Check for updates

OPEN ACCESS

EDITED BY Hubert Vaudry, Université de Rouen, France

REVIEWED BY

Mingmei Zhou, Shanghai University of Traditional Chinese Medicine, China Jacob Raber, Oregon Health and Science University, United States

*CORRESPONDENCE Jody A. Rusch jody.rusch@uct.ac.za

RECEIVED 23 December 2022 ACCEPTED 25 May 2023 PUBLISHED 19 June 2023

CITATION

Rusch JA, Layden BT and Dugas LR (2023) Signalling cognition: the gut microbiota and hypothalamic-pituitary-adrenal axis. *Front. Endocrinol.* 14:1130689. doi: 10.3389/fendo.2023.1130689

COPYRIGHT

© 2023 Rusch, Layden and Dugas. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Signalling cognition: the gut microbiota and hypothalamicpituitary-adrenal axis

Jody A. Rusch^{1,2*}, Brian T. Layden^{3,4} and Lara R. Dugas^{5,6}

¹Division of Chemical Pathology, Department of Pathology, University of Cape Town, Cape Town, South Africa, ²C17 Chemical Pathology Laboratory, Groote Schuur Hospital, National Health Laboratory Service, Cape Town, South Africa, ³Division of Endocrinology, Diabetes, and Metabolism, Department of Medicine, University of Illinois at Chicago, Chicago, IL, United States, ⁴Department of Medicine, Jesse Brown Veterans Affairs Medical Center, Chicago, IL, United States, ⁵Division of Epidemiology and Biostatistics, School of Public Health, University of Cape Town, Cape Town, South Africa, ⁶Public Health Sciences, Parkinson School of Health Sciences and Public Health, Loyola University Chicago, Maywood, IL, United States

Cognitive function in humans depends on the complex and interplay between multiple body systems, including the hypothalamic-pituitary-adrenal (HPA) axis. The gut microbiota, which vastly outnumbers human cells and has a genetic potential that exceeds that of the human genome, plays a crucial role in this interplay. The microbiota-gut-brain (MGB) axis is a bidirectional signalling pathway that operates through neural, endocrine, immune, and metabolic pathways. One of the major neuroendocrine systems responding to stress is the HPA axis which produces glucocorticoids such as cortisol in humans and corticosterone in rodents. Appropriate concentrations of cortisol are essential for normal neurodevelopment and function, as well as cognitive processes such as learning and memory, and studies have shown that microbes modulate the HPA axis throughout life. Stress can significantly impact the MGB axis via the HPA axis and other pathways. Animal research has advanced our understanding of these mechanisms and pathways, leading to a paradigm shift in conceptual thinking about the influence of the microbiota on human health and disease. Preclinical and human trials are currently underway to determine how these animal models translate to humans. In this review article, we summarize the current knowledge of the relationship between the gut microbiota, HPA axis, and cognition, and provide an overview of the main findings and conclusions in this broad field.

KEYWORDS

microbiota-gut-brain axis, hypothalamic-pituitary-adrenal axis, stress, cortisol, glucocorticoids, cognition

1 Introduction

Humans have a longstanding and intimate, life-long, relationship with microbes, collectively known as the human microbiota, which plays a key role in influencing bodily systems responsible for human health and disease (1-4). The gut microbiota (GM), which comprises a complex, heterogeneous ecosystem of microorganisms, including bacteria, archaea, fungi, protozoa, viruses, and parasites, is a vital component of the human microbiota (5-9). The bacterial population is the most extensively characterised subset of the GM. While initially believed that microbial cells outnumbered human cells by a factor of 10 to 1, the current estimate is approximately 1.3 to 1 (10). Notably, the Human Gut Microbiome (HRGM) has recently been expanded to include 232,098 non-redundant genomes for 5,414 representative prokaryotic species, with over 103 million unique proteins (11). While the human genome is essentially stable and limited in flexibility for the lifespan of the host, the vast genetic potential of the microbes is dynamic and responsive to the environment. This suggests that the GM is an important environmental factor for humans, with evolutionarily conserved roles in the metabolism, immunity, development, and behaviour of the host (2, 12-17). Recently, pivotal roles in endocrine and neural development and function have started to be elucidated (18, 19).

Emerging evidence suggests that the GM, the hypothalamicpituitary-adrenal (HPA) axis, and cognitive processes are linked bidirectionally *via* multiple pathways, including the vagus nerve, neurotransmitter and metabolite production, immune system and blood-brain barrier regulation, and hormone metabolism. Alterations in the GM, whether due to diet, antibiotics, or other factors, can impact the stress response, HPA axis activity, and overall cognitive health. This review aims to summarize the current understanding of the role of the GM in regulating the HPA axis, a key component of the gut-brain axis. The review further explores the mechanisms and pathways through which the GM can alter gutbrain communication, with an emphasis on the effects on the HPA axis and its influence on cognition.

2 The microbiota-gut-brain axis

The MGB axis refers to the bidirectional communication network between the CNS, the autonomic nervous system (ANS), the endocrine system, the immune system, and the GM (20–23). This system enables microbes to share information with the brain, and the brain to communicate with the gut (24, 25). Despite extensive data from animal and human studies supporting the role of the GM in the MGB axis, the mechanisms by which the GM impacts the human brain are not yet fully understood. What is known has primarily been learnt from studies using germ-free animal models, and studies examining the effects of specific microbial species, probiotics, antibiotics, and infections. Further, technological advancements in sequencing and metabolomics have enabled scientists to explore this topic more thoroughly.

As a key regulator, the GM can modulate host physiological processes through several proposed mechanisms (25, 26). These

include microbial constituents [e.g. lipopolysaccharides (LPS) and peptidoglycans (PG)] (25), microbial products [e.g. enzymes, short chain fatty acids (SCFAs), and neurotransmitters] (27, 28), hormone release (e.g. glucocorticoids) (29), and substrate metabolism [e.g. bile acids (BAs) and tryptophan] (30, 31). In the gut, the GM and its metabolites have been shown to modulate gastrointestinal functions *via* their effects on intestinal permeability (32–35), mucosal immune function (36–38), intestinal motility and sensitivity (39, 40), and the enteric nervous system (ENS) (21, 24, 25, 41). The GM can stimulate the release of peptides and hormones from enteroendocrine cells, which can have direct or indirect central effects (22, 39). Further neuroendocrine and metabolic pathways link the GM with the CNS (21).

The GM is critical to the development and functioning of the CNS. Studies in animals have demonstrated the effect that the GM has on neural development and neurochemistry in the host, influencing the stress system, behavior, and cognition (42–47). Conversely, the brain can affect intestinal function and the GM, for example, by HPA axis-mediated glucocorticoid modification of immunity in response to stress (29).

2.1 The role of the HPA axis in the MGB axis

The HPA axis plays a central role in mediating the stress response and regulating the interaction between the GM, gut, and brain (42).

Mechanistically, cortisol can impact the MGB axis through multiple pathways. Cortisol receptors are expressed on various cells of the gut, including epithelial cells, immune cells, and enteroendocrine cells, indicating a direct effect of cortisol on gut function (21, 29, 48). Cortisol can also affect the gut microbiota by altering gut transit time, intestinal permeability, and nutrient availability, which can in turn impact the composition and diversity of the GM (48). Furthermore, cortisol can impact the brain by binding to glucocorticoid receptors (GRs) located in various brain regions, including the hippocampus, amygdala, and prefrontal cortex. There is also evidence for signaling between the GM and CNS, since microbes residing in the gut can activate stress circuits in the CNS through the vagus nerve and sensory neurons of the ENS (21, 49–54).

Chronic or prolonged stress can lead to dysregulation of the HPA axis, which can have negative effects on various bodily systems, including the MGB axis (55). Elevated cortisol levels have been associated with alterations in GM composition and increased gut permeability, which can lead to inflammation and contribute to brain dysfunction and various CNS disorders (56).

The effects of dysregulation of the HPA axis has primarily been studied using various modalities of stress. Preclinical evidence suggests that GM-mediated mechanisms are likely involved in modulating brain processes, including brain biochemistry, response to stress, pain interpretation, feeding, emotional behaviors, and cognition (57–64). Initial clinical evidence of MGB interactions primarily stems from associations between dysbiosis and CNS disorders, such as autism, anxiety-depressive behaviors,

and functional gastrointestinal disorders (63, 65–72). Furthermore, recent studies have shown dramatic changes in the GM of patients with Alzheimer's disease, Parkinson's disease, multiple sclerosis, and schizophrenia (73–77). Fecal microbiota transplant (FMT) studies have added evidence for causality by inducing many of the symptoms of these diseases in germ-free animals (78).

2.2 Development and life course of the MGB axis

There exists some controversy regarding prenatal exposure to microbes and their importance to fetal development (79, 80). However, the acquisition of microbes occurs primarily at birth, with delivery through the birth canal exposing the infant to its mother's microbiota, resulting in vertical transmission of an initial maternal signature (81). Caesarean section delivery alters the initial microbial composition (82). After birth, several factors influence GM composition in early life, including breastfeeding, nutrition, infection and antibiotic use, environmental stressors, and host genetics (83). While initial microbial diversity is low, it escalates rapidly as a function of diet and environment, with an increase in the relative composition of strict anaerobes (84). A more stable and complex, adult-like microbiota starts to emerge as early as one to three years of age, although this development may continue as late as pre-adolescence (85, 86). The first year of life encompasses a critical "window period" of development in which the GM may be more susceptible to environmental influences and highly influential with regards to the overall health of the host. This critical window of GM development aligns with critical windows of development of other systems that are also more sensitive and vulnerable to environmental input, such as the immune system, HPA axis, and brain development in general.

The GM of children is characterized by relatively higher abundances of microbes with genes that function to support human development, including vitamin biosynthesis and polysaccharide and xenobiotic metabolism (84). Adolescence is characterized by an intense period of sexual development and growth, and recent data indicate that the GM undergoes progressive changes during this period likely due to hormonal surges, stressors, and other age-related factors (87). In adulthood, the core GM is relatively stable, and the main factors influencing its composition are diet, exercise, stress, disease, and medications (88). Confirming the role of the GM in cognitive function in midlife, Meyer et al. explored the relationship between gut microbial community composition and cognitive function in 597 middleaged adults, and found a significant positive association between β diversity and all higher cognitive functions tested (89).

The aging process of the host parallels a continuous aging progression of the GM (90). Stability begins to decline as we age, and studies suggest that there is greater inter-individual variation in older adults than younger adults (<65 years old) (91, 92) There is also evidence that the diversity levels of the GM differ with aging, with a reduction in the number of different bacterial species present and changes in the relative abundance of different bacterial phyla, with a decrease in beneficial bacteria such as Bifidobacteria and an

increase in potentially harmful bacteria such as Proteobacteria (93– 96). Microbial composition can predict human chronological age relatively accurately (97). In a study that used metagenomic profiles from over 4,000 healthy people aged 18–90, the authors were able to construct an algorithm that predicted individuals' ages within approximately six years of their actual age (98). Functionally, an associated shift towards a more pro-inflammatory state has been noted, along with a decline in immune function, making older individuals more susceptible to infections and other diseases (96, 99). The GM of older individuals may be less efficient at metabolizing certain nutrients, which could affect overall health (92). These alterations may contribute to age-related inflammation, oxidative stress, and neurodegeneration, and have been associated with age-related health issues such as frailty and cognitive decline (92, 100, 101).

There is growing evidence that age-related changes in the microbiota may contribute to cognitive decline and other neurodegenerative disorders (102, 103). Human studies comparing the composition of the GM of elderly participants suffering from cognitive impairment with healthy individuals indicate significant changes in GM composition. Specifically, there is an increase in pro-inflammatory taxa and a reduction in anti-inflammatory taxa (104–109).

The HPA axis also undergoes age-related changes including alterations in the sensitivity and responsiveness of the hypothalamus and pituitary gland to negative feedback by cortisol, changes in the levels and circadian rhythm of circulating cortisol, changes in the expression and function of CRH and glucocorticoid receptors in the hypothalamus, hippocampus, and prefrontal cortex, changes in the activity and connectivity of brain regions involved in stress regulation, including the amygdala, hippocampus, and prefrontal cortex, and changes in the microbiota, which can impact HPA axis function and contribute to inflammation and oxidative stress (110). Essentially, the HPA axis appears to become less responsive with aging, contributing to the dysregulation of stress responses and the resultant development of cognitive decline (111, 112).

Similar to aging, dysbiosis has been associated with increased cortisol secretion, decreased negative feedback at the level of the hypothalamus, and changes in the circadian rhythm of cortisol release (113).

2.3 The influence of the gut microbiota on neurodevelopment

Animal studies suggest that the GM plays a crucial role in regulating early brain development (19, 45, 60, 114–117). During key prenatal and postnatal periods, neurodevelopment depends on the integration of environmental cues, such as MGB axis molecular signaling (116). The maturation of neuronal networks is essential for the developing nervous system to form functional neural circuitry. Microglial cells, innate immune cells of the brain, play a critical role in the elimination of unnecessary synaptic connections required for this maturation process (118, 119). The GM appears to influence microglial development and maturation (120, 121), and

animal models have demonstrated downstream effects on various aspects of neurocognitive function (19, 45, 60, 114, 115). In contrast, early-life disruptions of gut colonization have been linked to CNS alterations (122).

Germ-free animal models have been crucial in developing an understanding of the role that the GM plays in neurodevelopment (123). The nervous systems of germ-free animals develop differently from those conventionally colonized, exhibiting key differences in multiple neurotransmitter systems and their receptors (25). They display increases in neurogenesis, hippocampal and amygdala volume, myelination, and myelin plasticity-related genes in the prefrontal cortex (25). Their dendrites are longer, and spines are denser, but there are fewer synaptic connections. In the amygdala, they have increased synaptic and neural plasticity-related genes and increased neuronal activity-related genes (124). They present with immature microglia and decreased immune system-related genes. Due to the decreased expression of tight junction proteins, they have a more permeable blood-brain barrier (BBB). Correspondingly, they have impaired immune systems, dysregulated hormone signalling, altered metabolism, and differences in neurotransmission (25). Finally, germ-free animal studies demonstrate that the GM is necessary for normal stress responsivity, anxiety-like behaviors, sociability, and cognition (123).

3 Mechanisms and pathways of the MGB axis

Although the precise mechanisms involved in the crosstalk between the GM and brain remain to be fully determined, there are several putative mechanisms and pathways (125). Microbes influence CNS processes *via* modulation of the nervous system (52), endocrine system (51, 126), and immune system (120), together with their ability to synthesize neurotransmitters (127–129) and produce metabolites (127, 130–132) (Figure 1). Together, these mechanisms and pathways illustrate the complex interplay between the GM and the brain, highlighting the importance of understanding the MGB axis in health and disease.

3.1 The HPA axis, glucocorticoids, and the stress response

The HPA axis is a major stress response system in the body, with neurons in the hypothalamic paraventricular nucleus (PVN) synthesizing and secreting corticotropin-releasing hormone (CRH) and antidiuretic hormone (ADH) in response to stress (133). These peptides stimulate the release of adrenocorticotropic hormone (ACTH) from corticotropic cells of the anterior pituitary gland, which in turn targets the zona fasciculata of the adrenal cortex to produce glucocorticoids, such as cortisol in humans and corticosterone in rodents (133–135). Cortisol acts on the hypothalamus and pituitary in a negative feedback loop to regulate the response (133). The PVN's activity is regulated by various afferent systems, including the sympathetic (SNS) and parasympathetic nervous system (PNS), and limbic circuits (136). The interplay of central and peripheral systems produces the characteristic behavioral, endocrine, autonomic, and immune responses to stress (135, 137). The release of cortisol is characterized by both circadian and 60- to 90-minute oscillations (138). The normal 24-hour profiles of both ACTH and cortisol show an early morning peak, decreasing concentrations throughout the day, a nadir around midnight, and an abrupt elevation during late sleep culminating in the early morning peak (138).

Glucocorticoids are essential for regulating cellular processes, including metabolism, growth, differentiation, and apoptosis, and act *via* intracellular receptors in the nuclear receptor superfamily (139). They regulate the transcription of target genes in organ systems that maintain homeostasis and help the body cope with physical and psychological stress (134, 135). Glucocorticoids are involved in several processes related to host defense, including immunity and inflammation, as well as metabolism, growth, cardiovascular function, water and electrolyte balance, reproduction, and mood and cognition (49, 140–148).

The brain is a crucial target organ for glucocorticoids, and their actions are mediated by the mineralocorticoid receptor (MR) and GR, which act as transcription factors and mediate non-genomic steroid effects (149). During the early stages of acute stress, MR activation is required for the appraisal process and memory retrieval, while GR promotes memory consolidation and behavioral adaptation (150). Glucocorticoids also play a vital role in central nervous system (CNS) development and are required for normal maturation (139). In adulthood, they contribute to neuronal plasticity, and have been implicated in neurodegenerative processes.

HPA axis dysregulation can result in hyper- or hypocortisolism, excessive or dampened reactivity to stressors, and circadian rhythm abnormalities (17). HPA axis dysfunction is linked to a decline in cognitive function, aging, immune system dysfunction, and systemic inflammation (135). Individuals with altered HPA axis function are also more likely to develop metabolic disorders such as cardiovascular disease, diabetes, and inflammatory bowel disease (IBD) (142, 151). Neuropsychiatric symptoms such as depression, mania, anxiety, and neurocognitive impairment are associated with both hyper- and hypocortisolism (143, 144). HPA axis dysfunction is also associated with many major psychopathologies, including autism, anxiety disorders, depression, and schizophrenia, as well as other cognitive disorders (145).

3.2 The nervous system

3.2.1 The autonomic nervous system

The ANS regulates involuntary physiological processes throughout the body, except for skeletal muscle, providing neural control (152). In the gastrointestinal system, both the SNS and PNS transmit afferent signals arising from the lumen to the CNS (via enteric, spinal and vagal pathways) and efferent signals from the

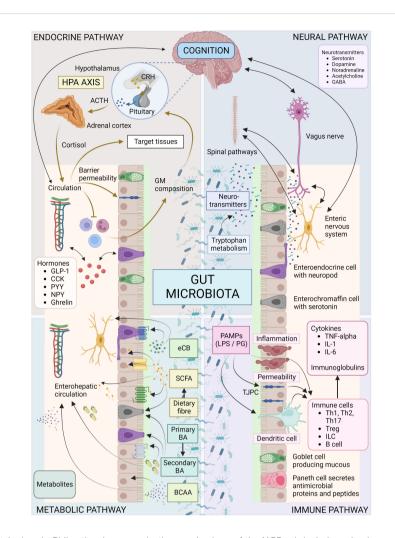


FIGURE 1

Overview of microbiota-gut-brain axis. Bidirectional communication mechanisms of the MGB axis include endocrine, neural, metabolic and immune system pathways. The hypothalamic-pituitary-adrenal axis is a major neuro-endocrine system responding to stress with the release of corticotrophin-releasing hormone (CRH) from the hypothalamus, and the subsequent release of ACTH from the pituitary, then cortisol from the adrenal cortex. Cortisol reaches target tissues through the circulation, modulates the immune system, and impacts on GM composition and gut permeability. The GM in turn is able to influence the stress response (for e.g., the HPA axis can be activated in response to increased circulating cytokines subsequent to bacterial translocation). Various GM and enteroendocrine cell interactions result in the release of hormones that work locally or on target tissues such as the brain, *via* the circulation. The vagus nerve, enteric nervous system, and spinal pathways provide rapid neural communication routes, while neurotransmitters or their precursors can be produced or metabolized by microbes. Metabolites such as SCFA, BA and eCB may be produced or modified by microbes and bind specific cell receptors in the gut or they may be absorbed into circulation and affect target tissues. Microbes and their products may interact with the immune cells with downstream pro-inflammatory or anti-inflammatory effects. ACTH, Adrenocorticotropic hormone; BA, bile acid; BCAA, branched chain amino acids; CCK, cholecystokinin; CRH, corticotropin-releasing hormone; eCB, endocannabinoid; GABA, γ -aminobutyric acid; GLP-1, glucagon-like peptide 1; GM, gut microbiota; HPA, hypothalamic-adrenal-pituitary; IL, interleukin; ILC, innate lymphoid cells; LPS, lipopolysaccharide; PYY, Peptide YY; NPY, neuropeptide Y; PAMP, Pathogen-associated molecular pattern; PG, peptidoglycan; SCFA, short chain fatty acid; Th, T helper cell; TJPC, tight junction protein complex; T reg, regulatory T cell; TNF- α , tumor necrosis factor- α . Figure created wi

CNS to the intestinal structures (153). The PNS, which includes the vagus nerve, provides both excitatory and inhibitory control over gastric, intestinal, and pancreatic functions (154). On the other hand, the SNS, predominantly inhibits gastrointestinal muscle and mucosal secretion and regulates blood flow through neural-dependent vasoconstriction. The ENS is the third and largest component of the ANS. The individual components of the MGB axis communicate bidirectionally within the ANS. Additionally, in combination with the HPA axis and neuroendocrine signaling, the ANS can induce CNS-modulated changes in the gut (155).

3.2.1.1 The enteric nervous system

The ENS is a mesh-like system of 200 to 600 million neurons embedded in the gastrointestinal system's lining, which facilitates communication between the brain and the GM (53). The ENS has several functions, including food propulsion, nutrient handling, blood flow regulation, and immunological defense (37, 153, 156). It is crucial in maintaining homeostasis and a stable gut microenvironment, in collaboration with the intestinal immune system, endocrine system, and the GM (53). Structurally, it is arranged into two ganglionated plexuses, the submucosal plexus (Meissner's plexus) and the myenteric plexus (Auerbach's Plexus), consisting of nitrergic (nitric oxide-dependent) and cholinergic (acetylcholine-dependent) enteric neurons (53).

The ENS can independently manage gastrointestinal function since it is equipped with intrinsic reflex microcircuits (157, 158). Moreover, the ENS produces more than 30 neurotransmitters, and the hormones and peptides it releases into circulation can cross the blood-brain barrier (BBB) and synergistically act with the vagus nerve (159). Neuropod cells, a recently discovered type of enteroendocrine cell, can transduce signals from the ENS to sensory neurons (160). There are many commonalities between the ENS and the CNS in terms of neurotransmitters, signaling pathways, and anatomical properties (53), which is why the ENS is referred to as the "little brain" (161). Although the ENS provides independent control over gastrointestinal function, the CNS provides extrinsic neural inputs that modulate, regulate and integrate these functions *via* the vagus nerve, thoracolumbar, and lumbosacral spinal cord (154).

Interactions between the GM and ENS have garnered significant attention in the past decade. Cooperative interactions between the ENS, GM, and intestinal immune and endocrine systems maintain host homeostasis (53). The GM can influence the development and function of the ENS directly and indirectly due to close proximity. For instance, early exposure to intestinal microbes is crucial for the postnatal development and organization of the ENS (53). Germ-free mice display abnormalities in ENS structure, such as reduced enteric neurons, with associated deficits in gut motility (53). They also exhibit attenuated intrinsic sensory signaling, defective influx of enteric glial cells into the intestinal mucosa, and altered neurochemistry (114, 162, 163).

Enterochromaffin cells of the ENS are another intermediary that facilitates communication with the GM (164). The biosynthesis of serotonin by enterochromaffin cells is promoted and enhanced by the GM, and is necessary for mucosal and platelet function (164). The GM can also produce neurotransmitters, such as serotonin, γ aminobutyric acid (GABA), histamine, catecholamines, and acetylcholine, further influencing ENS activity (18). Additionally, enteric neurons express toll-like receptors (TLRs), which recognize and respond to microbial molecules (e.g., LPS and PG) or viral RNA (54).

Recent studies have employed advanced technologies to investigate how the GM regulates neural programs by sensing cues from the environment and sending this information to the CNS (165, 166). For instance, a seminal study by Muller et al. used neuronal tracing techniques to demonstrate the modulation of neuronal pathways of the MGB axis by microbes (165). Specifically, the GM was shown to influence the functioning of enteric neurons through activation of aryl hydrocarbon receptors that regulate intestinal motility (166).

The ENS can influence the HPA axis through its regulation of gut hormones, neuropeptides, and cytokines, which can stimulate or inhibit HPA axis activity (155). Conversely, the HPA axis can affect the ENS by altering gut motility and secretion, as well as modulating the activity of enteric neurons and glial cells (21). This bidirectional communication suggests that the ENS and HPA axis are closely interconnected and play important roles in regulating stress responses and maintaining homeostasis.

3.2.1.2 The vagus nerve

The vagus nerve is the tenth cranial nerve that extends from its origin in the brainstem down to the visceral organs (159). It is a vital component of the PNS that connects the brain and gut to regulate homeostasis and cognitive areas function (21). The vagus nerve consists of both afferent and efferent neurons, making it the fastest and most direct pathway between the brain and gut. Evidence supports crucial roles in regulating inflammation, appetite, mood, and the stress response (167).

Some vagal endings synapse onto neurons from the ENS, and neuropod cells form fast excitatory synapses with vagal afferents using glutamate as a neurotransmitter (78, 168). This increase in the range of signals that can be transmitted by the vagus nerve enhances its ability to perform various functions. Vagal afferents express a plethora of receptors that detect various molecules such as nutrients, peptides, cytokines, hormones, and endotoxins (78). These sensory cues are transmitted rapidly to the nucleus tractus solitarius (NTS) of the brainstem (78), the primary projection site of gut-related vagal afferents in the brain (53).

Vagal brainstem nuclei then project to several regions of the brain, including nuclei involved in stress and cognition (167). For example, the hypothalamic PVN, an important hub for relaying signals from the vagus nerve, is involved in stress-induced gastrointestinal responses, including arousal, anxiety, and depression (169). Its projections to the pituitary and ventral tegmental area (VTA) provide the means to directly modulate the HPA axis and cognition, respectively. Further vagal projections to the arcuate nucleus integrate endocrine and behavioral aspects of gastrointestinal function, modulating food intake and satiety (167). Barrington's nucleus assimilates cognitive behaviors, while the locus coeruleus maintains arousal and attention, and integrates stress and cognitive inputs. In the forebrain, the amygdala integrates emotional and aversive inputs with learning and memory. The stria terminalis processes and consolidates emotions and behavior, and regulates the HPA axis and autonomic responses to stress. The cortex integrates affect, emotion, and memory with autonomic functions (167).

The gut is a vital control center for the immune system, and the vagus nerve displays immunomodulatory properties in the complex relationship between the gut, brain, and inflammation (170). In response to cytokines and endotoxins, the vagus nerve signals the brainstem, affecting fever and sickness behavior, as well as appetite and mood (171–174). The HPA axis reacts by providing a modulatory anti-inflammatory response (175–177). Moreover, the cholinergic anti-inflammatory pathway primarily signals through vagal efferents, serving as the effector limb of the "inflammatory reflex" and interfacing the nervous and immune systems (50, 170, 178–182) (Figure 1).

Animal studies have shown that disrupting the vagus nerve can lead to abnormalities in neurogenesis, stress reactivity, cognition, and anxiety- and fear-related behavior (183). On the other hand, stimulating the vagus nerve has been found to enhance memory (184, 185), facilitate hippocampal neurogenesis, increases expression of brain-derived neurotrophic factor (BDNF) (186, 187), and enhance synaptic plasticity (188). BDNF is an important plasticity-related protein that promotes neuronal growth, development, and survival, and plays a key role in learning, memory and mood regulation. Altered BDNF expression is associated with disruptions in cognitive function (189). These findings suggest that the vagus nerve promotes neurogenic and neurotrophic signaling. Indeed, vagal nerve stimulation is used to treat refractory epilepsy, Crohn's disease, refractory depression, chronic pain, and other conditions in humans (159).

This section highlights the role of the GM in regulating brain function through the vagus nerve. Animal and human studies have demonstrated that the vagus nerve serves as the primary and most direct signaling pathway between the GM and the brain (60, 78, 115, 190). The GM can activate vagal afferents directly or indirectly by releasing neuroactive mediators or by influencing the luminal concentration of molecules that vagal afferents detect (28, 78). Specific bacterial strains have been shown to influence vagus nerve signaling, to communicate with the brain, and alter cognition. For example, administration of Citrobacter rodentium, a pathogen, increased anxiety-like behaviors in mice, while Bifidobacterium longum (NC3001) produced anxiolytic effects in a vagus-dependent manner (191, 192). Similarly, Campylobacter jejuni administration resulted in increased levels of anxietyrelated behavior and activation of vagal afferents (178). Additionally, studies have shown that vagotomy prevented the positive effects of administration of a human milk oligosaccharide on long-term potentiation, learning, and memory in rats (193).

3.2.2 Neurotransmitters

Neurotransmitters provide additional communication mechanisms between the GM and nervous system (Figure 2). Microbes synthesize and metabolize several neurotransmitters, including dopamine, noradrenaline, serotonin, acetylcholine, histamine, and GABA (28). However, these neurotransmitters do not seem to cross the BBB and likely act indirectly to modulate brain function *via* the vagus nerve or ENS (78). Some neurotransmitter precursors synthesized in the gut may reach the CNS *via* the circulation and are able to cross the BBB *via* active transporters (194).

Animal studies provide evidence that microbial modulation of these neurotransmitters may impact host physiology, and preliminary human studies demonstrate that microbiota-based interventions can alter neurotransmitter concentrations (28). Germ-free mice studies have shown significant alterations in multiple neurotransmitter systems and their receptors in several brain regions (124). Similarly, antibiotic administration to deplete the GM can change the levels of neurotransmitters in the gut and blood (195, 196). Furthermore, microbial abundance has been shown to alter the expression of neurotransmitter receptors in the brain (51, 189, 191). Therefore, there is a growing body of evidence suggesting that the GM can ultimately influence the levels of neurotransmitters in the brain and alter brain function and cognition.

3.2.2.1 Tryptophan metabolism

Tryptophan is an essential amino acid. Its synthesis by microbes has been well described (197). In the gut, it may be further metabolized under direct or indirect control by the GM, giving rise to several compounds, such as serotonin, kynurenines, tryptamine, and indolic compounds, which participate in MGB communication (197, 198).

Although tryptophan is essential for serotonin synthesis, the dominant physiological pathway is the kynurenine pathway (Figure 3). Kynurenine is produced from tryptophan by the action of the hepatic enzyme, tryptophan-2,3-dioxygenase (TDO), or the ubiquitous indoleamine-2,3-dioxygenase (IDO) (199). Glucocorticoids and tryptophan induce TDO, while cytokines induce IDO (200). Kynurenine can cross the BBB, and is further metabolized along two separate arms to either kynurenic acid or quinolinic acid, and further, niacin and nicotinamide adenine dinucleotide (NAD⁺) (197). The balance between these two metabolites appears important in neural health and disease (31), as kynurenine pathway end-products are implicated in the regulation of biological processes involving neurotransmission, inflammation, and immunity (197). Moreover, kynurenic acid appears to exert mucosal protective and immunoregulatory effects in the gut (201). Activation of stress-responsive TDO or immuneresponsive IDO can limit the availability of tryptophan for serotonin synthesis and increase the downstream production of neurotoxic or neuroprotective metabolites (31).

In a recent study, the importance of the GM's ability to metabolize tryptophan into aryl hydrocarbon receptor ligands and, therefore, modulate gut inflammation, was demonstrated in patients with celiac disease (202). Several bacterial taxa can also affect tryptophan levels by direct utilization for growth or *via* tryptophanase expression (28, 203, 204), and these bacteria have been associated with the development of neuropsychiatric disorders, including autism spectrum disorders (205). Mounting evidence suggests that the GM modulates the tryptophan and kynurenine pathways and is a humoral route through which the GM may influence cognition at the level of the CNS (206–210).

3.2.2.2 Serotonin

Serotonin regulates pleiotropic physiological processes, including cognition, circadian rhythm, nociception, blood coagulation, cardiovascular homeostasis, and gastrointestinal secretion and peristalsis (211). Serotonin effects are mediated by the family of serotonin G-protein-coupled receptors (GPCRs) (212). Serotonin exerts both central and peripheral control. The vast majority of serotonin is found outside of the CNS, with 90-95% located in the gut, mostly within enterochromaffin cells (213, 214).

Serotonergic neurons have a significant influence on neuroendocrine function, and there is a dynamic interplay and extensive crosstalk between the serotonergic system and HPA axis (215). Serotonin has complex effects on the overall stress response, depending on the target cell and receptor type (216). Serotonin regulates upstream CRH signaling systems *via* the activation of serotonin 2C receptors of the hypothalamic PVN (217). Pregestational and early-life stress, with activation of the HPA axis,

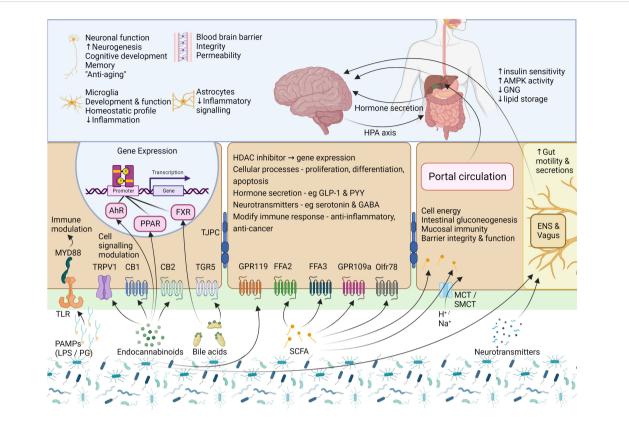


FIGURE 2

Signalling mechanisms – microbial products, metabolites, and neurotransmitters. Cells of the gut express a variety of receptors which are able to sense and transmit signals from the intestinal lumen and mucosa. To communicate, the GM uses factors which include several microbial products, eCBs, BAs, SCFAs, and neurotransmitters. PAMPs, such as LPS and PG, are small molecular microbial motifs that are recognized by TLRs, while this signal is transferred to intracellular signaling pathways (for e.g., immune cell activation) by MYD88. The eCB system is not limited to the activity of CB1 and CB2, and eCBs can also interact with other GPCRs, TRPV1, and the nuclear receptors PPAR-α and PPAR-γ. To modulate gut function, BAs interact with two main receptors, the GPCR named TGR5, and the nuclear receptor FXR. In the gut, SCFAs can activate FFA2, FFA3, GPR109a and Olfr78, but may also enter the cell via transporters or via passive diffusion where they modulate the activity of several enzymes and transcription factors or provide a source of energy for the cell. Small amounts of SCFAs are taken up into circulation where they may be transported to target tissues such as the liver, pancreas and brain. The binding of these GM-derived molecules with their respective receptors leads to the activation of cellular signaling pathways which then leads to alterations in cellular activity and gene expression, with downstream effects on host physiological processes. AhR, aryl hydrocarbon receptor; AMPK, AMP-activated protein kinase; BA, bile acid; CB1 and CB2,, cannabinoid receptor type 1 and 2; eCB, endocannabinoid; ENS, enteric nervous system; FFA2 and FFA3, free fatty acid receptor 2 and 3; FXR, farsenoid X receptor; GABA, γ aminobutyric acid; GLP-1, glucagon-like peptide 1; GNG, gluconeogenesis; GPR119 and GPR109a, G-protein coupled receptor 119 and 109a; HDAC, histone deacetylase; LPS, lipopolysaccharide; MCT, monocarboxylate transporter; MYD88, Myeloid differentiation primary response 88; Olfr78, Olfactory receptor 78; PAMP, Pathogen-associated molecular pattern; PG, peptidoglycan; PPAR α/γ , peroxisome proliferator-activated receptors α/γ ; PRRs, pattern recognition receptors; PYY, Peptide YY; SCFA, short chain fatty acid; SMCT, sodium-dependent monocarboxylate transporter; TGR5, Takeda G protein-coupled receptor 5; TJPC, tight junction protein complex; TLR, toll-like receptor; TPRV1, transient receptor potential cation channel subfamily V member 1. Figure created with BioRender.com.

have been linked with an altered serotonergic system, leading to interruptions in brain development and cognition (218–220). Indeed, the development of the serotonergic system within the MGB axis depends on a low-stress environment and early life events may be critical.

The GM can further regulate serotonin availability by signaling enterochromaffin cells to produce serotonin *via* expression of tryptophan hydroxylase, and by altering levels of SCFAs and BAs which can influence serotonin production (46, 164, 221–223). Germ-free mice exhibit reduced colonic serotonin production and decreased levels in the blood, which normalize with microbial colonization (164, 223). Moreover, male germ-free mice have increased hippocampal serotonin levels, which colonization immediately post-weaning does not reverse (124). When administered to rats, *B. infantis* results in reduced 5-HIAA (serotonin metabolite) concentrations in the frontal cortex, and a marked increase in blood concentrations of tryptophan and kynurenic acid (224). Thus, the GM can indirectly influence the activity of the serotonergic system, which in turn can affect the HPA axis and stress response.

3.2.2.3 Catecholamines

Catecholamines, such as dopamine and noradrenaline, regulate various body functions, including cognition, mood, and gut motility and integrity (225). Dopamine is a major neurotransmitter in reward-motivated behavior and is a precursor for other catecholamines, like noradrenaline and adrenaline. Noradrenaline is involved in arousal, alertness, sensory signal detection, behavior,

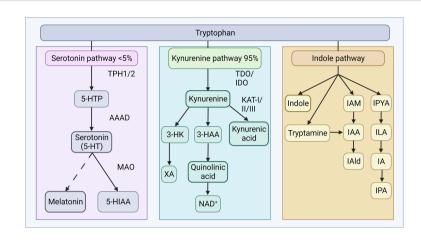


FIGURE 3

Tryptophan metabolism. Tryptophan metabolism occurs *via* the serotonin or kynurenine pathways to produce bioactive products. In the serotonin pathway, tryptophan is converted to 5-HTP by TPH1 in enterochromaffin cells, or TPH2 in neurons of the ENS or CNS. AAAD converts 5-HTP to serotonin, which can be further metabolized to melatonin, *via* a series of steps. The vast majority of tryptophan is, in fact, utilized in the kynurenine pathway, where tryptophan is converted to kynurenine by TDO in the liver (majority), or ubiquitously *via* IDO (including gut, brain, liver). Kynurenine can be converted to kynurenic acid by the KAT enzymes, quinolic acid and further NAD⁺, or XA. In the indole pathway, microbes of the gut metabolize tryptophan into indole and indole derivatives. 3-HAA, 3-hydroxyanthranilic acid; 3-HK, 3-hydroxykynurenine; 5-HIAA, 5-hydroxytryptophan; AAAD, aromatic amino acid decarboxylase; IA, anholocyclic acid; IAA, indole-3-acetia cid; 5-HT, 5-hydroxytryptamine; 5-HTP, 5-hydroxytryptophan; AAAD, aromatic amino acid decarboxylase; IA, anholocyclic acid; IAA, indole-3-acetic acid; IPA, indole-3-acetaldehyde; IAId, indole-3-aldehyde; IAM, indole-3-acetamide; IDO, indoleamine 2,3-dioxygenase; NAD⁺, nicotinamide adenine dinucleotide; TDO, tryptophan 2,3-dioxygenase; XA, xanthurenic acid. Figure created with BioRender.com.

cognition, and the acute stress response (226). It is well established that brainstem catecholaminergic centers play an important role in the regulation of the HPA axis (227) and noradrenergic neurons are required for normal activation in response to a variety of stressors. Noradrenaline, released within the intestinal wall during activation of the sympathetic nervous system in acute stress, has a wide variety of actions at the intestinal mucosa, such as modulating intestinal motility and transepithelial ion transport (228).

The GM also synthesize and respond to catecholamines. For example, bacteria produce noradrenaline as a quorum sensing molecule, and noradrenaline and adrenaline can promote pathogenesis and growth (229, 230) Germ-free mice show decreased noradrenaline in the cecal lumen and tissue, which is restored with colonization (231). Additionally, they have increased turnover rates of dopamine, noradrenaline, and serotonin in the brain (124).

3.2.2.4 γ -aminobutyric acid

 γ -aminobutyric acid (GABA) is an inhibitory neurotransmitter synthesized from glutamate by GABAergic neurons in the brain. GABA regulates various physiological processes and has been shown to play a central role in cognition by regulating and synchronizing neuronal signaling in the hippocampus (232). The HPA axis is also regulated by GABAergic signaling at the level of CRH, and nearly 50% of all synapses in the PVN are GABAergic (233). Although CRH neurons integrate information from many different brain regions involving several neurotransmitter systems, the activity of CRH neurons is ultimately regulated by GABAergic inhibition, mediated by GABA_A receptors (234). The HPA axis can also influence GABA production and signaling. Altered GABAergic profiles are associated with multiple diseases with cognitive dysfunction, such as dementia and depression.

The GM can metabolize GABA and recent research has shown that certain microbes can produce GABA, required for their growth (235). In addition, changes in GM composition have been associated with alterations in GABA receptor expression and GABA levels in the brain (191). Germ-free animals demonstrate decreased GABA concentrations in stool and blood, while fecal GABA levels can be modified with antibiotics. Remarkably, in a study of healthy women, levels of *Bacteroides*, identified as the major microbial producers of GABA, were associated with increased grey matter in the cerebellum, hippocampus, and frontal regions of the brain, as well as reduced levels of anxiety, distress, and irritability (236). GABA does not cross the BBB and so microbial-derived GABA would need to act locally on the ENS or vagus nerve to influence the CNS. However, SCFAs such as acetate can cross the BBB and be incorporated into the GABA metabolic cycle (237).

3.2.3 The endocannabinoid system

The endocannabinoid system (ECS) is a complex signaling system found throughout the body. The ECS is composed of endocannabinoids (eCBs), cannabinoid receptors, and enzymes involved in the synthesis and degradation of endocannabinoids. The two primary endocannabinoids are anandamide (AEA) and 2-arachidonoylglycerol (2-AG) (238). These bioactive lipid mediators are produced from the common phospholipid precursor arachidonic acid and released by various cell types in the body, including neurons, immune cells, and adipocytes (239). They bind high-affinity GPCRs, including cannabinoid receptors type 1 (CB1) and type 2 (CB2) (238). As neuromodulators, eCBs often act in

retrograde, released from postsynaptic cells and traveling backward across synapses, where they transiently inhibit the release of either inhibitory GABA or excitatory glutamate from presynaptic terminals (240) (Figure 4).

The ECS modulates a multitude of physiological processes, including the HPA axis (241), cognition, learning and memory (242), intestinal-barrier function (243), inflammation (244), energy metabolism (245), among others (reviewed recently (239)). In response to stress, eCB signaling modulates glucocorticoid and CRH signaling in the brain and is crucial in recovering homeostasis (241, 246, 247). The ECS is also widely expressed in neural tissue of the gut and is critically involved in the maintenance of intestinal homeostasis. It regulates barrier function and permeability through the immune system, epithelial tight junction proteins, and mucous secretion (248). Furthermore, it modulates myenteric neuron activity, SNS and vagal nerve function, and the release of neuropeptides such as ghrelin, leptin, and orexin (249).

The ECS and GM interact to regulate intestinal homeostasis resulting in relevant functional effects in the gut and CNS (238, 246, 248, 250). Dysbiosis affects eCB signaling, and vice versa (243). Germ-free animals demonstrate significant changes in the expression of CB1 and CB2, and synthetic and degradative enzymes throughout the gut (251). Vijay et al. studied the relationships between the ECS, inflammatory cytokines, and the GM using a six-week exercise intervention in humans (252). Changes in eCBs correlated with increased butyrate levels, and decreased TNF, IL-6 and IL-10. Hence, the anti-inflammatory effects of SCFAs may be partly mediated by the ECS. Healthy mice colonized with *Candida albicans* showed marked anxiety-like behavior and increased corticosterone concentrations that were inversely correlated with forebrain AEA, demonstrating disruption of the HPA axis through dysregulation of the ECS (253). Both animal and human studies have shown that the ECS and GM play a role in cognitive decline (254). Although microbes secrete eCBs, their role in host physiology remains unclear (255).

3.3 The immune system

The immune system's primary responsibility is to distinguish between "harmful" and "harmless" signals and respond appropriately. This is especially important in the gut, where immune cells are constantly in contact with microbes. The GM is therefore closely linked to the immune system, and they interact in several ways [reviewed recently (256, 257)]. Additionally, the immune system, HPA axis and CNS, and GM are closely interlinked.

One critical interaction involves the activation of pattern recognition receptors (PRRs), including TLRs, by microbial

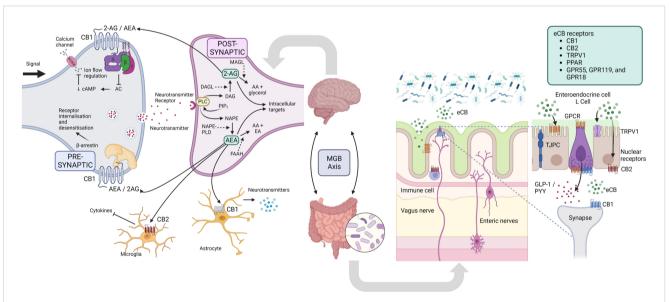


FIGURE 4

Endocannabinoid system. In the nervous system, presynaptic electrical impulses lead to calcium entry into the cell which drives the release of neurotransmitters into the synapse. Neurotransmitter receptors on the postsynaptic neurons are then activated and drive the action potential forward. The eCB system is a ubiquitous neuromodulatory system that functions throughout the body, including the nervous system to modulate cell signaling. DAG and NAPE are produced from phospholipid precursors, and are converted to the endocannabinoids (eCB) 2-AE and AEA by DAGL and NAPE-PLD, respectively. In retrograde signaling, these eCBs are mobilized from postsynaptic neurons and target presynaptic CB1 receptors to suppress neurotransmitter release by inhibiting AC, decreasing cAMP and therefore decreasing calcium ion flow into the cell, or alternatively influence receptor sensitivity and internalization *via* β-arrestin. eCB signaling in the CNS can also affect the functioning of microglia and astrocytes, with modulation of the release of cytokines and neurotransmitters, respectively. In the gut, eCBs secreted by certain microbes (or host cells) interact in microbiota-epithelial crosstalk, and include the immune and nervous systems, and metabolic, endocrine and barrier functions. 2-AG, 2-Arachidonoylglycerol; AA, arachidonic acid; AEA, N-arachidonoylethanolamine (aka anandamide); AC, adenylate cyclase; CNS, central nervous system; cAMP, cyclic AMP; DAG, diacylglycerol; DAGL, diacylglycerol lipase; EA, ethanolamine; FAAH, Fatty acid amide hydrolase; GLP-1, glucagon-like peptide 1; GPCR MAGL - monoacylglycerol; DAGL, diacylglycerol lipase; PLC, phospholipase C; PYY, Peptide YY; TPRV1, transient receptor potential cation channel subfamily V member 1. Figure created with BioRender.com.

associated molecular patterns (MAMPs) (54). These molecular signatures consist of microbial products such as the endotoxins LPS and polysaccharide A (on the cell surface of gram-negative bacteria) and PG (on gram-positive bacteria) (258). Each MAMP is detected by a specific PRR expressed on various cell types, including cells of the immune system (macrophages and natural killer cells) and nervous system (myenteric neurons and enteric glial cells) (53).

Cytokines and chemokines are another mechanism by which the immune system, GM, and CNS (including HPA axis) interact (259). Immune cells in the gut produce cytokines to maintain intestinal homeostasis, which in turn affects local microbial concentrations (260). Cytokines may also be produced locally in the CNS, or they may cross the BBB from the systemic circulation, and directly affect brain function.

Epithelial integrity is a crucial feature of gastrointestinal and nervous system homeostasis. It is essential to prevent the unregulated leakage of products across the barrier while allowing the transport of essential molecules. Furthermore, gut epithelial integrity is critical for maintaining the symbiotic relationship with the commensal microbes of the GM. This physical barrier includes the mucosa, epithelial cells, as well as tight junction proteins, such as occludin, claudins and zonula occludens (261). These tight junction protein complexes are dynamically modulated by intracellular signaling transduction systems and several extracellular stimuli, including cytokines, small GTPases, and post-translational modifications. When these regulatory mechanisms break down, barrier integrity may be compromised. Injury, infections and autoimmune diseases can influence the permeability of the gut and BBB (262). Microbes and microbial products then gain access to the circulation and may gain easier access to the CNS (262). Moreover, microbial products, cytokines, and other immune molecules released under the influence of the GM may further influence the BBB's integrity, alter BBB transport rates, and promote the release of neuroimmune molecules from the cells of the BBB (262). The GM can also alter BBB permeability by changing the expression of occludin and claudin 5 (263). These factors may lead to neuroinflammation, which is an important process shaping brain function.

Crosstalk between the GM and CNS is also essential for normal development and homeostatic functioning of the immune system, both innate and adaptive (256). While immune cell activation and cytokine production have a minor impact on the CNS during physiological perturbations, chronic systemic inflammation, mainly in the form of infections, has long been associated with behavioral alterations and cognitive dysfunction (264–266). Antibiotic-treated and germ-free mice have pro-inflammatory systemic and CNS immune system responses (120, 267). Perturbations in microbial diversity, secondary to antimicrobials, have been shown to influence pro-inflammatory cytokine secretion in the CNS and alter microglial morphology (268–270).

3.4 Microbial metabolites

In addition to the complex communication pathways between the GM and the host immune and nervous systems that have been described, there are several other mechanisms involving the production of small molecules that impact human function (271, 272).

3.4.1 Short chain fatty acids

Short chain fatty acids (SCFAs) are small organic monocarboxylic acids produced by bacterial fermentation of nondigestible polysaccharides in the large intestine. The main SCFAs are butyrate (C4), propionate (C3), and acetate (C2) (132). SCFAs are absorbed by colonocytes *via* monocarboxylate transporters (MCTs) or *via* non-ionic diffusion across the epithelium (131, 273).

SCFAs are a source of energy and trophic factors for cells of the colon and liver (274). Additionally, they can bind GPCRs, specifically the free-fatty-acid receptors FFA2, FFA3, Olfr78, and GPR109a, located throughout the body, including enteroendocrine, immune, and neural cells (275-278). This suggests that SCFAs play a key role in neuro-immuno-endocrine regulation (279-282) (Figure 4). Indeed, extensive evidence supports pleiotropic roles of SCFAs, which affect several host organs and systems, including the gut and CNS (127, 131, 132). SCFAs have several local effects that improve intestinal health, including the maintenance of intestinal barrier integrity, mucus production, and protection against inflammation (250). These processes are crucial to the gut's first line of defense. SCFAs promote immunity and suppress inflammatory responses in the intestine and other organs by regulating immune cells such as lymphoid cells, T cells, and B cells (283-285).

By inhibiting histone deacetylase (HDAC) activity, SCFAs also regulate systemic functions, promoting histone acetylation and gene expression in host cells (250). This epigenetic mechanism has been described in gastrointestinal, immune and neurological cells [reviewed (131)].

SCFAs appear to play a significant role in MGB communication (286). Research indicates that SCFAs can indirectly modulate the PNS through expression of FFA3 in the enteric neural plexus, portal nerve, and autonomic and sensory ganglia (131). Activation of FFA3 receptors on vagus nerve cells can result in the activation of various neurons in the CNS, including dynamic regulation of hypothalamic neuronal circuitry (287).

SCFA-induced activation of receptors on enteroendocrine cells can promote gut-brain signaling by inducing hormones such as glucagon-like peptide 1 (GLP1) and peptide YY (PYY), as well as neurotransmitters like GABA and serotonin (276). SCFA-signaling can also induce other hormones, including leptin from adipocytes, and insulin from pancreatic β -cells (288). Additionally, SCFAs can modulate the levels of neurotransmitters and neurotrophic factors and regulate the expression of tryptophan 5-hydroxylase, the enzyme involved in the synthesis of serotonin, and tyrosine hydroxylase, the enzyme involved in the rate-limiting step in dopamine, noradrenaline, and adrenaline synthesis (18, 132, 164, 222).

Moreover, the abundant expression of MCTs on endothelial cells suggests that SCFAs can cross the BBB, which is supported by the presence of SCFAs in human cerebrospinal fluid (CSF) and in brain uptake studies (286). Accumulating evidence supports the idea that SCFAs are necessary for the maintenance of CNS homeostasis, learning and cognition, and reward-associated

behaviors (25). SCFAs also influence the integrity of the BBB by upregulating the expression of tight junction proteins (263).

SCFAs may also improve neuronal homeostasis and function by influencing neurotrophic factors such as nerve growth factor, glial cell line-derived neurotrophic factor, and BDNF (119–122). These factors regulate the growth, survival, and differentiation of neurons and synapses in the CNS, and are important for learning and memory. SCFAs can modify neuroinflammation by affecting the morphology and function of glial cells (120, 286, 289, 290). By administering SCFAs to germ-free mice, Erny et al. were able to rescue deficits in microglial immaturity and morphology (120).

There is also evidence to suggest that SCFAs can modulate the HPA axis. In stressed mice, SCFA administration reduced HPA axis hyperactivity and intestinal permeability (291). In humans, a recent triple-blind, randomized, placebo-controlled intervention trial examined the effects of colonic SCFA-mixture delivery in men on responses to psychosocial stress and fear tasks (292). SCFA supplementation was shown to downregulate the HPA axis by significantly attenuating the cortisol response.

Altered SCFA production has also been demonstrated in a variety of neuropathologies (42, 116, 127, 130, 132, 292–295). These findings suggest that SCFAs regulate CNS processes through both direct and indirect mechanisms and may ultimately affect host cognition and response to stress.

3.4.2 Bile acids

Bile acids (BAs) are products of cholesterol metabolism primarily produced in the liver as primary BAs and modified by the GM into secondary BAs through processes such as deconjugation, dihydroxylation, dehydrogenation, and isomerization (Figure 5) (30, 296, 297). While their role in enterohepatic circulation as detergents for lipid digestion is well established, recent studies have also revealed their function as hormones via receptors such as farnesoid X receptor (FXR) and Takeda G protein-coupled receptor 5 (TGR5), with significant regulatory and signaling activities (298). BAs can also activate pregnane X receptors, vitamin D receptors, and glucocorticoid receptors (299). Their functions encompass regulation of motor, sensory, and secretory functions of the gut, intestinal barrier permeability, inflammatory response, and several metabolic processes, including lipid and glucose metabolism, and hepatic gluconeogenesis (300).

The effects of BAs extend beyond the gut, impacting various tissues throughout the host. BA receptors are present in the brain, and BAs can either be synthesized locally or actively transported across the BBB by BA transporters from circulation (297, 301, 302). Consequently, circulating BA levels significantly influence the CNS's BA profile (303). FXR knockout mice exhibit abnormal BA and neurotransmitter concentrations, resulting in impaired cognition and motor coordination (304). TGR5, expressed in brain and peripheral neurons, as well as glial and microglial cells, can be activated by several neurosteroids (305). Specific BAs demonstrate neuroprotective effects in cellular and animal models, with human clinical trials underway (306–308).

BAs play a role in regulating the HPA axis. BAs modulate HPA axis activity by inhibiting CRH release through FXR activation, expressed in the hypothalamus (309). Additionally, BAs can interact with TGR5, expressed in the hypothalamic PVN, stimulating the HPA axis by increasing CRH (310, 311). Cholestasis, associated with suppression of the HPA axis, likely due to BA interactions with glucocorticoid receptors in the brain (299, 312). The discovery of FXR and TGR5 receptors in the adrenal gland further connects BAs with glucocorticoid metabolism (311, 313-315). BAs might act via TGR5 in a cAMP/protein kinase A (PKA)-dependent fashion phosphorylating and thus activating steroidogenic acute regulatory protein (StAR) and hormone sensitive lipase (HSL) (316). FXR activation is known to regulate lipoprotein receptors and transporters, as well as enzymes in the steroidogenic pathway, and has been shown to increase corticosterone levels in mice (311).

The GM influences BA metabolism, and BAs affect the GM's composition (296). Specific microbes directly contribute to BA transformation, impacting the BA pool's composition and size (317). Some BAs serve as substrates for gut microbes, while others exhibit antimicrobial properties, actively shaping the GM at the highest taxonomic levels (318). The BA-microbiota axis modulates the immunoregulatory environment along the gut (303). Given this close bidirectional relationship between BAs and the GM, these metabolites have emerged as important modulators of the MGB axis, functioning directly *via* BA receptors in the ENS and brain or indirectly *via* GLP-1 or the FXR-FGF15/19 axis (297, 319). Changes in the GM's composition correspond to changes in blood and brain BA profiles, which are essential because specific BA ligands' distinct physicochemical properties determine the potency of BA receptor activation (320–323).

Altered BA profiles have been observed in several neuropathologies associated with cognitive decline, such as Alzheimer's disease (302, 324). BA disorders are also associated with neural symptoms (300). BAs may substantially affect cognitive function by their affinity to muscarinic receptors, as well as GABA and NMDA receptors (325). Germ-free mice excrete less fecal BAs, have a larger BA pool, and have different gene expression profiles involved in BA metabolism, than wild-type mice (323). Postnatal maturation of the GM in newborn mice was shown to be dependent on BAs and neonatal cholestasis is associated with dysbiosis in infants (326, 327). Bile duct ligation alters the GM composition, and increases the permeability of the BBB (328, 329). In a study of patients with depression and anxiety, BA profiles associated with altered GM composition were significantly different in those with more severe symptoms, and specific BA parameters were able to distinguish treatment failures from remitters (330).

3.4.3 Branched-chain amino acids

Branched-chain amino acids (BCAAs) are essential amino acids including leucine, isoleucine, and valine, which participate in various biochemical functions, including energy production, protein synthesis, insulin secretion, brain amino acid uptake, and immunity (331). The GM produces higher proportions of specific

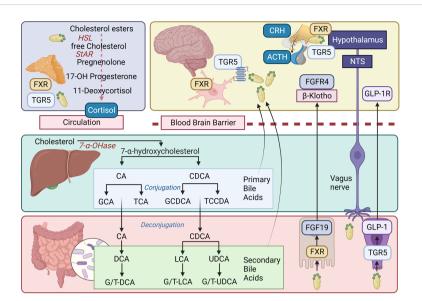


FIGURE 5

Bile acids, BA receptors, and signaling pathways. In the liver, the classical pathway of bile acid (BA) synthesis begins with the conversion of cholesterol into 7α -hydroxycholesterol by the rate-limiting enzyme cholesterol 7α -hydroxylase (7α -OHase; CYP7A1). The 7α -hydroxycholesterol is then further metabolized into cholic acid (CA) and chenodeoxycholic acid (CDCA) through a series of enzymatic reactions. Once synthesized, BAs are conjugated with either glycine or taurine, which increases their solubility and reduces their toxicity. The conjugated BAs are then secreted into bile canaliculi, stored in the gallbladder, and released into the small intestine following a meal. After completing their role, approximately 95% of BAs are reabsorbed in the ileum and transported back to the liver via the enterohepatic circulation. As BAs pass through the gastrointestinal tract, they encounter a diverse population of gut bacteria and the synthesis of secondary BAs occurs in the large intestine as a result of microbial biotransformation. Secondary BAs are important for maintaining the overall BA pool in the body and contribute to the regulation of cholesterol homeostasis, energy metabolism, and the immune system. BAs can also act as signaling molecules, interacting with specific receptors such as the nuclear receptor FXR and the cell membrane receptor TGR5 (expressed in various tissues, including the liver, gut, enteric nervous system, CNS, and adrenal glands) which are involved in the modulation of numerous physiological processes, including glucose metabolism, lipid metabolism, and the regulation of the gut-brain axis. In the gastrointestinal tract, BAs bind FXR in enterocytes and this activates the expression of FGF19, which is then secreted into the bloodstream and plays a crucial role in MGB communication. FGF19 acts as an endocrine signal crossing the BBB to reach the CNS and then binding to its cognate receptor, FGFR4, and co-receptor β-Klotho. This interaction leads to the activation of intracellular signaling cascades, such as the MAPK pathway and the PI3K/Akt pathway. These signaling pathways regulate various processes, including cell growth, differentiation, and metabolism, and contribute to the modulation of the gut-brain axis. Additionally, activation of TGR5 by BAs can lead to the release of GLP-1, an incretin hormone that modulates insulin secretion and glucose homeostasis. In the CNS, TGR5 activation has been implicated in the regulation of energy balance, neuroinflammation, and neuroprotection. BAs can influence the HPA axis through both direct and indirect mechanisms involving signaling pathways in the CNS and the adrenal glands. In the CNS, BAs can modulate the HPA axis by interacting with FXR and TGR5, which are expressed in various brain regions, including the hypothalamus and the hippocampus. Activation of these receptors by BAs can influence the release of CRH from the hypothalamus and ACTH from the pituitary gland, leading to the modulation of cortisol secretion from the adrenal cortex. Furthermore, BAs can directly affect the adrenal glands, influencing the release of cortisol. BA can alter adrenal steroidogenesis by modulating the expression and activity of key enzymes involved in the biosynthesis of cortisol, including HSL, StAR, and cytochrome P450 enzymes (e.g., CYP11A1, CYP11B1, and CYP11B2). Additionally, BAs can influence adrenal cell function by activating FXR and TGR5, which may regulate intracellular signaling pathways and gene expression patterns related to steroid hormone production, inflammation, and oxidative stress. Primary bile acids: CA, cholic acid; CDCA, chenodeoxycholic acid; GCA, glycocholic acid; TCA, taurocholic acid; GCDCA, glycochonedeoxycholic acid; TCCDA, taurochenodeoxycholic acid. Secondary bile acids: DCA, deoxycholic acid; G/T-DCA, glyco/tauro-deoxycholic acid; G/T-LCA, glyco/taurolithocholic acid; G/T-UDCA, glyco/tauro-ursodeoxycholic acid; UDCA, ursodeoxycholic acid; LCA, lithocholic acid; UDCA, ursodeoxycholic acid. ACTH, adrenocorticotropic hormone; Akt, protein kinase B; BA, bile acid; BBB, blood brain barrier; CNS, central nervous system; CRH, corticotrophin-releasing hormone; FGF19, fibroblast growth factor 19; FGFR1-4, fibroblast growth factor receptors 1 to 4; FXR, farnesoid X receptor; GLP-1, glucagon-like peptide 1; GLP-1R, glucagon-like peptide 1 receptor; HSL, hormone sensitive lipase; MAPK, mitogen-activated protein kinase; MGB, microbiota-gut-brain; PI3K, phosphatidylinositol 3-kinase; StAR, steroidogenic acute regulatory protein; TGR5, Takeda G protein-coupled receptor 5. Figure created with BioRender.com

BCAAs (valerate, isobutyrate, and isovalerate) relative to other amino acids, which have been shown to influence epithelial and mucosal homeostasis (332). Additionally, BCAAs can be utilized by microbes, potentially regulating intestinal microbial species, diversity, and metabolism (333, 334).

BCAAs regulate key signaling pathways, most notably the activation of mechanistic target of rapamycin (mTOR), which serves as the master regulator of cell growth and proliferation (331). Mice supplemented with a BCAA-enriched cocktail exhibited improved physical endurance and an extended lifespan (335).

In the CNS, BCAAs play roles in protein synthesis, food intake regulation, and serve as nitrogen donors involved in intercellular shuttling and the synthesis of the neurotransmitters glutamate and GABA (both modulate the HPA axis) (336). Mice deprived of leucine had increased HPA axis activation *via* CRH expression (337). Excessive BCAA concentrations are considered toxic and can cause tissue damage, particularly in the CNS (338). Although exploratory studies remain in their infancy, evidence suggests that BCAA modulation may be useful in cognition disorders (339–341). Further research is required to determine the relationship between the GM, BCAAs, HPA axis, and cognition.

4 The relationship between the microbiota, HPA axis, and cognition

4.1 Stress-related interactions between the HPA axis and gut microbiota

4.1.1 Evidence from animal studies

The microbiota and HPA axis develop rapidly and profoundly in the first years of life, and environmental stressors can affect both (146–148). Stress experienced during different periods of life can have varying physiological consequences. Early life stressors, and *in utero* stressors, can impact the development and function of the HPA axis (149). Stress during pregnancy disrupts the vertical transmission of microbes from mother to offspring, leading to alterations in the maternal microbiota, which are then transferred to the offspring (342). Jašarević et al. demonstrated changes in the microbiome of these offspring, as well as alterations in the metabolome of the gut and brain (343). In a subsequent study, it was shown that FMT from stressed dams into stress-naïve germfree mice was sufficient to instill the phenotype observed in stressexposed offspring (344).

Early life stress, such as maternal separation, activates the HPA axis with associated changes to the developing microbiota, ultimately leading to an imbalance in the GM and an inappropriate stress response (18). Several studies have demonstrated that neonatal stress can lead to short- and longterm alterations in the diversity and composition of the GM (345-347). Interestingly, these consequences appear to be age-dependent. In response to early life stress, younger rats had increased neurogenesis, decreased BDNF IV promoter histone methylation, with a complementary increase in hippocampal BDNF concentration, and associated improvements in spatial and nonspatial learning (348). In contrast, middle-aged rats demonstrated opposing changes, concomitant with impairments in hippocampaldependent cognitive tasks. These discordant results illustrate the biphasic consequences of early life stress and indicate a role for epigenetic modification of BDNF expression. Furthermore, chronic antidepressant treatment post-exposure was able to rescue the neurological decline observed in the middle-aged rats (348).

A landmark paper by Sudo et al. provided evidence for the essential role of the GM in programming the stress response by illustrating the differences in HPA axis hormones and receptors in germ-free mice when compared to specific pathogen free (SPF) mice (51). Germ-free mice had increased acetylcholine, ACTH, and corticosterone responses following acute stress, indicative of enhanced HPA axis activity. Moreover, these animals showed decreased expression of NMDA receptor subunits (NR-1) in the cortex, and of NR-2a in the cortex and hippocampus, while BDNF levels were lower in the cortex and hippocampus. Following chronic restraint stress, germ-free mice showed significantly greater HPA axis activity, whereas the SPF mice exhibited more anxiety-like behaviors under the same stress (349). These findings have subsequently been reproduced, with both male and female mice demonstrating enhanced stress reactivity to a novel environmental stressors (189, 350, 351). Indeed, germ-free animals show widespread neurodevelopmental changes, associated with alterations in monoaminergic neurotransmission in the CNS (189).

Animal models of chronic stress demonstrate altered intestinal physiology and GM composition, with an increased secretory state and permeability (352-355). Alterations in gut barrier integrity enable bacteria (and microbial-products) to translocate across the mucosa and epithelium, interfacing with immune and neuronal cells (356, 357). Mounting evidence suggests that chronic interactions can lead to systemic, low-grade inflammation contributing to the development of autoimmune, metabolic, and cognitive disorders (358). Moreover, exposure to chronic stress and the subsequent disruption of GM stability has been shown to increase host susceptibility to infection. Mice exposed to prolonged restraint stress demonstrated changes in GM composition, such as bacterial overgrowth and reductions in diversity and richness (359). When challenged orally with the enteric murine pathogen C. rodentium, chronically stressed mice had an increased pathogen load and increased colonic TNF- α expression. Probiotics rescued these changes in the GM and the associated host-microbe interactions. A study by Allen et al. demonstrated that the GM is necessary for stress-induced immunomodulation, with enhancement of splenic macrophage reactivity occurring in colonized controls but not in germ-free mice, in response to social disruption stress (360).

The impact of the GM on the HPA axis can further be interrogated following deliberate interventions. Chronic antibiotic treatment led to a decrease in CRH receptor mRNA levels in the brains of rats (361). The introduction of pathogenic bacteria reduced cognitive abilities and heightened anxiety-like behaviors (362, 363). Exposure of neonatal animals to low-dose endotoxins resulted in the activation of TLRs (364, 365). In addition, they showed long-term HPA axis alterations in activity, as evidenced by increased mean glucocorticoid concentrations resulting from an increase in glucocorticoid pulse frequency and amplitude. In an animal model of diet-induced obesity, anxious and depressive-like behaviors were associated with decreased hippocampal levels of glucocorticoid receptors and an exaggerated HPA axis-mediated stress response to acute physical and social stress (366). The probiotic B. pseudocatenulatum (CECT 7765) reversed the glucocorticoid receptor and stress response abnormalities, along with the neurobehavioral phenotype. Probiotic treatment with Lactobacillus sp. concurrent with early life maternal separation stress could normalize HPA activity (346). Similarly, pre-treatment with L. farciminis reduced HPA hyper-reactivity, intestinal permeability, and neuroinflammation resulting from restraint stress (367). When L. rhamnosus was administered to mice, region-dependent alterations in GABA receptor expression in the brain were found to parallel the reduction in stress-induced glucocorticoid levels (191). Notably, the neurochemical effects were not found in vagotomised mice, implicating the vagal pathway. Recently, the GM was shown to influence the expression of genes encoding proteins that participate in the HPA axis and the peripheral metabolism of glucocorticoids (368). A study by Mudd et al. reported a predictive relationship between levels of fecal Ruminococcus, serum cortisol, and brain Nacetylaspartate in young pigs (126).

4.1.2 Evidence from human studies

Although there remains a dearth of evidence from human studies, a recent pilot study of 34 healthy infants found that GM composition at one month (measured as alpha diversity) was positively associated with HPA axis reactivity following a painful stressor (369). In a larger cohort of 193 babies (aged 2.5 months), the cortisol stress response was weakly associated with alpha diversity (370). In healthy adults, the experimental administration of LPS in a randomized control trial (RCT) caused a transient physiological stress response, with dose-related increases in cortisol, noradrenaline, body temperature, pulse rate, and cytokines (371). This stress response was associated with increased anxiety and depressed mood. Alterations in cognition occurred both in the short- and long-term, confirming mechanisms for both the promotion and inhibition of cognitive performance during acute inflammatory stress.

In another RCT, a combination probiotic (L. helveticus (R0052) and B. longum (R0175)) administered to healthy volunteers was associated with beneficial psychological effects in participants and a decrease in 24-hour urinary cortisol, suggesting attenuation of the HPA axis in response to stressors (372). A recent meta-analysis of RCTs focusing on the efficacy of probiotics on stress in healthy individuals showed that probiotic use generally reduced subjective stress levels and appeared to alleviate stress-related sub-threshold anxiety and depression (373). However, cortisol levels were not significantly altered. In a small but detailed four-week study investigating the role of dietary fibre and fermented foods on the GM profile and function, including stress and overall health, subtle GM composition changes were associated with significant changes in several faecal lipids and urinary tryptophan metabolites (374). Participants reported reductions in perceived stress, but these were not significantly different to controls, and markers of stress were unaffected. However, the reduction in perceived stress was dosedependent, with higher dietary adherence resulting in larger reductions in stress.

4.2 HPA axis-related interactions between the gut microbiota and cognition

4.2.1 Cognition

Cognition is defined as the complex mental process of acquiring, understanding, and storing information through thought, experience, and the senses. Essentially, it is the ability to perceive and react, process and understand, store and retrieve, and make decisions and produce appropriate responses. Cognition is not a singular concept, and various 'domains' (functions) with several components have been identified. Cognitive dysfunction typically manifests as impairment in one or more aspects of memory, language, visuospatial, execution, computation, understanding, or judgment, amongst others. An increasing body of preclinical and human evidence demonstrates that the MGB axis plays important roles in the development and maintenance of various components of cognition (191, 362, 375).

4.2.2 Evidence from animal studies

Studies in germ-free mice have shown that, in the absence of microbes, the brain is markedly affected, exhibiting deficits in learning, memory formation and recognition, and social and emotional behaviors (58, 123, 124, 362). Gareau et al. demonstrated impaired short-term recognition and working memory in 5 to 6-week-old germ-free mice when compared to conventionally reared counterparts (362). Behavioral differences in germ-free mice include their anxiolytic-like manner when compared to SPF controls, although some studies report increased anxiety-like behavior in different species (124, 189, 351, 376). Microbial colonization has been shown to rescue these elements of cognition, but only if administered during early stages of life (124, 350). Germ-free mice present with social cognitive deficits, which may be associated with biochemical changes, such as decreased hippocampal BDNF and c-FOS expression, important in memory (124, 362, 376, 377). Further studies have demonstrated that reduced cognitive function is inversely associated with BDNF mRNA levels (124, 350).

Several metabolomic studies show alterations in the dopamine and serotonin systems (124, 189, 350, 378–380). These changes include increased hippocampal dopamine D1 receptor mRNA levels and decreased dopamine D1 receptor in the striatum and nucleus accumbens, as well as increased serotonin in the blood and hippocampus, with decreased serotonin receptor expression, respectively.

The role of sexual dimorphism must be noted, as several studies have documented sex-specific differences in the serotonergic system of germ-free animals (350). Female offspring displayed increased cognitive deficits and anxiety-like behavior following prenatal stress, which were associated with GM alterations in the pregnant females and increased IL-1 β and decreased BDNF levels *in utero* (381). In contrast, male offspring presented with deficits in social cognition only, without disturbances in the other behavior or cognitive parameters measured (146).

The use of broad-spectrum antibiotics provides another modality to study the cognitive deficits induced by dysbiosis. Administration has been shown to alter tryptophan metabolism and BDNF, NMDA receptor subunit 2B, serotonin transporter, neuropeptide Y, oxytocin, noradrenaline, and vasopressin expression (382, 383). Möhle et al. administered long-term antibiotics to adult mice, resulting in a decrease in hippocampal neurogenesis and memory retention (384). Confirming a role for the GM, these deficits were reversed by a combination of gut flora reconstitution with probiotics and voluntary exercise. The function of the GM was further demonstrated in a recent study comparing antibiotic-treated and germ-free mice with regards to fear extinction learning (385). Without a complex microbiota, both types of mice exhibited altered fear-associated behavior, changes in gene expression in brain cells, and alterations in the firing patterns and rewiring ability of neurons. Selective colonization revealed a critical developmental period, indicating that microbiota-derived signals were required during early post-natal and adult life for normal learning. Given the well-established

relationship between glucocorticoids and memory formation, studies like this suggest a complex relationship between the GM, HPA axis and cognitive processes (386).

Animal models can be further used to explore the role of prebiotics and probiotics on cognition and behavior, including changes in depression, anxiety, and stress associated with changes in immune markers, hippocampal synaptic efficacy, and tryptophan metabolism (376, 387). Administering Mycobacterium vaccae, a transient commensal microbe, as a probiotic or vaccine to young adult mice improved behavior, learning, and memory during cognitive testing, confirming the role of the immune and serotonergic systems in MGB pathways (388, 389). The probiotic B. longum (NCC3001) demonstrated vagal pathway-mediated anxiolytic effects in mice and B. longum (1714) showed improved learning and memory after 11 weeks of supplementation (192, 390). Supplementation with various Lactobacillus species resulted in improvements in cognitive abilities and social deficits and these changes were shown to occur alongside altered GABA expression in the brain, as well as significant changes in the oxytocin and vagus nerve pathways (191, 391). Six weeks of administration with the probiotic Clostridium butyricum restored cognitive function in a mouse model for vascular dementia, which was associated with increased butyrate levels in fecal and brain samples, and activation of the hippocampal BDNF-PI3K/Akt pathway (392).

Infection studies are another useful modality. Administration of *C. rodentium*, combined with acute stress, led to memory dysfunction in young adult germ-free mice (362). This deficit was prevented by daily administration of a *Lactobacillus* sp. probiotic combination before infection. Humann et al. showed that microbial PGs can traverse the mouse placenta, such that maternal administration of PGs could be detected in the fetal brain (64). The offspring exhibited decreased cognitive function related to TLR-2-mediated neuroproliferation *via* FoxG1 induction in the cortex. Neonatal LPS exposure leads to similar cognitive deficits (393). Bilbo et al. revealed that neonatal *E. coli* infection could impair memory in adult rats (394). Intriguingly, the deficit was only observed if an LPS challenge was administered at the time of learning and could be prevented by daily handling of the neonatal rats, which significantly altered their basal HPA axis activity (395).

Animal and human studies indicate an association between prenatal bacterial or viral exposure and the subsequent development of various disorders. Mid-gestation injection of maternal mice with low-dose immunostimulatory polyinosinic: polycytidylic acid (poly I:C) to mimic viral infection significantly impaired non-spatial memory, learning, and motor activity in the offspring at 3 and 9 weeks of age (396). Infection and/or the immune response to mimetics of infectious agents potentially have long-lasting effects on the cognitive abilities of offspring. In humans, the consequences of maternal infection for the microbiota of the offspring, and related cognitive abnormalities, remain unclear and warrant further investigation. These data highlight the interrelatedness of the microbes, the HPA axis, and cognition.

4.2.3 Evidence from human studies 4.2.3.1 Microbial colonization and microbiota development

Previously, cognition was thought to be regulated solely by the CNS. However, it has become clear that many other non-CNS factors, including the GM, also regulate and influence cognitive function (397). Indeed, it has been postulated that the development of higher cognition in humans may not have evolved in the absence of microbes (398). Considerable attention has recently been paid to unravelling the mechanisms by which the GM, human brain and cognition interrelate (Figure 6). Carlson et al. assessed microbiota composition in three groups of one-year-olds, characterized by global and regional brain volume using magnetic resonance imaging (MRI), and cognitive outcome tests, using cluster analysis (399). The group with the greatest abundance of Bacteroides outperformed the other two groups using the Mullen Scales of Early Learning assessment. They were also less likely to have been delivered by Caesarean-section. Other studies have also linked delivery mode with neurocognitive development and longterm poor immune and metabolic health outcomes, highlighting the importance of microbial colonization at birth (400, 401). New research suggests that the HPA axis may also be important for linking delivery by C-section to poor health outcomes later in life. In a longitudinal study of 136 infants, those delivered by C-section had lower cortisol concentrations at baseline and in response to a painful stress test at six-month follow-up, signifying an altered HPA axis (402). In a cohort of more than 7,000 neonates, Kiilerich et al. found that levels of inflammatory and stress markers were lower, and growth factor levels higher, in infants delivered via pre-labor Csection when compared to those delivered vaginally (403). Interestingly, these differences were not significant if C-section was performed during labor, suggesting that the labor process itself initiates important endocrine, physiological, and biochemical processes relevant to the neonatal immune system and stress response.

4.2.3.2 Antibiotic use

It is well established that the use of antibiotics is critical and necessary when warranted. However, inappropriate use is associated with several negative health outcomes, including the emergence of antimicrobial resistance. Certainly, the MGB axis has multiple sites for off-target activity, which may lead to positive or negative neurocognitive effects downstream, emphasizing the importance of antibiotic stewardship (404–407). Antimicrobial administration in high doses or for long periods can induce severe or irreversible alterations at both the intestinal and brain level (408, 409).

The infant microbiota is highly sensitive to various perturbations like stress and antimicrobials (408). Studies show that perinatal exposure to the various classes of antibiotics substantially alters the establishment of the neonate and infant microbiota, and antimicrobial prophylaxis for C-section delivery is

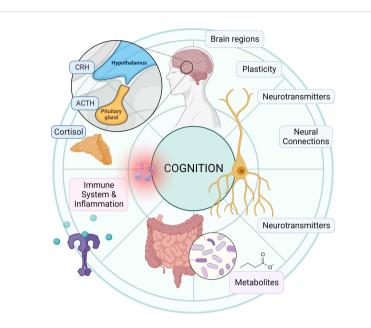


FIGURE 6

The gut microbiota, HPA axis and cognition. Schematic summary representation of the relationship between the gut microbiota, HPA axis, and cognition. The gut microbiota influences the HPA axis and cognition through the production of metabolites (e.g., SCFAs, and bile acids), neurotransmitters (e.g., serotonin, GABA, catecholamines), and immune system modulation. The HPA axis, comprising the hypothalamus, pituitary gland, and adrenal glands, regulates cortisol release, which in turn affects both gut microbiota and cognitive function. Cognitive processes involve various brain regions (e.g., hippocampus, amygdala, and prefrontal cortex), neurotransmitters, and plasticity, and are modulated by the interplay between the gut microbiota and HPA axis. ACTH, adrenocorticotropic hormone; CRH, corticotrophin-releasing hormone. Figure created with BioRender.com.

routine. In a RCT of 40 mother/infant pairs in which prophylactic antibiotics were administered before versus after clamping of the cord, the timing of the exposure was found to be critical for microbiota development (410). In a study of over 800 children, antibiotic use in the first year of life was associated with subsequent diminished neurocognitive outcomes (411). Indeed, behavioral difficulties associated with GM composition may persist to at least ten years of age (412). Early-life antibiotic exposure, therefore, appears to disrupt microbiota colonization and maturation, resulting in adverse child health outcomes (413).

Not only infants and children are at risk. In a large prospective cohort of 14,542 participants, chronic antibiotic use in midlife was associated with cognitive impairment later in life, such that antibiotic use was associated with an additional three years of aging, compared to those with no antibiotic exposure (407).

On the other hand, in certain conditions antibiotics may be useful in the treatment of symptoms related to a dysbiotic GM. For example, it is well established that the cognitive impairment arising due to hepatic encephalopathy is mediated by microbial disturbances and can be reversed with oral antibiotic treatment (414–417). Administration of non-absorbable antibiotics in patients with hepatic encephalopathy and mild cognitive impairment improves cognitive performance, subcortical brain activity, fronto-parietal connectivity, and serum metabolomic profile (418, 419).

4.2.3.3 Exogenous glucocorticoids and other drugs

While antibiotics are well known to alter GM composition and function, there are several other types of drugs that influence the GM

and may place an individual at risk for cognitive dysfunction (420, 421). Exogenous glucocorticoids are frequently prescribed to treat a multitude of disorders, because of their ability to suppress the immune system and decrease inflammation (422). While we have a cursory understanding of their impact on the GM, and their short- and long-term cognitive effects, a more thorough understanding is critically needed. Several studies have highlighted the deleterious effects of these drugs on cognition (423–428). Both endogenous and exogenous glucocorticoid excess on MR and GR expressed by neurons in the CNS have been shown directly alter the structure and functioning of the brain (429–431). Furthermore, glucocorticoids may modulate the brain indirectly through their effects on the immune system, metabolism, sleep, other hormones, and the GM (432, 433).

Regarding the effects of glucocorticoids on the composition and functioning of the microbiota, some studies have indicated microbiota disruption, while others have shown beneficial effects, which likely depend on the degree of exposure (dose and period) and the type of glucocorticoid studied (434-437) Furthermore, the use of exogenous corticosteroids is not always associated with deleterious effects on cognition, and some studies in animals and humans have failed to demonstrate an association, while some provide evidence of corticosteroid-induced cognitive enhancement (438-443). It appears that the physiological effects of glucocorticoids act in a curvilinear, or 'inverted U-shaped', manner on several cognitive systems, where moderate levels are optimal, while very low or high concentrations have distinct adverse cognitive outcomes (444, 445). More research is needed to clarify the deleterious effects and potential usefulness of corticosteroid treatment in MGB-related disorders.

Another group of drugs that has been shown to cause dysbiosis and is associated with cognitive decline is the proton pump inhibitors (PPI) (446, 447). Outcomes vary, with some studies showing no association. Intriguingly, gastric acid suppression by histamine-2 receptor antagonists has more reliably demonstrated associations between chronic use, dysbiosis, and cognitive decline (448).

The gut is often an unwarranted target of chemotherapeutic agents, with mucositis being a common complication (449). Women diagnosed with breast cancer undergoing chemotherapy experience disruption in GM diversity and composition, cognitive impairment, and symptoms of stress, such as anxiety and depression (450).

4.2.3.4 Exposure to Infection

Unsurprisingly, both bacterial and viral infections have been associated with cognitive impairment and functional decline in humans (451–453). *C. difficile* infection is an important example demonstrating these adverse health consequences, beyond acute intestinal dysbiosis (454). Evidence is emerging regarding the potential involvement of *C. difficile* in brain pathologies such as neurodegenerative diseases (e.g. Parkinson's disease, Alzheimer's disease), multiple sclerosis, and autism spectrum disorder (455). *C. difficile* has been shown to alter the metabolism of dopamine and interfere with cognitive functions that involve the neuromodulatory action of dopamine, such as motivation and memory consolidation (456). In two case reports in humans, FMT improved gastrointestinal symptoms, cognition, and mood, along with the eradication of *C. difficile* in patients with Alzheimer's disease (457, 458).

Evidence suggests that the cumulative effect of exposure to multiple infectious pathogens, both bacterial and viral, over several years is associated with changes in cognition, as well as multi-system biological dysfunction before clinical disease is apparent (459, 460). In a large multi-ethnic cohort, an elevated infectious burden, defined as a composite serologic measure of exposure to five specific common pathogens (e.g. cytomegalovirus, *Helicobacter pylori* and herpes simplex virus), was associated with cognitive impairment as assessed by the mini-mental state examination (461, 462). Thus, there appears to be a cumulative effect whereby past infections may contribute to later cognitive impairments. Furthermore, there is extensive evidence in humans showing that prenatal infectious exposure is associated with cognitive and other neurodevelopmental impairments (463, 464).

4.2.3.5 Probiotics and prebiotics

Over time, there has been an increasing recognition of the potential value of probiotics, prebiotics, and combinations thereof. These interventions may be beneficial for patients with dysbiosis, as well as for individuals presenting with other specific complaints or those taking them prophylactically.

Several studies in healthy participants have demonstrated reduced stress-induced cortisol levels and pro-inflammatory cytokines, along with improved cognitive test performance, perceptions of stress, anxiety, and mood (465–469). In a group of healthy women consuming a fermented milk product containing several probiotics, brain activity was found to be enhanced in specific regions associated with cognition (470). In a RCT, multispecies probiotic supplementation protected against the neurocognitive effects of acute stress in healthy women, as measured by neural changes in the frontal cortex using fMRI (471). These findings have been replicated and expanded upon, with studies exploring the relationships between stress, cognition, and the MGB axis. These studies have identified compositional and functional changes in the intestine and brain, along with distinct changes in brain morphology, resting-state brain function, brain activity and functional connectivity in regions known to regulate stress (472, 473). Mechanistic studies have further linked the stress response, cognition, and MGB axis, demonstrating a relative abundance of fibre-degrading bacteria that produce SCFAs, as well as changes in the serotonin and dopamine-norepinephrine pathways (474, 475).

Probiotics have been shown to improve cognitive symptoms and biochemical markers in participants with various dysbiosisrelated disorders, including Alzheimer's disease (476), fibromyalgia (477, 478), and irritable bowel syndrome (IBS) (479). Clinical studies have also suggested that probiotics may decrease anxiety and depression (480). In a study of non-obese and obese individuals, GM composition was found to be associated with scores of cognitive speed, attention, and flexibility, along with significant changes in neural activity in specific brain regions (thalamus, hypothalamus, and amygdala) (481). Coadministration of a probiotic containing L. rhamnosus, B. animalis, and magnesium chloride for nine weeks to obese male and female participants with depression did not rescue their altered cognition, mood, and intestinal integrity, but did decrease CRP levels (482). Administration of heat-killed M. vaccae to terminal lung cancer patients was shown to improve emotional health and cognitive function, thought to be as a result of the enhanced release of neurotransmitters such as serotonin, as part of the immune response to the microbial products (483).

In a study where healthy individuals were given the prebiotic inulin, subjective improvements in mood were noted, along with improved scores on a set of memory tasks (484). In another study, B-GOS supplementation was compared to placebo in healthy participants and found to decrease waking salivary cortisol concentrations and increase the positive processing of information in a dot-probe task (485). Therefore, prebiotics may modulate the HPA axis and benefit cognition.

Barrio et al. reviewed the association between the GM and conditions with neurocognitive impairment, finding that dysbiosis can predict the development of these disorders and influence their pathogenesis (486). Additionally, interventions such as dietary fiber supplementation or probiotics (*Lactobacillus* sp.) have been shown to improve cognitive function and modulate HPA axis activity (reducing cortisol response).

It is important to note that many probiotic trials show no significant effect, especially when performed in healthy populations. For instance, *L. rhamnosus* was not found to be superior to placebo in modifying stress, HPA response, inflammation, or cognitive performance, and the authors highlighted the challenges

associated with advancing promising preclinical studies in animals to the clinic (487). Notably, the participants were healthy adult males and not patients with stress-related disorders. A recent systematic review of interventions in children and adolescents to enhance cognitive functioning and emotional behavior found limited consistent effects in these developing populations, although study heterogeneity was seen as a major factor (488). Another systematic review evaluating the utility of prebiotics, probiotics, and fermented food interventions on cognitive performance had negative findings, possibly due to the limited number of small and short-term studies, as well as clinical heterogeneity relating to the population, cognitive tests, and interventions (489). Therefore, further clinical research using adequately powered samples and standardized protocols is warranted.

4.2.3.6 Stress

Chronic stress is associated with various cognitive deficits, systemic inflammation, premature aging, immune system dysfunction, and a higher likelihood of suffering from metabolic disorders such as diabetes, cardiovascular disease, and IBD (490–492). Stress is considered a trigger in patients with functional gastrointestinal disorders, such as IBS and functional dyspepsia, as well as IBDs like ulcerative colitis and Crohn's disease (493). These conditions exhibit cognitive and behavioral alterations, along with dysbiotic GM profiles.

A study using 3T MRI technology found thalamic volume to be smaller in the IBD group, when compared to healthy controls (494). Yamaoka et al. recently examined the relationship between the GM and stress-related brain functions in healthy subjects, using functional near-infrared spectroscopy (495). They found that the prefrontal cortex stress response correlated with the relative abundance of GM microbes, and that healthy participants with higher stress responses had an increased abundance of microbes found to be associated with depression.

Cognitive alterations may be present in individuals with IBS, and evidence is emerging that specific changes relate to hippocampal-mediated visuospatial memory deficits, which are linked to indices of HPA axis function (20, 496–498). IBS patients also show impairments in tests of cognitive flexibility and have abnormal brain activity in frontal regions of the brain (499).

4.3 The relationship between the gut microbiota and cognition, independent of the HPA axis

It is evident that the GM has a significant and profound influence on cognition. Although the HPA axis is centrally placed in linking the various components of the MGB axis with cognitive function, research has demonstrated GM effects on cognition that are independent of the HPA axis. Additionally, while GM signaling pathways and mechanisms involving the vagus nerve, immune system, microbial metabolites, hormones, and neurotransmitters may impact cognition either directly or indirectly through the HPA axis, their involvement might not always require the HPA axis (223).

For example, Gareau et al. demonstrated a lack of memory in germ-free mice in the T-maze test and novel object test in situations with or without stress (362). Frohlich et al. administered antibiotics to adult mice and demonstrated that the resultant cognitive deficits were associated with brain region-specific changes in BDNF, NMDA receptor subunit 2B, serotonin transporter, and NPY (383). Vagus nerve stimulation has been shown to improve cognitive processes and has been attributed to modulation of central noradrenergic and GABA systems, as well as neuronal adaptations within the amygdala, hippocampus, and prefrontal cortex (500). Microbes also create neurotoxic substances such as D-lactate, homocysteine, and ammonia, which can pass through the BBB and affect cognition (321).

Animal models of neurodegenerative diseases have demonstrated associations between the MGB axis and cognition that are likely independent of the HPA axis. In Alzheimer's disease, CCAAT/enhancer binding protein β /asparagine endopeptidase (C/ EBP β /AEP) signaling mediates disease progression by cleaving both β-amyloid precursor protein and Tau. A recent study demonstrated that gut dysbiosis was positively associated with C/EBPB/AEP messaging in the brain of a 5xFAD mouse model of Alzheimer's disease, concomitant with age-related progression of disease severity (501). Chronic antibiotic treatment subdued C/EBPB/ AEP signaling and diminished amyloidogenic processes, rescuing cognitive functions. In a D-galactose/aluminum chloride-induced model of Alzheimer's disease, selenium nanoparticles-enriched L. casei (ATCC 393) administered for thirteen weeks significantly improved cognitive dysfunction and minimized β-amyloid aggregation, hyperphosphorylation of Tau protein, and prevented neuronal death by modulating the Akt/cAMP-response element binding protein/BDNF signaling pathway (502). The intervention was shown to mitigate intestinal barrier dysfunction, inhibit the activation of microglia, and protect brain neurons from oxidative stress and neuroinflammation. Several microbial species can produce amyloid proteins, which can cross the intestinal and BBB when permeable due to inflammation, and promote β -amyloid protein formation and accumulation in the brain, thereby enhancing Alzheimer's disease pathogenesis in the elderly (503).

In a mouse model that overexpresses the protein α -synuclein, resulting in Parkinsonism, Sampson et al. were able to show that MGB axis signaling is required for motor deficits, microglia activation, and synucleinopathies (75). Oral administration of SCFAs to germ-free α -synuclein mice promoted neuroinflammation and motor dysfunction, and FMT with Parkinson's disease-affected patients enhanced the Parkinsonism compared to FMT from healthy human donors. In a study by Liu et al., administration of the SCFA butyrate improved cognitive impairments in a mouse model of vascular dementia (392). Huntington's disease is another neurodegenerative disorder involving psychiatric, cognitive, and motor symptoms. FMT from wild-type mice into a mouse model of Huntington's disease improved cognitive outcomes, particularly in females (504). Interestingly, simply transplanting the GM of aged rats into young rats induced structural and functional alterations in the brain, as well as behavioral changes indicative of cognitive decline (505).

The influence of diet and dietary habits on the GM is an extensive topic and a primary determinant of GM composition and function. A comprehensive review of this topic is beyond the scope of this review, however, a significant body of evidence demonstrates that consuming a GM-favorable diet can positively influence the GM, improve health status, and enhance cognitive function (506–510). This is likely due to the availability of prebiotics, probiotics, SCFAs, and polyphenols. Conversely, a poor diet and obesity are associated with GM alterations, cognitive dysfunction, and an increased risk for dementia [reviewed in (511)]. Proposed mechanisms include changes in GM composition and function, alterations in microbial metabolites, compromised barrier integrity, and peripheral and central inflammatory processes, as well as hormonal, glucoregulatory, and cardiovascular changes (502, 511, 512).

In obesity, dysbiosis has been associated with cognitive alterations such as hippocampal dysfunction, impaired memory, and reductions in attention and executive function (513). Notably, weight gain and obesity do not appear to be necessary for dietinduced cognitive impairment and age-related cognitive decline (514, 515). Mice colonized with GM from donors fed a high-fat diet demonstrated compositionally distinct microbiota, altered biochemical markers, and developed impairments in exploratory, cognitive, and stereotypical behaviors, in the absence of significant differences in body weight (516). In mice with diet-induced obesity, antibacterial treatment improved metabolic parameters, insulin signaling in the brain, and neurobehavioral changes indicative of anxiety and depression (517). These changes were associated with altered levels of tryptophan, GABA, BDNF, amino acids, and acylcarnitines, while these effects were transferable to germ-free mice by FMT. In this way, FMTs have provided more direct evidence for the role of dysbiotic microbiota in cognitive dysfunction, independent of the HPA axis.

Besides diet, other environmental factors can alter GM composition and heterogeneity, as well as behavioral phenotype and cognition. Jaric et al. showed that even subtle environmental variation, such as the rearing facility of the mice, significantly influenced epigenetic patterns in neuronal genes at the level of chromatin organization, with effects on nucleosome assembly, neuronal differentiation, synaptic plasticity, and regulation of behavior (518). Guo et al. showed that mice exposed to environmental low-dose radiation demonstrated long-term impairments in cognitive function, hippocampal synaptic ultrastructure, and signaling through the Akt/mTOR pathway (519). Indeed, abnormal GM composition may contribute to radiotherapy-induced cognitive decline seen in cancer treatment (520).

5 Future perspectives

It is clear that the development and maintenance of higher cognitive function in humans are influenced and modulated by bidirectional HPA axis and GM mechanisms in the complex interplay of the MGB axis. Recognizing these connections opens a plethora of opportunities for novel mechanisms and potential avenues for GM-based diagnostics and therapeutics. While the current evidence is promising, it is essential to acknowledge the need for more research to better understand the underlying mechanisms and the potential for clinical applications in treating conditions with cognitive impairments.

Although GM-based therapeutics are an exciting field and receive much attention, it is vital that rigorous consideration be paid to understanding the mechanistic pathways, as well as the critical window for the development of the GM, HPA axis, and neurological system. Possible therapeutic strategies that could modulate GM and potentially improve cognitive function in different neurological disorders include FMT, prebiotic and probiotic supplementation, and dietary interventions.

While the GM plays an important role in shaping brain function, including the activity of the HPA axis, its role with regards to the distinct structures of the HPA axis remains understudied. Only recently have a small number of studies begun to provide detailed mechanistic insights. Xiang et al. showed that nucleotide-binding oligomerization domain 1 (Nod1) ligands, derived from the GM, modulate catecholamine storage and secretion, and therefore intestinal bacteria modulate the adrenomedullary response through Nod1 sensing in chromaffin cells (521).

Extra-adrenal glucocorticoid production is known to occur in the gut and other tissues, and these glucocorticoids play a highly specific role in regulating local homeostasis, cell development, and immune activation (522–526). However, there is only preliminary evidence available demonstrating that under basal conditions the GM contributes to the regulation of intestinal glucocorticoid production (527, 528).

Furthermore, GM-based studies of adrenal insufficiency and Cushing's syndrome are also severely lacking, although, as has been demonstrated throughout this review, there are mechanistic reasons to believe that important associations may be uncovered. The most common cause of adrenal insufficiency is autoimmune Addison's disease, which is thought to be a hapless combination of susceptible genetics and environmental factors. Whether the GM influences the development or pathogenesis of autoimmune Addison's disease is currently unknown. Many pathogens are known to infect the adrenal cortex, and some of them (e.g., M. tuberculosis) can cause adrenal insufficiency (529). A study of chronic adrenal insufficiency patients highlighted that 30% of the participants suffered frequent and incapacitating gut symptoms, consistent with the Rome IV criteria for IBS, and that they had a decreased quality of life (530). However, no evaluation of the GM was performed in this study. Further GM-based research should be prioritized for these two HPA axis conditions.

Finally, a plethora of human diseases have increasingly been associated with changes in the GM. However, whether these changes are causal, consequential, or co-incidental remain unresolved. While animal models have been critical to our understanding of how microbes influence host development and function, and germ-free models are a cornerstone for studying alterations that arise from microbial perturbations, they may not be directly transferable to models of human function. Introducing human microbial communities into germ-free animals may represent a more clinically relevant model. If the field is to move forward, it is necessary that the focus shifts towards determining causal relationships in humans through rigorous and critical approaches and to further advance the translatability of animal and pre-clinical studies to human trials.

In conclusion, this review confirms that the interaction between microbes and the endocrine and neural systems, including the HPA axis, is essential for optimal cognitive function in humans.

Author contributions

Review written by JR and LD and edited by BL. All authors contributed to the article and approved the submitted version.

References

1. D'Argenio V, Salvatore F. The role of the gut microbiome in the healthy adult status. *Clin Chim Acta* (2015) 451:97–102. doi: 10.1016/j.cca.2015.01.003

2. Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell* (2012) 148:1258–70. doi: 10.1016/j.cell.2012.01.035

3. Herrema H, IJzerman RG, Nieuwdorp M. Emerging role of intestinal microbiota and microbial metabolites in metabolic control. *Diabetologia* (2017) 60:613–7. doi: 10.1007/s00125-016-4192-0

4. Chen Y, Zhou J, Wang L. Role and mechanism of gut microbiota in human disease. Front Cell Infect Microbiol (2021) 11:625913. doi: 10.3389/fcimb.2021.625913

5. Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, et al. Diversity of the human intestinal microbial flora. *Science* (2005) 308:1635-8. doi: 10.1126/science.1110591

6. Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JI. The human microbiome project. *Nature* (2007) 449:804–10. doi: 10.1038/nature06244

7. Gevers D, Knight R, Petrosino JF, Huang K, McGuire AL, Birren BW, et al. The human microbiome project: a community resource for the healthy human microbiome. *PloS Biol* (2012) 10:6–10. doi: 10.1371/journal.pbio.1001377

8. Sommer F, Bäckhed F. The gut microbiota-masters of host development and physiology. *Nat Rev Microbiol* (2013) 11:227–38. doi: 10.1038/nrmicro2974

9. Walker AW, Duncan SH, Louis P, Flint HJ. Phylogeny, culturing, and metagenomics of the human gut microbiota. *Trends Microbiol* (2014) 22:267–74. doi: 10.1016/j.tim.2014.03.001

10. Sender R, Fuchs S, Milo R. Are we really vastly outnumbered? revisiting the ratio of bacterial to host cells in humans. *Cell* (2016) 164:337–40. doi: 10.1016/j.cell.2016.01.013

11. Kim CY, Lee M, Yang S, Kim K, Yong D, Kim HR, et al. Human reference gut microbiome catalog including newly assembled genomes from under-represented Asian metagenomes. *Genome Med* (2021) 13:134. doi: 10.1186/s13073-021-00950-7

12. Boulangé CL, Neves AL, Chilloux J, Nicholson JK, Dumas ME. Impact of the gut microbiota on inflammation, obesity, and metabolic disease. *Genome Med* (2016) 8:1–12. doi: 10.1186/s13073-016-0303-2

13. Nieuwdorp M, Gilijamse PW, Pai N, Kaplan LM. Role of the microbiome in energy regulation and metabolism. *Gastroenterology* (2014) 146:1525–33. doi: 10.1053/j.gastro.2014.02.008

14. Fujimura KE, Lynch SV. Microbiota in allergy and asthma and the emerging relationship with the gut microbiome. *Cell Host Microbe* (2015) 17:592–602. doi: 10.1016/j.chom.2015.04.007

Acknowledgments

LD is supported by NIH grant's R01DK111848 and R01HL148271 and the AXA Research Fund. BL is supported by 2R01 DK104927, R01HL148271 and VA Merit1I01BX00382-01A1.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

15. Lane ER, Zisman TL, Suskind DL. The microbiota in inflammatory bowel disease: current and therapeutic insights. *J Inflammation Res* (2017) 10:63–73. doi: 10.2147/JIR.S116088

16. Yurkovetskiy LA, Pickard JM, Chervonsky AV. Microbiota and autoimmunity: exploring new avenues. *Cell Host Microbe* (2015) 17:548–52. doi: 10.1016/j.chom.2015.04.010

17. de Weerth C. Do bacteria shape our development? crosstalk between intestinal microbiota and HPA axis. *Neurosci Biobehav Rev* (2017) 83:458–71. doi: 10.1016/j.neubiorev.2017.09.016

 Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG. Minireview: gut microbiota: the neglected endocrine organ. *Mol Endocrinol* (2014) 28:1221–38. doi: 10.1210/me.2014-1108

19. Mayer EA, Knight R, Mazmanian SK, Cryan JF, Tillisch K. Gut microbes and the brain: paradigm shift in neuroscience. *J Neurosci* (2014) 34:15490–6. doi: 10.1523/JNEUROSCI.3299-14.2014

20. Mayer EA. Gut feelings: the emerging biology of gut-brain communication. *Nat Rev Neurosci* (2011) 12:453–66. doi: 10.1038/nrn3071

21. Carabotti M, Scirocco A, Maselli MA, Severi C. The gut-brain axis: interactions between enteric microbiota, central and enteric nervous systems. *Ann Gastroenterol* (2015) 28:203–9.

22. Burokas A, Moloney RD, Dinan TG, Cryan JF. Microbiota regulation of the mammalian gut-brain axis. (2015) 91:1-62. doi: 10.1016/bs.aambs.2015.02.001

23. Chakrabarti A, Geurts L, Hoyles L, Iozzo P, Kraneveld AD, La Fata G, et al. The microbiota–gut–brain axis: pathways to better brain health. perspectives on what we know, what we need to investigate and how to put knowledge into practice. *Cell Mol Life Sci* (2022) 79:1–15. doi: 10.1007/s00018-021-04060-w

24. Rhee SH, Pothoulakis C, Mayer EA. Principles and clinical implications of the brain-gut-enteric microbiota axis. *Nat Rev Gastroenterol Hepatol* (2009) 6:306–14. doi: 10.1038/nrgastro.2009.35

25. Cryan JF, O'riordan KJ, Cowan CSM, Sandhu KV, Bastiaanssen TFS, Boehme M, et al. The microbiota-gut-brain axis. *Physiol Rev* (2019) 99:1877–2013. doi: 10.1152/ physrev.00018.2018

26. Dinan TG, Cryan JF. The microbiome-Gut-Brain axis in health and disease. Gastroenterol Clin North Am (2017) 46:77–89. doi: 10.1016/j.gtc.2016.09.007

27. Mirzaei R, Bouzari B, Hosseini-Fard SR, Mazaheri M, Ahmadyousefi Y, Abdi M, et al. Role of microbiota-derived short-chain fatty acids in nervous system disorders. *BioMed Pharmacother* (2021) 139:111661. doi: 10.1016/j.biopha.2021.111661

28. Strandwitz P. Neurotransmitter modulation by the gut microbiota. Brain Res (2018) 1693:128-33. doi: 10.1016/j.brainres.2018.03.015

29. Tetel MJ, de Vries GJ, Melcangi RC, Panzica G, O'Mahony SM. Steroids, stress and the gut microbiome-brain axis. *J Neuroendocrinol* (2018) 30:e12548. doi: 10.1111/jne.12548

30. Staley C, Weingarden AR, Khoruts A, Sadowsky MJ. Interaction of gut microbiota with bile acid metabolism and its influence on disease states. *Appl Microbiol Biotechnol* (2017) 101:47–64. doi: 10.1007/s00253-016-8006-6

31. O'Mahony SM, Clarke G, Borre YE, Dinan TG, Cryan JF. Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. *Behav Brain Res* (2015) 277:32–48. doi: 10.1016/j.bbr.2014.07.027

32. Cani PD, Osto M, Geurts L, Everard A. Involvement of gut microbiota in the development of low-grade inflammation and type 2 diabetes associated with obesity. *Gut Microbes* (2012) 3:279–88. doi: 10.4161/gmic.19625

33. Rutsch A, Kantsjö JB, Ronchi F. The gut-brain axis: how microbiota and host inflammasome influence brain physiology and pathology. *Front Immunol* (2020) 11:604179. doi: 10.3389/fimmu.2020.604179

34. Frazier TH, DiBaise JK, McClain CJ. Gut microbiota, intestinal permeability, obesity-induced inflammation, and liver injury. *JPEN J Parenter Enteral Nutr* (2011) 35:14S–20S. doi: 10.1177/0148607111413772

35. Fasano A. Gut permeability, obesity, and metabolic disorders: who is the chicken and who is the egg? *Am J Clin Nutr* (2017) 105:3–4. doi: 10.3945/ajcn.116.148338

36. Round JL, Mazmanian SK. The gut microbiota shapes intestinal immune responses during health and disease. *Nat Rev Immunol* (2009) 9:313-23. doi: 10.1038/nri2515

37. Allaire JM, Crowley SM, Law HT, Chang S-Y, Ko H-J, Vallance BA. The intestinal epithelium: central coordinator of mucosal immunity. *Trends Immunol* (2018) 39:677–96. doi: 10.1016/j.it.2018.04.002

38. Wlodarska M, Kostic AD, Xavier RJ. An integrative view of microbiome-host interactions in inflammatory bowel diseases. *Cell Host Microbe* (2015) 17:577–91. doi: 10.1016/j.chom.2015.04.008

39. Cani PD, Everard A, Duparc T. Gut microbiota, enteroendocrine functions and metabolism. *Curr Opin Pharmacol* (2013) 13:935–40. doi: 10.1016/j.coph.2013.09.008

40. Valdez-Morales EE, Overington J, Guerrero-Alba R, Ochoa-Cortes F, Ibeakanma CO, Spreadbury I, et al. Sensitization of peripheral sensory nerves by mediators from colonic biopsies of diarrhea-predominant irritable bowel syndrome patients: a role for PAR2. *Am J Gastroenterol* (2013) 108:1634–43. doi: 10.1038/ajg.2013.241

41. Forsythe P, Kunze WA. Voices from within: gut microbes and the CNS. *Cell Mol Life Sci* (2013) 70:55–69. doi: 10.1007/s00018-012-1028-z

42. Dinan TG, Cryan JF. Gut instincts: microbiota as a key regulator of brain development, ageing and neurodegeneration. *J Physiol* (2017) 595:489-503. doi: 10.1113/JP273106

43. Moloney RD, Desbonnet L, Clarke G, Dinan TG, Cryan JF. The microbiome: stress, health and disease. *Mamm Genome* (2014) 25:49–74. doi: 10.1007/s00335-013-9488-5

44. Salami M. Interplay of good bacteria and central nervous system: cognitive aspects and mechanistic considerations. *Front Neurosci* (2021) 15:613120. doi: 10.3389/ fnins.2021.613120

45. Sampson TR, Mazmanian SK. Control of brain development, function, and behavior by the microbiome. *Cell Host Microbe* (2015) 17:565–76. doi: 10.1016/j.chom.2015.04.011

46. Vuong HE, Yano JM, Fung TC, Hsiao EY. The microbiome and host behavior. Annu Rev Neurosci (2017) 40:21–49. doi: 10.1146/annurev-neuro-072116-031347

47. Connell E, Le Gall G, Pontifex MG, Sami S, Cryan JF, Clarke G, et al. Microbialderived metabolites as a risk factor of age-related cognitive decline and dementia. *Mol Neurodegener* (2022) 17:43. doi: 10.1186/s13024-022-00548-6

48. Mayer EA. The neurobiology of stress and gastrointestinal disease. *Gut* (2000) 47:861–9. doi: 10.1136/gut.47.6.861

49. Lyte M, Li W, Opitz N, Gaykema RPA, Goehler LE. Induction of anxiety-like behavior in mice during the initial stages of infection with the agent of murine colonic hyperplasia citrobacter rodentium. *Physiol Behav* (2006) 89:350–7. doi: 10.1016/ j.physbeh.2006.06.019

50. Goehler LE, Park SM, Opitz N, Lyte M, Gaykema RPA. Campylobacter jejuni infection increases anxiety-like behavior in the holeboard: possible anatomical substrates for viscerosensory modulation of exploratory behavior. *Brain Behav Immun* (2008) 22:354–66. doi: 10.1016/j.bbi.2007.08.009

51. Sudo N, Chida Y, Aiba Y, Sonoda J, Oyama N, Yu XN, et al. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J Physiol* (2004) 558:263–75. doi: 10.1113/jphysiol.2004.063388

52. Forsythe P, Bienenstock J, Kunze WA. Vagal pathways for microbiome-Brain-Gut axis communication BT - microbial endocrinology: the microbiota-Gut-Brain axis in health and disease. Lyte M, Cryan JF, editors. New York, NY: Springer New York (2014) p. 115–33. doi: 10.1007/978-1-4939-0897-4_5

53. Geng ZH, Zhu Y, Li QL, Zhao C, Zhou PH. Enteric nervous system: the bridge between the gut microbiota and neurological disorders. *Front Aging Neurosci* (2022) 14:810483. doi: 10.3389/fnagi.2022.810483

54. Hyland NP, Cryan JF. Microbe-host interactions: influence of the gut microbiota on the enteric nervous system. *Dev Biol* (2016) 417:182–7. doi: 10.1016/j.ydbio.2016.06.027

55. Kelly JR, Borre Y, O' Brien C, Patterson E, El Aidy S, Deane J, et al. Transferring the blues: depression-associated gut microbiota induces neurobehavioural changes in the rat. *J Psychiatr Res* (2016) 82:109–18. doi: 10.1016/j.jpsychires.2016.07.019

56. Mangiola F, Ianiro G, Franceschi F, Fagiuoli S, Gasbarrini G, Gasbarrini A. Gut microbiota in autism and mood disorders. *World J Gastroenterol* (2016) 22:361–8. doi: 10.3748/wjg.v22.i1.361

57. Stilling RM, Dinan TG, Cryan JF. Microbial genes, brain & behaviour - epigenetic regulation of the gut-brain axis. *Genes Brain Behav* (2014) 13:69–86. doi: 10.1111/gbb.12109

58. Foster JA, Rinaman L, Cryan JF. Stress & the gut-brain axis: regulation by the microbiome. *Neurobiol Stress* (2017) 7:124–36. doi: 10.1016/j.ynstr.2017.03.001

59. Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci* (2012) 13:701–12. doi: 10.1038/ nrn3346

60. Bercik P, Collins SM, Verdu EF. Microbes and the gut-brain axis. Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc (2012) 24:405-13. doi: 10.1111/j.1365-2982.2012.01906.x

61. Alcock J, Maley CC, Aktipis CA. Is eating behavior manipulated by the gastrointestinal microbiota? evolutionary pressures and potential mechanisms. *Bioessays* (2014) 36:940–9. doi: 10.1002/bies.201400071

62. Lam YY, Maguire S, Palacios T, Caterson ID. Are the gut bacteria telling us to eat or not to eat? reviewing the role of gut microbiota in the etiology, disease progression and treatment of eating disorders. *Nutrients* (2017) 9(6):602. doi: 10.3390/nu9060602

63. Foster JA, McVey Neufeld KA. Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci* (2013) 36:305-12. doi: 10.1016/j.tins.2013.01.005

64. Humann J, Mann B, Gao G, Moresco P, Ramahi J, Loh LN, et al. Bacterial peptidoglycan traverses the placenta to induce fetal neuroproliferation and aberrant postnatal behavior. *Cell Host Microbe* (2016) 19:388–99. doi: 10.1016/j.chom.2016.02.009

65. Mayer EA, Padua D, Tillisch K. Altered brain-gut axis in autism: comorbidity or causative mechanisms? *Bioessays* (2014) 36:933–9. doi: 10.1002/bies.201400075

66. Song Y, Liu C, Finegold SM. Real-time PCR quantitation of clostridia in feces of autistic children. *Appl Environ Microbiol* (2004) 70:6459–65. doi: 10.1128/AEM.70.11.6459-6465.2004

67. Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, et al. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell* (2013) 155:1451-63. doi: 10.1016/j.cell.2013.11.024

68. Naseribafrouei A, Hestad K, Avershina E, Sekelja M, Linløkken A, Wilson R, et al. Correlation between the human fecal microbiota and depression. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2014) 26:1155–62. doi: 10.1111/nmo.12378

69. Du Y, Gao XR, Peng L, Ge JF. Crosstalk between the microbiota-gut-brain axis and depression. *Heliyon* (2020) 6(6):e04097. doi: 10.1016/j.heliyon.2020.e04097

70. Mayer EA, Tillisch K. The brain-gut axis in abdominal pain syndromes. Annu Rev Med (2011) 62:381–96. doi: 10.1146/annurev-med-012309-103958

71. Berrill JW, Gallacher J, Hood K, Green JT, Matthews SB, Campbell AK, et al. An observational study of cognitive function in patients with irritable bowel syndrome and inflammatory bowel disease. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2013) 25:918–e704. doi: 10.1111/nmo.12219

72. Simrén M, Barbara G, Flint HJ, Spiegel BMR, Spiller RC, Vanner S, et al. Intestinal microbiota in functional bowel disorders: a Rome foundation report. *Gut* (2013) 62:159–76. doi: 10.1136/gutjnl-2012-302167

73. Haran JP, Bhattarai SK, Foley SE, Dutta P, Ward DV, Bucci V, et al. Alzheimer's disease microbiome is associated with dysregulation of the anti-inflammatory p-glycoprotein pathway. *MBio* (2019) 10:e00632-19. doi: 10.1128/mBio.00632-19

74. Romano S, Savva GM, Bedarf JR, Charles IG, Hildebrand F, Narbad A. Metaanalysis of the parkinson's disease gut microbiome suggests alterations linked to intestinal inflammation. *NPJ Park Dis* (2021) 7:27. doi: 10.1038/s41531-021-00156-z

75. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, et al. Gut microbiota regulate motor deficits and neuroinflammation in a model of parkinson's disease. *Cell* (2016) 167:1469–1480.e12. doi: 10.1016/j.cell.2016.11.018

76. Correale J, Hohlfeld R, Baranzini SE. The role of the gut microbiota in multiple sclerosis. *Nat Rev Neurol* (2022) 18:544–58. doi: 10.1038/s41582-022-00697-8

77. Zheng P, Zeng B, Liu M, Chen J, Pan J, Han Y, et al. The gut microbiome from patients with schizophrenia modulates the glutamate-glutamine-GABA cycle and schizophrenia-relevant behaviors in mice. *Sci Adv* (2019) 5:eaau8317. doi: 10.1126/sciadv.au8317

 Fülling C, Dinan TG, Cryan JF. Gut microbe to brain signaling: what happens in vagus. Neuron (2019) 101:998–1002. doi: 10.1016/j.neuron.2019.02.008

79. Jiménez E, Marín ML, Martín R, Odriozola JM, Olivares M, Xaus J, et al. Is meconium from healthy newborns actually sterile? *Res Microbiol* (2008) 159:187–93. doi: 10.1016/j.resmic.2007.12.007

80. Collado MC, Rautava S, Aakko J, Isolauri E, Salminen S. Human gut colonisation may be initiated *in utero* by distinct microbial communities in the placenta and amniotic fluid. *Sci Rep* (2016) 6:1–13. doi: 10.1038/srep23129

81. Frese SA, Mills DA. Birth of the infant gut microbiome: moms deliver twice! *Cell Host Microbe* (2015) 17:543–4. doi: 10.1016/j.chom.2015.04.014

82. Zeng S, Wang S, Ross RP, Stanton C. The road not taken: host genetics in shaping intergenerational microbiomes. *Trends Genet* (2022) 38:1180–92. doi: 10.1016/j.tig.2022.05.016

83. Tamburini S, Shen N, Wu HC, Clemente JC. The microbiome in early life: implications for health outcomes. *Nat Med* (2016) 22:713–22. doi: 10.1038/nm.4142

84. Koenig JE, Spor A, Scalfone N, Fricker AD, Stombaugh J, Knight R, et al. Succession of microbial consortia in the developing infant gut microbiome. *Proc Natl Acad Sci U.S.A.* (2011) 108:4578–85. doi: 10.1073/pnas.1000081107

85. Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, et al. Human gut microbiome viewed across age and geography. *Nature* (2012) 486:222–7. doi: 10.1038/nature11053

86. Hollister EB, Riehle K, Luna RA, Weidler EM, Rubio-Gonzales M, Mistretta T-A, et al. Structure and function of the healthy pre-adolescent pediatric gut microbiome. *Microbiome* (2015) 3:36. doi: 10.1186/s40168-015-0101-x

87. Yuan X, Chen R, Zhang Y, Lin X, Yang X. Gut microbiota: effect of pubertal status. *BMC Microbiol* (2020) 20:334. doi: 10.1186/s12866-020-02021-0

88. Martino C, Dilmore AH, Burcham ZM, Metcalf JL, Jeste D, Knight R. Microbiota succession throughout life from the cradle to the grave. Nat Rev Microbiol (2022) 20:707–20. doi: 10.1038/s41579-022-00768-z

89. Meyer K, Lulla A, Debroy K, Shikany JM, Yaffe K, Meirelles O, et al. Association of the gut microbiota with cognitive function in midlife. *JAMA Netw Open* (2022) 5(2): e2143941. doi: 10.1001/jamanetworkopen.2021.43941

90. Xu C, Zhu H, Qiu P. Aging progression of human gut microbiota. BMC Microbiol (2019) 19:236. doi: 10.1186/s12866-019-1616-2

91. Jeffery IB, Lynch DB, O'Toole PW. Composition and temporal stability of the gut microbiota in older persons. *ISME J* (2016) 10:170–82. doi: 10.1038/ismej.2015.88

92. Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S, et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature* (2012) 488:178–84. doi: 10.1038/nature11319

93. Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, et al. Composition, variability, and temporal stability of the intestinal microbiota of the elderly. *Proc Natl Acad Sci U.S.A.* (2011) 108 Suppl 1:4586–91. doi: 10.1073/pnas.1000097107

94. O'Toole PW, Jeffery IB. Gut microbiota and aging. Science (2015) 350:1214-5. doi: 10.1126/science.aac8469

95. Odamaki T, Kato K, Sugahara H, Hashikura N, Takahashi S, Xiao J, et al. Agerelated changes in gut microbiota composition from newborn to centenarian: a crosssectional study. *BMC Microbiol* (2016) 16:90. doi: 10.1186/s12866-016-0708-5

96. Biagi E, Nylund L, Candela M, Ostan R, Bucci L, Pini E, et al. Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. *PloS One* (2010) 5:e10667. doi: 10.1371/journal.pone.0010667

97. Chen Y, Wang H, Lu W, Wu T, Yuan W, Zhu J, et al. Human gut microbiome aging clocks based on taxonomic and functional signatures through multi-view learning. *Gut Microbes* (2022) 14:2025016. doi: 10.1080/19490976.2021.2025016

98. Galkin F, Mamoshina P, Aliper A, Putin E, Moskalev V, Gladyshev VN, et al. Human gut microbiome aging clock based on taxonomic profiling and deep learning. *iScience* (2020) 23:101199. doi: 10.1016/j.isci.2020.101199

99. Franceschi C, Campisi J. Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *J Gerontol A Biol Sci Med Sci* (2014) 69 Suppl 1:S4–9. doi: 10.1093/gerona/glu057

100. O'Toole PW, Jeffery IB. Microbiome-health interactions in older people. Cell Mol Life Sci (2018) 75:119–28. doi: 10.1007/s00018-017-2673-z

101. Jackson MA, Jeffery IB, Beaumont M, Bell JT, Clark AG, Ley RE, et al. Signatures of early frailty in the gut microbiota. *Genome Med* (2016) 8:8. doi: 10.1186/s13073-016-0262-7

102. Liu S, Gao J, Zhu M, Liu K, Zhang H-L. Gut microbiota and dysbiosis in alzheimer's disease: implications for pathogenesis and treatment. *Mol Neurobiol* (2020) 57:5026–43. doi: 10.1007/s12035-020-02073-3

103. Gorecki AM, Preskey L, Bakeberg MC, Kenna JE, Gildenhuys C, MacDougall G, et al. Altered gut microbiome in parkinson's disease and the influence of lipopolysaccharide in a human α -synuclein over-expressing mouse model. *Front Neurosci* (2019) 13:839. doi: 10.3389/fnins.2019.00839

104. Cattaneo A, Cattane N, Galluzzi S, Provasi S, Lopizzo N, Festari C, et al. Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. *Neurobiol Aging* (2017) 49:60–8. doi: 10.1016/j.neurobiolaging.2016.08.019

105. Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC, et al. Gut microbiome alterations in alzheimer's disease. *Sci Rep* (2017) 7:13537. doi: 10.1038/s41598-017-13601-y

106. Manderino L, Carroll I, Azcarate-Peril MA, Rochette A, Heinberg L, Peat C, et al. Preliminary evidence for an association between the composition of the gut

microbiome and cognitive function in neurologically healthy older adults. J Int Neuropsychol Soc (2017) 23:700–5. doi: 10.1017/S1355617717000492

107. Zhuang Z-Q, Shen L-L, Li W-W, Fu X, Zeng F, Gui L, et al. Gut microbiota is altered in patients with alzheimer's disease. *J Alzheimers Dis* (2018) 63:1337–46. doi: 10.3233/JAD-180176

108. Saji N, Niida S, Murotani K, Hisada T, Tsuduki T, Sugimoto T, et al. Analysis of the relationship between the gut microbiome and dementia: a cross-sectional study conducted in Japan. *Sci Rep* (2019) 9:1008. doi: 10.1038/s41598-018-38218-7

109. Sheng C, Lin L, Lin H, Wang X, Han Y, Liu S-L. Altered gut microbiota in adults with subjective cognitive decline: the SILCODE study. *J Alzheimers Dis* (2021) 82:513–26. doi: 10.3233/JAD-210259

110. Gupta D, Morley JE. Hypothalamic-pituitary-adrenal (HPA) axis and aging. Compr Physiol (2014) 4:1495–510. doi: 10.1002/cphy.c130049

111. Lupien SJ, de Leon M, de Santi S, Convit A, Tarshish C, Nair NP, et al. Cortisol levels during human aging predict hippocampal atrophy and memory deficits. *Nat Neurosci* (1998) 1:69–73. doi: 10.1038/271

112. Hinkelmann K, Moritz S, Botzenhardt J, Muhtz C, Wiedemann K, Kellner M, et al. Changes in cortisol secretion during antidepressive treatment and cognitive improvement in patients with major depression: a longitudinal study. *Psychoneuroendocrinology* (2012) 37:685–92. doi: 10.1016/j.psyneuen.2011.08.012

113. Farzi A, Fröhlich EE, Holzer P. Gut microbiota and the neuroendocrine system. *Neurotherapeutics* (2018) 15:5–22. doi: 10.1007/s13311-017-0600-5

114. Collins J, Borojevic R, Verdu EF, Huizinga JD, Ratcliffe EM. Intestinal microbiota influence the early postnatal development of the enteric nervous system. *Neurogastroenterol Motil* (2014) 26:98–107. doi: 10.1111/nmo.12236

115. Borre YE, Moloney RD, Clarke G, Dinan TG, Cryan JF. The impact of microbiota on brain and behavior: mechanisms & therapeutic potential. *Adv Exp Med Biol* (2014) 817:373-403. doi: 10.1007/978-1-4939-0897-4_17

116. Sharon G, Sampson TR, Geschwind DH, Mazmanian SK. The central nervous system and the gut microbiome. *Cell* (2016) 167:915–32. doi: 10.1016/j.cell.2016.10.027

117. Sarkar A, Harty S, Johnson KVA, Moeller AH, Carmody RN, Lehto SM, et al. The role of the microbiome in the neurobiology of social behaviour. *Biol Rev* (2020) 95:1131–66. doi: 10.1111/brv.12603

118. Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* (2016) 352:712–6. doi: 10.1126/science.aad8373

119. Wilton DK, Dissing-Olesen L, Stevens B. Neuron-glia signaling in synapse elimination. *Annu Rev Neurosci* (2019) 42:107–27. doi: 10.1146/annurev-neuro-070918-050306

120. Erny D, Hrabě de Angelis AL, Jaitin D, Wieghofer P, Staszewski O, David E, et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci* (2015) 18:965–77. doi: 10.1038/nn.4030

121. Abdel-Haq R, Schlachetzki JCM, Glass CK, Mazmanian SK. Microbiomemicroglia connections *via* the gut-brain axis. *J Exp Med* (2019) 216:41–59. doi: 10.1084/ jem.20180794

122. Borre YE, O'Keeffe GW, Clarke G, Stanton C, Dinan TG, Cryan JF. Microbiota and neurodevelopmental windows: implications for brain disorders. *Trends Mol Med* (2014) 20:509–18. doi: 10.1016/j.molmed.2014.05.002

123. Luczynski P, Neufeld KAMV, Oriach CS, Clarke G, Dinan TG, Cryan JF. Growing up in a bubble: using germ-free animals to assess the influence of the gut microbiota on brain and behavior. *Int J Neuropsychopharmacol* (2016) 19:1–17. doi: 10.1093/ijnp/pyw020

124. Heijtz RD, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, et al. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci* (2011) 108:3047–52. doi: 10.1073/pnas.1010529108

125. Liu L, Huh JR, Shah K. Microbiota and the gut-brain-axis: implications for new therapeutic design in the CNS. *eBioMedicine* (2022) 77:103908. doi: 10.1016/j.ebiom.2022.103908

126. Mudd AT, Berding K, Wang M, Donovan SM, Dilger RN. Serum cortisol mediates the relationship between fecal ruminococcus and brain n-acetylaspartate in the young pig. *Gut Microbes* (2017) 8:589–600. doi: 10.1080/19490976.2017.1353849

127. Fung TC, Olson CA, Hsiao EY. Interactions between the microbiota, immune and nervous systems in health and disease. *Nat Neurosci* (2017) 20:145–55. doi: 10.1038/nn.4476

128. Sherwin E, Dinan TG, Cryan JF. Recent developments in understanding the role of the gut microbiota in brain health and disease. *Ann N Y Acad Sci* (2018) 1420:5–25. doi: 10.1111/nyas.13416

129. Calvani R, Picca A, Lo Monaco MR, Landi F, Bernabei R, Marzetti E. Of microbes and minds: a narrative review on the second brain aging. *Front Med* (2018) 5:53. doi: 10.3389/fmed.2018.00053

130. Tan J, McKenzie C, Potamitis M, Thorburn AN, Mackay CR, Macia L. The role of short-chain fatty acids in health and disease. *Adv Immunol* (2014) 121:91–119. doi: 10.1016/B978-0-12-800100-4.00003-9

131. Stilling RM, van de Wouw M, Clarke G, Stanton C, Dinan TG, Cryan JF. The neuropharmacology of butyrate: the bread and butter of the microbiota-gut-brain axis? *Neurochem Int* (2016) 99:110–32. doi: 10.1016/j.neuint.2016.06.011

132. Dalile B, Van Oudenhove L, Vervliet B, Verbeke K. The role of short-chain fatty acids in microbiota-gut-brain communication. *Nat Rev Gastroenterol Hepatol* (2019) 16:461–78. doi: 10.1038/s41575-019-0157-3

133. Rifai N, Horvath AR, Wittwer CT. Tietz textbook of clinical chemistry and molecular diagnostics. Sixth Edit. St. Louis, Missouri: Elsevier, Inc (2018).

134. Buckingham JC, Christian HC, Gillies GE, Philip JG, Taylor AD. The physiology of immunity. In: *Progress in Brain Research* (1996), 331–54.

135. Buckingham JC. Glucocorticoids: exemplars of multi-tasking. Br J Pharmacol (2006) 147 Suppl:S258-68. doi: 10.1038/sj.bjp.0706456

136. Smith SM, Vale WW. The role of the hypothalamic-pituitary-adrenal axis in neuroendocrine responses to stress. *Dialogues Clin Neurosci* (2006) 8:383–95. doi: 10.31887/DCNS.2006.8.4/ssmith

137. Koning ASCAM, Buurstede JC, Van Weert LTCM, Meijer OC. Glucocorticoid and mineralocorticoid receptors in the brain: a transcriptional perspective. *J Endocr Soc* (2019) 3:1917–30. doi: 10.1210/js.2019-00158

138. Oster H, Challet E, Ott V, Arvat E, de Kloet ER, Dijk DJ, et al. The functional and clinical significance of the 24-hour rhythm of circulating glucocorticoids. *Endocr Rev* (2017) 38:3–45. doi: 10.1210/er.2015-1080

139. Strehl C, Ehlers L, Gaber T, Buttgereit F. Glucocorticoids–All-Rounders tackling the versatile players of the immune system. *Front Immunol* (2019) 10:1744. doi: 10.3389/fimmu.2019.01744

140. Tsigos C, Chrousos GP. Hypothalamic-pituitary-adrenal axis, neuroendocrine factors and stress. J Psychosom Res (2002) 53:865–71. doi: 10.1016/S0022-3999(02)00429-4

141. Meijer OC, Buurstede JC, Schaaf MJM. Corticosteroid receptors in the brain: transcriptional mechanisms for specificity and context-dependent effects. *Cell Mol Neurobiol* (2019) 39:539–49. doi: 10.1007/s10571-018-0625-2

142. Mawdsley JE, Rampton DS. Psychological stress in IBD: new insights into pathogenic and therapeutic implications. *Gut* (2005) 54:1481 LP – 1491. doi: 10.1136/gut.2005.064261

143. Pivonello R, Simeoli C, De Martino MC, Cozzolino A, De Leo M, Iacuaniello D, et al. Neuropsychiatric disorders in cushing's syndrome. *Front Neurosci* (2015) 9:129. doi: 10.3389/fnins.2015.00129

144. Anglin RE, Rosebush PI, Mazurek MF. The neuropsychiatric profile of addison's disease: revisiting a forgotten phenomenon. J Neuropsychiatry Clin Neurosci (2006) 18:450–9. doi: 10.1176/jnp.2006.18.4.450

145. Misiak B, Łoniewski I, Marlicz W, Frydecka D, Szulc A, Rudzki L, et al. The HPA axis dysregulation in severe mental illness: can we shift the blame to gut microbiota? *Prog Neuropsychopharmacol Biol Psychiatry* (2020) 102:1-9. doi: 10.1016/j.pnpbp.2020.109951

146. Gur TL, Palkar AV, Rajasekera T, Allen J, Niraula A, Godbout J, et al. Prenatal stress disrupts social behavior, cortical neurobiology and commensal microbes in adult male offspring. *Behav Brain Res* (2019) 359:886–94. doi: 10.1016/j.bbr.2018.06.025

147. Zijlmans MAC, Korpela K, Riksen-Walraven JM, de Vos WM, de Weerth C. Maternal prenatal stress is associated with the infant intestinal microbiota. *Psychoneuroendocrinology* (2015) 53:233-45. doi: 10.1016/j.psyneuen.2015.01.006

148. Jang H-M, Lee K-E, Lee H-J, Kim D-H. Immobilization stress-induced escherichia coli causes anxiety by inducing NF-κB activation through gut microbiota disturbance. *Sci Rep* (2018) 8:1–14. doi: 10.1038/s41598-018-31764-0

149. Maniam J, Antoniadis C, Morris MJ. Early-life stress, HPA axis adaptation, and mechanisms contributing to later health outcomes. *Front Endocrinol* (2014) 5:73. doi: 10.3389/fendo.2014.00073

150. de Kloet ER, Joëls M, Holsboer F. Stress and the brain: from adaptation to disease. *Nat Rev Neurosci* (2005) 6:463–75. doi: 10.1038/nrn1683

151. Liu B, Zhang T-N, Knight JK, Goodwin JE. The glucocorticoid receptor in cardiovascular health and disease. *Cells* (2019) 8:1227. doi: 10.3390/cells8101227

152. Karemaker JM. An introduction into autonomic nervous function. *Physiol Meas* (2017) 38:R89-R118. doi: 10.1088/1361-6579/aa6782

153. Wehrwein EA, Orer HS, Barman SM. Overview of the anatomy, physiology, and pharmacology of the autonomic nervous system. *Compr Physiol* (2016) 6:1239–78. doi: 10.1002/cphy.c150037

154. Browning KN, Travagli RA. Central nervous system control of gastrointestinal motility and secretion and modulation of gastrointestinal functions. In: *Comprehensive physiology* (2014) 4(4):1339–68. doi: 10.1002/cphy.c130055

155. Mayer EA, Tillisch K, Gupta A. Gut/brain axis and the microbiota. J Clin Invest (2015) 125:926-38. doi: 10.1172/JCI76304

156. Rao M, Gershon MD. The bowel and beyond: the enteric nervous system in neurological disorders. *Nat Rev Gastroenterol Hepatol* (2016) 13:517–28. doi: 10.1038/ nrgastro.2016.107

157. Furness JB, Callaghan BP, Rivera LR, Cho H-J. The enteric nervous system and gastrointestinal innervation: integrated local and central control BT - microbial endocrinology: the microbiota-Gut-Brain axis in health and disease. In: *Advances in Experimental Medicine and Biology* Lyte M, Cryan JF, editors. New York, NY: Springer New York (2014). p. 39–71. doi: 10.1007/978-1-4939-0897-4_3

158. Fleming MA, Ehsan L, Moore SR, Levin DE. The enteric nervous system and its emerging role as a therapeutic target. *Gastroenterol Res Pract* (2020) 2020:1–13. doi: 10.1155/2020/8024171

159. Breit S, Kupferberg A, Rogler G, Hasler G. Vagus nerve as modulator of the brain-gut axis in psychiatric and inflammatory disorders. *Front Psychiatry* (2018) 9:44. doi: 10.3389/fpsyt.2018.00044

160. Bohórquez DV, Shahid RA, Erdmann A, Kreger AM, Wang Y, Calakos N, et al. Neuroepithelial circuit formed by innervation of sensory enteroendocrine cells. *J Clin Invest* (2015) 125:782–6. doi: 10.1172/JCI78361

161. Annahazi A, Schemann M The enteric nervous system: "a little brain in the gut." *Neuroforum* (2020) 26:31–42 doi: 10.1515/nf-2019-0027

162. McVey Neufeld KA, Perez-Burgos A, Mao YK, Bienenstock J, Kunze WA. The gut microbiome restores intrinsic and extrinsic nerve function in germ-free mice accompanied by changes in calbindin. *Neurogastroenterol Motil* (2015) 27:627–36. doi: 10.1111/nmo.12534

163. Kabouridis PS, Lasrado R, McCallum S, Chng SH, Snippert HJ, Clevers H, et al. Microbiota controls the homeostasis of glial cells in the gut lamina propria. *Neuron* (2015) 85:289–95. doi: 10.1016/j.neuron.2014.12.037

164. Yano JM, Yu K, Donaldson GP, Shastri GG, Ann P, Ma L, et al. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* (2015) 161:264–76. doi: 10.1016/j.cell.2015.02.047

165. Muller PA, Schneeberger M, Matheis F, Wang P, Kerner Z, Ilanges A, et al. Microbiota modulate sympathetic neurons *via* a gut-brain circuit. *Nature* (2020) 583:441-6. doi: 10.1038/s41586-020-2474-7

166. Obata Y, Castaño Á, Boeing S, Bon-Frauches AC, Fung C, Fallesen T, et al. Neuronal programming by microbiota regulates intestinal physiology. *Nature* (2020) 578:284–9. doi: 10.1038/s41586-020-1975-8

167. Browning KN, Verheijden S, Boeckxstaens GE. The vagus nerve in appetite regulation, mood, and intestinal inflammation. *Gastroenterology* (2017) 152:730–44. doi: 10.1053/j.gastro.2016.10.046

168. Kaelberer MM, Buchanan KL, Klein ME, Barth BB, Montoya MM, Shen X, et al. A gut-brain neural circuit for nutrient sensory transduction. *Science* (2018) 361: eaat5236. doi: 10.1126/science.aat5236

169. Bonaz B, Sinniger V, Pellissier S. The vagus nerve in the neuro-immune axis: implications in the pathology of the gastrointestinal tract. *Front Immunol* (2017) 8:1452. doi: 10.3389/fimmu.2017.01452

170. Goverse G, Stakenborg M, Matteoli G. The intestinal cholinergic antiinflammatory pathway. J Physiol (2016) 594:5771–80. doi: 10.1113/JP271537

171. Hansen MK, O'Connor KA, Goehler LE, Watkins LR, Maier SF. The contribution of the vagus nerve in interleukin- 1β -induced fever is dependent on dose. *Am J Physiol Integr Comp Physiol* (2001) 280:R929–34. doi: 10.1152/ajpregu.2001.280.4.R929

172. Marquette C, Linard C, Galonnier M, van Uye A, Mathieu J, Gourmelon P, et al. IL-1β, TNFα and IL-6 induction in the rat brain after partial-body irradiation: role of vagal afferents. *Int J Radiat Biol* (2003) 79:777–85. doi: 10.1080/09553000310001610998

173. de La Serre CB, de Lartigue G, Raybould HE. Chronic exposure to low dose bacterial lipopolysaccharide inhibits leptin signaling in vagal afferent neurons. *Physiol Behav* (2015) 139:188–94. doi: 10.1016/j.physbeh.2014.10.032

174. Goehler LE, Gaykema RPA, Hansen MK, Anderson K, Maier SF, Watkins LR. Vagal immune-to-brain communication: a visceral chemosensory pathway. *Auton Neurosci* (2000) 85:49–59. doi: 10.1016/S1566-0702(00)00219-8

175. Haddad JJ, Saadé NE, Safieh-Garabedian B. Cytokines and neuro-immuneendocrine interactions: a role for the hypothalamic-pituitary-adrenal revolving axis. J Neuroimmunol (2002) 133:1-19. doi: 10.1016/S0165-5728(02)00357-0

176. Silverman MN, Pearce BD, Biron CA, Miller AH. Immune modulation of the hypothalamic-Pituitary-Adrenal (HPA) axis during viral infection. *Viral Immunol* (2005) 18:41–78. doi: 10.1089/vim.2005.18.41

177. Bellinger DL, Millar BA, Perez S, Carter J, Wood C, ThyagaRajan S, et al. Sympathetic modulation of immunity: relevance to disease. *Cell Immunol* (2008) 252:27–56. doi: 10.1016/j.cellimm.2007.09.005

178. Goehler LE, Gaykema RPA, Opitz N, Reddaway R, Badr N, Lyte M. Activation in vagal afferents and central autonomic pathways: early responses to intestinal infection with campylobacter jejuni. *Brain Behav Immun* (2005) 19:334–44. doi: 10.1016/j.bbi.2004.09.002

179. Cailotto C, Costes LMM, van der Vliet J, VAN Bree SHW, VAN Heerikhuize JJ, Buijs RM, et al. Neuroanatomical evidence demonstrating the existence of the vagal anti-inflammatory reflex in the intestine. *Neurogastroenterol Motil* (2012) 24:191–e93. doi: 10.1111/j.1365-2982.2011.01824.x

 Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, et al. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* (2000) 405:458–62. doi: 10.1038/35013070

181. Tracey KJ. The inflammatory reflex. *Nature* (2002) 420:853-9. doi: 10.1038/ nature01321

182. Tracey KJ. Reflex control of immunity. Nat Rev Immunol (2009) 9:418–28. doi: 10.1038/nri2566

183. Suarez AN, Hsu TM, Liu CM, Noble EE, Cortella AM, Nakamoto EM, et al. Gut vagal sensory signaling regulates hippocampus function through multi-order pathways. *Nat Commun* (2018) 9:2181. doi: 10.1038/s41467-018-04639-1

184. Clark KB, Smith DC, Hassert DL, Browning RA, Naritoku DK, Jensen RA. Posttraining electrical stimulation of vagal afferents with concomitant vagal efferent inactivation enhances memory storage processes in the rat. *Neurobiol Learn Mem* (1998) 70:364–73. doi: 10.1006/nlme.1998.3863

185. Clark KB, Naritoku DK, Smith DC, Browning RA, Jensen RA. Enhanced recognition memory following vagus nerve stimulation in human subjects. *Nat Neurosci* (1999) 2:94–8. doi: 10.1038/4600

186. Follesa P, Biggio F, Gorini G, Caria S, Talani G, Dazzi L, et al. Vagus nerve stimulation increases norepinephrine concentration and the gene expression of BDNF and bFGF in the rat brain. *Brain Res* (2007) 1179:28–34. doi: 10.1016/j.brainres.2007.08.045

187. Biggio F, Gorini G, Utzeri C, Olla P, Marrosu F, Mocchetti I, et al. Chronic vagus nerve stimulation induces neuronal plasticity in the rat hippocampus. *Int J Neuropsychopharmacol* (2009) 12:1209–21. doi: 10.1017/S1461145709000200

188. Noble EE, Billington CJ, Kotz CM, Wang C. The lighter side of BDNF. Am J Physiol Integr Comp Physiol (2011) 300:R1053–69. doi: 10.1152/ajpregu.00776.2010

189. Neufeld KM, Kang N, Bienenstock J, Foster JA. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2011) 23:255–64, e119. doi: 10.1111/j.1365-2982.2010.01620.x

190. Bravo JA, Julio-Pieper M, Forsythe P, Kunze W, Dinan TG, Bienenstock J, et al. Communication between gastrointestinal bacteria and the nervous system. *Curr Opin Pharmacol* (2012) 12:667–72. doi: 10.1016/j.coph.2012.09.010

191. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse *via* the vagus nerve. *Proc Natl Acad Sci* (2011) 108:16050–5. doi: 10.1073/pnas.1102999108

192. Bercik P, Park AJ, Sinclair D, Khoshdel A, Lu J, Huang X, et al. The anxiolytic effect of bifdobacterium longum NCC3001 involves vagal pathways for gut-brain communication. *Neurogastroenterol Motil* (2011) 23:1132–9. doi: 10.1111/j.1365-2982.2011.01796.x

193. Vazquez E, Barranco A, Ramirez M, Gruart A, Delgado-Garcia JM, Jimenez ML, et al. Dietary 2'-fucosyllactose enhances operant conditioning and long-term potentiation *via* gut-brain communication through the vagus nerve in rodents. *PLoS ONE* (2016) 11(11):e0166070. doi: 10.1371/journal.pone.0166070

194. Zaragozá R. Transport of amino acids across the blood-brain barrier. *Front Physiol* (2020) 11:973. doi: 10.3389/fphys.2020.00973

195. Gao K, Pi Y, Mu C-L, Peng Y, Huang Z, Zhu W-Y. Antibiotics-induced modulation of large intestinal microbiota altered aromatic amino acid profile and expression of neurotransmitters in the hypothalamus of piglets. *J Neurochem* (2018) 146:219–34. doi: 10.1111/jnc.14333

196. Fujisaka S, Avila-Pacheco J, Soto M, Kostic A, Dreyfuss JM, Pan H, et al. Diet, genetics, and the gut microbiome drive dynamic changes in plasma metabolites. *Cell Rep* (2018) 22:3072–86. doi: 10.1016/j.celrep.2018.02.060

197. Agus A, Planchais J, Sokol H. Gut microbiota regulation of tryptophan metabolism in health and disease. *Cell Host Microbe* (2018) 23:716–24. doi: 10.1016/j.chom.2018.05.003

198. Kennedy PJ, Cryan JF, Dinan TG, Clarke G. Kynurenine pathway metabolism and the microbiota-gut-brain axis. *Neuropharmacology* (2017) 112:399-412. doi: 10.1016/j.neuropharm.2016.07.002

199. Stone TW, Forrest CM, Darlington LG. Kynurenine pathway inhibition as a therapeutic strategy for neuroprotection. *FEBS J* (2012) 279:1386–97. doi: 10.1111/j.1742-4658.2012.08487.x

200. Ruddick JP, Evans AK, Nutt DJ, Lightman SL, Rook GAW, Lowry CA. Tryptophan metabolism in the central nervous system: medical implications. *Expert Rev Mol Med* (2006) 8:1–27. doi: 10.1017/S1462399406000068

201. Gao J, Xu K, Liu H, Liu G, Bai M, Peng C, et al. Impact of the gut microbiota on intestinal immunity mediated by tryptophan metabolism. *Front Cell Infect Microbiol* (2018) 8:13. doi: 10.3389/fcimb.2018.00013

202. Lamas B, Hernandez-Galan L, Galipeau HJ, Constante M, Clarizio A, Jury J, et al. Aryl hydrocarbon receptor ligand production by the gut microbiota is decreased in celiac disease leading to intestinal inflammation. *Sci Transl Med* (2020) 12:eaba0624. doi: 10.1126/scitranslmed.aba0624

203. Lee J-H, Lee J. Indole as an intercellular signal in microbial communities. *FEMS Microbiol Rev* (2010) 34:426–44. doi: 10.1111/j.1574-6976.2009.00204.x

204. Li G, Young KD. Indole production by the tryptophanase TnaA in escherichia coli is determined by the amount of exogenous tryptophan. *Microbiology* (2013) 159:402–10. doi: 10.1099/mic.0.064139-0

205. Roth W, Zadeh K, Vekariya R, Ge Y, Mohamadzadeh M. Tryptophan metabolism and gut-brain homeostasis. *Int J Mol Sci* (2021) 22:2973. doi: 10.3390/ ijms22062973

206. Bosi A, Banfi D, Bistoletti M, Giaroni C, Baj A. Tryptophan metabolites along the microbiota-Gut-Brain axis: an interkingdom communication system influencing the gut in health and disease. *Int J Tryptophan Res* (2020) 13:1178646920928984. doi: 10.1177/1178646920928984

207. Kiser D, Steemers B, Branchi I, Homberg JR. The reciprocal interaction between serotonin and social behaviour. *Neurosci Biobehav Rev* (2012) 36:786–98. doi: 10.1016/j.neubiorev.2011.12.009

208. Homberg JR. Serotonin and decision making processes. *Neurosci Biobehav Rev* (2012) 36:218–36. doi: 10.1016/j.neubiorev.2011.06.001

209. Schwarcz R, Bruno JP, Muchowski PJ, Wu H-Q. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat Rev Neurosci* (2012) 13:465–77. doi: 10.1038/nrn3257

210. Szalardy L, Zadori D, Toldi J, Fulop F, Klivenyi P, Vecsei L. Manipulating kynurenic acid levels in the brain - on the edge between neuroprotection and cognitive dysfunction. *Curr Top Med Chem* (2012) 12:1797–806. doi: 10.2174/1568026611209061797

211. Masson J, Emerit MB, Hamon M, Darmon M. Serotonergic signaling: multiple effectors and pleiotropic effects. *Wiley Interdiscip Rev Membr Transp Signal* (2012) 1:685–713. doi: 10.1002/wmts.50

212. Mawe GM, Hoffman JM. Serotonin signalling in the gut-functions, dysfunctions and therapeutic targets. *Nat Rev Gastroenterol Hepatol* (2013) 10:473-86. doi: 10.1038/nrgastro.2013.105

213. Fung TC, Vuong HE, Luna CDG, Pronovost GN, Aleksandrova AA, Riley NG, et al. Intestinal serotonin and fluoxetine exposure modulate bacterial colonization in the gut. *Nat Microbiol* (2019) 4:2064–73. doi: 10.1038/s41564-019-0540-4

214. Hata T, Asano Y, Yoshihara K, Kimura-Todani T, Miyata N, Zhang X-T, et al. Regulation of gut luminal serotonin by commensal microbiota in mice. *PloS One* (2017) 12:e0180745. doi: 10.1371/journal.pone.0180745

215. Berger M, Gray JA, Roth BL. The expanded biology of serotonin. Annu Rev Med (2009) 60:355–66. doi: 10.1146/annurev.med.60.042307.110802

216. Carrasco GA, Van de Kar LD. Neuroendocrine pharmacology of stress. Eur J Pharmacol (2003) 463:235-72. doi: 10.1016/s0014-2999(03)01285-8

217. Heisler LK, Pronchuk N, Nonogaki K, Zhou L, Raber J, Tung L, et al. Serotonin activates the hypothalamic–Pituitary–Adrenal axis via serotonin 2C receptor stimulation. J Neurosci (2007) 27:6956 LP – 6964. doi: 10.1523/JNEUROSCI.2584-06.2007

218. Ohta K-I, Miki T, Warita K, Suzuki S, Kusaka T, Yakura T, et al. Prolonged maternal separation disturbs the serotonergic system during early brain development. *Int J Dev Neurosci Off J Int Soc Dev Neurosci* (2014) 33:15–21. doi: 10.1016/jijdevneu.2013.10.007

219. Sood A, Pati S, Bhattacharya A, Chaudhari K, Vaidya VA. Early emergence of altered 5-HT(2A) receptor-evoked behavior, neural activation and gene expression following maternal separation. *Int J Dev Neurosci Off J Int Soc Dev Neurosci* (2018) 65:21–8. doi: 10.1016/j.ijdevneu.2017.10.005

220. Huang Y, Xu H, Li H, Yang H, Chen Y, Shi X. Pre-gestational stress reduces the ratio of 5-HIAA to 5-HT and the expression of 5-HT1A receptor and serotonin transporter in the brain of foetal rat. *BMC Neurosci* (2012) 13:22. doi: 10.1186/1471-2202-13-22

221. Gershon MD, Tack J. The serotonin signaling system: from basic understanding to drug development for functional GI disorders. *Gastroenterology* (2007) 132:397-414. doi: 10.1053/j.gastro.2006.11.002

222. Reigstad CS, Salmonson CE, Rainey JF3rd, Szurszewski JH, Linden DR, Sonnenburg JL, et al. Gut microbes promote colonic serotonin production through an effect of short-chain fatty acids on enterochromaffin cells. FASEB J Off Publ Fed Am Soc Exp Biol (2015) 29:1395–403. doi: 10.1096/fj.14-259598

223. Wikoff WR, Anfora AT, Liu J, Schultz PG, Lesley SA, Peters EC, et al. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc Natl Acad Sci* (2009) 106:3698–703. doi: 10.1073/pnas.0812874106

224. Desbonnet L, Garrett L, Clarke G, Bienenstock J, Dinan TG. The probiotic bifidobacteria infantis: an assessment of potential antidepressant properties in the rat. *J Psychiatr Res* (2008) 43:164–74. doi: 10.1016/j.jpsychires.2008.03.009

225. Eisenhofer G, Kopin IJ, Goldstein DS. Catecholamine metabolism: a contemporary view with implications for physiology and medicine. *Pharmacol Rev* (2004) 56:331-49. doi: 10.1124/pr.56.3.1

226. Borodovitsyna O, Flamini M, Chandler D. Noradrenergic modulation of cognition in health and disease. *Neural Plast* (2017) 2017;6031478. doi: 10.1155/2017/6031478

227. Herman JP, McKlveen JM, Ghosal S, Kopp B, Wulsin A, Makinson R, et al. Regulation of the hypothalamic-Pituitary-Adrenocortical stress response. Compr Physiol (2016) 6(2):603-21. doi: 10.1002/cphy.c150015

228. Lyte M, Vulchanova L, Brown DR. Stress at the intestinal surface: catecholamines and mucosa-bacteria interactions. *Cell Tissue Res* (2011) 343:23-32. doi: 10.1007/s00441-010-1050-0

229. Sperandio V, Torres AG, Jarvis B, Nataro JP, Kaper JB. Bacteria-host communication: the language of hormones. *Proc Natl Acad Sci* (2003) 100:8951-6. doi: 10.1073/pnas.1537100100

230. Lyte M, Ernst S. Catecholamine induced growth of gram negative bacteria. *Life Sci* (1992) 50:203–12. doi: 10.1016/0024-3205(92)90273-r

231. Asano Y, Hiramoto T, Nishino R, Aiba Y, Kimura T, Yoshihara K, et al. Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. *Am J Physiol Gastrointest Liver Physiol* (2012) 303:G1288–95. doi: 10.1152/ajpgi.00341.2012

232. Jiménez-Balado J, Eich TS. GABAergic dysfunction, neural network hyperactivity and memory impairments in human aging and alzheimer's disease. *Semin Cell Dev Biol* (2021) 116:146–59. doi: 10.1016/j.semcdb.2021.01.005

233. Miklós IH, Kovács KJ. GABAergic innervation of corticotropin-releasing hormone (CRH)-secreting parvocellular neurons and its plasticity as demonstrated by quantitative immunoelectron microscopy. *Neuroscience* (2002) 113:581–92. doi: 10.1016/s0306-4522(02)00147-1

234. Camille Melón L, Maguire J. GABAergic regulation of the HPA and HPG axes and the impact of stress on reproductive function. *J Steroid Biochem Mol Biol* (2016) 160:196–203. doi: 10.1016/j.jsbmb.2015.11.019

235. Strandwitz P, Kim KH, Terekhova D, Liu JK, Sharma A, Levering J, et al. GABA-modulating bacteria of the human gut microbiota. *Nat Microbiol* (2019) 4:396–403. doi: 10.1038/s41564-018-0307-3

236. Tillisch K, Mayer EA, Gupta A, Gill Z, Brazeilles R, Le Nevé B, et al. Brain structure and response to emotional stimuli as related to gut microbial profiles in healthy women. *Psychosom Med* (2017) 79:905-13. doi: 10.1097/PSY.0000000000000493

237. Frost G, Sleeth ML, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, et al. The short-chain fatty acid acetate reduces appetite *via* a central homeostatic mechanism. *Nat Commun* (2014) 5:3611. doi: 10.1038/ncomms4611

238. Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, et al. Endocannabinoids–at the crossroads between the gut microbiota and host metabolism. *Nat Rev Endocrinol* (2016) 12:133–43. doi: 10.1038/nrendo.2015.211

239. Cristino L, Bisogno T, Di Marzo V. Cannabinoids and the expanded endocannabinoid system in neurological disorders. *Nat Rev Neurol* (2020) 16:9–29. doi: 10.1038/s41582-019-0284-z

240. Alger BE. Retrograde signaling in the regulation of synaptic transmission: focus on endocannabinoids. *Prog Neurobiol* (2002) 68:247–86. doi: 10.1016/s0301-0082(02) 00080-1

241. Micale V, Drago F. Endocannabinoid system, stress and HPA axis. Eur J Pharmacol (2018) 834:230–9. doi: 10.1016/j.ejphar.2018.07.039

242. Zanettini C, Panlilio LV, Alicki M, Goldberg SR, Haller J, Yasar S. Effects of endocannabinoid system modulation on cognitive and emotional behavior. *Front Behav Neurosci* (2011) 5:57. doi: 10.3389/fnbeh.2011.00057

243. Muccioli GG, Naslain D, Bäckhed F, Reigstad CS, Lambert DM, Delzenne NM, et al. The endocannabinoid system links gut microbiota to adipogenesis. *Mol Syst Biol* (2010) 6:392. doi: 10.1038/msb.2010.46

244. Donvito G, Nass SR, Wilkerson JL, Curry ZA, Schurman LD, Kinsey SG, et al. The endogenous cannabinoid system: a budding source of targets for treating inflammatory and neuropathic pain. *Neuropsychopharmacol Off Publ Am Coll Neuropsychopharmacol* (2018) 43:52–79. doi: 10.1038/npp.2017.204

245. Wang J, Ueda N. Role of the endocannabinoid system in metabolic control. Curr Opin Nephrol Hypertens (2008) 17:1-10. doi: 10.1097/MNH.0b013e3282f29071

246. Srivastava RK, Lutz B, Ruiz de Azua I. The microbiome and gut endocannabinoid system in the regulation of stress responses and metabolism. *Front Cell Neurosci* (2022) 16:867267. doi: 10.3389/fncel.2022.867267

247. Hillard CJ, Beatka M, Sarvaideo J. Endocannabinoid signaling and the hypothalamic-Pituitary-Adrenal axis. *Compr Physiol* (2016) 7:1–15. doi: 10.1002/cphy.c160005

248. Cuddihey H, MacNaughton WK, Sharkey KA. Role of the endocannabinoid system in the regulation of intestinal homeostasis. *Cell Mol Gastroenterol Hepatol* (2022) 14:947–63. doi: 10.1016/j.jcmgh.2022.05.015

249. Zou S, Kumar U. Cannabinoid receptors and the endocannabinoid system: signaling and function in the central nervous system. *Int J Mol Sci* (2018) 19:833. doi: 10.3390/ijms19030833

250. de Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: mechanistic insights. *Gut* (2022) 71:1020–32. doi: 10.1136/gutjnl-2021-326789

251. Manca C, Boubertakh B, Leblanc N, Deschênes T, Lacroix S, Martin C, et al. Germ-free mice exhibit profound gut microbiota-dependent alterations of intestinal endocannabinoidome signaling. *J Lipid Res* (2020) 61:70–85. doi: 10.1194/ jlr.RA119000424

252. Vijay A, Kouraki A, Gohir S, Turnbull J, Kelly A, Chapman V, et al. The antiinflammatory effect of bacterial short chain fatty acids is partially mediated by endocannabinoids. *Gut Microbes* (2021) 13:1997559. doi: 10.1080/ 19490976.2021.1997559

253. Markey L, Hooper A, Melon LC, Baglot S, Hill MN, Maguire J, et al. Colonization with the commensal fungus candida albicans perturbs the gut-brain axis through dysregulation of endocannabinoid signaling. *Psychoneuroendocrinology* (2020) 121:104808. doi: 10.1016/j.psyneuen.2020.104808

254. Bisogno T, Lauritano A, Piscitelli F. The endocannabinoid system: a bridge between alzheimer's disease and gut microbiota. *Life* (2021) 11:1–18. doi: 10.3390/ life11090934

255. Cohen LJ, Esterhazy D, Kim S-H, Lemetre C, Aguilar RR, Gordon EA, et al. Commensal bacteria make GPCR ligands that mimic human signalling molecules. *Nature* (2017) 549:48–53. doi: 10.1038/nature23874

256. Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. Cell Res (2020) 30:492-506. doi: 10.1038/s41422-020-0332-7

257. Zhang H, Zhang Z, Liao Y, Zhang W, Tang D. The complex link and disease between the gut microbiome and the immune system in infants. *Front Cell Infect Microbiol* (2022) 12:924119. doi: 10.3389/fcimb.2022.924119

258. Negi S, Das DK, Pahari S, Nadeem S, Agrewala JN. Potential role of gut microbiota in induction and regulation of innate immune memory. *Front Immunol* (2019) 10:2441. doi: 10.3389/fimmu.2019.02441

259. Mou Y, Du Y, Zhou L, Yue J, Hu X, Liu Y, et al. Gut microbiota interact with the brain through systemic chronic inflammation: implications on neuroinflammation, neurodegeneration, and aging. *Front Immunol* (2022) 13:796288. doi: 10.3389/fimmu.2022.796288

260. Cahenzli J, Balmer ML, McCoy KD. Microbial-immune cross-talk and regulation of the immune system. *Immunology* (2013) 138:12–22. doi: 10.1111/j.1365-2567.2012.03624.x

261. Chelakkot C, Ghim J, Ryu SH. Mechanisms regulating intestinal barrier integrity and its pathological implications. *Exp Mol Med* (2018) 50:1–9. doi: 10.1038/s12276-018-0126-x

262. Logsdon AF, Erickson MA, Rhea EM, Salameh TS, Banks WA. Gut reactions: how the blood-brain barrier connects the microbiome and the brain. *Exp Biol Med (Maywood)* (2018) 243:159–65. doi: 10.1177/1535370217743766

263. Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Tóth M, et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med* (2014) 6:263ra158. doi: 10.1126/scitranslmed.3009759

264. Perry VH, Cunningham C, Holmes C. Systemic infections and inflammation affect chronic neurodegeneration. *Nat Rev Immunol* (2007) 7:161–7. doi: 10.1038/ nri2015

265. Perry VH, Newman TA, Cunningham C. The impact of systemic infection on the progression of neurodegenerative disease. *Nat Rev Neurosci* (2003) 4:103–12. doi: 10.1038/nrn1032

266. Sankowski R, Mader S, Valdés-Ferrer SI. Systemic inflammation and the brain: novel roles of genetic, molecular, and environmental cues as drivers of neurodegeneration. *Front Cell Neurosci* (2015) 9:28. doi: 10.3389/fncel.2015.00028

267. Stanisavljević S, Čepić A, Bojić S, Veljović K, Mihajlović S, Đedović N, et al. Oral neonatal antibiotic treatment perturbs gut microbiota and aggravates central nervous system autoimmunity in dark agouti rats. *Sci Rep* (2019) 9:918. doi: 10.1038/ s41598-018-37505-7

268. Minter MR, Zhang C, Leone V, Ringus DL, Zhang X, Oyler-Castrillo P, et al. Antibiotic-induced perturbations in gut microbial diversity influences neuro-inflammation and amyloidosis in a murine model of alzheimer's disease. *Sci Rep* (2016) 6:30028. doi: 10.1038/srep30028

269. Minter MR, Hinterleitner R, Meisel M, Zhang C, Leone V, Zhang X, et al Antibiotic-induced perturbations in microbial diversity during post-natal development alters amyloid pathology in an aged APP SWE /PS1 ΔE9 murine model of alzheimer's disease OPEN. *Sci Rep* (2017) 7(1):10411. doi: 10.1038/s41598-017-11047-w

270. Jang H-M, Lee H-J, Jang S-E, Han MJ, Kim D-H. Evidence for interplay among antibacterial-induced gut microbiota disturbance, neuro-inflammation, and anxiety in mice. *Mucosal Immunol* (2018) 11:1386–97. doi: 10.1038/s41385-018-0042-3

271. Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, et al. Host-gut microbiota metabolic interactions. *Sci* (80-) (2012) 336:1262–7. doi: 10.1126/science.1223813

272. Holmes E, Li JV, Marchesi JR, Nicholson JK. Gut microbiota composition and activity in relation to host metabolic phenotype and disease risk. *Cell Metab* (2012) 16:559–64. doi: 10.1016/j.cmet.2012.10.007

273. Charney AN, Micic L, Egnor RW. Nonionic diffusion of short-chain fatty acids across rat colon. Am J Physiol (1998) 274:G518–24. doi: 10.1152/ajpgi.1998.274.3.G518

274. Kelly CJ, Zheng L, Campbell EL, Saeedi B, Scholz CC, Bayless AJ, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe* (2015) 17:662–71. doi: 10.1016/j.chom.2015.03.005

275. Sina C, Gavrilova O, Förster M, Till A, Derer S, Hildebrand F, et al. G Proteincoupled receptor 43 is essential for neutrophil recruitment during intestinal inflammation. J Immunol (2009) 183:7514 LP – 7522. doi: 10.4049/jimmunol.0900063

276. Tolhurst G, Heffron H, Lam YS, Parker HE, Habib AM, Diakogiannaki E, et al. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion *via* the G-Proteincoupled receptor FFAR2. *Diabetes* (2012) 61:364–71. doi: 10.2337/db11-1019

277. Nøhr MK, Egerod KL, Christiansen SH, Gille A, Offermanns S, Schwartz TW, et al. Expression of the short chain fatty acid receptor GPR41/FFAR3 in autonomic and somatic sensory ganglia. *Neuroscience* (2015) 290:126–37. doi: 10.1016/j.neuroscience.2015.01.040

278. Lu VB, Gribble FM, Reimann F. Free fatty acid receptors in enteroendocrine cells. *Endocrinology* (2018) 159:2826–35. doi: 10.1210/en.2018-00261

279. Priyadarshini M, Kotlo KU, Dudeja PK, Layden BT. Role of short chain fatty acid receptors in intestinal physiology and pathophysiology. *Compr Physiol* (2018) 8:1091–115. doi: 10.1002/cphy.c170050

280. Priyadarshini M, Navarro G, Layden BT. Gut microbiota: FFAR reaching effects on islets. *Endocrinology* (2018) 159:2495–505. doi: 10.1210/en.2018-00296

281. Chun E, Lavoie S, Fonseca-Pereira D, Bae S, Michaud M, Hoveyda HR, et al. Metabolite-sensing receptor Ffar2 regulates colonic group 3 innate lymphoid cells and gut immunity. *Immunity* (2019) 51:871–884.e6. doi: 10.1016/j.immuni.2019.09.014

282. Priyadarshini M, Lednovich K, Xu K, Gough S, Wicksteed B, Layden BT. FFAR from the gut microbiome crowd: SCFA receptors in T1D pathology. *Metabolites* (2021) 11:302. doi: 10.3390/metabo11050302

283. Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, et al. The microbial metabolites, short-chain fatty acids, regulate colonic treg cell homeostasis. *Science* (2013) 341:569–73. doi: 10.1126/science.1241165

284. Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* (2013) 504:451–5. doi: 10.1038/nature12726

285. Kim CH. Control of lymphocyte functions by gut microbiota-derived shortchain fatty acids. *Cell Mol Immunol* (2021) 18:1161–71. doi: 10.1038/s41423-020-00625-0

286. Silva YP, Bernardi A, Frozza RL. The role of short-chain fatty acids from gut microbiota in gut-brain communication. *Front Endocrinol* (2020) 11:25. doi: 10.3389/ fendo.2020.00025

287. Li Z, Yi C-X, Katiraei S, Kooijman S, Zhou E, Chung CK, et al. Butyrate reduces appetite and activates brown adipose tissue *via* the gut-brain neural circuit. *Gut* (2018) 67:1269–79. doi: 10.1136/gutjnl-2017-314050

288. Puddu A, Sanguineti R, Montecucco F, Viviani GL. Evidence for the gut microbiota short-chain fatty acids as key pathophysiological molecules improving diabetes. *Mediators Inflammation* (2014) 2014:162021. doi: 10.1155/2014/162021

289. Patnala R, Arumugam TV, Gupta N, Dheen ST. HDAC inhibitor sodium butyrate-mediated epigenetic regulation enhances neuroprotective function of microglia during ischemic stroke. *Mol Neurobiol* (2017) 54:6391–411. doi: 10.1007/s12035-016-0149-z

290. Wang P, Zhang Y, Gong Y, Yang R, Chen Z, Hu W, et al. Sodium butyrate triggers a functional elongation of microglial process *via* akt-small RhoGTPase activation and HDACs inhibition. *Neurobiol Dis* (2018) 111:12–25. doi: 10.1016/j.nbd.2017.12.006

291. van de Wouw M, Boehme M, Lyte JM, Wiley N, Strain C, O'Sullivan O, et al. Short-chain fatty acids: microbial metabolites that alleviate stress-induced brain-gut axis alterations. *J Physiol* (2018) 596:4923–44. doi: 10.1113/JP276431

292. Dalile B, Vervliet B, Bergonzelli G, Verbeke K, Van Oudenhove L. Colondelivered short-chain fatty acids attenuate the cortisol response to psychosocial stress in healthy men: a randomized, placebo-controlled trial. *Neuropsychopharmacology* (2020) 45:2257–66. doi: 10.1038/s41386-020-0732-x

293. Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA. Elevated fecal short chain fatty acid and ammonia concentrations in children with autism spectrum disorder. *Dig Dis Sci* (2012) 57:2096–102. doi: 10.1007/s10620-012-2167-7

294. Ho L, Ono K, Tsuji M, Mazzola P, Singh R, Pasinetti GM. Protective roles of intestinal microbiota derived short chain fatty acids in alzheimer's disease-type beta-amyloid neuropathological mechanisms. *Expert Rev Neurother* (2018) 18:83–90. doi: 10.1080/14737175.2018.1400909

295. Skonieczna-Żydecka K, Grochans E, Maciejewska D, Szkup M, Schneider-Matyka D, Jurczak A, et al. Faecal short chain fatty acids profile is changed in polish depressive women. *Nutrients* (2018) 10:1939. doi: 10.3390/nu10121939

296. Ramírez-Pérez O, Cruz-Ramón V, Chinchilla-López P, Méndez-Sánchez N. The role of the gut microbiota in bile acid metabolism. *Ann Hepatol* (2017) 16:s15–20. doi: 10.5604/01.3001.0010.5494

297. Mertens KL, Kalsbeek A, Soeters MR, Eggink HM. Bile acid signaling pathways from the enterohepatic circulation to the central nervous system. *Front Neurosci* (2017) 11:617. doi: 10.3389/fnins.2017.00617

298. Perino A, Demagny H, Velazquez-Villegas L, Schoonjans K. Molecular physiology of bile acid signaling in health, disease, and aging. *Physiol Rev* (2020) 101:683-731. doi: 10.1152/physrev.00049.2019

299. McMillin M, Frampton G, Quinn M, Divan A, Grant S, Patel N, et al. Suppression of the HPA axis during cholestasis can be attributed to hypothalamic bile acid signaling. *Mol Endocrinol* (2015) 29:1720–30. doi: 10.1210/me.2015-1087

300. Monteiro-Cardoso VF, Corlianò M, Singaraja RR. Bile acids: a communication channel in the gut-brain axis. *NeuroMolecular Med* (2021) 23:99–117. doi: 10.1007/s12017-020-08625-z

301. Trauner M, Boyer JL. Bile salt transporters: molecular characterization, function, and regulation. *Physiol Rev* (2003) 83:633–71. doi: 10.1152/physrev.00027.2002

302. Baloni P, Funk CC, Yan J, Yurkovich JT, Kueider-Paisley A, Nho K, et al. Metabolic network analysis reveals altered bile acid synthesis and metabolism in alzheimer's disease. *Cell Rep Med* (2020) 1:100138. doi: 10.1016/j.xcrm.2020.100138

303. Mulak A. Bile acids as key modulators of the brain-Gut-Microbiota axis in alzheimer's disease. J Alzheimer's Dis (2021) 84:461-77. doi: 10.3233/JAD-210608

304. Huang F, Wang T, Lan Y, Yang L, Pan W, Zhu Y, et al. Deletion of mouse FXR gene disturbs multiple neurotransmitter systems and alters neurobehavior. *Front Behav Neurosci* (2015) 9:70. doi: 10.3389/fnbeh.2015.00070

305. Keitel V, Görg B, Bidmon HJ, Zemtsova I, Spomer L, Zilles K, et al. The bile acid receptor TGR5 (Gpbar-1) acts as a neurosteroid receptor in brain. *Glia* (2010) 58:1794–805. doi: 10.1002/glia.21049

306. Yanovsky Y, Schubring SR, Yao Q, Zhao Y, Li S, May A, et al. Waking action of ursodeoxycholic acid (UDCA) involves histamine and GABAA receptor block. *PloS One* (2012) 7:e42512. doi: 10.1371/journal.pone.0042512

307. Huang F, Pariante CM, Borsini A. From dried bear bile to molecular investigation: a systematic review of the effect of bile acids on cell apoptosis,

oxidative stress and inflammation in the brain, across pre-clinical models of neurological, neurodegenerative and neuropsychiatric disorde. *Brain Behav Immun* (2022) 99:132-46. doi: 10.1016/j.bbi.2021.09.021

308. Yanguas-Casás N, Barreda-Manso MA, Nieto-Sampedro M, Romero-Ramírez L. TUDCA: an agonist of the bile acid receptor GPBAR1/TGR5 with anti-inflammatory effects in microglial cells. *J Cell Physiol* (2017) 232:2231–45. doi: 10.1002/jcp.25742

309. McMillin M, DeMorrow S. Effects of bile acids on neurological function and disease. FASEB J (2016) 30:3658–68. doi: 10.1096/fj.201600275R

310. Duboc H, Taché Y, Hofmann AF. The bile acid TGR5 membrane receptor: from basic research to clinical application. *Dig Liver Dis* (2014) 46:302–12. doi: 10.1016/j.dld.2013.10.021

311. Theiler-Schwetz V, Zaufel A, Schlager H, Obermayer-Pietsch B, Fickert P, Zollner G. Bile acids and glucocorticoid metabolism in health and disease. *Biochim Biophys Acta - Mol Basis Dis* (2019) 1865:243–51. doi: 10.1016/j.bbadis.2018.08.001

312. Perry RJ, Lee S, Ma L, Zhang D, Schlessinger J, Shulman GI. FGF1 and FGF19 reverse diabetes by suppression of the hypothalamic-pituitary-adrenal axis. *Nat Commun* (2015) 6:6980. doi: 10.1038/ncomms7980

313. Forman BM, Goode E, Chen J, Oro AE, Bradley DJ, Perlmann T, et al. Identification of a nuclear receptor that is activated by farnesol metabolites. *Cell* (1995) 81:687–93. doi: 10.1016/0092-8674(95)90530-8

314. Kawamata Y, Fujii R, Hosoya M, Harada M, Yoshida H, Miwa M, et al. A G protein-coupled receptor responsive to bile acids*. J Biol Chem (2003) 278:9435–40. doi: 10.1074/jbc.M209706200

315. Baptissart M, Vega A, Martinot E, Baron S, Lobaccaro J-MA, Volle DH. Farnesoid X receptor alpha: a molecular link between bile acids and steroid signaling? *Cell Mol Life Sci* (2013) 70:4511–26. doi: 10.1007/s00018-013-1387-0

316. Stocco DM, Clark BJ. Regulation of the acute production of steroids in steroidogenic cells. *Endocr Rev* (1996) 17:221-44. doi: 10.1210/edrv-17-3-221

317. Schoeler M, Caesar R. Dietary lipids, gut microbiota and lipid metabolism. *Rev Endocr Metab Disord* (2019) 20:461–72. doi: 10.1007/s11154-019-09512-0

318. Peter K, Koji K, Kanako M, Atsushi Y. Mechanism of growth inhibition by free bile acids in lactobacilli and bifidobacteria. *J Bacteriol* (2006) 188:1979–86. doi: 10.1128/JB.188.5.1979-1986.2006

319. Katafuchi T, Makishima M. Molecular basis of bile acid-FXR-FGF15/19 signaling axis. Int J Mol Sci (2022) 23:6046. doi: 10.3390/ijms23116046

320. Grant SM, DeMorrow S. Bile acid signaling in neurodegenerative and neurological disorders. Int J Mol Sci (2020) 21:5982. doi: 10.3390/ijms21175982

321. Jia W, Rajani C, Kaddurah-Daouk R, Li H. Expert insights: the potential role of the gut microbiome-bile acid-brain axis in the development and progression of alzheimer's disease and hepatic encephalopathy. *Med Res Rev* (2020) 40:1496–507. doi: 10.1002/med.21653

322. Makishima M, Okamoto AY, Repa JJ, Tu H, Learned RM, Luk A, et al. Identification of a nuclear receptor for bile acids. *Science* (1999) 284:1362–5. doi: 10.1126/science.284.5418.1362

323. Sayin SI, Wahlström A, Felin J, Jäntti S, Marschall H-U, Bamberg K, et al. Gut microbiota regulates bile acid metabolism by reducing the levels of tauro-betamuricholic acid, a naturally occurring FXR antagonist. *Cell Metab* (2013) 17:225–35. doi: 10.1016/j.cmet.2013.01.003

324. Bin WZ, Chen YR, Lv JT, Wang MX, Chen ZY, Zhou W, et al. A review of bile acid metabolism and signaling in cognitive dysfunction-related diseases. *Oxid Med Cell Longev* (2022) 2022:4289383. doi: 10.1155/2022/4289383

325. Schubring SR, Fleischer W, Lin JS, Haas HL, Sergeeva OA. The bile steroid chenodeoxycholate is a potent antagonist at NMDA and GABA(A) receptors. *Neurosci Lett* (2012) 506:322–6. doi: 10.1016/j.neulet.2011.11.036

326. van Best N, Rolle-Kampczyk U, Schaap FG, Basic M, Olde Damink SWM, Bleich A, et al. Bile acids drive the newborn's gut microbiota maturation. *Nat Commun* (2020) 11:3692. doi: 10.1038/s41467-020-17183-8

327. Li M, Liu S, Wang M, Hu H, Yin J, Liu C, et al. Gut microbiota dysbiosis associated with bile acid metabolism in neonatal cholestasis disease. *Sci Rep* (2020) 10:7686. doi: 10.1038/s41598-020-64728-4

328. Quinn M, McMillin M, Galindo C, Frampton G, Pae HY, DeMorrow S. Bile acids permeabilize the blood brain barrier after bile duct ligation in rats *via* Rac1-dependent mechanisms. *Dig Liver Dis* (2014) 46:527–34. doi: 10.1016/j.dld.2014.01.159

329. Cabrera-Rubio R, Patterson AM, Cotter PD, Beraza N. Cholestasis induced by bile duct ligation promotes changes in the intestinal microbiome in mice. *Sci Rep* (2019) 9:12324. doi: 10.1038/s41598-019-48784-z

330. MahmoudianDehkordi S, Bhattacharyya S, Brydges CR, Jia W, Fiehn O, Rush AJ, et al. Gut microbiome-linked metabolites in the pathobiology of major depression with or without anxiety-a role for bile acids. *Front Neurosci* (2022) 16:937906. doi: 10.3389/fnins.2022.937906

331. Zhang S, Zeng X, Ren M, Mao X, Qiao S. Novel metabolic and physiological functions of branched chain amino acids: a review. *J Anim Sci Biotechnol* (2017) 8:10. doi: 10.1186/s40104-016-0139-z

332. Blachier F, Mariotti F, Huneau JF, Tomé D. Effects of amino acid-derived luminal metabolites on the colonic epithelium and physiopathological consequences. *Amino Acids* (2007) 33:547–62. doi: 10.1007/s00726-006-0477-9

333. Dai Z-L, Zhang J, Wu G, Zhu W-Y. Utilization of amino acids by bacteria from the pig small intestine. *Amino Acids* (2010) 39:1201–15. doi: 10.1007/s00726-010-0556-9

334. Yang Z, Huang S, Zou D, Dong D, He X, Liu. N, et al. Metabolic shifts and structural changes in the gut microbiota upon branched-chain amino acid supplementation in middle-aged mice. *Amino Acids* (2016) 48:2731-45. doi: 10.1007/s00726-016-2308-y

335. D'Antona G, Ragni M, Cardile A, Tedesco L, Dossena M, Bruttini F, et al. Branched-chain amino acid supplementation promotes survival and supports cardiac and skeletal muscle mitochondrial biogenesis in middle-aged mice. *Cell Metab* (2010) 12:362–72. doi: 10.1016/j.cmet.2010.08.016

336. Sperringer JE, Addington A, Hutson SM. Branched-chain amino acids and brain metabolism. *Neurochem Res* (2017) 42:1697–709. doi: 10.1007/s11064-017-2261-5

337. Cheng Y, Zhang Q, Meng Q, Xia T, Huang Z, Wang C, et al. Leucine deprivation stimulates fat loss *via* increasing CRH expression in the hypothalamus and activating the sympathetic nervous system. *Mol Endocrinol* (2011) 25:1624–35. doi: 10.1210/me.2011-0028

338. O'Reilly D, Crushell E, Hughes J, Ryan S, Rogers Y, Borovickova I, et al. Maple syrup urine disease: clinical outcomes, metabolic control, and genotypes in a screened population after four decades of newborn bloodspot screening in the republic of Ireland. *J Inherit Metab Dis* (2021) 44:639–55. doi: 10.1002/jimd.12337

339. Shida Y, Endo H, Owada S, Inagaki Y, Sumiyoshi H, Kamiya A, et al. Branchedchain amino acids govern the high learning ability phenotype in tokai high avoider (THA) rats. *Sci Rep* (2021) 11:23104. doi: 10.1038/s41598-021-02591-7

340. Elliott JE, Keil AT, Mithani S, Gill JM, O'Neil ME, Cohen AS, et al. Dietary supplementation with branched chain amino acids to improve sleep in veterans with traumatic brain injury: a randomized double-blind placebo-controlled pilot and feasibility trial. *Front Syst Neurosci* (2022) 16:854874. doi: 10.3389/fnsys.2022.854874

341. Siddik MA, Mullins CA, Kramer A, Shah H, Gannaban RB, Zabet-Moghaddam M, et al. Branched-chain amino acids are linked with alzheimer's disease-related pathology and cognitive deficits. *Cells* (2022) 11:3523. doi: 10.3390/cells11213523

342. Dinan TG, Cryan JF. Regulation of the stress response by the gut microbiota: implications for psychoneuroendocrinology. *Psychoneuroendocrinology* (2012) 37:1369–78. doi: 10.1016/j.psyneuen.2012.03.007

343. Jašarević E, Howerton CL, Howard CD, Bale TL. Alterations in the vaginal microbiome by maternal stress are associated with metabolic reprogramming of the offspring gut and brain. *Endocrinology* (2015) 156:3265–76. doi: 10.1210/en.2015-1177

344. Jašarević E, Howard CD, Morrison K, Misic A, Weinkopff T, Scott P, et al. The maternal vaginal microbiome partially mediates the effects of prenatal stress on offspring gut and hypothalamus. *Nat Neurosci* (2018) 21:1061–71. doi: 10.1038/ s41593-018-0182-5

345. Gareau MG, Jury J, Yang PC, MacQueen G, Perdue MH. Neonatal maternal separation causes colonic dysfunction in rat pups including impaired host resistance. *Pediatr Res* (2006) 59:83–8. doi: 10.1203/01.pdr.0000190577.62426.45

346. Gareau MG, Jury J, MacQueen G, Sherman PM, Perdue MH. Probiotic treatment of rat pups normalises corticosterone release and ameliorates colonic dysfunction induced by maternal separation. *Gut* (2007) 56:1522–8. doi: 10.1136/gut.2006.117176

347. O'Mahony SM, Marchesi JR, Scully P, Codling C, Ceolho AM, Quigley EMM, et al. Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol Psychiatry* (2009) 65:263–7. doi: 10.1016/j.biopsych.2008.06.026

348. Suri D, Veenit V, Sarkar A, Thiagarajan D, Kumar A, Nestler EJ, et al. Early stress evokes age-dependent biphasic changes in hippocampal neurogenesis, BDNF expression, and cognition. *Biol Psychiatry* (2013) 73:658–66. doi: 10.1016/jbiopsych.2012.10.023

349. Huo R, Zeng B, Zeng L, Cheng K, Li B, Luo Y, et al. Microbiota modulate anxiety-like behavior and endocrine abnormalities in hypothalamic-pituitary-adrenal axis. *Front Cell Infect Microbiol* (2017) 7:489. doi: 10.3389/fcimb.2017.00489

350. Clarke G, Grenham S, Scully P, Fitzgerald P, Moloney RD, Shanahan F, et al. The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. *Mol Psychiatry* (2013) 18:666–73. doi: 10.1038/mp.2012.77

351. Crumeyrolle-Arias M, Jaglin M, Bruneau A, Vancassel S, Cardona A, Daugé V, et al. Absence of the gut microbiota enhances anxiety-like behavior and neuroendocrine response to acute stress in rats. *Psychoneuroendocrinology* (2014) 42:207–17. doi: 10.1016/j.psyneuen.2014.01.014

352. Söderholm JD, Yang P-C, Ceponis P, Vohra A, Riddell R, Sherman PM, et al. Chronic stress induces mast cell-dependent bacterial adherence and initiates mucosal inflammation in rat intestine. *Gastroenterology* (2002) 123:1099–108. doi: 10.1053/gast.2002.36019

353. Cameron HL, Perdue MH. Stress impairs murine intestinal barrier function: improvement by glucagon-like peptide-2. J Pharmacol Exp Ther (2005) 314:214–20. doi: 10.1124/jpet.105.085373

354. Sun Y, Zhang M, Chen C-C, Gillilland M3rd, Sun X, El-Zaatari M, et al. Stressinduced corticotropin-releasing hormone-mediated NLRP6 inflammasome inhibition and transmissible enteritis in mice. *Gastroenterology* (2013) 144:1478–87, 1487.e1–8. doi: 10.1053/j.gastro.2013.02.038 355. Lutgendorff F, Akkermans LMA, Söderholm JD. The role of microbiota and probiotics in stress-induced gastro-intestinal damage. *Curr Mol Med* (2008) 8:282–98. doi: 10.2174/156652408784533779

356. Gareau MG, Silva MA, Perdue MH. Pathophysiological mechanisms of stressinduced intestinal damage. *Curr Mol Med* (2008) 8:274–81. doi: 10.2174/ 156652408784533760

357. Teitelbaum AA, Gareau MG, Jury J, Yang PC, Perdue MH. Chronic peripheral administration of corticotropin-releasing factor causes colonic barrier dysfunction similar to psychological stress. *Am J Physiol Gastrointest Liver Physiol* (2008) 295: G452–9. doi: 10.1152/ajpgi.90210.2008

358. Martel J, Chang S-H, Ko Y-F, Hwang T-L, Young JD, Ojcius DM. Gut barrier disruption and chronic disease. *Trends Endocrinol Metab* (2022) 33:247–65. doi: 10.1016/j.tem.2022.01.002

359. Bailey MT, Dowd SE, Parry NMA, Galley JD, Schauer DB, Lyte M. Stressor exposure disrupts commensal microbial populations in the intestines and leads to increased colonization by citrobacter rodentium. *Infect Immun* (2010) 78:1509–19. doi: 10.1128/IAI.00862-09

360. Allen RG, Lafuse WP, Galley JD, Ali MM, Ahmer BMM, Bailey MT. The intestinal microbiota are necessary for stressor-induced enhancement of splenic macrophage microbicidal activity. *Brain Behav Immun* (2012) 26:371–82. doi: 10.1016/j.bbi.2011.11.002

361. Hoban AE, Moloney RD, Golubeva AV, McVey Neufeld KA, O'Sullivan O, Patterson E, et al. Behavioural and neurochemical consequences of chronic gut microbiota depletion during adulthood in the rat. *Neuroscience* (2016) 339:463–77. doi: 10.1016/j.neuroscience.2016.10.003

362. Gareau MG, Wine E, Rodrigues DM, Cho JH, Whary MT, Philpott DJ, et al. Bacterial infection causes stress-induced memory dysfunction in mice. *Gut* (2011) 60:307–17. doi: 10.1136/gut.2009.202515

363. Jiang PF, Zhu T, Gao JD, Gao F, Mao SS, Zhao WT, et al. The effect of maternal infection on cognitive development and hippocampus neuronal apoptosis, proliferation and differentiation in the neonatal rats. *Neuroscience* (2013) 246:422–34. doi: 10.1016/j.neuroscience.2013.04.021

364. Shanks N, Larocque S, Meaney MJ. Neonatal endotoxin exposure alters the development of the hypothalamic-pituitary-adrenal axis: early illness and later responsivity to stress. *J Neurosci Off J Soc Neurosci* (1995) 15:376–84. doi: 10.1523/JNEUROSCI.15-01-00376.1995

365. Shanks N, Windle RJ, Perks PA, Harbuz MS, Jessop DS, Ingram CD, et al. Early-life exposure to endotoxin alters hypothalamic-pituitary-adrenal function and predisposition to inflammation. *Proc Natl Acad Sci U.S.A.* (2000) 97:5645–50. doi: 10.1073/pnas.090571897

366. Agusti A, Moya-Pérez A, Campillo I, Montserrat-de la Paz S, Cerrudo V, Perez-Villalba A, et al. Bifidobacterium pseudocatenulatum CECT 7765 ameliorates neuroendocrine alterations associated with an exaggerated stress response and anhedonia in obese mice. *Mol Neurobiol* (2018) 55:5337–52. doi: 10.1007/s12035-017-0768-z

367. Ait-Belgnaoui A, Durand H, Cartier C, Chaumaz G, Eutamene H, Ferrier L, et al. Prevention of gut leakiness by a probiotic treatment leads to attenuated HPA response to an acute psychological stress in rats. *Psychoneuroendocrinology* (2012) 37:1885–95. doi: 10.1016/j.psyneuen.2012.03.024

368. Vodička M, Ergang P, Hrnčíř T, Mikulecká A, Kvapilová P, Vagnerová K, et al. Microbiota affects the expression of genes involved in HPA axis regulation and local metabolism of glucocorticoids in chronic psychosocial stress. *Brain Behav Immun* (2018) 73:615–24. doi: 10.1016/j.bbi.2018.07.007

369. Rosin S, Xia K, Azcarate-Peril MA, Carlson AL, Propper CB, Thompson AL, et al. A preliminary study of gut microbiome variation and HPA axis reactivity in healthy infants. *Psychoneuroendocrinology* (2021) 124:105046. doi: 10.1016/j.psyneuen.2020.105046

370. Keskitalo A, Aatsinki AK, Kortesluoma S, Pelto J, Korhonen L, Lahti L, et al. Gut microbiota diversity but not composition is related to saliva cortisol stress response at the age of 2.5 months. *Stress* (2021) 24:551–60. doi: 10.1080/10253890.2021.1895110

371. Grigoleit J-S, Kullmann JS, Wolf OT, Hammes F, Wegner A, Jablonowski S, et al. Dose-dependent effects of endotoxin on neurobehavioral functions in humans. *PloS One* (2011) 6:e28330. doi: 10.1371/journal.pone.0028330

372. Messaoudi M, Lalonde R, Violle N, Javelot H, Desor D, Nejdi A, et al. Assessment of psychotropic-like properties of a probiotic formulation (Lactobacillus helveticus R0052 and bifidobacterium longum R0175) in rats and human subjects. *Br J Nutr* (2011) 105:755–64. doi: 10.1017/S0007114510004319

373. Zhang N, Zhang Y, Li M, Wang W, Liu Z, Xi C, et al. Efficacy of probiotics on stress in healthy volunteers: a systematic review and meta-analysis based on randomized controlled trials. *Brain Behav* (2020) 10:e01699. doi: 10.1002/brb3.1699

374. Berding K, Bastiaanssen TFS, Moloney GM, Boscaini S, Strain CR, Anesi A, et al. Feed your microbes to deal with stress: a psychobiotic diet impacts microbial stability and perceived stress in a healthy adult population. *Mol Psychiatry* (2022) 28:601–10. doi: 10.1038/s41380-022-01817-y

375. Desbonnet L, Garrett L, Clarke G, Kiely B, Cryan JF, Dinan TG. Effects of the probiotic bifidobacterium infantis in the maternal separation model of depression. *Neuroscience* (2010) 170:1179-88. doi: 10.1016/j.neuroscience.2010.08.005

376. Bercik P, Denou E, Collins J, Jackson W, Lu J, Jury J, et al. The intestinal microbiota affect central levels of brain-derived neurotropic factor and behavior in mice. *Gastroenterology* (2011) 141:599–609. doi: 10.1053/j.gastro.2011.04.052

377. Desbonnet L, Clarke G, Shanahan F, Dinan TG, Cryan JF. Microbiota is essential for social development in the mouse. *Mol Psychiatry* (2014) 19:146-8. doi: 10.1038/mp.2013.65

378. Lukić I, Getselter D, Koren O, Elliott E. Role of tryptophan in microbiotainduced depressive-like behavior: evidence from tryptophan depletion study. *Front Behav Neurosci* (2019) 13:123. doi: 10.3389/fnbeh.2019.00123

379. Lyte JM, Gheorghe CE, Goodson MS, Kelley-Loughnane N, Dinan TG, Cryan JF, et al. Gut-brain axis serotonergic responses to acute stress exposure are microbiomedependent. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2020) 32:e13881. doi: 10.1111/nmo.13881

380. Matsumoto M, Kibe R, Ooga T, Aiba Y, Sawaki E, Koga Y, et al. Cerebral lowmolecular metabolites influenced by intestinal microbiota: a pilot study. *Front Syst Neurosci* (2013) 7:9. doi: 10.3389/fnsys.2013.00009

381. Gur TL, Shay L, Palkar AV, Fisher S, Varaljay VA, Dowd S, et al. Prenatal stress affects placental cytokines and neurotrophins, commensal microbes, and anxiety-like behavior in adult female offspring. *Brain Behav Immun* (2017) 64:50–8. doi: 10.1016/ j.bbi.2016.12.021

382. Desbonnet L, Clarke G, Traplin A, O'Sullivan O, Crispie F, Moloney RD, et al. Gut microbiota depletion from early adolescence in mice: implications for brain and behaviour. *Brain Behav Immun* (2015) 48:165–73. doi: 10.1016/j.bbi.2015.04.004

383. Fröhlich EE, Farzi A, Mayerhofer R, Reichmann F, Jačan A, Wagner B, et al. Cognitive impairment by antibiotic-induced gut dysbiosis: analysis of gut microbiotabrain communication. *Brain Behav Immun* (2016) 56:140–55. doi: 10.1016/ j.bbi.2016.02.020

384. Möhle L, Mattei D, Heimesaat MM, Bereswill S, Fischer A, Alutis M, et al. Ly6Chi monocytes provide a link between antibiotic-induced changes in gut microbiota and adult hippocampal neurogenesis. *Cell Rep* (2016) 15:1945–56. doi: 10.1016/j.celrep.2016.04.074

385. Chu C, Murdock MH, Jing D, Won TH, Chung H, Kressel AM, et al. The microbiota regulate neuronal function and fear extinction learning. *Nature* (2019) 574:543–8. doi: 10.1038/s41586-019-1644-y

386. Joëls M, Baram TZ. The neuro-symphony of stress. *Nat Rev Neurosci* (2009) 10:459–66. doi: 10.1038/nrn2632

387. Burokas A, Arboleya S, Moloney RD, Peterson VL, Murphy K, Clarke G, et al. Targeting the microbiota-Gut-Brain axis: prebiotics have anxiolytic and antidepressant-like effects and reverse the impact of chronic stress in mice. *Biol Psychiatry* (2017) 82:472–87. doi: 10.1016/j.biopsych.2016.12.031

388. Matthews DM, Jenks SM. Ingestion of mycobacterium vaccae decreases anxiety-related behavior and improves learning in mice. *Behav Processes* (2013) 96:27–35. doi: 10.1016/j.beproc.2013.02.007

389. Lowry CA, Hollis JH, de Vries A, Pan B, Brunet LR, Hunt JRF, et al. Identification of an immune-responsive mesolimbocortical serotonergic system: potential role in regulation of emotional behavior. *Neuroscience* (2007) 146:756–72. doi: 10.1016/j.neuroscience.2007.01.067

390. Savignac HM, Tramullas M, Kiely B, Dinan TG, Cryan JF. Bifidobacteria modulate cognitive processes in an anxious mouse strain. *Behav Brain Res* (2015) 287:59–72. doi: 10.1016/j.bbr.2015.02.044

391. Buffington SA, Di Prisco GV, Auchtung TA, Ajami NJ, Petrosino JF, Costa-Mattioli M. Microbial reconstitution reverses maternal diet-induced social and synaptic deficits in offspring. *Cell* (2016) 165:1762–75. doi: 10.1016/j.cell.2016.06.001

392. Liu J, Sun J, Wang F, Yu X, Ling Z, Li H, et al. Neuroprotective effects of clostridium butyricum against vascular dementia in mice *via* metabolic butyrate. *BioMed Res Int* (2015) 2015:412946. doi: 10.1155/2015/412946

393. Wang K-C, Fan L-W, Kaizaki A, Pang Y, Cai Z, Tien L-T. Neonatal lipopolysaccharide exposure induces long-lasting learning impairment, less anxiety-like response and hippocampal injury in adult rats. *Neuroscience* (2013) 234:146–57. doi: 10.1016/j.neuroscience.2012.12.049

394. Bilbo SD, Biedenkapp JC, Der-Avakian A, Watkins LR, Rudy JW, Maier SF. Neonatal infection-induced memory impairment after lipopolysaccharide in adulthood is prevented *via* caspase-1 inhibition. *J Neurosci* (2005) 25:8000–9. doi: 10.1523/ JNEUROSCI.1748-05.2005

395. Bilbo SD, Newsum NJ, Sprunger DB, Watkins LR, Rudy JW, Maier SF. Differential effects of neonatal handling on early life infection-induced alterations in cognition in adulthood. *Brain Behav Immun* (2007) 21:332–42. doi: 10.1016/j.bbi.2006.10.005

396. Ratnayake U, Quinn TA, Castillo-Melendez M, Dickinson H, Walker DW. Behaviour and hippocampus-specific changes in spiny mouse neonates after treatment of the mother with the viral-mimetic poly I:C at mid-pregnancy. *Brain Behav Immun* (2012) 26:1288–99. doi: 10.1016/j.bbi.2012.08.011

397. Zhu S, Jiang Y, Xu K, Cui M, Ye W, Zhao G, et al. The progress of gut microbiome research related to brain disorders. *J Neuroinflamm* (2020) 17:25. doi: 10.1186/s12974-020-1705-z

398. Montiel-Castro A, González-Cervantes R, Bravo-Ruiseco G, Pacheco-Lopez G. The microbiota-gut-brain axis: neurobehavioral correlates, health and sociality. *Front Integr Neurosci* (2013) 7:70. doi: 10.3389/fnint.2013.00070

399. Carlson AL, Xia K, Azcarate-Peril MA, Goldman BD, Ahn M, Styner MA, et al. Infant gut microbiome associated with cognitive development. *Biol Psychiatry* (2018) 83:148–59. doi: 10.1016/j.biopsych.2017.06.021

400. Polidano C, Zhu A, Bornstein JC. The relation between cesarean birth and child cognitive development. Sci Rep (2017) 7:11483. doi: 10.1038/s41598-017-10831-y

401. Blustein J, Liu J. Time to consider the risks of caesarean delivery for long term child health. BMJ Br Med J (2015) 350:h2410. doi: 10.1136/bmj.h2410

402. Martinez LD, Glynn LM, Sandman CA, Wing DA, Davis EP. Cesarean delivery and infant cortisol regulation. *Psychoneuroendocrinology* (2020) 122:104862. doi: 10.1016/j.psyneuen.2020.104862

403. Kiilerich P, Cortes R, Lausten-Thomsen U, Borbye-Lorenzen N, Holmgaard S, Skogstrand K. Delivery modality affect neonatal levels of inflammation, stress, and growth factors. *Front Pediatr* (2021) 9:709765. doi: 10.3389/fped.2021.709765

404. Halverson T, Alagiakrishnan K. Gut microbes in neurocognitive and mental health disorders. Ann Med (2020) 52:423-43. doi: 10.1080/07853890.2020.1808239

405. Dinan K, Dinan T. Antibiotics and mental health: the good, the bad and the ugly. J Intern Med (2022) 292:858-69. doi: 10.1111/joim.13543

406. Liu Z, Wei S, Chen X, Liu L, Wei Z, Liao Z, et al. The effect of long-term or repeated use of antibiotics in children and adolescents on cognitive impairment in middle-aged and older person(s) adults: a cohort study. *Front Aging Neurosci* (2022) 14:833365. doi: 10.3389/fnagi.2022.833365

407. Mehta RS, Lochhead P, Wang Y, Ma W, Nguyen LH, Kochar B, et al. Association of midlife antibiotic use with subsequent cognitive function in women. *PloS One* (2022) 17:e0264649. doi: 10.1371/journal.pone.0264649

408. Bokulich NA, Chung J, Battaglia T, Henderson N, Jay M, Li H, et al. Antibiotics, birth mode, and diet shape microbiome maturation during early life. *Sci Transl Med* (2016) 8:343ra82–343ra82. doi: 10.1126/scitranslmed.aad7121

409. Taggart H, Bergstrom L. An overview of the microbiome and the effects of antibiotics. J Nurse Pract (2014) 10:445–50. doi: 10.1016/j.nurpra.2014.05.005

410. Bossung V, Lupatsii M, Dashdorj L, Tassiello O, Jonassen S, Pagel J, et al. Timing of antimicrobial prophylaxis for cesarean section is critical for gut microbiome development in term born infants. *Gut Microbes* (2022) 14:2038855. doi: 10.1080/ 19409076.2022.2038855

411. Slykerman RF, Thompson J, Waldie KE, Murphy R, Wall C, Mitchell EA. Antibiotics in the first year of life and subsequent neurocognitive outcomes. *Acta Paediatr* (2017) 106:87–94. doi: 10.1111/apa.13613

412. Ou Y, Belzer C, Smidt H, de Weerth C. Development of the gut microbiota in healthy children in the first ten years of life: associations with internalizing and externalizing behavior. *Gut Microbes* (2022) 14:2038853. doi: 10.1080/19490976.2022.2038853

413. Thänert R, Sawhney SS, Schwartz DJ, Dantas G. The resistance within: antibiotic disruption of the gut microbiome and resistome dynamics in infancy. *Cell Host Microbe* (2022) 30:675–83. doi: 10.1016/j.chom.2022.03.013

414. Bajaj JS, Betrapally NS, Gillevet PM. Decompensated cirrhosis and microbiome interpretation. *Nature* (2015) 525:E1-2. doi: 10.1038/nature14851

415. Patidar KR, Bajaj JS. Antibiotics for the treatment of hepatic encephalopathy. *Metab Brain Dis* (2013) 28:307–12. doi: 10.1007/s11011-013-9383-5

416. Sung CM, Lin Y-F, Chen K-F, Ke H-M, Huang H-Y, Gong Y-N, et al. Predicting clinical outcomes of cirrhosis patients with hepatic encephalopathy from the fecal microbiome. *Cell Mol Gastroenterol Hepatol* (2019) 8:301–318.e2. doi: 10.1016/j.jcmgh.2019.04.008

417. Festi D, Mazzella G, Parini P, Ronchi M, Cipolla A, Orsini M, et al. Treatment of hepatic encephalopathy with non-absorbable antibiotics. *Ital J Gastroenterol* (1992) 24:14–6.

418. Ahluwalia V, Wade JB, Heuman DM, Hammeke TA, Sanyal AJ, Sterling RK, et al. Enhancement of functional connectivity, working memory and inhibitory control on multi-modal brain MR imaging with rifaximin in cirrhosis: implications for the gut-liver-brain axis. *Metab Brain Dis* (2014) 29:1017–25. doi: 10.1007/s11011-014-9507-6

419. Bajaj JS, Heuman DM, Sanyal AJ, Hylemon PB, Sterling RK, Stravitz RT, et al. Modulation of the metabiome by rifaximin in patients with cirrhosis and minimal hepatic encephalopathy. *PloS One* (2013) 8:e60042. doi: 10.1371/journal.pone.0060042

420. Stark CM, Susi A, Nylund CM. Early life exposure to microbiota-altering medications is associated with childhood psychotropic medication prescriptions. *Pediatrics* (2021) 147:1036–7. doi: 10.1542/peds.147.3MA11.1036

421. Liu L, Shang L, Jin D, Wu X, Long B. General anesthesia bullies the gut: a toxic relationship with dysbiosis and cognitive dysfunction. *Psychopharmacol (Berl)* (2022) 239:709–28. doi: 10.1007/s00213-022-06096-7

422. Judd LL, Schettler PJ, Brown ES, Wolkowitz OM, Sternberg EM, Bender BG, et al. Adverse consequences of glucocorticoid medication: psychological, cognitive, and behavioral effects. *Am J Psychiatry* (2014) 171:1045–51. doi: 10.1176/appi.ajp.2014.13091264

423. Varney NR, Alexander B, MacIndoe JH. Reversible steroid dementia in patients without steroid psychosis. *Am J Psychiatry* (1984) 141:369–72. doi: 10.1176/ajp.141.3.369

424. Stoudemire A, Anfinson T, Edwards J. Corticosteroid-induced delirium and dependency. *Gen Hosp Psychiatry* (1996) 18:196–202. doi: 10.1016/0163-8343(96) 00005-9

425. Newcomer JW, Craft S, Hershey T, Askins K, Bardgett ME. Glucocorticoidinduced impairment in declarative memory performance in adult humans. J Neurosci Off J Soc Neurosci (1994) 14:2047–53. doi: 10.1523/JNEUROSCI.14-04-02047.1994

426. Newcomer JW, Selke G, Melson AK, Hershey T, Craft S, Richards K, et al. Decreased memory performance in healthy humans induced by stress-level cortisol treatment. *Arch Gen Psychiatry* (1999) 56:527–33. doi: 10.1001/archpsyc.56.6.527

427. Bender BG, Lerner JA, Kollasch E. Mood and memory changes in asthmatic children receiving corticosteroids. *J Am Acad Child Adolesc Psychiatry* (1988) 27:720–5. doi: 10.1097/00004583-198811000-00010

428. Savas M, Vinkers CH, Rosmalen JGM, Hartman CA, Wester VL, van den Akker ELT, et al. Systemic and local corticosteroid use is associated with reduced executive cognition, and mood and anxiety disorders. *Neuroendocrinology* (2020) 110:282–91. doi: 10.1159/000501617

429. Dekkers AJ, Amaya JM, van der Meulen M, Biermasz NR, Meijer OC, Pereira AM. Long-term effects of glucocorticoid excess on the brain. *J Neuroendocrinol* (2022) 34:e13142. doi: 10.1111/jne.13142

430. Prado CE, Crowe SF. Corticosteroids and cognition: a meta-analysis. Neuropsychol Rev (2019) 29:288-312. doi: 10.1007/s11065-019-09405-8

431. Lupien SJ, Fiocco A, Wan N, Maheu F, Lord C, Schramek T, et al. Stress hormones and human memory function across the lifespan. *Psychoneuroendocrinology* (2005) 30:225–42. doi: 10.1016/j.psyneuen.2004.08.003

432. Bauduin SEEC, den Rooijen ILB, Meijer M, van der Werff SJA, Keo A, Dzyubachyk O, et al. Potential associations between immune signaling genes, deactivated microglia, and oligodendrocytes and cortical gray matter loss in patients with long-term remitted cushing's disease. *Psychoneuroendocrinology* (2021) 132:105334. doi: 10.1016/j.psyncuen.2021.105334

433. Henry M, Thomas KGF, Ross IL. Sleep, cognition and cortisol in addison's disease: a mechanistic relationship. *Front Endocrinol* (2021) 12:694046. doi: 10.3389/fendo.2021.694046

434. Wu T, Yang L, Jiang J, Ni Y, Zhu J, Zheng X, et al. Chronic glucocorticoid treatment induced circadian clock disorder leads to lipid metabolism and gut microbiota alterations in rats. *Life Sci* (2018) 192:173–82. doi: 10.1016/j.lfs.2017.11.049

435. Schepper JD, Collins F, Rios-Arce ND, Kang HJ, Schaefer L, Gardinier JD, et al. Involvement of the gut microbiota and barrier function in glucocorticoid-induced osteoporosis. J Bone Miner Res (2020) 35:801–20. doi: 10.1002/jbmr.3947

436. Wang M, Zhu Z, Lin X, Li H, Wen C, Bao J, et al. Gut microbiota mediated the therapeutic efficacies and the side effects of prednisone in the treatment of MRL/lpr mice. *Arthritis Res Ther* (2021) 23:1–10. doi: 10.1186/s13075-021-02620-w

437. Huang EY, Inoue T, Leone VA, Dalal S, Touw K, Wang Y, et al. Using corticosteroids to reshape the gut microbiome: implications for inflammatory bowel diseases. *Inflammation Bowel Dis* (2015) 21:963–72. doi: 10.1097/MIB.00000000000332

438. Lupien SJ, McEwen BS. The acute effects of corticosteroids on cognition: integration of animal and human model studies. *Brain Res Rev* (1997) 24:1-27. doi: 10.1016/S0165-0173(97)00004-0

439. Cottrell GA, Nakajima S. Effect of corticosteroids in the hippocampus on passive avoidance behavior in the rat. *Pharmacol Biochem Behav* (1977) 7:277-80. doi: 10.1016/0091-3057(77)90146-0

440. Sandi C, Loscertales M, Guaza C. Experience-dependent facilitating effect of corticosterone on spatial memory formation in the water maze. *Eur J Neurosci* (1997) 9:637–42. doi: 10.1111/j.1460-9568.1997.tb01412.x

441. Sandi C, Rose SPR. Corticosterone enhances long-term retention in one-dayold chicks trained in a weak passive avoidance learning paradigm. *Brain Res* (1994) 647:106–12. doi: 10.1016/0006-8993(94)91404-4

442. Buchanan TW, Lovallo WR. Enhanced memory for emotional material following stress-level cortisol treatment in humans. *Psychoneuroendocrinology* (2001) 26:307–17. doi: 10.1016/S0306-4530(00)00058-5

443. Lupien SJ, Wilkinson CW, Brière S, Ménard C, Ng Ying Kin NMK, Nair NPV. The modulatory effects of corticosteroids on cognition: studies in young human populations. *Psychoneuroendocrinology* (2002) 27:401–16. doi: 10.1016/S0306-4530 (01)00061-0

444. Beckwith BE, Petros TV, Scaglione C, Nelson J. Dose-dependent effects of hydrocortisone on memory in human males. *Physiol Behav* (1986) 36:283-6. doi: 10.1016/0031-9384(86)90017-x

445. Lupien SJ, McEwen BS, Gunnar MR, Heim C. Effects of stress throughout the lifespan on the brain, behaviour and cognition. *Nat Rev Neurosci* (2009) 10:434–45. doi: 10.1038/nrn2639

446. Bruno G, Zaccari P, Rocco G, Scalese G, Panetta C, Porowska B, et al. Proton pump inhibitors and dysbiosis: current knowledge and aspects to be clarified. *World J Gastroenterol* (2019) 25:2706–19. doi: 10.3748/wjg.v25.i22.2706

447. Lochhead P, Hagan K, Joshi AD, Khalili H, Nguyen LH, Grodstein F, et al. Association between proton pump inhibitor use and cognitive function in women. *Gastroenterology* (2017) 153:971–979.e4. doi: 10.1053/j.gastro.2017.06.061

448. Wu C-Y, Xiong LY, Ouk M, Rabin JS, Herrmann N, Lanctôt KL, et al. Gastric acid suppressants and cognitive decline in people with or without cognitive impairment. *Alzheimer's Dement (New York N Y)* (2022) 8:e12243. doi: 10.1002/trc2.12243

449. Bajic JE, Johnston IN, Howarth GS, Hutchinson MR. From the bottom-up: chemotherapy and gut-brain axis dysregulation. *Front Behav Neurosci* (2018) 12:104. doi: 10.3389/fnbeh.2018.00104

450. Bilenduke E, Sterrett JD, Ranby KW, Borges VF, Grigsby J, Carr AL, et al. Impacts of breast cancer and chemotherapy on gut microbiome, cognitive functioning, and mood relative to healthy controls. *Sci Rep* (2022) 12:19547. doi: 10.1038/s41598-022-23793-7

451. Benros ME, Sørensen HJ, Nielsen PR, Nordentoft M, Mortensen PB, Petersen L. The association between infections and general cognitive ability in young men – a nationwide study. *PloS One* (2015) 10:e0124005. doi: 10.1371/journal.pone.0124005

452. Muzambi R, Bhaskaran K, Brayne C, Smeeth L, Warren-Gash C. Common bacterial infections and risk of incident cognitive decline or dementia: a systematic review protocol. *BMJ Open* (2019) 9:e030874. doi: 10.1136/bmjopen-2019-030874

453. Damiano RF, Guedes BF, de Rocca CC, de Pádua Serafim A, Castro LHM, Munhoz CD, et al. Cognitive decline following acute viral infections: literature review and projections for post-COVID-19. *Eur Arch Psychiatry Clin Neurosci* (2022) 272:139–54. doi: 10.1007/s00406-021-01286-4

454. Fernandez-Cotarelo M-J, Nagy-Agren SE, Smolkin ME, Jimenez-Diez-Canseco L, Perez-Pomata M-T, Shenal BV, et al. Functional and cognitive status in clostridium difficile infection in the hospitalized elderly: a retrospective study of two sites. *J Gen Intern Med* (2019) 34:1392–3. doi: 10.1007/s11606-019-04935-6

455. Biazzo M, Allegra M, Deidda G. Clostridioides difficile and neurological disorders: new perspectives. *Front Neurosci* (2022) 16:946601. doi: 10.3389/fnins.2022.946601

456. Vinithakumari AA, Piyush P, Belen H, Je-Han LS, Aaron D-K, Lucas S, et al. Clostridioides difficile infection dysregulates brain dopamine metabolism. *Microbiol Spectr* (2022) 10:e00073–22. doi: 10.1128/spectrum.00073-22

457. Hazan S. Rapid improvement in alzheimer's disease symptoms following fecal microbiota transplantation: a case report. *J Int Med Res* (2020) 48:300060520925930. doi: 10.1177/0300060520925930

458. Park S-H, Lee JH, Shin J, Kim J-S, Cha B, Lee S, et al. Cognitive function improvement after fecal microbiota transplantation in alzheimer's dementia patient: a case report. *Curr Med Res Opin* (2021) 37:1739-44. doi: 10.1080/03007995.2021.1957807

459. Tarter KD, Simanek AM, Dowd JB, Aiello AE. Persistent viral pathogens and cognitive impairment across the life course in the third national health and nutrition examination survey. *J Infect Dis* (2014) 209:837–44. doi: 10.1093/infdis/jit616

460. Noppert GA, Aiello AE, O'Rand AM, Cohen HJ. Investigating pathogen burden in relation to a cumulative deficits index in a representative sample of US adults. *Epidemiol Infect* (2018) 146:1968–76. doi: 10.1017/S095026881800153X

461. Katan M, Moon YP, Paik MC, Sacco RL, Wright CB, Elkind MSV. Infectious burden and cognitive function: the northern Manhattan study. *Neurology* (2013) 80:1209–15. doi: 10.1212/WNL.0b013e3182896e79

462. Wright CB, Gardener H, Dong C, Yoshita M, DeCarli C, Sacco RL, et al. Infectious burden and cognitive decline in the northern Manhattan study. *J Am Geriatr Soc* (2015) 63:1540–5. doi: 10.1111/jgs.13557

463. Lee YH, Papandonatos GD, Savitz DA, Heindel WC, Buka SL. Effects of prenatal bacterial infection on cognitive performance in early childhood. *Paediatr Perinat Epidemiol* (2020) 34:70–9. doi: 10.1111/ppe.12603

464. O'Connor TG, Ciesla AA, Sefair AV, Thornburg LL, Brown AS, Glover V, et al. Maternal prenatal infection and anxiety predict neurodevelopmental outcomes in middle childhood. J Psychopathol Clin Sci (2022) 131:422–34. doi: 10.1037/abn0000746

465. Allen AP, Hutch W, Borre YE, Kennedy PJ, Temko A, Boylan G, et al. Bifidobacterium longum 1714 as a translational psychobiotic: modulation of stress, electrophysiology and neurocognition in healthy volunteers. *Transl Psychiatry* (2016) 6: e939. doi: 10.1038/tp.2016.191

466. Chung Y-C, Jin H-M, Cui Y, Kim DS, Jung JM, Park J-I, et al. Fermented milk of lactobacillus helveticus IDCC3801 improves cognitive functioning during cognitive fatigue tests in healthy older adults. *J Funct Foods* (2014) 10:465–74. doi: 10.1016/j.jff.2014.07.007

467. Lew L-C, Hor Y-Y, Yusoff NAA, Choi S-B, Yusoff MSB, Roslan NS, et al. Probiotic lactobacillus plantarum P8 alleviated stress and anxiety while enhancing memory and cognition in stressed adults: a randomised, double-blind, placebocontrolled study. *Clin Nutr* (2019) 38:2053–64. doi: 10.1016/j.clnu.2018.09.010

468. Steenbergen L, Sellaro R, van Hemert S, Bosch JA, Colzato LS. A randomized controlled trial to test the effect of multispecies probiotics on cognitive reactivity to sad mood. *Brain Behav Immun* (2015) 48:258–64. doi: 10.1016/j.bbi.2015.04.003

469. Dahiya D, Nigam PS. Clinical potential of microbial strains, used in fermentation for probiotic food, beverages and in synbiotic supplements, as psychobiotics for cognitive treatment through gut-brain signaling. *Microorganisms* (2022) 10:1687. doi: 10.3390/microorganisms10091687

470. Tillisch K, Labus J, Kilpatrick L, Jiang Z, Stains J, Ebrat B, et al. Consumption of fermented milk product with probiotic modulates brain activity. *Gastroenterology* (2013) 144:1394–401, 1401.e1–4. doi: 10.1053/j.gastro.2013.02.043

471. Papalini S, Michels F, Kohn N, Wegman J, van Hemert S, Roelofs K, et al. Stress matters: randomized controlled trial on the effect of probiotics on neurocognition. *Neurobiol Stress* (2019) 10:100141. doi: 10.1016/j.ynstr.2018.100141

472. Rode J, Edebol Carlman HMT, König J, Hutchinson AN, Thunberg P, Persson J, et al. Multi-strain probiotic mixture affects brain morphology and resting state brain function in healthy subjects: an RCT. *Cells* (2022) 11:2922. doi: 10.3390/cells11182922

473. Edebol Carlman HMT, Rode J, König J, Repsilber D, Hutchinson AN, Thunberg P, et al. Probiotic mixture containing lactobacillus helveticus, bifidobacterium longum and lactiplantibacillus plantarum affects brain responses to an arithmetic stress task in healthy subjects: a randomised clinical trial and proof-of-Concept study. *Nutrients* (2022) 14:1329. doi: 10.3390/nu14071329

474. Bloemendaal M, Szopinska-Tokov J, Belzer C, Boverhoff D, Papalini S, Michels F, et al. Probiotics-induced changes in gut microbial composition and its effects on cognitive performance after stress: exploratory analyses. *Transl Psychiatry* (2021) 11:300. doi: 10.1038/s41398-021-01404-9

475. Liu G, Chong H-X, Chung FY, Li Y, Liong M-T. Lactobacillus plantarum DR7 modulated bowel movement and gut microbiota associated with dopamine and serotonin pathways in stressed adults. *Int J Mol Sci* (2020) 21:4608. doi: 10.3390/ ijms21134608

476. Akbari E, Asemi Z, Daneshvar Kakhaki R, Bahmani F, Kouchaki E, Tamtaji OR, et al. Effect of probiotic supplementation on cognitive function and metabolic status in alzheimer's disease: a randomized, double-blind and controlled trial. *Front Aging Neurosci* (2016) 8:256. doi: 10.3389/fnagi.2016.00256

477. Roman P, Estévez AF, Miras A, Sánchez-Labraca N, Cañadas F, Vivas AB, et al. A pilot randomized controlled trial to explore cognitive and emotional effects of probiotics in fibromyalgia. *Sci Rep* (2018) 8:10965. doi: 10.1038/s41598-018-29388-5

478. Malatji BG, Mason S, Mienie LJ, Wevers RA, Meyer H, van Reenen M, et al. The GC-MS metabolomics signature in patients with fibromyalgia syndrome directs to dysbiosis as an aspect contributing factor of FMS pathophysiology. *Metabolomics* (2019) 15:54. doi: 10.1007/s11306-019-1513-6

479. Pinto-Sanchez MI, Hall GB, Ghajar K, Nardelli A, Bolino C, Lau JT, et al. Probiotic bifidobacterium longum NCC3001 reduces depression scores and alters brain activity: a pilot study in patients with irritable bowel syndrome. *Gastroenterology* (2017) 153:448–459.e8. doi: 10.1053/j.gastro.2017.05.003

480. Dinan TG, Cryan JF. Melancholic microbes: a link between gut microbiota and depression? *Neurogastroenterol Motil* (2013) 25:713–9. doi: 10.1111/nmo.12198

481. Fernandez-Real J-M, Serino M, Blasco G, Puig J, Daunis-i-Estadella J, Ricart W, et al. Gut microbiota interacts with brain microstructure and function. *J Clin Endocrinol Metab* (2015) 100:4505–13. doi: 10.1210/jc.2015-3076

482. Mahboobi S, Ghasvarian M, Ghaem H, Alipour H, Alipour S, Eftekhari MH. Effects of probiotic and magnesium co-supplementation on mood, cognition, intestinal barrier function and inflammation in individuals with obesity and depressed mood: a randomized, double-blind placebo-controlled clinical trial. *Front Nutr* (2022) 9:1018357. doi: 10.3389/fnut.2022.1018357

483. O'Brien MER, Anderson H, Kaukel E, O'Byrne K, Pawlicki M, Von Pawel J, et al. SRL172 (killed mycobacterium vaccae) in addition to standard chemotherapy improves quality of life without affecting survival, in patients with advanced non-small-cell lung cancer: phase III results. *Ann Oncol Off J Eur Soc Med Oncol* (2004) 15:906–14. doi: 10.1093/annonc/mdh220

484. Smith AP, Sutherland D, Hewlett P. An investigation of the acute effects of oligofructose-enriched inulin on subjective wellbeing, mood and cognitive performance. *Nutrients* (2015) 7:8887–96. doi: 10.3390/nu7115441

485. Schmidt K, Cowen PJ, Harmer CJ, Tzortzis G, Errington S, Burnet PWJ. Prebiotic intake reduces the waking cortisol response and alters emotional bias in healthy volunteers. *Psychopharmacology* (2015) 232:1793-801. doi: 10.1007/s00213-014-3810-0

486. Barrio C, Arias-Sánchez S, Martín-Monzón I. The gut microbiota-brain axis, psychobiotics and its influence on brain and behaviour: a systematic review. *Psychoneuroendocrinology* (2022) 137:105640. doi: 10.1016/j.psyneuen.2021.105640

487. Kelly JR, Allen AP, Temko A, Hutch W, Kennedy PJ, Farid N, et al. Lost in translation? the potential psychobiotic lactobacillus rhamnosus (JB-1) fails to modulate stress or cognitive performance in healthy male subjects. *Brain Behav Immun* (2017) 61:50–9. doi: 10.1016/j.bbi.2016.11.018

488. Basso M, Johnstone N, Knytl P, Nauta A, Groeneveld A, Cohen Kadosh K. A systematic review of psychobiotic interventions in children and adolescents to enhance cognitive functioning and emotional behavior. *Nutrients* (2022) 14:614. doi: 10.3390/nu14030614

489. Marx W, Scholey A, Firth J, D'Cunha NM, Lane M, Hockey M, et al. Prebiotics, probiotics, fermented foods and cognitive outcomes: a meta-analysis of randomized controlled trials. *Neurosci Biobehav Rev* (2020) 118:472–84. doi: 10.1016/j.neubiorev.2020.07.036

490. Sweis BM, Veverka KK, Dhillon ES, Urban JH, Lucas LR. Individual differences in the effects of chronic stress on memory: behavioral and neurochemical correlates of resiliency. *Neuroscience* (2013) 246:142–59. doi: 10.1016/j.neuroscience.2013.04.052

491. Roos LG, Janson J, Sturmbauer SC, Bennett JM, Rohleder N. Higher trait reappraisal predicts stronger HPA axis habituation to repeated stress. *Psychoneuroendocrinology* (2019) 101:12-8. doi: 10.1016/j.psyneuen.2018.10.018 492. Frankiensztajn LM, Elliott E, Koren O. The microbiota and the hypothalamuspituitary-adrenocortical (HPA) axis, implications for anxiety and stress disorders. *Curr Opin Neurobiol* (2020) 62:76–82. doi: 10.1016/j.conb.2019.12.003

493. Ge L, Liu S, Li S, Yang J, Hu G, Xu C, et al. Psychological stress in inflammatory bowel disease: psychoneuroimmunological insights into bidirectional gut-brain communications. *Front Immunol* (2022) 13:1016578. doi: 10.3389/fimmu.2022.1016578

494. Skrobisz K, Piotrowicz G, Rudnik A, Naumczyk P, Sabisz A, Markiet K, et al. Evaluation of subcortical structure volumes in patients with non-specific digestive diseases. *Diagnostics* (2022) 12:2199. doi: 10.3390/diagnostics12092199

495. Yamaoka K, Uotsu N, Hoshino E. Relationship between psychosocial stressinduced prefrontal cortex activity and gut microbiota in healthy participants-a functional near-infrared spectroscopy study. *Neurobiol Stress* (2022) 20:100479. doi: 10.1016/j.ynstr.2022.100479

496. Kennedy PJ, Clarke G, O'Neill A, Groeger JA, Quigley EM, Shanahan F, et al. An assessment of cognitive function in irritable bowel syndrome (IBS): are deficits in visuo-spatial memory stress-related and mediated by tryptophan metabolism along the kynurenine pathway? *Gastroenterology* (2012) 142:S-548. doi: 10.1016/S0016-5085(12)62105-2

497. Kennedy PJ, Clarke G, Quigley EMM, Groeger JA, Dinan TG, Cryan JF. Gut memories: towards a cognitive neurobiology of irritable bowel syndrome. *Neurosci Biobehav Rev* (2012) 36:310–40. doi: 10.1016/j.neubiorev.2011.07.001

498. Kennedy PJ, Clarke G, O'Neill A, Groeger JA, Quigley EMM, Shanahan F, et al. Cognitive performance in irritable bowel syndrome: evidence of a stress-related impairment in visuospatial memory. *Psychol Med* (2014) 44:1553–66. doi: 10.1017/ S0033291713002171

499. Aizawa E, Sato Y, Kochiyama T, Saito N, Izumiyama M, Morishita J, et al. Altered cognitive function of prefrontal cortex during error feedback in patients with irritable bowel syndrome, based on FMRI and dynamic causal modeling. *Gastroenterology* (2012) 143:1188–98. doi: 10.1053/j.gastro.2012.07.104

500. Klarer M, Weber-Stadlbauer U, Arnold M, Langhans W, Meyer U. Cognitive effects of subdiaphragmatic vagal deafferentation in rats. *Neurobiol Learn Mem* (2017) 142:190–9. doi: 10.1016/j.nlm.2017.05.006

501. Chen C, Ahn EH, Kang SS, Liu X, Alam A, Ye K. Gut dysbiosis contributes to amyloid pathology, associated with C/EBP β /AEP signaling activation in alzheimer's disease mouse model. *Sci Adv* (2022) 6:eaba0466. doi: 10.1126/sciadv.aba0466

502. Li W, Dowd SE, Scurlock B, Acosta-Martinez V, Lyte M. Memory and learning behavior in mice is temporally associated with diet-induced alterations in gut bacteria. *Physiol Behav* (2009) 96:557–67. doi: 10.1016/j.physbeh.2008.12.004

503. Kowalski K, Mulak A. Brain-Gut-Microbiota axis in alzheimer's disease. J Neurogastroenterol Motil (2019) 25:48-60. doi: 10.5056/jnm18087

504. Gubert C, Choo JM, Love CJ, Kodikara S, Masson BA, Liew JJM, et al. Faecal microbiota transplant ameliorates gut dysbiosis and cognitive deficits in huntington's disease mice. *Brain Commun* (2022) 4:fcac205. doi: 10.1093/braincomms/fcac205

505. Li Y, Ning L, Yin Y, Wang R, Zhang Z, Hao L, et al. Age-related shifts in gut microbiota contribute to cognitive decline in aged rats. Aging (2020) 12:7801–17. doi: 10.18632/aging.103093

506. Tilg H, Moschen AR. Food, immunity, and the microbiome. *Gastroenterology* (2015) 148:1107–19. doi: 10.1053/j.gastro.2014.12.036

507. Luciano M, Corley J, Cox SR, Valdés Hernández MC, Craig LCA, Dickie DA, et al. Mediterranean-Type diet and brain structural change from 73 to 76 years in a Scottish cohort. *Neurology* (2017) 88:449 LP – 455. doi: 10.1212/WNL.000000000003559

508. Ghosh TS, Rampelli S, Jeffery IB, Santoro A, Neto M, Capri M, et al. Mediterranean Diet intervention alters the gut microbiome in older people reducing frailty and improving health status: the NU-AGE 1-year dietary intervention across five European countries. *Gut* (2020) 69:1218–28. doi: 10.1136/gutjnl-2019-319654

509. Horn J, Mayer DE, Chen S, Mayer EA. Role of diet and its effects on the gut microbiome in the pathophysiology of mental disorders. *Transl Psychiatry* (2022) 12:164. doi: 10.1038/s41398-022-01922-0

510. Romanenko M, Kholin V, Koliada A, Vaiserman A. Nutrition, gut microbiota, and alzheimer's disease. *Front Psychiatry* (2021) 12:712673. doi: 10.3389/fpsyt.2021.712673

511. Leigh S-J, Morris MJ. Diet, inflammation and the gut microbiome: mechanisms for obesity-associated cognitive impairment. *Biochim Biophys Acta - Mol Basis Dis* (2020) 1866:165767. doi: 10.1016/j.bbadis.2020.165767

512. Ohland CL, Kish L, Bell H, Thiesen A, Hotte N, Pankiv E, et al. Effects of lactobacillus helveticus on murine behavior are dependent on diet and genotype and correlate with alterations in the gut microbiome. *Psychoneuroendocrinology* (2013) 38:1738–47. doi: 10.1016/j.psyneuen.2013.02.008

513. Agustí A, García-Pardo MP, López-Almela I, Campillo I, Maes M, Romaní-Pérez M, et al. Interplay between the gut-brain axis, obesity and cognitive function. *Front Neurosci* (2018) 12:155. doi: 10.3389/fnins.2018.00155

514. Proctor C, Thiennimitr P, Chattipakorn N, Chattipakorn SC. Diet, gut microbiota and cognition. *Metab Brain Dis* (2017) 32:1–17. doi: 10.1007/s11011-016-9917-8

515. Otaegui-Arrazola A, Amiano P, Elbusto A, Urdaneta E, Martínez-Lage P. Diet, cognition, and alzheimer's disease: food for thought. *Eur J Nutr* (2014) 53:1–23. doi: 10.1007/s00394-013-0561-3

516. Bruce-Keller AJ, Salbaum JM, Luo M, Blanchard E4, Taylor CM, Welsh DA, et al. Obese-type gut microbiota induce neurobehavioral changes in the absence of obesity. *Biol Psychiatry* (2015) 77:607–15. doi: 10.1016/j.biopsych.2014.07.012

517. Soto M, Herzog C, Pacheco JA, Fujisaka S, Bullock K, Clish CB, et al. Gut microbiota modulate neurobehavior through changes in brain insulin sensitivity and metabolism. *Mol Psychiatry* (2018) 23:2287–301. doi: 10.1038/s41380-018-0086-5

518. Jaric I, Voelkl B, Clerc M, Schmid MW, Novak J, Rosso M, et al. The rearing environment persistently modulates mouse phenotypes from the molecular to the behavioural level. *PloS Biol* (2022) 20:e3001837. doi: 10.1371/journal.pbio.3001837

519. Guo P, Lei M, Hu S, Xu Z, Zhou Y, Zhou P, et al. Long-term LDR exposure may induce cognitive impairments: a possible association through targeting gut microbiota-gutbrain axis. *Ecotoxicol Environ Saf* (2023) 249:114351. doi: 10.1016/j.ecoenv.2022.114351

520. Luo X-X, Yang C, Zhan G-F, Li S, Hua D-Y, Luo A-L, et al. Whole brain radiotherapy induces cognitive dysfunction in mice: key role of gut microbiota. *Psychopharmacology* (2020) 237:2089–101. doi: 10.1007/s00213-020-05520-0

521. Xiang C, Chen P, Zhang Q, Li Y, Pan Y, Xie W, et al. Intestinal microbiota modulates adrenomedullary response through Nod1 sensing in chromaffin cells. *iScience* (2021) 24:102849. doi: 10.1016/j.isci.2021.102849

522. Kostadinova F, Schwaderer J, Sebeo V, Brunner T. Why does the gut synthesize glucocorticoids? Ann Med (2014) 46:490–7. doi: 10.3109/07853890.2014.932920

523. Noti M, Sidler D, Brunner T. Extra-adrenal glucocorticoid synthesis in the intestinal epithelium: more than a drop in the ocean? *Semin Immunopathol* (2009) 31:237–48. doi: 10.1007/s00281-009-0159-2

524. Huang J, Jia R, Brunner T. Local synthesis of immunosuppressive glucocorticoids in the intestinal epithelium regulates anti-viral immune responses. *Cell Immunol* (2018) 334:1-10. doi: 10.1016/j.cellimm.2018.08.009

525. Merk VM, Phan TS, Brunner T. Regulation of tissue immune responses by local glucocorticoids at epithelial barriers and their impact on interorgan crosstalk. *Front Immunol* (2021) 12:672808. doi: 10.3389/fimmu.2021.672808

526. Talabér G, Jondal M, Okret S. Extra-adrenal glucocorticoid synthesis: immune regulation and aspects on local organ homeostasis. *Mol Cell Endocrinol* (2013) 380:89–98. doi: 10.1016/j.mce.2013.05.007

527. Ergang P, Vagnerová K, Hermanová P, Vodička M, Jágr M, Šrůtková D, et al. The gut microbiota affects corticosterone production in the murine small intestine. *Int J Mol Sci* (2021) 22:4229. doi: 10.3390/ijms22084229

528. Mukherji A, Kobiita A, Ye T, Chambon P. Homeostasis in intestinal epithelium is orchestrated by the circadian clock and microbiota cues transduced by TLRs. *Cell* (2013) 153:812–27. doi: 10.1016/j.cell.2013.04.020

529. Paolo WFJ, Nosanchuk JD. Adrenal infections. Int J Infect Dis IJID Off Publ Int Soc Infect Dis (2006) 10:343–53. doi: 10.1016/j.ijid.2005.08.001

530. Quénéhervé L, Drui D, Blin J, Péré M, Coron E, Barbara G, et al. Digestive symptoms in daily life of chronic adrenal insufficiency patients are similar to irritable bowel syndrome symptoms. *Sci Rep* (2021) 11:8077. doi: 10.1038/s41598-021-87158-2