Check for updates

OPEN ACCESS

EDITED BY Rosalba Senese, University of Campania Luigi Vanvitelli, Italy

REVIEWED BY

George Grant, University of Aberdeen, United Kingdom Kathiresh kumar Mani, Texas A&M Health Science Center, United States

*CORRESPONDENCE Wei Chen Chenwei8027@163.com

RECEIVED 15 June 2023 ACCEPTED 05 September 2023 PUBLISHED 26 September 2023

CITATION

Xie L, Zhao H and Chen W (2023) Relationship between gut microbiota and thyroid function: a two-sample Mendelian randomization study. *Front. Endocrinol.* 14:1240752. doi: 10.3389/fendo.2023.1240752

COPYRIGHT

© 2023 Xie, Zhao and Chen. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Relationship between gut microbiota and thyroid function: a two-sample Mendelian randomization study

Liangzhuo Xie¹, Huaye Zhao² and Wei Chen^{1,3*}

¹Graduate School, Liaoning University of Traditional Chinese Medicine, Shenyang, China, ²School of Tranditional Chinese, Liaoning University of Traditional Chinese Medicine, Shenyang, China, ³Department of Geriatrics, The Second Affiliated Hospital of Liaoning University of Traditional Chinese Medicine, Shenyang, China

Background: Numerous observational studies have indicated a link between the composition of gut microbiota and thyroid function. Nevertheless, the precise causal relationship between gut microbiota and thyroid function remains uncertain.

Methods: In this two-sample Mendelian randomization study, we utilized summary data from a genome-wide association study of gut microbiota composition in 18,340 participants from 24 cohorts, as well as summary statistics on thyroid hormones and thyroid-stimulating hormone from the ThyroidOmics Consortium and summary statistics on hypothyroidism and hyperthyroidism from the FinnGen R8 release. Five different methods, including inverse variance weighting, MR-Egger, weighted median, weighted mode, and simple mode, were employed to examine the causal relationship between gut microbiota and thyroid function. Reverse Mendelian randomization analysis was conducted for taxa identified as having a causal relationship with thyroid function in the Mendelian randomization analysis. To assess the robustness of the results, sensitivity analyses were conducted employing Cochran's Q test, MR-Egger intercept test, MR-PRESSO global test, and leave-one-out analysis.

Results: Through MR analysis of 211 microbial taxa and 4 phenotypes, we identified a total of 34 gut microbiota taxa that were associated with the outcomes. After using the bonferroni method for multiple testing correction, phylum *Actinobacteria* (id.400) had a protective effect on hypothyroidism (OR=0.883, 95% CI: 0.817-0.955, P=0.002), and class *Deltaproteobacteria* (id.3087) had a protective effect on hyperthyroidism (OR=0.549, 95% CI: 0.374-0.805, P=0.002). According to the results of reverse MR analysis, no significant causal effect of the four phenotypes was found on gut microbiota. No significant horizontal pleiotropy was detected based on MR-Egger intercept test and MR-PRESSO global test.

Conclusion: Through two-sample MR analysis, we identified specific gut microbiota taxa at the genetic level that are predicted to have a causal relationship with thyroid function, which may serve as useful biomarkers for early disease diagnosis.

KEYWORDS

gut microbiota, thyroid function, causal effect, hypothyroidism, hyperthyroidism, Mendelian randomization

1 Introduction

The thyroid gland is the largest endocrine gland in the human body, and its secreted thyroid hormones(TH) play a crucial role in growth, development, and metabolism (1). Thyroid-stimulating hormone (TSH) is secreted by the anterior pituitary gland and regulates thyroid hormone production by acting on the TSH receptor (TSH-R) on the basolateral membrane of thyroid follicular cells (2). In clinical practice, thyroid function is assessed by measuring circulating levels of TSH and free thyroxine (FT4). Elevated TSH levels indicate hypothyroidism, while decreased TSH levels suggest hyperthyroidism (3). The NHANES III study in the United States found that the overall prevalence of hyperthyroidism was 1.3% (0.5% overt hyperthyroidism and 0.7% subclinical hyperthyroidism), while the overall prevalence of hypothyroidism was 4.5% (0.3% overt hypothyroidism and 4.3% subclinical hypothyroidism) (4). Hypothyroidism is associated with cardiovascular dysfunction and is a risk factor for heart failure (5), as well as affecting the nervous, musculoskeletal, and gastrointestinal systems to varying degrees (6). Hyperthyroidism is associated with conditions such as atrial fibrillation, stroke, pulmonary embolism, and hypercoagulability (7). The etiology of hypothyroidism and hyperthyroidism is still not fully understood, but Graves' disease and nodular thyroid disease are considered major causes of hyperthyroidism (8), while immune abnormalities, thyroidectomy, and excessive treatment for hyperthyroidism are considered major causes of hypothyroidism (6).

The concept of the thyroid-gut axis has been proposed and studied in relation to thyroid diseases and gut microbiota (9-11). According to a study, patients with hyperthyroidism exhibited a notable decrease in the abundance of bifidobacteria and lactobacilli in their gut compared to healthy individuals, along with an increase in enterococci (9). Another study analyzed the differences in gut microbiota between patients with primary hypothyroidism and healthy individuals using 16S rRNA sequencing and found differences in taxa such as Veillonella, Paraprevotella, Neisseria, and Rheinheimera (10). Although these studies confirmed the association between the thyroid and gut microbiota, it is difficult to determine whether dysbiosis of the gut microbiota is the cause or result of thyroid dysfunction. Moreover, observational studies have limitations in ruling out confounding factors between thyroid function and gut microbiota, such as lifestyle, environment, and age, making it challenging to establish a causal relationship.

Mendelian randomization (MR) employs genetic variation as instrumental variables to infer causal connections between exposures and outcomes. Since genotypes are randomly assigned from parents to offspring, Compared to traditional observational studies, MR has the distinct advantage of effectively reducing the influence of confounding factors and reverse causality, leading to enhanced credibility of research findings (12). MR has been widely used to explore the causal relationships between thyroid function and various diseases, such as the relationship between thyroid function and atrial fibrillation (13), lipid metabolism (14), and Alzheimer's disease (15). In this study, we conducted a two-sample MR analysis using summary-level statistics from the MiBioGen Consortium, ThyroidOmics Consortium, and FinnGen Consortium genome-wide association studies (GWAS). The primary objective was to investigate the causal relationships between gut microbiota and FT4, TSH, hypothyroidism, and hyperthyroidism.

2 Methods

2.1 Study design

Mendelian randomization (MR) is an approach employed to evaluate causal connections between variables. This analysis necessitates meeting three core assumptions: (i) the instrumental variables (IVs) possess a robust association with the exposure factor, (ii) the IVs remain unaffected by confounding factors, and (iii) the IVs solely impact the outcome through the exposure factor (16). In this study, we used gut microbiota GWAS data as the exposure factor and FT4, TSH, hypothyroidism, and hyperthyroidism GWAS data as the outcomes. Based on inclusion criteria, we selected suitable SNPs as instrumental variables and conducted a twosample MR analysis to examine the causal relationships between gut microbiota and FT4, TSH, hypothyroidism, and hyperthyroidism. It is worth mentioning that this study strictly adhered to the requirements of the STROBE-MR guidelines (17).

2.2 Data sources

The GWAS data on the gut microbiota originates from the comprehensive meta-analysis conducted by the MiBioGen consortium (18). The study included 18,340 individuals from 24

distinct cohorts. The primary objective was to analyze the microbial composition by focusing on variable regions V4, V3-V4, and V1-V2 of the 16S rRNA gene (18). The direct taxonomic binning method was used to classify the microbiota and performed microbial quantitative trait locus (mbQTL) analysis to identify host genetic variation loci associated with the abundance levels of bacterial taxa in the gut. And the study identified genetic variations associated with 9 phyla, 16 classes, 20 orders, 35 families, and 131 genera (18).

Thyroid function-related genetic variant data were obtained from the ThyroidOmics Consortium (3). Among these, TSH data were derived from 54,288 individuals across 22 cohorts, while FT4 data were obtained from 49,269 individuals across 19 cohorts.

Thyroid disease datasets were obtained from the FinnGen consortium R8 release data (19). The data for hypothyroidism included 42,000 cases and 292,316 controls, whereas the data for hyperthyroidism included 1,621 cases and 255,931 controls. Supplementary Table 1 shows the details of the exposure and outcome analyzed in this MR study.

2.3 Selection of instrumental variables

To meet the requirements of Mendelian randomization (MR) studies, we first selected SNPs that were strongly associated with the exposure factor. Since very few instrumental variables (IVs) are obtained when the threshold is set at ($p < 5 \times 10^{-8}$), we chose a threshold of ($p < 1 \times 10^{-5}$) for SNP selection to obtain more IVs and obtain robust results.

Secondly, to account for linkage disequilibrium and avoid biased results, we set the linkage disequilibrium parameter (R^2) for SNPs at 0.001 and the genetic distance at 10,000 kb. This ensured that each IV was independently present. We set the minor allele frequency level at 0.01 and excluded palindromic SNPs and those that were not present in the outcome.

Considering that IVs used as instrumental variables need to be strongly correlated with the exposure, we assessed the correlation between instrumental variables and the exposure using the Fstatistic. We considered an F-statistic greater than 10 as indicating a strong correlation between the two. The formula for calculating the F-statistic is

$$F = \frac{R^2(N - K - 1)}{K(1 - R^2)}$$

,where

$$R^{2} = \frac{2 \times EAF \times (1 - EAF) \times Beta^{2}}{2 \times EAF \times (1 - EAF) \times Beta^{2} + [2 \times EAF \times (1 - EAF) \times N \times SE^{2}]}$$
(20).

2.4 Mendelian randomization analysis

The inverse-variance weighted (IVW) method is a primary approach used in Mendelian randomization (MR) analysis. This method uses a meta-analytic approach to synthesize the Wald estimates associated with each individual SNP, yielding an aggregate estimate of the collective influence of the gut microbiota on thyroid function. One fundamental assumption of Mendelian randomization (MR) is that there is no horizontal pleiotropy between the instrumental variables (genes) and the outcome variable. This means that the effects of these genes on the outcome are solely mediated through the exposure and not through any other pathways. If this assumption holds true, the IVW method can provide consistent and efficient estimates (21). If a causal relationship is identified by the IVW method (p<0.05), four additional methods, namely MR-Egger, weighted median method, simple mode method, and weighted mode method, are employed to complement the IVW results. The MR-Egger method does not enforce the intercept to be zero, allowing estimation of the causal effect even in the presence of invalid instruments (SNPs that can affect the outcome through non-exposure pathways), and the intercept can indicate the degree of horizontal pleiotropy (22). The weighted median method calculates the median of the ratio estimates after ranking the SNP estimates by their weights. This method requires a minimum of three SNPs, and if more than 50% of the SNPs have effects on the outcome that are not mediated through the exposure, the estimate provided by this method may be biased (23). The weighted mode method provides an consistent estimate if more than half of the SNPs were valid IVs (24). The simple mode method is similar to the weighted mode method, and if the causal effect estimates is derived from a majority of invalid SNPs, the estimate provided by this method may be biased (25). We set the significance threshold at p < 0.05. Additionally, we performed Bonferroni correction for multiple testing, and causal relationships were represented by odds ratios (OR) and 95% confidence intervals (CI). When the IVW method yielded significant results and the directions of the other four methods were consistent, we considered a causal relationship between the exposure and the outcome. In the results with causal relationships, unknown taxa were excluded, and sensitivity analyses were subsequently conducted to ensure the stability of the results.

2.5 Heterogeneity and pleiotropy testing

Heterogeneity was assessed using Cochran's Q method, with a significance level of p < 0.05 indicating significant heterogeneity (23, 26). The presence of horizontal pleiotropy was examined using the MR-Egger intercept test (27) and the MR-PRESSO global test (26). Outliers identified by the MR-PRESSO global test were excluded, and a robustness analysis was performed using the leave-one-out method to validate the results. A significance threshold of p < 0.05 was chosen for the analysis. To account for multiple testing, the Bonferroni method was employed. For each specific level, the significance threshold was adjusted to p < 0.05/n, where n represents the number of taxa present at that level. All statistical analyses and data visualizations were conducted using R software version 4.2.3¹.

¹ https://www.r-project.org/



3 Results

3.1 Selection of instrumental variables

According to the selection criteria for instrumental variables (IV), a total of 1764 SNPs were chosen as instrumental variables. The Fstatistics for the IVs are all greater than 10, indicating that the estimated coefficients are unlikely to be affected by weak instrument bias. Details of the selected IVs are shown in Supplementary Table 2.

3.2 FT4

The estimates calculated using the IVW test indicated that the genetically predicted relative abundance of genus *Lactobacillus* (id.1837) (OR=0.919, 95% CI: 0.847-0.996, P=0.040), genus *Lachnospiraceae UCG-001* (id.11321) (OR=0.943, 95%CI:0.891-0.998, P=0.043), genus *Ruminococcus gauvreauii group* (id.11342) (OR= 0.922, 95%CI: 0.860-0.988, P= 0.021) were associated with

decreased FT4 levels, while genus *Subdoligranulum* (id.2070) (OR= 1.123,95%CI: 1.039-1.214, P= 0.003)was associated with increased FT4 levels (Figure 1). The other four methods, MR-Egger, weighted median, simple mode, and weighted mode were used to supplement the results of IVW. For details, refer to the table (Supplementary Table 3). Similarly, the results were parallel to the IVW results (Supplementary Figure 1). The result of Cochrane's Q test showed that except for Lactobacillus (id.1837), which had a result of p< 0.05, indicating heterogeneity among SNPs, no significant heterogeneity was found among the other SNPs (p > 0.05).

3.3 TSH

The estimates calculated using the IVW test indicated that the genetically predicted relative abundance of order *Enterobacteriales* (id.3468) (OR=1.122, 95%CI: 1.032-1.220, P=0.007) family *Enterobacteriaceae* (id.3469) (OR=1.122, 95%CI: 1.032-1.220, P=7.11E-03), genus *Veillonella* (id.2198) (OR=1.078, 95%CI:



1.005-1.155, P=0.034) were associated with increased TSH levels, while family *Lachnospiraceae* (id.1987) (OR=0.941, 95%CI: 0.889-0.997, P=3.84E-02) and genus *Oscillospira* (id.2064) (OR=0.911, 95% CI: 0.848-0.979, P=0.011) were associated with decreased TSH levels (Figure 2). Similarly, the results of the other four methods can be seen in the table (Supplementary Table 4). And the results were parallel to the IVW results (Supplementary Figure 2). The result of Cochrane's Q test showed that no significant heterogeneity was found among the selected SNPs (p > 0.05).

3.4 Hypothyroidism

The estimates calculated using the IVW test indicated that the genetically predicted relative abundance of order Mollicutes RF9 (id.11579) (OR=1.092, 95%CI: 1.015-1.173, P=0.018), genus Desulfovibrio (id.3173) (OR=1.087, 95%CI: 1.009-1.172, P=0.028), genus Intestinimonas (id.2062) (OR=1.096, 95%CI: 1.032-1.164, P=0.003), genus Ruminiclostridium 5 (id.11355) (OR=1.099, 95% CI: 1.005-1.202, P=0.039), genus Ruminococcaceae UCG-005 (id.11363) (OR=1.097, 95%CI: 1.002-1.202, P=0.045) were positively associated with the risk of hypothyroidism, while phylum Actinobacteria (id.400) (OR=0.883, 95%CI: 0.817-0.955, P=0.002), family Alcaligenaceae (id.2875) (OR=0.894, 95%CI: 0.815-0.981, P=0.018), family Defluviitaleaceae (id.1924) (OR=0.926, 95%CI: 0.869-0.987, P=0.019), genus Butyrivibrio (id.1993) (OR=0.956, 95%CI: 0.924-0.989, P=0.009), genus Eggerthella (id.819) (OR=0.941, 95%CI: 0.891-0.994, P=0.030), genus Lachnospiraceae UCG-008 (id.11328) (OR=0.909, 95%CI: 0.841-0.983, P=0.017) were negatively associated with the risk of hypothyroidism (Figure 3). The results of the other four methods can be seen in the table (Supplementary Table 5). And the results were parallel to the IVW results (Supplementary Figure 3). The result of Cochrane's Q test showed that there was heterogeneity among the selected SNPs for Lachnospiraceae UCG-008 (id.11328) and Ruminococcaceae UCG-005 (id.11363), while no significant heterogeneity was found among the other selected SNPs (p > 0.05).

3.5 Hyperthyroidism

The estimates calculated using the IVW test indicated that the genetically predicted relative abundance of order Mollicutes RF9 (id.11579) (OR= 1.497, 95%CI: 1.107-2.025, P=0.009), genus Ruminococcaceae UCG-004 (id.11362) (OR= 1.384, 95%CI: 1.016-1.884, P=0.039), genus Ruminococcaceae NK4A214 group (id.11358) (OR= 1.502, 95%CI: 1.058-2.131, P= 0.023), genus Prevotella7 (id.11182) (OR= 1.259, 95%CI: 1.023-1.549, P=0.030), genus Collinsella (id.815) (OR= 1.858, 95%CI: 1.184-2.916, P=0.007), genus Catenibacterium (id.2153) (OR= 1.437, 95%CI: 1.057-1.954, P=0.021), genus Bilophila (id.3170) (OR= 1.486, 95% CI: 1.043-2.118, P=0.028) were positively associated with the risk of hyperthyroidism, while Phylum Verrucomicrobia (id.3982) (OR=0.697,95%CI: 0.508-0.957, P=0.026), class Deltaproteobacteria (id.3087) (OR=0.549, 95%CI: 0.374-0.805, P=0.002), family Desulfovibrionaceae (id.3169) (OR=0.621, 95% CI: 0.403-0.956, P=0.031), family Bacteroidaceae (id.917) (OR=0.552, 95%CI: 0.345-0.882, P=0.013), genus Parasutterella (id.2892) (OR=0.717, 95%CI: 0.546-0.941, P=0.016), genus Bacteroides (id.918) (OR=0.552, 95%CI: 0.345-0.882, P=0.013) were negatively associated with the risk of hyperthyroidism (Figure 4). And the results were parallel to the IVW results (Supplementary Figure 4). The results of the other four methods can be seen in the table (Supplementary Table 6). The result of Cochrane's Q test showed that no significant heterogeneity was found among the selected SNPs (p > 0.05).

3.6 Sensitivity analysis

The intercept tests of MR-Egger and the MR-PRESSO global test did not indicate the presence of horizontal pleiotropy (Tables 1, 2, 3, 4). Additionally, the leave-one-out analysis did not reveal any SNP that would significantly alter the overall results, ensuring the stability of the outcomes (Supplementary Figures 5-8). And the results of Cochrane's Q test can be found in Tables 1–4. The results

					01(35/601)	r-value
Phylum						
Actinobacteria	400	14	► -		0.883(0.817, 0.955)	0.002
Order						
Bifidobacteriales	432	11	⊢ – – –		0.930(0.851, 1.017)	0.112
MollicutesRF9	11579	12		F	1.092(1.015, 1.173)	0.018
Family						
Alcaligenaceae	2875	11	⊢ – – – •– – – – I		0.894(0.815, 0.981)	0.018
Defluviitaleaceae	1924	11	⊢∎		0.926(0.869, 0.987)	0.019
Genus						
Butyrivibrio	1993	15	⊢ -■ - 1		0.956(0.924, 0.989)	0.009
Desulfovibrio	3173	10		⊢−−	1.087(1.009, 1.172)	0.028
Eggerthella	819	9	⊢		0.941(0.891, 0.994)	0.030
Intestinimonas	2062	16		⊢+	1.096(1.032, 1.164)	0.003
LachnospiraceaeUCG008	11328	10	⊢−−∎−−⊣		0.909(0.841, 0.983)	0.017
Ruminiclostridium5	11355	i 11		⊢ – – – • – – – –	1.099(1.005, 1.202)	0.039
RuminococcaceaeUCG005	5 11363	14			- 1.097(1.002, 1.202)	0.045
	Actinobacteria Order Bifidobacteriales MollicutesRF9 Family Alcaligenaceae Defluviitaleaceae Genus Butyrivibrio Desulfovibrio Eggerthella Intestinimonas LachnospiraceaeUCG008 Ruminiclostridium5 RuminococcaceaeUCG005	Actinobacteria400OrderBifidobacteriales432Bifidobacteriales432MollicutesRF911579FamilyAlcaligenaceae2875Defluviitaleaceae1924GenusButyrivibrio1993Desulfovibrio3173Eggerthella819Intestinimonas2062LachnospiraceaeUCG00811326Ruminiclostridium511355RuminococcaceaeUCG00511363	Actinobacteria40014Order11Bifidobacteriales43211MollicutesRF91157912Family11Alcaligenaceae287511Defluviitaleaceae192411Genus199315Butyrivibrio199315Desulfovibrio317310Eggerthella8199Intestinimonas206216LachnospiraceaeUCG0081132810Ruminiclostridium51135511RuminococcaceaeUCG0051136314	Actinobacteria 400 14 ► Order Bifidobacteriales 432 11 ► Bifidobacteriales 432 11 ► MollicutesRF9 11579 12 Family Alcaligenaceae 2875 11 ►	Actinobacteria 400 14	Actinobacteria 400 14

FIGURE 3

Forest plot of GM taxa associated with hypothyroidism identified by IVW method.

Exposure	ID	No. of SNF)		OR(95%CI)	P-value
Phylum						
Verrucomicrobia	3982	12	⊢		0.697(0.508, 0.957)	0.026
Class						
Deltaproteobacteria	3087	12	⊢		0.549(0.374, 0.805)	0.002
Order						
MollicutesRF9	11579) 12		⊢− ►−−1	1.497(1.107, 2.025)	0.009
Family						
Desulfovibrionaceae	3169	9	⊢ • − I		0.621(0.403, 0.956)	0.031
Bacteroidaceae	917	8	⊢ - - I		0.552(0.345, 0.882)	0.013
Genus						
RuminococcaceaeUCG004	11362	2 11			1.384(1.016, 1.884)	0.039
Ruminococcaceae (NK4A214 group)	11358	3 13		⊨ – - – – – I	1.502(1.058, 2.131)	0.023
Prevotella7	11182	2 11		⊢ • − 1	1.259(1.023, 1.549)	0.030
Parasutterella	2892	14	H == 4		0.717(0.546, 0.941)	0.016
Collinsella	815	9			1.858(1.184, 2.916)	0.007
Catenibacterium	2153	4		⊨ – ⊨ – – I	1.437(1.057, 1.954)	0.021
Bilophila	3170	13		⊢ – - - – – – I	1.486(1.043, 2.118)	0.028
Bacteroides	918	8	⊢ - - i		0.552(0.345, 0.882)	0.013
			0.5 0.7 0.9	111.1 1.3 1.5 1.7 1.922.1 2.3 2.5 2.7 2.9 Odds Ratio(95%CI)	3	

of the Bonferroni correction test showed that phylum *Actinobacteria* (id.400) (OR=0.883, 95% CI: 0.817-0.955, P=0.002) exhibited a protective effect against hypothyroidism, while class *Deltaproteobacteria* (id.3087) (OR=0.549, 95% CI: 0.374-0.805, P=0.002) exhibited a protective effect against hyperthyroidism. Furthermore, the funnel plot for all the results can be seen in Supplementary Figures 9-12. The analysis of reverse Mendelian randomization showed no significant causal relationship. However, there were two nominally significant causal relationships that hyperthyroidism was associated with an increased risk of order *MollicutesRF9* (id.11579) (OR=1.067, 95% CI: 1.0003-1.138, P=0.0488), while hypothyroidism was associated with an increased risk of genus *Ruminococcaceae UCG005* (OR=1.038, 95% CI: 1.001-1.078, P=0.0497).

4 Discussion

This study comprehensively assessed the causal effects of 211 taxonomic groups within the domain of Genus and Species (from phylum to genus levels) on thyroid function, encompassing parameters such as FT4, TSH, hypothyroidism, and hyperthyroidism. Ultimately, a total of 34 causal relationships were identified, comprising 32 nominal causal associations and 2

strong causal connections. These findings underscore the significance of the gut microbiota in influencing thyroid functionality.

An increasing body of evidence suggests that the gut microbiota plays a pivotal role in the pathogenesis of thyroid disorders. Primarily, the gut microbiota may modulate thyroid function by influencing the uptake of thyroid-relevant micronutrients (28). The synthesis of thyroid hormones necessitates the presence of iodine, and inadequate iodine intake can impair thyroid function (29). Iodine is required to be absorbed through the gastrointestinal tract and transferred to the thyroid. Hence, the gut microbiota exerts a significant influence on iodine metabolism regulation. Within the human gastrointestinal tract, iodine uptake is predominantly facilitated by the sodium-iodide symporter (NIS) (30). Lipopolysaccharides (LPS) and short-chain fatty acids (SCFAs) released by the gut microbiota can modulate iodine uptake by affecting the expression and activity of NIS (31, 32). Additionally, lipopolysaccharides (LPS) can stimulate the synthesis of type 2 iodothyronine deiodinase (D2) in the rat hypothalamic paraventricular nucleus, thus promoting the conversion of T4 to T3 and inhibiting the secretion of pituitary TSH (33). Besides iodine, iron is also an essential trace element for thyroid hormone synthesis, and iron deficiency is a common symptom of hypothyroidism (34). Moreover, the gut microbiota may lower intestinal pH and enhance

Exposure	MR-Eg	ger intercep	ot test	MR-PRESSC	Cochran's Q	
	Egger_ intercept	SE	p-value	RSS obs	<i>p</i> -value	<i>p</i> -value
Genus						
Lactobacillus(ID:1837)	-5.56E-03	1.54E-02	0.730	21.053	0.050	0.027
Subdoligranulum(ID:2070)	1.32E-03	7.27E-03	0.860	9.163	0.703	0.673
LachnospiraceaeUCG001(ID:11321)	-4.84E-03	1.23E-02	0.702	12.957	0.466	0.454
Ruminococcusgauvreauiigroup(ID:11342)	-3.84E-03	1.22E-02	0.760	6.231	0.879	0.883

SE, standard error; RSS, residual sum of squares; Cochran's Q-derived.

Exposure	MR-Eg	iger intercept	t test	MR-PRESSO	Cochran's Q				
	Egger_ intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value	<i>p</i> -value			
Order									
Enterobacteriales(ID:3468)	-1.84E-02	1.89E-02	0.375	4.236	0.817	0.787			
Family									
Enterobacteriaceae(ID:3469)	-1.84E-02	1.89E-02	0.375	4.236	0.825	0.787			
Lachnospiraceae(ID:1987)	-9.33E-04	4.77E-03	0.848	9.527	0.931	0.930			
Genus									
Oscillospira(ID:2064)	3.06E-03	1.38E-02	0.833	4.115	0.829	0.792			
Veillonella(ID:2198)	-2.60E-02	2.26E-02	0.302	6.152	0.637	0.596			

TABLE 2 Horizontal pleiotropy analysis for IVs of 5 GM taxa associated with TSH.

SE, standard error; RSS, residual sum of squares; Cochran's Q-derived.

the bioavailability of colonic iron through the production of shortchain fatty acids (SCFAs), thereby influencing thyroid function (35). The thyroid, as the organ with the highest selenium content per unit tissue, contains selenium in the form of selenoproteins. Enzymes such as glutathione peroxidase, deiodinases (D1, D2, and D3), and thioredoxin reductase, which are selenium-dependent, maintain the stability and activity of thyroid hormones within the body (36). The gut microbiota can influence selenium metabolism and absorption in the colon (37). Furthermore, the gut microbiota can impact the secretion of the neurotransmitter dopamine in the brain and regulate the hypothalamic-pituitary-adrenal (HPA) axis (38). Dopamine, in turn, can inhibit the secretion of TSH within the body, thus affecting thyroid function (39). Autoimmune thyroid diseases (AITD) represent the most common group of autoimmune disorders, of which Hashimoto's thyroiditis (HT) and Graves' disease (GD) are major causes of hypothyroidism and hyperthyroidism, respectively. Research suggests that the gut microbiota and its metabolites might directly or indirectly impact thyroid immunity, thereby contributing

TABLE 3 Horizontal pleiotropy analysis for IVs of 12 GM taxa associated with HYPOTHYROIDISM.

Exposure	MR-Egg	jer interce	pt test	MR-PRESSO g	Cochran's Q	
	Egger_ intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value	<i>p</i> -value
Phylum						
Actinobacteria(ID:400)	0.016	0.010	0.135	13.397	0.579	0.569
Order						
Bifidobacteriales(ID:432)	-0.004	0.011	0.700	18.457	0.151	0.135
MollicutesRF9(ID:11579)	-0.008	0.009	0.398	15.689	0.308	0.267
Family						
Alcaligenaceae(ID:2875)	0.010	0.014	0.502	12.874	0.423	0.393
Defluviitaleaceae(ID:1924)	0.007	0.011	0.526	4.932	0.941	0.944
Genus						
Butyrivibrio(ID:1993)	-0.001	0.011	0.925	15.269	0.498	0.487
Desulfovibrio(ID:3173)	-0.010	0.011	0.403	12.710	0.385	0.332
Eggerthella(ID:819)	0.011	0.014	0.446	7.656	0.652	0.619
Intestinimonas(ID:2062)	-0.005	0.007	0.516	15.492	0.578	0.568
LachnospiraceaeUCG008(ID:11328)	0.044	0.016	0.026	21.512	0.061	0.050
Ruminiclostridium5(ID:11355)	-0.015	0.012	0.234	12.236	0.466	0.468
RuminococcaceaeUCG005(ID:11363)	0.008	0.011	0.465	26.541	0.055	0.039

SE, standard error; RSS, residual sum of squares; Cochran's Q-derived.

TABLE 4 Horizontal pleiotropy analysis for IVs of 13 GM taxa associated with HYPETHYROIDISM.

Exposure	MR-Egg	er interce	ept test	MR-PRESSC	Cochran's Q	
	Egger_ intercept	SE	<i>p</i> -value	RSS obs	<i>p</i> -value	<i>p</i> -value
Phylum						
Verrucomicrobia(ID:3982)	-0.001	0.037	0.977	12.373	0.539	0.572
Class						
Deltaproteobacteria(ID:3087)	-0.014	0.058	0.814	10.072	0.699	0.671
Order						
MollicutesRF9(ID:11579)	0.016	0.038	0.678	9.383	0.767	0.730
Family						
Desulfovibrionaceae(ID:3169)	0.006	0.061	0.928	6.658	0.738	0.729
Bacteroidaceae(ID:917)	-0.026	0.080	0.754	4.070	0.890	0.875
Genus						
RuminococcaceaeUCG004(ID:11362)	-0.013	0.077	0.873	12.308	0.447	0.424
RuminococcaceaeNK4A214group(ID:11358)	-0.063	0.043	0.173	13.66	0.493	0.472
Prevotella7(ID:11182)	-0.089	0.086	0.326	14.108	0.338	0.321
Parasutterella(ID:2892)	-0.012	0.031	0.713	8.297	0.912	0.910
Collinsella(ID:815)	-0.020	0.063	0.765	9.920	0.484	0.450
Catenibacterium(ID:2153)	-0.225	0.252	0.467	2.427	0.737	0.708
Bilophila(ID:3170)	-0.079	0.063	0.236	14.727	0.407	0.397
Bacteroides(ID:918)	-0.026	0.080	0.754	4.070	0.893	0.875

SE, standard error; RSS, residual sum of squares; Cochran's Q-derived.

to the development of AITD. These regulatory mechanisms may encompass inducing the shift from type 1 (Th1) to type 2 (Th2) Thelper cell responses (40), activation of toll-like receptor 4 by lipopolysaccharides (LPS) (41), and the induction of changes in transcriptional, proteomic, and metabolic changes.

In our study, we established two strong causal relationships. The phylum Actinobacteria, after Bonferroni correction, exhibited a significant reduction in the risk of hypothyroidism, while the class Deltaproteobacteria, post Bonferroni correction, demonstrated a significant reduction in the risk of hyperthyroidism. Actinobacteria is one of the four major phyla of the intestinal microbiota and is essential in maintaining intestinal homeostasis (42). Numerous Actinobacteria participate in the maintenance of microbial homeostasis, with some being considered probiotics that potentially exert an influence on thyroid function. Bifidobacterium, as a probiotic strain within the phylum Actinobacteria, is purported to possess the capacity to induce immune regulatory responses and mitigate inflammatory reactions (43). Several observational studies have suggested a diminished prevalence of Bifidobacteriaceae within individuals afflicted by hypothyroidism and hyperthyroidism (10, 44). And the administration of a mixture of Lactobacillus and Bifidobacterium in hypothyroid patients is effective in reducing the patient's need for LT-4 amounts, resulting in more stable

thyroid hormone levels (45, 46). Deltaproteobacteria, a class of Gram-negative proteobacteria, comprises obligate anaerobes, including numerous sulfate-reducing bacteria and sulfur-reducing bacteria (47). Among them, Desulfovibrionaceae is a prominent bacterial family within the class Deltaproteobacteria and is implicated in several diseases (48). In an animal experiment, iodine supplementation led to elevated levels of TT4, FT3, and TT3 in male ICR mice compared to the control group. However, in female mice, these values decreased in comparison to the control group. Subsequent analysis of intergroup fecal differences in mice revealed that iodine-supplemented female ICR mice exhibited higher abundance of Desulfovibrionaceae (49). Therefore, we postulate that the Desulfovibrionaceae family could potentially impact thyroid function through certain pathways. The specific mechanisms underlying this phenomenon should be further validated through experimental design.

Considering the potential for false negatives arising from Bonferroni correction, it becomes imperative to delve into the latent mechanisms underpinning the impact of the 32 nominal causal relationships on thyroid function. Preceding research has already elucidated a notable positive correlation between *Oscillospira* and diets low in fat and lean meat, as well as with human health. Moreover, *Oscillospira* has been demonstrated to

engender various short-chain fatty acids, prominently including butyrate (50). Concurrently, there exists a positive correlation between body mass index (BMI) and serum thyroid-stimulating hormone (TSH) levels (51). Based on these findings, it is conceivable that Oscillospira, by generating short-chain fatty acids and reducing BMI, might exert regulatory influence on TSH levels. Furthermore, Veillonella genus, a non-motile anaerobic Gramnegative diplococcus, resides within the human gastrointestinal tract (52). Discrepancies in microbial composition have been noted between patients afflicted by primary hypothyroidism and healthy individuals (10). An illustrative case-control study has evidenced a significant reduction in Veillonella abundance among children with attention deficit hyperactivity disorder (ADHD) (53). Given the association of ADHD with neurotransmitter imbalances like dopamine (53, 54), a plausible hypothesis emerges that Veillonella could potentially modulate the hypothalamic-pituitaryadrenal (HPA) axis and dopamine secretion, thereby exerting an inhibitory effect on TSH levels. Expanding further, in patients with hypothyroidism (HT) compared to their euthyroid counterparts, heightened abundances of microbial taxa including Lachnospiraceae incertae sedis, Lactonifactor, Alistipes, and Subdoligranulum have been observed (55). Integrating our research outcomes, we may conjecture that Subdoligranulum and Lachnospiraceae might influence free thyroxine (FT4) levels, thus impacting thyroid function. However, the intricate mechanisms necessitate in-depth investigation. The specific mechanisms require further investigation.

The present study has several advantages, firstly this is the first study to investigate the association between gut flora and thyroid function using GWAS data from the Finnish Biosample Repository and the Thyroidomics Consortium and the MiBiogen Consortium, secondly, MR method analysis can provide robust causality estimates by minimizing reverse causal effects or confounding factors, and finally we performed sensitivity analysis of the results by Mr-Egger intercept test and MR-PRESSO global test to ensure the robustness of the results. However, this study has some limitations, first we used summary level statistics, could not analyze specific disease subgroups, and did not explore the nonlinear relationship between exposure and outcome. In addition, we studied a European population, which limits generalizability to other ethnicities. Thyroid disease is more prevalent in female populations (56), however our study did not differentiate between genders, which may limit the understanding of the differential impact of gut flora on thyroid disease production by gender.

5 Conclusions

In conclusion, in this study, we conducted a two-sample MR analysis to investigate the causal relationships between gut microbiota and FT4, TSH, hypothyroidism, and hyperthyroidism. After using the bonferroni method for multiple testing correction, we identified 2 causal relationships and 32 nominally significant causal relationships. However, further research is needed to explore the specific effects of gut microbiota on thyroid function.

Data availability statement

The original contributions presented in the study are publicly available. This data can be found at the MiBioGen repository (https://mibiogen.gcc.rug.nl/), the FinnGen repository (https://r8. finngen.fi/) and the ThyroidOmics Consortium (https://transfer. sysepi.medizin.uni-greifswald.de/thyroidomics/datasets/).

Ethics statement

This research has been conducted using published studies and consortia providing publicly available summary statistics. Therefore, no additional separate ethical approval was required for this study.

Author contributions

LX designed the study, analyzed the data, and wrote the manuscript. HZ assisted in creating the tables and revising the manuscript. WC critically read and edited the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by National Natural Science Foundation of China (No. 82004340). WC is the project leader.

Acknowledgments

We gratefully thank The Thyroid Omics Consortium and MiBioGen Consortium for providing statistical data. And we want to acknowledge the participants and investigators of the FinnGen study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fendo.2023.1240752/ full#supplementary-material

References

1. Williams GR. Neurodevelopmental and neurophysiological actions of thyroid hormone. *J Neuroendocrinol* (2008) 20(6):784-794. doi: 10.1111/j.1365-2826.2008.01733.x

 Chiamolera MI, Wondisford FE. Minireview: Thyrotropin-releasing hormone and the thyroid hormone feedback mechanism. *Endocrinology* (2009) 150(3):1091– 1096. doi: 10.1210/en.2008-1795

3. Teumer AA-OX, Chaker L, Groeneweg S, Li Y, Di Munno C, Barbieri C, et al. Genome-wide analyses identify a role for SLC17A4 and AADAT in thyroid hormone regulation. *Nat Commun* (2018) 9(1):4455. doi: 10.1038/s41467-018-06356-1

4. Hollowell JG, Staehling Nw, Flanders Wd, Hannon Wh, Gunter Ew, Spencer Ca, et al. Serum TSH, T(4), and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). J Clin Endocrinol Metab (2002) 87(2):489–499. doi: 10.1210/jcem.87.2.8182

5. Abdel-Moneim AA-O, Gaber AM, Gouda S, Osama A, Othman SI, Allam G. Relationship of thyroid dysfunction with cardiovascular diseases: updated review on heart failure progression. *Hormones (Athens)* (2020) 19(3):301–309. doi: 10.1007/ s42000-020-00208-8

6. Chaker I., Bianco AC, Jonklaas J, Peeters RP. Hypothyroidism. Lancet (2017) 390 (10101):1550–1562. doi: 10.1016/S0140-6736(17)30703-1

7. Brandt F, Green A Fau - Hegedüs L, Hegedüs L Fau - Brix TH, Brix TH. A critical review and meta-analysis of the association between overt hyperthyroidism and mortality. *Eur J Endocrinol* (2011 165(4):491–7. doi: 10.1530/EJE-11-0299

8. Ross DS, Burch HB, Cooper DS, Greenlee MC, Laurberg P, Maia AL, et al. American thyroid association guidelines for diagnosis and management of hyperthyroidism and other causes of thyrotoxicosis. *Thyroid* (2016) 26(10):1343–421. doi: 10.1089/thy.2016.0229

9. Zhou L, Li X, Ahmed A, Wu D, Liu L, Qiu J, et al. Gut microbe analysis between hyperthyroid and healthy individuals. *Curr Microbiol* (2014) 69(5):675–680. doi: 10.1007/s00284-014-0640-6

10. Su X, Zhao Y, Li Y, Ma S, Wang Z. Gut dysbiosis is associated with primary hypothyroidism with interaction on gut-thyroid axis. *Clin Sci (Lond)* (2020) 134 (12):1521–35. doi: 10.1042/CS20200475

11. Lerner A, Jeremias P, Matthias T. Gut-thyroid axis and celiac disease. Endocr Connect (2017) 6(4):R52-R58. doi: 10.1530/EC-17-0021

12. Yao SA-O, Zhang MA-O, Dong SA-O, Wang JA-O, Zhang K, Guo JA-O, et al. Bidirectional two-sample Mendelian randomization analysis identifies causal associations between relative carbohydrate intake and depression. *Nat Hum Behav* (2022) 6(11):1569–76. doi: 10.1038/s41562-022-01412-9

13. Ellervik C, Roselli C, Christophersen IE, Alonso A, Pietzner M, Sitlani CM, et al. Assessment of the relationship between genetic determinants of thyroid function and atrial fibrillation: A mendelian randomization study. *JAMA Cardiol* (2019) 4(2):144–52. doi: 10.1001/jamacardio.2018.4635

14. Wang JJ, Zhuang ZH, Shao CL, Yu CQ, Wang WY, Zhang K, et al. Assessment of causal association between thyroid function and lipid metabolism: a Mendelian randomization study. *Chin Med J (Engl)* (2021) 134(9):1064–9. doi: 10.1097/CM9.00000000001505

15. Marouli E, Yusuf L, Kjaergaard AD, Omar R, Kuś A, Babajide O, et al. Thyroid function and the risk of alzheimer's disease: A mendelian randomization study. *Thyroid* (2021) 31(12):1794–9. doi: 10.1089/thy.2021.0321

16. Xiang SA-O, Jia TA-O, Xie C, Cheng W, Chaarani B, Banaschewski TA-O, et al. Association between vmPFC gray matter volume and smoking initiation in adolescents. *Nat Commun* (2023) 14(1):4684. doi: 10.1038/s41467-023-40079-2

17. Skrivankova VW, Richmond RC, Woolf BAR, Yarmolinsky J, Davies NM, Swanson SA, et al. Strengthening the reporting of observational studies in epidemiology using mendelian randomization: the STROBE-MR statement. *JAMA* (2021) 326(16):1614–21. doi: 10.1001/jama.2021.18236

18. Kurilshikov AA-O, Medina-Gomez CA-O, Bacigalupe RA-OX, Radjabzadeh D, Wang J, Demirkan A, et al. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat Genet* (2021) 53(2):156–165. doi: 10.1038/s41588-020-00763-1

19. Kurki MI, Karjalainen J, Palta P, Sipilä TP, Kristiansson K, Donner KM, et al. FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature* (2023) 613(7944):508–18. doi: 10.1038/s41586-022-05473-8

20. Burgess S, Thompson SG. Bias in causal estimates from Mendelian randomization studies with weak instruments. *Stat Med* (2011) 30(11):1312–23. doi: 10.1002/sim.4197

21. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. *Genet Epidemiol* (2013) 37 (7):658–65. doi: 10.1002/gepi.21758

22. Burgess SA-O, Thompson SG. Interpreting findings from Mendelian randomization using the MR-Egger method. *Eur J Epidemiol* (2017) 32(5):377–389. doi: 10.1007/s10654-017-0255-x

23. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in mendelian randomization with some invalid instruments using a weighted median estimator. *Genet Epidemiol* (2016) 40(4):304–14. doi: 10.1002/gepi.21965

24. Hartwig FP, Davey Smith G, Bowden J. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. *Int J Epidemiol* (2017) 46(6):1985–98. doi: 10.1093/ije/dyx102

25. Hemani GA-O, Zheng JA-O, Elsworth B, Wade KA-O, Haberland V, Baird DA-O, et al. The MR-Base platform supports systematic causal inference across the human phenome. *Elife.* (2018) 7:e34408. doi: 10.7554/eLife.34408

26. Greco MF, Minelli C, Sheehan NA, Thompson JR. Detecting pleiotropy in Mendelian randomisation studies with summary data and a continuous outcome. *Stat Med* (2015) 34(21):2926–2940. doi: 10.1002/sim.6522

27. Bowden J, Del Greco MF, Minelli C, Davey Smith G, Sheehan NA, Thompson JR. Assessing the suitability of summary data for two-sample Mendelian randomization analyses using MR-Egger regression: the role of the I2 statistic. *Int J Epidemiol* (2016) 45(6):1961–1974. doi: 10.1093/ije/dyw220

28. Fröhlich E, Wahl R. Microbiota and thyroid interaction in health and disease. *Trends Endocrinol Metab* (2019)30(8):479–90. doi: 10.1016/j.tem.2019.05.008

29. Zimmermann MB, Boelaert K. Iodine deficiency and thyroid disorders. Lancet Diabetes Endocrinol (2015) 3(4):286–95. doi: 10.1016/S2213-8587(14)70225-6

30. Köhrle JA-O. Selenium, iodine and iron-essential trace elements for thyroid hormone synthesis and metabolism. *Int J Mol Sci* (2023) 24(4):3393. doi: 10.3390/ ijms24043393

31. Nicola JP, Nazar M, Mascanfroni Id, Pellizas Cg, Masini-Repiso AM. NF-kappaB p65 subunit mediates lipopolysaccharide-induced Na(+)/I(-) symporter gene expression by involving functional interaction with the paired domain transcription factor Pax8. *Mol Endocrinol* (2010) 24(9):1846–62. doi: 10.1210/me.2010-0102

32. Sponziello M, Scipioni A, Durante C, Verrienti A, Maranghi M, Giacomelli L, et al. Regulation of sodium/iodide symporter and lactoperoxidase expression in four human breast cancer cell lines. *J Endocrinol Invest* (2010) 33(1):2–6. doi: 10.1007/ BF03346542

33. Fekete C, Gereben B, Doleschall M, Harney Jw, Dora Jm, Bianco Ac, et al. Lipopolysaccharide induces type 2 iodothyronine deiodinase in the mediobasal hypothalamus: implications for the nonthyroidal illness syndrome. *Endocrinology* (2004) 145(4):1649–55. doi: 10.1210/en.2003-1439

34. Zimmermann MB, Köhrle J. The impact of iron and selenium deficiencies on iodine and thyroid metabolism: biochemistry and relevance to public health. *Thyroid* (2002) 12(10):867–78. doi: 10.1089/105072502761016494

35. Knezevic J, Starchl C, Tmava Berisha A, Amrein K. Thyroid-gut-axis: How does the microbiota influence thyroid function? *Nutrients* (2020) 12(6):1769. doi: 10.3390/ nu12061769

36. Gorini FA-O, Sabatino LA-O, Pingitore AA-OX, Vassalle CA-O. Selenium: an element of life essential for thyroid function. Molecules (2021) 26(23):7084. doi: 10.3390/molecules26237084

37. Kasaikina MV, Kravtsova Ma, Lee Bc, Seravalli J, Peterson Da, Walter J, et al. Dietary selenium affects host selenoproteome expression by influencing the gut microbiota. *FASEB J* (2011) 25(7):2492–9. doi: 10.1096/fj.11-181990

38. Farzi AA-O, Fröhlich EE, Holzer P. Gut microbiota and the neuroendocrine system. *Neurotherapeutics* (2018) 15(1):5-22. doi: 10.1007/s13311-017-0600-5

39. Bowden SA, Goldis M. Congenital Hypothyroidism. *StatPearls*. Treasure Island (FL: StatPearls Publishing) (2023).

40. Wu B, Xu Y, Ban Y, Zhang M, Sun Z, Cai Y, et al. Correlation between the intestinal microflora and peripheral blood Th1/Th2 balance in hypothyroidism during the first half of pregnancy. *Front Cell Infect Microbiol* (2023) 13:1159238. doi: 10.3389/ fcimb.2023.1159238

41. Nicola JP, Vélez Ml, Lucero Am, Fozzatti L, Pellizas Cg, Masini-Repiso AM. Functional toll-like receptor 4 conferring lipopolysaccharide responsiveness is expressed in thyroid cells. *Endocrinology* (2009) 150(1):500–8. doi: 10.1210/en.2008-0345

42. Binda C, Lopetuso LR, Rizzatti G, Gibiino G, Cennamo V, Gasbarrini A. Actinobacteria: A relevant minority for the maintenance of gut homeostasis. *Dig Liver Dis* (2018) 50(5):421–8. doi: 10.1016/j.dld.2018.02.012

43. Derrien M, Turroni F, Ventura M, van Sinderen D. Insights into endogenous Bifidobacterium species in the human gut microbiota during adulthood. *Trends Microbiol* (2022) 30(10):940–7. doi: 10.1016/j.tim.2022.04.004

44. Bertani B, Ruiz N. Function and biogenesis of lipopolysaccharides. *EcoSal Plus* (2018) 8(1):10.1128/ecosalplus.ESP-0001-2018. doi: 10.1128/ecosalplus.ESP-0001-2018

45. Spaggiari G, Brigante G, De Vincentis S, Cattini U, Roli L, De Santis MC, et al. Probiotics ingestion does not directly affect thyroid hormonal parameters in hypothyroid patients on levothyroxine treatment. *Front Endocrinol (Lausanne)* (2017) 8:316. doi: 10.3389/fendo.2017.00316

46. Karimifar M, Heidari Z, Mohammadi H, Askari G. The effects of synbiotic supplementation on thyroid function and inflammation in hypothyroid patients: A randomized, double-blind, placebo-controlled trial. *Complement Ther Med* (2020) 48:102234. doi: 10.1016/j.ctim.2019.102234

47. Totzeck AA-O, Ramakrishnan E, Schlag M, Stolte B, Kizina K, Bolz S, et al. Gut bacterial microbiota in patients with myasthenia gravis: results from the MYBIOM

study. Ther Adv Neurol Disord (2021) 14:17562864211035657. doi: 10.1177/17562864211035657

48. Wang RA-O, Zhang LA-O, Chen LP, Wang JY, Zhang L, Xu YS, et al. Structural and functional modulation of gut microbiota by jiangzhi granules during the amelioration of nonalcoholic fatty liver disease. *Oxid Med Cell Longev* (2021) 2021:2234695. doi: 10.1155/2021/2234695

49. Shen H, Xu JA-O, Lu C, Han J, Zhou JA-O, Ming T, et al. Effects of the sex factor on mouse iodine intake: interactions between the gut microbiota composition and metabolic syndromes. *ACS Omega* (2021) 6(43):28569–78. doi: 10.1021/acsomega.1c02697

50. Yang JA-O, Li Y, Wen Z, Liu W, Meng L, Huang H. Oscillospira - a candidate for the next-generation probiotics. *Gut Microbes* (2021) 13(1):1987783. doi: 10.1080/19490976.2021.1987783

51. Babić Leko MA-O, Gunjača IA-O, Pleić NA-OX, Zemunik TA-O. Environmental factors affecting thyroid-stimulating hormone and thyroid hormone levels. *Int J Mol Sci* (2021) 22(12):6521. doi: 10.3390/ijms22126521

52. Karki S, Mainali A, Pandey S, Uprety N, Panigrahi K, Adhikari S. Veillonella bacteremia in a patient with metastatic colorectal carcinoma. *Cureus* (2023) 15(6): e41152. doi: 10.7759/cureus.41152

53. Wan L, Ge WR, Zhang S, Sun YL, Wang B, Yang G. Case-control study of the effects of gut microbiota composition on neurotransmitter metabolic pathways in children with attention deficit hyperactivity disorder. *Front Neurosci* (2020) 14:127. doi: 10.3389/fnins.2020.00127

54. Magula LA-O, Moxley KA-OX, Lachman AA-O. Iron deficiency in South African children and adolescents with attention deficit hyperactivity disorder. *J Child Adolesc Ment Health* (2019) 31(2):85–92. doi: 10.2989/17280583.2019.1637345

55. Liu S, An Y, Cao B, Sun R, Ke JA-O, Zhao DA-O. The composition of gut microbiota in patients bearing hashimoto's thyroiditis with euthyroidism and hypothyroidism. *Int J Endocrinol* (2020) 2020:5036959. doi: 10.1155/2020/5036959

56. Carbonero F, Benefiel Ac, Alizadeh-Ghamsari Ah, Gaskins HR. Microbial pathways in colonic sulfur metabolism and links with health and disease. *Front Physiol* (2012) 3:448. doi: 10.3389/fphys.2012.00448