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Waste to Wealth: The power of food-waste anaerobic digestion integrated with lactic acid fermentation

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Food waste (FW) costs the global economy \$1 trillion annually and is associated with 8% of anthropogenic greenhouse gas emissions. Anaerobic digestion (AD) is an effective technology for recycling organic waste, including FW, for energy and nutrient recovery. Current major revenue streams for AD include the sale of biogas/power, gate fees, and digestate (fertiliser). However, subsidies provided by governments are a major profit driver for commercial facilities and are generally required for profitability, limiting its widespread adoption. Lactic acid (LA) is a high value intermediate of the AD process and literature evidence has indicated the recovery of LA can significantly boost the revenue generated from FW-AD. Moreover, FW fermentation naturally tends towards LA accumulation, promotion of LA producing bacteria, and inhibition of alternate competing microbes, making LA attractive for commercial production from FW. The integration of LA production and recovery into FW-AD could improve its economic performance and reduce the need for subsidy support, providing a platform for global adoption of the AD technology. However, challenges, such as 1) the low LA yield on FW, 2) seasonality of the FW composition, 3) unknown influence of LA recovery on downstream AD, and 4) impact of standard operational procedures for AD on upstream LA production, still exist making this focus area for future research. Even so, literature has shown the benefits of the LA-AD biorefinery, detailing improved process economics, increased FW utilisation, and elimination of subsidy support. Therefore, this review focuses on exploring the integrating LA production into AD by examining the current status of AD, LA integration strategies, challenges associated with LA production from FW, and identifies key challenges and considerations associated with downstream AD of fermented waste.

KEYWORDS

lactic acid, fermentation, anaerobic digestion, biorefinery, integration

Abbreviations: AD, anaerobic digestion; BMP, biochemical methane potential; CCR, carbon catabolite repression; FDP, fructose 1,6-DiPhosphate; FVW, fruit and vegetable waste; FW, food waste; GRAS, generally regarded as safe; KW, kitchen waste; LA, lactic acid; LAB, lactic acid bacteria; LDH, lactate DeHydrogenase; OLR, organic loading rate; OP, optical purity; PICRUST, phylogenetic investigation of communities by reconstruction of unobserved states; PLA, poly-lactic Acid; TS, total solids; VS, volatile solids; wt, weight.

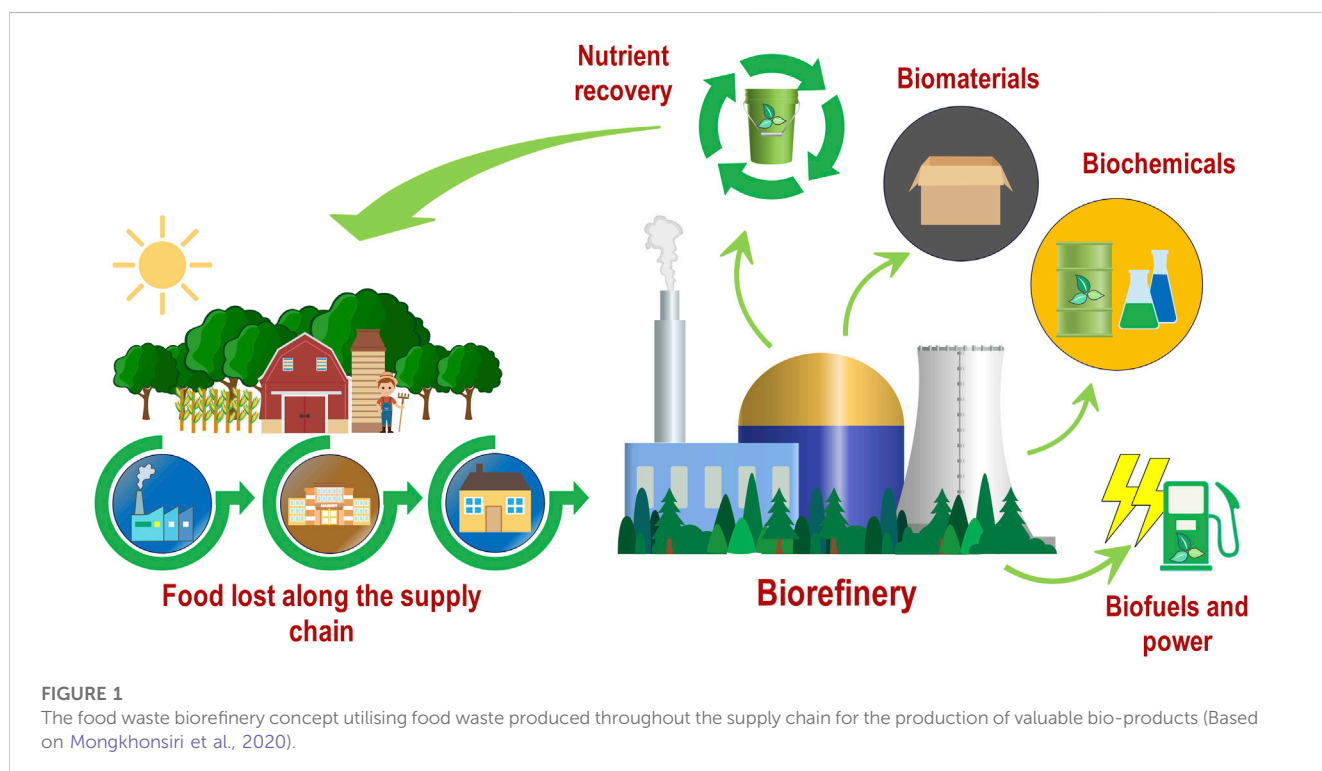
1 Introduction

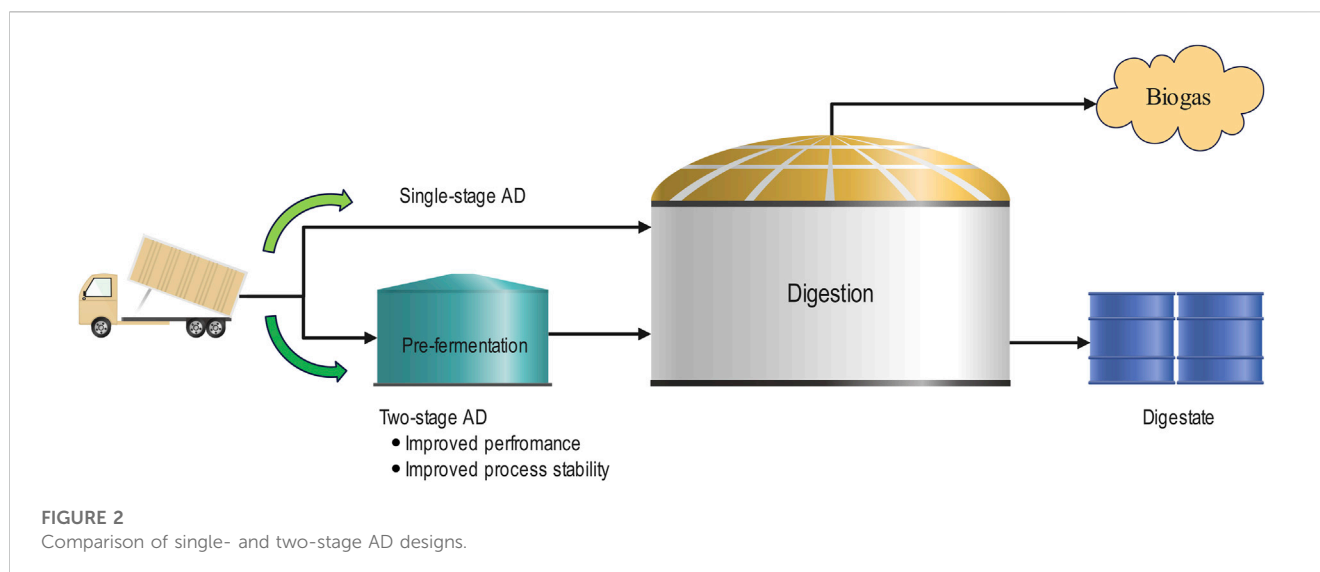
Recent estimates have valued cost of managing the annual 2.5 billion tonnes of globally produced FW at nearly \$230 billion (Safdie, 2023) and associated 8% of global anthropogenic greenhouse gas emissions with its production (WBA, 2018). FW encompasses a wide range of organic substrates, including dairy, fruit and vegetables, and distillery wastes produced along the whole supply chain (Kosseva, 2013), making its control a large scale complex issue. In an effort to reduce the FW production, the United Nations have set a goal to halve the production of FW *per capita* at the consumer and retail levels by 2030 (UN, 2015). Furthermore, the sustainable treatment of FW falls within multiple United Nations Sustainable Development Goals (UNSDGs), including the creation of sustainable production and consumption patterns, and taking action against climate change (UN, 2015). Consequently, the development of sustainable technologies for the recovery of value from this waste stream is becoming a focus point for modern research efforts.

Anaerobic digestion (AD) is a mature technology widely utilised for processing various organic wastes, including FW. In commercial applications, AD is commonly applied for processing large volumes of organic waste for the production of renewable energy (Comparetti et al., 2013). More recently, researchers have been investigating the applicability of AD on different waste streams, including lignocellulosic biomass, FW, sewage sludge, and animal manure (Carlsson et al., 2012; Mata-Alvarez et al., 2014; Sawatdeenarunat et al., 2015). In reference to the UNSDGs, the application of AD for FW recycling acts on numerous sustainability goals, including the promotion of sustainable agriculture (Goal 2), production of clean energy (Goal 6), development of sustainable cities (Goal 11),

ensuring sustainable production and consumption patterns (Goal 12), and combats climate change (Goal 13). However, the low economic value of the AD outputs (i.e., biogas and digestate (the liquid effluent)) and high cost of investment (capital expenditure), management (operational expenditure), and operation limit (e.g., biogas production) limit the financial feasibility of AD to farm-scale applications unless financial incentives are provided by governments (Gebrezgabher et al., 2010; Massaro et al., 2015). Alternatively, facilities may charge gate-fees for incoming waste streams, which has been shown to improve the financial feasibility of AD (Bastidas-Oyanedel and Schmidt, 2018), however, it is unrealistic to target gate-fees as a primary revenue driver as, not only can they vary significantly (Rolewicz-Kalińska et al., 2016), but competition between biogas plants could significantly reduce or eliminate gate-fees as a revenue stream. Consequently, there has been a growing body of literature aiming to value-add to the AD technology with additional technologies to recover additional high-value products (Kim et al., 2016; Choi et al., 2017; Demichelis et al., 2017; Chen et al., 2018; Demichelis et al., 2018; López et al., 2018; Sawatdeenarunat et al., 2018).

Biorefineries, analogous to oil refineries, produce a range of fuels, chemicals, and materials, but use biomass as their source material instead of fossil fuels (Figure 1). AD has the potential to produce a variety of high value biomaterials, biochemicals, and biofuels (Moraes et al., 2014; Sawatdeenarunat et al., 2016; Bastidas-Oyanedel and Schmidt, 2018) with lactic acid (LA) being of particular interest in recent research (Kim et al., 2016; Demichelis et al., 2017; Demichelis et al., 2018; Bühlmann et al., 2021; Bühlmann et al., 2022a; Bühlmann et al., 2022b). LA is a high-value chemical utilised in a variety of industries, including the food and pharmaceutical industries, with modern applications aiming to produce biodegradable





plastics (i.e., polylactic acid (PLA)) (Demichelis et al., 2017). Recent market estimates have forecast significant growth in the demand for LA over the coming years due to its applicability in multiple industries and the role it plays in the production of PLA (Bidddy et al., 2016; Castro-Aguirre et al., 2016; Alves de Oliveira et al., 2018; Ranjan and Baghel, 2018).

A growing body of recent key literature has explored the upstream fermentation of FW for LA followed by the downstream utilisation of the fermentation residues in AD for biogas production, aiming to generate greater financial feasibility (Kim et al., 2016; Demichelis et al., 2017; Bastidas-Oyanedel and Schmidt, 2018; Bühlmann et al., 2021; Bühlmann et al., 2022a; Bühlmann et al., 2022b). The two processes can be closely integrated to synergistically improve their performance. For example, AD can provide essential power and heat required to meet the energy needs of LA fermentation and downstream separation and recovery, while the residues from LA recovery can be utilised within FW-AD for biogas production (Figure 2). Furthermore, LA fermentation can improve the economic performance of the AD technology by providing an additional revenue stream. While Bühlmann et al. (2022a) reported a reduction in biogas yields, due to a reduced loading rate to digestion when recovering LA, an overall increase in revenue was identified. In fact, literature has suggested LA can boost revenue to the point where subsidy support is not required (Bastidas-Oyanedel and Schmidt, 2018). Moreover, LA recovery provides an opportunity to increase the overall plant capacity to account for the reduced organic loading to downstream AD, potentially further increasing overall plant revenue. However, challenges including 1) low LA yield on FW, 2) seasonality of FW composition within the commercial FW context leading to variable LA yields, 3) impact of LA recovery methods on downstream AD, and 4) microbial contamination, still exist and hinder the development of future FW LA-AD biorefineries. While reviews have explored the LA fermentation (Rawoof Salma Aathika et al., 2021; Song et al., 2022) and AD separately (Srisowmeya et al., 2020), no reviews, to the authors knowledge, have explored the integration challenges

between the two technologies. With this in mind, the following review explores the potential of integrating LA fermentation and recovery into FW-AD for improved profitability. The FW-AD context is examined from an industrial process perspective and the current status of LA fermentation and production is explored along with an examination of current key challenges in the LA-AD biorefinery context. Finally, important research prospects are outlined to identify critical areas that require investigation, which should lead to targeted research objectives.

2 Anaerobic digestion

AD is a complex multi-stage biological process which sequentially breaks down organic material, including a variety of high value intermediate metabolites, to its basic components (e.g., CO₂ and CH₄). AD has a long research history related to industrial applications, dating as far back as the 17th century when Van Helmont noted that decaying organic material produced flammable gases (Abbasi et al., 2012). Following over 300 years of development, and with the advancement of modern technology (including process control), it is possible to utilise AD for organic waste management (including FW) at large scale (Edwards et al., 2015). AD harnesses naturally occurring biological processes occurring in the absence of oxygen (i.e., anaerobic), which can be classified into four fundamental progressive stages, namely, hydrolysis, acidogenesis, acetogenesis, and methanogenesis. Each stage decomposes organic waste into progressively simpler metabolites (Melville et al., 2014; Sawatdeenarunat et al., 2015). Modern AD projects are generally developed to utilise one of two design modes (Figure 2); 1) Single-stage designs conduct all the above biological processes within a single vessel, and 2) two-stage designs aiming to split hydrolysis and acidogenesis (stage 1) from acetogenesis and methanogenesis (stage 2). In two-stage designs, stage 1 consists of a pre-fermentation vessel with a short hydraulic retention time (typically 2–3 days) prior to a much larger digester with much longer hydraulic retention time for the stage 2 digestion (typically 15–20 days) (Nabaterega et al., 2021). Several

modern AD facilities incorporate two-stage designs which allow stage 1 biological processes to be separately optimised from the stage 2 biological processes, yielding improved performance and stability (Nasr et al., 2012; Aslanzadeh et al., 2014; Schievano et al., 2014).

2.1 Current status and challenges

The development of new AD projects within developed and developing countries will depend upon the local waste feedstock availability and utilisation or value of biogas energy. Small-scale AD units are the most common in developing countries such as Bangladesh, India, and Nepal, with the biogas primarily utilised for cooking purposes (Zaman and Reynolds, 2015). The residuals following digestion are typically used for the fertilisation of crop soil and as fish feed in aquaculture through pond fertilisation and increased growth of organisms (e.g., microalgae, zooplankton) which fish consume, reducing demand on fish feed (Nhu et al., 2015; Sawatdeenarunat et al., 2016). In contrast, developed countries utilise AD for processing large volumes of organic waste while utilising the biogas for the production of power or, increasingly, compressed biomethane (Zhao et al., 2010; Comparetti et al., 2013; Hakawati et al., 2017). The operation of FW AD includes many challenges, such as inhibitor accumulation (e.g., volatile fatty acids, ammonium, etc.) and process instability (Li et al., 2018). However, the cost of developing new AD facilities and utilisation of the liquid effluent from AD can be major barriers to its adoption.

2.1.1 Financial incentives

A number of significant challenges complicate development of AD projects within developed countries, such as authorisation to connect to the electrical grid (as well as monetary fees), feedstock type and availability, physical construction, plant reliability, and the removal, management and costs of the liquid by-product, digestate (Insight, 2016). These challenges are primarily related to technical feasibility or acquiring approval from regulatory authorities. However, as a single digester can cost US\$ 953 per m³ of capacity (Chen et al., 2023), financing can also be a considerable challenge. AD also produces large volumes of effluent (e.g., digestate) which is low-value and can be a major cost burden to commercial facilities with transport (when used as an off-site fertiliser) and processing costs (primarily volume reduction) being substantial (O'Shea et al., 2022). Consequently, the success of commercial AD projects has heavily relied on subsidies (Gebrezgabher et al., 2010), limiting uptake of AD technology to date, because benefits and support provided by governmental policies can differ both internationally and nationally. For example, Europe is currently the pioneer of AD technology with 18,943 biogas plants in operation by the end of 2019 (EBA, 2021). However, the extensive implementation of AD around Europe is heavily driven by the financial subsidy policies provided by European governments to renewable energy producers and environmental regulations being strict but well developed, providing certainty to new projects (Linville et al., 2015; Vasco-Correa et al., 2018). In comparison, the Renewable Portfolio Standard in the US provides limited financial incentives for Anaerobic Digestion (AD). The subsidies tend to prioritize wind and solar energy over AD, which is biomass based. Moreover, these subsidies compete with the subsidies already given to fossil fuels (Edwards et al., 2015). Consequently, due to the limited financial support from governments, the financial viability of AD has been brought into question in the US, and as a result the adoption

of AD technology has been significantly slower, with approximately 2,200 AD plants currently in operation (Simet and Fletcher, 2017).

Australia is an example of virtually no uptake of the AD technology, having 242 known AD projects in operation, predominantly consisting of municipal sewage sludge digesters and landfill gas facilities (Tait et al., 2021). While government programs have aided the development of some AD projects for FW and abattoir waste processing (Wong, 2017), the lack of incentives within Australia to promote the development of the technology limit its widespread adoption. However, several policies do aim to support the development of new AD installations such as the Emission Reduction Fund (ERF) and the Renewable Energy Target (RET) scheme which aim to provide an additional revenue stream for facilities by generating and selling Carbon Credit Units, and Small-Scale Technology Certificates (STC) or Large-Scale Generation Certificates (LGC), depending on the size of the AD installation (STC < 100 kW < LGC) (Carlu et al., 2019). However, several barriers still harm the development of AD in Australia. ENEA, in collaboration with various Australian energy agencies and the Australian government, outlined several regulatory deficiencies which currently hinder the development of AD within Australia including; the absence of any national target for biogas production, financial uncertainties around power exports, and lack of uniform landfill levies among states (Carlu et al., 2019) which creates revenue uncertainty for new and existing AD projects.

2.1.2 Digestate

The liquid by-product produced from AD is commonly referred to as 'digestate' and, depending on the process and feedstock, can contain particulates in a wide range of 3.5%–13% (Plana and Noche, 2016). Generally, the solid fraction is composed of undigested material, such as lignin, while the liquid fraction contains mobile nutrients such as available nitrogen in the form of NH₄-N (approx. 1,000–5,000 ppm NH₄⁺-N) depending on the feedstock composition (Teglia et al., 2011; Akhbar, 2017; Coelho et al., 2018; Häfner et al., 2022). Depending on the feedstock composition, hydraulic retention time of the AD process, and process design (e.g., single- or two-stage), the characterisation of the solid and liquid fractions will vary (O'Connor et al., 2022).

To combat potential application of unsatisfactory digestate to land, standards, such as British standard "PAS110:2014", which is seen as the baseline quality specification for digestate, aim to ensure digestate is safe and reliable for land application (WRAP, 2017). While there are potential for digestate to displace synthetic fertilisers, digestate is generally seen as a cost-burden due to its bulky nature, dilute nutrient content (as compared to chemical fertilisers), and costs associated with its storage, transport, and land application (Turnley et al., 2016; Mickan et al., 2022). Even so, the use of digestate within agriculture has shown its benefit, improving plant growth and producing crop yields similar to synthetic nitrogen (Ren et al., 2020), and also with field application following soil injection (Riva et al., 2016; Zilio et al., 2021). However, FW derived digestate in the agricultural context still requires significant transport costs. Recently Mickan et al. (2022) investigated digestate as a nutrient base for potting media with positive responses, in terms of plant growth, whilst reducing the aforementioned transport constraints (Mickan et al., 2022). Furthermore, researchers have explored alternative uses for digestate, such as growing microalgae (Ayre et al., 2017; Chuka-ogwude et al.,

2022) and producing bioethanol (Sambusiti et al., 2016), with some researchers showing digestate could be a promising nutrient supplement and/or process water resource within the biotechnology industry to aid fermentation processes (Zhang et al., 2019; Ujor et al., 2020; Wang et al., 2021). Overall, while there are many potential use cases for digestate, no single solution is applicable in every commercial AD context, and sustained research efforts are required to continue exploring alternative end uses for AD digestate.

2.2 Anaerobic production of lactic acid

The sequential process by which AD breaks down biomass to biogas is complex and generates a variety of intermediate products, which if co-produced with biogas/power and digestate, could boost revenue from AD. LA is a promising high-value intermediate produced during the pre-fermentation stage of two-stage AD and has been the focus of a few recent studies (Kim et al., 2016; Bastidas-Oyanedel and Schmidt, 2018; Demichelis et al., 2018; Bühlmann et al., 2021; Bühlmann et al., 2022a; Bühlmann et al., 2022b; Chenebault et al., 2022). LA ($C_3H_6O_3$) is a three-carbon organic acid which is generally sold as an 88 wt% solution which is expected to experience significant market growth to 1,960 kton by 2024/2025 valued at USD 9.8 billion (Alves de Oliveira et al., 2018; Presswire, 2018). The price of LA generally follows the price of starch and sugar feedstocks, and recently has reached around 3.0–4.0 USD·kg⁻¹ (Alves de Oliveira et al., 2018). PLA (marketed as a compostable bioplastic) is anticipated to play a significant role in the expansion of the LA market with forecasts estimating, by 2025, 50% of all globally produced LA will be utilised for PLA production (Carcus, 2012; Djukić-Vuković et al., 2019), likely somewhat driven by the market for compostable plastics. The rapid growth of the LA market, high value of LA, and results of recent literature (Kim et al., 2016; Demichelis et al., 2017) suggest the development of the LA-AD biorefinery is a promising and potentially highly profitable method to improve the economic performance of AD.

Commercially, LA is primarily produced through fermentation with lactic acid bacteria (LAB) (Karp et al., 2011; Mazzoli et al., 2014; Juturu and Wu, 2016) utilising feedstocks consisting of pure sugars and nutrient-rich supplements (Hofvendahl and Hahn-Hägerdal, 2000; Abdel-Rahman et al., 2013). Both LA and feeds are commodity products, and the cost of the substrates heavily influences its market price (Reddy et al., 2016), resulting in the current production methods inflating the LA price and limiting its application for PLA production (Van Wouwe et al., 2016). Alternatively, waste feedstocks can reduce production costs by providing low-cost feedstock for LA production (Novik et al., 2017; Kwan et al., 2018). Furthermore, some waste streams, such as FW, contain nutrients that are essential for the growth of LAB and production of LA (Kim K. I. et al., 2003), reducing or potentially eliminating the need for nutrient supplements.

A large body of literature exists exploring LA fermentation and several reviews summarising the literature have been produced (Ghaffar et al., 2014; Alves de Oliveira et al., 2018; Eş et al., 2018). Fundamental research utilising waste streams for LA fermentation (Table 1) has identified that LA accumulation is promoted at low pH and short hydraulic retention time (HRT)

with the separation of hydrolysis and fermentation leading to further improved productivity and yield (Demichelis et al., 2017). However, conditions that promote LA production are generally avoided in AD because LA has a much lower pKa value (3.8) compared to other organic acids like acetic (4.76), propionic (4.87), and butyric acids (4.82), and LA and its degradation product, propionic acid, can inhibit methanogenesis (Bo et al., 2007; Tang et al., 2016; Gu et al., 2018). Consequently, as research generally aims to optimise biogas production (Khan et al., 2016), it is typically identified as a metabolic by-product that needs to be avoided.

While the conventional approach is to optimise AD for biogas production, recent literature has aimed to recover LA following fermentation (i.e., from the first-stage reactor) in two-stage AD processes. For example, Kim et al. (2016) and Demichelis et al. (2017) explored the recovery of LA from FW fermentation (simultaneous saccharification and fermentation) and its impacts on biogas production through subsequent AD of the solid fermentation residues. Both Kim et al. (2016) and Demichelis et al. (2017) reported a significant increase in value generated from FW processing (120%–180%), as compared to sole AD of FW, and noted the solid residues were suitable for methane production. A similar study by Bühlmann et al. (2022a) reported that, in addition to the solid fermentation residues, the liquid extraction residues following LA recovery (via ion exchange) were suitable for methane production and retained a significant (though 21% lower than raw feed) methane potential. While an overall net change in value generated from FW was not provided, the authors outlined a net 622 USD·day⁻¹ increase in relative value (i.e., difference in estimated revenue from LA sales and loss due to reduced biogas production from LA recovery) when implementing LA recovery within a commercial FW-AD context. Overall, the findings from the available literature indicates coupling LA fermentation with subsequent extraction of LA with FW-AD can increase revenue generated from FW processing while having a minor impact on AD biogas production.

3 Integration of lactic acid fermentation into anaerobic digestion

Proper integration of biorefinery technologies for LA production and recovery with AD requires the understanding of not only the individual fermentation and biogasification processes, but also the interaction between the two processes. Control of LA fermentation and utilisation of certain separation technologies may lead to unwanted effects on AD, while operation of AD may unintentionally impact LA fermentation (depending on the integration strategy). Therefore, it is important to examine the standard operation of the two processes and develop strategies to ensure optimal performance of fermentation and AD within an LA-AD biorefinery context.

3.1 Process integration

LA fermentation is most commonly conducted with highly fermentable carbohydrates such as glucose and sucrose obtained

TABLE 1 Summary of key research achievements from some researchers regarding the production of LA from fermentation in two-stage AD.

Feed	Operation mode	Aim	Key findings	References
KW ^c	Semi-continuous	Evaluate the influence of LA on methanogenesis	<ul style="list-style-type: none"> • LA is the main KW fermentation product • Propionic acid concentration in the methanogenic effluent is nearly linearly proportional to rate of LA loaded into the methane reactor • LA in the methanogenic influent should be avoided to improve two-phase AD performance 	Bo et al. (2007)
Simulated FVW ^b	Continuous	Operate the fermenter at pH 4.0 for LA production for its subsequent utilisation for methane production	<ul style="list-style-type: none"> • Low pH (4.0) promoted <i>Lactobacillus</i> • Low hydrogen partial pressure promoted LA degradation 	Wu et al. (2016)
FW	Semi-continuous	Co-produce LA and Biogas	<ul style="list-style-type: none"> • LA is predominantly produced at higher temperatures (>50°C) • Short HRTs (~1 day) promote LA production • 47 kg of LA and 54 m³ can be produced which has more than twice the economic value than that of conventional AD 	Kim et al. (2016)
Maize silage	Semi-continuous LBR for fermentation, CSTR for digestion	Examine the interrelationship between microbial communities and process parameters	<ul style="list-style-type: none"> • Fermentation alternates between LA production and gas production (hydrogen) • Alternating periods led to large changes in the microbial community structure • The biogas composition followed the metabolic dynamics of the fermentation phase • Stabilisation of the fermentation phase can lead to enhanced productivity for chemical and bioenergy production • LA played a key role in the presence of instabilities • Microbial chain elongation mechanisms may have played a role in fermentation instability 	Sträuber et al. (2016)
FW	Batch	Investigate the effects of organic loading rate, pH, and temperature on LA production	<ul style="list-style-type: none"> • Hydrolysis rate increased with pH • FW serves as an inoculum for LA fermentation • High temperatures favoured hydrolysis but reduced the rate of acidification • High OLR^d leads to systems instability, low LA yield, low VS destruction 	Tang et al. (2016)
FW	-	Increase LA and Biogas production by separating enzymatic hydrolysis and fermentation	<ul style="list-style-type: none"> • LA productivity and yield improved when separating hydrolysis and fermentation • Biogas yield improved by separating hydrolysis and fermentation 	Demichelis et al. (2017)
Simulated FW	Batch and semi-continuous LBR ^a	Reduce LA production from FW fermentation	<ul style="list-style-type: none"> • Acidic conditions promote (3.5–4.5) <i>Lactobacillus</i> (A key microorganism in LA production) • Short HRTs promote high LA concentrations • Batch fermentation promoted LA accumulation • Organic acid production is promoted at slightly acidic pH (5–6) 	Gu et al. (2018)
FW	Simulation	A techno-economic evaluation of different scenarios for LA and biogas co-production	<ul style="list-style-type: none"> • Conversion of FW into LA and biogas is economically feasible • Integrated biorefinery was more cost effective than sole production of either LA or biogas • Integrated biorefinery reduced waste generated, AD digester volume, minimised energy demand, and enhanced production of valuable products 	Demichelis et al. (2018)

a: Leach Bed Reactor, b: Fruit and Vegetable Waste, c: Kitchen Waste, d: Organic Loading Rate.

from corn, sugarcane, and cassava commonly used commercial substrates (Olszewska-Widrat et al., 2020). In contrast, the complex community which drives AD is capable of breaking down complex substrates for biogas production. In a FW context, FW contains many of the essential nutrients and carbon sources required for LA production (Kim et al., 2016; Tang et al., 2016; Pleissner et al., 2017), and thus is suitable for direct fermentation. Therefore, upstream fermentation for LA and downstream utilisation of fermentation residues for methane production is considered the most appropriate method to integrate the two

processes. This integration approach is beneficial for two primary reasons, 1) the highly available sugars and nutrients within FW may be directly utilised for LA production while the more complex substrates are retained for AD, 2) modern AD designs incorporate a two-stage design, with an upstream fermenter installed to separate hydrolysis/acidogenesis with methane formation (Section 2.0). This provides an opportunity for LA fermentation and recovery by integrating into existing facilities through retrofitting an LA separation and recovery into the first stage of the widely adopted two stage AD design process (Figure 3).

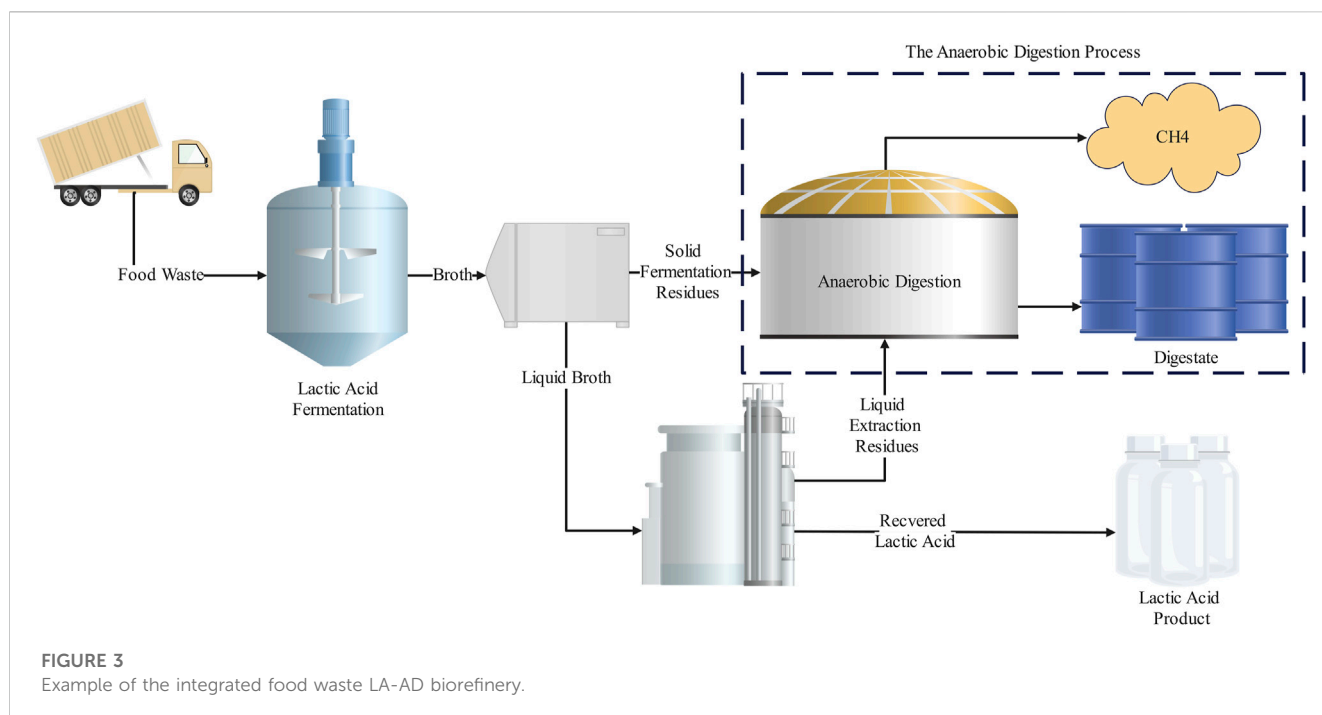


FIGURE 3
Example of the integrated food waste LA-AD biorefinery.

While this is a promising integration method, it is important to understand that the commercial FW context is highly variable and has yet to be fully explored for LA production. A recent study has shown the availability of various feedstocks are dependent on complex market forces, which can impact LA yields (Bühlmann et al., 2021). However, Bühlmann et al. (2021) reported high yields for an unoptimized FW fermenter and noted a surprising stability in LA was observed, suggesting commercial LA production from complex highly variable FW is technically feasible.

3.2 Lactic acid fermentation

Many LA producing microorganisms exist, including bacteria, filamentous fungi, and yeast. However, lactic acid bacteria (LAB) are generally utilised as they are 1) Generally Regarded As Safe (GRAS), with the exception of some pathogenic streptococci strains, 2) robust organisms already adapted to the stress of industrial processes, 3) are capable of metabolising numerous mono- and di-saccharides, 4) fast growing, and 5) produce a variety of high value metabolites (Mazzoli et al., 2014; Juturu and Wu, 2016).

Many bacteria produce LA either as a primary or secondary product, but those labelled as “LAB” are exclusively grouped in the order *Lactobacillales* which include; *Lactobacillus*, *Pediococcus*, *Carnobacterium*, *Aerococcus*, *Vagococcus*, *Enterococcus*, *Teragenococcus*, *Leuconostoc*, *Weissella*, *Streptococcus*, *Oenococcus*, and *Lactococcus* (Juturu and Wu, 2016). Bacteria outside of this group have been utilised for LA production (Table 2), but most commercialised LAB fall into the genus *Lactobacillus* as they are tolerant to acidic conditions and can be easily engineered to selectively produce LA; however, other organisms applied include *Streptococcus* and *Pediococcus* (Lee, 2015; Biddy et al., 2016).

The optimal conditions for LA production vary depending on the microorganism utilised as LAB can grow in the pH range of

3.5–10 and temperature range of 5–45°C (Abdel-Rahman et al., 2013) and at even higher temperatures in some cases (Table 2). However, most LA fermentation processes are conducted at a pH ranging from 5.0–6.0 and at mesophilic temperatures (Table 2). For continuous processes, short hydraulic retention times (HRTs) are generally preferred as they tend to promote LA accumulation (Komemoto et al., 2009; Tang et al., 2016). However, a universal value for the optimal HRT is difficult to determine as it varies depending on various factors, especially on fermenter design and operation mode, FW composition, and operational parameters (Kim M. et al., 2003; Tang et al., 2016; Gu et al., 2018). Similarly, organic loading rates (OLRs) utilised for LA production vary (Kim et al., 2012; Tang et al., 2016; Luongo et al., 2019) depending on similar process factors, but high OLRs generally favour the production of LA and assist in limiting the conversion of LA to other organic acids (Bo & Pin-jing, 2014; Luongo et al., 2019).

The presence of a chiral carbon in LAs chemical structure can complicate fermentation as different bacteria can produce L- and D-LA (the two isomers of LA) in varying quantities depending on their production of L-lactate dehydrogenase (L-LDH) or D-LDH (Garvie, 1980; Liu, 2003; Eiteman and Ramalingam, 2015), or even depending on the use of alternative metabolic pathways such as the methylglyoxal bypass which produces a racemic LA mixture (Mazumdar et al., 2013). A mixture of L- and D-LA can be troublesome as the optical purity (OP; ratio of L-LA to the total LA present) affects its applicability, particularly if it is to be used for the synthesis of PLA (OP >98%) (Gandolfi et al., 2015) as several properties including crystallinity and thermal stability are impacted by the relative quantities of L- and D-LA within the polymer blend (Eiteman and Ramalingam, 2015). Furthermore, certain industries may require a specific isomeric form, such as the food and pharmaceutical industries which generally require L-LA as D-LA, in high dosages, can be harmful to humans (Alves de Oliveira et al., 2018). As the food industry demands the majority of the LA

TABLE 2 Operating Conditions used for LA fermentation by some researchers (adopted from (Alves de Oliveira et al., 2018))

Dominant LAB	Product	Substrate	Temperature (°C)	Operation mode	pH control	LA productivity (g L ⁻¹ h ⁻¹)	Yield (g·g ⁻¹)
<i>Sporolactobacillus</i> sp. CASD	D-LA	Glucose	42	Fed-batch	5.0–6.0 CaCO ₃	4.4	0.84
<i>Bacillus</i> sp. WL-S20	L-LA	Glucose	45	Fed-batch	9.0 NaOH	1.04	0.993
<i>Bacillus coagulans</i> C106	L-LA	Xylose	50	Fed batch	6.0 Ca(OH) ₂	4	0.95
<i>Lactobacillus paracasei</i> 7BL (GMO)	L-LA	Glucose	37	Fed-batch	6.0 CaCO ₃	1.79	0.99
		Wood chips				2.25	0.96
		Rice straw				5.27	0.97
<i>Lactobacillus rhamnosus</i>	L-LA	Defatted rice bran	42	Fed-batch	6.25 Ca(OH) ₂	2.56	0.937
<i>Lactococcus lactis</i> ATCC19435	L-LA	Jerusalem artichoke	30	Fed-batch	6.0 NaOH	–	–
<i>Lactobacillus casei</i> G-02	L-LA	Jerusalem artichoke	40	Fed-batch (SSF)	6.5 CaCO ₃	4.7	0.963
<i>Bacillus coagulans</i> XZL4	L-LA	Hemp hurds (Glucose–xylose)	50	Batch	5.5 CaCO ₃	–	0.900 0.840
<i>Bacillus coagulans</i> LA204	LA	Pretreated corncob	50	Fed- batch	6.0 NaOH	1.37	0.77
				(SSF)			
<i>Lactobacillus agilis</i> LPB 56	L-LA	Soybean vinasse	30	Batch	6.0 Ca(OH) ₂	0.86	0.849
<i>Bacillus coagulans</i> LA1507	L-LA	Sweet sorghum bagasse	50	Open-fed-batch (SSF)	5.2–6.2 Ca(OH) ₂	1.59	0.437
<i>Pediococcus acidilactici</i> (GMO)	LA	Detoxified corn stover (Cellulose)	45	Batch (SSF)	5.5 NaOH	1.45	0.715
<i>Lactobacillus plantarum</i> (GMO)	D-LA	Delignified hardwood pulp	37	Batch (SSF)	6.0 NaOH	2.29	0.879
<i>Lactobacillus paracasei</i>	D-LA	<i>Curcuma longa</i> waste	34	Batch (SSF)	6.0 NH ₄ OH	2.08	0.69
<i>Lactobacillus coryniformis</i>	L-LA		37			2.7	0.65
<i>Lactobacillus pentosus</i>	LA	Corn stover	37	Fed-batch (SSF)	6	1.92	0.66
<i>Bacillus coagulans</i>]I12	L-LA	oil palm empty fruit bunch hydrolysate	50	Batch (SSF)	6.0 Ca(OH) ₂	3.4	–
<i>Lactobacillus rhamnosus</i> ATCC7469	LA	Recycled paper sludge	37	Batch (SSF)	5.5 CaCO ₃	2.9	0.97
<i>Streptococcus</i> sp	LA and biogas	FW	35	Batch	6	3.38	0.33
<i>Streptococcus</i> sp	LA	FW	35	Batch (SSF)	6.0 NaOH	2.16	0.81
<i>Rhizopus oryzae</i>	L-LA	Xylo-oligosaccharides manufacturing waste	40	Batch (SSF)	5.5 CaCO ₃	1	0.6
<i>Bacillus coagulans</i>	L-LA	Coffee mucilage	52	Batch	6.0 NaOH	4.4	0.77
<i>Bacillus coagulans</i>	L-LA	Coffee pulp	52	Batch	6.0 NaOH	4.02	0.78
<i>Lactobacillus paracasei</i> KM2 (GMO)	L-LA	Whole slurry of oil palm trunk	37	Batch	6.0 NH ₄ OH	–	0.895
<i>Geobacillus stearothermophilus</i> DSM494	L-LA	Raw potato starch	60	Batch	7.0 NaOH	1.8	0.66
<i>Lactobacillus delbrueckii</i>	LA, xylitol, activated carbon and phenolic acids	Brewer's spent grains (Cellulose)	37	Batch	6.0 NaOH	0.59	0.99
<i>Lactobacillus pentosus</i> DSM20314	LA Hetero-fermentation	Wheat bran	30	Batch	6.3 NaOH	0.3	0.73

(Continued on following page)

TABLE 2 (Continued) Operating Conditions used for LA fermentation by some researchers (adopted from (Alves de Oliveira et al., 2018))

Dominant LAB	Product	Substrate	Temperature (°C)	Operation mode	pH control	LA productivity (g L ⁻¹ h ⁻¹)	Yield (g·g ⁻¹)
<i>Lactobacillus delbrueckii</i> NBRC 3202	D-LA	Cassava fibrous waste	37	Batch	6.5 NaOH	0.9	0.5
<i>Lactobacillus casei</i> 12A	LA	De-oiled algal biomass + glucose	37	Batch	6.5	-	-

produced (~85%) (Ahmad et al., 2020), L-LA is generally the target for fermentation. Studies have shown the OP can be controlled through the manipulation of environmental conditions (Gu et al., 2014; Zhang et al., 2017), supplementation of nutrients (Zhang et al., 2020b), co-fermentation (Ma et al., 2021), or utilisation of specific LAB strains (Yuan et al., 2018; Alexandri et al., 2019). However, the response of OP with changes in fermentation conditions is not consistent between studies and may be related to the differing microbial communities which form during mixed-culture fermentation, different pure bacteria cultures, or differences in dominant metabolic pathways utilised for LA.

3.2.1 Pre-treatment

Pre-treatment processes, which may be classified into biological, chemical and physical processes, are primarily designed to improve hydrolysis of complex biomass and increase the release of macromolecular substances (Yang et al., 2013; Wang et al., 2022). Many pre-treatment methods are available including extrusion, alkali pre-treatment, organosolv (an organic solvent-based treatment), and ammonia fibre expansion (AFEX) (Table 3). As the composition of wastes vary significantly, there can be no universal pre-treatment method for every feedstock. However, it has been suggested that the selected pre-treatment method should aim to, 1) avoid the need for size reduction of biomass particles, 2) preserve the hemicellulose fraction for lignocellulosic biomass, 3) reduce/remove inhibitory components and minimise their formation, 4) improve accessibility to difficult components within the biomass, 5) minimise power consumption, 6) improve the properties of the biomass surface for improved microbial interactions, 7) improve the hydrolysis rate of lipids and proteins, and 8) utilise a low cost catalyst/method for recycling of the catalyst and regeneration of lignin for co-product production (for lignocellulose) (Kumar and Sharma, 2017; Parthiba Karthikeyan et al., 2018). The majority of pre-treatment methods aim to improve the biodegradability of agricultural residues (Table 3) due to the presence of lignin, which is a major barrier to the enzymatic saccharification (Xu et al., 2016), or to aid in the breakdown of polysaccharides for microbes which may not produce the required enzymes for saccharification. However, most of the available pre-treatment methods are yet to be commercialised due to the high cost of biomass pre-treatment, and many do not meet the requirements for commercial application (Xu et al., 2016).

As LAB struggle to ferment substrates composing of complex polymers, pre-treatment processes have shown their benefit, for example, with pulp mill residue (de Oliveira Moraes et al., 2016), corn stover (Ahring et al., 2016), and sugarcane bagasse (Wischrall et al., 2019). Though FW is inherently relatively

biodegradable, having been shown to produce substantial LA from direct fermentation (e.g., 0.46 gLA·gTS⁻¹ (Tang et al., 2016), 0.18 gLA·gTS⁻¹ (high TS content) (Yousuf et al., 2018), 0.55 gLA·gVS⁻¹ (Bühlmann et al., 2022b), and 0.42 gLA·gVS⁻¹ (Wang et al., 2021)), application of pre-treatment methods to FW have been shown to benefit LA fermentation by improving the final LA yield (Kim K. I. et al., 2003; Kwan et al., 2016; Pleissner et al., 2016; Demichelis et al., 2017; Ahmad et al., 2020).

3.2.2 Metabolic considerations

A number of different metabolic pathways can be utilised by bacteria for LA production, however, LAB are primarily described as utilising the glycolytic pathway (Embden–Meyerhof–Parnas pathway; Figure 4), or the phosphoketolase pathway for homo or hetero-fermentation, respectively (Wang et al., 2015). Homo-fermentative LAB produce LA as their primary end product (Papagianni, 2012; Wang et al., 2015), while hetero-fermentative LAB metabolise pentose producing equimolar amounts of LA, carbon dioxide, and ethanol or acetate (Wang et al., 2015). The specific pathway utilised by LAB is determined at the family level (Holzapfel and Wood, 2014), with crucial differences between these pathways provided by the presence of key enzymes utilised in each route; fructose 1,6-diphosphate (FDP) aldolase and phosphoketolase for the glycolytic and phosphoketolase pathways, respectively (Alves de Oliveira et al., 2018). Homo-fermentative LAB are generally preferred for LA production due to high production of LA with minimal by-products.

Some LAB resemble obligatory homo-fermentative bacteria as they possess the capability to produce FDP aldolase, however, others are also capable of synthesising phosphoketolase, allowing hexose and pentose to be utilised via the above mentioned pathways (Salminen et al., 2004; Holzapfel and Wood, 2014). These LAB are known as facultative hetero-fermentative LAB. In this case, the behaviour of these LAB (i.e., homo- or hetero-fermentation) are determined by the available carbon source or from certain environmental factors such as nutritional, osmotic, and/or thermal stress (Alves de Oliveira et al., 2018). LA production through the Embden–Meyerhof–Parnas pathway is preferable as LA is the only product, maximising its yield on biomass. However, overall community metabolic pathways for LA production are influenced by environmental conditions, operating parameters, and feed composition (Papagianni, 2012; Holzapfel and Wood, 2014). Consequently, it may be useful to monitor metabolic pathways present to identify potential undesired shifts in the pathways. A variety of molecular methods are available for monitoring metabolic pathways during LA fermentation such as shotgun metabolomics, lipidomics, metagenomics, and predictive

TABLE 3 Summary of some of pre-treatment methods for the treatment of various feedstocks (Modified from (Menon and Rao, 2012; Capolupo and Faraco, 2016))

Pre-treatment method (classification)	Feed stock	Sugar yield	Advantages	Disadvantages	Commercial standing
Acidic (Chemical)	<ul style="list-style-type: none"> • Corn stover, spruce, polar, and switchgrass 	-	<ul style="list-style-type: none"> • Hydrolyses hemicellulose and cellulose and alters the lignin structure 	<ul style="list-style-type: none"> • Hazardous, toxic, and corrosive chemicals used • High cost • Gypsum formation during neutralisation • Inhibitory by-product formation 	Commercially implemented. Generally dilute sulphuric acid is utilised (Menon and Rao, 2012)
Alkali (Chemical)	<ul style="list-style-type: none"> • Lignocellulosic biomass (Corn stover, bagasse, wheat straw, rice straw, and switchgrass) 	-	<ul style="list-style-type: none"> • Removes lignin and hemicellulose. Increases the accessible surface area 	<ul style="list-style-type: none"> • Long residence time and irrecoverable salts produced 	- ^c
Liquid hot water (Physico-chemical)	<ul style="list-style-type: none"> • Agricultural residues (sugarcane bagasse, corn stover, wheat straw, and sunflower stalks) • MSW 	80%–94% reducing sugars	<ul style="list-style-type: none"> • Removal of cellulose making enzymes • Recovery of almost pure hemicellulose • No catalyst or other chemicals • Hydrolysis of hemicellulose • No need for size reduction • High sugar recovery • Low formation of inhibitors 	<ul style="list-style-type: none"> • Long residence time, lower removal of lignin • High energy demand • Remaining solids will need to be processed 	Demonstration plant (Zheng and Rehmann, 2014)
Organosolv (Chemical)	<ul style="list-style-type: none"> • Agricultural residues (Wheat straw, sugarcane bagasse) 	Up to 60% of reducing sugars	<ul style="list-style-type: none"> • Hydrolysis of lignin and hemicellulose • Pure lignin removal as by-product 	<ul style="list-style-type: none"> • A condenser is required for solvent recovery • Costly process 	- ^c
Ozonolysis (Chemical)	<ul style="list-style-type: none"> • Agricultural residues (wheat straw, bagasse, and peanut and poplar sawdust) 	45%–90% reducing sugars	<ul style="list-style-type: none"> • Reduces lignin content while not producing toxic residues • Moderate reaction conditions • Efficient lignin degradation 	<ul style="list-style-type: none"> • Large quantities of ozone are needed • Costly process 	- ^c
CO ₂ Explosion (Physico-chemical)	<ul style="list-style-type: none"> • Agricultural residues (Wheat straw, sugarcane bagasse) 	Up to 90% of reducing sugars	<ul style="list-style-type: none"> • Hemicellulose removal, cellulose decrystallization, cost-effective • Increases accessible surface area • Does not imply toxic chemical generation 	<ul style="list-style-type: none"> • Does not modify the lignin structure • Costly process • Very high pressures required 	- ^c
Steam Explosion (Physico-chemical)	<ul style="list-style-type: none"> • Agricultural residues (corn stalk, wheat straw, and sugarcane) • MSW • Hardwood • Forest residues 	50%–70% reducing sugars	<ul style="list-style-type: none"> • Hemicellulose removal and alters the lignin structure • Good sugar recovery • Low cost • Less hazardous process 	<ul style="list-style-type: none"> • Incomplete destruction of the lignin-carbohydrate complex • Partial hemicellulose degradation • Generation of inhibitor compounds 	Demonstrated at commercial scale at the Masonite plants (Menon and Rao, 2012) Commercialised for ethanol production (Zheng and Rehmann, 2014)
AFEX ^a (Physico-chemical)	<ul style="list-style-type: none"> • Agricultural residues (wheat straw, corn stover, bagasse, and rice straw) • MSW 	Up to 80%–90% of reducing sugars	<ul style="list-style-type: none"> • Removes lignin and hemicellulose • Low formation of inhibitor compounds • Moderate process conditions 	<ul style="list-style-type: none"> • Not efficient for biomass with a high lignin content • High-cost process 	- ^c
Ionic Liquids (Chemical)	<ul style="list-style-type: none"> • Agricultural residues (Wheat straw, sugarcane bagasse, peanut and poplar sawdust, and corn stover) 	60%–85% reducing sugars	<ul style="list-style-type: none"> • Dissolution of cellulose and increases the amenability to cellulase 	<ul style="list-style-type: none"> • In early stages of development • Chemicals are expensive • Solutions are viscous and difficult to handle 	- ^c

(Continued on following page)

TABLE 3 (Continued) Summary of some of pre-treatment methods for the treatment of various feedstocks (Modified from (Menon and Rao, 2012; Capolupo and Faraco, 2016))

Pre-treatment method (classification)	Feed stock	Sugar yield	Advantages	Disadvantages	Commercial standing
Biological (Biological)	<ul style="list-style-type: none"> • Agricultural residues (Wheat straw, rice straw) • Soft wood 	20%–50% reducing sugars	<ul style="list-style-type: none"> • Low energy input • Moderate reactor conditions • No catalyst or chemical additives • Do not imply toxic chemical generation • Low cost 	<ul style="list-style-type: none"> • Low hydrolysis rate • Large area required 	Unlikely to commercialised (Menon and Rao, 2012)
Extrusion (Physical)	<ul style="list-style-type: none"> • Agricultural residues (Rice straw, wheat straw, and corn stover) 	50%–75% reducing sugars	<ul style="list-style-type: none"> • Moderate temperatures • Good sugar yields • High flexibility for many process modifications • Less hazardous process 	<ul style="list-style-type: none"> • Partial hemicellulose degradation • Generation of inhibitor compounds • Incomplete destruction of lignin-carbohydrate matrix 	Systems are already commercially available (Capolupo and Faraco, 2016) Commercialised counter current extrusion reactor (Zheng and Rehmann, 2014)

a: ammonia Fibre Explosion/Expansion, b: the method can be applied to different feed stocks, c: Commercial implementation not found.

techniques based on DNA amplicon sequencing (e.g., PICRSU, PICRUSt2, Tax4Fun, and Faprotax). Each differs in its approach, and associated advantages and disadvantages (Table 4). In-silico gene inference techniques stand out compared to the other three methods as, instead of directly measuring components within sample, taxonomic compositions, which are inferred from amplicon sequencing, are used to predict microbial functional genes (Sun et al., 2020). The major advantage of *in silico* techniques is their low cost compared to others which fully sequence the genome. For example, the cost of PICRUSt analyses can be 5–15 times lower than shotgun metagenomics (Mukherjee et al., 2017). However, these techniques are limited to the genomes listed in utilised databases, which are currently highly biased towards microorganisms associated with human health and associated biotechnology (Sun et al., 2020).

The method recommended for use in an industrial setting will primarily depend on its cost and complexity. For LA fermentation, it is expected that the microbial community will have a relatively low diversity (Kim et al., 2016; Tang et al., 2016), especially compared to communities within downstream digesters (Wu et al., 2016). Consequently, *in silico* gene inference analysis could be used as a low-cost method to monitor the metabolic activities of the biological system which could be intermittently confirmed with more complex sequencing techniques (Table 4) to maintain optimal LA production.

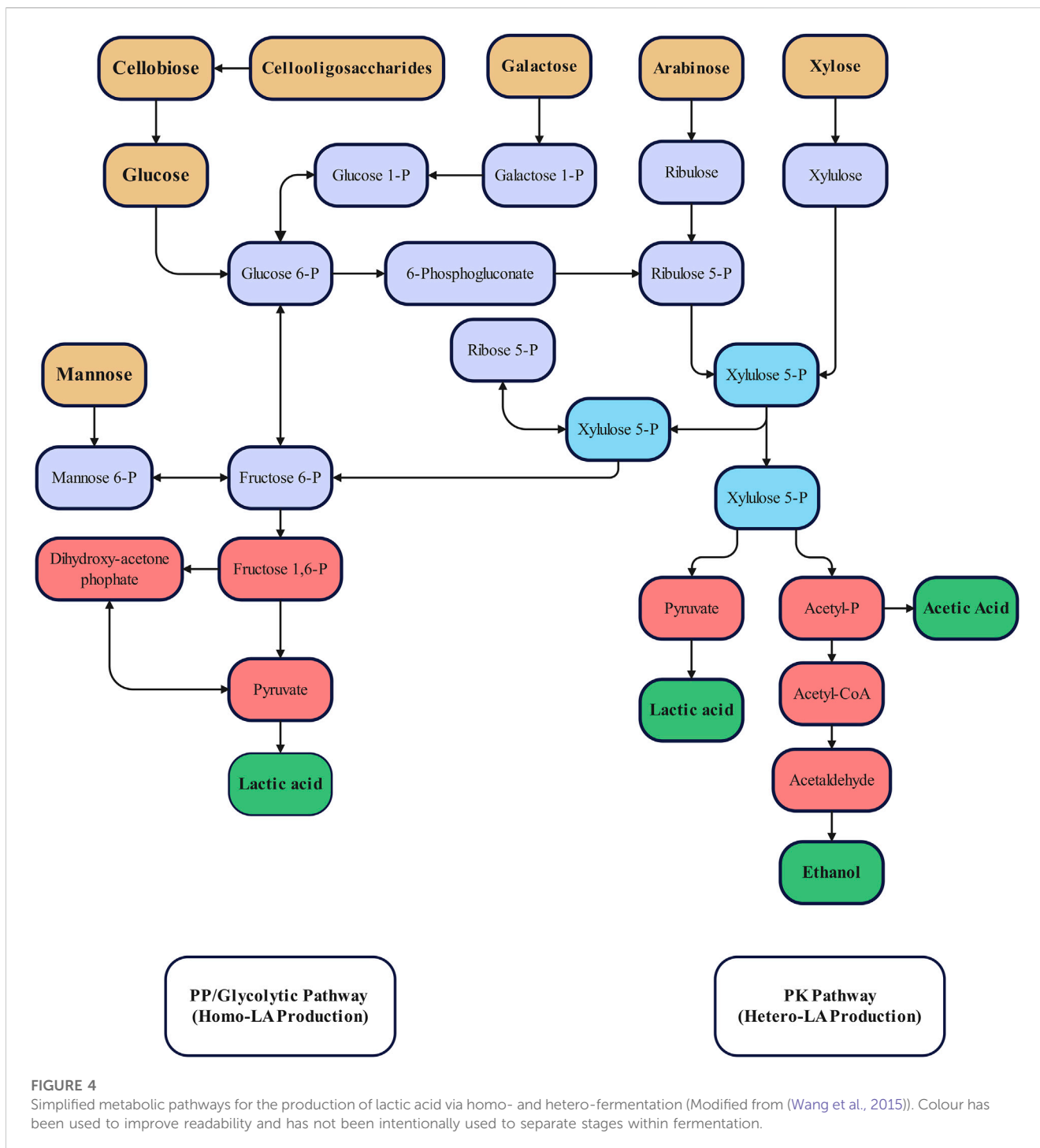
3.2.3 Nutritional requirements

LAB are complex, fastidious microorganisms which require rich and complex nutrients for growth (e.g., amino acids, vitamins, carbohydrates, and minerals) with some requiring specific growth factors, such as whey, and tomato juice (Holzapfel and Wood, 2014). A variety of amino acids have been identified as essential for the growth of LAB including glutamic acid, valine, isoleucine, and leucine, which are required by nearly all LAB, while many require methionine, tryptophan, and tyrosine (Garvie, 1967; Ledesma et al., 1977). A variety of vitamins such as pantothenate, niacin and biotin and the metals, Mg²⁺, Mn²⁺, and Zn²⁺, are also essential for many LAB (Holzapfel and Wood, 2014), albeit, metal ions are usually only

required for enzymatic reactions (Archibald, 1986). While many nutrients have been identified as essential for many LAB, specific nutrient requirements are dependent on strain (Holzapfel and Wood, 2014).

During the fermentation of refined sugars, nutrient supplements are essential for fermentation as LAB lack many biosynthetic capabilities to synthesize nutrients for their own use (Hofvendahl and Hahn-Hägerdal, 2000; Abdel-Rahman et al., 2013). It has been hypothesised these requirements resulted from the bacteria evolving in nutrient-rich media, such as meat and milk, leading them to develop without the need for the bio-processes to synthesize these nutrients (Alves de Oliveira et al., 2018). Therefore, to make these nutrients available, yeast extract, peptone, meat extract, corn steep liquor, and malt extract are commonly utilised (Yang et al., 2013; Wang et al., 2015). As mentioned previously FWs contain many of the essential nutrients required by LAB for growth. Even so, literature has shown FWs may be deficient in some essential nutrients, such as nitrogen, which could limit LA production (Zhang et al., 2020a; Zhang et al., 2020b).

It is important to note the FW classification covers a vast range of different solid and liquid substrates. Generally, liquid wastes consist of process waters with elevated concentrations of suspended solids and soluble organics (e.g., carbohydrates, proteins, and lipids), while solid wastes are generally composed of the solid fraction of various foods, such as fruit and vegetable peels, bones, skins, and seeds (Thakur et al., 2021). However, specific waste streams, including meats, fruit and vegetables, and distillery wastes (Table 1 and 2), all of which may be further categorised into diverse waste streams, can vary substantially in composition (Kosseva, 2013). Consequently, selection of specific FW substrates to maximise LA yield and productivity can be challenging. Targeted selection of certain FW substrates for co-fermentation, such as dairy wastes, could be highly beneficial in this regard. Dairy wastes are known to be natural environments for LAB (Sar et al., 2022) and have been shown to have the potential to yield high LA concentrations (Abedi and Hashemi, 2020), suggesting dairy, or similar wastes, could form beneficial co-substrates for FW fermentation. This



approach allows co-fermented substrates to provide essential nutrients which other co-substrates could be lacking, improving LA production.

Within an integrated LA-AD biorefinery, alternate nutrient supplementation approaches could be available. While literature has demonstrated the nutritional benefits of FW digestate in agriculture (Dutta et al., 2021; Mickan et al., 2022), recent research has identified LA fermentation may benefit from these nutrients present within digestate. Due to the nature of the FW substrate, FW digestate naturally contains elevated ammonium concentrations

(Banks et al., 2011; Serna-Maza et al., 2015; Buhlmann et al., 2018), which may be suitable for LA fermentation. Limited available research has shown the benefits of digestate on FW fermentation, improving pH stability, increasing microbial diversity, and maintaining a low oxidation reduction potential (Wang et al., 2021), and has even been shown to be suitable as process water following pre-treatment (Zhang et al., 2019). Implementing partial digestate recirculation to LA fermentation within an LA-AD biorefinery may be an effective way to promote LA production and provide an alternative on-site use for the low-value digestate. However, additional research is

TABLE 4 Advantages and disadvantages of different metabolic pathway monitoring methods.

Method	Techniques	Advantages	Disadvantages	References
Predictive	PICRUSt PICRUSt2 Tax4Fun FaproTax	<ul style="list-style-type: none"> • Users can customise the tool to meet the needs of their system • Predictive power will improve with time • Low cost 	<ul style="list-style-type: none"> • Only 16S marker genes for bacteria and archaea are currently included • Quality of predictions are dependent on input data used • Biased towards human health and biotechnology 	Langille et al. (2013), Sun et al. (2020)
Metabolomics	Nuclear magnetic resonance	<ul style="list-style-type: none"> • Fast and highly reproducible • Minimal sample preparation • Non-destructive • Provides information on metabolite concentration and chemical structure 	<ul style="list-style-type: none"> • Insensitivity • Magnetic fields required affect surrounding equipment • High instrument cost 	Chatham and Blackband (2001), Oldiges et al. (2007), Nagana Gowda et al. (2008), Scalbert et al. (2009), Alonso et al. (2015)
	Mass spectrometry	<ul style="list-style-type: none"> • Can measure metabolites in complex bio-fluids • Can distinguish between isotopes 	<ul style="list-style-type: none"> • Data post-processing is time intensive • Requires standards for metabolite identification • Prone to matrix effects • Can have a high cost • Requires tedious sample preparation 	
Lipidomics	Raman spectroscopy	<ul style="list-style-type: none"> • Non-destructive • Non-invasive • Ability to obtain complex information • Obtain spectral and spatial information • No specific sample requirements 	<ul style="list-style-type: none"> • Measurements may affect compounds within the samples • Slow • Requires advanced chemometric tools for data analysis • Had to adapt for quantitative analysis 	Jurowski et al. (2017)
	Fourier transform infrared spectroscopy	<ul style="list-style-type: none"> • Non-destructive • Non-invasive • Ability to obtain complex information • Obtain spectral and spatial information • No specific sample requirements 	<ul style="list-style-type: none"> • Lower spatial resolution than Raman spectroscopy • Requires absence of water • can be slow • Requires advanced chemometric tools for data analysis 	
	Nuclear magnetic resonance	<ul style="list-style-type: none"> • Non-destructive • large range of samples are applicable (^{31}P)^a • Structural analysis of purified compounds (^1H)^a 	<ul style="list-style-type: none"> • Low selectivity • Sensitive to motion • High cost • Magnetic fields required affect surrounding equipment 	
Shotgun Metagenomics	-	<ul style="list-style-type: none"> • Avoids amplification bias • Full community analysis 	<ul style="list-style-type: none"> • High cost • Analysis of bioinformatics data is computationally intensive and complex • Provides lower taxonomic resolution than 16s rRNA data 	Langille et al. (2013), De Filippis et al. (2017), Mukherjee et al. (2017)

a: (^{31}P) phosphorous atom is utilised, (^1H) hydrogen atom is utilised.

required to understand the impact digestate may have on LA fermentation, the product spectrum, microbial community, and degradation pathways for LA production.

4 Integration challenges/considerations

While integrating LA production into AD has been suggested to improve the overall process economics (Section 2.2), care should be taken to ensure optimal operation of both processes. For this, specific feed requirements and operational parameters and chemical/nutrient

supplements should be considered, emphasising their possible impact on downstream processes. Furthermore, special care should be taken when selecting the LA recovery method to ensure the usability of fermentation waste in AD.

4.1 Challenges using FW as a substrate for fermentation

While approximately 90% of worldwide LA production is achieved through fermentation (Karp et al., 2011), these processes primarily utilise food based feedstocks (Section 3.1)

which are not only influenced by seasonal variability, but high cost coupled with the high cost of nutrient supplements increases the price of the LA production. In contrast, the versatile AD technology is capable of processing a variety of wastes including FW, cardboard, grease trap residues, and fat oils (Edwards et al., 2015). This capacity to receive various waste streams provides a buffer from feedstock supply uncertainties and seasonal changes in waste availability, though depending on location, surrounding industries, climate, season, population density and socio-demographics, and government policies such as landfill waste diversion, available feedstocks are expected to somewhat vary (Ghatak, 2011; Bühlmann et al., 2021). Utilisation of FW feedstocks for LA fermentation can reduce costs associated with acquiring feedstocks, however, fermentation of complex substrates, availability of mixed sugars, and introduction of alternate microbes from the complex FW feedstock may impact LA production.

4.1.1 Feedstock composition, and potential pre-treatment requirements

The maximum yield achieved and feedstock cost heavily influences the economics of fermentation (Manandhar and Shah, 2020). Even though waste streams can reduce operational costs associated with obtaining feedstocks, yields are still a challenge. Fermentation of waste to LA is primarily limited by the available carbohydrate fraction within the substrate. While carbohydrates make up a significant fraction of FW (Demichelis et al., 2017), LAB struggle to fully utilise the substrate. Pre-treatment of the FW via enzymatic, fungal, acidic, or alkali pre-treatments can effectively improve the LA yield on FWs (Kim K. I. et al., 2003; Kwan et al., 2016; Pleissner et al., 2016; Demichelis et al., 2017; Ahmad et al., 2020), but are costly and generally produce large quantities of solid and liquid wastes which require further treatment prior to disposal (Surendra et al., 2015). A diverse community adapted for LA accumulation could be beneficial for FW fermentation, allowing the increased utilisation of the diverse FW substrate from the presence of diverse hydrolytic bacteria. Moreover, utilisation of a functionally redundant community could improve process stability to changes in feedstock composition. However, while studies have explored the community structure during LA fermentation (Bühlmann et al., 2021; Wang et al., 2021) the functional stability of these cultures have not been fully explored.

Some wastes, such as lignocellulose (following pre-treatment), can release mixed sugars which may lead to carbon catabolite repression (CCR) (Abdel-Rahman and Sonomoto, 2016). The presence of a variety of different carbon sources may complicate fermentation as some bacteria limit the utilisation of secondary carbon sources when a primary source is present (i.e., CCR), which is a problem for most microbial producers (Görke and Stülke, 2008; Abdel-Rahman and Sonomoto, 2016). For example, *Escherichia coli* prefer glucose over lactose as a carbon source while *Streptococcus thermophilus* prefer lactose over glucose (Brückner and Titgemeyer, 2002). These bacteria will metabolise their preferred substrate before utilising the secondary source. Mixed sugar fermentation could result in increased costs associated with separation and purification stages (Wang et al., 2015) due to lower LA yields and increased by-product formation. This behaviour can be problematic when fermenting substrates containing various carbon sources as certain sugars may require the utilisation of hetero-fermentative pathways for LA production (Figure 4).

4.1.2 Microbial contamination

Microbial contamination is also a significant risk which can complicate microbial LA production, especially when utilising waste feedstocks, but is not widely explored for LA fermentation. AD facilities receive and utilise a variety of waste feed stocks which are primarily unsterilised. Though this does not pose a problem for AD, it can be a significant risk to the economics of LA fermentation. Many fermentation systems with optimised process conditions (e.g., pH, temperature, HRT, and OLR) tend to selectively promote LAB and LA production (Kim et al., 2016; Tang et al., 2016; Bühlmann et al., 2022b). However, FWs contain a variety of alternate bacteria, and are naturally enriched with many LAB (Kim et al., 2016), which may compete with target strains for substrate, reducing LA yields and selectivity. Pasteurisation is generally applied to eliminate the risk of microbial contamination, however, due to its high energy demands (Bolzonella et al., 2018; Liu et al., 2019), many studies aim to utilise FW without pasteurisation (Kim et al., 2016; Tang et al., 2016; Feng et al., 2018; Zhang et al., 2020a; Zhang et al., 2020b). An LA-AD biorefinery could hold a distinct advantage in this regard, producing large quantities of waste heat generated from biogas combustion which could be utilised for upstream pasteurisation. Future LA-AD biorefineries will likely utilise pasteurisation or finely tuned operational conditions to selectively promote the growth of target strains to ensure consistent LA production from fermentation.

4.2 Separation and recovery considerations

The viability of biologically derived products is heavily dictated by the cost of downstream processes required to isolate the target compound (Saboe et al., 2018). However, the separation and recovery of LA is difficult due to its low vapour pressure, high affinity to water, and tendency to undergo self-esterification (Sun et al., 2006). Consequently, separation and final purification can represent up to 50% of the production costs (Komesu et al., 2017a).

Traditionally, LA is recovered via gypsum precipitation and esterification, for the commercial production of high purity LA (Figure 5) (Lee, 2015; Komesu et al., 2017a). LA generally exists as a salt in the fermentation broth due to the addition of neutralising agents (CaCO_3 , Ca(OH)_2 , NaOH , NH_3) to maintain optimal pH (5–7) (normally CaCO_3 , Ca(OH)_2) (Lee, 2015; Komesu et al., 2017b; Alves de Oliveira et al., 2018). Following fermentation, the broth may be adjusted to a pH of 10 and heated to 80 °C in order to solubilise the calcium lactate and coagulate proteins within the broth to simplify filtration (Lee, 2015). The broth is then filtered and re-acidified with sulphuric acid to produce LA and precipitate gypsum. The resultant mixture is then filtered, producing a technical grade LA mixture (22%–44%) (Komesu et al., 2017a). This product can be further refined to produce high purity LA, through esterification with methanol, distillation, and hydrolysis (Figure 5).

While this recovery process is effective and a proven technology, it is costly, produces large quantities of gypsum waste, and requires large volumes of sulphuric acid (Komesu et al., 2017c; Jantasee et al., 2017; Alves de Oliveira et al., 2018; Singhvi et al., 2018). Consequently, research has been focused on developing and testing alternate

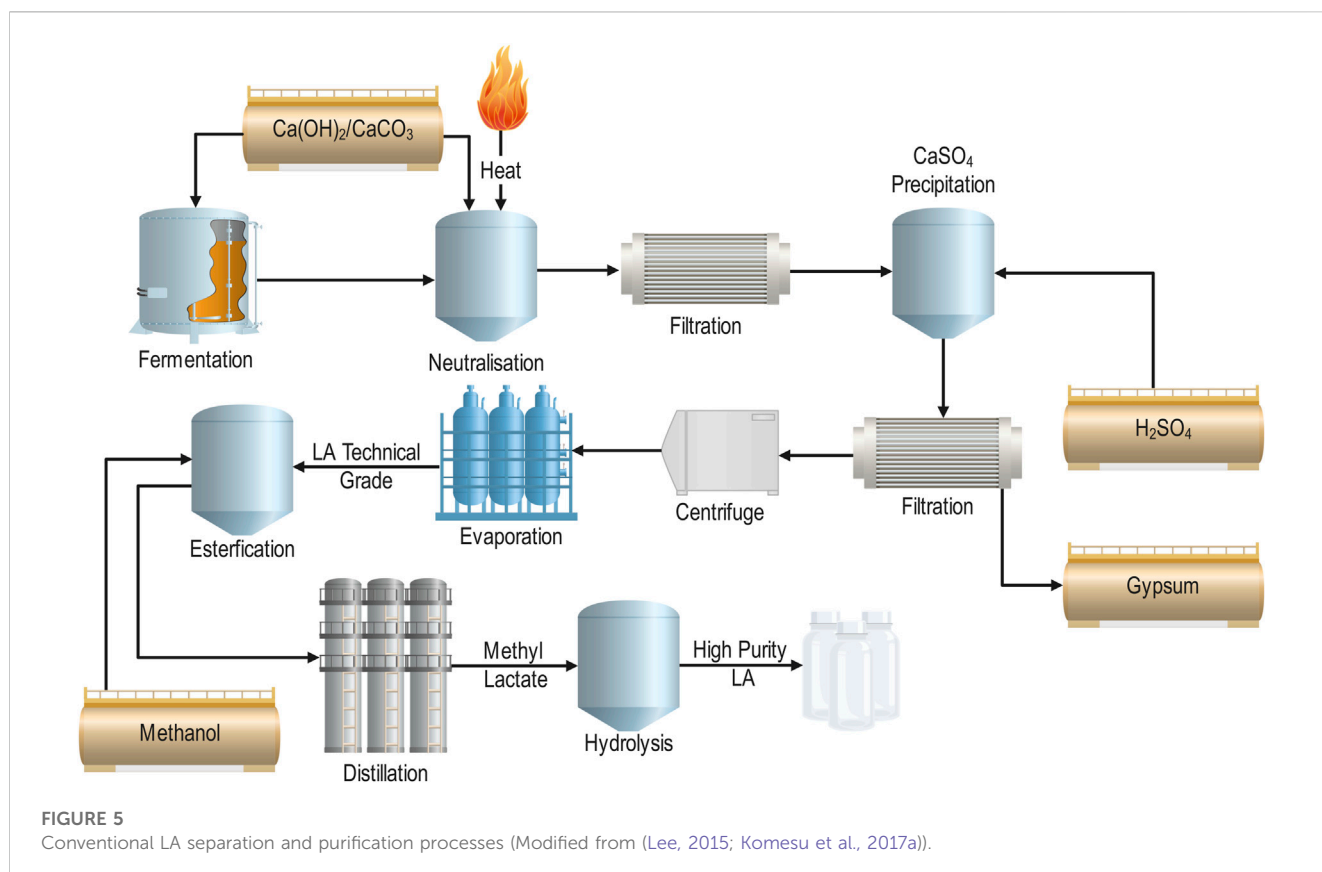


TABLE 5 Advantages and disadvantages of separation processes for the recovery of lactic acid (Adopted from (Komesu et al., 2017b)).

Separation process	Advantages	Disadvantages
Precipitation	<ul style="list-style-type: none"> Easily applicable in industry 	<ul style="list-style-type: none"> High sulphuric acid consumption Generates large quantities of gypsum Low product purity
Liquid-liquid extraction	<ul style="list-style-type: none"> No gypsum generation Reduced risk of thermal decomposition 	<ul style="list-style-type: none"> Extractant requires stripping and regeneration stages Low product purity Conventional extraction agents show unfavourable activity coefficients
Membrane processes	<ul style="list-style-type: none"> Great flexibility in production scale High selectivity High levels of purification Potential integration with conventional fermenters 	<ul style="list-style-type: none"> Membranes have a high cost Fouling of membranes Polarization issues Difficulties in upscaling
Molecular distillation	<ul style="list-style-type: none"> Reduced risk of thermal decomposition High purification levels No solvents No further purification stages needed 	<ul style="list-style-type: none"> Difficulties in upscaling Requires high vacuum conditions
Reactive distillation	<ul style="list-style-type: none"> Integrates reaction and separation into the same apparatus High purification levels Lower energy consumption 	<ul style="list-style-type: none"> Process is complex specifically applied to reversible reactions in the liquid phase Requires high reaction rates Separation and reaction temperatures need to be relatively close together Homogeneous catalyst leads to corrosion and separation issues

separation and purification technologies for LA including; distillation, solvent extraction, adsorption, and membrane separation processes (reverse osmosis, ultrafiltration, and electrodialysis) (Komesu et al., 2017a). Ideally, the separation process selected for LA recovery should be based on the efficient and economical usage of these processes, along with a consideration of their individual advantages and disadvantages (Table 5) (Wasewar, 2005).

4.2.1 Advances in separation technologies

The current commercial method for LA separation and purification has many economic and environmental ramifications when obtaining pure LA. Consequently, alternative methods for the recovery and purification of LA have been the focus of recent literature (Table 5). Recovery techniques which can be utilised *in situ* show promise in reducing environmental impacts and reagent

addition associated with fermentation and LA recovery. These technologies couple fermentation with separation for the continuous production and extraction of LA. A variety of technologies exist, each with its own associated advantages and disadvantages (Table 6). Two promising *in situ* techniques include liquid-liquid extraction and ion exchange due to their high efficiency and selectivity in recovering LA from fermentation broth.

4.2.2 Liquid-liquid extraction

In-situ liquid-liquid extraction is a promising method for LA recovery. A variety of solvents can be utilised including, water insoluble amines, quaternary ammonium salts, esters, or ketones. Due their high selectivity and efficiency of tertiary amines, as well as their poor solubility in the aqueous phase, they are also appropriate for LA extraction (Jantasee et al., 2017). A variety of tertiary amines have been tested utilising a number of different alcohols as diluents but trioctylamine in 1-octanol remains the extractant-diluent combination that provides the highest LA distribution (Krzyżaniak et al., 2013). However, it is important to note that these solvents are toxic to microorganisms (Singhvi et al., 2018), which is closely related to the hydrophobicity of the solvent, although the range of tolerable concentrations is depended by the type of microorganism (Matsumoto et al., 2004). Optimisation of the solvent concentration can reduce this toxic effect as shown by Gao et al. (2009).

4.2.3 Adsorption

Adsorption is another promising recovery technique and is widely used within industrial biotechnology as it is robust and relatively easy to operate (da Silva and Miranda, 2013). Various resins have been applied for LA recovery (Table 7) but weakly basic resins are generally

preferred as they do not require powerful regeneration steps and have a much higher resistance to organic fouling compared to strongly basic resins (Tung and Judson King, 1994; Gluszc et al., 2004). Several reports have explored anionic resins for LA recovery and notably Boonmee et al. (2016) and Ataei and Vasheghani-Farahani (2008) reported a 2.1–6.8 and 5 fold increase in LA productivity, respectively, when applying anion exchange to LA fermentation. Furthermore, Zhang et al. (2018) identified the cost to apply *in situ* anion exchange for LA recovery was similar to that of CaCO₃ for pH control. Therefore, application of ion exchange for LA recovery can enhance the production rate of LA, allowing reduced process vessel volumes and capital costs, while simultaneously reducing the need for neutralising agents following fermentation.

4.2.4 Recovery method considerations

While a variety of LA recovery techniques have been explored in literature (Table 6), they are predominantly concerned with the recovery of LA and tend not to consider implications for downstream processes. Within an integrated LA-AD biorefinery, this would not only concern further purification stages, but also downstream AD which would likely utilise the extraction residues within AD for disposal and methane generation. This would likely include the solid and liquid fraction fractions. For this, the recovery method should be carefully considered to ensure LA is selectively extracted, not only to reduce downstream costs, but also to maximise the remaining organic fraction within the residues to maximise biogas production within AD. Furthermore, the extraction method should minimise the broths exposure to toxic or inhibitory compounds which may follow the extraction residues to downstream AD and inhibit methane formation or restrict digestate use within agriculture. For

TABLE 6 Summary of advantages and disadvantages of different *in situ* recovery techniques (Modified from (Van Hecke et al., 2014))

Recovery technique	Operation concept	Advantages	Disadvantages	References
Liquid-liquid extraction	LA is dissolved in the hydrophobic solvent via a reaction mechanism that varies from proton transfer to ion exchange	<ul style="list-style-type: none"> • High efficiency and selectivity 	<ul style="list-style-type: none"> • Solvents and impurities are toxic to microorganisms • Solvent impurities may be toxic to microorganisms • Different in optimal pH for fermentation and LA extraction 	Yankov et al. (2005), Gao et al. (2009)
Ion exchange resins	Ion exchange resins extract dissociated LA from the aqueous phase by exchanging an anion for the dissociated lactate anion	<ul style="list-style-type: none"> • Improved LA productivity • Higher substrate loading • Reduced inhibition • Elimination of need for neutralising agent 	<ul style="list-style-type: none"> • Requires elution and regeneration stages • Prone to organic fouling 	Bornak (2012), Boonmee et al. (2016), Zhang et al. (2018)
Electrodialysis	Cation and anion exchange membranes allow the selective transfer of ions depending on their charge, allowing their selective removal into separate compartments	<ul style="list-style-type: none"> • Based produced from water splitting electrodialysis can be used for pH control • LA is removed in a concentrated form 	<ul style="list-style-type: none"> • Membrane fouling • Corrosion • High capital expenditure • High power consumption • Strict pH requirements 	Li et al. (2004), Arora et al. (2007), Ramaswamy et al. (2013), Cassano (2016)
Electrodeionization	Electrically driven separation process, similar to electrodialysis, which incorporates ion exchange resins to provide a pathway for improved ion migration	<ul style="list-style-type: none"> • High separation factor 	<ul style="list-style-type: none"> • Nutrients from growth media may reduce separation efficiency, product purity and lead to lactate salt formation 	Arora et al. (2007), Pan et al. (2017)
Crystallization	Calcium lactate is precipitated straight from the fermentation broth without the removal of biomass or cell mass	<ul style="list-style-type: none"> • Significantly improved LA productivity • High LA yield on glucose 	<ul style="list-style-type: none"> • Requires Ca(OH)₂ addition • Requires separate reactor for crystallisation due to difference in fermentation and crystallisation temperatures 	Xu and Xu (2014)

TABLE 7 Summary of LA removal from via the use of anion exchange resins.

Resin	Mode	Loading ($\text{g}_{\text{LA}} \cdot \text{g}_{\text{sorbent}}^{-1}$)	LA production	Notes	References
Dowex MWA-1	Synthetic solution	0.3–0.18 (pH 5–6)	-	Identified as best candidate for LA recovery	Tung and Judson King (1994)
Reillex 425	Synthetic solution	<0.05 (pH 5–6)	-		Tung and Judson King (1994)
Duolite A7	Synthetic solution	0.18–0.08 (pH 5–6)	-		Tung and Judson King (1994)
Amberlite IRA-910	Synthetic solution	0.3 (pH 5–6)	-	Requires pH increase to >11 for LA elution	Tung and Judson King (1994)
Amberlite IRA-35	Synthetic solution	0.35–0.29 (pH 5–6)	-	Identified as best candidate for LA recovery	Tung and Judson King (1994)
Amberlite IRA-67	<i>In situ recovery</i>	0.15 (pH 6.5)	Improved LA productivity by 2.1 to 6.8 fold compared to batch fermentation		Boonmee et al. (2016)
Amberlite IRA 67	<i>In situ recovery</i>	80 mg/mL (pH 5.0)	-		John et al. (2008)
Amberlite XAD1600 (Neutral resin)	Separation from grass silage juice	-(<3.78)	-	Undissociated LA is adsorbed onto the resin while inorganic salts and sugars were not	Thang and Novalin (2008)
Amberlite resin (IRA-400, Cl ⁻)	<i>In situ recovery</i>	-	LA productivity was 5-times higher than the conventional system		Ataei and Vasheghani-Farahani (2008)
335	<i>In situ recovery</i>	0.23 (pH 5.5)	Similar to conventional fermentation	Did not demonstrate any adsorption capacity for glucose	Zhang et al. (2018)

example, while liquid-liquid extraction processes can be effective for LA recovery (Section 4.2.2), a portion of the solvent will remain within the extracted broth and accumulate within AD. Depending on the microbial tolerances and the section of the community impacted, the retained solvent could impact AD process performance and stability, and impact the quality the digestate produced.

While only a handful of literature has examined the feasibility of utilising LA fermentation waste within AD (Dreschke et al., 2015; Kim et al., 2016; Demichelis et al., 2017; Bühlmann et al., 2022a), the results are promising. For example, Demichelis et al. (2017) outlined the solid fermentation residues were appropriate for methane formation, while, a similar study by Bühlmann et al. (2022a) reported the liquid extraction residues, following LA recovery via ion exchange, were also suitable for use within AD along with the solid fraction. Overall, AD can effectively utilise extraction residues from LA recovery, however, future work should explore the impacts of alternative recovery methods on the liquid fraction aiming to minimise downstream impacts to AD.

5 Concluding remarks and recommendations

Overall, the current literature review identified challenges with integrating LA production into commercial FW-AD; however, it was clear that LA production from waste streams could boost the economic performance of two-stage AD systems, providing a high-value by-product and better harnessing existing capital. FW, being composed of the primary carbohydrates and nutrients required for LA production, is a promising substrate for LA production with recent

literature having shown solid FW fermentation residues and liquid extraction residues can be utilised within existing AD systems engineered for methane production. While these reports are promising for the LA-AD biorefinery, the commercial FW context is highly variable and complex, and has yet to be fully explored, particularly in areas related to waste availability, its diverse and varying composition, and its potential impact on LA fermentation and recovery. Furthermore, literature has yet to fully explore the integration into existing two-stage AD infrastructure. For this, the pre-fermenter would be converted to an LA fermenter which, to minimise start-up and operational costs, should be regulated through the control levers of pH and temperature. Moreover, different operational modes and reactor designs should be explored to elucidate their effects on LA-AD biorefinery concept. A pilot scale study would be highly beneficial in this regard, providing a platform for the study of LA production from diverse, complex, and variable FW, study stability of LA producing communities, how LA recovery impacts the BMP of the fermented FW, while also providing an opportunity for unknown process influences to be studied.

Limited literature has shown an industrial inoculum can be directed to target LA, however, further research is required to identify methods to redirect commercial fermentation to target and maximise LA. Furthermore, studies should explore community functional stability and explore methods to ensure consistent LA production. Such a study could be conducted on a continuous fermenter fermenting real diverse FW, while monitoring community composition, function, and LA production. Although literature has identified the solid and liquid residues are suitable for methane production, it is unclear how different recovery methods

may impact the liquid fraction and resulting AD. Therefore, to continue the development of the LA-AD biorefinery, it is necessary to explore the above-mentioned research areas while aiming to maximise LA production and minimise the negative impacts imposed on downstream AD.

Author contributions

CB: Conceptualization, Writing—original draft, Writing—review and editing, Investigation, Project administration, Visualization. BM: Funding acquisition, Supervision, Writing—review and editing. ST: Supervision, Writing—review and editing. DB: Writing—review and editing. PB: Funding acquisition, Supervision, Writing—review and editing.

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