independent manner during inflammation, causing pain, CGRP release, and vasodilation (50). Therefore, it was hypothesized that TRP channels may be able to directly or indirectly interact with microbiota or their products in the gut, thus influencing the release of neuropeptides and contributing to the maintenance of gut homeostasis.

The activation of TRPV4 increased intracellular Ca²⁺ concentration in LPS-treated macrophages and potentiated macrophage (51), while downregulation of TRPV4 subsequently

impaired the phagocytosis of macrophages (52). In neutrophils, TRPV4 was essential for inflammatory responses, such as the neutrophil adhesion, chemotaxis, and formation of reactive oxygen species (53). TRPV4-mediated Ca²⁺ inflow in T cells was also capable of inducing the proliferation of T cells and the secretion of TNF- α , IFN- γ , and IL-2 *in vitro* (54). Recent data revealed that TRPV4 could promote the phagocytosis of mouse CD11c⁺ bone marrow-derived cells (55). These findings clearly highlight the critical pro-inflammatory role of TRPV4 in immune cells.

Regarding TRPM2, it was demonstrated that the lack of this channel in LPS-stimulated monocytes cell line reduced the release of TNF- α , IL-6, IL-8, and IL-10 (56). TRPM2-associated Ca²⁺ signaling was essential in the transmigration and cytotoxicity of neutrophils (57, 58), the proliferation of T cells, and the release of pro-inflammatory cytokines (59). For TRPM8, the activation by menthol in murine peritoneal macrophages increased IL-10 expression and decreased TNF- α release, thus exerting an anti-inflammatory effect (60). TRPM8-knockout (*Trpm8*^{-/-}) peritoneal macrophages exhibited an impaired phagocytic activity while the phagocytosis was enhanced in WT peritoneal macrophages after the activation of TRPM8 (60). Consistently, the activation of TRPM8 was reported to restrain the release of pro-inflammatory mediators in monocytes and lymphocytes, and *Trpm8*^{-/-} CD11c⁺ DCs showed hyperinflammatory responses to TLR-stimulation (61, 62). In T cells, the inhibition of TRPM8 suppressed murine T-cell activation and the release of IL-2 and IL-6 (63). Overall, TRPM2 appears to potentiate inflammatory effects of immune cells while TRPM8 often performs anti-inflammatory roles.

TRP CHANNELS IN INFLAMMATORY VISCERAL HYPERSENSITIVITY OF IBD

Due to their immunomodulatory function via neuropeptides and immune cells, TRP channels are associated with GI immunity and inflammation. Notably, it is found that their expression has been altered in IBD patients and colitis models (Table 1), suggesting an involvement of TRP channels in IBD. In particular, IBD patients are associated with a visceral hypersensitivity (VHS), which is featured of an aberrant and chronic visceral pain (5, 12). As visceral nociceptors, TRP channels are proposed to be responsible for VHS in IBD. Since TRP channels serve as secondary transducers for GPCRs, some mediators that act on GPCRs subsequently activate or sensitize TRP channels, resulting in aberrant sensation. Through this mechanism, pro-inflammatory mediators secreted during colitis, such as bradykinin, serotonin (5-hydroxytryptamine, 5-HT), cytokines, adenosine triphosphate, prostaglandins, and epinephrine can lead to the inflammatory VHS (5). A number of researches have been conducted to explore the definite role of TRP channels in colitis and VHS (Table 2).

TRPV1

TRPV1 channel is closely linked to VHS. It was found that some patients with quiescent IBD still complained about

TABLE 1 | The expression of TRP channels in colonic tissue of IBD patients and colitis models.

	TRPV1	TRPA1	TRPV4	TRPM2	TRPM8
IBD patients					
UC	UP (64–68) NS (69) DOWN (70, 71)	UP (68)	UP (70, 72) NS (73)	NA	NA
CD	UP (64–66) DOWN (68, 71)	UP (68, 71, 74, 75)	UP (72, 76) NS (73)	NA	UP (77)
Animal models					
DSS-treated mice	UP (68, 78–80) NS (71)	UP (71)	UP (73, 81)	NA	UP (77, 82)
DSS-treated rats	UP (83)	NA	NA	NA	NA
TNBS-treated mice	UP (84)	NA	UP (72)	NA	UP (77, 82)
DNBS-treated mice	NA	UP (85)	NA	NA	NA
TNBS-treated rats	UP (86) NS (87)	UP (88, 89)	NA	UP (90)	NA
Mustard oil-treated mice	NS (91)	UP (91)	NA	NA	NA

UC, ulcerative colitis; CD, Crohn's disease; DSS, dextran sulfate sodium; TNBS, 2,4,6-trinitrobenzenesulfonic acid; DNBS, dinitrobenzene sulfonic acid; UP, upregulated; DOWN, downregulated; NS, no significant difference; NA, not available.

TABLE 2 | The function of TRP channels in VHS of colitis models.

Pro-VHS function			
TRP channel	Colitis model	Result	References
TRPV1	DSS-treated mice	Increased VHS which could be enhanced by the agonist for TRPV1	(92)
	DSS- and TNBS- treated rat	Increased VHS which could be relieved by the antagonist for TRPV1	(83, 86, 87, 93)
	Trpv1 ^{-/-} mice with DSS-induced colitis	Decreased VHS compared to WT mice	(78)
TRPA1	DSS-treated mice	Increased VHS which could be enhanced by the agonist or be relieved by the antagonist for TRPA1	(94)
	TNBS-treated rat	Increased VHS which could be enhanced by the agonist or be relieved by the antagonist for TRPA1	(88, 89, 95, 96)
	Trpa1 ^{-/-} mice with TNBS-induced colitis	Decreased VHS compared to WT mice	(97, 98)
TRPV4	TNBS-treated mice	Increased VHS which could be relieved by the antagonist for TRPV4	(72)
	Trpv4 ^{-/-} mice	Decreased VHS compared to WT mice	(99)
TRPM2	TNBS-treated rat	Increased VHS which could be relieved by the antagonist for TRPM2	(90)
	Trpm2 ^{-/-} mice with TNBS-induced colitis	Decreased VHS compared to WT mice	(90)
TRPM8	DSS- and TNBS- treated mice	Increased VHS which could be enhanced by the agonist or be relieved by the antagonist for TRPM8	(82)
	Trpm8 ^{-/-} mice	VHS was only decreased under higher level of stimuli compared to Trpv1 ^{-/-} and Trpv4 ^{-/-} mice	(100)
Anti-VHS function			
TRPM8	WT mice	The function of TRPV1 and TRPA1 was inhibited by TRPM8 activation	(101)
	TNBS-treated rat	Increased VHS which could be relieved by the agonist for TRPM8	(102)

VHS, visceral hypersensitivity; DSS, dextran sulfate sodium; TNBS, 2,4,6-trinitrobenzenesulfonic acid; WT, wild-type.

abdominal pain, and the severity of their symptoms was correlated to the increased TRPV1⁺ fibers in colonic mucosa (64). In animal studies, the behavioral responses to intracolonic capsaicin administration and the expression of spinal cord neuronal c-Fos, which is a marker of neuronal excitation, were increased in DSS-treated mice (92). Yang et al. (83) reported that an oral administration of curcumin, which is

clinically valuable for the treatment of IBD (103), in DSS-treated rats could significantly ameliorate visceral hyperalgesia through inhibiting phosphorylation of TRPV1, indicating a nociceptive effect of TRPV1. Likewise, Phillis et al. (93) revealed that TRPV1 antagonist remarkably reduced the mechanosensory response to the stimulus in a dose-dependent manner in rats with DSS-induced colitis. Visceral hyperalgesia and increased

visceromotor response (VMR) were also confirmed in rats with 2,4,6-trinitrobenzenesulfonic acid (TNBS)-induced colitis (86, 87), while TRPV1 antagonist (JYL1421) could prevent and relieve the VHS (86). Additional studies exhibited that TRPV1-knockout (*Trpv1*^{-/-}) mice conferred a resistance to colorectal distension (78, 104), and VHS was enhanced by inflammatory mediators, such as bradykinin, 5-HT, histamine, and prostaglandin E2 (PGE2) in WT mice but not in *Trpv1*^{-/-} mice (12). SP was demonstrated to enhance the sensitivity and function of TRPV1 in DSS-induced colitis and *in vitro* (78), suggesting the engagement of neuropeptides in VHS. Noteworthy, the augmented activity of pelvic nerve afferents after TRPV1 activation in DSS-treated rats was more prominent on the first day post DSS-treatment, in comparison to the eighth day (92). Similarly, the levels of TRPV1 and TRPA1 messenger RNA (mRNA) in mice were upregulated in mustard oil (MO)-induced colitis within 6 h but decreased 24- and 72-h after MO-injection (91). Therefore, it can be hypothesized that the excitatory mechanism modulated by TRPV1 mainly particulate during early stage of experimental colitis.

TRPA1

TRPA1 could contribute to colorectal contraction and enhanced VMR to intracolonic AITC, which were detectable in TNBS-induced colitis. These actions could be suppressed by intrathecal pretreatment with a TRPA1 antisense oligodeoxynucleotide, and were absent in *Trpa1*^{-/-} mice (88, 89, 97, 98). Likewise, AITC enhanced the sensitivity of colon and the expression of c-Fos in spinal cord of DSS-treated mice (94). During TNBS-induced colitis, the production of hydrogen peroxide (H₂O₂) was enhanced due to the infiltration of white blood cells and the presence of oxidative stress. The increased H₂O₂ then activated TRPA1 and led to the hypersensitivity of VMR (95). The aberrant GI motility might result from the effects of PGE2 induced by TRPA1 activation (105). Similarly, inflammatory mediators, such as bradykinin and 5-HT could lead to an increased visceral mechano-sensitivity in a TRPA1-associated manner (97, 106). These data suggest a close link between TRPA1 and pro-inflammatory cytokines, both of which contribute to visceral hyperalgesia. Of interest, Vermeulen et al. (96) reported that a combined application of antagonists for TRPV1 and TRPA1 could reduce the VMR more effectively in TNBS-induced colitis, in comparison to targeting either TRPV1 or TRPA1 alone. Such evidence appears to provide an inspiring therapeutic method for inflammatory VHS.

TRPV4

TRPV4 is vital for a mechanically-evoked visceral pain in the GI tract (76). It was found that TRPV4 co-expressed with PAR-2 and pretreatment of PAR-2 agonist enhanced TRPV4 activity and hypersensitivity, while the inhibition of PKA and PKC restrained this effect (99, 107). Also, 5-HT and histamine improved TRPV4-induced hypersensitivity for colorectal distention in mice (108). These results indicate the responses of TRPV4 to inflammatory mediators through GPCR signaling pathway. A selective blockade of TRPV4 was subsequently evident to alleviate intestinal inflammation and pain in TNBS-treated mice (72).

Therefore, the pro-hypersensitivity function of TRPV4 during colitis is relatively clear.

TRPM2

In TNBS-induced rat colitis, the VMR was enhanced and could be ameliorated by an oral administration of TRPM2 antagonist (econazole) (90). However, in TRPM2-knockout (*Trpm2*^{-/-}) mice, the TNBS-induced VMR was less severe compared to WT mice (90), showing a potential facilitating role of TRPM2 in VHS.

TRPM8

Menthol, serving as the agonist for TRPM8, has been applied to relieve abdominal discomfort and pain in traditional Chinese medicine, suggesting that TRPM8 activation can diminish visceral pain perception (12). In TNBS-induced colitis, the colonic mechano-hypersensitivity was remarkably suppressed by a combined adoption of peppermint and caraway oil, which are agonists for TRPM8 (102). Harrington et al. (101) demonstrated that TRPM8 activation restrained the downstream chemosensory and mechanosensory actions of TRPA1 and TRPV1 to agonists in colonic afferents, stating the potential function of TRPM8 for inhibiting TRPV1 and TRPA1. In contrast, it was showed that the TRPM8 agonist (WS-12) enhanced visceral pain response while a pretreatment of TRPM8 antagonist inhibited the hypersensitivity (82). Another study reported that in *Trpm8*^{-/-} mice, VMR only decreased when the pressure level of colorectal distension was quite high; but in *Trpv1*^{-/-} and *Trpv4*^{-/-} mice, VMR was remarkably decreased in all pressure ranges (100). Of note, both bradykinin and histamine were found to suppress TRPM8 mainly via the G-protein subunit G α which inhibited ion channel activity of TRPM8 (109), indicating the ability of inflammatory mediators to desensitize TRPM8 and inhibit its function. Such a mechanism may account for the TRPM8-associated enhanced-VHS during inflammation.

TRP CHANNELS IN IMMUNE PATHOGENESIS OF IBD

In addition to the roles in inflammatory VHS, the potential engagement of TRP channels in the immune pathogenesis of IBD has been highlighted in human and animal studies (Table 3).

TRPV1

The TRPV1⁺ fibers were increased in the colonic mucosa of IBD patients, along with non-neuronal TRPV1 immunoreactivity (65, 66). Further study confirmed an increased expression of TRPV1 in inflamed tissue of active UC patients compared with non-inflamed tissue, being associated with a relapse and continuous activity of disease (64, 67). However, a downregulated expression of TRPV1 was also revealed in colonic biopsies from UC and CD patients (68, 70, 71), and Rizopoulos et al. (70) found no significant correlation between TRPV1 expression and clinical features in UC patients. In experimental colitis models, TRPV1 expression was also found to be altered (68, 78–80, 83, 84, 86) (Table 1). Kihara et al. (110) subcutaneously injected noxious-dose capsaicin into neonatal rats to chemically denervate the TRPV1 channel, revealing that the denervated rats exhibited a

TABLE 3 | The function of TRP channels in pathophysiology of colitis models.

Pro-inflammatory function			
TRP channel	Colitis model	Result	References
TRPV1	DSS-treated mice	Chemically denervation of TRPV1, the antagonist for TRPV1, and the Trpa1-knockout alleviated colitis	(110–114)
	TNBS-treated rat	The antagonist for TRPV1 alleviated colitis	(115)
	TLR-4 ^{-/-} mice with TNBS-induced colitis	Downregulated TRPV1 expression and alleviated colitis compared to WT mice	(84)
	Toxin-A treated isolated rat ileal segment	Aggravated inflammation which could be enhanced by the agonist or be alleviated by the antagonist for TRPV1	(116)
	T cell-transfer mice colitis	Genetic or pharmacological inhibition of TRPV1 in T cell or colonic tissue resulted in less severe colitis	(117, 118)
TRPA1	DSS-treated mice	The Trpa1-knockout and the antagonist for TRPA1 alleviated colitis	(119)
	TNBS-treated mice	The Trpa1-knockout and the antagonist for TRPA1 alleviated colitis	(119)
TRPV4	DSS-treated mice	The agonist for TRPV4 aggravated colitis and the Trpv4-knockout alleviated colitis	(73, 81)
	TNBS-treated mice	The antagonist for TRPV4 alleviated colitis	(72)
TRPM2	DSS-treated mice	The Trpm2-knockout alleviated colitis	(120)
Anti-inflammatory function			
TRPV1	DSS-treated rat	The agonist for TRPV1 alleviated colitis and chemically denervation of TRPV1 aggravated colitis	(121)
	TNBS-treated rat	The agonist for TRPV1 alleviated colitis	(122, 123)
	DNBS-treated mice	The Trpv1-knockout aggravated colitis	(124)
	Oxazolone-treated mice	Chemically denervation of TRPV1 aggravated colitis	(125)
	Iodoacetamide-treated rat	Chemically denervation of TRPV1 aggravated colitis	(126)
	Formalin-treated rabbit	Chemically denervation of TRPV1 aggravated colitis	(127)
TRPA1	DSS-treated mice	The Trpa1-knockout and the antagonist for TRPA1 aggravated colitis; the agonist for TRPA1 alleviated colitis	(71, 85)
	T cell-transfer mice colitis	TRPV1 ⁺ TRPA1 ⁻ T cells induced more severe colitis compared to TRPV1 ⁺ TRPA1 ⁺ T cells	(68)
	TNBS-treated mice	The Trpa1-knockout aggravated the fibrosis in colitis	(57, 67)
TRPM8	DNBS-treated mice	The agonist for TRPA1 alleviated colitis	(85)
	TNBS-treated mice	The agonist for TRPM8 alleviated colitis	(77)
	DSS-treated mice	The Trpm8-knockout aggravated colitis and the agonist for TRPM8 alleviated colitis; adoptive transfer of TRPM8 ^{-/-} macrophages in mice induced more severe colitis compared to WT macrophages	(60, 77)

DSS, dextran sulfate sodium; TNBS, 2,4,6-trinitrobenzenesulfonic acid; DNBS, dinitrobenzene sulfonic acid; WT, wild-type.

lower severity of DSS-induced colitis compared with the control group. Similarly, it was showed that an oral administration of capsaizepine (CPZ), which is a specific antagonist for TRPV1, significantly reduced the overall macroscopic epithelial damage in mice colonic tissue after intraperitoneal DSS-administration (111). In *Trpv1*^{-/-} mice, the DSS-induced colitis was less severe (112, 113), and a DSS-associated upregulation of SP-positive fibers was reduced (114), demonstrating a crosstalk between TRPV1 and neurogenic inflammation in colitis. In addition, it was reported that rats with TNBS-induced colitis exhibited a reduction of macroscopic damage score and MPO activity after CPZ enema (115). Recent data pointed that TLR4-knockout mice showed a less inflammatory infiltration and a decreased expression of TRPV1 in TNBS-induced colitis, indicating one possible function of TLR4 for mediating TRPV1 signaling under inflammatory conditions (84). As for other animal models, McVey et al. (116) suggested that an intraluminal administration of capsaicin in isolated ileal segments of rats led to an intestinal

inflammation which could be reduced by CPZ. In T-cell-transfer colitis model, the activation of TRPV1 tended to exacerbate the intestinal inflammation, while the colitis was less severe when the TRPV1 in T cell was genetically or pharmacologically inhibited. The pro-inflammatory property of TRPV1 in T cells may be associated with the release of TNF- α , IFN- γ , IL-2, IL-4, IL-10, and IL-17 (117). In another study, capsaicin-induced TRPV1⁺ fibers-denervation ameliorated the intestinal inflammation in the T-cell-transfer colitis model (118), but the suppressive effect of noxious-dose capsaicin pretreatment only existed in 7–8 weeks old mice for several weeks after T-cell transfer, and these mice eventually developed colitis (128). It was further revealed that the severity of TNBS-induced colitis in the TRPV1⁺ fibers-denervated rats was drastically increased within 3–7 days after TNBS administration. Nevertheless, no significant difference of the colitis was found between denervated rats and normal rats in 14–21 days (129), reinforcing the concept that TRPV1⁺ fibers are involved in the early steps of colitis. Taken

together, these reports indicate that the activation of TRPV1 in colon is essential for the propagation of intestinal inflammation, and it might be a proximal event in the inflammatory process.

Noteworthy, several publications have reported the protective effects of TRPV1 in experimental colitis. It was exhibited that a local application of capsaicin and exogenous administration of CGRP ameliorated the colonic lesions in TNBS-induced rat colitis (122, 123). Massa et al. (124) stated that *Trpv1*^{-/-} mice exhibited a worse outcome of colitis and lower expression of anti-inflammatory neuropeptides, such as vasoactive intestinal peptide (VIP) and pituitary adenylate cyclase-activating peptide (PACAP), while the NF- κ B and STAT3 signaling pathways were demonstrated to be enhanced (130). Moreover, TRPV1 was reported to restrain the initiation and progression of colon cancer (130). In DSS-treated rats, a daily administration of capsaicin was able to reduce the severity of colitis, while a desensitization of TRPV1⁺-fibers dramatically worsened the inflammation (121). A protective role of TRPV1 was also identified in oxazolone-induced mice colitis, iodoacetamide-induced rat acute colitis, and formalin-induced rabbit acute colitis (125–127). The rather ambiguous findings concerning the roles of TRPV1 in colitis required further scrutiny.

Intriguingly, the expression of the TRPV1 was increased in the distal colon and rectum compared to the proximal colon in mice, and similar proximodistal gradient of CGRP/SP was detected (79, 131). Considering the anatomical distribution pattern of UC that often exhibits an ascending inflammation from rectal to proximal colon (4), the increased activity of TRPV1 and neuropeptides in distal colon might give rise to the increased susceptibility of distal colon to colitis and promote the spread of ascending inflammation. Such observations hint at the correlation between the diverse expression of TRP channels in the GI tract and the anatomical distribution pattern of IBD. Differences of microbial composition in certain gut regions and the crosstalk between microbiota and TRP channels are also likely to underlie the IBD anatomical distribution (6, 132). Likewise, it is reasonable to hypothesize that the diverse function of TRP channels in immune cells may be responsible for the distinct pathological pattern of UC and CD. A clear elucidation of this issue can facilitate a better understanding of the TRP channels and pathogenesis involved in IBD.

TRPA1

The studies regarding the role of TRPA1 in IBD all showed a upregulated TRPA1 expression in the colonic tissue of IBD patients (Table 1). In animal studies, mice with experimental colitis exhibited an increased TRPA1-mediated colonic neuropeptide release, while the experimental colitis appeared to be less severe after the inhibition of TRPA1 by the antagonist or genetic depletion (119). Additional studies suggested a protective role of TRPA1 in the GI tract. Pagano et al. (85) demonstrated that Cannabidiol, a potent agonist of TRPA1, was able to attenuate the intestinal inflammation in biopsies from pediatric patients with active UC. In dinitrobenzene sulfonic acid (DNBS)- and DSS-treated mice, Cannabidiol could also ameliorate neutrophil infiltration, intestinal permeability, cytokine production, and alter the dysregulation of gut

microbiota (85). Kun et al. (71) reported that the ablation of TRPA1 aggravated DSS-induced colitis and the activation of TRPA1 reduced the release of neuropeptides, cytokines, and chemokines, such as IL-1 β and macrophage chemoattractant protein-1 (MCP-1). Further support showed that TRPA1 activation reduced the level of TNF- α in colitis (2). Given that macrophage is the major producer for TNF- α , it may be that the TRPA1 in macrophages can suppress the release of TNF- α and modulate the anti-colitogenic effect, albeit the definite mechanism remains unclear. In addition, some evidence indicated that the expression of TRPA1 was increased in colonic stenotic regions of CD patients. The extent of intestinal inflammation and fibrotic changes in TNBS-treated TRPA1^{-/-} mice were more prominent compare to WT mice and the fibrosis could not be suppressed by inhibitors. The underlying mechanism was considered to be based on the anti-fibrotic role of TRPA1 in intestinal myofibroblasts (74, 75). These observations hint a novel therapeutic target to relieve the fibrosis in IBD.

Whilst considering the highly co-expressive nature of TRPV1 and TRPA1 in colonic afferents, it is interesting to shed light on the interaction between TRPV1 and TRPA1 in colitis. A stimulation of TRPA1 in dorsal root ganglia could result in the activation of PKA and subsequent phosphorylation of TRPV1 (133), while the activation of TRPV1 in afferents could desensitize TRPA1 through phosphatidylinositol biphosphate (PIP₂) depletion (134). In IBD patients, a vast infiltration of TRPV1⁺TRPA1⁺ T cells had been identified in inflamed colonic tissue (68). Bertin et al. (68) found that TRPV1⁺TRPA1⁻ T cells were able to enhance T-cell receptor-induced Ca²⁺ influx and aggravated intestinal inflammation in IL-10 knockout mice and T-cell-transfer colitis models compared to TRPV1⁺TRPA1⁺ T cells. However, the colitogenic properties of TRPV1⁺TRPA1⁻ T cells were abrogated with pharmacological inhibition or genetic deletion of TRPV1 (68, 117), suggesting that TRPA1 inhibited TRPV1 activity in CD4⁺ T cells and consequently restrained the activity of CD4⁺ T cells. Thus, the role of TRPA1 in colitis could be either protective or damaging.

TRPV4

The TRPV4 mRNA expression and TRPV4 immunoreactivity in colon were remarkably upregulated in IBD patients (70, 72, 76), in particular, serosal blood vessels with active inflammation were more densely innervated by TRPV4-positive fibers, which often co-localized with the infiltrating CD45⁺ cells (73, 76). Meanwhile, TRPV4 activation could recruit macrophages and other immune cells through the induction of chemokines, such as IL-8 and MCP-1 (73). D'Aldebert et al. (73) indicated the upregulated colonic TRPV4 expression in DSS-treated mice. Intracolonic administration of the TRPV4 agonists (4 α -phorbol-12,13-didecanoate or GSK1016790A) in mice activated NF- κ B and activator protein 1 (AP-1) signaling pathway, resulting in exacerbated DSS-induced colitis and even transiently increased the paracellular permeability of epithelium and blood vessel, while TRPV4-knockout mice conferred a strong resistance to the colitis (73, 81). These results prove the deleterious effects of TRPV4 on mucosal inflammation. Conversely, a systemic or local administration of RN1734, a selective TRPV4

antagonist, remarkably relieved the TNBS-induced colitis (72), suggesting the benefit of attenuating inflammation through blocking TRPV4. The medications aiming at TRPV4 might be capable of alleviating intestinal inflammation in IBD.

TRPM2

The expression of TRPM2 in distal colon was increased in TNBS-treated rats (90). In *Trpm2*^{-/-} mice, Yamamoto et al. (120) exhibited that the infiltration of immune cells and the severity of intestinal inflammation were ameliorated in DSS-induced colitis. The underlying mechanism might be that the Ca²⁺ influx was impaired in *Trpm2*^{-/-} macrophages, thus affecting the activation of NF-κB pathway (120). This evidence reminds us that TRPM2 can exert pro-inflammatory effects in the colitis via its essential role in macrophages and NF-κB signaling pathway.

TRPM8

TRPM8 expression was demonstrated to be upregulated in IBD patients and in DSS- or TNBS-treated mice (77). The activation of TRPM8 with icilin significantly attenuated the experimental colitis, but *Trpm8*^{-/-} mice were quite susceptible to colitis (62, 77). It was considered that TRPM8 performed its protective role in the intestine via restraining the release of TNF-α, IL-1, IL-6, and MCP-1, and inducing the release of CGRP (62, 77). TRPM8 activation could also reduce the TRPV1-dependent CGRP release in the gut (77), showing the ability of TRPM8 to suppress the TRPV1-associated inflammatory cascade. The reconstitution of *Trpm8*^{-/-} macrophages in mice exerted a deleterious effect on DSS-induced colitis (60), exhibiting a protective property of TRPM8 in macrophages. These findings reinforce the anti-colitogenic function of TRPM8. Agonists for TRPM8 possibly serve as therapeutic strategies for alleviating intestinal inflammation.

POSSIBLE FACTORS BEHIND PRIOR CONTRADICTORY RESULTS

According to aforementioned researches, the roles of a certain type of TRP channels in IBD and experimental colitis tended to be bidirectional or even conflicting.

The human studies mainly concentrated on the expression of TRP channels in the colonic tissue of IBD patients, however, the results of these studies appeared to be contradictory, especially concerning the expression of TRPV1. Actually, TRP channels are widely but anatomically distinctly distributed in various tissues and cell types in the GI tract (6). The expression and function of TRP channels may also be diverse in different subtypes and phases of IBD, and vary among individuals (79). However, the tissue samples of previous studies were acquired at multiple sites of the GI tract and the sample sizes were relatively small. Therefore, further researches that collect sufficient samples from a certain GI region and separately analyze the expression of TRP channels in UC and CD are warranted. Noteworthy, in IBD genome-wide association studies (GWAS), no single nucleotide polymorphism of TRP channel-related genes has been identified in correlation with IBD (135). However, the functions of TRP channels in the GI tract are tightly associated with the content in

GI lumen and molecules that possess significant polymorphisms in IBD GWAS, such as TLR4 (136). Additional IBD GWAS studies are needed to uncover specific factors including dietary intake or microbiota in IBD patients in order to explore the definite role of TRP channels polymorphisms in IBD.

Amongst the animal studies regarding TRP channels, researches on TRPV1 and TRPA1 were dominated, thus the majority of conflicting data was related to the functions of these two channels in experimental colitis. Many elements were probably responsible for the paradox.

First, the limitations of the animal models and experimental methods applied in the studies should be considered. Unfortunately, the ideal IBD models that completely mimic the multifactorial chronic disease do not exist and the pathophysiological mechanisms underlying different models are diverse. Also, animals of different strains, species, or ages have their distinct susceptibility to the stimulus, resulting in various demonstrations in experimental colitis (137). Due to the variety of animals and colitis models being used in prior studies, the animal models with distinct characteristics *per se* might accidentally account for the discrepant actions of TRP channels. Meanwhile, the different experimental methodology and drug administration could lead to opposing results. For example, capsaicin, the agonist for TRPV1, has dual effects that the low-dose capsaicin only affects a variable number of TRPV1-expressing nerves, while the high-dose capsaicin results in nerve desensitization (6), indicating the influence of the dose of stimuli on TRP channels. Moreover, the function of TRP channels might be affected by the changes in the microenvironment of the gut induced by agonists or antagonists (138), thus masking the true effects of TRP channels. It was revealed that TRP channels activation could be achieved via overexpression, phosphorylation, or recruitment to the plasma membrane (68). Additional experiments regarding the mode of TRP channels activation induced by specific stimulus may provide a rational view on the interaction between the stimulus and TRP channels.

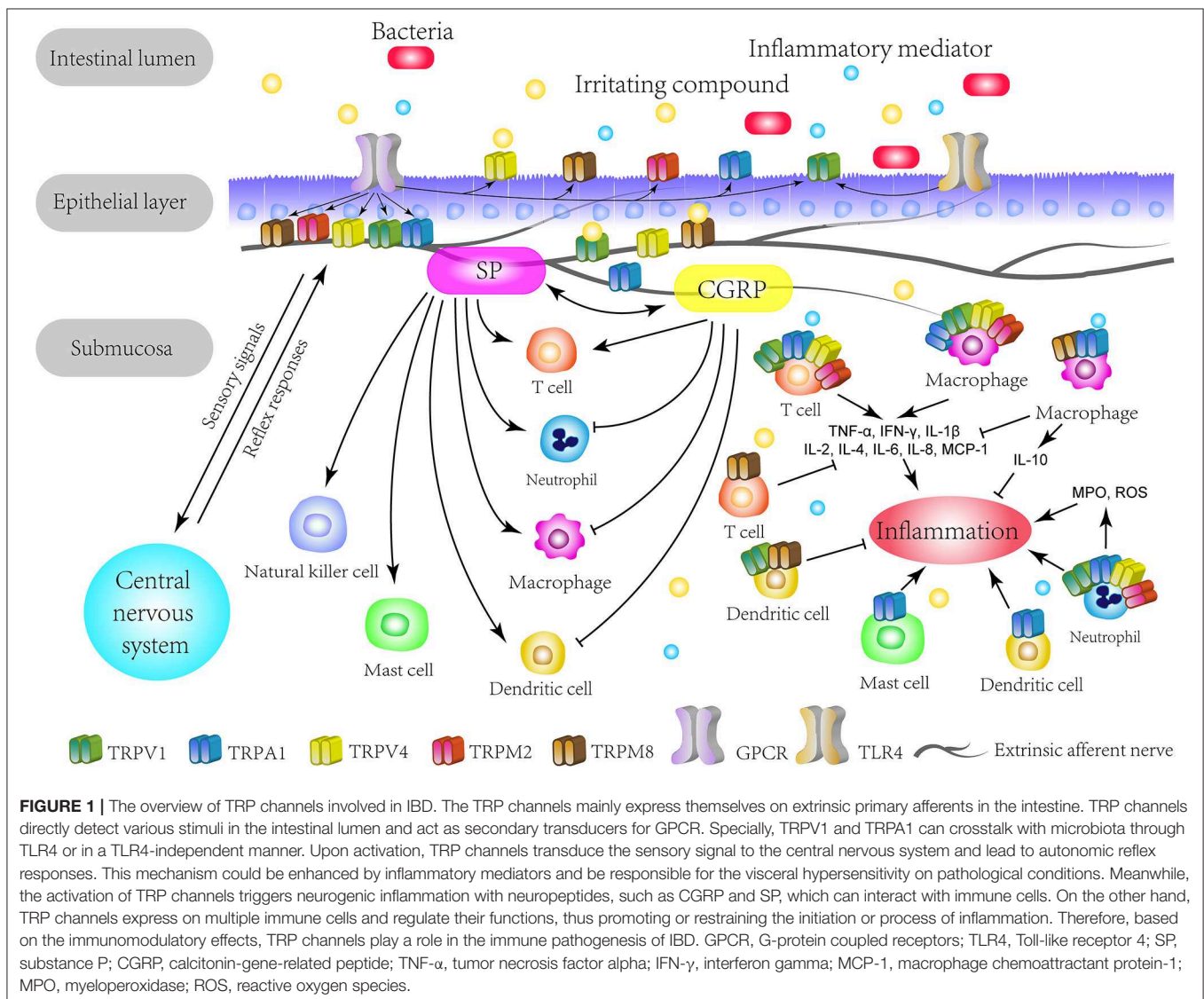
Second, in addition to the exogenous stimuli applied in studies, there appears to be various endogenous ligands acting on TRP channels, thus influencing the results of experiments. Compounds, such as prostaglandin metabolites, nerve growth factor, and products of oxidative stress can mediate TRPV1 and TRPA1 (5, 95, 139), making it difficult to attribute the results observed in studies to the stimulation of exogenous chemicals or to the stimulation of endogenous mediators. Actually, besides the administration of exogenous stimuli for TRP channels, the activation of TRP channels in IBD is also based on the stimulating effects of multiple endogenous mediators which are synthesized and released within the progress of colitis. Some of these compounds may potentialize TRP channels via the GPCR pathway (5). Hence, it is likely that TRP channels play a role not only in the initiation but also in the regulation of the intestinal inflammation, while the exact mechanism is unclear and needs further explorations.

Third, the functions of neurogenic inflammation and immune responses triggered by TRP channels activation are complicated. The neurogenic inflammation is featured of the release of

CGRP and SP, but the effects of these two neuropeptides on intestinal inflammation were not clearly elucidated and tended to be contradictory. The differences in the concentration of neuropeptides and the expression of receptors might contribute to the discrepancy (4). SP was reported to sensitize TRPV1 during colitis and affect the functions of TRPV1 (78), suggesting a possible feedback sensitization loop between neuropeptides and TRP channels. In addition, a range of evidence showed that neuropeptides, such as somatostatin, galanin, opioid peptides, VIP, and PACAP could participate in the inflammation and regulate the inflammatory responses (130, 140). It is warranted to explore whether there is an association between these neuropeptides and TRP channels in colitis. As for immunity, besides the TRP channels-expressing immune cells, some non-immune cells may have TRP channels in the colitis. For example, the expression and function of TRPA1 were identified in fibroblasts which could transform into myofibroblasts and contribute to the regulation of intestinal inflammation (74,

75, 105). However, the definite involvement of myofibroblasts in colitis was poorly understood. Additional explorations are necessary to reveal other TRP channels-expressing cells that play a role in colitis.

Fourth, the TRP channels may interact with various cellular pathways. For instance, the inhibition of TRPV1 could lead to an increased availability of anandamide, and then induced downstream effects on NF- κ B and TNF- α and affected bowel motility via the receptor for anandamide (78, 124). Meanwhile, anandamide could also act on TRPV1 and regulate a protection against intestinal inflammation (78, 124), suggesting a potential synergy between TRP channels and other cellular pathways in some settings. Similar to the interaction between TRPV1 and TRPA1, the possible crosstalk between other subtypes of TRP channels is also worth noting. Further studies are warranted to elucidate the comprehensive regulatory network induced by the stimulation of TRP channels in colitis.



In general, the stimulation of TRP channels and a serial cascade of events are like double-edged swords in the intestinal inflammation that exert colitogenic or anti-colitogenic effects in different situations, influenced by a myriad of interactions amongst stimuli, neuropeptides, and the immunity. It is a challenge to figure out the accurate action of a certain type of TRP channels in a specific immune or cellular pathway. Intriguingly, Cohen et al. (22) applied *in vivo* optogenetic strategy to selectively stimulate cutaneous TRPV1⁺-neuron and showed that the afferent activation was based on a nerve reflex. In this study, the activation of TRPV1 was in the absence of other inflammatory stimuli, thus specifically demonstrating the precise role of TRPV1 in the afferents and its immunity-triggering effects. Considering the intense interactions amongst various factors in previous researches, utilizing novel technologies that can efficiently eliminate interferences is a promising strategy for further studies on TRP channels.

THERAPEUTIC VALUES OF TRP CHANNELS FOR IBD

As discussed, the effects of TRP channels in IBD have been increasingly appreciated, it is intriguing for researchers to explore their therapeutic values for relieving inflammatory VHS

and intestinal inflammation. Pharmacologically, the modulating agents for TRP channels include antagonists and stimulant agonists (141). In particular, antagonists for TRP channels exert a specific effect on modification of ion channel, and stimulant agonists facilitate the desensitization of sensitive afferents (142). However, owing to the wide distribution and various physiological roles of TRP channels within and outside the GI tract (143), the modulation of TRP channels may result in pronounced side effects, such as hyper-thermic effect and impaired injurious-heat perception generated by TRPV1 antagonists (144, 145). Therefore, it is vital to develop the stimulus-specific blockers for TRP channels that specifically act on the aberrant function while sparing the physiological function.

Besides targeting TRP channels directly, it is worth noting that aiming at the stimulus and downstream pathways for GPCRs tends to be another valuable method of restraining the action of TRP channels, especially in the inflammatory process. A novel class of endogenous lipid mediators named resolvin, which are generated from immune cells, such as eosinophils and neutrophils, are of particular interest and have the ability to suppress the function of TRP channels including TRPV1, TRPA1, and TRPV4 (146, 147). The anti-inflammatory effects of resolvin are likely based on the activation of inhibitory GPCR that subsequently suppresses the GPCR-associated sensitization or activation of TRP channels (148), showing the feasibility for

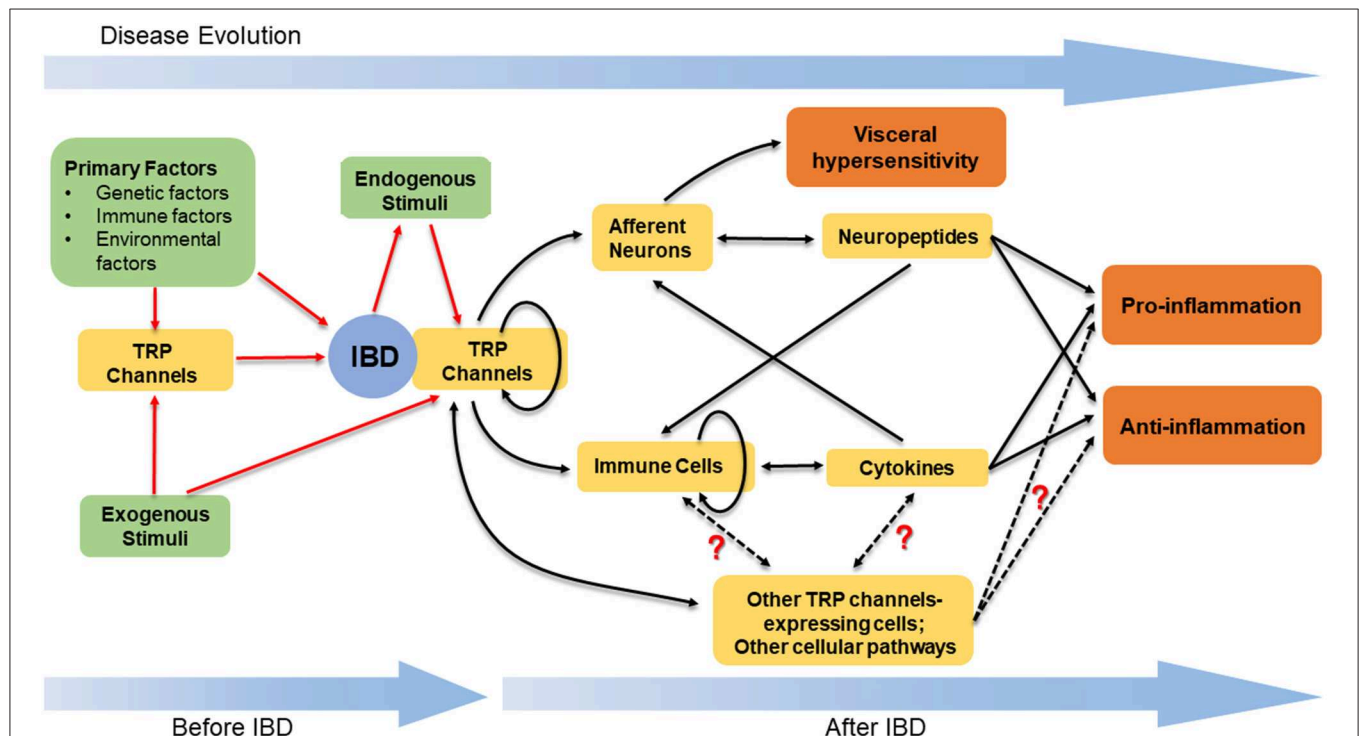


FIGURE 2 | The stimulation of TRP channels and the downstream regulatory network in IBD. TRP channels, together with other primary factors, play a role in the pathogenesis of IBD. The stimulation of TRP channels is based on the exogenous stimuli and the endogenous stimuli. The latter mainly refer to the mediators synthesized and released within the progress of colitis. The activated TRP channels can induce the release of neuropeptides and cytokines, thus leading to the pro- or anti-inflammatory effects. In addition to neurons and immune cells, there are other TRP channels-expressing cells and cellular pathways contributing to regulate the intestinal inflammation, while the definite functions of these cells and pathways are unclear. The complicated crosstalk amongst the neuropeptides, cytokines, TRP channels-expressing cells, and diverse cellular pathways results in the various but elusive effects induced by the stimulation of TRP channels in IBD.

inhibiting TRP channels through regulating GPCRs. Overall, treatment strategies targeting TRP channels and their signaling pathways predict a promising future for alleviating the symptoms and improving the prognosis of IBD. More studies are warranted to identify the efficacy and safety of these therapeutic approaches.

To conclude, TRP channels are not only widely distributed on neurons in the GI tract, functioning as detectors for stimuli and triggers for neurogenic inflammation, but also expressed in multiple immune cells and modulate immune responses (Figure 1). Accumulated evidence has supported an important association between TRP channels and IBD. Although different types of TRP channels exert distinct effects, it is evident that TRP channels are involved in the VHS and the pathogenesis of IBD through a complicated and elusive regulatory network (Figure 2). The inhibition or activation of selected TRP channels can restrain the development of VHS and inflammation in the context of colitis. Therefore, TRP antagonists and agonists tend

to constitute an attractive target in IBD treatment and need further attention.

AUTHOR CONTRIBUTIONS

YC and HZ reviewed the literature and outlined the overall manuscript. YC wrote the manuscript. JM and MZ helped to write the manuscript. AM and HZ revised the manuscript. HZ supervised the preparation of the draft and edited it and worked as a corresponding author. All authors approved the final version.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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