



Does HIV Exploit the Inflammatory Milieu of the Male Genital Tract for Successful Infection?

Rachel T. Esra¹, Abraham J. Olivier¹, Jo-Ann S. Passmore^{2,3}, Heather B. Jaspan¹,
Rushil Harryparsad¹ and Clive M. Gray^{1,3*}

¹Department of Pathology, Division of Immunology, Faculty of Health Sciences, Institute of Infectious Disease and Molecular Medicine, University of Cape Town, Cape Town, South Africa, ²Department of Pathology, Division of Virology, Faculty of Health Sciences, Institute of Infectious Disease and Molecular Medicine, University of Cape Town, Cape Town, South Africa, ³National Health Laboratory Services, Cape Town, South Africa

OPEN ACCESS

Edited by:

Philip Norris,
Blood Systems Research Institute,
USA

Reviewed by:

Francisco Veas,
Institut de recherche pour le
Développement (IRD), France
Roberto Nisini,
Istituto Superiore di Sanita', Italy

*Correspondence:

Clive M. Gray
clive.gray@uct.ac.za

Specialty section:

This article was submitted
to HIV and AIDS,
a section of the journal
Frontiers in Immunology

Received: 31 March 2016

Accepted: 10 June 2016

Published: 24 June 2016

Citation:

Esra RT, Olivier AJ, Passmore J-AS,
Jaspan HB, Harryparsad R and
Gray CM (2016) Does HIV Exploit the
Inflammatory Milieu of the Male
Genital Tract for Successful Infection?
Front. Immunol. 7:245.
doi: 10.3389/fimmu.2016.00245

In many parts of the World, medical male circumcision (MMC) is used as standard prevention of care against HIV infection. This is based on seminal reports made over 10 years ago that removal of the foreskin provides up to 60% protection against HIV infection in males and seems currently the best antiretroviral-free prevention strategy yet against the global epidemic. We explore the potential mechanisms by which MMC protects against HIV-1 acquisition and that one of the oldest, albeit re-invented, rituals of removing a foreskin underscores the exploitative nature of HIV on the anatomy and tissue of the uncircumcised penis. Furthermore, foreskin removal also reveals how males acquire HIV, and in reality, the underlying mechanisms of MMC are not known. We argue that the normal sequelae of inflammation in the male genital tract (MGT) for protection from sexually transmitted infections (STI)-induced pathology represents a perfect immune and microbial ecosystem for HIV acquisition. The accumulation of HIV-1 target cells in foreskin tissue and within the urethra in response to STIs, both during and after resolution of infection, suggests that acquisition of HIV-1, through sexual contact, makes use of the natural immune milieu of the MGT. Understanding immunity in the MGT, the movement of HIV-1 target cells to the urethra and foreskin tissue upon encounter with microbial signals would provide more insight into viral acquisition and lay the foundation for further prevention strategies in males that would be critical to curb the epidemic in all sexual partners at risk of infection. The global female-centric focus of HIV-1 transmission and acquisition research has tended to leave gaps in our knowledge of what determines HIV-1 acquisition in men and such understanding would provide a more balanced and complete view of viral acquisition.

Keywords: HIV-1, medical male circumcision, sexually transmitted infections, inflammation, risk factors, acquisition, exploitation

INTRODUCTION

There are approximately 35 million people living with HIV globally with Sub-Saharan Africa (SSA) accounting for more than two-thirds of global HIV/AIDS infections (1). Adolescent girls and young women are disproportionately affected by HIV in SSA, and 58% of the population living with HIV are estimated to be women (1). The high prevalence of HIV acquisition in females is

most likely to be multifactorial, associated with the presence of STIs, genital inflammation, and exogenous hormone contraceptive use (2, 3). There are also sociodemographic factors that may underlie higher prevalence in women, such as age discrepancies in relationships, gender-power imbalances, and gender-based violence that contribute the higher rates of infection in young women (2, 3). What is perhaps missing in these equations is how males acquire HIV, as male-to-female transmission is likely the source of transmission and a driver behind the high rates of infection measured in women. Men who have sex with men (MSM) are 19 times more likely to be living with HIV than the general population (4) and the prevalence of HIV in MSM is rising in several parts of the world (5). Little is known about the biological determinants of HIV transmission in male-to-male, male-to-female, or female-to-male sexual networks underlying the high prevalence of HIV in high-risk groups such as adolescents in SSA and MSM. One other possible determinant, perhaps overlooked, is that a high proportion MSM in Africa also report recent female sexual partners (6). HIV genotype studies at the early stages of the epidemic in South Africa showed that subtype B HIV-1 was confined mostly to the MSM community while subtype C circulated mostly in the heterosexual population, suggesting two separate HIV-1 introductions into this region (7). More recent studies in South Africa, Kenya and Senegal show that this has changed with MSM predominately being infected with the same variants as the heterosexual population (8–10). Conceivably, this might suggest a homogenization of epidemics between heterosexual and MSM sexual networks in the regions with the highest global prevalence of HIV (8, 10). Although other reviews focus on factors that influence female risks of HIV acquisition, few have focused on possible determinants of risk factors in males, which likely provides a niche in the male genital tract (MGT) conducive for HIV acquisition.

The low estimates of HIV transmission risk per sex act between males and females do little to explain the magnitude of the HIV epidemic, both globally and in SSA. Recent meta-analyses have reported the risk of heterosexual HIV acquisition in developed countries (expressed per 10,000 sexual exposures) to be four for insertive and eight for receptive penile–vaginal intercourse (11). The same analysis reports a significantly higher risk estimate for the MSM population with the risk of HIV acquisition to be 138 for receptive and 11 for insertive anal intercourse per 10,000 exposures (11). Estimates of the risk of HIV acquisition in SSA are difficult to accurately determine due to substantial heterogeneity in published data. Meta analyses from 14 sites in eastern and southern Africa have estimated that both men and women are at a higher risk of HIV acquisition through heterosexual contact when compared to data in developed countries: 10 and 9 per 10,000 exposures, respectively (12). These estimates raise relevant and important questions. Why is the magnitude of the HIV epidemic so high in SSA? Are there cofactors, both biological and/or social, that might explain the high rates of global heterosexual acquisition of HIV? HIV acquisition risk is unlikely to be defined by a “one size fits all” transmission probability; where age, other sexually transmitted infections (STIs), circumcision status, disease stage, antiretroviral (ARV) use, viral load, and viral subtype, all contribute to the efficiency of viral transmission (13).

There is no doubt that current research on HIV acquisition and prevention in females is of utmost importance, but this focus leaves large gaps of knowledge in the determinants of HIV acquisition, and hence transmission, in males. We know that medical male circumcision (MMC) is 56–61% effective at preventing HIV acquisition in men (13–15), but we do not fully understand how circumcision protects against HIV and other STIs at a structural, cellular, and molecular level. That such an ancient ritual of circumcision, albeit modernized, can have such a profound effect on preventing viral acquisition in males underscores the exploitative nature of HIV on the anatomy and tissue of the uncircumcised penis. How does HIV use the MGT environment and anatomical structures to promote its acquisition in men?

FORESKIN BARRIER INTEGRITY AND IMMUNITY

The protective effect of circumcision against HIV infection indicates that the foreskin is an important entry point of HIV in the MGT. The MGT consists of the penile urethra and the testes and comprises both simple and stratified epithelia (16). Although not necessary for normal penile functioning, the foreskin is thought to confer physical and immunological protection to the sensitive glans penis (**Figure 1A**) (17). The foreskin is a common site of entry for other STIs and is susceptible to micro-abrasions during intercourse, which likely leads to inflammation and resident HIV target cell exposure making it an important immunological area of the MGT (18). In the same way that HIV has evolved to exploit lymphoid structures to efficiently propagate systemically in the body (19), HIV utilizes the structures in the foreskin and urethra, along with the underlying immune cells, as a portal into lymphoid structures and systemic circulation.

The foreskin consists of a double layer of stratified epithelium containing squamous keratinocytes that covers the glans/corona and meatus of a flaccid penis (18). It is believed that the removal of the majority of the foreskin epithelium through circumcision results in a “dry” keratinized epithelium of the glans/corona that is thought to be more resistant to microabrasions during sexual intercourse and limiting the chances for HIV to contact target cells residing in dermis and epidermis, compared the “wet” mucosal epithelium of the glans/corona covered by the foreskin in uncircumcised men (25). The keratin layer is believed to provide the first line of innate defence against infection of the penile tissue although published data differs with regards to the degree of foreskin keratinization, and the difference between keratin thickness of the inner and outer foreskin tissues (26). In an uncircumcised penis, the foreskin retracts during erection, exposing the glans penis as well as the inner foreskin, which is thought to be more susceptible to viral acquisition compared to other penile tissues (27). This is based on observations from explant studies that have shown the inner foreskin to exhibit higher inflammatory cytokine responses and HIV target cell activation compared to the outer foreskin, leading to the hypothesis that HIV target cells in the inner foreskin have increased interaction with external factors (27, 28). During sexual intercourse, the inner foreskin is exposed to vaginal and/or rectal secretions and

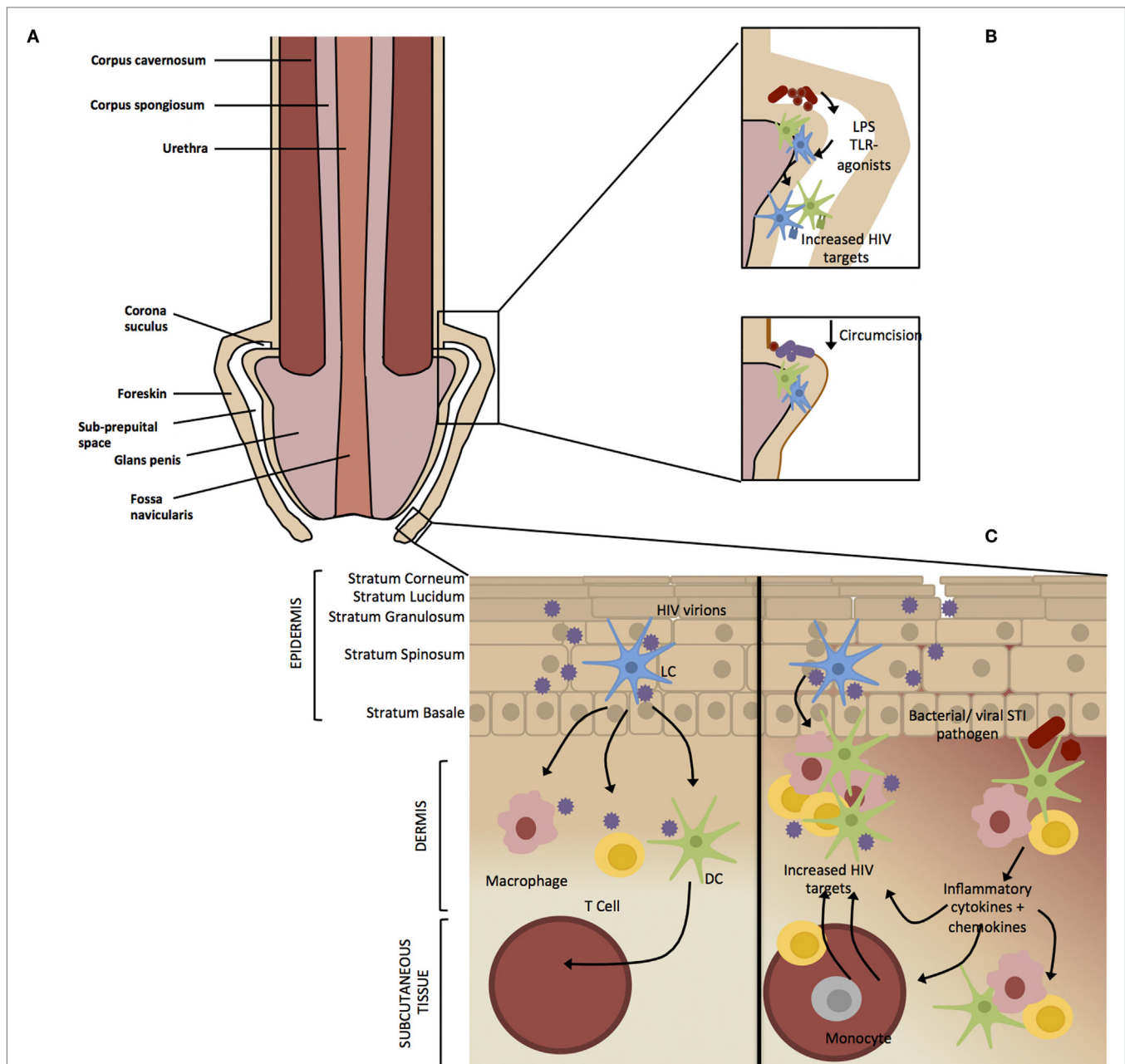


FIGURE 1 | Factors affecting HIV infection of the male genital tract. (A) The foreskin consists of a double layer of stratified epithelium that covers the glans/corona and meatus of a flaccid penis. **(B)** Circumcision results in the removal of the majority of the foreskin epithelium leaving a “dry” keratinized surface that is assumed to be resistant to HIV infection. Non-STI genital microbial populations have been shown to modulate genital inflammation through the antigen recognition, which may result in migration or activation of HIV target cells into the foreskin. Circumcision results in a removal of the moist sub-preputial space and decrease in anaerobic bacterial species, which are likely pro-inflammatory. **(C)** The foreskin is a stratified epithelium consisting of six epidermal layers namely stratum corneum (SC), stratum granulosum (SG), stratum spinosum (SS), and stratum basale (SB). HIV virions crosses the keratinized (light brown) epithelial border through micro abrasions that occur during sexual intercourse or through the formation of viral synapses between HIV-infected cells and epithelial cells (20). Langerhans’ Cells (LCs) reside within the epidermis where they are the first to encounter the virus (21). Activated LCs migrate into the lower dermal tissues, transferring the virus to resident dermal immune cells such as T cells, macrophages, and epithelial dendritic cells (DCs). Dendritic cells act as professional antigen presenting cells, phagocytosing virions, and migrating to the draining lymph nodes where they present HIV antigens to immature T cells and B cells (22). Intact virus can be trafficked into the lymph nodes by DCs and transferred to CD4⁺ T cells (23) where the virus replicates and is disseminated throughout the body (24). Asymptomatic STIs do not present clinically but inflammation still occurs on a cellular level. The release of pro-inflammatory cytokines by keratinocytes and dermal immune cells in response to an STI would result in recruitment and activation immune cells to the site of infection. This inflammatory environment would result in an accumulation of HIV target cells and therefore increase the efficiency of viral transmission in the case of an HIV infection.

is susceptible to micro-abrasions, possibly creating additional portals for HIV entry.

The foreskin was shown to contain a high density of the HIV target cells (29, 30), and a higher density of CD4⁺ T cells and langerin expressing Langerhans' cells (LCs) were identified in the inner compared to the outer foreskin (30, 31). Prodder et al. described an "inflammatory immune microenvironment" within the foreskin where they observed an upregulation of HIV co-receptors on CD4⁺ T cells as well as increased production of inflammatory cytokines, TNF- α , and IFN- γ by CD8⁺ T cells (32). A similar observation was made by Lemos et al., suggesting that the protective effects of circumcision may be due to the removal of inflamed tissue that is not only more permeable to HIV but would possess a higher density of HIV target cells (33).

Stratified epithelium, typically found on surfaces exposed to the environment, consists of multiple layers of epithelial cells forming a physical barrier against external pathogens (34). High numbers of immune cells, such as LCs, which serve as specialized antigen presenting cells, are typically present in stratified epithelium (35). LCs have been identified in the upper layers of the epidermis and are proposed to be the first cells to come into contact with HIV upon infection (36). Other HIV target cells such as CD4⁺ T cells, dendritic cells (DCs), and macrophages expressing C-type lectins reside in the lower dermis. The gp120 subunit of HIV Env binds the C-type lectin langerin expressed by circulating LCs where after the virus is either internalized and degraded or transferred to a CD4⁺ T cell, depending on viral load (21, 31). DCs express the C-type lectin DC-SIGN, which is also able to bind to gp120 and may promote transfer of HIV-1 from DCs to CD4⁺ T cells (23, 37). Macrophages are able to selectively capture HIV-1-infected CD4⁺ T-cells leading to the infection and efficient transfer of HIV-1 from cell-to-cell (38). These cells are found in surfaces typical of the meatus (distal end of penile urethra), fossa navicularis (opening), and foreskin (39). Keratinized stratified squamous epithelium spans the exterior of the glans penis and foreskin and transitions into non-keratinized stratified squamous epithelium in the fossa navicularis (**Figure 1A**) (16).

URETHRA: THE MUCOSAL SURFACE OF THE MGT

In contrast to the stratified squamous epithelium of the foreskin, a simple layer of non-keratinized pseudo stratified columnar epithelial cells is found along the penile urethra and provides a lower level of protection against HIV infection (**Figure 1A**) (39). The urethral epithelium has a high density of intraepithelial immune cells including CD8 T cells, natural killer cells, as well as HIV-1 targets CD4 T cells, DCs, and macrophages (16). As the urethra is the primary site of infection for many bacterial and viral STIs, this mucosal surface represents an important component of the penile protective immune tract, but also implicating it as a portal of entry to HIV and other STI that use these cells for productive infection during or after sexual intercourse (40, 41). Present knowledge of HIV infection through the urethra is incomplete due to the limited availability of fresh urethral samples, although cells expressing HIV-1 co-receptors have been identified at the urethral

opening (42). It is thus conceivable that the urethral opening may be an underestimated vulnerable site of HIV infection in circumcised men (25). An *ex vivo* study on penile tissue obtained following gender reassignment surgery found tissue explants from the foreskin, glans, meatus, and urethra to be equally susceptible to R5 HIV-1 infection (43, 44). Another study using penile tissues from cadavers found the inner foreskin to be more susceptible to HIV-1 infection compared to the outer foreskin, glans, shaft and urethral opening (44). Through the use of urethral tissue explants and the development of a novel *in vitro* reconstructed urethral mucosa, Ganor et al. identified the urethra as an entry site for HIV-1 infection, where urethral macrophages, and not T cells or LCs, were the initial HIV-1 target cells at the urethral opening (45). It is known that approximately 40% of men are not protected after MMC, and evidence pointing to the urethra as an HIV-1 portal suggests that further focus should be made at understanding viral acquisition at this unique mucosal surface.

THE IMPACT OF MMC AND OTHER STIs ON HIV RISK

Data from MMC trials give strong evidence for the protective effect of circumcision against HIV acquisition (13–15), although the findings were not as conclusive concerning other STIs. The Uganda trial showed that MMC of adolescent boys and men resulted in a 45% overall reduction in genital ulcerative disease (GUD) (46), and both Ugandan and South African trials showed a significant reduction in Herpes Simplex Virus-2 (HSV-2) acquisition (46, 47). Contradictory results emerged from the Kenyan trial indicating no significant decrease in HSV-2 infections as a result of MMC (48), suggesting that the reduction in HIV due to circumcision was independent of HSV-2 and GUD (48). Weiss et al. found that circumcision significantly reduced the risk of chancroid and syphilis but was only weakly associated with a reduction of HSV-2 infection (49). It was suggested that the warm, moist area beneath the foreskin encourages pathogenic growth and that stricter hygiene of the pre-nuptial space (between the foreskin and glans penis) is associated with lower HIV prevalence. This would indicate that the dynamic environment occupying this space has an effect on HIV risk (50, 51). Thus the removal of such an environmental niche for these pathogens is more protective against diseases that commonly cause lesions on the foreskin tissue, such as chancroid, compared to lesions caused by syphilis and HSV-2, both of which present more widely across the male genitalia (52). Follow up from the Kenyan trial demonstrated that circumcision did not decrease the risk of *Neisseria gonorrhoea* (NG), *Chlamydia trachomatis* (CT), and *Trichomonas vaginalis* and is thus not protective against non-ulcerative genital diseases (NUDs) (53). While there has been epidemiological evidence of decreased prevalence of cervical neoplasia if females with circumcised partners (54–56), observational studies have found inconsistent results with regards to impact of circumcision on the transmission of HPV to female partners (57, 58). Results from the Ugandan circumcision trial found a lower incidence of high-risk HPV infection in women with circumcised partners, leading the authors to hypothesize that male circumcision

involves a reduction of penile HPV carriage (59). Data on the effect of circumcision on HPV prevalence in men are inconsistent (60), and various studies have reported an increased prevalence of HPV infection in uncircumcised men (61). Hernandez et al. found that circumcision did not increase the risk HPV acquisition, but rather reduced viral clearance for both oncogenic and non-oncogenic forms of the virus (62). The higher prevalence of HPV in uncircumcised men therefore could be attributed to a longer duration of infection rather than increased acquisition (62), highlighting that pathogen-specific factors need to be considered in the epidemiology of STIs in men.

Despite the seeming disparity on the effectiveness of MMC in reducing the risk for other STIs, there is evidence to indicate reductions in the incidence of GUDs following MMC although the biological mechanism through which this would impact HIV risk is unclear. It is biologically plausible and likely that mucosal disruptions as result of ulcerative STIs would provide additional routes of transmission for HIV acquisition in males. In addition, inflammation from STIs could increase the efficiency of HIV infection by recruiting and activating HIV target cells that reside and migrate to the foreskin tissues (33). This inflammatory effect may persist after clearance of the infection as demonstrated by Donoval et al. who found a higher proportion of HIV target cells in the foreskins of men with a history of STIs compared to those who had no STI history (63). Sbazzo et al. have also suggested that the reduction of the highly vascularized frenulum, which is susceptible to trauma during sexual intercourse as well as ulcerative lesions, is one of the mechanisms by which circumcision prevents the synergy found between HIV and others STIs (64).

ARE ASYMPTOMATIC STIs ELEVATING HIV ACQUISITION RISK IN MALES?

Sexually transmitted infections have been identified as a significant factor increasing the risk of transmission and acquisition of HIV (65, 66), where pre-existing STIs increase the risk of HIV acquisition by two to threefold (67) and that ulcerative STIs are an even higher risk factor than non-ulcerative STIs. These findings led to the hypothesis that STI treatment may be an effective HIV intervention strategy in populations where HIV and other STIs are prevalent. Initial results from a randomized controlled trial (RCT) in Mwanza, Tanzania showing a 38% reduction of HIV incidence as a result of treating STIs (68). However, subsequent data from nine RCTs could not reproduce this finding and there was no significant reduction of HIV incidence when symptomatic STIs were treated (65). These incongruous results have brought into question whether symptomatic STI treatment should be seen as an HIV intervention (69). Such a syndrome-based approach to STI management, employed by multiple countries in SSA, may underestimate the impact of subclinical inflammation due to asymptomatic and non-ulcerative STIs that may contribute to HIV susceptibility. Regardless of visible ulcerations, HSV-2 has been linked to an approximately threefold increased risk of HIV acquisition.

A case for persistent immune activation was observed after successful treatment of HSV-2 with Acyclovir, where increased

expression of mucosal CCR5⁺ CD4⁺ T-cells remained at the sight of herpetic ulcers long after infection had been cleared (70). These findings would suggest that an initial immune activation event results in the persistence of HIV target cells in the MGT long after HSV-2 ulcerative resolution. Prodder et al. observed a higher proportion of HIV target CCR5⁺ T cells in the foreskins of Ugandan men with asymptomatic HSV-2 (71), supporting previous findings that HSV-2 infection increases the density of HIV target cells in the foreskin (72). Collectively, these data support the hypothesis that an asymptomatic or cleared HSV-2 infection may increase the susceptibility of the foreskin to HIV infection. In addition to an increased density of HIV target cells, asymptomatic HSV-2 has been associated with decreased expression of the epithelial junction protein Claudin in the foreskin, creating a compromised epithelial barrier that may be more susceptible to HIV-1 infection (73). Experiments using genital epithelial monolayers demonstrated that exposure to HIV-1 directly impairs mucosal barrier integrity through the disruption of tight junction markers, namely ZO-1 and Occludin enhancing HIV viral entry to sub mucosal targets (74). The disruption of the mucosal barrier, as a result of direct HIV exposure or coinfection with another STI, and the resultant epithelial cell induced inflammation (74) may be another mechanism HIV utilizes to gain access to target cells in the genital sub mucosa.

THE PENILE MICROBIOME AND IMMUNE INTEGRITY

The link between the microbiome and immunity has clearly been shown in the intestine (75, 76) and the way by which commensal microbiota, and released metabolites, shapes immunity in the skin is a focus of current research (77–79). Circumcision has been shown to alter the diversity and prevalence of the penile microbiota – a potential mechanism explaining the protective effect of circumcision against HIV (80, 81). Non-STI-genital microbial populations have been shown to modulate genital mucosal inflammation through antigen recognition and thus increase HIV risk through the activation of HIV target cells (82, 83). Price et al. showed that the microbiota of the coronal sulcus, the junction between the shaft and glans of the penis, predominately consists of anaerobic and putative vaginal taxa prior to circumcision and that anaerobes and skin taxa dominate following circumcision (81). Similarly distinct microbiota were shown to differentiate the coronal sulcus of circumcised and uncircumcised men (84). There was no difference between the microbial diversity within first pass urine samples, as a measure of urethral microbiome, collected from the same men, before and after circumcision, indicating that circumcision has no impact on the microbial diversity of the urethra (84). The moist sub-preputial space below the foreskin is thought to provide an anoxic microenvironment that harbors anaerobic bacterial species (81, 85). Predominantly anaerobic vaginal microbiota associated with bacterial vaginosis (BV) in women increases inflammation and HIV susceptibility (86, 87) in the female genital tract. Circumcision has been associated with a reduction of BV in female partners (88, 89) as well as a reduction of anaerobic bacterial species colonizing the coronal

suculus (81). The pro-inflammatory microbial community of the sub-preputial space may have a knock-on effect on other possible points of HIV entry as the foreskin covers the urethral opening in the majority of uncircumcised men (85). Based on this, it is postulated that the anaerobes on the uncircumcised penis are likewise pro-inflammatory, and thus result in migration or activation of HIV target cells into the foreskin (81), thereby increasing the likelihood of HIV infection through the urethra and foreskin in uncircumcised men (25) (**Figure 1B**).

DOES HIV-1 EXPLOIT THE MGT?

The role of subclinical inflammation in increasing susceptibility of the foreskin to HIV infection is unclear, especially in developing countries where a syndrome-based approach to STI management is followed. A high prevalence of commonly asymptomatic STIs such as CT and NG have been reported in high-risk MSM cohorts (90, 91), although there is little data on the prevalence of asymptomatic STIs in the general population. Asymptomatic STIs in men may be the driver of elevated pro-inflammatory cytokines and chemokines in foreskin tissue and the urethra, thereby recruiting activated HIV target cells to the site of infection (**Figure 1C**). Endocervical epithelial cells are the predominant niche for initial infection of *Chlamydia*, a primarily asymptomatic STI (92) and is one of the most prevalent STI in both men and women. *Chlamydia* infection leads to an inflammatory cascade associated with an influx of HIV-1 target cells as a result of the release of pro-inflammatory cytokines by epithelial cells (93). Buckner et al. showed that HIV infection of CD4⁺ CCR5⁺ cell lines is enhanced by exposure to supernatant from *Chlamydia*-infected epithelial cells. This infers that *Chlamydia* infection may facilitate viral infection in the local environment (94).

We thus posit that asymptomatic STIs increase the risk of HIV infection through subclinical inflammation that is modulated by epithelial dysbiosis. The inflammatory events in the MGT upon STI infection are a natural sequelae for protective immunity around the penile tissue. However, such subclinical inflammation in the foreskin/urethra of uncircumcised males and the urethra in circumcised men represent a perfect niche for HIV-1 to exploit in establishing a successful productive infection. Similar to the female genital tract and all human mucosal surfaces, innate immune defences such as mucus production, pattern recognition receptors, and antimicrobial peptide production are present in the MGT [Reviewed in Ref. (16)]. There is evidence that penile immunity is quite active in the FS, urethra, and within the epithelial tissues (95) and it would seem intuitive that immune protection within the MGT is a critical survival advantage to the host and therefore used to a survival advantage by HIV-1.

REFERENCES

1. UNAIDS. *UNAIDS: The Gap Report 26*. Geneva: Joint United Nations Programme on HIV/AIDS (2014).
2. Kelly RJ, Gray RH, Sewankambo NK, Serwadda D, Wabwire-Mangen F, Lutalo T, et al. Age differences in sexual partners and risk of HIV-1 infection in rural Uganda. *J Acquir Immune Defic Syndr* (2003) 32:446–51. doi:10.1097/00126334-200304010-00016

CONCLUSION

Medical male circumcision has been widely accepted as an effective intervention strategy for the heterosexual transmission of HIV (96, 97) and has been shown to be a more effective intervention strategy than STI treatments, vaccines, and microbicides (29). Complications as a result of MMC were low after the MMC trials (between 1.7 and 7.6%) and were mostly of minor clinical significance (98, 99). The prevalence of traditional circumcision performed in a non-clinical setting is thought to be between 20 and 80% in Eastern and Southern Africa (54) and has been associated with serious medical complications (100). Although the World Health Organisation reported 9.1 million MMCs performed between 2007 and 2014 in priority countries in East and Southern Africa (97), traditional circumcision is still prevalent in these regions for reasons that are both cultural and based on health service capacity (100). More research is needed to evaluate the impact of traditional circumcision on HIV transmission rates and well as on the acceptability of MMC in communities where traditional circumcision is practiced.

Little is known about mechanisms by which HIV gains access to the MGT. While circumcision has been shown to be an effective HIV intervention strategy, 40% of circumcised men are not protected after MMC, and condom usage is still regarded the best HIV prevention method, despite low adherence globally (101–103). The efficiency of MMC indicates that the removal of the foreskin reduces a natural environmental niche for HIV-1 acquisition in the MGT. Due to limited access to penile tissue, there are large gaps of knowledge regarding routes of HIV acquisition in males other than the foreskin. More studies on how HIV-1 has evolved to exploit the MGT architecture – utilizing subclinical inflammation and microbial dysbiosis to its advantage – are required to inform targeted intervention strategies that can prevent acquisition. In addition, a multidisciplinary approach marrying the biology of transmission and acquisition with the identity of sexual networks between MSM and heterosexual populations is a way to understand the HIV-1 epidemic and ways to mitigate transmission.

AUTHOR CONTRIBUTIONS

RE, devised and co-wrote the review. AO, co-wrote the review. J-AP, co-wrote the review. HJ, co-wrote the review. RH, co-wrote the review. CG, shaped and co-wrote the review.

FUNDING

Support for this work was from a Strategic Primer Award from the European and Developing Countries Trials Partnership (SP.2011.41304.038).

3. Langen TT. Gender power imbalance on women's capacity to negotiate self-protection against HIV/AIDS in Botswana and South Africa. *Afr Health Sci* (2005) 5:188–97.
4. UNAIDS. *Global AIDS response progress reporting 2014: Construction of Core Indicators for Monitoring the 2011 UN Political Declaration on HIV/AIDS*. Geneva: Joint United Nations Programme on HIV/AIDS (2014).
5. Beyrer C, Baral SD, Van Griensven F, Goodreau SM, Chariyaletsak S, Wirtz AL, et al. Global epidemiology of HIV infection in men who have

- sex with men. *Lancet* (2012) 380:367–77. doi:10.1016/S0140-6736(12)60821-6
6. Abara WE, Garba I. HIV epidemic and human rights among men who have sex with in sub-Saharan Africa: implications for HIV prevention, care, and surveillance. *Glob Public Health* (2015) 1692:1–14. doi:10.1080/17441692.2015.1094107
 7. van Harmelen J, Wood R, Lambrick M, Rybicki EP, Williamson AL, Williamson C. An association between HIV-1 subtypes and mode of transmission in Cape Town, South Africa. *AIDS* (1997) 11:81–7. doi:10.1097/00002030-199701000-00012
 8. Middelkoop K, Rademeyer C, Brown BB, Cashmore TJ, Marais JC, Scheibe AP, et al. Epidemiology of HIV-1 subtypes among men who have sex with men in Cape Town, South Africa. *J Acquir Immune Defic Syndr* (2014) 65:473. doi:10.1097/QAI.0000000000000067
 9. Ndiaye HD, Toure-Kane C, Vidal N, Niama FR, Niang-Diallo PA, Dièye T, et al. Surprisingly high prevalence of subtype C and specific HIV-1 subtype/CRF distribution in men having sex with men in Senegal. *J Acquir Immune Defic Syndr* (2009) 52:249–52. doi:10.1097/QAI.0b013e3181af70a4
 10. Tovnanabutra S, Sanders EJ, Graham SM, Mwangome M, Peshu N, McClelland RS, et al. Evaluation of HIV type 1 strains in men having sex with men and in female sex workers in Mombasa, Kenya. *AIDS Res Hum Retroviruses* (2010) 26:123–31. doi:10.1089/aid.2009.0115
 11. Patel P, Borkowf CB, Brooks JT, Lasry A, Lansky A, Mermin J. Estimating per-act HIV transmission risk. *AIDS* (2014) 28:1509–19. doi:10.1097/QAD.0000000000000298
 12. Hughes JP, Baeten JM, Lingappa JR, Magaret AS, Wald A, De Bruyn G, et al. Determinants of per-coital-act HIV-1 infectivity among African HIV-1-serodiscordant couples. *J Infect Dis* (2012) 205:358–65. doi:10.1093/infdis/jir747
 13. Auvert B, Taljaard D, Lagarde E, Sobngwi-Tambekou J, Sitta R, Puren A. Randomized, controlled intervention trial of male circumcision for reduction of HIV infection risk: the ANRS 1265 trial. *PLoS Med* (2005) 2:e298. doi:10.1371/journal.pmed.0020298
 14. Bailey RC, Moses S, Parker CB, Agot K, Maclean I, Krieger JN, et al. Male circumcision for HIV prevention in young men in Kisumu, Kenya: a randomised controlled trial. *Eur Urol* (2007) 52:605–6. doi:10.1016/j.eururo.2007.04.090
 15. Gray RH, Kigozi G, Serwadda D, Makumbi F, Watya S, Nalugoda F, et al. Male circumcision for HIV prevention in men in Rakai, Uganda: a randomised trial. *Lancet* (2007) 369:657–66. doi:10.1016/S0140-6736(07)60313-4
 16. Nguyen PV, Kafka JK, Ferreira VH, Roth K, Kaushic C. Innate and adaptive immune responses in male and female reproductive tracts in homeostasis and following HIV infection. *Cell Mol Immunol* (2014) 11:410–27. doi:10.1038/cmi.2014.41
 17. Taves DR. The intromission function of the foreskin. *Med Hypotheses* (2002) 59:180–2. doi:10.1016/S0306-9877(02)00250-5
 18. McCoombe SG, Short RV. Potential HIV-1 target cells in the human penis. *AIDS* (2006) 20:1491–5. doi:10.1097/01.aids.0000237364.11123.98
 19. Schacker TW, Brencley JM, Gregory J, Reilly C, Pambuccian SE, Taylor J, et al. Lymphatic tissue fibrosis is associated with reduced numbers of naïve CD4 + T cells in human immunodeficiency virus type 1 infection. *Clin Vaccine Immunol* (2006) 13:556–60. doi:10.1128/CVI.13.5.556-560.2006
 20. Bomsel M. Transcytosis of infectious human immunodeficiency virus across a tight human epithelial cell line barrier. *Nat Med* (1997) 3:42–7. doi:10.1038/nm0197-42
 21. De Witte L, Nabatov A, Pion M, Fluitsma D, de Jong MA, de Gruijl T, et al. Langerin is a natural barrier to HIV-1 transmission by Langerhans cells. *Nat Med* (2007) 13:367–71. doi:10.1038/nm1541
 22. Wu L. Biology of HIV mucosal transmission. *Curr Opin HIV AIDS* (2008) 3:534–40. doi:10.1097/COH.0b013e32830634c6
 23. Wu L, KewalRamani VN. Dendritic-cell interactions with HIV: infection and viral dissemination. *Nat Rev Immunol* (2006) 6:859–68. doi:10.1038/nri1960
 24. Shattock RJ, Moore JP. Inhibiting sexual transmission of HIV-1 infection. *Nat Rev Microbiol* (2003) 1:25–34. doi:10.1038/nrmicro729
 25. Anderson D, Politch JA, Pudney J. HIV infection and immune defense of the penis. *Am J Reprod Immunol* (2011) 65:220–9. doi:10.1111/j.1600-0897.2010.00941.x
 26. Jayathunge PHM, McBride WJH, Maclaren D, Kaldor J, Vallely A, Turville S. Male circumcision and HIV transmission; what do we know? *Open AIDS J* (2014) 8:31–44. doi:10.2174/1874613601408010031
 27. Fahrbach KM, Barry SM, Anderson MR, Hope TJ. Enhanced cellular responses and environmental sampling within inner foreskin explants: implications for the foreskin's role in HIV transmission. *Mucosal Immunol* (2010) 3:410–8. doi:10.1038/mi.2010.18
 28. Zhou Z, De Longchamps NB, Schmitt A, Zerbib M, Ganor Y, Bomsel M. HIV-1 efficient entry in inner foreskin is mediated by elevated CCL5/RANTES that recruits T cells and fuels conjugate formation with Langerhans cells. *PLoS Pathog* (2011) 7:e1002100. doi:10.1371/journal.ppat.1002100
 29. Padian NS, McLoy SI, Balkus JE, Wasserheit NJ. Weighing the gold in the gold standard: challenges in HIV prevention research. *Changes* (2012) 24:621–35. doi:10.1097/QAD.0b013e328337798a
 30. Hirbod T, Bailey RC, Agot K, Moses S, Ndinya-achola J, Murugu R, et al. Abundant expression of HIV target cells and C-type lectin receptors in the foreskin tissue of young Kenyan men. *Am J Pathol* (2010) 176:2798–805. doi:10.2353/ajpath.2010.090926
 31. Ganor Y, Zhou Z, Tudor D, Schmitt A, Vacher-Lavenu M-C, Gibault L, et al. Within 1 h, HIV-1 uses viral synapses to enter efficiently the inner, but not outer, foreskin mucosa and engages Langerhans-T cell conjugates. *Mucosal Immunol* (2010) 3:506–22. doi:10.1038/mi.2010.32
 32. Prodger JL, Gray R, Kigozi G, Nalugoda F, Galiwango R, Hirbod T, et al. Foreskin T cell subsets differ substantially from blood with respect to HIV co-receptor expression, inflammatory profile and memory status. *Mucosal Immunol* (2012) 5:121–8. doi:10.1038/mi.2011.56
 33. Lemos MP, Lama JR, Karuna ST, Fong Y, Montano SM, Ganoza C, et al. The inner foreskin of healthy males at risk of HIV infection harbors epithelial CD4 + CCR5 + cells and has features of an inflamed epidermal barrier. *PLoS One* (2014) 9:e108954. doi:10.1371/journal.pone.0108954
 34. Ross HM, Romrell LJKG. *Histology, a Text an Atlas*. 3rd ed. Baltimore: Williams & Wilkins (1995).
 35. Merad M, Sathe P, Helft J, Miller J, Mortha A. The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. *Annu Rev Immunol* (2013) 31:1146. doi:10.1146/annurev-immunol-020711-074950
 36. Hladik F, McElrath MJ. Setting the stage – HIV host invasion. *Nat Rev Immunol* (2008) 8:447–57. doi:10.1038/nri2302
 37. Geijtenbeek TB, Kwon DS, Torensma R, van Vliet SJ, van Duijnhoven GC, Middel J, et al. DC-SIGN, a dendritic cell-specific HIV-1-binding protein that enhances trans-infection of T cells. *Cell* (2000) 100:587–97. doi:10.1016/S0092-8674(00)80694-7
 38. Baxter AE, Russell RA, Duncan CJ, Moore MD, Willberg CB, Pablos JL, et al. Macrophage infection via selective capture of HIV-1-infected CD4+ T cells. *Cell Host Microbe* (2014) 16:711–21. doi:10.1016/j.chom.2014.10.010
 39. Anderson DJ, Pudney J. Human male genital tract immunity and experimental models. *Mucosal Immunol* (2005) 2:1647–59. doi:10.1016/B978-012491543-5/50100-5
 40. Hogan RJ, Mathews SA, Mukhopadhyay S, Summersgill JT, Timms P. Chlamydial persistence: beyond the biphasic paradigm. *Infect Immun* (2004) 72:1843–55. doi:10.1128/IAI.72.4.1843-1855.2004
 41. Edwards JL, Apicella MA. The molecular mechanisms used by *Neisseria gonorrhoeae* to initiate infection differ between men and women: the molecular mechanisms used by *Neisseria gonorrhoeae* to initiate infection differ between men and women. *Clin Microbiol Rev* (2004) 17:965–81. doi:10.1128/CMR.17.4.965-981.2004
 42. McClure CP, Tighe PJ, Robins RA, Bansal D, Bowman CA, Kingston M, et al. HIV coreceptor and chemokine ligand gene expression in the male urethra and female cervix. *AIDS* (2005) 19:1257–65. doi:10.1097/01.aids.0000180096.50393.96
 43. Fischetti L, Barry SM, Hope TJ, Shattock RJ. HIV-1 infection of human penile explant tissue and protection by candidate microbicides. *AIDS* (2009) 23:319–28. doi:10.1097/QAD.0b013e328321b778
 44. Dinh MH, Anderson MR, McRaven MD, Cianci GC, McCoombe SG, Kelley ZL, et al. Visualization of HIV-1 interactions with penile and foreskin epithelia: clues for female-to-male HIV transmission. *PLoS Pathog* (2015) 11:e1004729. doi:10.1371/journal.ppat.1004729
 45. Ganor Y, Zhou Z, Bodo J, Tudor D, Leibowitch J, Mathez D, et al. The adult penile urethra is a novel entry site for HIV-1 that preferentially targets resident urethral macrophages. *Mucosal Immunol* (2013) 6:776–86. doi:10.1038/mi.2012.116

46. Tobian AAR, Kigozi DS, Quinn TC, Kigozi G, Gravitt PE, Laeyendecker O, et al. Male circumcision for the prevention of HSV-2 and HPV infections and syphilis. *N Engl J Med* (2009) 360:1298–309. doi:10.1056/NEJMoa0802556
47. Sobngwi-Tambekou J, Taljaard D, Lissouba P, Zarca K, Puren A, Lagarde E, et al. Effect of HSV-2 serostatus on acquisition of HIV by young men: results of a longitudinal study in Orange Farm, South Africa. *J Infect Dis* (2009) 199:958–64. doi:10.1086/597208
48. Mehta SD, Moss S, Parker CB, Agot K, Maclean I, Bailey RC. Circumcision status and incident herpes simplex virus type 2 infection, genital ulcer disease, and HIV infection. *AIDS* (2012) 29:997–1003. doi:10.1097/QAD.0b013e328352d116
49. Weiss HA, Thomas SL, Munabi SK, Hayes RJ. Male circumcision and risk of syphilis, chancroid, and genital herpes: a systematic review and meta-analysis. *Sex Transm Infect* (2006) 82:101–9. doi:10.1136/sti.2005.017442
50. O'Farrell N, Morison L, Chung C-K. Low prevalence of penile wetness among male sexually transmitted infection clinic attendees in London. *Sex Transm Dis* (2007) 34:408–9. doi:10.1097/01.olq.0000240294.79426.a3
51. O'Farrell N, Morison L, Moodley P, Pillay K, Vanmali T, Quigley M, et al. Association between HIV and subpreputial penile wetness in uncircumcised men in South Africa. *J Acquir Immune Defic Syndr* (2006) 43:69–77. doi:10.1097/01.qai.0000225014.61192.98
52. Nsanze H, Fast MV, D'Costa LJ, Tukei P, Curran J, Ronald A. Genital ulcers in Kenya. Clinical and laboratory study. *Br J Vener Dis* (1981) 57:378–81.
53. Mehta SD, Moses S, Agot K, Parker C, Ndinya-achola JO, Maclean I, et al. Adult male circumcision does not reduce risk of incident *Neisseria gonorrhoeae*, *Chlamydia trachomatis*, and *Trichomonas vaginalis*: results from a randomized controlled trial in Kenya. *J Infect Dis* (2009) 200:370–8. doi:10.1086/600074
54. Drain PK, Halperin DT, Hughes JP, Klausner JD, Bailey RC. Male circumcision, religion, and infectious diseases: an ecologic analysis of 118 developing countries. *BMC Infect Dis* (2006) 6:172. doi:10.1186/1471-2334-6-85
55. Castellsagué X, Bosch F, Muñoz N, Meijer CJ, Shah KV, de Sanjose S. Male circumcision, penile human papillomavirus infection, and cervical cancer in female partners. *N Engl J Med* (2002) 346:1105–12. doi:10.1056/NEJMoa011688
56. Kjaer SK, de Villiers EM, Dahl C, Engholm G, Bock JE, Vestergaard BF, et al. Case-control study of risk factors for cervical neoplasia in Denmark. I: role of the “male factor” in women with one lifetime sexual partner. *Int J Cancer* (1991) 48:39–44. doi:10.1002/ijc.2910480108
57. Brinton LA, Reeves WC, Brenes MM, Herrero R, Gaitan E, Tenorio F, et al. The male factor in the etiology of cervical cancer among sexually monogamous women. *Int J Cancer* (1989) 44:199–203. doi:10.1002/ijc.2910440202
58. Dickson NP, Ryding J, Van Roode T, Paul C, Herbison P, Dillner J, et al. Male circumcision and serologically determined human papillomavirus infection in a birth cohort. *Cancer Epidemiol Biomarkers Prev* (2009) 18:177–83. doi:10.1158/1055-9965.EPI-08-0353
59. Wawer MJ, Tobian AA, Kigozi G, Kong X, Gravitt PE, Serwadda D, et al. Effect of circumcision of HIV-negative men on transmission of human papillomavirus to HIV-negative women: a randomized trial in Rakai, Uganda. *Lancet* (2011) 377:209–18. doi:10.1016/S0140-6736(10)61967-8
60. Zhu YP, Jia ZW, Dai B, Ye DW, Kong YY, Chang K, et al. Relationship between circumcision and human papillomavirus infection: a systematic review and meta-analysis. *Asian J Androl* (2016) 18:1–7. doi:10.4103/1008-682X.175092
61. Hernandez BY, Wilkens LR, Zhu X, McDuffie K, Thompson P, Shvetsov YB, et al. Circumcision and human papillomavirus infection in men: a site-specific comparison. *J Infect Dis* (2008) 197:787–94. doi:10.1086/528379
62. Hernandez BY, Shvetsov YB, Goodman MT, Wilkens LR, Thomson P, Zhu X, et al. Reduced clearance of penile human papillomavirus infection in uncircumcised men. *J Infect Dis* (2010) 201:1340–3. doi:10.1086/651607
63. Donoval BA, Landay AL, Moses S, Agot K, Ndinya-Achola JO, Nyagaya EA, et al. HIV-1 target cells in foreskin of African men with varying histories of sexually transmitted infections. *Am J Clin Pathol* (2006) 125:386–91. doi:10.1309/JVHQVDJYDKM58EPH
64. Sbazro R, Short RV. How does male circumcision protect against HIV infection? *BMJ* (2000) 320:1592–4. doi:10.1136/bmj.320.7249.1592
65. Stillwaggon E, Sowers L. Rush to judgment: the STI-treatment trials and HIV in sub-Saharan Africa. *J Int AIDS Soc* (2015) 18:19844. doi:10.7448/IAS.18.1.19844
66. Barnabas RV, Webb EL, Weiss HA, Wasserheit JN. The role of coinfections in HIV epidemic trajectory and positive prevention. *AIDS* (2011) 25:1559–73. doi:10.1097/QAD.0b013e3283491e3e
67. Sexton J, Garnett G, Röttingen J-A. Metaanalysis and meta-regression in interpreting study variability in the impact of sexually transmitted diseases on susceptibility to HIV infection. *Sex Transm Dis* (2005) 32:351–7. doi:10.1097/01.olq.0000154504.54686.d1
68. Grosskurth H, Mosha F, Todd J, Mwijarubi E, Klokke A, Senkoro K, et al. Impact of improved treatment of sexually transmitted diseases on HIV infection in rural Tanzania: randomised controlled trial. *Lancet* (1995) 346(8974):530–6. doi:10.1016/S0140-6736(95)91380-7
69. Gray RH, Wawer MJ. Reassessing the hypothesis on STI control for HIV prevention. *Lancet* (2008) 371:2064–5. doi:10.1016/S0140-6736(08)60896-X
70. Zhu J, Hladik F, Woodward A, Klock A, Peng T, Johnston C, et al. Persistence of HIV-1 receptor-positive cells after HSV-2 reactivation is a potential mechanism for increased HIV-1 acquisition. *Nat Med* (2009) 15:886–92. doi:10.1038/nm.2006
71. Procter JL, Gray R, Kigozi G, Nalugoda F, Nehemiah K, Kakanga M, et al. Impact of asymptomatic herpes simplex virus-2 infection on T cell function in the foreskin of men from Rakai, Uganda. *AIDS* (2014) 26:1319–22. doi:10.1097/QAD.0b013e328354675c
72. Johnson KE, Redd AD, Quinn TC, Collinson-streng AN, Cornish T, Kong X, et al. Effects of HIV-1 and herpes simplex virus type 2 infection on lymphocyte and dendritic cell density in adult foreskins from Rakai, Uganda. *J Infect Dis* (2011) 203:602–9. doi:10.1093/infdis/jiq091
73. Rohl M, Tjernlund A, Mehta SD, Pettersson P, Bailey RC, Broliden K. Comparable mRNA expression of inflammatory markers but lower claudin-1 mRNA levels in foreskin tissue of HSV-2 seropositive versus seronegative asymptomatic Kenyan young men. *BMJ Open* (2015) 5:e006627. doi:10.1136/bmjopen-2014-006627
74. Nazli A, Chan O, Dobson-Belaire WN, Ouellet M, Tremblay MJ, Gray-Owen SD, et al. Exposure to HIV-1 directly impairs mucosal epithelial barrier integrity allowing microbial translocation. *PLoS Pathog* (2010) 6:e1000852. doi:10.1371/journal.ppat.1000852
75. Kau AL, Ahern PP, Griffin NW, Goodman AL, Gordon JL. Human nutrition, the gut microbiome and the immune system. *Nature* (2011) 474:327–36. doi:10.1038/nature10213
76. Young VB. The intestinal microbiota in health and disease. *Curr Opin Gastroenterol* (2012) 28:63–9. doi:10.1097/MOG.0b013e32834d61e9
77. Weyrich LS, Dixit S, Farrer AG, Cooper AJ, Cooper AJ. The skin microbiome: associations between altered microbial communities and disease. *Australas J Dermatol* (2015) 56(4):268–74. doi:10.1111/ajd.12253
78. Skabytska Y, Kaesler S, Volz T, Biedermann T. How the innate immune system trains immunity: lessons from studying atopic dermatitis and cutaneous bacteria. *J Dtsch Dermatol Ges* (2016) 14:153–6. doi:10.1111/ddg.12843
79. Barnard E, Li H. Shaping of cutaneous function by encounters with commensals. *J Physiol* (2016) 594:1–17. doi:10.1111/jfj.12213
80. Liu CM, Hungate BA, Tobian AAR, Serwadda D, Ravel J, Lester R, et al. Male circumcision significantly reduces prevalence and load of genital anaerobic bacteria. *MBio* (2013) 4:1–9. doi:10.1128/mBio.00076-13
81. Price LB, Liu CM, Johnson KE, Aziz M, Lau MK, Bowers J, et al. The effects of circumcision on the penis microbiome. *PLoS One* (2010) 5:e8422. doi:10.1371/journal.pone.0008422
82. De Jong MA, Geijtenbeek TB. Human immunodeficiency virus-1 acquisition in genital mucosa: Langerhans cells as key-players. *J Intern Med* (2009) 265:18–28. doi:10.1111/j.1365-2796.2008.02046.x
83. Ogawa Y, Kawamura T, Kimura T, Ito M, Blauvelt A, Shimada S. Gram-positive bacteria enhance HIV-1 susceptibility in Langerhans cells, but not in dendritic cells, via toll-like receptor activation. *Blood* (2009) 113:5157–67. doi:10.1182/blood-2008-10-185728
84. Nelson DE, Dong Q, van der Pol B, Toh E, Fan B, Katz BP, et al. Bacterial communities of the coronal sulcus and distal urethra of adolescent males. *PLoS One* (2012) 7:e36298. doi:10.1371/journal.pone.0036298
85. O'Farrell N, Chung CK, Weiss HA. Foreskin length in uncircumcised men is associated with subpreputial wetness. *Int J STD AIDS* (2008) 19:821–3. doi:10.1258/ijsa.2008.008106
86. Atashili J, Poole C, Ndumbe PM, Adimora AA, Smith JS. Bacterial vaginosis and HIV acquisition: a meta-analysis of published studies. *AIDS* (2008) 22:1493–501. doi:10.1097/QAD.0b013e3283021a37

87. Myer L, Denny L, Telerant R, de Souza M, Wright TC Jr, Kuhn L. Bacterial vaginosis and susceptibility to HIV infection in South African women: a nested case-control study. *J Infect Dis* (2005) 192:1372–80. doi:10.1086/462427
88. Gray RH, Kigozi G, Serwadda D, Makumbi F, Nalugoda F, Watya S, et al. The effects of male circumcision on female partners' genital tract symptoms and vaginal infections in a randomized trial in Rakai, Uganda. *Am J Obstet Gynecol* (2009) 200:42.e1–7. doi:10.1016/j.ajog.2008.07.069
89. Liu CM, Hungate BA, Tobian AAR, Ravel J, Prodder JL, Serwadda D, et al. Penile microbiota and female partner bacterial vaginosis in Rakai, Uganda. *MBio* (2015) 6:17–20. doi:10.1128/mBio.00589-15
90. Lutz AR. Screening for asymptomatic extragenital gonorrhoea and chlamydia in men who have sex with men: significance, recommendations, and options for overcoming barriers to testing. *LGBT Health* (2015) 2:27–34. doi:10.1089/lgbt.2014.0056
91. Rebe K, Lewis D, Myer L, de Swardt G, Struthers H, Kamkuemah M, et al. A cross sectional analysis of gonococcal and chlamydial infections among men-who-have-sex-with-men in Cape Town, South Africa. *PLoS One* (2015) 10:e0138315. doi:10.1371/journal.pone.0138315
92. Irwin K. Observations from the CDC the silent epidemic of *Chlamydia trachomatis*: the urgent need for detection and treatment in women the problem: a highly prevalent asymptomatic infection linked. *J Womens Health Gender-Based Med* (2009) 9:339–43.
93. Ficarra M, Ibana JS, Poretta C, Ma L, Myers L, Taylor SN, et al. A distinct cellular profile is seen in the human endocervix during *Chlamydia trachomatis* infection. *Am J Reprod Immunol* (2008) 60:415–25. doi:10.1111/j.1600-0897.2008.00639.x
94. Buckner LR, Amedee AM, Albritton HL, Kozlowski PA, Lacour N, McGowin CL, et al. *Chlamydia trachomatis* infection of endocervical epithelial cells enhances early HIV transmission events. *PLoS One* (2016) 11:e0146663. doi:10.1371/journal.pone.0146663
95. Anderson D, Politch JA, Pudney J. HIV Infection of the penis. *Am J Reprod Immunol* (2010) 65:220–9. doi:10.1111/j.1600-0897.2010.00941.x
96. World Health Organisation, Joint United Nations Programme on HIV and AIDS. *WHO and UNAIDS Announce Recommendations from Expert Consultation on Male Circumcision for HIV Prevention*. Vol. 1. Geneva: World Health Organisation (2007).
97. WHO. *WHO Progress Brief – Voluntary Medical Male Circumcision for HIV Prevention in Priority Countries of East and Southern Africa*. Geneva: World Health Organisation (2014).
98. Weiss HA, Halperin D, Hayes RJ, Schmid G, Hankins CA. Male circumcision for HIV prevention: from evidence to action? *AIDS* (2008) 22:567–74. doi:10.1097/QAD.0b013e3282f3f406
99. Muula AS, Prozesky HW, Mataya RH, Ikechebelu JI. Prevalence of complications of male circumcision in anglophone Africa: a systematic review. *BMC Urol* (2007) 7:4. doi:10.1186/1471-2490-7-4
100. Wilcken A, Keil T, Dick B. Traditional male circumcision in eastern and southern Africa: a systematic review of prevalence and complications. *Bull World Health Organ* (2010) 88:907–14. doi:10.2471/BLT.09.072975
101. Bedimo AL, Bennett M, Kissinger P, Clark RA. Understanding barriers to condom usage among HIV-infected African American women. *J Assoc Nurses AIDS Care* (1998) 9:48–58. doi:10.1016/S1055-3290(98)80019-8
102. Ahmed S, Lutalo T, Wawer MJ, Serwadda D, Sewankambo NK, Nalugoda F, et al. HIV incidence and sexually transmitted disease prevalence associated with condom use: a population study in Rakai, Uganda. *AIDS* (2001) 15:2171–9. doi:10.1097/00002030-200111090-00013
103. Weller S, Davis-Beatty K. Condom effectiveness in reducing heterosexual HIV transmission. *Cochrane Database Syst Rev* (2002) 1:1–22. doi:10.1002/14651858.CD003255

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Esra, Olivier, Passmore, Jaspán, Harryparsad and Gray. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.