



Toxin-Antitoxin Systems - A New Player in Morphological Transformation of Antibiotic-Exposed *Helicobacter pylori*?

Paweł Krzyżek*

Department of Microbiology, Faculty of Medicine, Wrocław Medical University, Wrocław, Poland

Keywords: *Helicobacter pylori*, toxin-antitoxin system, coccoid forms, morphological transformation, stress response

INTRODUCTION

The recently published original article by Mortaji et al. (2020) characterized for the first time the function of a type I toxin-antitoxin (TA) system in the gastric pathogen *Helicobacter pylori*. It was found that the high expression of an AapA1 toxin, which is part of this system, causes a drastic decrease in the amount of culturable *H. pylori* cells and their transformation from a spiral to a coccoid morphotype. It was also established that AapA1 is a hydrophobic peptide disrupting cell division and that oxidative stress is an inducer of the toxin expression.

The development of genomics and bioinformatics in recent years has contributed to the discovery of a high frequency of TA systems in microorganisms, which was a strong stimulus for the intensification of research on their structure and function (Lee and Lee, 2016; Yang and Walsh, 2017). Prokaryotic TA modules are genetic elements that encode information about a toxin involved in inhibiting growth of the bacterial producer and an antitoxin that counteracts the activity of the former. Toxins belonging to TA systems restrict microbial replication by targeting key processes for cell physiology, including replication, transcription, translation and/or cell wall synthesis (Harms et al., 2018). Attention is being paid increasingly to the participation of these systems in suppressing the microbial multiplication and the stimulatory effect on adaptation to stressful conditions, i.e., nutritional starvation, exposure to antimicrobial substances or immune system cells' attack (Lee and Lee, 2016; Yang and Walsh, 2017).

To date, five type II TA systems of *H. pylori* have been identified. These include chromosomally encoded HP0892-HP0893 (Han et al., 2013), HP0894-HP0895 (Han et al., 2011), HP0315-HP0316 (Kwon et al., 2012), and HP0967-HP0968 (Cárdenas-Mondragón et al., 2016), and the newly identified TfiT-TfiA (Boampong et al., 2020), which is encoded on mobile genetic fragments. The expression of toxins belonging to the above modules arrest the growth of bacterial producers and cause the reduction of their number (expressed in CFU/mL). Similar observations were made in 2017 by Arnion et al. (2017), who first identified the existence of the type I TA system in *H. pylori* (called AapA1-IsoA1), and noted that the expression of the toxin significantly decreases the amount of culturable *H. pylori* cells. At this point it is worth mentioning that Mortaji et al. (2020) deepened the knowledge related to the above phenomenon. They proved in their next original article that this decline was caused by a reduction in the culturability (observed as the optical density of the culture) but not the viability of *H. pylori* (preserved cell membrane integrity and a stable ATP level), and was

OPEN ACCESS

Edited by:

D. Scott Merrell,
Uniformed Services University,
United States

Reviewed by:

Timothy Cover,
Vanderbilt University, United States

*Correspondence:

Paweł Krzyżek
krojcerpawel@gmail.com

Specialty section:

This article was submitted to
Molecular Bacterial Pathogenesis,
a section of the journal
Frontiers in Cellular
and Infection Microbiology

Received: 22 February 2021

Accepted: 06 April 2021

Published: 26 April 2021

Citation:

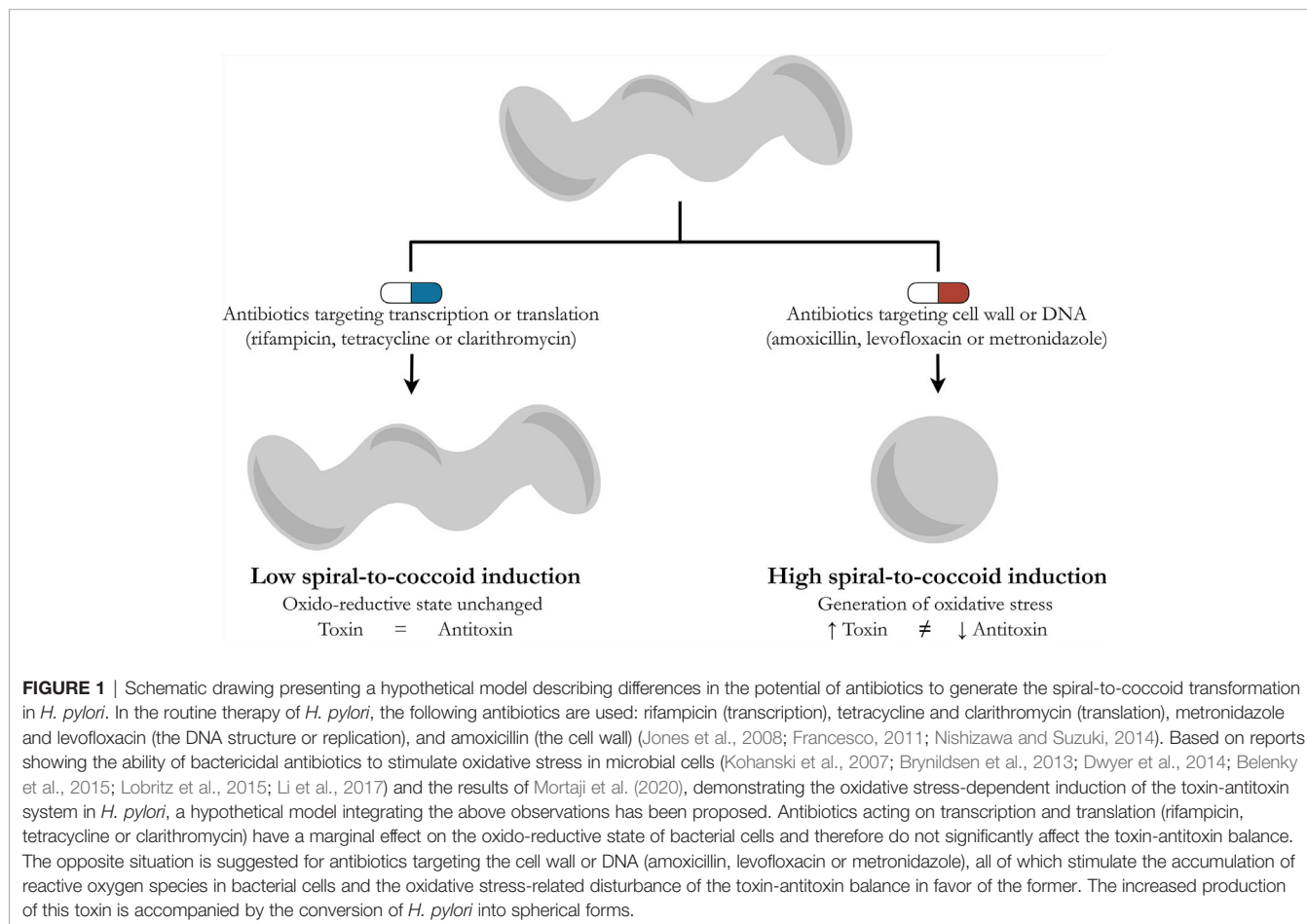
Krzyżek P (2021) Toxin-Antitoxin
Systems - A New Player
in Morphological Transformation
of Antibiotic-Exposed
Helicobacter pylori?
Front. Cell. Infect. Microbiol. 11:670677.
doi: 10.3389/fcimb.2021.670677

accompanied by the transition of morphology from spiral to coccoidal (Mortaji et al., 2020). This observation is very valuable from the scientific point of view and confirms the postulates presented by our research group, pointing to difficulties in the correct interpretation of the *H. pylori* viability (understood as the sum of various cell parameters suggesting its physiological activity) and frequent mistakes made by scientists taking the culturability (detected by culture optical density or CFU/mL) as the only determinant of the viability of this pathogen (Krzyżek and Grande, 2020).

An additional valuable cognitive element shown by Mortaji et al. (2020) was a proof that oxidative stress was an inducer of the *aapA1* expression in *H. pylori*, and thus a trigger for the spiral-to-coccoid transition. Exposure to high concentrations of oxygen, understood here as oxidative stress, is a well-known stress factor for *H. pylori* determining its intensive transformation into spherical forms (Chuang et al., 2005; Zeng et al., 2008). Thus, Mortaji et al. (2020) neatly revealed a possible molecular mechanism governing this process. In regard to this, it is also worth paying attention to the results presented by many research teams that have shown that bactericidal antibiotics, unlike bacteriostatic ones, stimulate the formation of oxygen free radicals and oxidative stress in bacterial cells, regardless of their target site (Kohanski et al., 2007; Brynildsen et al., 2013; Dwyer

et al., 2014; Belenky et al., 2015; Lobritz et al., 2015; Li et al., 2017). According to Lobritz et al. (2015), this effect was particularly visible with the use of antibiotics acting on the microbial cell wall and DNA, but neither translation nor transcription. The above information, in conjunction with the results provided by Mortaji et al. (2020), seem to be extremely interesting, as they may explain why bactericidal antibiotics (amoxicillin, levofloxacin or metronidazole) induce morphological transformation into spherical forms in *H. pylori* significantly faster than bacteriostatic antibiotics (Sörberg et al., 1997; Sörberg et al., 1998; Akada et al., 1999; Faghri et al., 2014; Krzyżek et al., 2019a; Krzyżek et al., 2019b). Still, it should be remembered that the process of cell death and/or formation of coccoids by *H. pylori* during the exposure to bactericidal antibiotics may depend on many factors simultaneously or be independent of oxidative stress.

In the original article by Mortaji et al. (2020), *H. pylori* was exposed to one of two antibiotics: rifampicin or tetracycline targeting transcription or translation, respectively. The authors did not observe any significant increase in the *aapA1* expression in rifampicin- or tetracycline-treated cells, concluding that exposure of *H. pylori* to antibiotics did not affect the expression of this toxin. In the light of the above presented deduction, however, it seems that divergent results may arise for



other antibiotics used in the therapy of *H. pylori*, especially those with a strong bactericidal activity, e.g., amoxicillin, levofloxacin or metronidazole. Extending research to include these antibiotics would allow it to be established whether the hypothesis presented by an author of this commentary about the inducing effect of bactericidal antibiotics and their oxidative stress-dependent generation of morphological transition into spherical forms by *H. pylori* is correct (Figure 1).

Finally, it is worth noting that the results presented by Mortaji et al. (2020) may have clinically significant implications, especially in the context of the eradication of difficult-to-treat, recurrent *H. pylori* infections. Recently, Morales-Espinosa et al. (2020) showed that the expression of HP0315, one of the components of the type II TA systems, is expressed significantly higher in intracellular *H. pylori* subpopulations and that the expression of this gene was accompanied by the formation of coccoid forms by these bacteria. Therefore, it seems very interesting to determine whether this type of relationship

can also be demonstrated for other TA modules, including AapA1-IsoA1, and whether lowering the expression of the toxin or increasing the expression of the antitoxin would positively influence the frequency of *H. pylori* eradication.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

FUNDING

The study was supported by the Wrocław Medical University grant No: SUB.A130.21.031. The funder had no role in the preparation of the manuscript.

REFERENCES

- Akada, J. K., Shirai, M., Fujii, K., Okita, K., and Nakazawa, T. (1999). In Vitro Anti-*Helicobacter pylori* Activities of New Rifamycin Derivatives, KRM-1648 and KRM-1657. *Antimicrob. Agents Chemother.* 43, 1072–1076. doi: 10.1128/aac.43.5.1072
- Arnion, H., Korkut, D. N., Gelo, S. M., Chabas, S., Reigner, J., Iost, I., et al. (2017). Mechanistic Insights Into Type I Toxin Antitoxin Systems in *Helicobacter pylori*: The Importance of mRNA Folding in Controlling Toxin Expression. *Nucleic Acids Res.* 45, 4782–4795. doi: 10.1093/nar/gkw1343
- Belenky, P., Ye, J. D., Porter, C. B. M., Cohen, N. R., Lobritz, M. A., Ferrante, T., et al. (2015). Bactericidal Antibiotics Induce Toxic Metabolic Perturbations That Lead to Cellular Damage. *Cell Rep.* 13, 968–980. doi: 10.1016/j.celrep.2015.09.059
- Boampong, K., Smith, S. L., and Delahay, R. M. (2020). Rapid Growth Inhibitory Activity of a YafQ-Family Endonuclease Toxin of the *Helicobacter pylori* Tfs4 Integrative and Conjugative Element. *Sci. Rep.* 10, 18171. doi: 10.1038/s41598-020-72063-x
- Brynildsen, M. P., Winkler, J. A., Spina, C. S., MacDonald, I. C., and Collins, J. J. (2013). Potentiating Antibacterial Activity by Predictably Enhancing Endogenous Microbial ROS Production. *Nat. Biotechnol.* 31, 160–165. doi: 10.1038/nbt.2458
- Cárdenas-Mondragón, M. G., Ares, M. A., Panunzi, L. G., Pacheco, S., Camorlinga-Ponce, M., Girón, J. A., et al. (2016). Transcriptional Profiling of Type II Toxin-Antitoxin Genes of *Helicobacter pylori* Under Different Environmental Conditions: Identification of HP0967-HP0968 System. *Front. Microbiol.* 7, 1872. doi: 10.3389/fmicb.2016.01872
- Chuang, M.-H., Wu, M.-S., Lin, J.-T., and Chiou, S.-H. (2005). Proteomic Analysis of Proteins Expressed by *Helicobacter pylori* Under Oxidative Stress. *Proteomics* 5, 3895–3901. doi: 10.1002/pmic.200401232
- Dwyer, D. J., Belenky, P. A., Yang, J. H., Cody MacDonald, I., Martell, J. D., Takahashi, N., et al. (2014). Antibiotics Induce Redox-Related Physiological Alterations as Part of Their Lethality. *Proc. Natl. Acad. Sci. U. S. A.* 111, E2100–E2109. doi: 10.1073/pnas.1401876111
- Faghri, J., Poursina, F., Moghim, S., Zarkesh Esfahani, H., Nasr Esfahani, B., Fazeli, H., et al. (2014). Morphological and Bactericidal Effects of Different Antibiotics on *Helicobacter pylori*. *Jundishapur. J. Microbiol.* 7, e8704. doi: 10.5812/jjm.8704
- Francesco, V. (2011). Mechanisms of *Helicobacter pylori* Antibiotic Resistance: An Updated Appraisal. *World J. Gastrointest. Pathophysiol.* 2, 41. doi: 10.4291/wjgp.v2.i3.35
- Han, K. D., Ahn, D. H., Lee, S. A., Min, Y. H., Kwon, A. R., Ahn, H. C., et al. (2013). Identification of Chromosomal HP0892-HP0893 Toxin-Antitoxin Proteins in *Helicobacter pylori* and Structural Elucidation of Their Protein-Protein Interaction. *J. Biol. Chem.* 288, 6004–6013. doi: 10.1074/jbc.M111.322784
- Han, K. D., Matsuura, A., Ahn, H. C., Kwon, A. R., Min, Y. H., Park, H. J., et al. (2011). Functional Identification of Toxin-Antitoxin Molecules From *Helicobacter pylori* 26695 and Structural Elucidation of the Molecular Interactions. *J. Biol. Chem.* 286, 4842–4853. doi: 10.1074/jbc.M109.097840
- Harms, A., Brodersen, D. E., Mitarai, N., and Gerdes, K. (2018). Toxins, Targets, and Triggers: An Overview of Toxin-Antitoxin Biology. *Mol. Cell* 70, 768–784. doi: 10.1016/j.molcel.2018.01.003
- Jones, K. R., Cha, J.-H., and Merrell, D. S. (2008). Who's Winning the War? Molecular Mechanisms of Antibiotic Resistance in *Helicobacter pylori*. *Curr. Drug Ther.* 3, 190–203. doi: 10.2174/157488508785747899
- Kohanski, M. A., Dwyer, D. J., Hayete, B., Lawrence, C. A., and Collins, J. J. (2007). A Common Mechanism of Cellular Death Induced by Bactericidal Antibiotics. *Cell* 130, 797–810. doi: 10.1016/j.cell.2007.06.049
- Krzyżek, P., Franciczek, R., Krzyżanowska, B., Łączmański, Ł., Migdał, P., and Gościński, G. (2019a). In Vitro Activity of 3-Bromopyruvate, an Anti-Cancer Compound, Against Antibiotic-Susceptible and Antibiotic-Resistant *Helicobacter pylori* Strains. *Cancers (Basel)* 11, 229. doi: 10.3390/cancers11020229
- Krzyżek, P., Franciczek, R., Krzyżanowska, B., Łączmański, Ł., Migdał, P., and Gościński, G. (2019b). In Vitro Activity of Sertraline, an Antidepressant, Against Antibiotic-Susceptible and Antibiotic-Resistant *Helicobacter pylori* Strains. *Pathogens* 8, 228. doi: 10.3390/pathogens8040228
- Krzyżek, P., and Grande, R. (2020). Transformation of *Helicobacter pylori* Into Coccoid Forms as a Challenge for Research Determining Activity of Antimicrobial Substances. *Pathogens* 9, 184. doi: 10.3390/pathogens9030184
- Kwon, A. R., Kim, J. H., Park, S. J., Lee, K. Y., Min, Y. H., Im, H., et al. (2012). Structural and Biochemical Characterization of HP0315 From *Helicobacter pylori* as a VapD Protein With an Endoribonuclease Activity. *Nucleic Acids Res.* 40, 4216–4228. doi: 10.1093/nar/gkr1305
- Lee, K. Y., and Lee, B. J. (2016). Structure, Biology, and Therapeutic Application of Toxin-Antitoxin Systems in Pathogenic Bacteria. *Toxins (Basel)* 8, 305. doi: 10.3390/toxins8100305
- Li, Z., Tan, J., Shao, L., Dong, X., Ye, R. D., and Chen, D. (2017). Selenium-Mediated Protection in Reversing the Sensitivity of Bacterium to the Bactericidal Antibiotics. *J. Trace Elem. Med. Biol.* 41, 23–31. doi: 10.1016/j.jtemb.2017.02.007
- Lobritz, M. A., Belenky, P., Porter, C. B. M., Gutierrez, A., Yang, J. H., Schwarz, E. G., et al. (2015). Antibiotic Efficacy is Linked to Bacterial Cellular Respiration. *Proc. Natl. Acad. Sci. U. S. A.* 112, 8173–8180. doi: 10.1073/pnas.1509743112
- Morales-Espinosa, R., Delgado, G., Serrano, L. R., Castillo, E., Santiago, C. A., Hernández-Castro, R., et al. (2020). High Expression of *Helicobacter pylori* VapD

- in Both the Intracellular Environment and Biopsies From Gastric Patients With Severity. *PLoS One* 15, e0230220. doi: 10.1371/journal.pone.0230220
- Mortaji, L., Tejada-Arranz, A., Rifflet, A., Boneca, I. G., Pehau-Arnaudet, G., Radicella, J. P., et al. (2020). A Peptide of a Type I Toxin-Antitoxin System Induces *Helicobacter pylori* Morphological Transformation From Spiral Shape to Cocci. *Proc. Natl. Acad. Sci. U. S. A.* 117, 31398–31409. doi: 10.1073/pnas.2016195117
- Nishizawa, T., and Suzuki, H. (2014). Mechanisms of *Helicobacter pylori* Antibiotic Resistance and Molecular Testing. *Front. Mol. Biosci.* 1:19. doi: 10.3389/fmolb.2014.00019
- Sörberg, M., Hanberger, H., Nilsson, M., Björkman, A., and Nilsson, L. E. (1998). Risk of Development of In Vitro Resistance to Amoxicillin, Clarithromycin, and Metronidazole in *Helicobacter pylori*. *Antimicrob. Agents Chemother.* 42, 1228. doi: 10.1128/AAC.42.5.1222
- Sörberg, M., Hanberger, H., Nilsson, M., and Nilsson, L. E. (1997). Pharmacodynamic Effects of Antibiotics and Acid Pump Inhibitors on *Helicobacter pylori*. *Antimicrob. Agents Chemother.* 41, 2218–2223. doi: 10.1128/aac.41.10.2218
- Yang, Q. E., and Walsh, T. R. (2017). Toxin-Antitoxin Systems and Their Role in Disseminating and Maintaining Antimicrobial Resistance. *FEMS Microbiol. Rev.* 41, 343–353. doi: 10.1093/femsre/fux006
- Zeng, H., Guo, G., Mao, X. H., De Tong, W., and Zou, Q. M. (2008). Proteomic Insights Into *Helicobacter pylori* Coccioid Forms Under Oxidative Stress. *Curr. Microbiol.* 57, 281–286. doi: 10.1007/s00284-008-9190-0

Conflict of Interest: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Krzyżek. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.