Check for updates

OPEN ACCESS

EDITED BY Gang Ye, Sichuan Agricultural University, China

REVIEWED BY

Xingxing Yuan, Heilongjiang Academy of Traditional Chinese Medicine, China Hao Chi, Macau University of Science and Technology, Macao SAR, China

*CORRESPONDENCE Ming Pei tjtcmpeiming@126.com

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 06 November 2023 ACCEPTED 14 December 2023 PUBLISHED 11 January 2024

CITATION

Wang L, Xu A, Wang J, Fan G, Liu R, Wei L and Pei M (2024) The effect and mechanism of Fushen Granule on gut microbiome in the prevention and treatment of chronic renal failure. *Front. Cell. Infect. Microbiol.* 13:1334213. doi: 10.3389/fcimb.2023.1334213

COPYRIGHT

© 2024 Wang, Xu, Wang, Fan, Liu, Wei and Pei. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The effect and mechanism of Fushen Granule on gut microbiome in the prevention and treatment of chronic renal failure

Lin Wang^{1,2†}, Ao Xu^{1,2†}, Jinxiang Wang^{3†}, Guorong Fan^{1,2}, Ruiqi Liu⁴, Lijuan Wei^{1,2} and Ming Pei^{1,2*}

¹Nephrology Department, First Teaching Hospital of Tianjin University of Traditional Chinese Medicine, Tianjin, China, ²Graduate School, Tianjin University of Traditional Chinese Medicine, Tianjin, China, ³Guangdong Provincial Key Laboratory of Digestive Cancer Research, Precision Medicine Center, Scientific Research Center, The Seventh Affiliated Hospital, Sun Yat-Sen University, Shenzhen, China, ⁴Nephrology Department, Tianjin Academy of Traditional Chinese Medicine Affiliated Hospital, Tianjin, China

Background: Fushen Granule is an improved granule based on the classic formula Fushen Formula, which is used for the treatment of peritoneal dialysis-related intestinal dysfunction in patients with end-stage renal disease. However, the effect and mechanism of this granule on the prevention and treatment of chronic renal failure have not been fully elucidated.

Methods: A 5/6 nephrectomy model of CRF was induced and Fushen Granule was administered at low and high doses to observe its effects on renal function, D-lactate, serum endotoxin, and intestinal-derived metabolic toxins. The 16SrRNA sequencing method was used to analyze the abundance and structure of the intestinal flora of CRF rats. A FMT assay was also used to evaluate the effects of transplantation of Fushen Granule fecal bacteria on renal-related functional parameters and metabolic toxins in CRF rats.

Results: Gavage administration of Fushen Granule at low and high doses downregulated creatinine, urea nitrogen, 24-h urine microalbumin, D-lactate, endotoxin, and the intestinal-derived toxins indophenol sulphateand p-cresol sulphate in CRF rats. Compared with the sham-operated group in the same period, CRF rats had a decreased abundance of the firmicutes phylum and an increased abundance of the bacteroidetes phylum at the phylum level, and a decreasing trend of the lactobacillus genus at the genus level. Fushen Granule intervention increased the abundance of the firmicutes phylum, decreased the abundance of the bacteroidetes phylum, and increased the abundance of the lactobacillus genus. The transplantation of Fushen Granule fecal bacteria significantly reduced creatinine(Cr), blood urea nitrogen(Bun), uric acid(UA), 24-h urinary microalbumin, D-lactate, serum endotoxin, and enterogenic metabolic toxins in CRF rats. Compared with the sham-operated group, the transplantation of Fushen Granule fecal bacteria modulated the Firmicutes and Bacteroidetes phyla and the Lactobacillus genus. **Conclusion:** Fushen Granule improved renal function and intestinal barrier function by regulating intestinal flora, inhibiting renal fibrosis, and delaying the progression of chronic renal failure.

KEYWORDS

chronic renal failure, intestinal microecology, fecal microbiota transplantation, renal-intestinal axis, Fushen Granule



1 Introduction

Chronic renal failure (CRF) is a progressive disease resulting from various chronic kidney diseases (CKD), leading to abnormalities in kidney tissue structure and function until eventual failure (Fang et al., 2023). Its development into the final stage, end-stage renal disease (ESRD), poses a significant threat to human health and consumes considerable health resources(Wouters et al., 2015) (Dai et al., 2021; Wouk, 2021). Statistics show that CKD affects approximately 15-20% of the global population (Matsushita et al., 2022), with the number of cases increasing by about 20 million per year. The prevalence of CRF stands at 15.1% among adults in the US and 10.8% in China (Wang et al., 2021). As a late complication of CKD (Webster et al., 2017), ESRD is a major disease with high morbidity and mortality rates worldwide (Zhang et al., 2012; Liyanage et al., 2015). The cost of treating ESRD is currently staggering; in the US alone, it is estimated to be around US\$34 billion per year (Saran et al., 2018). The burden of CRF on healthcare systems and health insurance costs has become a tremendous challenge for public health systems worldwide. Therefore, determining how to delay the disease process and improve patients' quality of life during the chronic kidney failure phase is a crucial issue in preventing and treating chronic kidney disease and end-stage renal disease.

The treatment of CRF still has no specific method in the international medical community, mainly to control protein intake, anti-oxidative stress, blood pressure, blood glucose and lipid control and other risk factors that may lead to the progression of the disease to slow down the deterioration (Ruiz-Ortega et al., 2020; Faria and de Pinho, 2021); Although there has been great progress in the study of molecular mechanisms and related markers, the clinical translation is poor. In recent years, more and more studies have demonstrated that the development of CRF is closely related to the imbalance of intestinal microecology, and that disorders of intestinal flora and disruption of intestinal barrier structure and function play an important role in disease progression (Yang et al., 2018; Pluznick, 2020). Intestinal dysfunction can be seen in the early stages of CRF, mainly in the form of digestive symptoms such as impaired digestion and absorption in the intestine, as well as varying degrees of impairment of intestinal barrier function. The presence of the intestinal barrier prevents bacteria, pathogens and other harmful substances in the intestinal lumen from entering the bloodstream to maintain the stability of the internal environment. After the intestinal barrier is damaged, endotoxins and intestinal flora can enter the blood circulation, resulting in enterogenic endotoxemia, which aggravates the microinflammatory state of the body. CRF and peritoneal dialysis patients are

Abbreviations: BUN, Blood urea nitrogen; CKD, Chronic kidney diseases; CR, Creatinine; CRF, Chronic renal failure; ESRD, End-stage renal disease; FMT, Fecal microbiota transplantation; IS, Indophenol sulphate; NMDS, Non-metric Multidimensional Scaling Analysis; PCA, Principal Component Analysis; PCOA, Principal Coordinate Analysis; PCS, P-cresol sulphate; SPF, Specific pathogenfree; TCM, Traditional Chinese medicine; UA, Uric acid.

combined with varying degrees of endotoxemia (Gonçalves et al., 2006; Szeto et al., 2008; Feroze et al., 2012), and uremic rats are more prone to lateral translocation of intestinal bacteria (de Almeida Duarte et al., 2004), this is a reflection of the impaired intestinal barrier in CRF. In addition, the accumulation of intestinal proteins in CRF alters the structure of the intestinal flora and increases the number of enteric proteolytic bacteria. Tryptophan is fermented in the intestine to produce indole, which is absorbed into the blood via the intestine and converted to IS in the liver; tyrosine and phenylalanine are fermented in the intestine to produce p-cresol, which is converted to PCS in the intestinal epithelium (Liabeuf et al., 2011). The latter is converted to PCS in the intestinal epithelium. Since IS and PCS are bound to albumin in the blood and cannot be excreted through dialysis, their accumulation in the body can accelerate the progression of renal failure by aggravating glomerulosclerosis and interstitial fibrosis (Miyazaki et al., 1997; Gelasco and Raymond, 2006; Watanabe et al., 2013). Therefore, repairing the intestinal barrier and reversing the imbalance of intestinal flora are considered as potential therapeutic targets and new targets for intervention in the treatment of CRF.

Traditional Chinese medicine (TCM) holds a unique advantage in the treatment of CRF. CRF involves a complex interplay among various organs, with a particularly intricate relationship between the spleen and kidney systems in TCM. The fundamental mechanism underlying CRF is often described as "deficiency of the spleen and kidney, accumulation of turbid toxins." Guided by this medical principle, the Department of Nephrology at the First Affiliated Hospital of Tianjin University of Traditional Chinese Medicine has tailored its approach to the pathogenic patterns of CRF. Draw upon extensive clinical practice and years of specialized case data, and referencing the optimized compilation and efficacy validation of nationwide TCM protocols for CRF, this has ultimately evolved into the "Fushen Formula." This formula serves as the basis for a corresponding composite herbal preparation named "Fushen Granule." The granule preparation has received hospital formulation approval (Batch Number: 140928) and is primarily employed to treat intestinal dysfunctions related to peritoneal dialysis in ESRD patients. It has been shown to achieve definite therapeutic effects in clinical practice. In clinical application, the treatment group receiving Fushen Granule demonstrated superior improvement in renal function and reduction in blood creatinine (BCR) and blood urea nitrogen(BUN) levels compared to the control group, with no adverse reactions observed (Sun and Liu, 2010). Among elderly CKD stage 4 patients, oral administration of Fushen Granule significantly enhanced kidney function and nutritional status, ameliorated symptoms, and elevated quality of life (Yang and Yang, 2015). Another study revealed that the combination of blood purification and oral administration of Fushen Granule for treating CRF significantly improved kidney function and quality of life, reduced hospitalization time, lowered mortality rates, and alleviated economic burdens (Lei, 2016). Furthermore, the comprehensive efficacy of oral administration of Fushen Granule combined with acupuncture at specific points was found to surpass that of conventional acupuncture alone in CKD stage 4 patients (Zhang and Yang, 2016). Building upon clinical practice, fundamental research on Fushen Granule has also been initiated.

Results from basic experiments demonstrate that Fushen Granule can slow down the progression of renal function impairment in a rat model of CRF. It reduces the expression of mild inflammatory factors (such as hs-CRP, IL-6, and TNF- α), lowers blood urea nitrogen (BUN) and blood creatinine(BCR) levels, thereby improving kidney function and delaying the progression of renal deterioration (Zhang and Yang, 2018a; Zhang and Yang, 2018b). Fushen Granules are not only suitable for patients with CRF, but also for conditions such as peritoneal dialysis in CRF and renal interstitial fibrosis (Zhao et al., 2022). A multicenter clinical study involving 240 peritoneal dialysis patients over a 6-month period demonstrated significant enhancements attributed to Fushen Granules. These enhancements include improved TCM syndromes, enhanced dialysis efficiency, increased removal of toxins and excess water, slowed decline of residual kidney function, and an overall improvement in quality of life (Yang et al., 2012a; Yang et al., 2013). Additionally, Fushen Granules exhibited positive effects on the prognosis of peritonitis (Jiang et al., 2020a), gastrointestinal function in peritoneal dialysis patients (Chen, 2016), and maintenance of protein nutritional status (Yang et al., 2014; Lei and Yang, 2018; Lei et al., 2018a). In peritoneal dialysis rats, research reveals that Fushen Granules improve renal function damage through multiple pathways, including modulating Glo-1 expression levels and reducing serum AGEs accumulation, thereby safeguarding residual kidney function(Tang et al., 2017; Tang et al., 2018b). Furthermore, it alleviates ultrafiltration volume and glucose transport, enhancing peritoneal dialysis efficacy(Yang et al., 2021b). Fundamental research also indicates that Fushen Granules regulate Dcn, inhibit activation of the TGF-B pathway, disrupt extracellular matrix accumulation, modulate glucose and lipid metabolism, suppress expression of fibrosis-promoting factors (such as TGF-B1, TGF-BRII, CTGF, IL-6, CTGF, VEGF), and elevate expression of anti-fibrotic factors (HGF, BMP-7), thereby restraining peritoneal collagen formation and inhibiting mesothelial cell transdifferentiation. As a result, it slows the progression of peritoneal fibrosis (Yang et al., 2012b; Dou, 2015; Lei et al., 2018b; Lei et al., 2018c; Jiang et al., 2020b; Li et al., 2020; Yang et al., 2021a). Additionally, Fushen Granules reduce the accumulation of prooxidative factors like MG and MDA, upregulate the expression of the antioxidant GSH, subsequently ameliorating anemic conditions in CRF peritoneal dialysis rats (Tang et al., 2018a).

Fecal microbiota transplantation (FMT) has recently gained renewed clinical interest as a long-established therapy for rebuilding the intestinal flora. FMT, i.e. the isolation of functional donor or autologous intestinal flora and transplantation into the patient's gut, has been applied to treat a range of gastrointestinal/nongastrointestinal disorders associated with intestinal flora and is a breakthrough medical advance in recent years (Cui et al., 2016). In addition, FMT is also used in the renal field for kidney transplantation (Ahmad and Bromberg, 2016). Although the correlation between intestinal flora and CKD/CRF is currently a hot topic of research, the use of FMT in the treatment of CKD/CRF has not yet been reported. Therefore, within the scope of affirming Fushen Granules' ability to retard peritoneal fibrosis progression, enhance dialysis efficacy, and improve quality of life for peritoneal dialysis patients—particularly in ameliorating intestinal functional disorders—we have integrated Fushen Granules with FMT. This integration enables a more profound and direct impact of postintervention microbiota reinfusion on intestinal microbiota dynamics. From this perspective, the pivotal role and significance of gut microbiota in CRF are harnessed, establishing novel avenues and intervention targets for preventing and treating CRF progression.

2 Materials and methods

2.1 Preparation of the experimental drug, Fushen Granule

Fushen Granules (Batch number: 140928) were provided by the First Teaching Hospital of Tianjin University of Traditional Chinese Medicine (Tianjin, China) in strict compliance with the Good Manufacturing Practice and Good Laboratory Practice guidelines for Pharmaceutical Manufacturers. Fushen Granules are primarily composed of eight different herbal ingredients. The names of these herbs have been revision in the plant list (http:// www.theplantlist.org), and the details can be found in Table 1. Through UPLC/Q-TOF analysis, the main active components of the Fushen Granules were characterized. We have preliminarily identified 55 different compounds in FSG, as detailed in the attached document. The granules were dissolved in distilled water, and the dosage for rats was adjusted based on the body surface area of humans, with equal amounts administered as low doses and 10 times the amount given as high doses.

2.2 Laboratory animals

Specific pathogen-free (SPF) healthy male Sprague-Dawley (SD) rats (weight 180 \pm 20g) were purchased from the Experimental Animal Centre of the Chinese People's Liberation Army Academy of Military Medical Sciences. They were housed in the Experimental Animal Centre of Tianjin University of Chinese Medicine. Throughout the experiment, the rats were fed normal

chow (no yeast, no probiotics) and had ad libitum access to water. All animal procedures were conducted in accordance with the NIH Guide for the Care and Use of Laboratory Animals and the Chinese Laboratory Animal Management Methods. The ethical approval for this study was No. IRM-DWLL-2020100.

2.3 Experimental reagents and apparatus

The following reagents and apparatus were used in the study: Rat Indoxyl Sulfate (IS) ELISA Kit (Jianglai Bio, JL44174), Rat P-Cresol Sulfate (PCS) ELISA Kit (Jianglai Bio, JL48836), Lactate Test Kit (Nanjing Jiancheng, A019-2-1), Endotoxin Assay Kit (Xiamen Horseshoe Crab Bio, EC80545), Urea Nitrogen Assay Kit (Changchun Huili, C010), Creatinine Assay Kit (Changchun Huili, C074), Uric Acid Assay Kit (Changchun Huili, C075), and DNA Isolation Kit (MoBio, Carlsbad, CA, USA). An Illumina HiSeq platform was provided by Novogene Bioinformatics Co. Ltd., Beijing, China.

Equipment used in the study included a vertical refrigerated display cabinet (Star), a horizontal freezer (Meiling), an electronic balance (Mettler-Toledo Instruments (Shanghai) Co., Ltd.), a highspeed tissue grinder (Servicebio), a benchtop high-speed frozen centrifuge (ThermoFisher), and an enzyme-linked immunosorbent assay (BioTek).

3 Methods

3.1 Chronic renal failure rat model construction and sample collection

In this study, a 5/6 nephrectomy was performed to establish a chronic renal failure(CRF) model in rats. The left kidney was initially resected by 5/6 nephrectomy, and after a 1-week recovery period, the right kidney was completely resected. Blood and urine biochemical tests were conducted at the end of the modeling period. Fresh feces from each group were collected in sterile lyophilized tubes, immediately placed in liquid nitrogen, and then stored at -80°C.

Num	Scientific name	Material	Latin name	Chinese name	Mass Ratio
1	Astragalus membranaceus	Root	Hedysarum Multijugum Maxim	Huangqi	15g
2	Angelica sinensis	Root	Angelica sinensis (Oliv.) Diels	Danggui	10g
3	Epimedium brevicornu Maxim	Root	Epimedium brevicornu Maxim	Xianlingpi	15g
4	Citrus reticulata Blanco	outer pericarp	Citrus reticulata Blanco	Chenpi	10g
5	Pinellia ternata	Roots and rootstalk	Pinellia ternata (Thunb.) Makino	Banxia	15g
6	Salvia miltiorrhiza Bunge	Roots and rootstalk	Salvia miltiorrhiza Bunge	Danshen	30g
7	Rheum	Roots and rootstalk	Rheum palmatum L.	Dahuang	10g
8	Siebold	Winged shoots or branch wings	Euonymus alatus (Thunb.) Siebold	Guijianyu	30g

TABLE 1 The constituents of Fushen Granule.

10.3389/fcimb.2023.1334213

3.2 Observation on the intervention effect of Fushen Granule on intestinal flora in CRF

For in vivo studies, 150 specific pathogen-free (SPF) healthy male Sprague-Dawley (SD) rats were acclimatized for 1 week. The rats were randomly divided into the following groups: 30 rats in the sham-operated group (Group A), 30 rats in the sham-operated Chinese medicine group (Group B), and 90 rats in the modeling group. The modeling group was further divided into a low-dose Chinese medicine group (Group C), a high-dose Chinese medicine group (Group D), and a control model group (Group E), with 30 animals in each group. Groups B and C received a low dose of Fushen Granule via gavage; Group D received 10 times the high dose of Fushen Granule via gavage; Groups A and E were given 2ml·d-1 distilled water via gavage. Six animals from each group were euthanized for examination at the beginning of drug intervention, at the end of the second week, and at the end of the fourth week. All remaining animals were euthanized at the end of the sixth week.

3.3 Fecal microbiota transplant experiment

After 1 week of acclimatization, 55 specific pathogen-free (SPF) healthy male Sprague-Dawley (SD) rats were randomly divided into a 5/6 nephrectomized kidney failure rat model recipient group (15 x 3) and a donor rat group (5 x 2, approximately 1/3 the number of the recipient group). The donor group was further divided into the Fushen Granule donor group and the normal donor group, while the recipient group was divided into the model group + Fushen Granule donor FMT group, the model group + normal donor FMT group, and the model group + saline sham operation group. In the Fushen Granule donor group, a low dose of Fushen Granule was administered for 6 weeks from the first day of right nephrectomy, based on the effect of the previous experimental intervention. No intervention was made in the normal donor group. Fresh, uncontaminated feces were collected daily, and intestinal microbial extracts were prepared in saline. Fecal microbiota transplantation(FMT) was completed within 4 hours of feces collection. The recipient groups, model group + Fushen Granule donor and model group + normal donor, underwent FMT with intestinal microbial extracts from the Fushen Granule donor group and the normal donor group, after determining successful modeling through dynamic observation of kidney function. The model group + saline sham group underwent a procedure with an equal amount of saline equivalent as the control group.

3.4 Observation indicators

The animals were dynamically observed, and their body weight was assessed every 5 days. The focus was on the general status of the rats at the beginning of drug intervention and at the end of the 2nd, 4th, and 6th weeks (general status in terms of behavioral activity, body weight, phenotypic signs, and survival rate), renal function indicators (Cr, BUN, UA, eGFR, and urinary microalbumin), serum endotoxin, D-lactate, metabolic toxins of intestinal origin (IS, PCS), and analysis of intestinal bacterial diversity.

3.5 Analysis of intestinal bacterial diversity

In this study, 16S rRNA was used to assess intestinal bacterial diversity, but the timing of fecal sample collection differed between the two experiments. Fresh fecal samples were collected at around 10 am at the beginning of the intervention and at the end of the second, fourth, and sixth weeks. Fecal samples were collected from the three model groups at two time points: before the start of the FMT experiment and 15 days after the transplantation (Note: The feces were collected for the same amount of time each instance). Fresh fecal samples collected were immediately frozen in liquid nitrogen for 5 minutes and then stored at -80°C. DNA was extracted from feces using the Power Fecal DNA Isolation Kit (MoBio, Carlsbad, CA, USA). DNA was recovered using 30 mL of the buffer included in the kit. Sequencing principles were used for synthetic sequencing with the Illumina HiSeq platform (Novogene Bioinformatics Co., Beijing, China). Taxonomic composition of the flora was analyzed to evaluate bacterial abundance and compositional diversity, using phylum, order, family, and genus classifications.

3.6 Statistical analysis

SPSS 19.0 for Windows statistical software was used for data analysis. Data were expressed as mean \pm SEM relative to the number of samples in each group (n). Analysis of variance (ANOVA), Wilcoxon rank-sum test, Tukey's t-test, and Student's t-test were used to determine statistical significance between multiple treatment groups. The Kaplan-Meier survival test was used to analyze survival rates. Results with p< 0.05 were considered statistically significant.

4 Results

4.1 Effect of Fushen Granule on the status of rats with CRF

4.1.1 General state

During the modeling phase, rats in the model group typically exhibited irritability, emotional instability, and agitated reactions when injected or given the drug via gavage. Over time, rats in the sham-operated group gained weight, appeared in good spirits, moved freely, were responsive, and had well-groomed, moisturized fur. In contrast, rats in the model group gradually lost weight, ate less than the sham-operated group, appeared more depressed, squinted, were less active, less responsive, and had disheveled and unkempt fur. Rats in the drug intervention group demonstrated better performance in terms of mental status, body weight, activity, responsiveness, and fur appearance after treatment, but still showed slight differences compared to the shamoperated group.

4.1.2 Observation of renal function indicators

Compared with the sham-operated group, the model group showed a statistically significant increase in Cr, BUN, and 24-h urinary microalbumin at weeks 2, 4, and 6 (P< 0.01), indicating successful modeling of renal failure (see Table 2). UA also tended to increase, but the difference was not statistically significant. Compared with the model group, the 24-h urine microalbumin in the Chinese medicine low-dose group decreased significantly at weeks 2 and 4 (P< 0.001), BUN decreased significantly at week 4 (P< 0.05), and Cr decreased significantly at week 6 (P< 0.05). The Chinese medicine high-dose group showed significant decreases in Cr and 24-h urine microalbumin at weeks 2, 4, and 6 (P< 0.01), and BUN at week 4 (P< 0.05) (Table 2). In conclusion, the herbal intervention group had a positive effect on the renal function indices of rats with CRF, and the efficacy of the high-dose herbal group was better.

4.1.3 Observation of serum endotoxins and metabolic toxin-like indicators of enteric origin

Compared with the sham-operated group, rats in the model group showed significantly higher IS, PCS, D-lactic acid, and endotoxin levels at weeks 2, 4, and 6 (P< 0.05), indicating successful modeling of renal failure. Compared with the model group, the Chinese medicine low-dose group showed significant decreases in IS, PCS, D-lactate, and endotoxin at weeks 2, 4, and 6 (P< 0.05). The high-dose group of Chinese medicine showed a significant decrease in PCS (P< 0.01) at the beginning of the drug intervention and in IS, PCS, D-lactic acid, and endotoxin at weeks 2, 4, and 6 (P< 0.05) (Table 3). This indicates that both the low and high dose groups of Chinese medicine demonstrated improvement in D-lactate, endotoxin, and enteric-derived metabolic toxin-like indicators. Compared with the low-dose group, the high-dose group generally had a better effect on D-lactate, endotoxin, and intestinal-derived metabolic toxins.

4.1.4 Analysis of intestinal bacterial diversity 4.1.4.1 Sequencing data quality and OTU analysis

The IonS5TMXL sequencing platform was utilized to analyze the diversity of intestinal bacteria. Using a single-end sequencing

TABLE 2	Indicators	of ki	dney	function.
---------	------------	-------	------	-----------

Group	Point in time	Cr (mmol/L)	BUN (µmol/l)	UA (µmol/l)	24-h urine microalbumin (mg/L)
	Initial	48.25 ± 4.28	18.29 ± 2.15	108.35 ± 4.79	23.28 ± 4.82
Group in Sham-operated Group A 2 Sham-operated Group A 2 Fushen Granule low dose control (Group B) 2 Model (Group E) 2 Fushen Granule at low doses (Group C) 2 Fushen Granule at high doses (Group D) 2	2 weeks	42.45 ± 3.35	14.28 ± 2.23	104.22 ± 16.38	26.75 ± 3.24
	4 weeks	65.59 ± 10.07	16.58 ± 0.61	107.53 ± 20.37	29.36 ± 4.91
	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	25.18 ± 3.52			
Fushen Granule low dose control (Group B)	Initial	50.23 ± 5.97	20.34 ± 2.46	110.25 ± 5.32	25.17 ± 3.34
	2 weeks	55.9 ± 5.23	13.98 ± 2.80	121.00 ± 20.57	25.37 ± 5.82
	4 weeks	65.72 ± 9.82**	16.04 ± 1.16	109.29 ± 14.96	27.54 ± 3.26
	6 weeks	72.34 ± 8.14	38.47 ± 0.67	139.85 ± 40.34	24.34 ± 2.84
Model (Group E)	Initial	47.93 ± 4.74	19.23 ± 2.73	115.32 ± 5.28	27.43 ± 3.44
	2 weeks	109.23 ± 4.56***	50.24 ± 4.56***	110.25 ± 11.39	128.37 ± 12.37***
	4 weeks	103.75 ± 2.57**	45.47 ± 0.95**	104.42 ± 16.04	379.85 ± 28.37***
	6 weeks	107.79 ± 12.37**	44.33 ± 2.1**	127.40 ± 18.15	583.92 ± 45.29***
	Initial	51.2 ± 5.22	20.12 ± 1.94	109.73 ± 4.91	24.82 ± 4.19
Fushen Granule at low doses	2 weeks	106.78 ± 1.25	45.91 ± 3.12	108.13 ± 26.28	88.24 ± 6.74###
(Group C)	4 weeks	96.46 ± 3.67**	25.48 ± 2.63#	107.68 ± 24.15	128.49 ± 18.58###
	6 weeks	95.41 ± 3.91**#	$44.63 \pm 1.45^{**}$	136 ± 34.74	73.95 ± 10.28###
	Initial	49.37 ± 4.60	18.32 ± 2.31	111.20 ± 4.83	26.37 ± 2.58
Fushan Cranula at high dagaa	2 weeks	95.24 ± 3.13##	44.15 ± 4.50	103.67 ± 30.12	75.63 ± 6.53###
(Group D)	4 weeks	85.1 ± 9.7**##	$41.69 \pm 0.9^{**}$ #	120.97 ± 24.09	102.53 ± 13.45###
	6 weeks	90.41 + 9.77**##	44.03 ± 2.24**	119.87 ± 15.49	58.24 ± 8.94###

Compared to the sham-operated group: * indicates p< 0.05, ** indicates p< 0.01, *** indicates p< 0.001; compared to the model group, # indicates p< 0.05, ## indicates p< 0.01, ### p< 0.001.

Group	Point in time	Metabolic toxins of enteric origin		Serum endotoxin		
· ·	AroupPoint in timeImage: Image: Image	IS (µg/ml)	PCS (µg/ml)	D-Lactic acid (µg/ml)	Endotoxin (EU/ml)	
	Initial	1.02 ± 0.12	0.03 ± 0.01	7.68 ± 0.27	0.019 ± 0.003	
	2 weeks	1.35 ± 0.34	0.05 ± 0.01	7.59 ± 0.31	0.029 ± 0.004	
Sham-operated (Group A)	4 weeks	1.66 ± 1.12	0.08 ± 0.01	7.51 ± 0.21	0.027 ± 0.002	
	6 weeks	1.71 ± 0.47	0.13 ± 0.07	7.66 ± 0.30	0.032 ± 0.004	
	Initial	0.85 ± 0.23	0.02 ± 0.01	7.52 ± 0.31	0.021 ± 0.002	
Fushen Granule low dose control	2 weeks	0.90 ± 0.11	0.04 ± 0.01	7.45 ± 0.36	0.025 ± 0.003	
(Group B)	4 weeks	0.96 ± 0.47	0.07 ± 0.01	7.22 ± 0.25	0.026 ± 0.006	
	6 weeks	3.04 ± 0.6	0.06 ± 0.02	7.82 ± 0.48	0.021 ± 0.001	
	Initial	1.05 ± 0.19	$0.05 \pm 0.01^*$	7.61 ± 0.27	0.023 ± 0.003	
Madd (Course F)	2 weeks	3.67 ± 0.65***	0.43 ± 0.11***	15.92 ± 0.28***	0.316 ± 0.027***	
Model (Group E)	4 weeks	6.02 ± 1.58**	0.58 ± 0.38**	20.31 ± 0.48***	0.429 ± 0.031***	
	6 weeks	6.35 ± 2.24**	$1.74 \pm 0.34^{*}$	14.76 ± 0.34***	0.496 ± 0.038***	
	Initial	0.98 ± 0.10	0.04 ± 0.01	7.76 ± 0.24	0.026 ± 0.003	
	2 weeks	2.59 ± 0.35##	0.20 ± 0.04###	10.22 ± 0.47###	0.209 ± 0.023###	
Fushen Granule at low doses (Group C)	4 weeks	3.89 ± 0.78**##	0.26 ± 0.1	9.03 ± 1.02###	0.297 ± 0.027###	
	6 weeks	3.54 ± 1.97#	0.3 ± 0.23#	11.76 ± 0.13###	0.234 ± 0.025###	
	Initial	0.95 ± 0.15	0.02 ± 0.01##	7.53 ± 0.25	0.025 ± 0.001	
Fusher Crearle et high desse (Creare D)	2 weeks	2.48 ± 0.32##	0.13 ± 0.02###	9.84 ± 0.52###	0.174 ± 0.018###	
rushen Granule at nign doses (Group D)	4 weeks	3.55 ± 0.79*##	0.15 ± 0.04#	10.79 ± 0.63###	0.216 ± 0.023###	
	6 weeks	3.19 ± 1.56#	0.39 ± 0.24#	12.01 ± 0.44###	0.183 ± 0.017###	

TABLE 3 Indicators for D-lactate, endotoxin and enteric-derived metabolic toxin categories.

Compared to the sham-operated group: * indicates p< 0.05, ** indicates p< 0.01, *** indicates p< 0.001; compared to the model group, # indicates p< 0.05, ## indicates p< 0.01, ### p< 0.001.

method, an average of 83,925 reads per sample were measured by shear filtering of Reads, and an average of 78,965 valid data were obtained after quality control, with a quality control efficiency of 94.15%. The sequences were clustered into OTUs (Operational Taxonomic Units) with 97% agreement, yielding a total of 4,955 OTUs. The OTUs sequences were then analyzed against the Silva132 database for species annotation to understand the composition and differences between intestinal flora. A total of 4,955 OTUs were counted according to different taxonomic levels, of which the number of OTUs that could be annotated to the database was 4,954 (99.98%). Dilution curves were constructed by the number of OTUs detected at each sequencing depth of the experimental data, and it was found that as the sequencing depth gradually increased, the sparsity curve also gradually smoothed out, indicating that the current sequencing depth is essentially sufficient to reflect the microbial diversity contained in this community sample (Figures 1A, B). The smoothness of the Rank Abundance curve reflects the uniformity of the species distribution, and from the figure (Figure 1C), it can be observed that the distribution of microbial species contained in this sequenced sample was poorly distributed. Species accumulation box plots were used for sample

size and diversity analysis of the biota. As seen in the box line plot (Figure 1D), the box positions no longer rose sharply but leveled off as the sample size expanded, indicating that the current sample size was abundant and adequate, and largely representative of the community's flora characteristics.

By clustering, sequences can be grouped into various categories based on their similarity to each other, with each category representing a taxonomic OTU. OTUs with a 97% similarity level are typically analyzed for bioinformatic statistics. In this study, we analyzed intestinal bacterial diversity in 5/6 nephrectomized rats. The results showed that the number of OTUs shared between the model and sham-operated groups was highest at week 0 (1465). However, the number of OTUs shared by the five groups gradually decreased at weeks 2 and 4, with 1197 and 541, respectively. This indicates that 5/6 nephrectomy can cause a decrease in the number of OTUs in the group. In terms of unique OTUs, the model group had 247, 186, and 66 unique OTUs at weeks 2, 4, and 6, respectively. The Chinese medicine high-dose treatment group had 393, 357, and 821 unique OTUs, while the low-dose treatment group had 87, 177, and 55 unique OTUs in that order. These findings suggest that Fushen Granule may alleviate intestinal flora damage and increase



Analysis of intestinal bacterial diversity. (A, B) Sparsity curve of the sample to be tested; (C) Rank Abundance curve; (D) Cumulative box plot of species.

the number of OTUs in rats with 5/6 nephrectomy to some extent. Compared with the same time points, the herbal high-dose treatment groups at weeks 2, 4, and 6 contained the highest number of unique OTUs. This indicates that the Chinese medicine high-dose group was more effective in restoring intestinal flora diversity in 5/6 nephrectomized rats than the lowdose treatment group (Figure 2).

4.1.4.2 Species annotation and taxonomic analysis

Species abundance tables at different taxonomic levels were generated using QIIME software, and then mapped into community structure at each taxonomic level of the samples using R language tools. The results showed that the dominant species at the phylum level were Firmicutes, Bacteroidetes, and Proteobacteria; at the genus level, the dominant species were Lactobacillus, Enterococcus, and Stenotrophomonas. The phylum Firmicutes and the phylum Bacteroidetes are the most abundant bacterial phyla in the intestinal flora, accounting for more than 90% of all bacteria. An analysis of the relative abundance of the intestinal flora at the phylum and genus taxonomic levels is shown in Figure 3A, B. In the sham-operated group, the levels of the two bacteria, Phylum Firmicutes and Phylum Bacteroidetes, were

relatively stable and did not change over time. The relative abundance of Phylum Firmicutes was 70%, 69%, 68%, and 63%, and the relative abundance of Phylum Bacteroidetes bacteria was 24%, 24%, 22%, and 30% at the four time points of 0, 2, 4, and 6 weeks, respectively. The ratio of Phylum Firmicutes to Phylum Bacteroidetes bacteria remained roughly between 2 and 3. In contrast, in the 5/6 nephrectomy model group, the relative abundance of the Firmicutes phylum decreased, and the relative abundance of the Bacteroidetes increased, resulting in a decrease in the Firmicutes phylum/Bacteroidetes phylum ratio. Compared to the sham-operated group during the same period, the relative abundance of Firmicutes phylum bacteria in the model group decreased from 81% at week 0 to 54% at week 6, while the relative abundance of Bacteroidetes phylum increased from 10% at week 0 to nearly 40% at week 6, with a decreasing trend in the ratio of the two contents (Table 4). Compared with the model group, the Chinese medicine low-dose and Chinese medicine highdose groups could differently reduce this alteration to some extent and restore the Firmicutes phylum and Bacteroidetes phylum composition ratio. For example, at the second week, the Firmicutes phylum and Bacteroidetes phylum ratio was 2.88 in the sham-operated group, and the ratio was 1.1, 2.27, and 1.88 in the



model, low-dose, and high-dose Chinese medicine groups, respectively (Table 5). In terms of genus level, Lactobacillus was the main dominant genus, and compared with the sham-operated group during the same period, the level of Lactobacillus in the model group showed a decreasing trend, at 25% at 2 weeks, 23% at 4 weeks, and 10% at 6 weeks, respectively. Compared with the model group, the low dose of Chinese medicine and high dose of Chinese medicine treatment could increase the content of Lactobacillus to different degrees (Table 6).

4.2 Using FMT as a technical platform to study the Improvement of Intestinal Flora in CRF with Fushen Granule

4.2.1 General state observation

The rats in the control donor group had significantly increased body weight, good mental condition, free movement, responsiveness, and neat and moist hair. In contrast, the rats in the model group gradually lost weight, ate less, appeared more depressed, squinted, were less active, less responsive, and had loose body hair and shaggy fur. The Fushen Granule donor group or FMT group had a better mental status, weight, activity, reaction, and coat color after treatment, but was still worse than the control group.

4.2.2 Observation of renal function indicators

Compared with the normal donor group, Cr, BUN, UA, and 24h urine microalbumin were significantly higher in the nephrectomy + saline group at weeks 2, 4, and 6, with statistically significant differences (P< 0.05), indicating successful modeling of renal failure. Compared with the nephrectomy + saline group, BUN and 24-h urine microalbumin decreased significantly (P< 0.05) at week 2, and Cr, BUN, UA, and 24-h urine microalbumin also decreased significantly (P< 0.05) at weeks 4 and 6 in the nephrectomy + Fushen Granule group. In contrast, BUN, UA, and 24-h urine microalbumin also decreased significantly in the nephrectomy + normal f group at week 2 (P< 0.05), and Cr, BUN, UA, and 24-h urine microalbumin decreased significantly at weeks 4 and 6 (P< 0.05). The improvement in 24-h urinary microalbumin at weeks 2, 4, and 6 did not significantly differ between the kidney cut + Fushen Granule group and the kidney cut + normal f group (P > 0.05) (Table 7).

4.2.3 Observation of serum endotoxins and metabolic toxin-like indicators of enteric origin

Compared to the normal donor group, the nephrectomy + saline group showed a statistically significant increase (P< 0.05) in IS, PCS, D-lactic acid, and endotoxin at weeks 2, 4, and 6, indicating successful modeling of renal failure. Compared with the nephrectomy + saline group, the nephrectomy + Fushen Granule group showed a significant decrease in PCS, D-lactic acid, and endotoxin at weeks 2 and 4 (P< 0.05), and a significant decrease in IS at week 6 (P< 0.05). Similarly, compared to the nephrectomy + saline group, the nephrectomy + normal f group showed significant decreases in PCS, D-lactic acid, and endotoxin at weeks 2, 4, and 6 (P< 0.05), and a significant decrease in IS at weeks 4 and 6 (P< 0.05). The improvement in D-lactate and endotoxin in the kidney cut + Fushen Granule group at weeks 2, 4, and 6 was not significantly different from that in the kidney cut + normal f group (P > 0.05) (Table 8).



4.2.4 Analysis of intestinal bacterial diversity 4.2.4.1 Sequencing data quality and OTU analysis

In this study, the coverage of each sample library was assessed by calculating the Coverage Index, and values of the Coverage Index were obtained to be >0.99, indicating comprehensive coverage of each sample library and a high detection rate of the sample sequences (Table 9). Hierarchical abundance curves were used to simultaneously interpret the richness and evenness of the species

TABLE 4	Levels of	Firmicutes a	and Bacteroides	of rats in	sham-operated	and model	groups at 0,	, 2, 4 and 6 v	weeks.
---------	-----------	--------------	-----------------	------------	---------------	-----------	--------------	----------------	--------

Group	Time Point (weeks)	Firmicutes (%)	Bacteroides (%)	Firmicutes/Bacteroidetes ratio
	0	70	24	2.97
Sham-operated	2	69	24	2.88
	4	68	22	3.1
	6	63	30	2.1
	0	81	10	8.1
M. 1.1	2	51	46	1.11
Model	4	60	26	2.31
	6	54	40	1.35

TABLE 5 Levels of Firmicutes and Bacteroides of rats in each group at week 2.

Group	Firmicutes (%)	Bacteroidetes (%)	Firmicutes/ Bacteroidetes ratio
Sham	69%	24%	2.88
Sham- operated	56%	37%	1.53
Model	51%	46%	1.11
Fushen Granule at low doses	66%	29%	2.27
Fushen Granule at high doses	62%	33%	1.88

TABLE 6 Levels of Lactobacillus of rats in each group at 2, 4 and 6 weeks.

Group	Lactobacillus 2 weeks	Lactobacillus 4 weeks	Lactobacillus 6 weeks
Sham	0.46	0.25	0.25
Sham- operated	0.30	0.19	0.25
Model	0.25	0.23	0.10
Fushen Granule at low doses	0.36	0.33	0.17
Fushen Granule at high doses	0.20	0.27	0.16

contained in the samples. The graph shows that the distribution of microbial species in the sequenced samples, excluding sample B2, was reasonably even, it is possible that the B2 sample was contaminated during the sampling process, resulting in a significantly different species abundance and proportion from the pre-intervention samples (Figures 4A, B). Dilution curves were used to verify that the sequencing data reflected the actual biodiversity in the samples and indirectly the species richness in the samples. The results showed that the curve levelled off as the number of sequences increased, indicating that the depth of sequencing had largely covered all species in the samples (Figures 4C, D). In addition, the results of the number of OTUs of each sample obtained by clustering using usearch software showed that the number of rat intestinal flora characteristics in groups FD, FE and FF after Fecal microbiota transplantation(FMT) showed an overall increasing trend compared to groups FA, FB and FC before FMT. In terms of the number of rat intestinal flora characteristics after FMT, the kidney cut + normal group FMT (FD) > kidney cut + Fushen Granule FMT (FF) > kidney cut + saline group FMT (FE) (Figure 4E).

4.2.4.2 Alpha diversity analysis

Alpha diversity reflects the richness and diversity of species in a single sample and is measured by various indices such as Chao, Ace, Shannon, Simpson, Coverage, and peritoneal dialysis - whole_tree. The Chao and Ace indices measure species richness, i.e., the number of species, and larger values of the Shannon and Simpson indices are used to indicate the species diversity of a sample (Grice et al., 2009). Using QIIME2 software, the Alpha Diversity Index of the samples was assessed, and the values for each sample were tallied in Table 9. In terms of flora richness, the Ace and Chao indices increased in groups FD, FE, and FF compared to groups FA, FB, and FC, indicating an increase in flora richness in rats after fecal microbiota transplantation(FMT), with statistically significant differences. In terms of bacterial diversity, the Shannon and Simpson indices were used to estimate the diversity of the flora, and the higher the value, the higher the community diversity. It was found that the Shannon and Simpson indices in groups FD, FE, and FF were higher than those in groups FA, FB, and FC. This indicates that the diversity of the rat intestinal flora was restored after FMT. In conclusion, FMT helped to restore the richness and diversity of the rat intestinal flora. In the comparison between the groups, the normal group had the best results for FMT, followed by the FMT of Fushen Granule, and finally the saline sham operation group.

4.2.4.3 Beta diversity analysis

Beta diversity analysis was conducted in this study using QIIME software to compare the extent to which different samples were similar in terms of species diversity. Principal Component Analysis (PCA), Principal Coordinate Analysis (PCoA), and Non-metric Multidimensional Scaling Analysis (NMDS) were used to examine differences between samples. PCA uses variance decomposition to reflect differences between multiple data sets on a two-dimensional coordinate plot, with the axes representing the two eigenvalues that best reflect the variance. PCoA and PCA plots showed that groups FA, FB, and FC were closer together, indicating high consistency and less variability in the samples. On the other hand, samples FD, FE, and FF after 15 days were less similar in terms of species diversity compared to groups FA, FB, and FC, suggesting that the intervention caused a change in species variability. NMDS analysis had a reliable Stress value of 0.0445 (less than 0.2), indicating its accuracy. Overall, the results suggest that the transplantation of Fushen Granule and normal flora had a moderating effect on the changes in the composition of the intestinal flora in 5/6 nephrectomized rats (Figures 5A-C).

4.2.5 Annotation and variability analysis of intestinal flora species

The primary bacteria that dominate at the phylum level include firmicutes, bacteroidetes, proteobacteria, spirochaetes, actinobacteria, patescibacteria, tenericutes, cyanobacteria, and verrucomicrobia. The relative abundance of intestinal flora at the phylum taxonomic level was analyzed in Figure 6. Compared to rats before FMT, the model + saline group, the model + Fushen Granule Donor group, and the model + normal Donor group showed a decreasing trend for Firmicutes bacteria and an increasing trend for

TABLE 7 Indicators of renal function.

Group	Point in time	Cr	Bun	UA (μmol/L)	24-hour urine microalbumin (mg/L)
Normal Donor	Initial	44.39±0.81	20.43±0.44	102.27±1.08	20.28±2.89
	2 weeks	47.80±3.55	23.53±2.35	112.23±9.48	23.57±2.74
	4 weeks	45.89±0.98	22.89±0.95	104.83±2.04	28.32±2.53
	6 weeks	45.89±0.98	25.93±4.39	128.32±8.85	30.28±2.73
Kidney cut + saline	Initial	42.34±1.97	24.45±0.73	105.26±1.15	22.53±4.20
	2 weeks	95.78±1.96**	37.20±0.28*	145.60±1.19**	288.34±21.49***
	4 weeks	110.39±26.23**	33.33±8.98*	146.46±2.00**	482.95±32.94***
	6 weeks	110.39±26.23**	55.82±2.89*	190.33±45.55**	638.29±44.76***
Kidney cut + Fushen Decoction donor	Initial	45.12±2.49	20.81±0.64	105.87±2.73	21.15±2.46
	2 weeks	55.78±3.61###	26.06±3.51 [#]	121.37±6.30 ^{###}	75.24±7.93 ^{###}
	4 weeks	38.15±1.22 ^{###}	32.58±2.09	130.70±20.98 ^{###}	129.27±10.89 ^{###}
	6 weeks	38.15±1.22###	29.70±3.99 [#]	128.50±5.41 ^{###}	91.13±9.27 ^{###}
Kidney cut + Fushen Decoction f	Initial	43.70±1.01	22.38±0.68	104.25±1.37	23.47±3.19
	2 weeks	94.04±3.57	22.56±5.68 [#]	142.57±4.42	100.66±12.18 ^{###, Δ}
	4 weeks	62.45±1.87 ^{###}	23.48±1.12 [#]	111.40±18.23 ^{###}	178.82±12.34 ^{###, Δ}
	6 weeks	62.45±1.87 ^{###}	37.31±0.86 [#]	93.28±3.56 ^{###}	103.28±8.42 ^{###, Δ}
Kidney cut + normal f	Initial	42.29±3.04	22.66±1.39	105.10±1.77	24.72±2.50
	2 weeks	98.63±0.32	24.57±1.40 [#]	137.03±6.89 ^{###}	83.47±8.82 ^{###}
	4 weeks	57.04±2.81 ^{###}	29.01±4.23 [#]	63.47±1.93 ^{###}	153.94±13.76 ^{###}
	6 weeks	57.04±2.81 ^{###}	43.92±0.51 [#]	104.57±15.06 ^{###}	98.73±7.51 ^{###}

Compared with normal donor group,** P< 0.01,*** P< 0.001; compared with kidney cut + saline group, #P< 0.05, ### P< 0.001; compared with kidney cut + normal f group, Δ P > 0.05.

TABLE 8 Indicators for D-lactate, endotoxin and enteric-derived metabolic toxin categories.

Group	Point in time	Metabolic toxins of enteric origin		Serum endotoxin		
		IS (μg/mL)	PCS (µg/mL)	D-Lactic acid (µg/mL)	Endotoxin (EU/ml)	
Normal Donor	Initial	0.91±0.13	0.17±0.02	7.34±0.28	0.023±0.003	
	2 weeks	0.93±0.06	0.15±0.1	7.52±0.36	0.028±0.003	
	4 weeks	0.85±0.21	0.13±0.02	7.88±0.42	0.032±0.002	
	6 weeks	0.86±0.23	0.15±0.01	7.52±0.28	0.029±0.004	
Kidney cut + saline	Initial	0.83±0.12	0.14 ± 0.01	7.58±0.19	0.030 ± 0.004	
	2 weeks	5.69±2.87**	1.73±0.22***	18.19±0.54***	0.387±0.032***	
	4 weeks	5.24±0.73***	1.8±0.21***	15.28±1.25***	0.498±0.028***	
	6 weeks	6.68±2.17***	2.12±0.62***	15.28±1.25***	0.518±0.028***	
Kidney Cut + Fushen Decoction Donor	Initial	0.94±0.17	0.18±0.03	7.28±0.32	0.026±0.005	
	2 weeks	4.98±1.25	1.28±0.31#	12.33±0.17 ^{###}	0.224±0.021 ^{###}	
	4 weeks	4.83±1.00	0.37±0.06 ^{###}	10.25±0.93 ^{###}	0.243±0.017 ^{###}	
	6 weeks	3.64±1.10 [#]	0.39±0.08 ^{###}	10.25±0.93 ^{###}	0.217±0.014 ^{###}	

(Continued)

TABLE 8 Continued

Group	Point in time	Metabolic toxins of enteric origin		Serum endotoxin	
		IS (μg/mL)	PCS (µg/mL)	D-Lactic acid (µg/mL)	Endotoxin (EU/ml)
Kidney cut + Fushen Decoction f	Initial	0.89±0.12	0.16±0.02	7.69±0.33	0.028±0.002
	2 weeks	4.90±1.31	1.03±0.19 ^{##}	14.28±1.23 ^{###, Δ}	0.258±0.028 ^{###, Δ}
	4 weeks	4.22±0.92	0.93±0.15 ^{###}	12.15±1.02 ^{##, Δ}	0.305±0.020 ^{###, ΔΔ}
	6 weeks	4.08±1.36 [#]	0.86±0.21 ^{###}	12.15±1.02 ^{###, Δ}	0.293±0.021 ^{###, Δ}
Kidney cut + normal f	Initial	0.86±0.09	0.15±0.01	7.21±0.21	0.027±0.003
	2 weeks	3.92±1.19	0.88±0.09 ^{###}	13.57±0.67 ^{###}	0.206±0.019 ^{###}
	4 weeks	3.49±1.25 [#]	0.75±0.11 ^{###}	12.57±0.78 ^{##}	0.251±0.019 ^{###}
	6 weeks	2.78±0.64 ^{##}	0.64±0.09 ^{###}	12.57±0.78 ^{###}	0.264±0.029 ^{###}

Compared to the normal donor group, *** indicates P < 0.001; compared to the nephrectomy + saline group, # indicates P < 0.05, ## indicates P < 0.01, ### indicates P < 0.001; compared to the nephrectomy + normal f group, Δ indicates P > 0.05.

Bacteroidetes. Compared with the model + saline sham-operated group, the relative abundance of these two groups decreased less in the model + Fushen Granule Donor group and the model + normal Donor group, suggesting that the Fushen Granule and normal flora transplantation had a moderating effect on the relative abundance of colonic flora at the phylum level. At the genus level, the model group exhibited a decreasing trend of Lactobacillus after FMT. Compared with the Kidney-cut + saline group during the same period, the Kidney-cut + Fushen Granule Donor group and the Kidney-cut + normal Donor group had some degree of restorative effect on Lactobacillus, and the Fushen Granule FMT was superior to the other two groups.

4.2.6 LefSe analysis

In order to screen for colonies with significant differences between groups, this study employed a novel metagenomic analysis method called LefSe (Linear Discriminant Analysis Size Effect). This method is a statistical approach used in the fields of genetics and microbiology to identify high-dimensional biomarkers and reveal genomic features. LefSe analysis combines linear discriminant analysis with non-parametric tests to identify multiple biomarkers and enable comparisons between multiple groups. In this study, LDA values greater than 3.5 were used as screening criteria to examine the abundance of the flora. In an evolutionary branching tree diagram, different circles represent various taxonomic levels from inner to outer, including domain, kingdom, phylum, class, order, family, genus, and species. Different nodes in the diagram represent dominant genera, while the connections between nodes indicate their correlation. The thickness of the lines reflects the strength of the correlation; thicker lines represent stronger correlations. Additionally, the size of the nodes represents the number of associations with other microorganisms; the more associations, the larger the node. The results showed that Model Group FA had significant differences in a) uncultured_bacterium_g_Prevotella_1 and b) g_prevotella_1. Group FE exhibited significant differences in c) o_Clostridiales after FMT (Figure 7).

5 Discussion

As a progressive, irreversible disease with a complex pathogenesis, limited therapeutic modalities, and issues of tolerance and dependence, the development of effective drugs to treat or alleviate CRF has become an urgent task (Eirin and Lerman, 2014). The gut's microbial composition is a key player in renal

TABLE 9	Displays the analysis of	of intestinal flora	diversity in each	group of rats (\bar{x} ±	s, n = 3).
---------	--------------------------	---------------------	-------------------	-----------------------------	------------

Group	Chao	Ace	Shannon	Simpson	Coverage
FA	502.8182 ± 10.0702	495.2557 ± 9.7063	5.5293 ± 0.1972	0.9363 ± 0.0128	0.9987 ± 0
FB	564.9597 ± 10.163	569.3319 ± 19.4272	6.2448 ± 0.7023	0.951 ± 0.0204	0.9983 ± 0
FC	548.2892 ± 15.9462	535.5732 ± 13.0593	5.1845 ± 0.3452	0.9081 ± 0.0152	0.9983 ± 0.0003
FD	658.0036 ± 9.4833	645.3728 ± 9.0772	6.04 ± 0.5148	0.9244 ± 0.0399	0.9989 ± 0
FE	582.801 ± 7.0036	578.1836 ± 6.6149	5.6382 ± 0.4512	0.9257 ± 0.0281	0.9985 ± 0
FF	603.0205 ± 5.3112	583.5407 ± 15.2415	5.8779 ± 0.5683	0.9368 ± 0.0241	0.9984 ± 0.0005

FA before fecal microbiota transplantation(FMT) in the model group + Donor group; FB before FMT in the model group + control Donor group; FC before model group + saline intervention; FD after FMT in the model group + control Donor group; FE after model group + saline intervention; FF after FMT in the model group + Donor group.



coordinate is the number of randomly selected sequencing strips, and the vertical coordinate is the number of features obtained based on the number of sequencing strips. Each curve represents a sample and is marked with a different color. **(E)** Plot of the number of features for each group of samples. The horizontal coordinate is the name of the sample, the vertical coordinate is the number of features, and the number above the bar is the number of OTUs for the corresponding sample.

disease and is closely related to the development of CRF (Meijers et al., 2019; Zhu et al., 2021; Krukowski et al., 2023). Based on our findings, the use of Fushen Granule can regulate the intestinal microbiota and control the progression of CRF. And we demonstrated that FMT in rats in the Fushen Granule group reversed the severe taxonomic and functional imbalance caused by CRF to a certain extent. This represents the first published study to date utilizing the combined treatment of Fushen Granules and FMT for CRF. The Fushen Granule and its compound "Fushen Granules" have been used clinically in the treatment of various kidney diseases such as peritoneal dialysis and interstitial fibrosis (Chen, 2021; Ding, 2022). Modern pharmacological studies have also shown that Astragalus, the primary component of Fushen Granule, has the ability to reduce urinary protein (Zhang et al., 2014) and protect kidney function (Zhou et al., 2017). The combination of Astragalus and Salvia miltiorrhiza can slow down



FIGURE 5

The beta diversity analysis. (A) PCA. The dots indicate each sample with different colors representing different groupings. The horizontal and vertical coordinates represent the two eigenvalues that cause the greatest variation between samples, with the percentage indicating the contribution of each principal component to the variation of the samples. (B) PCoA. Points represent each sample separately, with different colors representing different groupings. The horizontal and vertical coordinates represent the two characteristic values that cause the greatest variation between samples, and the degree of influence is represented as a percentage. The closer the samples are on the coordinate plot, the greater the similarity. (C) NMDS. The dots represent each sample separately, with different groupings, and the distance between the dots indicates the degree of difference. A Stress value of less than 0.2 indicates that NMDS analysis has a certain degree of reliability, and the closer the samples are on the coordinate graph, the higher the similarity.



FIGURE 6

Histogram of the relative abundance of species composition of multiple samples for community composition analysis. (A) phylum level, (B) genus level. FA before FMT in the model group + Donor group; FB before FMT in the model group + control Donor group; FC before model group + saline intervention; FD after FMT in the model group + control Donor group; FE after model group + saline intervention; FF after FMT in the model group + Donor group; FE after model group + saline intervention; FF after FMT in the model group + Donor group.



kidney fibrosis (Han et al., 2021) and suggests a high level of drug safety. In vitro and clinical trials have demonstrated that Fushen Granule can, to a certain extent, correct intestinal dysfunction in peritoneal dialysis patients and delay the process of peritoneal fibrosis. Meanwhile, there is increasing evidence suggesting that traditional Chinese medicine plays a beneficial role in the treatment of chronic kidney disease by modulating dysbiosis of the intestinal microbiota (Gao et al., 2021; Ming et al., 2021). However, the effect of Fushen Granule on CRF and the specific potential mechanism remain to be elucidated. Therefore, in this experiment, the effect of Fushen Granule on CRF rats was investigated using a 5/6 nephrectomized CRF rat model. The experimental results showed that the basic conditions of CRF rats were alleviated after different doses of Fushen Granule, and the levels of the intestinal toxins indoxyl sulfate (IS) and p-cresyl sulfate (PCS), as well as D-lactic acid and endotoxin, which are indicators of intestinal barrier damage, were reduced in a dose-related manner. At the same time, the number of operational taxonomic units (OTUs), the ratio of Firmicutes to Bacteroidetes, and the number of lactic acid bacteria increased after Fushen Granule treatment, which restored to some extent the changes in the abundance and structure of the intestinal flora of the rats caused by slow renal failure.

CRF is accompanied by a progressive decrease in glomerular filtration rate and the continued accumulation of metabolic waste products that can accelerate the progression of CRF and affect its prognosis. The enteric-derived toxins indoxyl sulfate (IS) and p-cresol sulfate(PCS) are known as protein-bound toxins due to their proteinbinding properties and are currently not filtered out by dialysis. In normal renal function, these two substances are secreted into the renal tubules and excreted from the body. In CRF, the intestinal tract undergoes a series of changes, including a lack of gastrointestinal motility and reduced digestive capacity of the small intestine, which increases the amount and retention time of dietary protein in the colon, which further contributes to the increase in the number of colonic proteolytic bacteria, resulting in increased production of the IS and PCS precursors indole and p-cresol, and ultimately an increase in IS and PCS generation. At the same time, renal filtration function is reduced, resulting in the accumulation of metabolic toxins such as IS and PCS in the body. The long-term retention of IS and PCS in the body can further aggravate glomerulosclerosis and renal fibrosis, leading to a vicious circle (Ito et al., 2010; Lekawanvijit et al., 2010; Niwa, 2010; Liabeuf et al., 2011; Shimizu et al., 2011; Lin et al., 2012; Liu et al., 2012). Therefore, reducing the level of IS and PCS is one of the most important problems to be solved. There is a lack of effective means of IS and PCS clearance, and most of the research on reducing IS and PCS has been conducted in the intestinal tract from the perspective of inhibiting their production, such as with AST120 (Shimoishi et al., 2007; Kikuchi et al., 2010; Asai et al., 2019). Since enteric-derived toxin precursors are primarily produced by the enzymatic processes of the intestinal flora, it is possible to reduce their production by adjusting the structure of the intestinal flora. In the present study, Fushen Granule demonstrated a notable downregulating effect on the levels of intestine-derived metabolic toxins IS and PCS in CRF rats. It is hypothesized that the mechanism may be related to improving the intestinal tract status and promoting the excretion of toxin precursors.

Serum endotoxin is a metabolite or component of intestinal bacteria that can translocate into the circulation when the barrier is not functioning properly (Violi et al., 2023), it is used as an indicator of intestinal barrier function (Vancamelbeke and Vermeire, 2017). Fushen Granule can improve renal function indicators by repairing intestinal barrier function, reducing metabolic toxins in the serum, and enhancing renal function. D-lactate is a bacterial metabolite produced by the intestinal flora, and its level is low and relatively stable under normal conditions. Elevated levels may reveal the extent of damage to the intestinal mucosa (Guo et al., 2019). Endotoxin, a major component of the outer membrane of Gram-negative bacteria, can translocate into the body's circulation when intestinal barrier function is compromised, transforming host immune defenses into a pro-inflammatory state (Violi et al., 2023). The endotoxin itself can further contribute to the deterioration of the mucosal barrier function (Tang et al., 2019). In this study, serum endotoxin and D-lactate levels were measured in CRF rats to assess intestinal barrier function. In the sham-operated group, serum endotoxin levels were stable, whereas in the model group, serum endotoxin levels increased with the progression of CRF. The model rats also showed a significant increase in serum D-lactate, both of which are indicative of impaired intestinal barrier function. Fushen Granule reduced the variation of endotoxin and down-regulated the serum D-lactate level, which had a protective effect on the intestinal barrier function and thus delayed the progression of CRF.

Changes in the intestinal flora play a key role in accelerating the progression of CRF, and this study found that Fushen Granule was able to modify the structure of intestinal flora in rats to slow down the progression of CRF. Previous research has reported that in patients with ESRD, the number of Firmicutes, Actinobacteria, Proteobacteria, and Lactobacillus decreased(Vaziri et al., 2013; Simões-Silva et al., 2020), while the number of Bacteroides increased (Crespo-Salgado et al., 2016). In patients with CKD, there was a lower abundance of Lactobacillus and an increased proportion of Enterobacteriaceae (Mahmoodpoor et al., 2017; Lau et al., 2018). It is evident that there is an imbalance between the intestinal flora and the host in the disease state, which may lead to the accumulation of uremic toxins while limiting the beneficial functions and products conferred by the normal flora, thereby accelerating the progression of renal disease (Vaziri et al., 2013). In contrast, the homeostatic gut microbiota serves the host through a wide range of physiological activities, including the prevention of pathogens, maintenance of the function and integrity of the intestinal epithelium, and regulation of the host immune system (Bian et al., 2022). 16S rRNA or DNA sequencing is the common method to evaluate microbial diversity and identify differential microbes in patients compared with healthy control subjects (Cao et al., 2022). The present study, combined with 16S rRNA gene sequencing experiments, aimed to investigate the mechanism of action of Fushen Granule in improving CRF from the perspective of structural changes in the intestinal flora. The results showed that the rats in the model group exhibited a decrease in the number of OTUs in the intestinal flora. At the phylum level, the relative abundance of bacteria in the Firmicutes phylum decreased in the model group after 5/6 nephrectomy compared to the sham-operated group during the same period, while the content of the Bacteroidetes phylum increased, and the ratio of the Firmicutes phylum to the Bacteroidetes phylum showed a decreasing trend. After Fushen Granule intervention, the low-dose and high-dose groups of Chinese medicine reduced this alteration to different degrees, thus restoring the normal ratio of Firmicutes and Bacteroidetes to a certain extent and repairing the alteration of the composition ratio of intestinal flora caused by slow renal failure. At the genus level, the relative abundance comparison showed that Lactobacillus was the predominant genus. This genus is a probiotic strain that can slow down the progression of kidney disease by improving the intestinal

environment (Yoshifuji et al., 2016). The relative abundance of Lactobacillus in the model group showed a decreasing trend compared to the sham-operated group during the same period. In contrast, the relative abundance of Lactobacillus in the low-dose and high-dose groups of Chinese medicine showed an increasing trend after treatment. This indicates that Fushen Granule can, to a certain extent, reverse the effect of 5/6 nephrectomy on the intestinal flora of rats, correct the disorder of flora, and make it converge to the normal group, and has the effect of improving the intestinal flora of CRF rats. In conclusion, the restoration of the altered intestinal flora in the CRF state by Fushen Granule may be another mechanism of its action in delaying renal failure.

To further investigate whether Fushen Granule exerts its therapeutic effect through the regulation of intestinal flora, a FMT experiment was conducted to verify the effect of Fushen Granule. Multiple studies indicate that the transplantation of a healthy gut microbiota plays a therapeutic role in various renal diseases. A clinical case study report suggests that FMT reduces the accumulation of PBUTs in the host by modulating the intestinal microbiota's amino acid metabolism, consequently mitigating the progression of CKD (Liu et al., 2022). In investigations on IgAN, it has been reported that FMT-induced microbial transfer regulates the IgAN phenotype, thereby paving the way for new treatment modalities for IgAN patients (Lauriero et al., 2021). Furthermore, in various published studies, FMT has consistently been a valuable tool in verifying the correlation between gut microbiota dysbiosis and the progression of CRF, indicating its value in restoring the intestinal microbiota in CRF. However, there is currently limited understanding of the application of FMT in the treatment of CRF. In this study, we used a rat model to comprehensively evaluate the effects of Fushen Granule on the improvement of serum intestinal endotoxins, intestinal flora structure and diversity, and renal function in CRF with the aid of FMT technology. The results of the study showed that both the Fushen Granule Donor Group and the FMT Group could effectively improve the general status of CRF rats, enhance renal function, reduce serum levels of toxins, and improve intestinal status to promote excretion of toxin precursors, reduce D-lactate, narrow the rise of endotoxin, and protect intestinal barrier function after treatment. Alpha and beta diversity were assessed in the feces collected from each group of rats at the end of fecal microbiota transplantation(FMT). Alpha diversity was assessed mainly from the Ace, Chao, Shannon, and Simpson indices. The results showed that all the above indices exhibited different degrees of increase in the model group after FMT, suggesting that the diversity of rat intestinal flora was restored and the abundance tended to increase. The Ace, Chao, Shannon, and Simpson indices of the model + control Donor group and the model + Fushen Granule Donor group were very close to each other compared to the model + saline sham group. β diversity was demonstrated by PCA, PCoA, and NMDS, and the results showed that there was little overlap between the three groups, suggesting significant differences in the structure of the flora between the groups. Further analysis of the composition and species abundance of the intestinal flora of each group revealed that Fushen Granule and normal flora transplantation had a moderating effect on the structure and relative abundance of the intestinal flora of CRF rats.

6 Conclusion

Based on the 16SrRNA amplification and sequencing technology, this study investigated the mechanism of action of Fushen Granule in the prevention and treatment of CRF in rats with 5/6 nephrectomy as a classical renal failure animal model. The results showed that Fushen Granule was effective in the treatment of CRF rats, potentially delaying the process of CRF by repairing intestinal barrier function, reducing metabolic toxins in serum, improving renal function indicators, and adjusting the structure of intestinal flora. In addition, through FMT experiments, we found that the transplantation of feces from the Fushen Granule Group was able to replicate the intestinal flora structure of the original Fushen Granule Group to a certain extent in the subject rats, and improve the renal function and intestinal status of the CRF rats. This further demonstrates that Fushen Granule can delay the progression of CRF by regulating the "renal-intestinal axis" and improving the intestinal flora structure. This study demonstrates that the Fushen Granules are advantageous in enhancing the structure of the intestinal microbiota and play a constructive role in the prevention and treatment of CRF, thereby offering a scientific basis for its clinical application. The results also suggest that reshaping the homeostasis of the intestinal flora may be an important therapeutic target to slow down the progression of CRF.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

The animal study was approved by Institute of Radiology, Chinese Academy of Medical Sciences. The study was conducted in accordance with the local legislation and institutional requirements.

References

Ahmad, S., and Bromberg, J. S. (2016). Current status of the microbiome in renal transplantation. *Curr. Opin. Nephrol. Hypertens.* 25 (6), 570–576. doi: 10.1097/mnh.0000000000262

Asai, M., Kumakura, S., and Kikuchi, M. (2019). Review of the efficacy of AST-120 (KREMEZIN([®])) on renal function in chronic kidney disease patients. *Ren Fail* 41 (1), 47–56. doi: 10.1080/0886022x.2018.1561376

Bian, J., Liebert, A., Bicknell, B., Chen, X. M., Huang, C., and Pollock, C. A. (2022). Faecal microbiota transplantation and chronic kidney disease. *Nutrients* 14 (12), 2. doi: 10.3390/nu14122528

Cao, C., Zhu, H., Yao, Y., and Zeng, R. (2022). Gut dysbiosis and kidney diseases. Front. Med. (Lausanne) 9. doi: 10.3389/fmed.2022.829349

Chen, J. (2021). A study on the intervention of TGF- β 1/BMP-7/Gremlin pathway on peritoneal fibrosis based on Chinese medicine and method to explore the regulation of Fu Ren formula. Masters (Tianjin University of Chinese Medicine). doi: 10.27368/d.cnki.gtzyy.2021.000070

Chen, X. (2016). "Effect of kidney support granules on peritoneal dialysis intestinal dysfunction"," in 2016 Academic Annual Meeting of the Renal Disease Specialized Committee of the Chinese Society of Integrative Medicine. 381.

Author contributions

LW: Methodology, Visualization, Writing – original draft. AX: Software, Writing – original draft. JW: Software, Writing – review & editing. GF: Writing – review & editing. RL: Formal analysis, Methodology, Writing – review & editing. LJW: Data curation, Writing – review & editing. MP: Conceptualization, Funding acquisition, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work is supported by the National Natural Science Foundation of China (2019YFC1709401).

Acknowledgments

The authors are thankful to Tianjin University of Traditional Chinese Medicine for the help in conducting this study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Crespo-Salgado, J., Vehaskari, V. M., Stewart, T., Ferris, M., Zhang, Q., Wang, G., et al. (2016). Intestinal microbiota in pediatric patients with end stage renal disease: a Midwest Pediatric Nephrology Consortium study. *Microbiome* 4 (1), 50. doi: 10.1186/ s40168-016-0195-9

Cui, M., Xiao, H., Luo, D., Zhang, X., Zhao, S., Zheng, Q., et al. (2016). Circadian rhythm shapes the gut microbiota affecting host radiosensitivity. *Int. J. Mol. Sci.* 17 (11), 1-2,7. doi: 10.3390/ijms17111786

Dai, P., Chang, W., Xin, Z., Cheng, H., Ouyang, W., and Luo, A. (2021). Retrospective study on the influencing factors and prediction of hospitalization expenses for chronic renal failure in China based on random forest and LASSO regression. *Front. Public Health* 9. doi: 10.3389/fpubh.2021.678276

de Almeida Duarte, J. B., de Aguilar-Nascimento, J. E., Nascimento, M., and Nochi, R. J. Jr. (2004). Bacterial translocation in experimental uremia. *Urol Res.* 32 (4), 266–270. doi: 10.1007/s00240-003-0381-7

Ding, F. (2022). Investigating the mechanism of Fu Ren Formula for the prevention and treatment of peritoneal dialysis-associated peritonitis based on TLR4/NF- κ B signaling pathway. Masters (Tianjin University of Chinese Medicine). doi: 10.27368/d.cnki.gtzyy.2022.000005

Dou, Y. (2015). Effect of kidney support granules on the expression of activation factors related to peritoneal fibrosis in rats on peritoneal dialysis," in 2015 academic annual meeting of renal disease specialized committee of chinese society of integrative medicine and western medicine. 1200–1201.

Eirin, A., and Lerman, L. O. (2014). Mesenchymal stem cell treatment for chronic renal failure. *Stem Cell Res. Ther.* 5 (4), 83. doi: 10.1186/scrt472

Fang, J., Guo, Y., Yin, W., Zhang, L., Li, G., Ma, J., et al. (2023). Neoxanthin alleviates the chronic renal failure-induced aging and fibrosis by regulating inflammatory process. *Int. Immunopharmacol.* 114, 109429. doi: 10.1016/j.intimp.2022.109429

Faria, M., and de Pinho, M. N. (2021). Challenges of reducing protein-bound uremic toxin levels in chronic kidney disease and end stage renal disease. *Transl. Res.* 229, 115–134. doi: 10.1016/j.trsl.2020.09.001

Feroze, U., Kalantar-Zadeh, K., Sterling, K. A., Molnar, M. Z., Noori, N., Benner, D., et al. (2012). Examining associations of circulating endotoxin with nutritional status, inflammation, and mortality in hemodialysis patients. *J. Ren Nutr.* 22 (3), 317–326. doi: 10.1053/j.jrn.2011.05.004

Gao, Y., Yang, R., Guo, L., Wang, Y., Liu, W. J., Ai, S., et al. (2021). Qing-re-xiaozheng formula modulates gut microbiota and inhibits inflammation in mice with diabetic kidney disease. *Front. Med. (Lausanne)* 8. doi: 10.3389/fmed.2021.719950

Gelasco, A. K., and Raymond, J. R. (2006). Indoxyl sulfate induces complex redox alterations in mesangial cells. *Am. J. Physiol. Renal Physiol.* 290 (6), F1551–F1558. doi: 10.1152/ajprenal.00281.2004

Gonçalves, S., Pecoits-Filho, R., Perreto, S., Barberato, S. H., Stinghen, A. E., Lima, E. G., et al. (2006). Associations between renal function, volume status and endotoxaemia in chronic kidney disease patients. *Nephrol. Dial Transplant.* 21 (10), 2788–2794. doi: 10.1093/ndt/gfl273

Grice, E. A., Kong, H. H., Conlan, S., Deming, C. B., Davis, J., Young, A. C., et al. (2009). Topographical and temporal diversity of the human skin microbiome. *Science* 324 (5931), 1190–1192. doi: 10.1126/science.1171700

Guo, Y., Li, H., Liu, Z., Li, C., Chen, Y., Jiang, C., et al. (2019). Impaired intestinal barrier function in a mouse model of hyperuricemia. *Mol. Med. Rep.* 20 (4), 3292–3300. doi: 10.3892/mmr.2019.10586

Han, C., Jiang, Y. H., Li, W., and Liu, Y. (2021). Astragalus membranaceus and Salvia miltiorrhiza ameliorates cyclosporin A-induced chronic nephrotoxicity through the "gut-kidney axis". J. Ethnopharmacol. 269, 113768. doi: 10.1016/j.jep.2020.113768

Ito, S., Osaka, M., Higuchi, Y., Nishijima, F., Ishii, H., and Yoshida, M. (2010). Indoxyl sulfate induces leukocyte-endothelial interactions through up-regulation of Eselectin. J. Biol. Chem. 285 (50), 38869–38875. doi: 10.1074/jbc.M110.166686

Jiang, C., Jiao, S., and Yang, H. (2020a). Effect of kidney support granules on 1-year prognosis of patients with peritoneal dialysis-associated peritonitis. *Chin. J. Integrated Traditional West. Med.* 40 (11), 1333–1338.

Jiang, C., Lin, W., Wang, L., Lv, Y., Song, Y., Chen, X., et al. (2020b). Fushen Granule, A Traditional Chinese Medicine, ameliorates intestinal mucosal dysfunction in peritoneal dialysis rat model by regulating p38MAPK signaling pathway. *J. Ethnopharmacol.* 251, 112501. doi: 10.1016/j.jep.2019.112501

Kikuchi, K., Itoh, Y., Tateoka, R., Ezawa, A., Murakami, K., and Niwa, T. (2010). Metabolomic search for uremic toxins as indicators of the effect of an oral sorbent AST-120 by liquid chromatography/tandem mass spectrometry. *J. Chromatogr. B Anal. Technol. BioMed. Life Sci.* 878 (29), 2997–3002. doi: 10.1016/j.jchromb.2010.09.006

Krukowski, H., Valkenburg, S., Madella, A. M., Garssen, J., van Bergenhenegouwen, J., Overbeek, S. A., et al. (2023). Gut microbiome studies in CKD: opportunities, pitfalls and therapeutic potential. *Nat. Rev. Nephrol.* 19 (2), 87–101. doi: 10.1038/s41581-022-00647-z

Lau, W. L., Savoj, J., Nakata, M. B., and Vaziri, N. D. (2018). Altered microbiome in chronic kidney disease: systemic effects of gut-derived uremic toxins. *Clin. Sci. (Lond)* 132 (5), 509–522. doi: 10.1042/cs20171107

Lauriero, G., Abbad, L., Vacca, M., Celano, G., Chemouny, J. M., Calasso, M., et al. (2021). Fecal microbiota transplantation modulates renal phenotype in the humanized mouse model of igA nephropathy. *Front. Immunol.* 12. doi: 10.3389/fimmu.2021.694787

Lei, Y. (2016). "Observations on the clinical efficacy of blood purification and kidney support granules in the treatment of chronic renal failure and the impact on patients' quality of life.", in: 2016 Academic Annual Meeting of the Renal Disease Specialized Committee of the Chinese Society of Integrative Medicine), 1.

Lei, Y., and Yang, H. (2018). Efficacy of kidney support granules combined with colonic enema on intestinal dysfunction after peritoneal dialysis catheterization. *Chin. J. Surg. Integrated Traditional West. Med.* 24 (03), 294–298.

Lei, Y., Yang, B., Ming, P., and Yang, H. (2018a). Forty-six cases of gastrointestinal dysfunction in peritoneal dialysis patients treated by enema combined with kidney support granules. *Chin. J. Surg. Integrated Traditional West. Med.* 24 (04), 441–446.

Lei, Y., Yang, B., Pei, M., Lin, Y., and Yang, H. (2018b). Effects of Kidney Support Formula on the expression of TGF- β 1, E-cadherin and α -SMA in peritoneal mesenchyme of uremic rats. *Tianjin J. Traditional Chin. Med.* 35 (07), 521–525.

Lei, Y., Yang, B., Wang, M., Qiao, Y., Zhao, Y., and Yang, H. (2018c). Discussion on the effect and mechanism of Kidney Supporting Formula on the transdifferentiation of rat peritoneal mesothelial cells. *Liaoning J. Traditional Chin. Med.* 45 (07), 1500–1503. doi: 10.13192/j.issn.1000-1719.2018.07.052

Lekawanvijit, S., Adrahtas, A., Kelly, D. J., Kompa, A. R., Wang, B. H., and Krum, H. (2010). Does indoxyl sulfate, a uraemic toxin, have direct effects on cardiac fibroblasts and myocytes? *Eur. Heart J.* 31 (14), 1771–1779. doi: 10.1093/eurheartj/ehp574

Li, J., Yang, B., and Yang, H. (2020). Overview of the mechanism of action of renal pellets in regulating decorin interference with peritoneal collagen stability. *Inner Mongolia J. Traditional Chin. Med.* 39 (02), 129–131. doi: 10.16040/j.cnki.cn15-1101.2020.02.081

Liabeuf, S., Drüeke, T. B., and Massy, Z. A. (2011). Protein-bound uremic toxins: new insight from clinical studies. *Toxins (Basel)* 3 (7), 911–919. doi: 10.3390/toxins3070911

Lin, C. J., Liu, H. L., Pan, C. F., Chuang, C. K., Jayakumar, T., Wang, T. J., et al. (2012). Indoxyl sulfate predicts cardiovascular disease and renal function deterioration in advanced chronic kidney disease. *Arch. Med. Res.* 43 (6), 451–456. doi: 10.1016/j.arcmed.2012.08.002

Liu, S., Wang, B. H., Kompa, A. R., Lekawanvijit, S., and Krum, H. (2012). Antagonists of organic anion transporters 1 and 3 ameliorate adverse cardiac remodelling induced by uremic toxin indoxyl sulfate. *Int. J. Cardiol.* 158 (3), 457–458. doi: 10.1016/j.ijcard.2012.05.022

Liu, X., Zhang, M., Wang, X., Liu, P., Wang, L., Li, Y., et al. (2022). Fecal microbiota transplantation restores normal fecal composition and delays Malignant development of mild chronic kidney disease in rats. *Front. Microbiol.* 13. doi: 10.3389/fmicb.2022.1037257

Liyanage, T., Ninomiya, T., Jha, V., Neal, B., Patrice, H. M., Okpechi, I., et al. (2015). Worldwide access to treatment for end-stage kidney disease: a systematic review. *Lancet* 385 (9981), 1975–1982. doi: 10.1016/s0140-6736(14)61601-9

Mahmoodpoor, F., Rahbar Saadat, Y., Barzegari, A., Ardalan, M., and Zununi Vahed, S. (2017). The impact of gut microbiota on kidney function and pathogenesis. *BioMed. Pharmacother.* 93, 412–419. doi: 10.1016/j.biopha.2017.06.066

Matsushita, K., Ballew, S. H., Wang, A. Y., Kalyesubula, R., Schaeffner, E., and Agarwal, R. (2022). Epidemiology and risk of cardiovascular disease in populations with chronic kidney disease. *Nat. Rev. Nephrol.* 18 (11), 696–707. doi: 10.1038/s41581-022-00616-6

Meijers, B., Evenepoel, P., and Anders, H. J. (2019). Intestinal microbiome and fitness in kidney disease. *Nat. Rev. Nephrol.* 15 (9), 531–545. doi: 10.1038/s41581-019-0172-1

Ming, Y., Cheng, S., Long, W., Wang, H. L., Xu, C., Liu, X., et al. (2021). The herbal formula granule prescription mahuang decoction ameliorated chronic kidney disease which was associated with restoration of dysbiosis of intestinal microbiota in rats. *Evid Based Complement Alternat Med.* 2021, 4602612. doi: 10.1155/2021/4602612

Miyazaki, T., Ise, M., Hirata, M., Endo, K., Ito, Y., Seo, H., et al. (1997). Indoxyl sulfate stimulates renal synthesis of transforming growth factor-beta 1 and progression of renal failure. *Kidney Int. Suppl.* 63, S211–S214.

Niwa, T. (2010). Indoxyl sulfate is a nephro-vascular toxin. J. Ren Nutr. 20 (5 Suppl), S2–S6. doi: 10.1053/j.jrn.2010.05.002

Pluznick, J. L. (2020). The gut microbiota in kidney disease. Science 369 (6510), 1426–1427. doi: 10.1126/science.abd8344

Ruiz-Ortega, M., Rayego-Mateos, S., Lamas, S., Ortiz, A., and Rodrigues-Diez, R. R. (2020). Targeting the progression of chronic kidney disease. *Nat. Rev. Nephrol.* 16 (5), 269–288. doi: 10.1038/s41581-019-0248-y

Saran, R., Robinson, B., Abbott, K. C., Agodoa, L. Y. C., Bhave, N., Bragg-Gresham, J., et al. (2018). US renal data system 2017 annual data report: epidemiology of kidney disease in the United States. *Am. J. Kidney Dis.* 71 (3 Suppl 1), A7. doi: 10.1053/j.ajkd.2018.01.002

Shimizu, H., Bolati, D., Adijiang, A., Adelibieke, Y., Muteliefu, G., Enomoto, A., et al. (2011). Indoxyl sulfate downregulates renal expression of Klotho through production of ROS and activation of nuclear factor-κB. *Am. J. Nephrol.* 33 (4), 319–324. doi: 10.1159/000324885

Shimoishi, K., Anraku, M., Kitamura, K., Tasaki, Y., Taguchi, K., Hashimoto, M., et al. (2007). An oral adsorbent, AST-120 protects against the progression of oxidative stress by reducing the accumulation of indoxyl sulfate in the systemic circulation in renal failure. *Pharm. Res.* 24 (7), 1283–1289. doi: 10.1007/s11095-007-9248-x

Simões-Silva, L., Araujo, R., Pestana, M., Soares-Silva, I., and Sampaio-Maia, B. (2020). Peritoneal microbiome in end-stage renal disease patients and the impact of peritoneal dialysis therapy. *Microorganisms* 8 (2), 4–9. doi: 10.3390/microorganisms8020173

Sun, A., and Liu, Z. (2010). Therapeutic efficacy of enema with western medicine in the treatment of chronic renal failure by Fu Ren Tang and Enema Formula. *Shaanxi J. Traditional Chin. Med.* 31 (04), 397–398.

Szeto, C. C., Kwan, B. C., Chow, K. M., Lai, K. B., Chung, K. Y., Leung, C. B., et al. (2008). Endotoxemia is related to systemic inflammation and atherosclerosis in peritoneal dialysis patients. *Clin. J. Am. Soc. Nephrol.* 3 (2), 431–436. doi: 10.2215/ cjn.03600807

Tang, G., Yang, H., Lin, Y., Bu, Y., and Chen, D. (2018a). Mechanism study on the improvement of renal anemia in rats with abdominal dialysis model by supporting kidney granules. *Liaoning J. Traditional Chin. Med.* 45 (09), 1971–1973. doi: 10.13192/j.issn.1000-1719.2018.09.057

Tang, G., Yang, H., Lin, Y., Guo, S., Bu, Y., and Mei, X. (2017). Molecular mechanism of improvement of peritoneal ultrafiltration function by kidney support granules. *Chin. J. Exp. Traditional Med. Formulae* 23 (23), 103–108. doi: 10.13422/j.cnki.syfjx. 2017230103

Tang, G., Yang, H., Lin, Y., Lv, Y., Wang, C., Guo, S., et al. (2018b). Study on the mechanism of inhibition of peritoneal fibrosis in peritoneal dialysis rats by regulating the production of AGEs by Fukui Kidney Granules. *China J. Traditional Chin. Med. Pharm.* 33 (07), 3052–3055.

Tang, W. H. W., Li, D. Y., and Hazen, S. L. (2019). Dietary metabolism, the gut microbiome, and heart failure. *Nat. Rev. Cardiol.* 16 (3), 137–154. doi: 10.1038/s41569-018-0108-7

Vancamelbeke, M., and Vermeire, S. (2017). The intestinal barrier: a fundamental role in health and disease. *Expert Rev. Gastroenterol. Hepatol.* 11 (9), 821–834. doi: 10.1080/17474124.2017.1343143

Vaziri, N. D., Wong, J., Pahl, M., Piceno, Y. M., Yuan, J., DeSantis, T. Z., et al. (2013). Chronic kidney disease alters intestinal microbial flora. *Kidney Int.* 83 (2), 308–315. doi: 10.1038/ki.2012.345

Violi, F., Cammisotto, V., Bartimoccia, S., Pignatelli, P., Carnevale, R., and Nocella, C. (2023). Gut-derived low-grade endotoxaemia, atherothrombosis and cardiovascular disease. *Nat. Rev. Cardiol.* 20 (1), 24–37. doi: 10.1038/s41569-022-00737-2

Wang, Z., Zhang, S., Zheng, X., and Zhang, L. (2021). Efficacy and safety of colonic dialysis combined with traditional Chinese medicine retention enema in the treatment of chronic renal failure: A protocol for systematic review and meta-analysis. *Med.* (*Baltimore*) 100 (50), e28082. doi: 10.1097/md.00000000028082

Watanabe, H., Miyamoto, Y., Honda, D., Tanaka, H., Wu, Q., Endo, M., et al. (2013). p-Cresyl sulfate causes renal tubular cell damage by inducing oxidative stress by activation of NADPH oxidase. *Kidney Int.* 83 (4), 582–592. doi: 10.1038/ki.2012.448

Webster, A. C., Nagler, E. V., Morton, R. L., and Masson, P. (2017). Chronic kidney disease. Lancet 389 (10075), 1238–1252. doi: 10.1016/s0140-6736(16)32064-5

Wouk, N. (2021). End-stage renal disease: medical management. Am. Fam Phys. 104 (5), 493–499.

Wouters, O. J., O'Donoghue, D. J., Ritchie, J., Kanavos, P. G., and Narva, A. S. (2015). Early chronic kidney disease: diagnosis, management and models of care. *Nat. Rev. Nephrol.* 11 (8), 491–502. doi: 10.1038/nrneph.2015.85

Yang, H., Cao, S., Zhao, J., Lin, Y., Yang, B., Fan, S., et al. (2013). Study related to the improvement of survival quality and prevention of peritoneal fibrosis in patients with abdominal dialysis by supporting kidney granules. *Tianjin J. Traditional Chin. Med.* 30 (11), 659.

Yang, H., Lu, Y., Deng, Y., Fang, J., Liu, G., Jiang, A., et al. (2012a). A multicenter clinical study of Fukui Kidney Granules for evidence-based intervention in peritoneal dialysis patients. *Chin. J. Integrated Traditional West. Nephrol.* 13 (05), 410–412.

Yang, B., Ren, T., Li, J., Jiang, C., Xing, H., Zhang, L., et al. (2014). Clinical study on the improvement of malnutrition in peritoneal dialysis patients by supporting kidney granules. *Chin. J. Integrated Traditional West. Nephrol.* 15 (03), 234–236.

Yang, T., Richards, E. M., Pepine, C. J., and Raizada, M. K. (2018). The gut microbiota and the brain-gut-kidney axis in hypertension and chronic kidney disease. *Nat. Rev. Nephrol.* 14 (7), 442–456. doi: 10.1038/s41581-018-0018-2

Yang, B., Wang, M., Sun, L., Li, J., Qiao, Y., and Yang, H. (2021a). Effects of Kidney Support Formula on peritoneal function and VEGF, TNF- α , IL-12 and IFN- γ in uremic peritoneal dialysis rats. *Chin. J. Comp. Med.* 31 (04), 15–20.

Yang, B., Wang, M., Sun, L., Zhong, K., Li, J., and Yang, H. (2021b). Effects of Kidney Support Formula on peritoneal ultrafiltration function and VEGF-Notch signaling pathway in uremic peritoneal dialysis rats. *Tianjin Med. J.* 49 (07), 699–705.

Yang, B., and Yang, H. (2015). "The effect of kidney support granules on the nutritional status of elderly patients with chronic kidney disease stage 4," in 2015 Academic Annual Meeting of Renal Disease Specialized Committee of Chinese Society of Integrative Medicine and Western Medicine. 1194–1195.

Yang, H., Zhang, J., Zhao, J., and Yang, B. (2012b). Experimental study on the effect of kidney support granules on peritoneal dialysis-associated peritoneal fibrosis and its mechanism of action. *Chin. J. Integrated Traditional West. Nephrol.* 13 (06), 482–486.

Yoshifuji, A., Wakino, S., Irie, J., Tajima, T., Hasegawa, K., Kanda, T., et al. (2016). Gut Lactobacillus protects against the progression of renal damage by modulating the gut environment in rats. *Nephrol. Dial Transplant.* 31 (3), 401–412. doi: 10.1093/ndt/gfv353

Zhang, H. W., Lin, Z. X., Xu, C., Leung, C., and Chan, L. S. (2014). Astragalus (a traditional Chinese medicine) for treating chronic kidney disease. *Cochrane Database Syst. Rev.* 10), Cd008369. doi: 10.1002/14651858.CD008369.pub2

Zhang, L., Wang, F., Wang, L., Wang, W., Liu, B., Liu, J., et al. (2012). Prevalence of chronic kidney disease in China: a cross-sectional survey. *Lancet* 379 (9818), 815–822. doi: 10.1016/s0140-6736(12)60033-6

Zhang, L., and Yang, H. (2016). "Clinical study on the integrated treatment of chronic kidney disease stage 4 with traditional Chinese medicine," in 2016 Academic Annual Meeting of the Renal Disease Specialized Committee of the Chinese Society of Integrative Medicine. 347.

Zhang, L., and Yang, H. (2018a). Experimental study on the modulation of microinflammatory state by fusiform renal fluid in rats with chronic renal failure. *Liaoning J. Traditional Chin. Med.* 45 (08), 1740–1743. doi: 10.13192/j.issn.1000-1719.2018.08.059

Zhang, L., and Yang, H. (2018b). "Study on the inhibition of microinflammatory state in rats with chronic renal insufficiency model by Compound Kidney Supporting Liquid," in 2018 Academic Annual Meeting of Renal Disease Specialized Committee of Chinese Society of Integrative Medicine and Western Medicine. 428.

Zhao, X., Zhang, X., Li, M., and Wang, Y. (2022). Effects of Kidney Supporting Formula on renal pathology and expression of CTGF, FN and Vimentin in UUO rats. *Tianjin J. Traditional Chin. Med.* 39 (02), 232–237.

Zhou, X., Sun, X., Gong, X., Yang, Y., Chen, C., Shan, G., et al. (2017). Astragaloside IV from Astragalus membranaceus ameliorates renal interstitial fibrosis by inhibiting inflammation via TLR4/NF-κB in vivo and in vitro. Int. Immunopharmacol. 42, 18–24. doi: 10.1016/j.intimp.2016.11.006

Zhu, H., Cao, C., Wu, Z., Zhang, H., Sun, Z., Wang, M., et al. (2021). The probiotic L. casei Zhang slows the progression of acute and chronic kidney disease. *Cell Metab.* 33 (10), 1926–1942.e1928. doi: 10.1016/j.cmet.2021.06.014