



Longevity in Cave Animals

Enrico Lunghi and Helena Bilandžija*

Division of Molecular Biology, Institut Ruder Bošković, Zagreb, Croatia

An extraordinary longevity has been observed in some cave species, and this raised the hypothesis that a longer lifespan may be considered one of the characteristic traits of these animals. However, only a few cave species have been studied thus far, and a firm conclusion remains to be drawn. Here we review the available knowledge on the longevity of subterranean species, point out the limitations of previous studies, and provide suggestions for future studies to answer important questions regarding the longevity in cave animals, its adaptive value and the related promoting factors. We also argue that studying the longevity in cave animals will contribute to the field of aging, especially to understanding the evolution of this phenomenon.

Keywords: adaptation, biospeleology, cave biology, lifespan, senescence, subterranean, troglobite, convergent evolution

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*Correspondence:

Helena Bilandžija
helena.bilandzija@irb.hr

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INTRODUCTION

With their unique ecological setting, subterranean environments are theaters of evolutionary processes that lead to fascinating adaptive traits in the species able to colonize them (Mammola, 2019). All subsurface habitats are characterized by a common feature, the absence of light (Culver and Pipan, 2014, 2019). Light can only penetrate and affect the area surrounding their connections to the surface (Lunghi et al., 2015; Culver and Pipan, 2019), limiting the effects of seasonality and the circadian cycle in deeper zones, a condition that contributes to highly stable microclimate characterized by high humidity and constant temperatures (Biswas, 2009; Lunghi et al., 2015). Without sunlight, photosynthetic processes are absent (Culver and Pipan, 2019), what leads to a strong reduction of the available organic matter, and therefore the subterranean food web is largely dependent on inputs of allochthonous organic material from the surface (Schneider et al., 2011; Barzaghi et al., 2017). This scarcity of food resources results in reduced abundance of species at all levels of the food web (Venarsky et al., 2014; Culver and Pipan, 2019; Manenti et al., 2020).

Colonization of subterranean environments by aboveground species is accompanied by numerous changes in their phenotypic traits (Hervant et al., 2001; Bilandžija et al., 2020; Lunghi and Zhao, 2020). The hallmarks of adaptation to subterranean environments are loss of eyes and pigmentation, which are so common in subterranean taxa that they are used as diagnostic traits to assess the degree of adaptation to these environments (Howarth and Moldovan, 2018; but see Lunghi et al., 2014). Some of the other conspicuous adaptive traits in subterranean species include elongation of body appendages, increase in fat stores,

overexpression of mechanosensory and chemosensory systems (Jeffery, 2009; Bilandžija et al., 2012; Gonzalez et al., 2017). Although most obvious, morphological adaptations often represent the final chapter in a story that began with the behavioral and physiological changes that accompanied the early stages of cave colonization (Culver et al., 1995; Pigliucci et al., 2006; Bilandžija et al., 2020).

In this review we explore a poorly known aspect of subterranean species: their longevity. Among subterranean taxa, individuals with exceptional longevity have occasionally been reported (e.g., Voituron et al., 2011; Puljas et al., 2014), raising the idea that the dark passages within the earth harbor animals with extended lifespans. Over the years, most studies have aimed to estimate the age and lifespan of cave species (e.g., Trajano, 1991; Voituron et al., 2011; Puljas et al., 2014), but very few in a comparative framework that included a surface relative (Poulson, 1963). Ultimately, it has yet to be proven whether higher longevity is indeed a characterizing trait of subterranean animals, and eventually whether such trait is adaptive. Here, we review what is known about the longevity of subterranean species and provide recommendations for future studies to experimentally test whether these animals show higher longevity compared to their surface relatives and what the potential promoting factors might be.

LONGEVITY IN SUBTERRANEAN SPECIES

The limited information on the lifespan of cave species mostly comes from the few species that have been successfully reared in laboratory facilities (Poulson, 1963; Miaud and Guillaume, 2005; Hinaux et al., 2011; Voituron et al., 2011; Bichuette and Trajano, 2021). Animals kept under controlled conditions enable tracking their growth and development of specific growth rate curves can provide age estimates for wild populations (Brunkow and Collins, 1996; Gallo and Jeffery, 2012). However, controlled conditions may differ from those found in natural environments and may alter growth rates of individuals (Trajano, 1997; Simon et al., 2017). Luckily, growth rates can be estimated also in the wild for species that are marked (e.g., Lunghi and Bruni, 2018; Lunghi et al., 2019) and recaptured over time (Taddei Ruggiero, 2001; Venarsky et al., 2012; Balázs et al., 2020; Lunghi et al., 2022), or when individuals show reliable age marks (Simon et al., 2017; Riddle et al., 2018).

One of the most iconic cave species is the olm, *Proteus anguinus*, an aquatic salamander distributed in the Dinaric karst in the South-Eastern Europe (Sket, 1997; Gorički et al., 2017). Because it is extremely difficult to study *Proteus* in the wild (Buzzacott et al., 2009; Balázs et al., 2020), most of the available knowledge comes from captive observations (Juberthie et al., 1996; Ipsen and Knolle, 2017; Aljančič, 2019). This neotenic salamander has a slow development, becoming sexually mature after 14 years and reproducing every 12.5 for at least 50/60 years (Voituron et al., 2011;

Ipsen and Knolle, 2017). The oldest known individual has an age between 48 and 58 years (Voituron et al., 2011). Given its slow growth rate and low reproductive activity, the average lifespan of the olm should be around 70 years, although some individuals can live more than 100 years (Bulog et al., 2000; Voituron et al., 2011). In comparison, the closest aboveground relatives are from the North American genus *Necturus*, and the longevity of *Necturus maculosus* has been estimated at 34 years (Petranka, 1998). Interestingly, the olm's small size, metabolic rate comparable to other salamanders, and lack of increased antioxidant activity are in stark contrast to the predictions of aging theory, which foresee higher lifespan in larger species with low metabolic rate and high antioxidant activity (Blanco and Sherman, 2005; Voituron et al., 2011).

Cavefish are within the best-studied subterranean animals, yet only a few studies on their lifespan exist. For example, the Mexican tetra, *Astyanax mexicanus*, is probably one of the widest used model species in cave biology (Wilkens, 1988; Rétaux and Casane, 2013; Keene et al., 2015; Jeffery, 2020), but reliable information on the potential lifespan of this species is missing. We know from captive breeding that both surface and cave forms of *A. mexicanus* can reach the age of 15 (Riddle et al., 2018). There is a divergence in growth rate and age distribution between several populations of cave and surface forms (Simon et al., 2017). Specifically, the range of estimated ages of fish was greater in cave populations (2–8 years) than in surface populations (2–5 years), and there is variability in age distribution even between cave populations (Simon et al., 2017). In another comparative study, Poulson (1963) used the scale and otolith to age five amblyopsid fishes, two surface species, *Chologaster cornuta* and *Forbesichthys agassizi*, and three subsurface, *Typhlichthys subterraneus*, *Amblyopsis spelaea*, and *A. rosae*. The results showed a higher lifespan in cave fishes, where the age of older individuals ranged from 3 to 7 years, up to three times more than for the surface fishes (Poulson, 1963). A similar age for *A. rosae* (about 4–5 years) has been estimated in another study (Brown and Johnson, 2001). However, more up to date techniques indicate a potential lifespan of 20–30 years for these cavefish (Poulson, 2001). For the Brazilian *Ancistrus cryptophthalmus*, an initial estimate determined a lifespan of 8–10 years (Trajano and Bichuette, 2007), while a subsequent study extended it to more than 15 years and provided evidence that the longest-lived individuals can live up to 20–25 years (Secutti and Trajano, 2009). An estimate of lifespan based on growth rate exists also for some other cavefish from the tropics: *Pimelodella kronei* (10–15 years; Trajano, 1991), *Trichomycterus itacarambiensis* (7 years; Trajano, 1997), *Caecobarbus geertsii* (9–15 years; Heuts, 1952; Proudlove and Romero, 2001), and *Ituglanis passensis* (>10; Secutti and Trajano, 2021). No comparable information on closely related surface species exists for these Brazilian cavefish; however, in comparison with other cyprinids (data from AnAge database; Tacutu et al., 2018) these species are among the longer living ones.

Studies in cave invertebrates also yielded extraordinary estimates of their lifespan. An analysis of the growth line

formation in the shells of the cave bivalve *Congeria kusceri* found that the average lifespan was about 30 years. Importantly, authors also showed that growth lines form annually despite animals residing in stable underground waters (Puljas et al., 2014). The oldest individual in this study, a 14.3 mm long male, had an impressive lifespan of 53 years (Puljas et al., 2014). However, other studies have found much larger individuals, so the lifespan of this and possibly other *Congeria* species is likely much higher (Morton, 1969; Bilandžija et al., 2013). The average lifespan of other dreissenid species ranges between 2 and 5 years (Puljas et al., 2014), implying that cave bivalves have evolved lifespans 10 times longer than their surface relatives. As with *Proteus*, this is contrary to predictions which suggest that larger animals have longer lifespans (Blanco and Sherman, 2005), as *Congeria* is one of the smaller dreissenids. However, its slow growth rate and low investment in reproduction (Morton and Puljas, 2013) are consistent with disposable soma theory of aging (Kirkwood, 1977) which suggests a tradeoff between the investment in the reproduction and somatic maintenance; the latter leading to lifespan increase. Similarly, analysis of growth rate of the brachiopod *Neocrania anomala* from marine caves estimated that its lifespan may exceed 40 years (Taddei Ruggiero, 2001). The lifespan of *N. anomala* is about three times higher than *Waltonia inconspicua* and *Terebratalia transversa*, brachiopods that can reach a maximum age of 15 and 13 years, respectively, (Paine, 1969; Rickwood, 1977). Age estimation of an extinct brachiopod, *Gigantoproductus okensis*, indicates that this species

could have lived up to 20 years (Angiolini et al., 2012), which is half of the lifespan estimated for the cave *N. anomala* (Taddei Ruggiero, 2001).

A capture-mark-recapture study conducted over 5 years provided important information on the lifespan of the crayfish *Orconectes australis* (Venarsky et al., 2012). Using growth rate data, the authors were able to re-evaluate the lifespan for this species, which was incorrectly estimated to be 100 years, and found out that the species can live over 22 years (Venarsky et al., 2012). In addition, these authors compiled a list of crayfish species for which lifespan estimates exist (see Table 2 in Venarsky et al., 2012), including two additional cave species: *O. inermis* (9–10 years), and *Procamburus erythropros* (>16 years). Of the 10 surface cambarids included in the list, one has an estimated lifespan of 13 years, while the others ≤ 7 years. Similar results have been obtained for other cave crustaceans. Based on the distribution of body size, Pacioglu et al. (2020) suggested that the lifespan of *Gammarus balcanicus* might exceed that of epigeic gammarids, which have lifespan estimates of about 1 year. In another study, Magniez (1975) estimated that the cave *Stenasellus virei* can reach an age of 12–15, a lifespan up to 20 times longer than epigeic congeneric species. Furthermore, a 40-year study performed by Carpenter (2021) showed that *Bahalana geracei* has probably the longest known lifespan (ranging from 24.5 to 35 years) among isopods. Capture mark recapture studies showed that the cave beetle *Laemostenus schreibersi* can reach the remarkable age of >6.5 (Rusdea, 1994), almost doubling

TABLE 1 | Summary of the lifespan data for cave species collected in this review.

Species	Average lifespan	Maximum lifespan	References
Vertebrates			
<i>Proteus anguinus</i>	70	>100	Voituron et al., 2011
<i>Speleomantes italicus</i>		25	Lunghi, 2022
<i>Calotriton asper</i>	~12	16–19	Miaud and Guillaume, 2005
<i>Astyanax mexicanus</i>	2–8	15	Simon et al., 2017, Riddle et al., 2018
<i>Typhlichthys subterraneus</i>	1–2	4 (20–30)	Poulson, 1963, Poulson, 2001
<i>Amblyopsis spelaea</i>	2–5	7 (20–30)	Poulson, 1963, 2001
<i>Amblyopsis rosae</i>	2–3	4 (20–30)	Poulson, 1963, 2001
<i>Ancistrus cryptophthalmus</i>	15	20–25	Secutti and Trajano, 2009
<i>Pimelodella kronei</i>	6	10–15	Trajano, 1991
<i>Trichomycterus itacarambiensis</i>	7	7	Trajano, 1997
<i>Ituglanis passensis</i>		>10	Secutti and Trajano, 2021
<i>Caecobarbus geertsii</i>		9–13 (<15)	Heuts, 1952, Proudlove and Romero, 2001
Invertebrates			
<i>Congeria kusceri</i>	30	53	Puljas et al., 2014
<i>Neocrania anomala</i>		>40	Taddei Ruggiero, 2001
<i>Orconectes australis</i>		>22	Venarsky et al., 2012
<i>Orconectes inermis</i>		9–10	Retrieved from Venarsky et al. (2012)
<i>Procamburus erythropros</i>		>16	Retrieved from Venarsky et al. (2012)
<i>Gammarus balcanicus</i>		>1	Pacioglu et al., 2020
<i>Stenasellus virei</i>		12–15	Magniez, 1975
<i>Bahalana geracei</i>		24.5–35	Carpenter, 2021
<i>Laemostenus schreibersi</i>		>6.5	Rusdea, 1994

Data is shown in years and refer to both observed and predicted species average and maximum lifespan.

the maximum lifespan observed for epigeal ground beetles (Lövei and Sunderland, 1996).

All information on cave species lifespan collected in this review are summarized in **Table 1**.

Although bats are not strictly bound to subterranean habitats, as they use caves primarily for sheltering purposes and do not exhibit classic cave-related adaptations (Howarth and Moldovan, 2018; Sadier et al., 2020), it is worth mentioning this animal group in the context of longevity, as they are the longest-lived mammals relative to their mass (Podlutzky et al., 2005; Wilkinson and Adams, 2019). Interestingly, cave use predicts the evolution of longevity in several bat lineages (Wilkinson and South, 2002; Salmon et al., 2009).

CONCLUSION AND FUTURE DIRECTIONS

The information assembled here suggests that longevity can indeed be considered a characteristic of cave animals. However, additional systematic studies encompassing a broader range of animal groups in a comparative framework which includes surface and cave relatives are needed to fortify this conclusion. The main remaining questions are: (1) is longevity adaptive for subterranean lifestyle or did it evolve as a physiological consequence of other adaptive traits, (2) what components of the subterranean environment contribute to the evolution of life extension in resident animals, and (3) what are the molecular and developmental mechanisms that lead to convergent evolution of longevity in cave organisms. Several hypotheses can be made regarding the links of aging with other cave adaptive traits and the characteristics of subterranean environments that may be promoting it. For example, adaptive traits in caves such as slower growth, lower metabolic rate and lower investment in reproduction, have been associated with increased lifespan (Flatt and Schmidt, 2009). Further, many environmental features of caves such as limited food resources (Aspiras et al., 2015), lower extrinsic mortality (lack of predators; Plath and Schlupp, 2008), hypoxia (Boggs and Gross, 2021; van der Weele and Jeffery, 2022) and lack of UV irradiation (Körner et al., 2006), are consistent with known ecological predictors of longevity (Speakman and Selman, 2011; Flament et al., 2013; Omotoso et al., 2021). These nonexclusive hypotheses can be tested using model systems that have closely related species or populations in caves and on the surface, have a reliable method for estimating their age, and can survive in the laboratory to be used for experimental perturbations.

Although important to understand the life history characteristics of subterranean species (e.g., sexual maturity, reproductive potential; Poulson, 1963; Lunghi et al., 2018; Lunghi, 2022), growth rate curves may not be the best method to estimate species ages. First, species generally do not grow indefinitely and their growth rate can only be estimated during a particular phase of their life cycle (Venarsky et al., 2012; Lunghi, 2022). A clear example of this is the aforementioned crayfish *Orconectes australis*, whose lifespan has been reduced by a factor of four using

better-tuned methods (Venarsky et al., 2012). Another limitation to using this method stems from the ecological differences between cave and surface populations. For example, low availability of nutrients in subterranean environments (Huntsman et al., 2011; Culver and Pipan, 2019; Pacheco et al., 2020) strongly influences species growth (Simon et al., 2017). Without rearing cave and surface relatives under the same conditions, it is difficult to distinguish intrinsic from extrinsic effects on lifespan.

The use of molecular biomarkers of aging (e.g., SA- β -Gal, 4-HNE, or lipofuscin staining; Nelson et al., 2012; Dimri et al., 1995) may overcome some of these limitations. In addition to allowing reliable assessment of lifespan differences between subterranean and surface species, the use of markers for relevant physiological and molecular processes will enable following the basic processes underlying aging as well as the assessment of variability in aging on population or species levels (Robins et al., 2017).

Studies on cave animals may also provide important insights into the biology of aging, particularly in the areas of evolution and genetics of aging. One of the greatest challenges in the field of aging is to uncover the genes and processes that cause lifespan differences among species (Partridge and Gems, 2006). Because of the known direction of evolution -from surface to cave- and the ability to compare ancestral form with derived form, cave dwellers provide a rich source of information about mechanisms that lead to slowing rates of aging in nature. Knowledge of these mechanisms could be of great benefit and have implications for human welfare. Because of well-defined environmental characteristics and numerous convergently evolved physiological and life-history adaptations, the study of aging in cave dwellers can shed light on how longevity covaries and co-evolves with other organismal and environmental traits.

Aging is a complex phenotype, so it is important to incorporate research on nontraditional and atypical species and integrate approaches from multiple biological disciplines (Cohen, 2018). How and why senescence evolves in natural populations and how to maintain health in old age is still a major issue in contemporary biology. We propose that studies of aging in cave-adapted animals can provide new and original insights into how nature has solved the problem of extending lifespan.

AUTHOR CONTRIBUTIONS

EL drafted the manuscript. Both authors conceived the study and critically revised the manuscript.

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REFERENCES

- Aljančič, G. (2019). History of research on *Proteus anguinus* Laurenti 1768 in Slovenia. *Folia Biol. Geol.* 60, 39–69. doi: 10.3986/fbg0050
- Angiolini, L., Stephenson, M., Leng, M. J., Jadoul, F., Millward, D., Aldridge, A., et al. (2012). Heterogeneity, cyclicity and diagenesis in a Mississippian brachiopod shell of palaeoequatorial Britain. *Terra Nova* 24, 16–26. doi: 10.1111/j.1365-3121.2011.01032.x
- Aspiras, A. C., Rohner, N., Martineau, B., Borowsky, R. L., and Tabin, C. J. (2015). Melanocortin 4 receptor mutations contribute to the adaptation of cavefish to nutrient-poor conditions. *Proc. Natl. Acad. Sci. U.S.A.* 112, 9668–9673. doi: 10.1073/pnas.1510802112
- Balázs, G., Lewarne, B., and Herczeg, G. (2020). Extreme site fidelity of the olm (*Proteus anguinus*) revealed by a long-term capture–mark–recapture study. *J. Zool.* 311, 99–105. doi: 10.1111/jzo.12760
- Barzaghi, B., Ficetola, G. F., Pennati, R., and Manenti, R. (2017). Biphase predators provide biomass subsidies in small freshwater habitats: a case study of spring and cave pools. *Freshw. Biol.* 62, 1637–1644. doi: 10.1111/fwb.12975
- Bichuette, M. E., and Trajano, E. (2021). Monitoring Brazilian cavefish: Ecology and conservation of four threatened catfish of genus ituglanis (siluriformes: Trichomycteridae) from central Brazil. *Diversity* 13:91. doi: 10.3390/d13020091
- Bilandžija, H., Četković, H., and Jeffery, W. R. (2012). Evolution of albinism in cave planthoppers by a convergent defect in the first step of melanin biosynthesis. *Evol. Dev.* 14, 196–203. doi: 10.1111/j.1525-142X.2012.00535.x
- Bilandžija, H., Morton, B., Podnar, M., and Četković, H. (2013). Evolutionary history of relict *Congerina* (Bivalvia: Dreissenidae): unearthing the subterranean biodiversity of the Dinaric Karst. *Front. Zool.* 10:5. doi: 10.1186/1742-9994-10-5
- Bilandžija, H., Hollifield, B., Steck, M., Meng, G., Ng, M., Koch, A. D., et al. (2020). Phenotypic plasticity as a mechanism of cave colonization and adaptation. *eLife* 9:e51830. doi: 10.7554/eLife.51830
- Biswas, J. (2009). Kotumsar Cave biodiversity: a review of cavernicoles and their troglomorphic traits. *Biodivers. Conserv.* 19, 275–289. doi: 10.1007/s10531-009-9710-7
- Blanco, M. A., and Sherman, P. W. (2005). Maximum longevity of chemically protected and non-protected fishes, reptiles, and amphibians support evolutionary hypotheses of aging. *Mech. Ageing Dev.* 126, 794–803. doi: 10.1016/j.mad.2005.02.006
- Boggs, T., and Gross, J. (2021). Reduced oxygen as an environmental pressure in the evolution of the blind Mexican cavefish. *Diversity* 13:26. doi: 10.3390/d13010026
- Brown, J. Z., and Johnson, J. E. (2001). Population biology and growth of Ozark cavefish in Logan Cave National Wildlife Refuge, Arkansas. *Environ. Biol. Fishes* 62, 161–169. doi: 10.1023/a:1011860821932
- Brunkow, P. E., and Collins, J. P. (1996). Effects of individual variation in size on growth and development of larval salamanders. *Ecology* 77, 1483–1492. doi: 10.2307/2265545
- Bulog, B., Bizjakmal, L., Kos, M., Mihajl, K., Prelovsek, P.-M., and Aljančič, G. (2000). Biology and functional morphology of *Proteus anguinus* (Amphibia. Caudata). *Acta Biol. Slov.* 43, 85–102.
- Buzzacott, P. L., Zeigler, E., Denoble, P., and Vann, R. (2009). American cave diving fatalities 1969–2007. *Int. J. Aquat. Res. Educ.* 3, 162–177.
- Carpenter, J. H. (2021). Forty-year natural history study of *Bahalana geracei* Carpenter, 1981, an anchialine cave-dwelling isopod (Crustacea, Isopoda, Cirolanidae) from San Salvador Island, Bahamas: reproduction, growth, longevity, and population structure. *Subterr. Biol.* 37, 105–156. doi: 10.3897/subtbl.37.60653
- Cohen, A. A. (2018). Aging across the tree of life: The importance of a comparative perspective for the use of animal models in aging. *Biochim. Biophys. Acta Mol. Basis Dis.* 1864, 2680–2689. doi: 10.1016/j.bbdis.2017.05.028
- Culver, D. C., Kane, T. C., and Fong, D. W. (1995). *Adaptation and Natural Selection in Caves. The Evolution of Gammarus Minus*. Cambridge, MA: Harvard University Press.
- Culver, D. C., and Pipan, T. (2014). *Shallow Subterranean Habitats: Ecology, Evolution, and Conservation*. Oxford: Oxford University Press.
- Culver, D. C., and Pipan, T. (2019). *The Biology of Caves and Other Subterranean Habitats*. Oxford: Oxford University Press.
- Dimri, G. P., Leet, X., Basile, G., Acosta, M., Scortt, G., Roskelley, C., et al. (1995). A biomarker that identifies senescent human cells in culture and in aging skin in vivo. *Proc. Natl. Acad. Sci. U.S.A.* 92, 9363–9367.
- Flament, F., Bazin, R., Laquieze, S., Rubert, V., Simonpietri, E., and Piot, B. (2013). Effect of the sun on visible clinical signs of aging in Caucasian skin. *Clin. Cosmet. Investig. Dermatol.* 6, 221–232. doi: 10.2147/CCID.S44686
- Flatt, T., and Schmidt, P. S. (2009). Integrating evolutionary and molecular genetics of aging. *Biochim. Biophys. Acta* 1790, 951–962. doi: 10.1016/j.bbagen.2009.07.010
- Gallo, N. D., and Jeffery, W. R. (2012). Evolution of space dependent growth in the Teleost *Astyanax mexicanus*. *PLoS One* 7:e41443. doi: 10.1371/journal.pone.0041443
- Gonzalez, B. C., Worsaae, K., Fontaneto, D., and Martínez, A. (2017). Anophthalmia and elongation of body appendages in cave scale worms (Annelida: Aphroditiformia). *Zool. Scr.* 47, 106–121. doi: 10.1111/zsc.12258
- Goričić, S., Stanković, D., Snoj, A., Kuntner, M., Jeffery, W. R., Trontelj, P., et al. (2017). Environmental DNA in subterranean biology: range extension and taxonomic implications for *Proteus*. *Sci. Rep.* 7:45054. doi: 10.1038/srep45054
- Hervant, F., Mathieu, J., and Durand, J. P. (2001). Behavioural, physiological and metabolic responses to long-term starvation and refeeding in a blind cave-dwelling (*Proteus anguinus*) and a surface-dwelling (*Euproctus asper*) salamander. *J. Exp. Biol.* 204, 269–281. doi: 10.1242/jeb.204.2.269
- Heuts, M. J. (1952). Ecology, variation and adaptation of the blind African cave fish *Caecobarbus geertsi* Blgr. *Ann. Soc. Roy. Zool. Belg.* 82, 155–230.
- Hinaux, H., Pottin, K., Chalhoub, H., Pèrè, S., Elipot, Y., Legendre, L., et al. (2011). A developmental staging table for *Astyanax mexicanus* surface fish and Pachón cavefish. *Zebrafish* 8, 155–165. doi: 10.1089/zeb.2011.0713
- Howarth, F. G., and Moldovan, O. T. (2018). “The ecological classification of cave animals and their adaptations,” in *Cave Ecology*, eds Moldovan, L. Kováč, and S. Halse (Berlin: Springer), 41–67. doi: 10.1007/978-3-319-98852-8_4
- Huntsman, B. M., Venarsky, M. P., Benstead, J. P., and Huryn, A. D. (2011). Effects of organic matter availability on the life history and production of a top vertebrate predator (Plethodontidae: *Gyrinophilus palleucus*) in two cave streams. *Freshw. Biol.* 56, 1746–1760.
- Ipsen, A., and Knolle, F. (2017). The olm of Hermann’s Cave, Harz Mountains, Germany – eggs laid after more than 80 years. *Nat. Slov.* 19, 51–52.
- Jeffery, W. R. (2009). Regressive evolution in *Astyanax* cavefish. *Annu. Rev. Genet.* 43, 25–47. doi: 10.1146/annurev-genet-102108-134216
- Jeffery, W. R. (2020). *Astyanax* surface and cave fish morphs. *EvoDevo* 11:14. doi: 10.1186/s13227-020-00159-6
- Juberthie, C., Durand, J., and Dupuy, M. (1996). La reproduction des Protées (*Proteus anguinus*): bilan de 35 ans d’élevage dans les grottes-laboratoires de Moulis et d’Aulignac. *Mém. Biospéol.* 23, 53–56.
- Keene, A., Yoshizawa, M., and McGaugh, S. E. (2015). *Biology and Evolution of the Mexican Cavefish*. Cambridge, MA: Academic Press.
- Kirkwood, T. (1977). Evolution of ageing. *Nature* 270, 301–304.
- Körner, K. E., Schlupp, I., Plath, M., and Loew, E. R. (2006). Spectral sensitivity of mollies: comparing surface- and cave-dwelling Atlantic mollies, *Poecilia mexicana*. *J. Fish Biol.* 69, 54–65. doi: 10.1111/j.1095-8649.2006.01056.x
- Lövei, G. L., and Sunderland, K. D. (1996). Ecology and behavior of ground beetles (Coleoptera: Carabidae). *Annu. Rev. Entomol.* 41, 231–256. doi: 10.1146/annurev.en.41.010196.001311
- Lunghi, E. (2022). Doubling the lifespan of European plethodontid salamanders. *Ecology* 103:e03581. doi: 10.1002/ecy.3581
- Lunghi, E., and Bruni, G. (2018). Long-term reliability of Visual Implant Elastomers in the Italian cave salamander (*Hydromantes italicus*). *Salamandra* 54, 283–286.
- Lunghi, E., Corti, C., Biaggini, M., Merilli, S., Manenti, R., Zhao, Y., et al. (2022). Capture-mark-recapture data on the strictly protected *Speleomantes italicus*. *Ecology* doi: 10.1002/ecy.3641 [Epub ahead of print].
- Lunghi, E., Corti, C., Manenti, R., Barzaghi, B., Buschetti, S., Canedoli, C., et al. (2018). Comparative reproductive biology of European cave salamanders (genus *Hydromantes*): nesting selection and multiple annual breeding. *Salamandra* 54, 101–108.

- Lunghi, E., Manenti, R., and Ficetola, G. F. (2014). Do cave features affect underground habitat exploitation by non-troglobite species? *Acta Oecol.* 55, 29–35.
- Lunghi, E., Manenti, R., and Ficetola, G. F. (2015). Seasonal variation in microhabitat of salamanders: environmental variation or shift of habitat selection? *PeerJ* 3:e1122.
- Lunghi, E., Romeo, D., Mulargia, M., Cogoni, R., Manenti, R., Corti, C., et al. (2019). On the stability of the dorsal pattern of European cave salamanders (genus *Hydromantes*). *Herpetozoa* 32, 249–253. doi: 10.3897/herpetozoa.32.e39030
- Lunghi, E., and Zhao, Y. (2020). Do Chinese cavefish show intraspecific variability in morphological traits? *Ecol. Evol.* 10, 7723–7730. doi: 10.1002/ece3.6495
- Magniez, G. (1975). Observations sur la biologie de *Stenasellus virei* (Crustacea Isopoda Asellota) des eaux souterraines. *Int. J. Speleol.* 7, 79–228. doi: 10.5038/1827-806x.7.1.8
- Mammola, S. (2019). Finding answers in the dark: caves as models in ecology fifty years after Poulson and White. *Ecography* 42, 1331–1351. doi: 10.1111/ecog.03905
- Manenti, R., Melotto, A., Guillaume, O., Ficetola, G. F., and Lunghi, E. (2020). Switching from mesopredator to apex predator: how do responses vary in amphibians adapted to cave living? *Behav. Ecol. Sociobiol.* 74:126.
- Miaud, C., and Guillaume, O. (2005). Variation in age, body size and growth among surface and cave-dwelling populations of the Pyrenean newt, *Euproctus asper* (Amphibia; Urodela). *Herpetologica* 61, 241–249. doi: 10.1655/04-29.1
- Morton, B. (1969). Studies on the biology of *Dreissena polymorpha* Pall. (III). Population dynamics. *Proc. Malacol. Soc. Lond.* 38, 471–482.
- Morton, B., and Puljas, S. (2013). Life-history strategy, with ctenidial and pallial larval brooding, of the troglodytic 'living fossil' *Congeria kusceri* (Bivalvia: Dreissenidae) from the subterranean Dinaric Alpine karst of Croatia. *Biol. J. Linn. Soc.* 108, 294–314. doi: 10.1111/j.1095-8312.2012.02020.x
- Nelson, P. T., Alafuzoff, I., Bigio, E. H., Bouras, C., Braak, H., Cairns, N. J., et al. (2012). Correlation of alzheimer disease neuropathologic changes with cognitive status: A review of the literature. *J. Neuropathol. Exp. Neurol.* 71, 362–381. doi: 10.1097/NEN.0b013e31825018f7
- Omotoso, A. O., Reyer, H., Oster, M., Ponsuksili, S., Trakooljul, N., Muráni, E., et al. (2021). Jejunal transcriptomic profiling of two layer strains throughout the entire production period. *Sci. Rep.* 11:20086. doi: 10.1038/s41598-021-99566-5
- Pacheco, G. S. M., Souza Silva, M., Cano, E., and Ferreira, R. L. (2020). The role of microhabitats in structuring cave invertebrate communities in Guatemala. *Int. J. Speleol.* 49, 161–169. doi: 10.5038/1827-806x.49.2.2333
- Pacioglu, O., Strungaru, S.-A., Ianovici, N., Filimon, M. N., Sinitean, A., Iacob, G., et al. (2020). Ecophysiological and life-history adaptations of *Gammarus balcanicus* (Schäferna, 1922) in a sinking-cave stream from Western Carpathians (Romania). *Zoology* 139:125754. doi: 10.1016/j.zool.2020.125754
- Paine, R. T. (1969). Growth and size distribution of the brachiopod *Terebratalia transversa* Sowerby. *Pac. Sci.* 23, 337–343.
- Partridge, L., and Gems, D. (2006). Beyond the evolutionary theory of ageing: from functional genomics to evo-gero. *Trends Ecol. Evol.* 21, 334–340. doi: 10.1016/j.tree.2006.02.008
- Petranka, J. W. (1998). *Salamanders of the United States and Canada*. Washington, DC: Smithsonian press.
- Pigliucci, M., Murren, C. J., and Schlichting, C. D. (2006). Phenotypic plasticity and evolution by genetic assimilation. *J. Exp. Biol.* 209, 2362–2367. doi: 10.1242/jeb.02070
- Plath, M., and Schlupp, I. (2008). Parallel evolution leads to reduced shoaling behavior in two cave dwelling populations of Atlantic mollies (*Poecilia mexicana*, Poeciliidae, Teleostei). *Environ. Biol. Fish.* 82, 289–297. doi: 10.1007/s10641-007-9291-9
- Podlutzky, A., Khritankov, A. M., Ovodov, N. D., and Austad, S. N. (2005). A new field record for bat longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* 60, 1366–1368. doi: 10.1093/gerona/60.11.1366
- Poulson, T. L. (1963). Cave adaptation in Amblyopsid fishes. *Amer. Midl. Nat.* 70, 257–290. doi: 10.2307/2423056
- Poulson, T. L. (2001). Adaptations of cave fishes with some comparisons to deep-sea fishes. *Environ. Biol. Fishes* 62, 345–364. doi: 10.1007/978-94-015-9795-1_28
- Proudlove, G. S., and Romero, A. (2001). Threatened fishes of the world: *Caecobarbus geertsii* Boulenger, 1921 (Cyprinidae). *Environ. Biol. Fishes* 62, 238–238. doi: 10.1023/a:1011803519256
- Puljas, S., Peharda, M., Morton, B., Štambuk Giljanović, N., and Jurić, I. (2014). Growth and longevity of the "living fossil" *Congeria kusceri* (Bivalvia: Dreissenidae) from the subterranean Dinaric karst of Croatia. *Malacologia* 57, 353–364.
- Rétaux, S., and Casane, D. (2013). Evolution of eye development in the darkness of caves: adaptation, drift, or both? *EvoDevo* 4:26. doi: 10.1186/2041-9139-4-26
- Rickwood, A. E. (1977). Age, growth and shape of the intertidal brachiopod *Waltonia inconspicua* Sowerby, from New Zealand. *Am. Zool.* 17, 63–73. doi: 10.1093/icb/17.1.63
- Riddle, M. R., Aspiras, A. C., Gaudenz, K., Peuß, R., Sung, J. Y., Martineau, B., et al. (2018). Insulin resistance in cavefish as an adaptation to a nutrient-limited environment. *Nature* 555, 647–651. doi: 10.1038/nature26136
- Robins, C., McRae, A. F., Powell, J. E., Wiener, H. W., Aslibekyan, S., Kennedy, E. M., et al. (2017). Testing two evolutionary theories of human aging with DNA methylation data. *Genetics* 207, 1547–1560. doi: 10.1534/genetics.117.300217
- Rusdea, E. (1994). "Population dynamics of *Laemostenus schreibersi* (Carabidae) in a cave in Carinthia (Austria)," in *Carabid Beetles: Ecology and Evolution (Series Entomologica 51)*, eds K. Desender et al. (Dordrecht: Springer Science), 219–225.
- Sadier, A., Urban, D. J., Anthwal, N., Howenstine, A. O., Sinha, I., and Sears, K. E. (2020). Making a bat: The developmental basis of bat evolution. *Genet. Mol. Biol.* 43:e20190146. doi: 10.1590/1678-4685-GMB-2019-0146
- Salmon, A. B., Leonard, S., Masamsetti, V., Pierce, A., Podlutzky, A. J., Podlutzkaya, N., et al. (2009). The long lifespan of two bat species is correlated with resistance to protein oxidation and enhanced protein homeostasis. *FASEB J.* 23, 2317–2326. doi: 10.1096/fj.08-122523
- Schneider, K., Christman, M. C., and Fagan, W. F. (2011). The influence of resource subsidies on cave invertebrates: results from an ecosystem-level manipulation experiment. *Ecology* 92, 765–776. doi: 10.1890/10-0157.1
- Secutti, S., and Trajano, E. (2009). Reproductive behavior, development and eye regression in the cave armored catfish, *Ancistrus cryptophthalmus* Reis, 1987 (Siluriformes: Loricariidae), breed in laboratory. *Neotrop. Ichthyol.* 7, 479–490. doi: 10.1590/s1679-62252009000300016
- Secutti, S., and Trajano, E. (2021). Reproduction, development, asymmetry and late eye regression in the Brazilian cave catfish *Ituglanis passensis* (Siluriformes, Trichomycteridae): evidence contributing to the neutral mutation theory. *Subterr. Biol.* 38, 91–112. doi: 10.3897/subtbl.31.60691
- Simon, V., Elleboode, R., Mahé, K., Legendre, L., OrnelasGarcia, P., Espinasa, L., et al. (2017). Comparing growth in surface and cave morphs of the species *Astybanax mexicanus*: insights from scales. *EvoDevo* 8:23. doi: 10.1186/s13227-017-0086-6
- Sket, B. (1997). Distribution of *Proteus* (Amphibia: Urodela: Proteidae) and its possible explanation. *J. Biogeogr.* 24, 263–280. doi: 10.1046/j.1365-2699.1997.00103.x
- Speakman, J. R., and Selman, C. (2011). The free-radical damage theory: Accumulating evidence against a simple link of oxidative stress to ageing and lifespan. *Bioessays* 33, 255–259. doi: 10.1002/bies.201000132
- Tacutu, R., Thornton, D., Johnson, E., Budovsky, A., Barardo, D., Craig, T., et al. (2018). Human ageing genomic resources: new and updated databases. *Nucleic Acids Res.* 46, D1083–D1090. doi: 10.1093/nar/gkx1042
- Taddei Ruggiero, E. (2001). "Brachiopods of the Isca submarine cave: observations during ten years," in *Brachiopods*, eds H. Brunton, L. R. M. Cocks, and S. L. Long (Milton Park: Taylor & Francis), 261–267. doi: 10.1201/9780203210437.pt4
- Trajano, E. (1991). Population ecology of *Pimelodella kroniei*, troglitic catfish from Southeastern Brazil (Siluriformes, Pimelodidae). *Environ. Biol. Fishes* 30, 407–421. doi: 10.1007/bf02027984
- Trajano, E. (1997). Population ecology of *Trichomycterus itacarambiensis*, a cave catfish from eastern Brazil (Siluriformes, Trichomycteridae). *Environ. Biol. Fishes* 50, 357–369. doi: 10.1023/a:1007366119261
- Trajano, E., and Bichuette, M. E. (2007). Population ecology of cave armoured catfish, *Ancistrus cryptophthalmus* Reis 1987, from central Brazil (Siluriformes: Loricariidae). *Ecol. Freshw. Fish* 16, 105–115. doi: 10.1111/j.1600-0633.2006.00196.x

- van der Weele, C. M., and Jeffery, W. R. (2022). Cavefish cope with environmental hypoxia by developing more erythrocytes and overexpression of hypoxia-inducible genes. *eLife* 11:e69109. doi: 10.7554/eLife.69109
- Venarsky, M. P., Huntsman, B. M., Huryn, A. D., Benstead, J. P., and Kuhajda, B. R. (2014). Quantitative food web analysis supports the energy limitation hypothesis in cave stream ecosystems. *Oecologia* 176, 859–869. doi: 10.1007/s00442-014-3042-3
- Venarsky, M. P., Huryn, A. D., and Benstead, J. P. (2012). Re-examining extreme longevity of the cave crayfish *Orconectes australis* using new mark-recapture data: a lesson on the limitations of iterative size-at-age models. *Freshw. Biol.* 57, 1471–1481. doi: 10.1111/j.1365-2427.2012.02812.x
- Voituron, Y., de Fraipont, M., Issartel, J., Guillaume, O., and Clobert, J. (2011). Extreme lifespan of the human fish (*Proteus anguinus*): a challenge for ageing mechanisms. *Biol. Lett.* 7, 105–107. doi: 10.1098/rsbl.2010.0539
- Wilkens, H. (1988). Evolution and genetics of epigeal and cave *Astyanax fasciatus* (Characidae, Pisces) support for the neutral mutation theory. *Evol. Biol.* 23, 271–367. doi: 10.1007/978-1-4613-1043-3_8
- Wilkinson, G. S., and Adams, D. M. (2019). Recurrent evolution of extreme longevity in bats. *Biol. Lett.* 15:20180860. doi: 10.1098/rsbl.2018.0860
- Wilkinson, G. S., and South, J. M. (2002). Life history, ecology and longevity in bats. *Aging Cell* 1, 124–131. doi: 10.1046/j.1474-9728.2002.00020.x
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