



The Selective Autophagy Receptor Optineurin in Crohn's Disease

Markus Tschurtschenthaler^{1,2*} and Timon Erik Adolph^{3*}

¹ Center for Translational Cancer Research (TranslaTUM), Technical University of Munich, Munich, Germany, ² Department of Internal Medicine II, Klinikum Rechts der Isar, Technical University of Munich, Munich, Germany, ³ Department of Medicine I (Gastroenterology, Endocrinology and Metabolism), Medical University Innsbruck, Innsbruck, Austria

Autophagy is a pathway that allows cells to target organelles, protein complexes, or invading microorganisms for lysosomal degradation. The specificity of autophagic processes is becoming increasingly recognized and is conferred by selective autophagy receptors such as Optineurin (OPTN). As an autophagy receptor, OPTN controls the clearance of *Salmonella* infection and mediates mitochondrial turnover. Recent studies demonstrated that OPTN is critically required for pathogen clearance and an appropriate cytokine response in macrophages. Moreover, OPTN emerges as a critical regulator of inflammation emanating from epithelial cells in the intestine. OPTN directly interacts with and promotes the removal of inositol-requiring enzyme 1 α , a central inflammatory signaling hub of the stressed endoplasmic reticulum (ER). Perturbations of ER and autophagy functions have been linked to inflammatory bowel disease (IBD) and specifically Crohn's disease. Collectively, these studies may explain how perturbations at the ER can be resolved by selective autophagy to restrain inflammatory processes in the intestine and turn the spotlight on OPTN as a key autophagy receptor. This review covers a timely perspective on the regulation and function of OPTN in health and IBD.

Keywords: optineurin, inflammatory bowel disease, Crohn's disease, endoplasmic reticulum stress, selective autophagy

INTRODUCTION

Autophagy is an evolutionary conserved self-cannibalistic pathway that leads to the degradation of bulk cytoplasm (macroautophagy) in order to generate energy and to maintain cell homeostasis (1). However, researchers are increasingly appreciating that receptors specifically guide autophagic degradation as exemplified by the removal of damaged organelles [e.g., mitophagy of mitochondria, ER-phagy of endoplasmic reticulum (ER), pexophagy of peroxisomes], bacteria (xenophagy), lipid droplets (lipophagy), protein aggregates (aggrephagy), and other cytosolic constituents (2, 3). This rather selective autophagic process may be controlled by Optineurin (OPTN) besides other autophagy receptors including p62/SQSTM1, NBR1, CALCOCO2/NDP52, and TAX1BP1 (3, 4). These receptors recognize ubiquitinated cargo via their ubiquitin-binding domains (UBA, UBZ or UBAN) and tether it to the autophagosomal membranes by their LC3-interacting regions (LIRs) (5). However, OPTN does not only guide selective autophagy but also controls tumor necrosis factor (TNF), nuclear factor κ B (NF- κ B), and type I interferon (IFN) signaling (6–8). OPTN has been implicated in a variety of human diseases including glaucoma (9), amyotrophic lateral sclerosis (10, 11), Paget's disease (12, 13), and recently, inflammatory bowel disease (IBD) (14, 15). IBD comprise a spectrum of complex diseases that affect the gastrointestinal tract and organs beyond the intestine (e.g., eye, skin, joints). IBD is clinically distinguished into two major

OPEN ACCESS

Edited by:

Folma Buss,
University of Cambridge,
United Kingdom

Reviewed by:

Krzysztof Guzik,
Jagiellonian University, Poland
Giuseppe Sciumè,
Sapienza Università di
Roma, Italy

*Correspondence:

Markus Tschurtschenthaler
markus.tschurtschenthaler@tum.de;
Timon Erik Adolph
timon-erik.adolph@i-med.ac.at

Specialty section:

This article was submitted
to Inflammation,
a section of the journal
Frontiers in Immunology

Received: 29 January 2018

Accepted: 27 March 2018

Published: 10 April 2018

Citation:

Tschurtschenthaler M and Adolph TE
(2018) The Selective Autophagy
Receptor Optineurin in Crohn's
Disease.
Front. Immunol. 9:766.
doi: 10.3389/fimmu.2018.00766

phenotypes: ulcerative colitis (UC) and Crohn's disease (CD). Although these two diseases share some genetic risk they are considered separate disease entities due to their localization, clinical presentation and response to therapy (16). The pathophysiology of these diseases involves environmental factors and their impact on the intestinal microbiota which may orchestrate a chronic remittent form of inflammation in genetically susceptible hosts (17–19). Genetic variation in the autophagy gene *ATG16L1* has been linked to CD (20, 21) which leads to an impaired autophagic response due to caspase 3-mediated cleavage of the mutant *ATG16L1* variant (22). Impaired autophagy function especially in intestinal epithelial cells results in the susceptibility to small and large intestinal inflammation (23–27). As such, it is conceivable that the selective autophagy receptor OPTN may regulate inflammatory processes in the intestine. Evidence for the regulation and function of OPTN in intestinal inflammation and specifically IBD is covered in this review.

THE SELECTIVE AUTOPHAGY RECEPTOR OPTN IN HEALTH

The gene encoding OPTN is evolutionary conserved and expressed in most tissues of the human body (28). OPTN was initially discovered in 1998 in a yeast two-hybrid screen as a binding partner of the adenovirus protein E3-14.7K (early region 3 of group C human adenoviruses 14.7 kDa), and was thereafter named as FIP-2 (for 14.7 kDa interacting protein) (28). Later, this gene was identified to have a strong homology with NEMO (NF- κ B essential modulator) and was subsequently denoted as NEMO-related protein (29). But it also became known as transcription factor IIIA-interacting protein, Huntingtin-interacting protein 7, and Huntingtin yeast partner L (30). Eventually, the multifunctional protein was renamed to OPTN (“optic neuropathy inducing”), as it was found to play a major neuroprotective role and mutations in this gene were shown to be causative for the development of primary open-angle glaucoma, a leading cause of blindness (9).

The human *OPTN* gene is located at chromosome 10 and consists of three non-coding exons in the 5'UTR and 13 exons that encode a 577 amino acid protein with a size of 66 kDa. The mouse *Optn* gene is located at chromosome 2 and also contains 13 exons, which encodes a full-length protein of similar size that shows 78% sequence similarity to the human protein (6, 30). The OPTN protein consists of several functional domains including a basic leucine zipper motif (bZIP), a microtubule-associated protein 1 light-chain LIR, a ubiquitin-binding domain (UBAN), multiple coiled-coil motifs as well as a ubiquitin-binding zinc-finger domain at the C-terminus (6). Notably, NEMO, a central regulator of NF- κ B activation shares 53% similarity with OPTN and only lacks a fragment of 166 amino acids at the N-terminal region containing a putative leucine zipper domain (30). However, despite this similarity, OPTN is (unlike NEMO) not a regulatory subunit of the I κ B kinase (IKK) complex that is essential for NF- κ B activation (29). OPTN was shown to block the ability of NEMO to bind ubiquitinated receptor-interacting protein kinase 1 (RIPK1), which resulted in a suppression of

TNF-induced NF- κ B signaling (8). Similarly, OPTN also inhibits NF- κ B signaling through interaction with CYLD that leads to deubiquitination of NEMO and RIPK1 (31). Notably, OPTN is induced by TNF receptor signaling and can thus function as a negative-feedback regulator for NF- κ B (32). As such, OPTN negatively regulates TNF α -mediated NF- κ B signaling which is critically involved in the regulation of immune responses and cell death signaling. In contrast, *in vivo* experiments using *Optn* knock-out and *Optn*^{470T} knock-in mice suggest that OPTN plays no role in the regulation of NF- κ B signaling (33, 34).

Furthermore, OPTN was shown to be regulated by and control type I IFN responses (7, 29, 35). Production of type I IFNs is the primary response to bacterial and viral infections (36). Specifically, upon recognition of pathogen-associated molecular patterns by toll-like receptors or RIG-I-like receptors, IFN regulatory factor 3 (IRF3) becomes phosphorylated by TANK-binding kinase 1 (TBK1) and translocates to the nucleus, which leads to the transcription of type I IFN response genes (7). OPTN binds to TBK1 to support IRF3 activation and production of type I IFNs (34). In contrast to this notion, however, OPTN was shown to suppress virus-induced IRF3 signaling (37).

More recently, OPTN was identified as a selective autophagy receptor required for autophagic clearance of *Salmonella enterica* (38), removal of damaged mitochondria (39) and degradation of protein clusters at the ER (24). Selective autophagy receptors, i.e., OPTN, NDP52, p62, and TAX1BP1 recognize ubiquitinated cargo and link it to the autophagosomal membrane (3). Before autophagic clearance of the ubiquitinated cargo, TBK1 activates OPTN by phosphorylation in order to enhance its binding capacity to LC3, a conjugate of the autophagosomal membrane (38, 40). A similar mechanism has for example been demonstrated for mitophagy (41). Only recently, a mechanistic link between OPTN and autophagy has been provided. Bansal and colleagues demonstrated an interaction of OPTN with the core autophagy machinery forming around ATG16L1. More specifically, the authors demonstrated that OPTN was required for the recruitment of the ATG12/ATG5/ATG16L1 complex to phagophores for autophagosomal elongation in starvation-induced autophagy (42).

THE SELECTIVE AUTOPHAGY RECEPTOR OPTN IN DISEASE

OPTN Is Required for Pathogen Clearance and an Inflammatory Response in Macrophages

The Segal group and colleagues contributed to our understanding of OPTN in intestinal disease processes (15, 43). Bone marrow-derived macrophages from OPTN-deficient mice exhibited a decreased capacity to respond with TNF- α and IL-6 secretion upon stimulation with heat-inactivated *Escherichia coli*. Defective bacterial handling in OPTN-deficient macrophages was paralleled by a more severe *Citrobacter rodentium*-induced colitis and *E. coli* peritonitis. The more severe phenotype may be explained by an inappropriate immune response at the site of infection which increased mortality of OPTN-deficient animals in both

models (43). In line with this, OPTN-deficient HeLa cells exhibited a bacterial handling defect after *Salmonella* infection (38) similar to a more severe phenotype after *Salmonella* infection in mice that was independent from NF- κ B or type I IFN responses in macrophages (33). Of note, activation of inflammatory signaling cascades may be determined by distinct ubiquitin chains on bacteria as it was recently shown for the ubiquitin coat on *S. typhimurium* that provides a platform for NF- κ B (44). Activation of NF- κ B resulted in secretion of pro-inflammatory cytokines and reduced bacterial proliferation (44).

Collectively, these data demonstrate that OPTN limits bacterial infection in the intestine likely by mediating selective autophagy and pathogen clearance.

Macrophage OPTN Expression Is Reduced in a Proportion of CD Patients

The Segal group also analyzed monocyte-derived macrophages from ~40 patients with CD and UC. They noted that—similar to their findings in OPTN-deficient mice—CD macrophages exhibited impaired immune responses (i.e., TNF- α and IFN- γ secretion) upon stimulation with inactivated *E. coli* when compared to healthy controls which could not be explained by the transcriptional profile (15). The authors identified a CD subgroup which composed 10% of their cohort that expressed reduced OPTN in macrophages. Reduced expression may be explained by genetic variation as the authors observed an association with a single nucleotide polymorphism rs12415716 that exhibited a minor allele frequency of ~18%. Indeed, siRNA silencing reduced the inflammatory response of OPTN-deficient macrophages by 25% (43). These two studies highlight that OPTN is required for an appropriate immune response of macrophages upon exposure to bacterial antigens. Whether reduced OPTN expression and an impaired cytokine response is a cause of or consequence from IBDs deserves further attention. Furthermore, it will be interesting to decipher how OPTN modulates immune responses and how this may be related to autophagic processes (43).

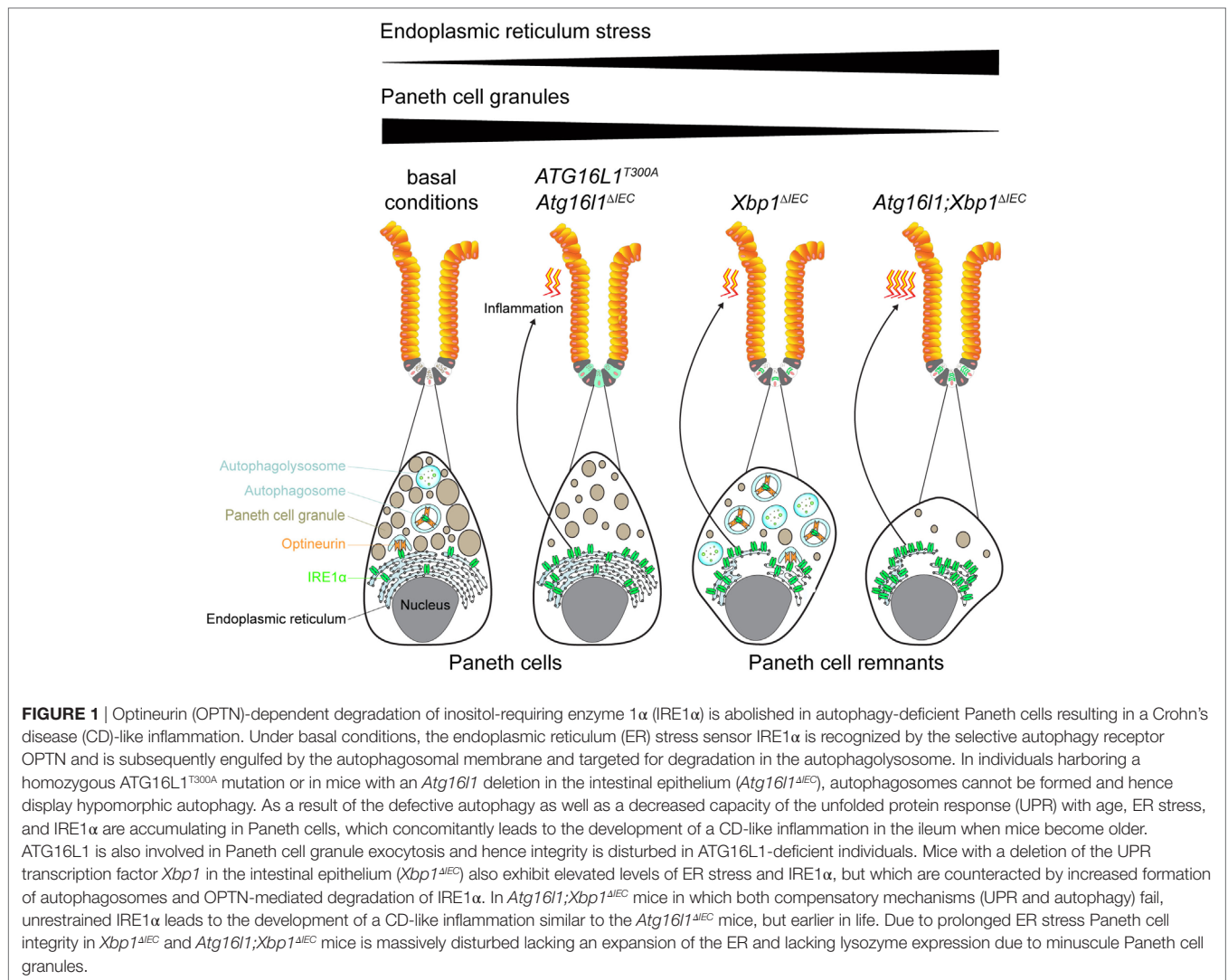
OPTN Limits the Accumulation of an ER-Based Inflammatory Signaling Hub in Intestinal Epithelial Cells

The ER is a cellular organelle which hosts protein synthesis and folding and which instigates trafficking for secretory purposes (45). These processes are fundamentally important for cellular homeostasis which is why they are tightly controlled by redundant mechanisms. One of these mechanisms is the unfolded protein response (UPR) that is equipped with three major sensors of stress at the ER: inositol-requiring enzyme 1, protein kinase RNA-like endoplasmic reticulum kinase, and activating transcription factor 6. These sensors are engaged upon accumulation of unfolded, misfolded, and aggregating proteins in the ER, a condition termed endoplasmic reticulum stress (ER stress) (46). The UPR generally aims at the resolution of ER stress; however, unabated stress at the ER may instigate inflammatory (danger) signaling (47–49). This may be executed for example by the formation of inositol-requiring enzyme 1 α (IRE1 α) oligomers which cluster and may not be suitable for

proteasomal degradation (23, 24). IRE1 α is expressed in intestinal epithelial cells and particularly Paneth cells that heavily rely on the UPR due to a high secretory burden (17, 24). IRE1 α is a transmembrane receptor that harbors a kinase and endoribonuclease domain, which allows splicing and activation of the transcription factor X-box-binding protein 1 (*Xbp1*) to instigate the UPR and maintain ER homeostasis (50). In turn, unabated ER stress induced by genetic deletion of *Xbp1* in *Xbp1 Δ IEC* mice hyperactivates IRE1 α (23, 51, 52). Importantly, ER stress-induced IRE1 α hyperactivation is restrained by autophagy as co-deletion of autophagy-related 16-like 1 (*Atg16l1*) increased the level of IRE1 α activation (Figure 1) (23). Defective removal and hyperactivation of IRE1 α in *Atg16l1*;*Xbp1 Δ IEC* mice was paralleled by a spontaneous CD-like inflammatory phenotype restricted to the small intestine with a fissuring transmural character (23). Indeed, genetic co-deletion of IRE1 α ameliorated CD-like inflammation in *Atg16l1*;*Xbp1 Δ IEC* mice (24) demonstrating that autophagy-restricted IRE1 α activity critically controlled inflammation that emanated from intestinal epithelial cells (23, 24). In these studies, the selective autophagy receptor OPTN emerged as critical regulator of IRE1 α degradation in the setting of unabated ER stress (24). These observations suggest that OPTN targets IRE1 α , possibly by a ubiquitin signal (53, 54), for autophagosomal degradation to remove an inflammatory signaling hub from the stressed ER (Figure 1). Notably, CD patients harboring the ATG16L1 T300A autophagy-deficient variant (22) exhibited increased IRE1 α accumulation in Paneth cells (24), a site of epithelial ER stress (23, 55). Collectively, these data suggest that autophagy controls ER stress and inflammation specifically in Paneth cells of CD patients harboring the ATG16L1^{T300A} variant. This notion is supported by a model of Paneth cell-specific ER stress which led to the development of a spontaneous enteritis (23). In summary, these studies suggest that IRE1 α degradation is dependent on ATG16L1-mediated autophagy and possibly on OPTN as an autophagy receptor (24, 42). These data advocate a role for OPTN in inflammatory processes consequent to ER stress, but further studies are needed to corroborate a role for OPTN and selective autophagy during intestinal inflammatory processes.

DISCUSSION

Optineurin controls autophagic processes by selectively targeting ubiquitinated molecules for autophagic degradation (3, 4). A direct interaction of OPTN with IRE1 α in intestinal epithelial cells and the requirement of OPTN for the removal of this inflammatory signaling hub may set a basis for our understanding of how autophagy can resolve ER stress-induced inflammation. We suggest that IRE1 α is targeted by OPTN for autophagosomal degradation under conditions of ER stress to restrain IRE1 α -mediated danger signaling and inflammation (24). Similarly, ER-phagy of stressed ER membranes also leads to the resolution of ER stress (56) which suggests once more that a tight control of the ER is indispensable for cellular homeostasis (57–61). Furthermore, OPTN is required to target a critical autophagy hub containing ATG16L1 to the forming phagophore (42). However, we acknowledge that distinct mechanisms other than selective



autophagy may control IRE1α activity and an inflammatory threshold (62).

Understanding the selectivity of OPTN-mediated autophagy would be highly informative. For example, definition of an unbiased OPTN interactome could help to understand two major biological functions of OPTN (i.e., selective autophagy and regulation of inflammatory pathways) and to define their relationship in health and disease. More specifically, it may be critical to discriminate OPTN-mediated autophagy functions from those that are independent of autophagy as it currently unclear how they are interconnected. Some literature would support the notion that receptors of selective autophagy are critically involved in inflammatory processes (63) similar to a genetic variant in *NDP52* with CD (64). Furthermore, autophagy receptors may control microbial dissemination (65, 66), a concept that becomes increasingly relevant in dysbiotic situations as seen in IBD (67). Reduction of *OPTN* expression in macrophages of some CD patients may not just result in diminished cytokine secretion upon bacterial infections (15), but may also lead to a decreased autophagic containment of pathogens

(or commensals) and degradation of inflammatory molecules as exemplified for IRE1α in Paneth cells (24). However, it may well be that microbial control and ER stress are interrelated pathways (68). The impact of these observations in IBD deserve further attention and direct evidence for the regulation and function of OPTN in epithelial cells of CD patients is eagerly awaited.

In case of OPTN-mediated autophagy, we are only beginning to appreciate a role in intestinal inflammatory disease processes. However, we propose broad implications for OPTN in ER stress-related diseases within and beyond the intestine (24, 69–71). A driving force (besides genetic variation and environmental cues) may be cellular senescence with a declining capacity of the UPR during aging (72).

CONCLUDING REMARKS

Evidence accumulates for a role of OPTN in disease processes within and beyond the intestine. Direct evidence for a role of OPTN in CD is limited. However a concept arises, in which OPTN is required for the removal of inflammatory molecules

from the ER and invading bacteria (14, 15, 24), which may be governed by OPTN-mediated selective autophagy. As such, OPTN emerges as a critical link between ER disturbances and the resolution by autophagy (24). This observation is of note as ER stress is commonly observed in IBD patients and especially those harboring prominent genetic risk factors (e.g., ATG16L1 and NOD2) (22, 73, 74), which reflects one facet in this complex inflammatory condition (17, 19). Pharmacologic targeting of autophagy may indeed be beneficial in IBD which could depend on the ability of the host to launch an appropriate autophagic response (75). Moreover, clinically established drugs may exert their beneficial effects through modulation of autophagy (75). As such, understanding the biology of OPTN in CD may help to establish or guide future therapies.

REFERENCES

- Galluzzi L, Baehrecke EH, Ballabio A, Boya P, Bravo-San Pedro JM, Cecconi F, et al. Molecular definitions of autophagy and related processes. *EMBO J* (2017) 36:1811–36. doi:10.15252/embj.201796697
- Klionsky DJ, Abdelmohsen K, Abe A, Abedin MJ, Abeliovich H, Acevedo Arozena A, et al. Guidelines for the use and interpretation of assays for monitoring autophagy (3rd edition). *Autophagy* (2016) 12:1–222. doi:10.1080/15548627.2015.1100356
- Stolz A, Ernst A, Dikic I. Cargo recognition and trafficking in selective autophagy. *Nat Cell Biol* (2014) 16:495–501. doi:10.1038/ncb2979
- Rogov V, Dotsch V, Johansen T, Kirkin V. Interactions between autophagy receptors and ubiquitin-like proteins form the molecular basis for selective autophagy. *Mol Cell* (2014) 53:167–78. doi:10.1016/j.molcel.2013.12.014
- Kim BW, Kwon DH, Song HK. Structure biology of selective autophagy receptors. *BMB Rep* (2016) 49:73–80. doi:10.5483/BMBRep.2016.49.2.265
- Slowicka K, Vereecke L, van Loo G. Cellular functions of optineurin in health and disease. *Trends Immunol* (2016) 37:621–33. doi:10.1016/j.it.2016.07.002
- Meena NP, Zhu G, Mittelstadt PR, Giardino Torchia ML, Pourcelot M, Arnoult D, et al. The TBK1-binding domain of optineurin promotes type I interferon responses. *FEBS Lett* (2016) 590:1498–508. doi:10.1002/1873-3468.12176
- Zhu G, Wu CJ, Zhao Y, Ashwell JD. Optineurin negatively regulates TNF α -induced NF- κ B activation by competing with NEMO for ubiquitinated RIP. *Curr Biol* (2007) 17:1438–43. doi:10.1016/j.cub.2007.07.041
- Rezaie T, Child A, Hitchings R, Brice G, Miller L, Coca-Prados M, et al. Adult-onset primary open-angle glaucoma caused by mutations in optineurin. *Science* (2002) 295:1077–9. doi:10.1126/science.1066901
- Maruyama H, Morino H, Ito H, Izumi Y, Kato H, Watanabe Y, et al. Mutations of optineurin in amyotrophic lateral sclerosis. *Nature* (2010) 465:223–6. doi:10.1038/nature08971
- Bury JJ, Highley JR, Cooper-Knock J, Goodall EF, Higginbottom A, McDermott CJ, et al. Oligogenic inheritance of optineurin (OPTN) and C9ORF72 mutations in ALS highlights localisation of OPTN in the TDP-43-negative inclusions of C9ORF72-ALS. *Neuropathology* (2016) 36:125–34. doi:10.1111/neup.12240
- Albagha OM, Visconti MR, Alonso N, Langston AL, Cundy T, Dargie R, et al. Genome-wide association study identifies variants at CSF1, OPTN and TNFRSF11A as genetic risk factors for Paget's disease of bone. *Nat Genet* (2010) 42:520–4. doi:10.1038/ng.562
- Chung PY, Beyens G, Boonen S, Papapoulos S, Geusens P, Karperien M, et al. The majority of the genetic risk for Paget's disease of bone is explained by genetic variants close to the CSF1, OPTN, TM7SF4, and TNFRSF11A genes. *Hum Genet* (2010) 128:615–26. doi:10.1007/s00439-010-0888-2
- Smith AM, Rahman FZ, Hayee B, Graham SJ, Marks DJ, Sewell GW, et al. Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *J Exp Med* (2009) 206:1883–97. doi:10.1084/jem.20091233090209c
- Smith AM, Sewell GW, Levine AP, Chew TS, Dunne J, O'Shea NR, et al. Disruption of macrophage pro-inflammatory cytokine release in Crohn's

AUTHOR CONTRIBUTIONS

MT and TA contributed equally to this manuscript.

FUNDING

This work was supported by the European Union's Framework Programme for Research and Innovation Horizon 2020 (2014–2020) under the Marie Skłodowska-Curie Grant Agreement No. 753058 (to MT) and an EMBO Long-Term Fellowship (ALTF 1290-2016 to MT). This work was further supported by the Austrian Science Fund (FWF, P 29379-B28), the Austrian Society of Gastroenterology and Hepatology (ÖGGH) and the European Crohn's and Colitis Organisation (ECCO) (to TA).

- disease is associated with reduced optineurin expression in a subset of patients. *Immunology* (2015) 144:45–55. doi:10.1111/imm.12338
- Abraham C, Cho JH. Inflammatory bowel disease. *N Engl J Med* (2009) 361:2066–78. doi:10.1056/NEJMra0804647
 - Kaser A, Zeissig S, Blumberg RS. Inflammatory bowel disease. *Annu Rev Immunol* (2010) 28:573–621. doi:10.1146/annurev-immunol-030409-101225
 - de Souza HS, Fiocchi C. Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol* (2016) 13:13–27. doi:10.1038/nrgastro.2015.186
 - Neurath MF. Cytokines in inflammatory bowel disease. *Nat Rev Immunol* (2014) 14:329–42. doi:10.1038/nri3661
 - Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet* (2007) 39:207–11. doi:10.1038/ng1954
 - Jostins L, Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* (2012) 491:119–24. doi:10.1038/nature11582
 - Murthy A, Li Y, Peng I, Reichelt M, Katakam AK, Noubade R, et al. A Crohn's disease variant in Atg16l1 enhances its degradation by caspase 3. *Nature* (2014) 506:456–62. doi:10.1038/nature13044
 - Adolph TE, Tomczak MF, Niederreiter L, Ko HJ, Bock J, Martinez-Naves E, et al. Paneth cells as a site of origin for intestinal inflammation. *Nature* (2013) 503:272–6. doi:10.1038/nature12599
 - Tschurtschenthaler M, Adolph TE, Ashcroft JW, Niederreiter L, Bharti R, Saveljeva S, et al. Defective ATG16L1-mediated removal of IRE1 α drives Crohn's disease-like ileitis. *J Exp Med* (2017) 214:401–22. doi:10.1084/jem.20160791
 - Diamanti MA, Gupta J, Bennecke M, De Oliveira T, Ramakrishnan M, Braczynski AK, et al. IKK α controls ATG16L1 degradation to prevent ER stress during inflammation. *J Exp Med* (2017) 214:423–37. doi:10.1084/jem.20161867
 - Cadwell K, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, et al. A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal Paneth cells. *Nature* (2008) 456:259–63. doi:10.1038/nature07416
 - Cadwell K, Patel KK, Maloney NS, Liu TC, Ng AC, Storer CE, et al. Virus-plus-susceptibility gene interaction determines Crohn's disease gene ATG16L1 phenotypes in intestine. *Cell* (2010) 141:1135–45. doi:10.1016/j.cell.2010.05.009
 - Li Y, Kang J, Horwitz MS. Interaction of an adenovirus E3 14.7-kilodalton protein with a novel tumor necrosis factor α -inducible cellular protein containing leucine zipper domains. *Mol Cell Biol* (1998) 18:1601–10. doi:10.1128/MCB.18.3.1601
 - Schwamborn K, Weil R, Courtois G, Whiteside ST, Israel A. Phorbol esters and cytokines regulate the expression of the NEMO-related protein, a molecule involved in a NF- κ B-independent pathway. *J Biol Chem* (2000) 275:22780–9. doi:10.1074/jbc.M001500200
 - Kachaner D, Genin P, Laplantine E, Weil R. Toward an integrative view of optineurin functions. *Cell Cycle* (2012) 11:2808–18. doi:10.4161/cc.20946
 - Nagabhushana A, Bansal M, Swarup G. Optineurin is required for CYLD-dependent inhibition of TNF α -induced NF- κ B activation. *PLoS One* (2011) 6:e17477. doi:10.1371/journal.pone.0017477

32. Sudhakar C, Nagabhushana A, Jain N, Swarup G. NF-kappaB mediates tumor necrosis factor alpha-induced expression of optineurin, a negative regulator of NF-kappaB. *PLoS One* (2009) 4:e5114. doi:10.1371/journal.pone.0005114
33. Slowicka K, Vereecke L, Mc Guire C, Sze M, Maelfait J, Kolpe A, et al. Optineurin deficiency in mice is associated with increased sensitivity to *Salmonella* but does not affect proinflammatory NF-kappaB signaling. *Eur J Immunol* (2016) 46:971–80. doi:10.1002/eji.201545863
34. Munitic I, Giardino Torchia ML, Meena NP, Zhu G, Li CC, Ashwell JD. Optineurin insufficiency impairs IRF3 but not NF-kappaB activation in immune cells. *J Immunol* (2013) 191:6231–40. doi:10.4049/jimmunol.1301696
35. Genin P, Cuvelier F, Lambin S, Corte-Real Filipe J, Autrusseau E, Laurent C, et al. Optineurin regulates the interferon response in a cell cycle-dependent manner. *PLoS Pathog* (2015) 11:e1004971. doi:10.1371/journal.ppat.1004971
36. Perry AK, Chen G, Zheng D, Tang H, Cheng G. The host type I interferon response to viral and bacterial infections. *Cell Res* (2005) 15:407–22. doi:10.1038/sj.cr.7290309
37. Mankouri J, Fragkoudis R, Richards KH, Wetherill LF, Harris M, Kohl A, et al. Optineurin independently regulates the induction of IFNbeta in response to RNA virus infection. *PLoS Pathog* (2010) 6:e1000778. doi:10.1371/journal.ppat.1000778
38. Wild P, Farhan H, McEwan DG, Wagner S, Rogov VV, Brady NR, et al. Phosphorylation of the autophagy receptor optineurin restricts *Salmonella* growth. *Science* (2011) 333:228–33. doi:10.1126/science.1205405
39. Wong YC, Holzbaur EL. Optineurin is an autophagy receptor for damaged mitochondria in parkin-mediated mitophagy that is disrupted by an ALS-linked mutation. *Proc Natl Acad Sci U S A* (2014) 111:E4439–48. doi:10.1073/pnas.1405752111
40. Richter B, Sliter DA, Herhaus L, Stolz A, Wang C, Beli P, et al. Phosphorylation of OPTN by TBK1 enhances its binding to Ub chains and promotes selective autophagy of damaged mitochondria. *Proc Natl Acad Sci U S A* (2016) 113:4039–44. doi:10.1073/pnas.1523926113
41. Heo JM, Ordureau A, Paulo JA, Rinehart J, Harper JW. The PINK1-PARKIN mitochondrial ubiquitylation pathway drives a program of OPTN/NDP52 recruitment and TBK1 activation to promote mitophagy. *Mol Cell* (2015) 60:7–20. doi:10.1016/j.molcel.2015.08.016
42. Bansal M, Moharir SC, Sailasree SP, Sirohi K, Sudhakar C, Sarathi DP, et al. Optineurin promotes autophagosome formation by recruiting the autophagy-related Atg12-5-16L1 complex to phagophores containing the Wipi2 protein. *J Biol Chem* (2018) 293:132–47. doi:10.1074/jbc.M117.801944
43. Chew TS, O'Shea NR, Sewell GW, Oehlers SH, Mulvey CM, Crosier PS, et al. Optineurin deficiency in mice contributes to impaired cytokine secretion and neutrophil recruitment in bacteria-driven colitis. *Dis Model Mech* (2015) 8:817–29. doi:10.1242/dmm.020362
44. van Wijk SJL, Fricke F, Herhaus L, Gupta J, Hotte K, Pampaloni F, et al. Linear ubiquitination of cytosolic *Salmonella* Typhimurium activates NF-kappaB and restricts bacterial proliferation. *Nat Microbiol* (2017) 2:17066. doi:10.1038/nmicrobiol.2017.66
45. Bravo R, Parra V, Gatica D, Rodriguez AE, Torrealba N, Paredes F, et al. Endoplasmic reticulum and the unfolded protein response: dynamics and metabolic integration. *Int Rev Cell Mol Biol* (2013) 301:215–90. doi:10.1016/B978-0-12-407704-1.00005-1
46. Hetz C, Papa FR. The unfolded protein response and cell fate control. *Mol Cell* (2018) 69:169–81. doi:10.1016/j.molcel.2017.06.017
47. Todd DJ, Lee AH, Glimcher LH. The endoplasmic reticulum stress response in immunity and autoimmunity. *Nat Rev Immunol* (2008) 8:663–74. doi:10.1038/nri2359
48. Hotamisligil GS. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell* (2010) 140:900–17. doi:10.1016/j.cell.2010.02.034
49. Hetz C, Martinon F, Rodriguez D, Glimcher LH. The unfolded protein response: integrating stress signals through the stress sensor IRE1alpha. *Physiol Rev* (2011) 91:1219–43. doi:10.1152/physrev.00001.2011
50. Urano F, Wang X, Bertolotti A, Zhang Y, Chung P, Harding HP, et al. Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1. *Science* (2000) 287:664–6. doi:10.1126/science.287.5453.664
51. Kaser A, Lee AH, Franke A, Glickman JN, Zeissig S, Tilg H, et al. XBP1 links ER stress to intestinal inflammation and confers genetic risk for human inflammatory bowel disease. *Cell* (2008) 134:743–56. doi:10.1016/j.cell.2008.07.021
52. Niederreiter L, Fritz TM, Adolph TE, Krismer AM, Offner FA, Tschurtschenthaler M, et al. ER stress transcription factor Xbp1 suppresses intestinal tumorigenesis and directs intestinal stem cells. *J Exp Med* (2013) 210:2041–56. doi:10.1084/jem.20122341
53. Zhu X, Zhang J, Sun H, Jiang C, Dong Y, Shan Q, et al. Ubiquitination of inositol-requiring enzyme 1 (IRE1) by the E3 ligase CHIP mediates the IRE1/TRAF2/JNK pathway. *J Biol Chem* (2014) 289:30567–77. doi:10.1074/jbc.M114.562868
54. Gao B, Lee SM, Chen A, Zhang J, Zhang DD, Kannan K, et al. Synovialin promotes IRE1 ubiquitination and degradation in synovial fibroblasts from mice with collagen-induced arthritis. *EMBO Rep* (2008) 9:480–5. doi:10.1038/embor.2008.37
55. Deuring JJ, Fuhler GM, Konstantinov SR, Peppelenbosch MP, Kuipers EJ, de Haar C, et al. Genomic ATG16L1 risk allele-restricted Paneth cell ER stress in quiescent Crohn's disease. *Gut* (2014) 63:1081–91. doi:10.1136/gutjnl-2012-303527
56. Moretti J, Roy S, Bozec D, Martinez J, Chapman JR, Ueberheide B, et al. STING senses microbial viability to orchestrate stress-mediated autophagy of the endoplasmic reticulum. *Cell* (2017) 171:809–23.e13. doi:10.1016/j.cell.2017.09.034
57. Khaminets A, Heinrich T, Mari M, Grumati P, Huebner AK, Akutsu M, et al. Regulation of endoplasmic reticulum turnover by selective autophagy. *Nature* (2015) 522:354–8. doi:10.1038/nature14498
58. Schuck S, Gallagher CM, Walter P. ER-phagy mediates selective degradation of endoplasmic reticulum independently of the core autophagy machinery. *J Cell Sci* (2014) 127:4078–88. doi:10.1242/jcs.154716
59. Bernales S, Schuck S, Walter P. ER-phagy: selective autophagy of the endoplasmic reticulum. *Autophagy* (2007) 3:285–7. doi:10.4161/auto.3930
60. Mochida K, Oikawa Y, Kimura Y, Kirisako H, Hirano H, Ohsumi Y, et al. Receptor-mediated selective autophagy degrades the endoplasmic reticulum and the nucleus. *Nature* (2015) 522:359–62. doi:10.1038/nature14506
61. Song S, Tan J, Miao Y, Zhang Q. Crosstalk of ER stress-mediated autophagy and ER-phagy: involvement of UPR and the core autophagy machinery. *J Cell Physiol* (2017) 233(5):3867–74. doi:10.1002/jcp.26137
62. Sun S, Shi G, Sha H, Ji Y, Han X, Shu X, et al. IRE1alpha is an endogenous substrate of endoplasmic-reticulum-associated degradation. *Nat Cell Biol* (2015) 17:1546–55. doi:10.1038/ncb3266
63. Samie M, Lim J, Verschueren E, Baughman JM, Peng I, Wong A, et al. Selective autophagy of the adaptor TRIF regulates innate inflammatory signaling. *Nat Immunol* (2018) 19(3):246–54. doi:10.1038/s41590-017-0042-6
64. Ellinghaus D, Zhang H, Zeissig S, Lipinski S, Till A, Jiang T, et al. Association between variants of PRDM1 and NDP52 and Crohn's disease, based on exome sequencing and functional studies. *Gastroenterology* (2013) 145:339–47. doi:10.1053/j.gastro.2013.04.040
65. Thurston TL, Ryzhakov G, Bloor S, von Muhlinen N, Randow F. The TBK1 adaptor and autophagy receptor NDP52 restricts the proliferation of ubiquitin-coated bacteria. *Nat Immunol* (2009) 10:1215–21. doi:10.1038/ni.1800
66. Thurston TL, Wandel MP, von Muhlinen N, Foeglein A, Randow F. Galectin 8 targets damaged vesicles for autophagy to defend cells against bacterial invasion. *Nature* (2012) 482:414–8. doi:10.1038/nature10744
67. Ni J, Wu GD, Albenberg L, Tomov VT. Gut microbiota and IBD: causation or correlation? *Nat Rev Gastroenterol Hepatol* (2017) 14:573–84. doi:10.1038/nrgastro.2017.88
68. Bel S, Pends M, Wang Y, Li Y, Ruhn KA, Hassell B, et al. Paneth cells secrete lysozyme via secretory autophagy during bacterial infection of the intestine. *Science* (2017) 357:1047–52. doi:10.1126/science.aal4677
69. Zode GS, Sharma AB, Lin X, Searby CC, Bugge K, Kim GH, et al. Ocular-specific ER stress reduction rescues glaucoma in murine glucocorticoid-induced glaucoma. *J Clin Invest* (2014) 124:1956–65. doi:10.1172/JCI69774
70. Hetz C, Saxena S. ER stress and the unfolded protein response in neurodegeneration. *Nat Rev Neurol* (2017) 13:477–91. doi:10.1038/nrneuro.2017.99
71. Wang T, Xu W, Qin M, Yang Y, Bao P, Shen F, et al. Pathogenic mutations in the valosin-containing protein/p97(VCP) N-domain inhibit the SUMOylation of VCP and lead to impaired stress response. *J Biol Chem* (2016) 291:14373–84. doi:10.1074/jbc.M116.729343

72. Taylor RC, Dillin A. XBP-1 is a cell-nonautonomous regulator of stress resistance and longevity. *Cell* (2013) 153:1435–47. doi:10.1016/j.cell.2013.05.042
73. Fritz T, Niederreiter L, Adolph T, Blumberg RS, Kaser A. Crohn's disease: NOD2, autophagy and ER stress converge. *Gut* (2011) 60:1580–8. doi:10.1136/gut.2009.206466
74. Keestra-Gounder AM, Byndloss MX, Seyffert N, Young BM, Chavez-Arroyo A, Tsai AY, et al. NOD1 and NOD2 signalling links ER stress with inflammation. *Nature* (2016) 532:394–7. doi:10.1038/nature17631
75. Hooper KM, Barlow PG, Stevens C, Henderson P. Inflammatory bowel disease drugs: a focus on autophagy. *J Crohns Colitis* (2017) 11:118–27. doi:10.1093/ecco-jcc/jjx002.208

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Tschurtschenthaler and Adolph. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.