



Activation of PAD4 in NET formation

Amanda S. Rohrbach¹, Daniel J. Slade², Paul R. Thompson^{2*} and Kerri A. Mowen^{1*}

¹ Department of Chemical Physiology, The Scripps Research Institute, La Jolla, CA, USA

² Department of Chemistry, The Scripps Research Institute, Jupiter, FL, USA

Edited by:

Marko Radic, University of Tennessee, USA

Reviewed by:

Margaret E. Bauer, Indiana University School of Medicine, USA

Cecilia Garlanda, Istituto Clinico Humanitas, Italy

Paul Proost, Rega Institute, K. U. Leuven, Belgium

*Correspondence:

Paul R. Thompson, Department of Chemistry, The Scripps Research Institute, Scripps Florida, 130 Scripps Way, Jupiter, FL 33458, USA.

e-mail: pthompson@scripps.edu;

Kerri A. Mowen, Department of Chemical Physiology, The Scripps Research Institute, 10550 North Torrey Pines Road, La Jolla, CA 92037, USA.

e-mail: kmowen@scripps.edu

Peptidylarginine deiminases, or PADs, convert arginine residues to the non-ribosomally encoded amino acid citrulline in a variety of protein substrates. PAD4 is expressed in granulocytes and is essential for the formation of neutrophil extracellular traps (NETs) via PAD4-mediated histone citrullination. Citrullination of histones is thought to promote NET formation by inducing chromatin decondensation and facilitating the expulsion of chromosomal DNA that is coated with antimicrobial molecules. Numerous stimuli have been reported to lead to PAD4 activation and NET formation. However, how this signaling process proceeds and how PAD4 becomes activated in cells is largely unknown. Herein, we describe the various stimuli and signaling pathways that have been implicated in PAD4 activation and NET formation, including the role of reactive oxygen species generation. To provide a foundation for the above discussion, we first describe PAD4 structure and function, and how these studies led to the development of PAD-specific inhibitors. A comprehensive survey of the receptors and signaling pathways that regulate PAD4 activation will be important for our understanding of innate immunity, and the identification of signaling intermediates in PAD4 activation may also lead to the generation of pharmaceuticals to target NET-related pathogenesis.

Keywords: PAD4, citrullination, deimination, neutrophil, NET

THE PEPTIDYL ARGININE DEIMINASE FAMILY

The mammalian genome encodes 20 natural amino acids; however, this diversity is greatly increased by posttranslational modification of the original set to yield more than one hundred unique amino acids (Uy and Wold, 1977). Citrullination, or deimination, is the posttranslational modification of an arginine to a citrulline residue. Hydrolysis of the guanidino group of the arginine yields a ureido group and the loss of an ammonia (Figure 1). Citrullination is catalyzed by the peptidyl arginine deiminase family of enzymes, or PADs. This process results in the loss of positive charge and an approximately 1 Da increase in mass. While this modification seems quite modest, the loss of positive charge, and hydrogen bond acceptors, can have dramatic effects on cell signaling because these types of interactions are critical for stabilizing protein–protein, protein–DNA, and protein–RNA interactions. Additionally, this PTM may disrupt intra-molecular interactions, which could trigger major conformational changes in a protein, potentially altering intermolecular interactions and decreasing protein stability (Vossenaar et al., 2003).

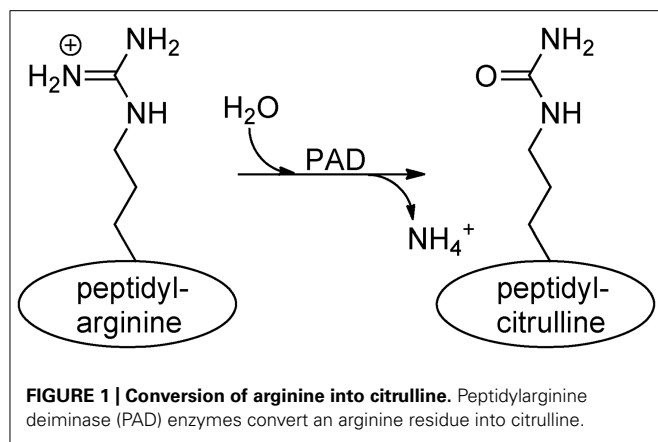
Five PAD enzymes are expressed in humans and mice, and the major difference between these isozymes appears to be tissue localization. PADs 1, 3, and 6 are expressed in the skin and uterus, hair follicles, and egg, ovary and embryo, respectively (Vossenaar et al., 2003; Wang et al., 2012). PAD4 expression has been reported in granulocytes, as well as some cancerous cell lines and tumors (Vossenaar et al., 2003; Chang et al., 2009; Jones et al., 2009; Hemmers et al., 2011; Wang et al., 2012), and most recently in mammalian oocytes and the preimplantation embryo (Brahmajosyula and Miyake, 2011). PAD2 has a much broader tissue expression profile and can be found in the CNS, skeletal muscle,

and cells of the immune system (Vossenaar et al., 2003). PADs 1, 2, and 4 are the only PADs expressed in the hematopoietic lineage, and, thus, are especially of immunological interest.

PADI genes, encoding the PAD enzymes, are located in a single gene cluster on chromosome 1p36.1 in humans and chromosome 4pE1 in mice (Vossenaar et al., 2003). The regions encoding the *PADI* locus in humans and mice span 334.7 and 230.4 kb, respectively (Vossenaar et al., 2003). All PAD enzymes are highly conserved, sharing at least 50% sequence homology among isozymes and 70% or greater homology of each vertebrate ortholog (Vossenaar et al., 2003). Eighteen per cent sequence identity is shared among all PADs (Vossenaar et al., 2003). Catalysis by all of these enzymes is calcium dependent, and, at least *in vitro*, requires calcium concentrations that are higher than that available in homeostatic cytoplasm, indicating calcium flux or a calcium-producing event is necessary to induce activity (Arita et al., 2004; Kearney et al., 2005; Raijmakers et al., 2007; Knuckley et al., 2010). Alternatively, a PTM or interacting protein may decrease the calcium concentration required for activation to physiologic levels.

PAD4

Of all of the PADs, PAD4 is of specific interest because of its importance in innate immunity and its putative role in a variety of pathogenic states, including autoimmune diseases, such as rheumatoid arthritis (RA), multiple sclerosis (MS), ulcerative colitis (UC), and systemic lupus erythematosus (SLE), and other inflammatory conditions, such as sepsis and thrombosis (Jones et al., 2009). Many autoantibodies in RA are directed against citrullinated proteins. In fact, the presence of anti-citrullinated protein



antibodies (ACPA) is a better predictor of RA than rheumatoid factor (Vossenaar et al., 2003), and, in 2011, ACPA were included in the new classification criteria for RA (Willemze et al., 2012). A genome-wide association study identified a PAD4 haplotype that is associated with RA in a Japanese population, albeit with a low odds ratio (OR = 1.14; Suzuki et al., 2003). The mutations in the PAD4 gene appear to confer prolonged stability to the transcript, leading to a model where increased expression of PAD4 in these populations would favor the generation of citrullinated self-epitopes to prime the autoimmune response (Suzuki et al., 2003). Though this association has been confirmed in other Asian populations, it has not been replicated in studies using patients from all Western European populations, indicating that the RA-associated PAD4 haplotype found in Asian RA patients can not explain the presence of ACPA's in all ethnicities (Klareskog et al., 2008). Interestingly, the PAD4 RA-associated disease haplotype has also been found in some Japanese patients with UC (Chen et al., 2008). MS patients have increased levels of the citrullinated form of myelin basic protein (Wood et al., 1996; Moscarello et al., 2007), and both PAD2 and PAD4 are overexpressed in the brains of MS patients (Wood et al., 2008). Finally, as will be discussed later in this review, in response to microbes, neutrophils can extrude their nuclear contents to form antimicrobial neutrophil extracellular traps (NETs; Brinkmann and Zychlinsky, 2007). Since PAD4 is essential for the formation of NETs (Li et al., 2010; Hemmers et al., 2011), PAD4 has also been implicated in NET-related pathologies, such as SLE and thrombosis, where NETs presumably promote deleterious inflammatory responses (Kessenbrock et al., 2009; Logters et al., 2009; Fuchs et al., 2010; Hakkim et al., 2010; Garcia-Romo et al., 2011; Villanueva et al., 2011). Thus, PAD4 may be a relevant target for several disease indications.

PAD4 is a 74 kDa protein that exists as a head-to-tail dimer (Arita et al., 2004; Liu et al., 2011). Each monomer consists of two N-terminal immunoglobulin (Ig) domains, formed by Ig subdomain 1, which contains nine β -sheets, and Ig subdomain 2, which contains 10 β -sheets and four short α -helices. The C-terminal catalytic domain adopts the α/β propeller fold that is characteristic of the deiminase superfamily (Shirai et al., 2001; Arita et al., 2004). The C-terminal catalytic domain is the most highly conserved area of the molecule (Vossenaar et al., 2003), suggesting that the active sites are likely quite similar among PADs. While a high degree of

conservation exists among PADs, PAD4 is the only family member to contain a classic nuclear localization sequence (56-PPAKKKST-63), found in Ig1 near the N-terminus, and, thus, is trafficked to the nucleus (Nakashima et al., 2002; Vossenaar et al., 2003; Arita et al., 2004). However, it is worth noting that recent data indicates that other PADs, most notably PAD2, are localized to the nucleus (Zhang et al., 2012).

PAD4 binds five calcium molecules, designated Ca1–Ca5, in a cooperative manner (Arita et al., 2004; Kearney et al., 2005; Liu et al., 2011). Ca1 and Ca2 bind in the C-terminal catalytic domain, and their binding induces major conformational changes that move several active site residues into positions that are competent for catalysis (Arita et al., 2004). This calcium-induced formation of the active site is unique to the PADs, and represents a novel mechanism of enzyme activation (Arita et al., 2004). Calcium binding also induces large structural changes in the N-terminus of the protein. For example, binding of Ca3, Ca4, and Ca5, along with two water molecules, induces the formation of the $\alpha 1$ α -helix, which is disordered in the apoenzyme (Arita et al., 2004). These conformational changes may provide, or remove, docking sites for other proteins, which may serve to further regulate PAD activity.

BIOCHEMICAL ACTIVATION OF PAD4

While it is unknown whether all PAD enzymes are capable of multimerizing, the dimer has been suggested to be required for PAD4 activity (Arita et al., 2004; Liu et al., 2011). However, the effects on enzyme activity and the calcium dependence of the enzyme are relatively minor (approximately twofold), and we routinely see robust enzyme activity at concentrations of protein that are an order of magnitude below the K_d of the dimer (Kearney et al., 2005). Nevertheless, dimer formation may represent a possible regulatory mechanism (Liu et al., 2011). Dimerization is mediated by both hydrophobic interactions and salt bridges between adjacent monomers (Arita et al., 2004; Liu et al., 2011).

The PADs display limited substrate specificity and citrullinate many proteins *in vitro*, preferring to modify arginine residues present in unstructured regions; the rate of substrate turnover is inversely proportional to the structural order of the substrate (Arita et al., 2006; Knuckley et al., 2010). Structurally, PAD4 interacts with the backbone atoms surrounding the site deimination, i.e., R-2, R-1, R0, and (R + 1), with few, if any, contacts with the side chains (Arita et al., 2006). The only requirement appears to be a small side chain at the R-2 position so as to avoid steric contacts with the active site (Arita et al., 2006). Upon binding to PAD4 the backbone of the substrate adopts a β -turn-like conformation within the substrate binding cleft (Arita et al., 2006), thereby explaining why the enzymes show such a high level of substrate promiscuity. In contrast to the situation *in vitro*, the PADs are believed to show greater substrate specificity *in vivo*. Presumably, interacting proteins modulate the substrate specificity of the enzyme or spatially target the enzyme to specific regions of the cell. For example, PAD4 is present in the nucleus and may be targeted to chromatin where it citrullinates a number of nuclear proteins, including the histones and protein arginine methyltransferase 1 (PRMT1; Vossenaar et al., 2003; Slack et al., 2011b). Although PAD4 was reported to convert monomethylated arginine residues to citrulline (Wang et al., 2004), this modification

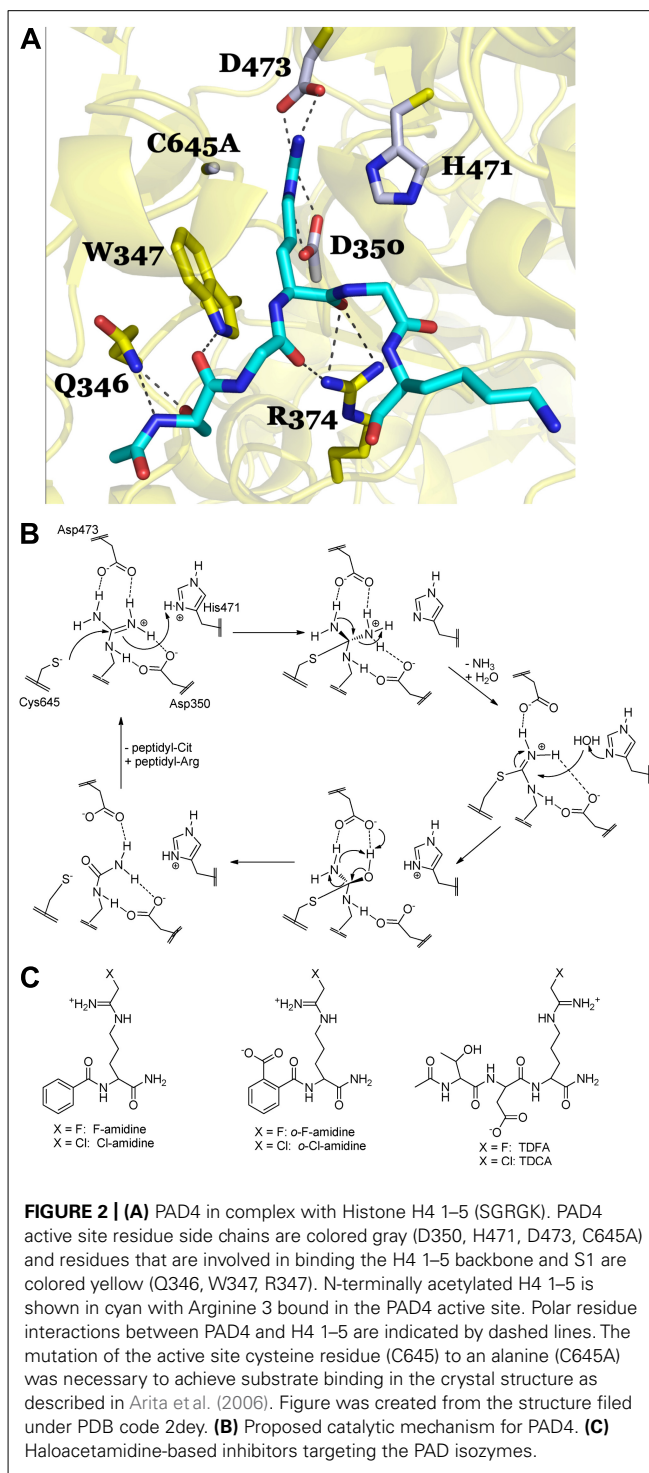
occurs at rates that are 10^2 - to 10^3 -fold slower than an unmodified arginine, suggesting that the, so-called “demethyliminium” reaction is not physiologically relevant (Hidaka et al., 2005; Kearney et al., 2005; Rajmakers et al., 2007; Thompson and Fast, 2006) and that citrullination simply antagonizes arginine methylation as originally suggested by Cuthbert et al. (2004).

In addition to the aforementioned protein substrates, PAD4, as well as the other PADs, autocitrullinate at several sites on the enzyme (Andrade et al., 2010; Mechin et al., 2010; Slack et al., 2011a). Although autocitrullination has been reported to directly modulate PAD4 activity (Andrade et al., 2010; Mechin et al., 2010), in our hands, this self-modification has no direct effect on enzyme activity, but it does appear to modulate protein–protein interactions (Slack et al., 2011b). For example, Slack et al. (2011b) demonstrated that citrullination of PAD4 reduces its ability to interact with PRMT1 and histone deacetylase (HDAC) 1, perhaps modulating its ability to alter gene transcription.

PAD MECHANISM AND INHIBITION

Given the substrate promiscuity of the PADs, it is unsurprising that the PADs also citrullinate a number of small molecule arginine mimics, including benzoyl arginine ethyl ester (BAEE) and benzoyl arginine amide (BAA). In fact, these compounds have served as important mechanistic probes of PAD4 catalysis and provided the molecular scaffold for the construction of the first highly potent PAD4 inhibitors. Below we highlight key mechanistic insights that guided the design of these inhibitors.

Briefly, there are four key catalytic residues, Asp350, His471, Asp473, and Cys645. Asp473 binds to both ω -nitrogens and Asp350 coordinates to one ω -nitrogen and the δ -nitrogen (Figure 2A). Cys645 and His471 lie on opposite sides of the guanidinium group and are appropriately positioned to promote catalysis via nucleophilic attack on the guanidinium carbon (Cys645) and protonation of the developing tetrahedral intermediate (His471; Figure 2B). Collapse of this intermediate leads to the loss of ammonia and the formation of the stable S-alkyl thiouronium intermediate that is subsequently hydrolyzed via a second tetrahedral intermediate to form citrulline; His471 likely activates the water molecule for nucleophilic attack (Figure 2B). Mechanistic studies (Knuckley et al., 2007, 2010), including mutagenesis, pH rate profile, solvent isotope effects, and solvent viscosity effects, as well as crystal structures of PAD4 bound to several substrates (i.e., BAA; Arita et al., 2006), and histone H3 and histone H4 tail analogs; Arita et al., 2004, 2006), and inhibitors (i.e., F-amidines, Cl-amidines, *o*-F-amidines, *o*-Cl-amidines, and TDFA; Luo et al., 2006a; Causey et al., 2011; Jones et al., 2012), provide strong support for the above mechanism and helped drive our thoughts about inhibitor design. For example, the presence of a reactive active site Cys prompted us to consider the synthesis of irreversible inhibitors (Knuckley et al., 2007, 2010). Additionally, the fact that mono-methylated arginine residues were exceptionally poor PAD substrates, as well as the steric restraints of the active site (Arita et al., 2004; Kearney et al., 2005; Rajmakers et al., 2007), told us that the reactive moiety would have to be relatively isosteric with respect to the substrate guanidinium. Furthermore, mutagenesis studies on the two active site aspartyl groups in PAD4, Asp350 and Asp473, showed that these residues are critical for



catalysis (Knuckley et al., 2007), indicating that proper positioning, hydrogen bonding, and electrostatic interactions between these residues and the substrate guanidinium are critical determinants for efficient substrate turnover, and would have to be maintained for efficient enzyme inactivation. As such, we initially focused our efforts on synthesizing F-amidines and Cl-amidines (Figure 2C), two haloacetamidines-containing compounds that we

hypothesized, and later confirmed, would inactivate the PADs by alkylating Cys645 (Luo et al., 2006a,b). These initial inhibitors, F-amidine and Cl-amidine, as well as our second generation compounds *o*-F-amidine, *o*-Cl-amidine, and TDFA (Knuckley et al., 2010; Causey et al., 2011), which show enhanced potency and selectivity, are bioavailable and have been used to show that the PADs play important roles in controlling gene transcription (Li et al., 2008; Yao et al., 2008; Jones et al., 2012; Zhang et al., 2011, 2012), fertility (Kan et al., 2012), differentiation (Slack et al., 2011c), and NET formation (Wang et al., 2009; Li et al., 2010). Additionally, Cl-amidine, or a Cl-amidine analog, decrease disease severity in animal models of RA, UC, nerve damage, and cancer (Chumanevich et al., 2011; Lange et al., 2011; Willis et al., 2011; Wang et al., 2012). Specifically, we were the first to show that the PAD inhibitor, Cl-amidine dose dependently decreased inflammation by up to 55% in the mouse collagen-induced arthritis (CIA) model of RA. Concomitant with the decreased severity there were significant decreases in the levels of citrullinated proteins, complement deposition, and epitope spreading (Willis et al., 2011). Similar dose-dependent effects were observed in the dextran sodium sulfate (DSS) model of UC where dosing of up to 75 mg/kg after the onset of disease led to significant reductions in weight loss, inflammation score, and colon lengthening (Chumanevich et al., 2011). The effects of Cl-amidine on nerve damage was examined in a chick embryo model of spinal cord injury where treatment with Cl-amidine reduced the abundance of deiminated histone 3, consistent with inhibition of PAD activity, and significantly reduced apoptosis and tissue loss following injury at embryonic day 15 (Lange et al., 2011). Finally, Wang et al. (2012) showed that a Cl-amidine analog, YW3-56, decreased tumor growth in a mouse sarcoma S-180 cell-derived solid tumor model and that additive effects on growth inhibition were observed when this compound was combined with the histone deacetylase inhibitor SAHA. We have observed similar effects with Cl-amidine in xenografts model of ductal carcinoma *in situ* (McElwee et al., 2012).

THE FUNCTION OF PAD4 IN NEUTROPHILS

Neutrophils are terminally differentiated granulocytes, which differentiate from hematopoietic stem cells in the bone marrow, and make up to 75% of white blood cells in the circulation (Ermer et al., 2009b). Mature neutrophils are released into circulation, where they have a very short life span of several hours to one day before undergoing apoptosis (Borregaard, 2010). Neutrophils are an important component of the innate immune system and form the first line of defense against invading pathogens, such as bacteria and fungi (Borregaard, 2010). Neutrophils contain an arsenal of antimicrobial proteins and peptides in primary (or azurophilic), secondary (or specific), and tertiary granules. Primary granules are mostly composed of proteases, such as myeloperoxidase and neutrophil elastase (NE), and antimicrobial peptides, such as defensins (Ermer et al., 2009b). Secondary and tertiary granules contain lactoferrin and gelatinase, respectively (Mantovani et al., 2011). Secretory granules also harbor stores of membrane proteins, such as the NADPH oxidase machinery (Nox; Borregaard, 2010), which can be trafficked to the surface of the cell quickly when necessary. In response to chemoattractants, neutrophils are guided to

areas of infection, where they respond with several effector mechanisms to invading pathogens, including phagocytosis, release of bactericidal products, and ROS production (Borregaard, 2010). Neutropenia, or the state of having too few neutrophils, leads to extreme immunosuppression and susceptibility to bacterial infections, which can be fatal (Janeway, 2005).

In 2004, Brinkmann et al. (2004) recognized the formation of NET structures, which were extruded by neutrophils in response to bacteria. NETs are composed of nuclear DNA that are decorated with a variety of nuclear and granular proteins, actively thrown out into the extracellular space, and result in death of the NET-producing cell (Brinkmann et al., 2004). Cell death by this mechanism is unique from apoptosis and necrosis and has been termed “NETosis” (Gupta et al., 2010). NETs ensnare extracellular bacteria, which are killed by the inherent antimicrobial properties of NET-associated proteins, such as histones (Parseghian and Luhrs, 2006), NE (Papayannopoulos and Zychlinsky, 2009), and lactoferrin (Papayannopoulos and Zychlinsky, 2009). These structures represent a novel method for pathogen killing, independent of both degranulation and phagocytosis, and have been shown to effectively kill a variety of pathogens, including bacteria, fungi, and protozoa (Papayannopoulos and Zychlinsky, 2009; Remijsen et al., 2011a). NETs have also been reported to occur in response to viral infection; however, they not appear to show any observable anti-viral capabilities (Hemmers et al., 2011). NETs may represent a killing mechanism for pathogens too large for the neutrophil to phagocytose, such as fungal hyphae or helminthes (Urban et al., 2006). Interestingly, bacteria have adapted defense mechanisms to NET formation. For example, the Group A *Streptococcus* express a DNase enzyme that can degrade NETs (Buchanan et al., 2006), and *Pseudomonas aeruginosa* expresses surface molecules that can prevent neutrophil activation and NET formation (Khatua et al., 2012).

Histone citrullination is thought to promote NET formation by inducing chromatin decondensation and facilitating the expulsion of chromosomal DNA coated with antimicrobial molecules (Neeli et al., 2008; Wang et al., 2009; Li et al., 2010). In fact, chemical inhibition of PAD4 activity significantly reduces histone decondensation and NET formation in response to either the calcium ionophore ionomycin or the bacterium *Shigella flexneri* (Wang et al., 2009). Our group and the Wang group have independently created PAD4-deficient mice (Li et al., 2010; Hemmers et al., 2011). Neutrophils isolated from PAD4-deficient mice are unable to citrullinate histones, decondense chromatin, and generate NETs (Li et al., 2010; Hemmers et al., 2011). Because of their inability to form NETs, PAD4 KO mice were shown to be more susceptible to bacterial infection (Li et al., 2010), and, thus, PAD4 is an important mediator of innate immunity.

Neutrophil elastase, a neutrophil serine protease, resides in the azurophilic granules and is a component of NETs (Borregaard, 2010; Amulic et al., 2012). The cleavage of microbial virulence factors by NE is essential for the clearance of specific Gram-negative bacteria (Pham, 2006). NE also cleaves histones to drive chromatin decondensation during NET formation (Papayannopoulos et al., 2010). Indeed, NE is essential for the process of NETosis (Papayannopoulos et al., 2010), and it is interesting to speculate that histone citrullination, by PAD4, promotes a relaxing

of the chromatin structure, allowing NE to gain access. Thus, the activity of NE and PAD4 may converge upon the chromatin decondensation process and NET formation. Neutrophils isolated from PAD4-deficient mice will be useful to delineate the hierarchy between PAD4 and other molecules, like NE, that are required for NETosis.

STIMULATION OF PAD4 ACTIVITY

A number of stimuli, including live and heat-killed bacteria, fungi, protozoa, and chemokines have been reported to induce NET formation (Guimaraes-Costa et al., 2012). Because NET formation is PAD4-dependent (Li et al., 2010; Hemmers et al., 2011), these same stimuli likely also induce PAD4 activation. However, the activity of PAD4 in relation to each stimuli must be assessed by looking for citrullination of histones, which is both a hallmark of PAD activity (Neeli et al., 2008, 2009) and a component of NETs (Li et al., 2010; Hemmers et al., 2011). Only a handful these, including live bacteria, the Gram-negative bacterial cell wall component lipopolysaccharide (LPS), the Gram-positive bacterial cell wall component lipoteichoic acid (LTA), the fungal cell wall component zymosan, the proinflammatory cytokine TNF α , and H₂O₂ have been shown to induce PAD4 activity and histone citrullination (Neeli et al., 2008, 2009; Li et al., 2010; Hemmers et al., 2011). As discussed earlier, PAD4 is calcium-dependent, and it is thought that PAD4 requires calcium levels higher than are found in the homeostatic cell to be active (Vossenaar et al., 2004). Not surprisingly, the calcium ionophore ionomycin activates PAD4 to induce histone citrullination and elicit NET formation (Neeli et al., 2008, 2009; Wang et al., 2009; Li et al., 2010). **Table 1** catalogs the variety of stimuli reported to stimulate NET formation.

Although little is known about the downstream signaling pathways required for PAD4 activation in neutrophils, cytoskeletal activity may be involved in PAD4 activation. Pretreatment of cells with nocodazole or cytochalasin D, which inhibit microtubule polymerization, prior to LPS stimulation leads to a reduction of histone citrullination and NET formation (Neeli et al., 2009). Additionally, blockade of integrin signaling through Mac-1 and cytohesin-1 impeded PAD4 activity and NET formation (Neeli et al., 2009). How cytoskeletal signaling impacts PAD4 is unknown; however, it has been proposed that the same receptors establish whether a cell will undergo phagocytosis or NET formation (Neeli et al., 2009). Indeed, studies have indicated that neutrophils initiate NET formation when phagocytosis of a large particle fails (Urban et al., 2006). Perhaps cytoskeletal activity and PAD4-mediated citrullination are linked because the initiation of NET formation represents a back-up killing mechanism following unsuccessful phagocytosis.

PAD4 ACTIVITY AND ROS

The generation of reactive oxygen species (ROS) is initiated by a wide variety of neutrophil stimuli, including phagocytosis of pathogens and signaling by LPS and TNF (Kohchi et al., 2009), which are also NET-inducing stimuli. Indeed, ROS generation is required for NET formation, and thus, it is likely that ROS generation is required for PAD4 activation as well. In neutrophils, superoxide (O₂^{•-}) is generated by the NADPH complex (Nox) and

by the electron transport chain in mitochondria (Kohchi et al., 2009). O₂^{•-} is then converted to H₂O₂ spontaneously or by the enzyme superoxide dismutase (SOD; Kohchi et al., 2009). H₂O₂ acts directly on target cells and is converted to additional effectors by enzymes such as myeloperoxidase (MPO). Interestingly, the addition of H₂O₂ to primary murine or human neutrophils induces PAD4-dependent histone citrullination (Neeli et al., 2008; Li et al., 2010). ROS molecules are highly cytotoxic and act as antimicrobial agents, but they can also play a dual role as reversible signal transduction mediators to regulate redox-sensitive target proteins (Amulic et al., 2012).

The link between ROS and NET formation was first recognized by the fact that patients with chronic granulomas disease (CGD), who are missing the Nox2 protein essential for NADPH assembly and, thus, cannot form ROS. Neutrophils isolated from CGD patients do not make NETs in response to *S. aureus* or phorbol myristate acetate (PMA; Fuchs et al., 2007). This phenotype is rescued by addition of H₂O₂ or exogenous glucose oxidase, which generates H₂O₂ (Fuchs et al., 2007), indicating that the ROS production facilitated by Nox2 is necessary for NETs. Catalase removes intracellular H₂O₂ by reduction to water and oxygen (Kohchi et al., 2009), and catalase inhibition increases intracellular H₂O₂, leading to increased NET production in healthy neutrophils (Fuchs et al., 2007). Subsequent studies have demonstrated that ROS generation is upstream of chromatin decondensation (Remijsen et al., 2011b), suggesting that NADPH oxidase activation may also be a prerequisite for PAD4 activation. Indeed, LPS-induced citrullination of histone H4 is decreased when cells are pre-incubated with the NADPH oxidase inhibitor apocynin (Neeli et al., 2009). Although, to our knowledge, the activity of PAD4 in CGD neutrophils has not yet been directly explored, since chromatin decondensation is not observed in CGD neutrophils, we would predict PAD4-mediated histone citrullination is also impaired. Since H₂O₂ treatment can activate PAD4-mediated histone deimination in primary murine and human neutrophils (Neeli et al., 2008; Li et al., 2010), and since NADPH activation seems to be upstream of NET formation (Neeli et al., 2009), we speculate that PAD4 activation may be downstream of NADPH (**Figure 3**).

The contribution of specific ROS molecules to NET formation has also been examined. NADPH oxidase or mitochondrial ROS selective inhibitors established a requirement for NADPH oxidase generated O₂^{•-} (Nishinaka et al., 2011), but excluded a need for mitochondrial ROS in PMA-induced NET generation (Kirchner et al., 2012). MPO catalyzes the oxidation of Cl⁻ ions to generate hypochlorous acid (HOCl) and other reactive products using H₂O₂ as a cosubstrate (Arnhold and Flemmig, 2010). In the absence of MPO activity, NET generation is absent (Papayannopoulos et al., 2010; Metzler et al., 2011; Kirchner et al., 2012), but this phenotype can be rescued by addition of HOCl (Palmer et al., 2012). In fact, the HOCl product of MPO has also been found to be both necessary and sufficient for NET formation, and in CGD neutrophils, the addition of HOCl is also sufficient to initiate NET formation (Palmer et al., 2012). Taurine is a cellular antioxidant capable of reducing HOCl and H₂O₂ to promote cell survival (Palmer et al., 2012). Accordingly, preincubation of neutrophils with taurine prior to PMA or HOCl stimulation reduces NET formation (Palmer et al., 2012). Additionally, while SOD

Table 1 | NET/PAD4 stimuli.

NET stimuli	Shown to activate PAD4	Reference
Activated endothelial cells	n.d.	Gupta et al. (2010)
<i>Aspergillus fumigatus</i>	n.d.	Bruns et al. (2010), McCormick et al. (2010)
<i>Candida albicans</i>	n.d.	Ermert et al. (2009a), Urban et al. (2009), Yost et al. (2009), Hakkim et al. (2010), Svobodova et al. (2012)
Oposonized <i>Candida albicans</i>	n.d.	Metzler et al. (2011)
<i>Cryptococcus</i> species	n.d.	Urban et al. (2009), Springer et al. (2010)
<i>Escherichia coli</i>	Yes	Grinberg et al. (2008), Wang et al. (2009), Yost et al. (2009), Yan et al. (2012)
f-MLP	Yes	Neeli et al. (2008)
H ₂ O ₂	Yes	Neeli et al. (2008), Li et al. (2010)
<i>Haemophilus influenzae</i>	n.d.	Juneau et al. (2011)
IL-8+ <i>Shigella flexneri</i>	Yes	Wang et al. (2009)
IL-8 ¹	n.d.	Brinkmann et al. (2004), Gupta et al. (2005)
Ca ²⁺ ionophore	Yes	Wang et al. (2009)
<i>Klebsiella pneumoniae</i>	n.d.	Papayannopoulos et al. (2010)
<i>Leishmania</i> species	n.d.	Guimaraes-Costa et al. (2009), Gabriel et al. (2010)
<i>Listeria monocytogenes</i>	n.d.	Ermert et al. (2009a), Munafo et al. (2009)
LPS	Yes	Neeli et al. (2008), Li et al. (2010), Munafo et al. (2009)
Lipoteichoic acid	Yes	Neeli et al. (2009)
<i>Mycobacterium</i> species	n.d.	Ramos-Kichik et al. (2009)
Nitric Oxide	n.d.	Patel et al. (2010)
Platelet activating factor	n.d.	Yost et al. (2009), Fuchs et al. (2010)
Platelet TLR-4	n.d.	Clark et al. (2007)
Phorbol-12-myristate-13-acetate (PMA)	Yes	Brinkmann et al. (2004), Ermert et al. (2009a), Munafo et al. (2009), Neeli et al. (2009), Chow et al. (2010), Li et al. (2010), Papayannopoulos et al. (2010), Pilszczek et al. (2010), Lin et al. (2011), Metzler et al. (2011), Remijsen et al. (2011b), Villanueva et al. (2011), Menegazzi et al. (2012), Palmer et al. (2012), Parker et al. (2012), Yan et al. (2012)
<i>Pseudomonas aeruginosa</i>	n.d.	Pilszczek et al. (2010)
<i>Salmonella typhimurium</i>	n.d.	Brinkmann et al. (2004)
<i>Shigella flexneri</i>	Yes	Brinkmann et al. (2004), Li et al. (2010)
<i>Staphylococcus aureus</i>	n.d.	Brinkmann et al. (2004), Chow et al. (2010), Pilszczek et al. (2010)
Oposonized <i>Staphylococcus aureus</i>	n.d.	Palmer et al. (2012)
<i>Staphylococcus epidermidis</i> δ -toxin	n.d.	Cogen et al. (2010)
<i>Streptococcus</i> species	Yes ²	Beiter et al. (2006), Buchanan et al. (2006), Lauth et al. (2009), Oehmcke et al. (2009), Crotty Alexander et al. (2010), Li et al. (2010), Pilszczek et al. (2010)
<i>Streptococcus pneumoniae</i>	n.d.	Mori et al. (2012)
α -Enolase		
TNF α	Yes	Neeli et al. (2009), Wang et al. (2009)
<i>Toxoplasma gondii</i>	n.d.	Abi Abdallah et al. (2012)
<i>Yersinia enterocolitica</i>	n.d.	Casutt-Meyer et al. (2010)
Zymosan	Yes	Neeli et al. (2009)

n.d., not determined.

¹ Some investigators have reported that IL-8-induced NET formation may be sensitive to cell culture conditions (Marcos et al., 2011).

² M1 GAS deficient in an extracellular DNase (*Sda1*) could induce PAD4-dependent NETs (Li et al., 2010).

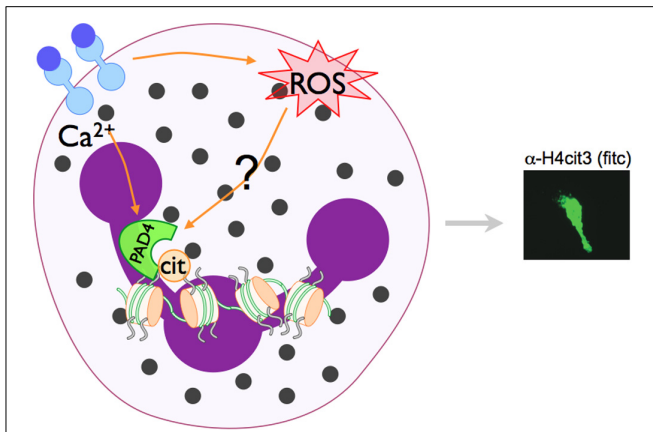


FIGURE 3 | Model of PAD4 activation in NET formation. Pathways that activate NET formation are less defined than phagocytic pathways, but are known to require NADPH oxidase activity and the activation of PAD4 and subsequent histone citrullination. PAD enzymes are Ca^{2+} -dependent. Since PAD4-mediated histone citrullination is abrogated by the NADPH inhibitor apocynin (Neeli et al., 2009), we speculate that NADPH regulated ROS generation and increase Ca^{2+} levels may converge to activate PAD4 in neutrophils.

inhibition does not impede NET formation, addition of exogenous SOD does seem to increase NET production, perhaps owing to the increase in available H_2O_2 (Palmer et al., 2012). These studies indicate a model in which NADPH oxidase activity generates $\text{O}_2^{\bullet-}$, which then dismutates to H_2O_2 either spontaneously or with the help of SOD, and is then used by MPO to generate HOCl, which is necessary and sufficient to induce NET formation. It will be interesting to determine whether HOCl can also directly regulate PAD4 activation.

PAD4 AND AUTOPHAGY

Like ROS, autophagy has been shown to be required for chromatin decondensation during NET generation (Remijsen et al., 2011b); however, these two processes seem to be independent of each other (Remijsen et al., 2011b). Blockade of PI3K with wortmannin inhibits autophagy, and pretreatment of PMA stimulated neutrophils with wortmannin prevented chromatin decondensation (Remijsen et al., 2011b). However, no direct role between autophagy, PI3K and citrullination has been shown. Recently, newly developed PAD4 inhibitors were found to reduce autophagy processes in an osteosarcoma cell line (Wang et al., 2012), further providing evidence of a link between PAD4 activity and autophagy.

REFERENCES

- Abi Abdallah, D. S., Lin, C., Ball, C. J., King, M. R., Duhamel, G. E., and Denkers, E. Y. (2012). *Toxoplasma gondii* triggers release of human and mouse neutrophil extracellular traps. *Infect. Immun.* 80, 768–777.
- Amulic, B., Cazalet, C., Hayes, G. L., Metzler, K. D., and Zychlinsky, A. (2012). Neutrophil function: from mechanisms to disease. *Annu. Rev. Immunol.* 30, 459–489.
- Andrade, F., Darrah, E., Gucek, M., Cole, R. N., Rosen, A., and Zhu, X. (2010). Autocitrullination of human peptidyl arginine deiminase type 4 regulates protein citrullination during cell activation. *Arthritis Rheum.* 62, 1630–1640.
- Arita, K., Hashimoto, H., Shimizu, T., Nakashima, K., Yamada, M., and Sato, M. (2004). Structural basis for Ca^{2+} -induced activation of human PAD4. *Nat. Struct. Mol. Biol.* 11, 777–783.
- Arita, K., Shimizu, T., Hashimoto, H., Hidaka, Y., Yamada, M., and Sato, M. (2006). Structural basis for histone N-terminal recognition by human peptidylarginine deiminase 4. *Proc. Natl. Acad. Sci. U.S.A.* 103, 5291–5296.
- Arnhold, J., and Flemmig, J. (2010). Human myeloperoxidase in innate and acquired immunity. *Arch. Biochem. Biophys.* 500, 92–106.
- Beiter, K., Wartha, F., Albiger, B., Normark, S., Zychlinsky, A., and Henriques-Normark, B. (2006). An endonuclease allows *Streptococcus pneumoniae* to escape from neutrophil extracellular traps. *Curr. Biol.* 16, 401–407.
- Borraagaard, N. (2010). Neutrophils, from marrow to microbes. *Immunity* 33, 657–670.
- Brahmajosyula, M., and Miyake, M. (2011). Localization and expression of peptidylarginine deiminase 4 (PAD4) in mammalian oocytes and preimplantation embryos. *Zygote*

CONCLUSION

Neutrophil extracellular traps have been reported in several pathological scenarios, including SLE, sepsis, thrombosis, and infectious disease, and may induce or exacerbate inflammation through prolonged inflammatory response, tissue damage, and presentation of neo-antigens. Indeed, our group recently described the PAD4-dependent formation of NETs in a murine model of the effector phase of RA; however, PAD4 was dispensable for disease in this model (Rohrbach et al., 2012). Pathology caused or exacerbated by NETs is an expanding field of research. Because PAD4 is required for NET formation, inhibition of PAD4 activity may improve clinical outcomes in patients experiencing these inflammatory diseases. In fact, PAD inhibitors have demonstrated efficacy in a variety of immune pathologies (Chumanevich et al., 2011; Lange et al., 2011; Slack et al., 2011c; Wang et al., 2012; Willis et al., 2011). Of course, because PAD4 is required for microbial-induced NET formation (Li et al., 2010; Hemmers et al., 2011), the side effects of PAD4-targeted therapeutics may also include increased susceptibility to infectious diseases.

PAD4 is an important component in the innate immune system, however its activity has been linked to a wide variety of disease states, including cancer, autoimmunity, and other inflammatory conditions. Despite its significance, little is understood about how the PAD4 enzyme becomes active in order to impart its helpful and harmful effects. At the protein level, calcium binding, dimerization, and autocitrullination may help regulate its activity. ROS may also play a role in regulating PAD4 activation (see model in Figure 3), and recently associations between PAD4 activity and autophagy have been proposed. Despite these efforts, much is left to understand about PAD4 enzyme regulation. Anti-PAD4 therapies have been proposed for inflammatory conditions and cancer, thus a more comprehensive understanding of the pathways that activate PAD4 in neutrophils will be important.

ACKNOWLEDGMENTS

The work in Dr. Mowen's laboratory was supported grants GM085117 and AI067460 from the National Institutes of Health (Kerri A. Mowen). The work in Dr. Thompson's laboratory was supported by grants GM079357 and CA151304 from the National Institutes of Health as well as an American College of Rheumatology Research Foundation Within Our Reach Grant. We apologize to investigators whose important contributions were not included due to space limitations. This is manuscript #21941 from The Scripps Research Institute.

- 1–11. doi: 10.1017/S0967199411000633 [Epub ahead of print].
- Brinkmann, V., and Zychlinsky, A. (2007). Beneficial suicide: why neutrophils die to make NETs. *Nat. Rev. Microbiol.* 5, 577–582.
- Brinkmann, V., Reichard, U., Goosmann, C., Fauler, B., Uhlemann, Y., Weiss, D. S., et al. (2004). Neutrophil extracellular traps kill bacteria. *Science* 303, 1532–1535.
- Bruns, S., Kniemeyer, O., Hasenberg, M., Aimaniananda, V., Nietzsche, S., Thywissen, A., et al. (2010). Production of extracellular traps against *Aspergillus fumigatus* in vitro and in infected lung tissue is dependent on invading neutrophils and influenced by hydrophobin RodA. *PLoS Pathog.* 6, e1000873. doi: 10.1371/journal.ppat.1000873
- Buchanan, J. T., Simpson, A. J., Aziz, R. K., Liu, G. Y., Kristian, S. A., Kotb, M., et al. (2006). DNase expression allows the pathogen group A *Streptococcus* to escape killing in neutrophil extracellular traps. *Curr. Biol.* 16, 396–400.
- Casutt-Meyer, S., Renzi, F., Schmalzer, M., Jann, N. J., Amstutz, M., and Cornelis, G. R. (2010). Oligomeric coiled-coil adhesin YadA is a double-edged sword. *PLoS ONE* 5, e15159. doi: 10.1371/journal.pone.0015159
- Causey, C. P., Jones, J. E., Slack, J. L., Kamei, D., Jones, L. E., Subramanian, V., et al. (2011). The development of *N*-alpha-(2-carboxyl)benzoyl-*N*(5)-(2-fluoro-1-iminoethyl)-*L*-ornithine amide (*o*-F-amidine) and *N*-alpha-(2-carboxyl)benzoyl-*N*(5)-(2-chloro-1-iminoethyl)-*L*-ornithine amide (*o*-Cl-amidine) as second generation protein arginine deiminase (PAD) inhibitors. *J. Med. Chem.* 54, 6919–6935.
- Chang, X., Han, J., Pang, L., Zhao, Y., Yang, Y., and Shen, Z. (2009). Increased PADI4 expression in blood and tissues of patients with malignant tumors. *BMC Cancer* 9, 40. doi: 10.1186/1471-2407-9-40
- Chen, C. C., Isomoto, H., Narumi, Y., Sato, K., Oishi, Y., Kobayashi, T., et al. (2008). Haplotypes of PADI4 susceptible to rheumatoid arthritis are also associated with ulcerative colitis in the Japanese population. *Clin. Immunol.* 126, 165–171.
- Chow, O. A., von Kockritz-Blickwede, M., Bright, A. T., Hensler, M. E., Zinkernagel, A. S., Cogen, A. L., et al. (2010). Statins enhance formation of phagocyte extracellular traps. *Cell Host Microbe* 8, 445–454.
- Chumanevich, A. A., Causey, C. P., Knuckley, B. A., Jones, J. E., Poudyal, D., Chumanevich, A. P., et al. (2011). Suppression of colitis in mice by Cl-amidine: a novel peptidylarginine deiminase inhibitor. *Am. J. Physiol. Gastrointest. Liver Physiol.* 300, G929–G938.
- Clark, S. R., Ma, A. C., Tavener, S. A., McDonald, B., Goodarzi, Z., Kelly, M. M., et al. (2007). Platelet TLR4 activates neutrophil extracellular traps to ensnare bacteria in septic blood. *Nat. Med.* 13, 463–469.
- Cogen, A. L., Yamasaki, K., Muto, J., Sanchez, K. M., Crotty Alexander, L., Tanios, J., et al. (2010). *Staphylococcus epidermidis* antimicrobial delta-toxin (phenol-soluble modulins-gamma) cooperates with host antimicrobial peptides to kill group A *Streptococcus*. *PLoS ONE* 5, e8557. doi: 10.1371/journal.pone.0008557
- Crotty Alexander, L. E., Maisey, H. C., Timmer, A. M., Rooijackers, S. H., Gallo, R. L., von Kockritz-Blickwede, M., et al. (2010). M1T1 group A streptococcal pili promote epithelial colonization but diminish systemic virulence through neutrophil extracellular entrapment. *J. Mol. Med. (Berl.)* 88, 371–381.
- Cuthbert, G. L., Daujat, S., Snowden, A. W., Erdjument-Bromage, H., Hagiwara, T., Yamada, M., et al. (2004). Histone deimination antagonizes arginine methylation. *Cell* 118, 545–553.
- Ermert, D., Urban, C. F., Laube, B., Goosmann, C., Zychlinsky, A., and Brinkmann, V. (2009a). Mouse neutrophil extracellular traps in microbial infections. *J. Innate Immun.* 1, 181–193.
- Ermert, D., Zychlinsky, A., and Urban, C. (2009b). Fungal and bacterial killing by neutrophils. *Methods Mol. Biol.* 470, 293–312.
- Fuchs, T. A., Abed, U., Goosmann, C., Hurwitz, R., Schulze, I., Wahn, V., et al. (2007). Novel cell death program leads to neutrophil extracellular traps. *J. Cell Biol.* 176, 231–241.
- Fuchs, T. A., Brill, A., Duerschmied, D., Schatzberg, D., Monestier, M., Myers, D. D. Jr., et al. (2010). Extracellular DNA traps promote thrombosis. *Proc. Natl. Acad. Sci. U.S.A.* 107, 15880–15885.
- Gabriel, C., McMaster, W. R., Girard, D., and Descoteaux, A. (2010). *Leishmania donovani* promastigotes evade the antimicrobial activity of neutrophil extracellular traps. *J. Immunol.* 185, 4319–4327.
- Garcia-Romo, G. S., Caielli, S., Vega, B., Connolly, J., Allantaz, F., Xu, Z., et al. (2011). Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci. Transl. Med.* 3, 73ra20.
- Grinberg, N., Elazar, S., Rosenshine, I., and Shpigel, N. Y. (2008). Beta-hydroxybutyrate abrogates formation of bovine neutrophil extracellular traps and bactericidal activity against mammary pathogenic *Escherichia coli*. *Infect. Immun.* 76, 2802–2807.
- Guimaraes-Costa, A. B., Nascimento, M. T., Froment, G. S., Soares, R. P., Morgado, F. N., Conceicao-Silva, F., et al. (2009). *Leishmania amazonensis* promastigotes induce and are killed by neutrophil extracellular traps. *Proc. Natl. Acad. Sci. U.S.A.* 106, 6748–6753.
- Guimaraes-Costa, A. B., Nascimento, M. T., Wardini, A. B., Pinto-da-Silva, L. H., and Saraiva, E. M. (2012). ETosis: a microbicidal mechanism beyond cell death. *J. Parasitol. Res.* 2012, 929743.
- Gupta, A. K., Hasler, P., Holzgreve, W., Gebhardt, S., and Hahn, S. (2005). Induction of neutrophil extracellular DNA lattices by placental microparticles and IL-8 and their presence in preeclampsia. *Hum. Immunol.* 66, 1146–1154.
- Gupta, A. K., Joshi, M. B., Philippova, M., Erne, P., Hasler, P., Hahn, S., et al. (2010). Activated endothelial cells induce neutrophil extracellular traps and are susceptible to NETosis-mediated cell death. *FEBS Lett.* 584, 3193–3197.
- Hakkim, A., Furnrohr, B. G., Amann, K., Laube, B., Abed, U. A., Brinkmann, V., et al. (2010). Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc. Natl. Acad. Sci. U.S.A.* 107, 9813–9818.
- Hemmers, S., Teijaro, J. R., Arandjelovic, S., and Mowen, K. A. (2011). PAD4-mediated neutrophil extracellular trap formation is not required for immunity against influenza infection. *PLoS ONE* 6, e22043. doi: 10.1371/journal.pone.0022043
- Hidaka, Y., Hagiwara, T., and Yamada, M. (2005). Methylation of the guanidino group of arginine residues prevents citrullination by peptidylarginine deiminase IV. *FEBS Lett.* 579, 4088–4092.
- Janeway, C. (2005). *Immunobiology: The Immune System in Health and Disease*, 6th Edn. New York: Garland Science.
- Jones, J. E., Causey, C. P., Knuckley, B., Slack-Noyes, J. L., and Thompson, P. R. (2009). Protein arginine deiminase 4 (PAD4): current understanding and future therapeutic potential. *Curr. Opin. Drug Discov. Dev.* 12, 616–627.
- Jones, J. E., Slack, J. L., Fang, P., Zhang, X., Subramanian, V., Causey, C. P., et al. (2012). Synthesis and screening of a haloacetamide containing library to identify PAD4 selective inhibitors. *ACS Chem. Biol.* 7, 160–165.
- Juneau, R. A., Pang, B., Weimer, K. E., Armbruster, C. E., and Swords, W. E. (2011). Nontypeable *Haemophilus influenzae* initiates formation of neutrophil extracellular traps. *Infect. Immun.* 79, 431–438.
- Kan, R., Jin, M., Subramanian, V., Causey, C. P., Thompson, P. R., and Coonrod, S. A. (2012). Potential role for PADI-mediated histone citrullination in preimplantation development. *BMC Dev. Biol.* 12, 19. doi: 10.1186/1471-213X-12-19
- Kearney, P. L., Bhatia, M., Jones, N. G., Yuan, L., Glascock, M. C., Catchings, K. L., et al. (2005). Kinetic characterization of protein arginine deiminase 4: a transcriptional corepressor implicated in the onset and progression of rheumatoid arthritis. *Biochemistry* 44, 10570–10582.
- Kessenbrock, K., Krumbholz, M., Schönermarck, U., Back, W., Gross, W. L., Werb, Z., et al. (2009). Netting neutrophils in autoimmune small-vessel vasculitis. *Nat. Med.* 15, 623–625.
- Khatua, B., Bhattacharya, K., and Mandal, C. (2012). Sialoglycoproteins adsorbed by *Pseudomonas aeruginosa* facilitate their survival by impeding neutrophil extracellular trap through siglec-9. *J. Leukoc. Biol.* 91, 641–655.
- Kirchner, T., Moller, S., Klinger, M., Solbach, W., Laskay, T., and Behnen, M. (2012). The impact of various reactive oxygen species on the formation of neutrophil extracellular traps. *Mediators Inflamm.* 2012, 849136.
- Klareskog, L., Ronnelid, J., Lundberg, K., Padyukov, L., and Alfredsson, L. (2008). Immunity to citrullinated proteins in rheumatoid arthritis. *Annu. Rev. Immunol.* 26, 651–675.
- Knuckley, B., Bhatia, M., and Thompson, P. R. (2007). Protein arginine deiminase 4: evidence for a reverse protonation mechanism. *Biochemistry* 46, 6578–6587.
- Knuckley, B., Causey, C. P., Jones, J. E., Bhatia, M., Dreyton, C. J., Osborne, T. C., et al. (2010). Substrate specificity and kinetic studies of PADs 1, 3, and 4 identify potent and selective inhibitors of protein arginine deiminase 3. *Biochemistry* 49, 1–28.
- Kohchi, C., Inagawa, H., Nishizawa, T., and Soma, G. (2009). ROS and

- innate immunity. *Anticancer Res.* 29, 817–821.
- Lange, S., Gogel, S., Leung, K. Y., Vernay, B., Nicholas, A. P., Causey, C. P., Thompson, P. R., et al. (2011). Protein deiminases: new players in the developmentally regulated loss of neural regenerative ability. *Dev. Biol.* 355, 205–214.
- Lauth, X., von Kockritz-Blickwede, M., McNamara, C. W., Myskowski, S., Zinkernagel, A. S., Beall, B., et al. (2009). M1 protein allows Group A streptococcal survival in phagocyte extracellular traps through cathelicidin inhibition. *J. Innate Immun.* 1, 202–214.
- Li, P., Li, M., Lindberg, M. R., Kennett, M. J., Xiong, N., and Wang, Y. (2010). PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J. Exp. Med.* 207, 1853–1862.
- Li, P., Yao, H., Zhang, Z., Li, M., Luo, Y., Thompson, P. R., et al. (2008). Regulation of p53 target gene expression by peptidylarginine deiminase 4. *Mol. Cell Biol.* 28, 4745–4758.
- Lin, A. M., Rubin, C. J., Khandpur, R., Wang, J. Y., Riblett, M., Yalavarthi, S., et al. (2011). Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. *J. Immunol.* 187, 490–500.
- Liu, Y. L., Chiang, Y. H., Liu, G. Y., and Hung, H. C. (2011). Functional role of dimerization of human peptidylarginine deiminase 4 (PAD4). *PLoS ONE* 6, e21314. doi: 10.1371/journal.pone.0021314
- Logters, T., Margraf, S., Altrichter, J., Cinatl, J., Mitzner, S., Windolf, J., et al. (2009). The clinical value of neutrophil extracellular traps. *Med. Microbiol. Immunol.* 198, 211–219.
- Luo, Y., Arita, K., Bhatia, M., Knuckley, B., Lee, Y. H., Stallcup, M. R., et al. (2006a) Inhibitors and inactivators of protein arginine deiminase 4: functional and structural characterization. *Biochemistry* 45, 11727–11736.
- Luo, Y., Knuckley, B., Bhatia, M., Pellechia, P. J., and Thompson, P. R. (2006b) Activity-based protein profiling reagents for protein arginine deiminase 4 (PAD4): synthesis and *in vitro* evaluation of a fluorescently labeled probe. *J. Am. Chem. Soc.* 128, 14468–14469.
- Mantovani, A., Cassatella, M. A., Costantini, C., and Jaillon, S. (2011). Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat. Rev. Immunol.* 11, 519–531.
- Marcos, V., Zhou, Z., Yildirim, A. O., Bohla, A., Hector, A., Vitkov, L., et al. (2011). Retraction: CXCR2 mediates NADPH oxidase-independent neutrophil extracellular trap formation in cystic fibrosis airway inflammation. *Nat. Med.* 17, 899.
- McCormick, A., Heesemann, L., Wagener, J., Marcos, V., Hartl, D., Loeffler, J., et al. (2010). NETs formed by human neutrophils inhibit growth of the pathogenic mold *Aspergillus fumigatus*. *Microbes Infect.* 12, 928–936.
- McElwee, J. L., Mohanan, S., Griffith, O. L., Breuer, H. C., Anguish, L. J., Cherrington, B. D., et al. (2012). Identification of PADI2 as a potential breast cancer biomarker and therapeutic target. *BMC Cancer* 12, 500. doi:10.1186/1471-2407-12-500 [Epub ahead of print].
- Mechin, M. C., Coudane, F., Adoue, V., Arnaud, J., Duplan, H., Charveron, M., et al. (2010). Deimination is regulated at multiple levels including auto-deimination of peptidylarginine deiminases. *Cell. Mol. Life Sci.* 67, 1491–1503.
- Menegazzi, R., Declava, E., and Dri, P. (2012). Killing by neutrophil extracellular traps: fact or folklore? *Blood* 119, 1214–1216.
- Metzler, K. D., Fuchs, T. A., Nauseef, W. M., Reumaux, D., Roesler, J., Schulze, I., et al. (2011). Myeloperoxidase is required for neutrophil extracellular trap formation: implications for innate immunity. *Blood* 117, 953–959.
- Mori, Y., Yamaguchi, M., Terao, Y., Hamada, S., Ooshima, T., and Kawabata, S. (2012). alpha-Enolase of *Streptococcus pneumoniae* induces formation of neutrophil extracellular traps. *J. Biol. Chem.* 287, 10472–10481.
- Moscarello, M. A., Mastronardi, F. G., and Wood, D. D. (2007). The role of citrullinated proteins suggests a novel mechanism in the pathogenesis of multiple sclerosis. *Neurochem. Res.* 32, 251–256.
- Munafa, D. B., Johnson, J. L., Brzezinska, A. A., Ellis, B. A., Wood, M. R., and Catz, S. D. (2009). DNase I inhibits a late phase of reactive oxygen species production in neutrophils. *J. Innate Immun.* 1, 527–542.
- Nakashima, K., Hagiwara, T., and Yamada, M. (2002). Nuclear localization of peptidylarginine deiminase V and histone deimination in granulocytes. *J. Biol. Chem.* 277, 49562–49568.
- Neeli, I., Dwivedi, N., Khan, S., and Radic, M. (2009). Regulation of extracellular chromatin release from neutrophils. *J. Innate Immun.* 1, 194–201.
- Neeli, I., Khan, S. N., and Radic, M. (2008). Histone deimination as a response to inflammatory stimuli in neutrophils. *J. Immunol.* 180, 1895–1902.
- Nishinaka, Y., Arai, T., Adachi, S., Takaori-Kondo, A., and Yamashita, K. (2011). Singlet oxygen is essential for neutrophil extracellular trap formation. *Biochem. Biophys. Res. Commun.* 413, 75–79.
- Oehmcke, S., Morgelin, M., and Herwald, H. (2009). Activation of the human contact system on neutrophil extracellular traps. *J. Innate Immun.* 1, 225–230.
- Palmer, L. J., Cooper, P. R., Ling, M. R., Wright, H. J., Huissoon, A., and Chapple, I. L. (2012). Hypochlorous acid regulates neutrophil extracellular trap release in humans. *Clin. Exp. Immunol.* 167, 261–268.
- Papayannopoulos, V., and Zychlinsky, A. (2009). NETs: a new strategy for using old weapons. *Trends Immunol.* 30, 513–521.
- Papayannopoulos, V., Metzler, K. D., Hakkim, A., and Zychlinsky, A. (2010). Neutrophil elastase and myeloperoxidase regulate the formation of neutrophil extracellular traps. *J. Cell Biol.* 191, 677–691.
- Parker, H., Albrett, A. M., Kettle, A. J., and Winterbourn, C. C. (2012). Myeloperoxidase associated with neutrophil extracellular traps is active and mediates bacterial killing in the presence of hydrogen peroxide. *J. Leukoc. Biol.* 91, 369–376.
- Parseghian, M. H., and Luhrs, K. A. (2006). Beyond the walls of the nucleus: the role of histones in cellular signaling and innate immunity. *Biochem. Cell Biol.* 84, 589–604.
- Patel, S., Kumar, S., Jyoti, A., Srinag, B. S., Keshari, R. S., Saluja, R., et al. (2010). Nitric oxide donors release extracellular traps from human neutrophils by augmenting free radical generation. *Nitric Oxide* 22, 226–234.
- Pham, C. T. (2006). Neutrophil serine proteases: specific regulators of inflammation. *Nat. Rev. Immunol.* 6, 541–550.
- Pilszczek, F. H., Salina, D., Poon, K. K., Fahey, C., Yipp, B. G., Sibley, C. D., et al. (2010). A novel mechanism of rapid nuclear neutrophil extracellular trap formation in response to *Staphylococcus aureus*. *J. Immunol.* 185, 7413–7425.
- Raijmakers, R., Zendman, A. J., Egberts, W. V., Vossenaar, E. R., Raats, J., Soede-Huijbregts, C., et al. (2007). Methylation of arginine residues interferes with citrullination by peptidylarginine deiminases *in vitro*. *J. Mol. Biol.* 367, 1118–1129.
- Ramos-Kichik, V., Mondragon-Flores, R., Mondragon-Castelan, M., Gonzalez-Pozos, S., Muniz-Hernandez, S., Rojas-Espinosa, O., et al. (2009). Neutrophil extracellular traps are induced by *Mycobacterium tuberculosis*. *Tuberculosis (Edinb.)* 89, 29–37.
- Remijsen, Q., Kuijpers, T. W., Wirawan, E., Lippens, S., Vandenabeele, P., and Vanden Berghe, T. (2011a). Dying for a cause: NETosis, mechanisms behind an antimicrobial cell death modality. *Cell Death Differ.* 18, 581–588.
- Remijsen, Q., Vanden Berghe, T., Wirawan, E., Asselbergh, B., Parthoens, E., De Rycke, R., et al. (2011b). Neutrophil extracellular trap cell death requires both autophagy and superoxide generation. *Cell Res.* 21, 290–304.
- Rohrbach, A. S., Hemmers, S., Arandjelovic, S., Corr, M., and Mowen, K. A. (2012). PAD4 is not essential for disease in the K/BxN murine autoantibody-mediated model of arthritis. *Arthritis Res. Ther.* 14, R104.
- Shirai, H., Blundell, T. L., and Mizuguchi, K. (2001). A novel superfamily of enzymes that catalyze the modification of guanidino groups. *Trends Biochem. Sci.* 26, 465–468.
- Slack, J. L., Causey, C. P., Luo, Y., and Thompson, P. R. (2011a). Development and use of clickable activity based protein profiling agents for protein arginine deiminase 4. *ACS Chem. Biol.* 6, 466–476.
- Slack, J. L., Jones, L. E. Jr., Bhatia, M. M., and Thompson, P. R. (2011b). Autodeimination of protein arginine deiminase 4 alters protein-protein interactions but not activity. *Biochemistry* 50, 3997–4010.
- Slack, J. L., Causey, C. P., and Thompson, P. R. (2011c). Protein arginine deiminase 4: a target for an epigenetic cancer therapy. *Cell. Mol. Life Sci.* 68, 709–720.
- Springer, D. J., Ren, P., Raina, R., Dong, Y., Behr, M. J., McEwen, B. F., et al. (2010). Extracellular fibrils of pathogenic yeast *Cryptococcus gattii* are important for ecological niche, murine virulence and human neutrophil interactions. *PLoS ONE* 5, e10978. doi: 10.1371/journal.pone.0010978
- Suzuki, A., Yamada, R., Chang, X., Tokuhira, S., Sawada, T., Suzuki, M., et al. (2003). Functional haplotypes of PADI4, encoding citrullinating enzyme peptidylarginine deiminase 4, are associated with rheumatoid arthritis. *Nat. Genet.* 34, 395–402.
- Svobodova, E., Staib, P., Losse, J., Hennis, F., Barz, D., and Jozsi, M. (2012). Differential interaction of

- the two related fungal species *Candida albicans* and *Candida dubliniensis* with human neutrophils. *J. Immunol.* 189, 2502–2511.
- Thompson, P. R., and Fast, W. (2006). Histone citrullination by protein arginine deiminase: is arginine methylation a green light or a roadblock? *ACS Chem. Biol.* 1, 433–441.
- Urban, C. F., Ermert, D., Schmid, M., Abu-Abed, U., Goosmann, C., Nacken, W., et al. (2009). Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against *Candida albicans*. *PLoS Pathog.* 5, e1000639. doi: 10.1371/journal.ppat.1000639
- Urban, C. F., Reichard, U., Brinkmann, V., and Zychlinsky, A. (2006). Neutrophil extracellular traps capture and kill *Candida albicans* yeast and hyphal forms. *Cell. Microbiol.* 8, 668–676.
- Uy, R., and Wold, F. (1977). Post-translational covalent modification of proteins. *Science* 198, 890–896.
- Villanueva, E., Yalavarthi, S., Berthier, C. C., Hodgins, J. B., Khandpur, R., Lin, A. M., et al. (2011). Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J. Immunol.* 187, 538–552.
- Vossenaar, E. R., Radstake, T. R., van der Heijden, A., van Mansum, M. A., Dieteren, C., de Rooij, D. J., et al. (2004). Expression and activity of citrullinating peptidylarginine deiminase enzymes in monocytes and macrophages. *Ann. Rheum. Dis.* 63, 373–381.
- Vossenaar, E. R., Zendman, A. J., van Venrooij, W. J., and Pruijn, G. J. (2003). PAD, a growing family of citrullinating enzymes: genes, features and involvement in disease. *Bioessays* 25, 1106–1118.
- Wang, Y., Li, M., Stadler, S., Correll, S., Li, P., Wang, D., et al. (2009). Histone hypercitrullination mediates chromatin decondensation and neutrophil extracellular trap formation. *J. Cell Biol.* 184, 205–213.
- Wang, Y., Li, P., Wang, S., Hu, J., Chen, X. A., Wu, J., et al. (2012). Anticancer peptidylarginine deiminase (PAD) inhibitors regulate the autophagy flux and the mammalian target of rapamycin complex 1 activity. *J. Biol. Chem.* 287, 25941–25953.
- Wang, Y., Wysocka, J., Sayegh, J., Lee, Y. H., Perlin, J. R., Leonelli, L., et al. (2004). Human PAD4 regulates histone arginine methylation levels via demethylation. *Science* 306, 279–283.
- Willemze, A., Trouw, L. A., Toes, R. E., and Huizinga, T. W. (2012). The influence of ACPA status and characteristics on the course of RA. *Nat. Rev. Rheumatol.* 8, 144–152.
- Willis, V. C., Gizinski, A. M., Banda, N. K., Causey, C. P., Knuckley, B., Cordova, K. N., et al. (2011). N-[alpha]-benzoyl-N5-(2-chloro-1-iminoethyl)-L-ornithine amide, a protein arginine deiminase inhibitor, reduces the severity of murine collagen-induced arthritis. *J. Immunol.* 186, 4396–4404.
- Wood, D. D., Ackerley, C. A., Brand, B., Zhang, L., Raijmakers, R., Mastronardi, F. G., et al. (2008). Myelin localization of peptidylarginine deiminases 2 and 4: comparison of PAD2 and PAD4 activities. *Lab. Invest.* 88, 354–364.
- Wood, D. D., Bilbao, J. M., O'Connors, P., and Moscarello, M. A. (1996). Acute multiple sclerosis (Marburg type) is associated with developmentally immature myelin basic protein. *Ann. Neurol.* 40, 18–24.
- Yan, J., Meng, X., Wancket, L. M., Lintner, K., Nelin, L. D., Chen, B., et al. (2012). Glutathione reductase facilitates host defense by sustaining phagocytic oxidative burst and promoting the development of neutrophil extracellular traps. *J. Immunol.* 188, 2316–2327.
- Yao, H., Li, P., Venters, B. J., Zheng, S., Thompson, P. R., Pugh, B. F., et al. (2008). Histone Arg modifications and p53 regulate the expression of OKL38, a mediator of apoptosis. *J. Biol. Chem.* 283, 20060–20068.
- Yost, C. C., Cody, M. J., Harris, E. S., Thornton, N. L., McInturff, A. M., Martinez, M. L., et al. (2009). Impaired neutrophil extracellular trap (NET) formation: a novel innate immune deficiency of human neonates. *Blood* 113, 6419–6427.
- Zhang, X., Bolt, M., Guertin, M. J., Chen, W., Zhang, S., Cherrington, B. D., et al. (2012). Peptidylarginine deiminase 2-catalyzed histone H3 arginine 26 citrullination facilitates estrogen receptor alpha target gene activation. *Proc. Natl. Acad. Sci. U.S.A.* 109, 13331–13336.
- Zhang, X., Gamble, M. J., Stadler, S., Cherrington, B. D., Causey, C. P., Thompson, P. R., et al. (2011). Genome-wide analysis reveals PAD4 cooperates with Elk-1 to activate c-Fos expression in breast cancer cells. *PLoS Genet.* 7, e1002112. doi: 10.1371/journal.pgen.1002112

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 17 September 2012; paper pending published: 04 October 2012; accepted: 11 November 2012; published online: 29 November 2012.

Citation: Rohrbach AS, Slade DJ, Thompson PR and Mowen KA (2012) Activation of PAD4 in NET formation. *Front. Immun.* 3:360. doi: 10.3389/fimmu.2012.00360

This article was submitted to *Frontiers in Molecular Innate Immunity*, a specialty of *Frontiers in Immunology*.

Copyright © 2012 Rohrbach, Slade, Thompson and Mowen. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.