

Pheochromocytomas and paragangliomas: clinical and genetic approaches

Marcia Helena Soares Costa¹, Tania M. Ortiga-Carvalho², Alice Dutra Violante³ and Mario Vaisman^{3*}

¹ Division of Endocrinology, Federal University of the State of Rio de Janeiro, Rio de Janeiro, Brazil, ² Laboratory of Translational Endocrinology, Carlos Chagas Filho Biophysics Institute, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil, ³ Division of Endocrinology, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil

OPEN ACCESS

Edited by:

Antongiulio Faggiano,
University of Naples Federico II, Italy

Reviewed by:

Massimo Mannelli,
University of Florence, Italy
Rossella Libé,
Hôpital Cochin, France

*Correspondence:

Mario Vaisman,
Serviço de Endocrinologia, HUCFF,
Rua Rodolpho Paulo Rocco,
255 Cidade Universitária,
Rio de Janeiro, RJ CEP 21941-913,
Brazil
vaisman@hucff.ufrj.br

Specialty section:

This article was submitted to Cancer
Endocrinology, a section of the
journal *Frontiers in Endocrinology*

Received: 20 May 2015

Accepted: 27 July 2015

Published: 17 August 2015

Citation:

Costa MHS, Ortiga-Carvalho TM,
Violante AD and Vaisman M (2015)
Pheochromocytomas and
paragangliomas: clinical and
genetic approaches.
Front. Endocrinol. 6:126.
doi: 10.3389/fendo.2015.00126

Pheochromocytomas (PCCs) and paragangliomas (PGLs) are neuroendocrine tumors derived from the chromaffin tissue. Diagnosis of these tumors is extremely important as they are linked to the hypertension syndrome with great cardiovascular morbidity and mortality. A great majority of PCCs and PGLs are sporadic and benign tumors; however, the classic idea of 10% exception of these features is changing. The description of new genes linked to familial forms of PCC/PGLs, such as succinate dehydrogenase (SDH) complex subunits, *KIF1B β* , *EGLN1*, *TMEM127*, and *MAX*, added to the well-known PCC familial syndrome (MEN2, VHL, and neurofibromatosis type 1) presents new challenges for diagnosis. In this review, we discuss the diversity of clinical and genetic approaches to this syndrome as well the diverse criteria that should guide genetic investigation.

Keywords: pheochromocytomas, MEN2, succinate dehydrogenase, VHL, neurofibromatosis

Introduction

Pheochromocytomas (PCCs) and paragangliomas (PGLs) are neuroendocrine tumors derived from the chromaffin tissue of the adrenal medulla or from extra-adrenal sympathetic and parasympathetic paraganglia. Parasympathetic PGLs are mostly non-secreting, whereas sympathetic PGLs generally produce catecholamine (1, 2).

The diagnosis of these neoplasias is extremely important, as they are associated with hypertension syndrome and great cardiovascular morbidity and mortality. The prevalence of PCC/PGLs in general hypertensive population is on average 0.1–0.6% and even higher in children (1.7%), with 5% of incidentally discovered adrenal mass (1, 3, 4). However, with modern imaging, 20–30% of these tumors are incidentally discovered (5). Therefore, high suspicion is necessary to avoid misdiagnosis.

Clinical Diagnosis Based on Genetic Aspects

The American Endocrine Society guidelines advocate that it is important to be aware of the main clinical features for proper diagnosis of PCC/PGLs (5). The clinical presentation of these tumors can overlap with other clinical conditions with non-specific symptoms, such as tachycardia, diaphoresis, pallor, anxiety, headaches, panic attacks, and sensation of sudden death. Hypertension may or may not be present; as previously described by an Italian group where 21% of patients with PCC were normotensive, and it can appear as a sustained sign or during paroxysm (6). In humans, blood pressure depends on many factors including the sympathetic nervous system and

the renin–angiotensin–aldosterone system among other factors. In patients with PCC, however, the behavior of hypertension depends also on the circulating catecholamine levels and eventually by other cardiovascular active substances secreted by the tumor. Moreover, production of vasodilator substances, such as dopamine or prostaglandins, downregulation of alpha-1 adrenergic receptors, and the presence of hypovolemia can contribute to this variability. Norepinephrine (NE)-secreting tumors usually cause sustained hypertension, while tumors that produce epinephrine (E) co-secreted with NE commonly result in episodic hypertension. Pure E-secreting tumors can present hypotension instead of high pressure as a symptom (7). Large cystic tumors are often asymptomatic because part of the catecholamine produced is metabolized inside the tumors before being released into circulation (8).

The heterogeneity of PCCs/PGLs clinical presentation explains the reason why this disease frequently is not considered as a possible diagnose condition, though can provoke a life-threatening crisis. At that point, any patient with the clinical manifestations cited before should be included as a case suspicion and must be properly screened for PCCs. These tumors occur in any age, but predominantly between 40 and 50 years, with approximately equal sex distribution (9, 10). However, the age of appearance can change in hereditary tumors, and they often occur at an earlier age. A great majority of PCCs and PGLs are sporadic and benign tumors; however, the classic idea of the traditional 10% rule for these features is changing. Occurrence of hereditary-associated tumors has increased to 30%. Also, diagnosis of several syndromes, such as multiple endocrine neoplasia type 2 (MEN2), von Hippel–Lindau disease (VHL), and neurofibromatosis type 1, has increased. Recently, additional mutations in the succinate dehydrogenase (SDH) complex subunits, *SDHA*, *SDHB*, *SDHC*, *SDHD*, and *SDHFA2*, related to familial PCC–PGL and novel susceptibility genes, such as *KIF1Bβ*, *EGLN1*, *TMEM127*, and *MAX*, have led to the diagnosis of new phenotypes (11, 12). Patient criteria, such as younger age, positive familial history for PCC/PGL, tumor bilaterally/multifocal, and recurrence or malignancy, could guide selection of patients who may be under greater risk of genetic abnormality (Figure 1). Genetic tests are expensive, and therefore, the knowledge of links between specific clinical and biochemical phenotype and PCC/PGLs-related genes and syndromes could reduce the cost of genetic screening significantly. This review aims to provide an overview of the genetic susceptibility of these tumors, clarifying and reinforcing the importance of their diagnosis.

Genes and Syndromes Related to PCC/PGL

Multiple Endocrine Neoplasia

Multiple endocrine neoplasia type 2 is described by medullary thyroid carcinomas (MTC) in association with PCC. This syndrome has three clinical variants: MEN2A (55% of cases), familial medullary thyroid carcinoma (FMTC – 35–40%), and MEN2B (5–10%) (13). The disease is caused by the germ-line mutation of the proto-oncogene, *RET* (rearranged during transfection). This 21-exon proto-oncogene is located on chromosome 10 and encodes a receptor tyrosine kinase that works in conjunction with glial-derived neurotrophic factor family ligands. The interaction

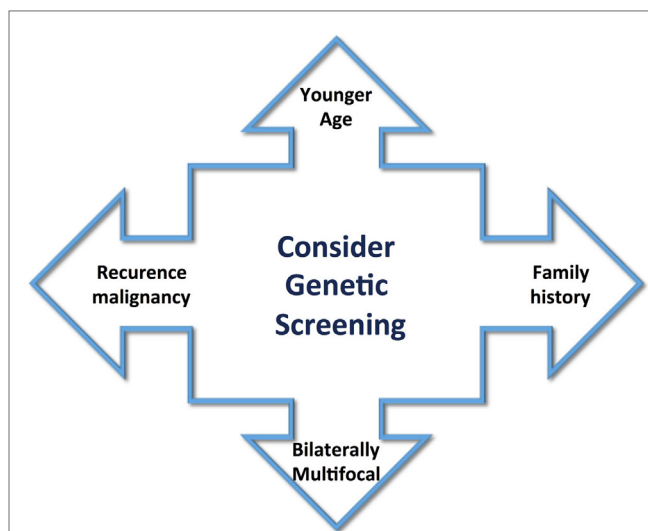


FIGURE 1 | Clinical aspects to guide the screening for genetic abnormality include younger age of tumor appearance, positive family history, bilaterally multifocal tumors, and recurrence or malignancy.

of these ligands results in the dimerization of the *RET* receptor, auto-phosphorylation of tyrosine residues, and finally cell growth and survival mediated by the mitogen-activated protein kinase intracellular signaling cascade. One germ-line activating mutation of this gene, generally missense and located in exons 10, 11, 13, 14, 15, 16, has been implicated in this syndrome (14, 15). MEN2A is also associated with hyperparathyroidism in 20–30% of the patients, unlike MEN2B, in which this feature is absent and marfanoid habitus calls attention. MEN2B tumors are more aggressive, with an earlier onset and higher mortality.

MEN2 types carry approximately 100% risk for MTC and at 50% risk for PCC. FMTC tumors are less aggressive and, by definition, without incidence of other endocrine neoplasia. There is a strong correlation between the phenotype and genotype in the clinical subtypes of MEN2 with respect to age of onset and aggressiveness of MTC. These aspects are used to determine the time of prophylactic thyroidectomy and whether patients should be screened for PCC and hyperparathyroidism. In this way, positive results for a specific *RET* mutation can influence the management of an apparently sporadic MTC.

Pheochromocytomas in MEN2A and B are usually benign, bilateral in half of the patients, and can rarely be extra-adrenal. The malignancy affects <5% of PCCs in this syndrome (16, 17). Mutation in codons 618, 620, and 634 of *RET* is representative of MEN2A in approximately 80% of patients. A single missense mutation in codon 918 is found in nearly 90% of MEN2B cases. Based on this genotype and phenotype correlation, the American Thyroid association has proposed the classification of four risk-categorized mutations and recommends the screening for PCC by the age of 20 years for MEN2A and by 8 years for MEN2B (18). One group has reported a high penetrance of PCCs, especially bilateral and large tumors, associated with a double C634/Y719F germ-line mutation in the *RET* protooncogene (19).

Von Hippel–Lindau Syndrome

Von Hippel–Lindau syndrome is an autosomal dominant disease associated with inactivating mutations in the *VHL* tumor suppressor gene located on chromosome 3. This disease is characterized by a multiplicity of benign and malignant tumors, including retinal and central nervous system (CNS) hemangioblastomas, pancreatic islet cell tumors and pancreatic cysts, epididymal cystadenomas, endolymphatic sac tumors, clear-cell renal carcinomas and PCCs, and PGLs (20–23).

This syndrome is divided into type 1 or type 2 based on the absence or presence of PCC, respectively. The *VHL* type 2 is further classified as 2A for patient with low risk of developing clear renal cell carcinoma, type 2B for patients with high risk of these tumors, and 2C for patients with only PCCs in the absence of the others classical tumors of the syndrome (23, 24). *VHL* mutations result in unilateral or bilateral PCCs and rarely in PGLs. Approximately half of the cases are bilateral and benign tumors, with malignant phenotype present in <5% of the patients (10, 25). The mean age at diagnosis of these tumors is in the second decade, generally approximately 30 years, and the absence of signs and symptoms is frequent. PCCs and PGLs are the first manifestation in approximately 30–50% of the cases. Due to these aspects, it has been proposed that catecholamine screening should be started at approximately 5 years old, especially in patients with a positive familial history of PCCs (12, 26).

There are some correlations between the genotype and the phenotype in this syndrome. The *VHL* gene comprise three exons, and its mutations are distributed throughout the coding sequence; missense, nonsense, splice-site mutations, micro deletions, and insertions are found in two-thirds of patients, while large deletions are described in only one-third of patients with this syndrome (20, 27). Deletions, nonsense, and frame shift mutations are regularly found in *VHL* type 1, whereas missense mutations are more common in *VHL* type 2. A majority of patients with PCC have missense mutations. Maher et al. (20) analyzed the genotype–phenotype correlations in 573 individuals with *VHL* disease and demonstrated that classifying missense substitutions according to their predicted effect on pVHL structure enhances the ability to predict pheochromocytoma risk. Although genetic screening has been able to identify mutations in the majority of patients with *VHL* disease, some patients (20%) without genetic diagnosis probably develop *de novo* mutations (28).

Neurofibromatosis Type 1

Neurofibromatosis type 1 (NF1) or Von Recklinghausens's disease is an autosomal dominant syndrome characterized by *café au lait* macules, cutaneous or subcutaneous neurofibromas, skinfold freckling, and iris Lisch nodules. These characteristics occur in an average of 90% of all NF1 adult patients, but the number of lesions is quite variable. Several other tumors, such as nerve glioma, dysplasia of the long bones, peripheral nerve sheath tumors, gastrointestinal stromal tumors (GIBT), rhabdomyosarcomas, breast cancer, and chronic myeloid leukemia, have also been described. Some patients can also present cognitive impairment (29, 30).

Pheochromocytomas are a rare finding in NF1, being identified in <6% of cases; however, they have a higher prevalence (13%) in autopsy series. The preferential mean age of presentation is in

the fourth decade. A majority of these PCCs are unilateral and benign, while approximately 10% can be bilateral or malignant. The presence of PGLs is not common in NF1 (31, 32). Considering these aspects, the main features of PCCs in NF1 are very similar to those of sporadic PCCs.

NF1 is a tumor suppressor large gene with 60 exons located on chromosome 17 with described missense, nonsense, and splice-site mutations, indels, and chromosomal rearrangements (33). These mutations are spread all over the gene without a hot spot, and the majority are located in the exons with variable penetrance and expression (32).

The *NF1* protein, neurofibromin, is a GTPase that suppresses cell proliferation by inactivating RAS and inhibiting the RAS/RAF/MAPK signaling pathways. When this gene is mutated, the cascade including PI3K and mTor is activated, subsequently resulting in uncontrolled cellular growth and differentiation (34). Some studies have suggested a possible novel potential NF1 genotype–phenotype correlation, especially with splice-site mutations associated with an increased tendency to develop neoplasms as CNS gliomas and malignant peripheral nerve sheath tumors (12). However, the diagnosis of the NF1 is established mainly based on the clinical aspects because it is difficult to genetically screen in the absence of hot spot mutations.

Familial PCC/PGLs

SDHx-Associated PCCs and PGLs

In the last decade, the genetics of hereditary PCC/PGLs has started to emerge with the description of germ-line mutations in the SDHx (SDH) genes related to this syndrome. The SDH complex is described as a heterotetrameric protein involved in the electron transport in the Krebs's cycle and mitochondrial respiratory chain. This enzyme is composed of four subunits (SDHA, SDHB, SDHC, and SDHD) that catalyze the oxidation of succinate to fumarate. The catalytic subunits, SDHA and SHDB, are inserted in mitochondrial matrix, and they are affixed to the inner membrane by subunits SDHC and SDHD. The SDHx genes are supposed to work as classical tumors suppressors, as tumors frequently show loss of heterozygosity (LOH) of the non-mutated allele.

The relations between SDH and PCC/PGLs emerged in the 2000s when germ-line *SDHD* mutations were associated to the occurrence of PCC/PGLs (35). *SDHD* is located on chromosome locus 11q23, has four exons, and is maternally imprinted (36, 37). The disease susceptibility occurs when the mutation is inherited from the father with rare exceptions.

SDHD mutations are linked to the development of familial PGLs and PCCs (PGL1), although they have been reported in apparently sporadic tumors; they are often associated with the risk of multiple tumors, especially head and neck PGLs (38, 39). PCC can occur usually unilateral (40). Malignancy associated with *SDHD*-derived PCC/PGL is quite rare.

The PGL2 syndrome is associated with *SDHAF2* mutations, also known as SDH5. The *SDHA* mutation has been described with Leigh's disease, a severe type of encephalopathy. The *SDHA* maps to chromosome 5p15, its products undergo post-translational modification by *SDHAF2*, and the gene of this protein has been implicated in development of hereditary head and neck PGLs (41, 42). Mutations in *SDHAF2* are considered rare; in a study with

443 patients with head and neck PGL evaluated, mutations on the *SDHAF2* gene were negative.

Mutations in *SDHC* are implicated in PGL3 syndrome. The gene is located on chromosome 1q23.3 and presents six exons. A mutation in *SDHC* is less frequent when compared to *SDHD* mutations; it is usually associated with benign solitary head and neck PGLs. Also, extra-adrenal PGLs and PCCs have been described in this syndrome. Malignancy associated with the *SDHC* gene is extremely rare (43).

Mutation in *SDHB* is implicated in PGL 4 syndrome, characterized mostly by abdominal, pelvic, and thoracic catecholamine-secreting familial PGL. The *SDHB* gene is located on chromosome 1p36.1, with eight exons. The risk of malignancy is great with this disease; less frequently, these mutations are found in PCCs or parasympathetic PGLs. The associated risk of malignancy is difficult to determine, especially because metastases can occur up to 20–30 years after primary tumors, and the applied definitions of malignancy vary between different studies (44).

Furthermore, *SDHB* mutations may result in susceptibility to other malignant tumors, such as renal cell carcinomas, papillary thyroid tumors, neuroblastoma, or gastrointestinal stromal tumor (GIST). Average 20% of patients with *SDHB* mutation will present malignant PGLs, and more than 50% of the individuals with malignant PGLs will have a *SDHB* mutation (40, 44). Recently, mutations in this gene and in *SDHD* and *SDHC* were described in association with Carney–Stratakis syndrome, an autosomal dominant disorder characterized by PGLs and GIST, but they have not been found in other forms of the Carney triad, suggesting that other genetic alterations may be implicated in this syndrome (45).

More efforts are needed to define the risk of other cancers in association with these mutations in order to help clinicians better design a phenotype and genotype correlation in PCC/PGLs and understand the complexity of these diseases.

Carney Triad and Carney–Stratakis Syndrome

Carney triad is a syndrome of tumors involving at least five organs, the stomach, the lung, the paraganglionic system, the adrenal (cortex and medulla), and the esophagus. The stomach features GIST, the lung chondroma, the paraganglionic system paraganglioma, the adrenal adenoma and pheochromocytoma, and the esophagus leiomyoma (46).

The etiology of the Carney triad syndrome is unknown; no candidate gene has been discovered until now and the disease does not appear in a family pattern. The younger age and female predilection of affected individuals do implicate in a link with an inherited genetic defect; however, the screening of *SDHA*, *SDHB*, *SDHC*, and *SDHD* mutations has been always negative. The mutated genes, often associated to GIST (*KIT* and *PDGFRA*), are also not affected.

Differently, Carney–Stratakis syndrome is inherited in an autosomal-dominant manner that includes PGLs and GIST without pulmonary chondromas (47). This disease affects equally men and women and the majority of patients present germ-line mutations in *SDHB*, *SDHC*, or *SDHD* supporting another molecular mechanism linked to GIST besides *KIT* and *PDGFRA* mutations usually described (48).

The associated PCC and PGLs in these syndromes can be different. In Carney triad disease, the majority of patients present with PGLs, including sympathetic and parasympathetic tumors, with 28 years old of mean age presentation; 11% of the patients present metastasis (49). In Carney–Stratakis syndrome, the majority of patients have a sympathetic or parasympathetic PGLs, with a mean age of 33 years old; however, no malignant tumors were described (47).

New PCCs and PGLs Susceptibility Genes

Despite identification of multiple genetic causes of PGLs and PCCs in the last decade, around two-thirds of these tumors remain without molecular diagnosis, suggesting that other susceptibility genes could be implicated. Using a wide-genomic approach, PCC/PGLs were clustered in two major groups depending on their global transcription profiles. Cluster 1 includes *VHL*, *SDHx*, FH-mutated tumors and a part of the sporadic PCC/PGLs. This group was composed of tumors without clear mutation or sporadic tumors; they showed signatures of pseudo hypoxia, angiogenesis, and decreased oxidoreductase response. This profile links these tumors with hypoxia-inducible factor (HIF) role, and overexpression of HIF1 α and 2 α were found in this cluster.

A second group (cluster 2) covered PCCs with *RET*, *NF1*, *KIF1B β* , and undefined tumors enriched for kinase receptor signaling pathways, translation initiation, protein synthesis, and genes involved in neural/neuroendocrine identity (50–52). Novel genes, such as *TMEM127*, were also identified in this cluster (53). Also, deregulation of another pathway involving egl nine homologs was suggested (54).

KIF1B β is a tumor suppressor gene necessary for neuronal apoptosis. It is located on chromosome 1 (55). Mutation in this gene was described in patients with PCC without other predisposing mutations. No specific phenotype has been identified yet; however, patients with *KIF1B β* seem to be prone to PCC and neuroblastomas. Ganglioneuromas, leiomyosarcomas, and lung adenocarcinomas have also been reported (50).

In 2008, a germ-line mutation in *EGLN1*, also called *PHD2*, was reported in a patient with erythrocytosis and recurrent para-aortic PGL. *EGLN1* is a member of EGLN prolyl hydroxylase family that plays a major role regulating the HIF, which is involved in angiogenesis, erythropoiesis, cell metabolism, and proliferation. This gene was linked to familial erythrocytosis but not in association with tumors. A mutation analysis of 82 patients with inherited PCC did not detect mutations in *EGLN1*, *EGLN2*, or *EGLN3*, suggesting that mutations in these genes are not a frequent cause of inherited PCC (56).

TMEM127 was recently identified as a PCC/PGLs susceptibility gene using linkage analysis and transcription profiling by microarray and copy number analysis of these tumors. It is possible that *TMEM127*, located on chromosome locus 2q11.2, could be a tumor suppressor gene playing a role in cell signaling associated with kinase receptor signals and as a negative regulator of mTOR, which promotes cell growth and protein translation (53). The most common clinical presentation of the mutation carriers is quite similar to sporadic PCC patient, single adrenal tumor with mean age of 40 years (57, 58), although multiple head and neck PGLs and retroperitoneal PGLs have been described.

Unilateral and bilateral tumors have also been found, but malignancy is rare.

MYC-associated factor X (MAX) is a transcription factor that was recently linked to PCC/PGL susceptibility. MAX is located on chromosome locus 14q23 and was associated with other cancers, such as neuroblastomas, acting as an oncogene or interacting with MYC family as a transcriptional suppressor (59, 60). Whole genome sequencing identified MAX germ-line mutations in familial cases of PCC without alterations in known genes. Comino-Mendez et al. studied 12 PCC patients with MAX mutations; three of the cases were unrelated individuals with hereditary PCC (59). The absence of MAX protein in the tumors and LOH of the wild-type allele in these tumors confirmed the tumor suppressor role of this gene in humans. Eight of these patients (67%) had bilateral PCCs, and 25% presented metastasis at diagnosis, suggesting that MAX mutation can be associated to multicentricity and malignant risk.

Genes and Sporadic PCC/PGLS

A problem that concerns clinicians is whether all sporadic PCCs should be screened for unsuspected germ-line mutations in genes associated with hereditary PGLs or PCCs. This aspect is not well clarified in the literature; the studies usually not include the analyses of all genes implicated in the familial forms. In a review by Jiménez et al., the authors provide a compilation of several studies, and the results suggest that 20% of patients with apparently sporadic PCCs have a germ-line mutation of one of the genes known to cause hereditary PCC/PGLs. When the authors excluded patients with bilateral or multicentric tumors with larger probability of familial disease, the amount was reduced to 17% (5.04% for *VHL*, 6.38% for *PGL4*, 3.72% for *PGL1* e 1.55% for *RET*) (61).

In one study developed by Neumann et al., it was found that 24% of the patients with apparently non-syndromic pheochromocytomas and no family history of the disease had mutations in *VHL*, *RET*, *SDHD*, and *SDHB* genes; these patients presented at a younger age (average of 24 years), and they had multiple tumors and 28% extra-adrenal tumors (62). Another study developed by the same group reinforced that up to 35% of patients with PCC/PGLs can be associated to an inherited mutation in these genes (32).

A Spanish group analyzed 135 apparently sporadic patients with a single tumor for the five susceptibility genes: *VHL*, *RET*, *SDHB*, *SDHC*, and *SDHD*. They found that 14% of the cases harbor germ-line mutations; a majority (98%) of them had tumors with younger onset, not more than 45 years old (63).

Demographic, clinical, and genetic evaluation was performed in a series of 71 patients with PCC/PGLs. Among 59 patients with apparently sporadic disease, unsuspected germ-line mutations occurred in 8 cases (13.6%). The authors did not find any difference between hereditary and sporadic disease concerning age, sex, and tumor size; bilateral and recurrent diseases were most common in hereditary cases (64).

Currently, the overall hereditary predisposition of PCCs is estimated to be approximately 20–35%. The high prevalence of unsuspected mutations indicates a demand for more extensive genetic tests for patients with these tumors (65). However, we have to be aware of the aspects that can influence the prevalence

of the PCC/PGL mutations: tertiary care centers are more prone to evaluate patients with multiple and more complicated PCCs. Specific genetic disorders are more prevalent in some geographic regions, and major effort is being currently contributed to easier recognition of these syndromes. An Endocrine Society taskforce that compiled 31 studies including 5031 patients with apparently sporadic PCC/PGLs screened for numerous genes, and the taskforce reinforces this idea, as they found approximately 11–13% germ-line mutations in this population (66).

Somatic mutations in sporadic pheochromocytoma/paraganglioma has been identified in *VHL*, *RET*, *SDHB*, and *SDHD* but their frequency was reported to be low. The COMETE cohort has demonstrated the occurrence of somatic mutations in *VHL* and *RET* genes in 14% of sporadic pheochromocytoma/paraganglioma, one germ-line, and three somatic mutations in the MAX were described by the same cohort (67, 68). Burnichon et al. have found that 41% (25/61) of the sporadic tumors in their population harbor an inactivating somatic mutation in the NF1 gene associated with loss of the wild-type allele in 84% (21/25) of cases (69). These findings reinforce that somatic mutation in sporadic PCC/PGLs can be more often than previously considered.

Genetic Screening Strategy in Patients with PCC and PGL

The American Society of Clinical Oncology suggests that all patients with PCC and PGLs should be submitted to genetic screening. It is currently accepted from the literature that approximately 30–35% of the PCC/PGLs are associated with germ-line mutations in one of the genes described above (70).

Molecular analysis has revealed that a significant number of PCC/PGLs, previously considered as sporadic tumors, have a genetic predisposition. However, in the last decade, several new genes were implicated in this disease, and we have considered the cost and availability of genetic tests before using them as diagnostic tools in PCC/PGLs.

Some characteristics, including personal or family history of PCC/PGLs, are very important. A detailed and medical family history and sometimes a specialized genetic consultation provide the potential consequences of genetic testing. In the absence of any family history, the description of sudden death without other cardiovascular disease should call attention. Clinical manifestations in the patient or other family members can point to a particular hereditary condition. For example, subtle retinal vascular lesions can indicate the possibility of *VHL* disease.

Generally, hereditary tumors occur at a younger age than the sporadic ones; age of diagnostics younger than 40 years is an important factor to call for a genetic test. An interesting study suggests that patients with epinephrine-producing tumors (mean age 50 years) were diagnosed 11 years later than those with tumors lacking appreciable epinephrine production (mean age 42 years). This finding reinforces that the cut-off age for genetic screening could consider tumor catecholamine phenotypes (71). However, finding nearly 36% germ-line mutations in children make these tests necessary in this group of patients (72). Presence of head and neck PGLs, bilateral or multifocal tumors, and the presence of malignancy should raise the suspicion of a possible hereditary abnormality (73, 74).

For family diseases, such as MEN2A/B, NF1, and VHL, a phenotype suspicion, because they are commonly associated to other tumors, helps to direct genetic studies. The presence of hemangioblastomas calls attention for VHL disease, as the association of medullary thyroid carcinoma for MEN2. MEN2-associated

tumors always produce epinephrine and can be uni- or bilateral, but malignant disease and extra-adrenal tumors are rare in this syndrome. PCCs in VHL always secrete NE and can be extra-adrenal. The biochemical profile, as shown in **Figure 2**, is very relevant to help and guide the genetic test. Diagnosis of NF1 is based

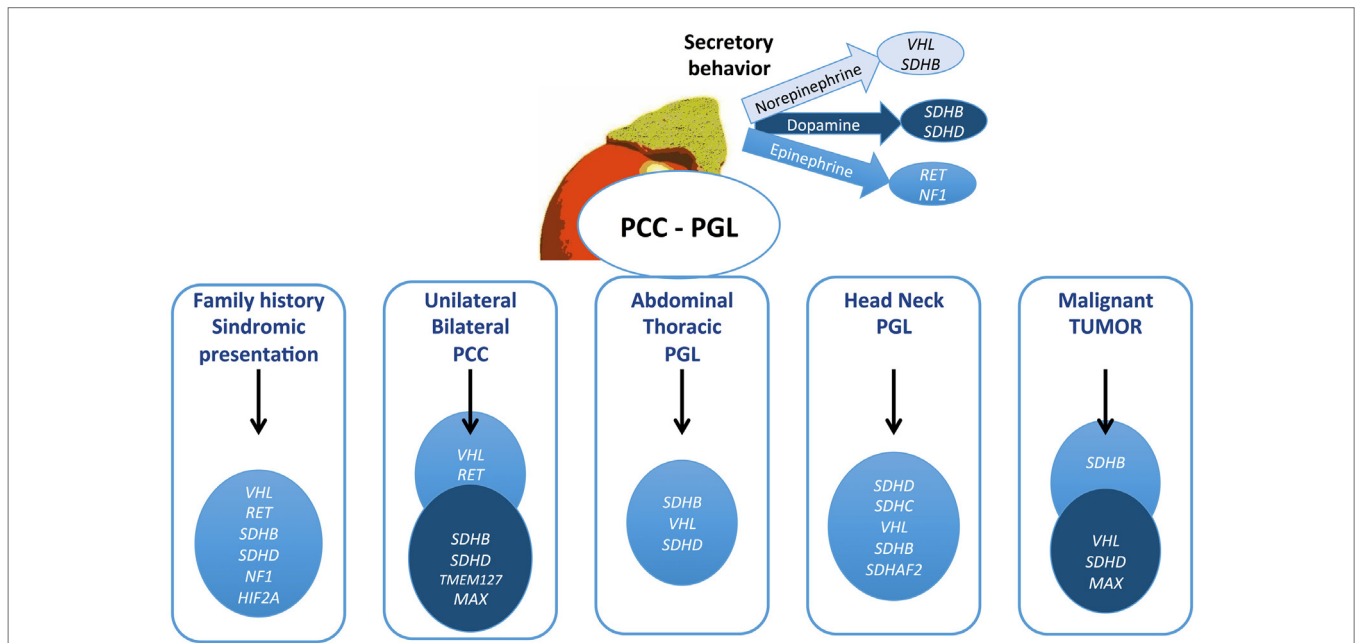


FIGURE 2 | Algorithm proposed for genetic testing for pheochromocytomas and paragangliomas patients. The gene described might be considered for testing in this order. Mutations in *TMEM127*, *MAX*, *HIF2A*, and *SDHAF2* are quite rare and should be contemplated in patients with negative test for the other genes.

TABLE 1 | Clinical and genetic features of the most frequent pheochromocytomas and paragangliomas-related syndromes.

| Gene | Loci | Protein function | Syndrome | Clinical manifestations or associated TU | Inheritance | Primary location | Malignancy rate |
|----------------|---------|-------------------------------|----------------|---|-------------|------------------------------------|-----------------|
| <i>RET</i> | 10q11.2 | Transmembrane tyrosine kinase | MEN2A MEN2B | MTC, Hyperparathyroidism, cutaneous lichen amyloidosis MTC, mucosal neuromas, marfanoid habitus | AD | PCC (single ou bilateral) | <5% |
| <i>VHL</i> | 3p25–26 | Ubiquitin ligase 3E activity | VHL | Hemangioblastomas, clear-cell renal cell carcinomas; endolymphatic sac tumors, cystadenomas; pancreatic islet cell tumor | AD | PCC (>bilateral) | <5% |
| <i>NF1</i> | 17q11.2 | GTPase | NF1 | Café au lait macules, neurofibromas, iris Lisch nodules, optic nerve gliomas, dysplasia of the long bones, astrocytomas, soft tissue sarcomas, CML, childhood learning disabilities, seizures | AD | PCC (>single) | 12% |
| <i>SDHD</i> | 11q23.1 | Complex II anchoring subunit | PGL1 | Papillary thyroid cancer; GIST | AD; PI | HNPGL/MPGL | <5% |
| <i>SDHAF2</i> | 11q13.1 | Cofactor for complex II | PGL2 | GIST | AD; PI | HNPGL/MPGL | Low |
| <i>SDHC</i> | 1q23.3 | Complex II-anchoring subunit | PGL3 | GIST | AD | HNPGL > SPCC = TAPGL = MPGL | Low |
| <i>SDHB</i> | 1p36.1 | Complex II catalytic subunit | PGL4 | Renal cell carcinomas; GIST, thyroid tumors, neuroblastomas | AD | TAPGL > HNPGL = SPGL = MPGL < SPCC | 30–70% |
| <i>SDHA</i> | 5p15 | Complex II catalytic subunit | | | AD | TAPGL/HNPGL | Low |
| <i>TMEM127</i> | 2q11.2 | Transmembrane protein | | | AD | SPCC > BPCC > TAPGL = HNPGL = MPGL | Low |
| <i>MAX</i> | 14q23–3 | BHL HLZ transcription factor | | | ADPI | SPCC = BPCC > TAPGL | 10% |

AD, autosomal dominant; PI, paternal inheritance; TAPGL, thoracic or abdominal PGL; HNPGL, head and neck PGL; SPCC/PGL, single PCC/PGL; BPC/PGL, bilateral PCC/PGL; MPGL, multiplal PGL.

on clinical features, and genetic testing is usually not required, because *NFI* is a large gene, costly, and difficult to screen.

Adrenal (unilateral or bilateral) PCCs are frequently associated with *VHL* and *RET*, while extra-adrenal sympathetic PGLs (abdominal or thoracic) are usually caused by *SDHB*, *SDHD*, and *VHL* mutations (75–77). Head and neck tumors are mainly caused by *SDH* mutations, especially *SDHD* and *B*, and less often by *VHL* and *SDHC* (25, 40). *SDHC* mutations have been described exclusively in patients with parasympathetic PGLs and quite rare in association with sympathetic PCC/PGLs (43). Tests for this gene are only indicated in these cases. Genetic test for *SDHB* is indicated in all patients with malignant PGL or PCC, and up of 50% of these tumors have an *SDHB* mutation. Elevation of dopamine with NE has been found in these tumors. Unlike classical syndromes, only 10% of the *SDHB* patients have a positive family history.

TMEM127 is customarily associated with benign PCC but not with PGL. Ordinarily, these tumors occur in old age, mimicking a truly sporadic PCC (57, 64).

Routine screening for germ-line mutations in patients with PCC/PGLs is costly and time consuming. Although some algorithms have been proposed, no routine guideline is available (73). For these reasons, the genetic screening directions are mostly based on clinical expertise, including clinical signs, the biochemical phenotype, and symptoms (Figure 2; Table 1). Understanding the genetic profile of these tumors and considering the patient's risk for developing multifocal lesions, malignant PCC/PGLs, and other malignancies provide a very important tool for following patients with this condition. Regarding these aspects, we suggest to screening all patients with multiple tumors, especially with extra-adrenal and head and neck tumors. Age is an important factor; nevertheless, some studies have demonstrated that the age range can be quite wide, even for familial forms. The genes might be screened according to clinical presentation; then cost and effective analyses, screening of *SDHB* and *VHL* should be considered. The most important clinical aspects related to different genes are provided in Table 1. Head and neck PGLs should be first screened for *SDHD* and *SDHC* mutations.

References

- Lenders JW, Eisenhofer G, Mannelli M, Pacak K. Pheochromocytoma. *Lancet* (2005) **366**:665–75. doi:10.1016/S0140-6736(05)67139-5
- Cohen DL, Fraker D, Townsend RR. Lack of symptoms in patients with histologic evidence of pheochromocytoma: a diagnostic challenge. *Ann N Y Acad Sci* (2006) **1073**:47–51. doi:10.1196/annals.1353.005
- Swiglo BA, Murad MH, Schünemann HJ, Kunz R, Vigersky RA, Guyatt GH, et al. A case for clarity, consistency, and helpfulness: state-of-the-art clinical practice guidelines in endocrinology using the grading of recommendations, assessment, development, and evaluation system. *J Clin Endocrinol Metab* (2008) **93**:666–73. doi:10.1210/jc.2007-1907
- Sinclair AM, Isles CG, Brown I, Cameron H, Murray GD, Robertson JW. Secondary hypertension in a blood pressure clinic. *Arch Intern Med* (1987) **147**:1289–93. doi:10.1001/archinte.147.7.1289
- Lenders JWM, Duh Q-Y, Eisenhofer G, Gimenez-Roqueplo A-P, Grebe SKG, Murad MH, et al. Pheochromocytoma and paraganglioma: an endocrine society clinical practice guideline. *J Clin Endocrinol Metab* (2014) **99**:1915–42. doi:10.1210/jc.2014-1498
- Mannelli M, Ianni L, Cilotti A, Conti A. Pheochromocytoma in Italy: a multicentric retrospective. *Eur J Endocrinol* (1999) **141**:619–24. doi:10.1530/eje.0.1410619
- Bravo EL, Tagle R. Pheochromocytoma: state-of-the-art and future prospects. *Endocr Rev* (2003) **24**:539–53. doi:10.1210/er.2002-0013
- Bravo EL, Tarazi RC, Fouad FM, Textor SC, Gifford RW, Vidt DG. Blood pressure regulation in pheochromocytoma. *Hypertension* (1982) **4**:193–9. doi:10.1161/01.HYP.4.3.193
- Young WF. Adrenal causes of hypertension: pheochromocytoma and primary aldosteronism. *Rev Endocr Metab Disord* (2007) **8**(4):309–20. doi:10.1007/s11154-007-9055-z
- Mannelli M, Castellano M, Schiavi F, Filetti S, Giacchè M, Mori L, et al. Clinically guided genetic screening in a large cohort of Italian patients with pheochromocytomas and/or functional or nonfunctional paragangliomas. *J Clin Endocrinol Metab* (2009) **94**:1541–7. doi:10.1210/jc.2008-2419
- Welander J, Söderkvist P, Gimm O. Genetics and clinical characteristics of hereditary pheochromocytomas and paragangliomas. *Endocr Relat Cancer* (2011) **18**:R253–76. doi:10.1530/ERC-11-0170
- Fishbein L, Nathanson KL. Pheochromocytoma and paraganglioma: understanding the complexities of the genetic background. *Cancer Genet* (2012) **205**:1–11. doi:10.1016/j.cancergen.2012.01.009
- Thakker RV. Multiple endocrine neoplasia. *Horm Res* (2001) **56**:67–72. doi:10.1159/000048138
- Brandi ML, Gagel RF, Angeli A, Bilezikian JP, Beck-Peccoz P, Bordi C, et al. Consensus: guidelines for diagnosis and therapy of MEN type 1

Currently, it is recommended that if a mutation is found, it is not necessary to search for additional genomic alterations; however, with the description of new genes, we have to be aware of new possibilities. Identification of a gene mutation in patients with PCC/PGLs results in precocious diagnosis and better surveillance, resulting in better prognosis. In specific cases, *RET* screening and genotype and phenotype correlation permits prophylactic surgery for an aggressive tumor. Nevertheless, further clinical studies are indispensable to better correlate genotype and phenotype features with the new genes.

Conclusion

To guide better screening strategy and optimize therapeutic options, the knowledge of PCC/PGL-implicated syndromes and genes is incredibly important. This review summarizes the main syndromes and the gene associated with these diseases. Further studies are needed for wider genotype to phenotype correlations, especially for new genes and pathways recently described for these tumors.

Author Contributions

MC, TO-C, AV, and MV contributed to the conception and design of the collection of the data for the revision, drafted the work, and critically revised the intellectual content. All the authors approved the last version of the manuscript and agreed to be accountable for all aspects of the work.

Acknowledgments

We thank the staff at the Endocrinology Division of Federal University of Rio de Janeiro and the staff at the Laboratory of Translational Endocrinology (UFRJ). This work was supported by FAPERJ (Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro, Grant: E26/11.845/2012, 112.063/2012, 102.873/2012) and CNPq (303734/2012-4).

- and type 2. *J Clin Endocrinol Metab* (2001) **86**:5658–71. doi:10.1210/jcem.86.12.8070
15. Eng C, Clayton D, Schuffenecker I, Lenoir G, Cote G, Gagel RF, et al. The relationship between specific RET proto-oncogene mutations and disease phenotype in multiple endocrine neoplasia type 2. International RET mutation consortium analysis. *JAMA* (1996) **276**:1575–9. doi:10.1001/jama.276.19.1575
 16. Quayle FJ, Fialkowski EA, Benveniste R, Moley JF. Pheochromocytoma penetrance varies by RET mutation in MEN 2A. *Surgery* (2007) **142**:800–5. doi:10.1016/j.surg.2007.09.013
 17. Rodriguez JM, Balsalobre M, Ponce JL, Rios A, Torregrosa NM, Tebar J, et al. Pheochromocytoma in MEN 2A syndrome. Study of 54 patients. *World J Surg* (2008) **32**:2520–6. doi:10.1007/s00268-008-9734-2
 18. American Thyroid Association Guidelines Task Force, Kloos RT, Eng C, Evans DB, Francis GL, Gagel RF, et al. Medullary thyroid cancer: management guidelines of the American thyroid association. *Thyroid* (2009) **19**:565–612. doi:10.1089/thy.2008.0403
 19. Toledo RA, Wagner SM, Coutinho FL, Lourenço DM, Azevedo JA, Longuini VC, et al. High penetrance of pheochromocytoma associated with the novel C634Y/Y791F double germline mutation in the RET protooncogene. *J Clin Endocrinol Metab* (2010) **95**:1318–27. doi:10.1210/jc.2009-1355
 20. Maher ER, Webster AR, Richards FM, Green JS, Crossey PA, Payne SJ, et al. Phenotypic expression in Von Hippel-Lindau disease: correlations with germline VHL gene mutations. *J Med Genet* (1996) **33**:328–32. doi:10.1136/jmg.33.4.328
 21. Benn DE, Gimenez-Roqueplo A-P, Reilly JR, Bertherat J, Burgess J, Byth K, et al. Clinical presentation and penetrance of pheochromocytoma/paraganglioma syndromes. *J Clin Endocrinol Metab* (2006) **91**:827–36. doi:10.1210/jc.2005-1862
 22. Zbar B, Kishida T, Chen F, Schmidt L, Maher ER, Richards FM, et al. Germline mutations in the Von Hippel-Lindau disease (VHL) gene in families from North America, Europe, and Japan. *Hum Mutat* (1996) **8**:348–57. doi:10.1002/(SICI)1098-1004(1996)8:4<348::AID-HUMU8>3.0.CO;2-3
 23. Cruz JB, Fernandes LPS, Clara SA, Conde SJ, Perone D, Kopp P, et al. Molecular analysis of the Von Hippel-Lindau (VHL) gene in a family with non-syndromic pheochromocytoma: the importance of genetic testing. *Arq Bras Endocrinol Metabol* (2007) **51**:1463–7. doi:10.1590/S0004-27302007000900008
 24. Weirich G, Klein B, Wöhl T, Engelhardt D, Brauch H. VHL2C phenotype in a German Von Hippel-Lindau family with concurrent VHL germline mutations P81S and L188V. *J Clin Endocrinol Metab* (2002) **87**:5241–6. doi:10.1210/jc.2002-020651
 25. Amar L, Bertherat J, Baudin E, Ajzenberg C, Bressac-de Paillerets B, Chabre O, et al. Genetic testing in pheochromocytoma or functional paraganglioma. *J Clin Oncol* (2005) **23**:8812–8. doi:10.1200/JCO.2005.03.1484
 26. Delman KA, Shapiro SE, Jonasch EW, Lee JE, Curley SA, Evans DB, et al. Abdominal visceral lesions in Von Hippel-Lindau disease: incidence and clinical behavior of pancreatic and adrenal lesions at a single center. *World J Surg* (2006) **30**:665–9. doi:10.1007/s00268-005-0359-4
 27. Neumann HP, Bender BU. Genotype-phenotype correlations in Von Hippel-Lindau disease. *J Intern Med* (1998) **243**:541–5. doi:10.1046/j.1365-2796.1998.00336.x
 28. Galan SR, Kann PH. Genetics and molecular pathogenesis of pheochromocytoma and paraganglioma. *Clin Endocrinol (Oxf)* (2013) **78**:165–75. doi:10.1111/cen.12071
 29. Ferner RE, Huson SM, Thomas N, Moss C, Willshaw H, Evans DG, et al. Guidelines for the diagnosis and management of individuals with neurofibromatosis 1. *J Med Genet* (2007) **44**:81–8. doi:10.1136/jmg.2006.045906
 30. Carey JC, Baty BJ, Johnson JP, Morrison T, Skolnick M, Kivlin J. The genetic aspects of neurofibromatosis. *Ann N Y Acad Sci* (1986) **486**:45–56. doi:10.1111/1/j.1749-6632.1986.tb48061.x
 31. Williams VC, Lucas J, Babcock MA, Gutmann DH, Korf B, Maria BL. Neurofibromatosis type 1 revisited. *Pediatrics* (2009) **123**:124–33. doi:10.1542/peds.2007-3204
 32. Bausch B, Borozdin W, Neumann HP, European-American Pheochromocytoma Study Group. Clinical and genetic characteristics of patients with neurofibromatosis type 1 and pheochromocytoma. *N Engl J Med* (2006) **354**:2729–31. doi:10.1056/NEJMc066006
 33. Boyd KP, Korf BR, Theos A. Neurofibromatosis type 1. *J Am Acad Dermatol* (2009) **61**:1–14. doi:10.1016/j.jaad.2008.12.051
 34. Johannessen CM, Reczek EE, James MF, Brems H, Legius E, Cichowski K. The NF1 tumor suppressor critically regulates TSC2 and mTOR. *Proc Natl Acad Sci U S A* (2005) **102**:8573–8. doi:10.1073/pnas.0503224102
 35. Baysal BE, Ferrell RE, Willett-Brozick JE, Lawrence EC, Myssiorek D, Bosch A, et al. Mutations in SDHD, a mitochondrial complex II gene, in hereditary paraganglioma. *Science* (2000) **287**:848–51. doi:10.1126/science.287.5454.848
 36. Burnichon N, Rohmer V, Amar L, Herman P, Leboulleux S, Darrouzet V, et al. The succinate dehydrogenase genetic testing in a large prospective series of patients with paragangliomas. *J Clin Endocrinol Metab* (2009) **94**:2817–27. doi:10.1210/jc.2008-2504
 37. Ricketts CJ, Forman JR, Rattenberry E, Bradshaw N, Lalloo F, Izatt L, et al. Tumor risks and genotype-phenotype-prototype analysis in 358 patients with germline mutations in SDHB and SDHD. *Hum Mutat* (2010) **31**:41–51. doi:10.1002/humu.21136
 38. Pigny P, Vincent A, Cardot Bateurs C, Bertrand M, de Montpreville VT, Crepin M, et al. Paraganglioma after maternal transmission of a succinate dehydrogenase gene mutation. *J Clin Endocrinol Metab* (2008) **93**:1609–15. doi:10.1210/jc.2007-1989
 39. Yamashita R, Usui T, Hashimoto S, Suzuki H, Takahashi M, Honkura K, et al. Predominant expression of mutated allele of the succinate dehydrogenase D (SDHD) gene in the SDHD-related paragangliomas. *Endocr J* (2009) **56**:1129–35. doi:10.1507/endocrj.K09E-111
 40. Neumann HPH, Pawlu C, Peczkowska M, Bausch B, McWhinney SR, Muresan M, et al. Distinct clinical features of paraganglioma syndromes associated with SDHB and SDHD gene mutations. *JAMA* (2004) **292**:943–51. doi:10.1001/jama.292.8.943
 41. Hao H-X, Khalimonchuk O, Schraders M, Dephoure N, Bayley J-P, Kunst H, et al. SDH5, a gene required for flavination of succinate dehydrogenase, is mutated in paraganglioma. *Science* (2009) **325**:1139–42. doi:10.1126/science.1175689
 42. Bayley J-P, Kunst HPM, Cascon A, Sampietro ML, Gaal J, Korpershoek E, et al. SDHAF2 mutations in familial and sporadic paraganglioma and pheochromocytoma. *Lancet Oncol* (2010) **11**:366–72. doi:10.1016/S1470-2045(10)70007-3
 43. Mannelli M, Ercolino T, Giach V, Simi C, Cirami C, Parenti G. Genetic screening for pheochromocytoma: should SDHC gene analysis be included? *J Med Genet* (2007) **44**:586–7. doi:10.1136/jmg.2007.051045
 44. Gimenez-Roqueplo A-P, Favier J, Rustin P, Rieubland C, Crespin M, Nau V, et al. Mutations in the SDHB gene are associated with extra-adrenal and/or malignant pheochromocytomas. *Cancer Res* (2003) **63**:5615–21.
 45. Stratakis CA, Carney JA. The triad of paragangliomas, gastric stromal tumours and pulmonary chondromas (Carney triad), and the dyad of paragangliomas and gastric stromal sarcomas (Carney-Stratakis syndrome): molecular genetics and clinical implications. *J Intern Med* (2009) **266**:43–52. doi:10.1111/j.1365-2796.2009.02110.x
 46. Carney JA. Carney triad: a syndrome featuring paraganglionic, adrenocortical, and possibly other endocrine tumors. *J Clin Endocrinol Metab* (2009) **94**:3656–62. doi:10.1210/jc.2009-1156
 47. Carney JA, Stratakis CA. Familial paraganglioma and gastric stromal sarcoma: a new syndrome distinct from the Carney triad. *Am J Med Genet* (2002) **108**:132–9. doi:10.1002/ajmg.10235
 48. Pasini B, McWhinney SR, Bei T, Matyakhina L, Stergiopoulos S, Muchow M, et al. Clinical and molecular genetics of patients with the Carney-Stratakis syndrome and germline mutations of the genes coding for the succinate dehydrogenase subunits SDHB, SDHC, and SDHD. *Eur J Hum Genet* (2008) **16**:79–88. doi:10.1038/sj.ejhg.5201904
 49. Carney JA. Gastric stromal sarcoma, pulmonary chondroma, and extra-adrenal paraganglioma (Carney triad): natural history, adrenocortical component, and possible familial occurrence. *Mayo Clin Proc* (1999) **74**:543–52. doi:10.4065/74.6.543
 50. Yeh I-T, Lenci RE, Qin Y, Buddavarapu K, Ligon AH, Leteurtre E, et al. A germline mutation of the KIF1B beta gene on 1p36 in a family with neural and nonneural tumors. *Hum Genet* (2008) **124**:279–85. doi:10.1007/s00439-008-0553-1
 51. Dahia PLM, Ross KN, Wright ME, Hayashida CY, Santagata S, Barontini M, et al. A HIF1alpha regulatory loop links hypoxia and mitochondrial signals in pheochromocytomas. *PLoS Genet* (2005) **1**:72–80. doi:10.1371/journal.pgen.0010008

52. Powers JF, Evinger MJ, Zhi J, Picard KL, Tischler AS. Pheochromocytomas in Nfl knockout mice express a neural progenitor gene expression profile. *Neuroscience* (2007) **147**:928–37. doi:10.1016/j.neuroscience.2007.05.008
53. Qin Y, Yao L, King EE, Buddavarapu K, Lenci RE, Chocron ES, et al. Germline mutations in TMEM127 confer susceptibility to pheochromocytoma. *Nat Genet* (2010) **42**:229–33. doi:10.1038/ng.533
54. Lee S, Nakamura E, Yang H, Wei W, Linggi MS, Sajan MP, et al. Neuronal apoptosis linked to EglN3 prolyl hydroxylase and familial pheochromocytoma genes: developmental culling and cancer. *Cancer Cell* (2005) **8**:155–67. doi:10.1016/j.ccr.2005.06.015
55. Schlisio S, Kenchappa RS, Vredevelde LCW, George RE, Stewart R, Greulich H, et al. The kinesin KIF1Bbeta acts downstream from EglN3 to induce apoptosis and is a potential 1p36 tumor suppressor. *Genes Dev* (2008) **22**:884–93. doi:10.1101/gad.1648608
56. Astuti D, Ricketts CJ, Chowdhury R, McDonough MA, Gentle D, Kirby G, et al. Mutation analysis of HIF prolyl hydroxylases (PHD/EGLN) in individuals with features of pheochromocytoma and renal cell carcinoma susceptibility. *Endocr Relat Cancer* (2011) **18**:73–83. doi:10.1677/ERC-10-0113
57. Yao L, Schiavi F, Cascon A, Qin Y, Inglada-Pérez L, King EE, et al. Spectrum and prevalence of FP/TMEM127 gene mutations in pheochromocytomas and paragangliomas. *JAMA* (2010) **304**:2611–9. doi:10.1001/jama.2010.1830
58. Abermil N, Guillaud-Bataille M, Burnichon N, Venisse A, Manivet P, Guignat L, et al. TMEM127 screening in a large cohort of patients with pheochromocytoma and/or paraganglioma. *J Clin Endocrinol Metab* (2012) **97**:E805–9. doi:10.1210/jc.2011-3360
59. Comino-Méndez I, Gracia-Aznárez FJ, Schiavi F, Landa I, Leandro-García LJ, Letón R, et al. Exome sequencing identifies MAX mutations as a cause of hereditary pheochromocytoma. *Nat Genet* (2011) **43**:663–7. doi:10.1038/ng.861
60. Blackwood EM, Lüscher B, Kretzner L, Eisenman RN. The Myc: max protein complex and cell growth regulation. *Cold Spring Harb Symp Quant Biol* (1991) **56**:109–17. doi:10.1101/SQB.1991.056.01.015
61. Jiménez C, Cote G, Arnold A, Gagel RF. Review: should patients with apparently sporadic pheochromocytomas or paragangliomas be screened for hereditary syndromes? *J Clin Endocrinol Metab* (2006) **91**:2851–8. doi:10.1210/jc.2005-2178
62. Neumann HPH, Bausch B, McWhinney SR, Bender BU, Gimm O, Franke G, et al. Germ-line mutations in nonsyndromic pheochromocytoma. *N Engl J Med* (2002) **346**:1459–66. doi:10.1056/NEJMoa020152
63. Cascón A, López-Jiménez E, Landa I, Leskelä S, Leandro-García LJ, Maliszewska A, et al. Rationalization of genetic testing in patients with apparently sporadic pheochromocytoma/paraganglioma. *Horm Metab Res* (2009) **41**:672–5. doi:10.1055/s-0029-1202814
64. Iacobone M, Schiavi F, Bottussi M, Taschin E, Bobisse S, Fassina A, et al. Is genetic screening indicated in apparently sporadic pheochromocytomas and paragangliomas? *Surgery* (2011) **150**:1194–201. doi:10.1016/j.surg.2011.09.024
65. Pacak K, Eisenhofer G, Ahlman H, Bornstein SR, Gimenez-Roqueplo A-P, Grossman AB, et al. Pheochromocytoma: recommendations for clinical practice from the first international symposium. *Nat Clin Pract Endocrinol Metab* (2007) **3**:92–102. doi:10.1038/ncpendmet0396
66. Brito JP, Asi N, Bancos I, Gionfriddo MR, Zeballos-Palacios CL, Leppin AL, et al. Testing for germline mutations in sporadic pheochromocytoma/paraganglioma: a systematic review. *Clin Endocrinol (Oxf)* (2015) **82**:338–45. doi:10.1111/cen.12530
67. Burnichon N, Vescovo L, Amar L, Libé R, de Reynies A, Venisse A, et al. Integrative genomic analysis reveals somatic mutations in pheochromocytoma and paraganglioma. *Hum Mol Genet* (2011) **20**:3974–85. doi:10.1093/hmg/ddr324
68. Burnichon N, Cascón A, Schiavi F, Morales NP, Comino-Méndez I, Abermil N, et al. MAX mutations cause hereditary and sporadic pheochromocytoma and paraganglioma. *Clin Cancer Res* (2012) **18**:2828–37. doi:10.1158/1078-0432.CCR-12-0160
69. Burnichon N, Buffet A, Parfait B, Letouzé E, Laurendeau I, Lorient C, et al. Somatic NF1 inactivation is a frequent event in sporadic pheochromocytoma. *Hum Mol Genet* (2012) **21**:5397–405. doi:10.1093/hmg/dds374
70. Martins R, Bugalho MJ. Paragangliomas/pheochromocytomas: clinically oriented genetic testing. *Int J Endocrinol* (2014) **2014**:794187. doi:10.1155/2014/794187
71. Eisenhofer G, Timmers HJ, Lenders JWM, Bornstein SR, Tiebel O, Mannelli M, et al. Age at diagnosis of pheochromocytoma differs according to catecholamine phenotype and tumor location. *J Clin Endocrinol Metab* (2011) **96**:375–84. doi:10.1210/jc.2010-1588
72. Pigny P, Cardot-Bauters C, Cao CD, Vantyghem MC, Carnaille B, Pattou F, et al. Should genetic testing be performed in each patient with sporadic pheochromocytoma at presentation? *Eur J Endocrinol* (2009) **160**:227–31. doi:10.1530/EJE-08-0574
73. Erlic Z, Rybicki L, Peczkowska M, Golcher H, Kann PH, Brauckhoff M, et al. Clinical predictors and algorithm for the genetic diagnosis of pheochromocytoma patients. *Clin Cancer Res* (2009) **15**:6378–85. doi:10.1158/1078-0432.CCR-09-1237
74. Shuch B, Ricketts CJ, Metwalli AR, Pacak K, Linehan WM. The genetic basis of pheochromocytoma and paraganglioma: implications for management. *Urology* (2014) **83**:1225–32. doi:10.1016/j.urology.2014.01.007
75. Walther MM, Reiter R, Keiser HR, Choyke PL, Venzon D, Hurley K, et al. Clinical and genetic characterization of pheochromocytoma in Von Hippel-Lindau families: comparison with sporadic pheochromocytoma gives insight into natural history of pheochromocytoma. *J Urol* (1999) **162**:659–64. doi:10.1097/00005392-199909010-00004
76. Boedeker CC, Erlic Z, Richard S, Kontny U, Gimenez-Roqueplo A-P, Cascon A, et al. Head and neck paragangliomas in Von Hippel-Lindau disease and multiple endocrine neoplasia type 2. *J Clin Endocrinol Metab* (2009) **94**:1938–44. doi:10.1210/jc.2009-0354
77. Schiavi F, Boedeker CC, Bausch B, Peczkowska M, Gomez CF, Strassburg T, et al. Predictors and prevalence of paraganglioma syndrome associated with mutations of the SDHC gene. *JAMA* (2005) **294**:2057–63. doi:10.1001/jama.294.16.2057

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Costa, Ortiga-Carvalho, Violante and Vaisman. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.