Check for updates

OPEN ACCESS

EDITED BY Siegfried Ussar, Helmholtz Association of German Research Centres (HZ), Germany

REVIEWED BY Christian Dani, INSERM, France

RECEIVED 17 March 2023 ACCEPTED 14 April 2023 PUBLISHED 26 April 2023

CITATION

White U (2023), Adipose tissue expansion in obesity, health, and disease. *Front. Cell Dev. Biol.* 11:1188844. doi: 10.3389/fcell.2023.1188844

COPYRIGHT

© 2023 White. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Adipose tissue expansion in obesity, health, and disease

Ursula White*

Clinical Science Division, LSU Pennington Biomedical Research Center, Baton Rouge, LA, United States

White adipose tissue (WAT) expands under physiological conditions via an increase in adipocyte size (hypertrophy) and/or number (hyperplasia; adipogenesis), and the ability of WAT to expand to accommodate energy demands is a significant determinant of metabolic health status. Obesity is associated with impaired WAT expansion and remodeling, which results in the deposition of lipids to other nonadipose organs, leading to metabolic derangements. Although increased hyperplasia has been implicated as a cornerstone in promoting healthy WAT expansion, recent developments suggest that the role of adipogenesis as a contributing factor in the transition from impaired subcutaneous WAT expansion to impaired metabolic health remains up for debate. This minireview will summarize recent developments and highlight emerging concepts on the features of WAT expansion and turnover, and the significance in obesity, health, and disease.

KEYWORDS

adipose tissue, adipose expansion, adipocyte, adipogenesis, obesity, human, metabolic health

1 Introduction

White adipose tissue (WAT) is the primary site for energy storage in the form of triacylglycerols (TGs) and is an important regulator of whole-body energy homeostasis (Cypess, 2022). Due to its high plasticity, WAT retains the ability to expand, reduce, and remodel during adulthood to accommodate changes in energy balance in response to a variety of metabolic stimuli, including obesity, diet, and exercise. The patterns of WAT expansion vary amongst the population and during metabolic perturbations, and these mechanisms continue to be uncovered. WAT mass is regulated by dynamic changes in adipocyte volume, via TG synthesis and breakdown, as well as adipocyte formation (i.e., hyperplasia; adipogenesis) and death. It is now appreciated that there is constant turnover (synthesis/formation and removal/death) of WAT TGs (Strawford et al., 2004; Arner et al., 2011) and adipocytes (Strawford et al., 2004; Spalding et al., 2008).

WAT expansion under physiological conditions involves via both hypertrophy (enlargement of existing adipocytes' size) and hyperplasia (increase in adipocyte number; adipogenesis), and evidence suggests that the capacity of subcutaneous WAT to expand is a key determinant of obesity-related metabolic dysregulation (Danforth, 2000; Sethi and Vidal-Puig, 2007). The adipose tissue expandability hypothesis postulates that impaired subcutaneous WAT expansion in response to energy demands can lead to ectopic lipid deposition in non-adipose organs and contribute to the development of insulin resistance and type 2 diabetes (Virtue and Vidal-Puig, 2010). Arner et al. reported that greater hypertrophy was linked to lower adipocyte number and insulin resistance, independent of sex and adiposity, while greater hyperplastic expansion was associated with better insulin sensitivity (Arner et al., 2010). Additional studies also suggest that impaired adipogenesis, or restricted hyperplasia, is linked to metabolic dysfunction (Weyer et al., 2000; Lessard et al., 2014), while a higher population of small adipocytes (e.g., adipogenesis) is associated with better glycemic and lipid profiles (Arner et al., 2010; Hoffstedt et al., 2010). However, the role of impaired adipogenesis as a contributing factor in the pathogenesis of obesity-related disorders remains debatable (White and Ravussin, 2019). Other studies reported that individuals with insulin resistance or T2DM have a higher proportion of small adipocytes, suggesting hyperplastic expansion, as compared to healthy individuals (McLaughlin et al., 2007; Pasarica et al., 2009; McLaughlin et al., 2014), and these observations imply that impaired expansion of small adipocytes is associated with insulin resistance (McLaughlin et al., 2007; McLaughlin et al., 2014; White et al., 2022).

Despite the significant role of WAT expansion in human health and pathology, the mechanistic underpinnings, notably the kinetics of adipose tissue components (adipocytes and TGs), are not fully understood. This is due, in part, to the slow turnover rate of WAT components. This mini review will discuss the dynamics of WAT expansion in humans during obesity, changes in energy balance (e.g., weight gain and loss), and exercise as relates to metabolic health and disease in humans, as well as some emerging, state-ofthe-art methodologies to assess the features of WAT expansion.

2 Challenges and novel approaches to measure adipose tissue expansion

The expansion and turnover of WAT (lipids and cells) have been difficult to study given the lack of appropriate methods; thus, the underlying mechanisms are not fully understood. Indirect measures, such as fat cell size and molecular markers of cell proliferation and death, are informative, but do not provide an integrative evaluation of turnover. Furthermore, the unspecific labeling of nucleotides with ³H-thymidine or BrdU, as well as the toxicity of the label, make these approaches inapplicable to humans. A more recent in vitro approach is the use of three-dimensional (3D) adipose tissue culture models, which can utilize human WAT stem cells that differentiate and organize into adipose organ-like structures or adipose explants in culture, thus, providing a more physiological setting to investigate adipocyte function (Murphy et al., 2019). Nevertheless, in vivo approaches are necessary to capture the dynamic changes that occur during the various facets of WAT expansion and turnover within the natural environment of the adipose tissue.

Spalding et al. (2008) introduced an *in vivo* method to study fat cell and lipid turnover in humans by measuring the incorporation of atmospheric ¹⁴C, derived from above ground nuclear bomb tests, into the adipocytes (Arner et al., 2011). The Hellerstein group developed another innovative method (Busch et al., 2007) using the incorporation of the stable isotope deuterium (²H) into adipose cells and lipids that has been validated to provide physiological, quantitative measures of WAT turnover *in vivo* (Neese et al., 2002; Strawford et al., 2004). While the ¹⁴C-labeling method has provided informative retrospective assessments of WAT expansion and turnover, the ²H-labeling approach can measure dynamic changes in WAT during prospective intervention studies (White and Ravussin, 2019). Studies using metabolic labeling with stable

isotopes can fill a substantial knowledge gap surrounding the dynamics of WAT remodeling and plasticity in humans.

3 Adipose tissue expansion in obesity

Obesity, characterized by excessive adiposity, can lead to dysregulation of WAT expansion and function, resulting in lipotoxicity and obesity-related comorbidities (Virtue and Vidal-Puig, 2010). Because in vitro data provide evidence to support the adipose tissue expandability hypothesis, one group investigated the relationship between the estimated manner of WAT expansion and the *in vivo* generation of adipocytes, as assessed by the ¹⁴C-labeling approach, in both lean subjects and individuals with obesity (Arner et al., 2010). Individuals displaying more hypertrophic fat expansion produced fewer adipocytes in vivo per year than individuals displaying more hyperplastic expansion. Another study assessed in vivo kinetics in the subcutaneous abdominal and femoral WAT depots of women with obesity using the ²H-labeling protocol and reported that higher in vivo adipocyte formation rates were positively correlated with increased adiposity [body mass index (BMI) and % body fat] (White et al., 2016). Interestingly, higher (not lower) in vivo adipogenesis was positively associated with visceral adipose tissue content and negatively associated with insulin sensitivity (White et al., 2017). These data challenge the adipose tissue expandability hypothesis and provide the first evidence of an association between facets of impaired metabolic health and higher, not lower, in vivo adipose cell formation.

TGs are a metabolically active pool and estimated to be replaced (i.e., turnover) ~6 times during the ~10 years lifespan of a healthy adipocyte. However, obesity, which is associated with impaired lipid metabolism, is characterized by decreased adipocyte TG turnover, as TGs are estimated to be replaced only ~3 times during the lifespan an adipocyte in individuals with obesity (Arner et al., 2011). This implies that high TG storage coupled with low removal (via lipolysis), or low TG turnover, may be important determinants of obesity (Arner et al., 2011). Other observations using the ²H-labeling approach reported that Black women had lower TG synthesis rates as compared to White women with obesity (White et al., 2018), who were also shown to have enhanced insulin sensitivity vs. Black women (DeLany et al., 2014). Another group reported that TG synthesis rates were significantly higher in insulin-sensitive vs. insulin-resistant individuals (Allister et al., 2015). Overall, these data suggest that higher adipose TG turnover is associated with favorable metabolic outcomes. Notably, a recently published study (Arner et al., 2018) implicated lipolysis as an important predictor of future weight gain and impaired glucose metabolism.

4 Adipose tissue expansion during changes in energy balance

To date, there have been no experimental overfeeding studies to examine the dynamics of WAT expandability *in vivo* during weight gain. Two studies (Johannsen et al., 2014; McLaughlin et al., 2016) have reported that individuals with smaller mean adipocyte size at baseline had the most impaired metabolic health outcomes (e.g., greater decline in insulin sensitivity) in response to overfeeding and

weight gain, as compared to those with a larger adipocyte size. In addition, one group recently reported that an increased proportion of small adipocytes (i.e., hyperplasia) in subcutaneous WAT is associated with impaired (not improved) metabolic health outcomes, specifically visceral and ectopic fat accumulation in the liver, during weight gain in response to overfeeding (White et al., 2022). These findings imply the presence of small adipocytes with a decreased capacity to accommodate lipid. Interestingly, although few studies have assessed depot differences in WAT expansion in humans, one study suggested depot-specific fat expansion in response to overfeeding, with mainly hypertrophy in the subcutaneous abdominal WAT and primarily hyperplasia in the subcutaneous femoral (Tchoukalova et al., 2010). Additional in vivo assessments during overfeeding interventions are necessary to better characterize the influence of WAT expansion during positive energy balance in the pathogenesis of metabolic disorders.

There is a paucity of data on the dynamics of WAT plasticity and remodelling during weight loss. Interestingly, one investigation reported that adipose cell formation in the subcutaneous WAT was negatively associated with the change in body weight during the ²H-labeling period, suggesting that women with greater weight loss had higher *in vivo* adipogenesis (White et al., 2017). Further investigations are necessary to better understand WAT expansion and remodelling during weight loss (i.e., caloric restriction, bariatric surgery, etc.) and determine how these adaptations could influence weight regain and weight cycling.

5 Adipose tissue expansion and exercise

The favorable effects of exercise on cardiovascular health and skeletal muscle are well-established, but rodent studies suggest that exercise-induced adaptations in the WAT may also influence overall metabolic health (Stanford et al., 2015). Although some important WAT adaptations in response to exercise have been reported, including changes in morphology and decreased adipocyte size (Despres et al., 1984; Mauriege et al., 1997), no studies have reported the effects of exercise on WAT turnover in vivo in humans. One study using the ²H-labeling protocol demonstrated that exercise (4 weeks of voluntary wheel running) significantly reduced new adipocyte formation (e.g., adipogenesis) in the WAT of both male and female mice in vivo (Allerton et al., 2021). Despite the very limited data available, these results suggest that exercise induces WAT remodeling in such a manner that reduces the need for new adipocyte formation, possibly due to enhanced metabolic efficiency and lifespan of existing adipocytes. Of note, higher adipogenesis could indicate a need for new adipocyte formation, due to existing adipocytes' fragility and death, which can lead to the recruitment of macrophages, unfavorable remodeling and inflammation (Strissel et al., 2007). Hence, lower hyperplasia could be a critical and metabolically favorable exercise-mediated WAT adaptation. Human studies to assess the effects of exercise on the dynamics of WAT expansion in vivo have yet to be reported.

6 Discussion

Given that the capacity for WAT expansion is a significant determinant of obesity-related complications, further investigations are necessary to better elucidate the important facets of WAT expandability and turnover in humans. Of note, more recent studies have identified the presence of white adipocyte subpopulations in WAT with distinct functions (Bilson et al., 2023), suggesting that adipocytes are not a uniform cell type. This presents new challenges to better understand WAT biology, as changes in the presence, function, and/or turnover of adipocyte subpopulations can contribute to obesity-related metabolic diseases.

Isotopic labeling methodologies to assess *in vivo* lipid and cell kinetics are a substantive departure from indirect and *in vitro* methods; and new insights derived from these emerging, cuttingedge approaches will advance our understanding of the important changes in WAT function that occur during changes in energy balance and in conditions of obesity and disease. Notably, the ²H-labeling method can provide comprehensive, quantitative measures of WAT turnover during a variety of prospective intervention studies, such as diet, exercise, and pharmacological treatments. This knowledge can facilitate the future development of therapeutic targets and treatments for obesity and related disorders that are focused on WAT.

Author contributions

UW conceived the topic, designed, and wrote all sections of the manuscript. UW conducted manuscript revision, read, and approved the submitted version.

Funding

UW is supported by R01DK121944 and partially by a NORC Center Grant P30DK072476 from the National Institute of Diabetes and Digestive and Kidney Diseases of the National Institutes of Health.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher. Allerton, T. D., Savoie, J. J., Fitch, M. D., Hellerstein, M. K., Stephens, J. M., and White, U. (2021). Exercise reduced the formation of new adipocytes in the adipose tissue of mice *in vivo*. *PLoS One* 16 (1), e0244804. Epub 2021/01/21. doi:10.1371/journal.pone.0244804

Allister, C. A., Liu, L. F., Lamendola, C. A., Craig, C. M., Cushman, S. W., Hellerstein, M. K., et al. (2015). *In vivo* 2H2O administration reveals impaired triglyceride storage in adipose tissue of insulin-resistant humans. *J. lipid Res.* 56 (2), 435–439. Epub 2014/11/ 25. doi:10.1194/jlr.M052860

Arner, E., Westermark, P. O., Spalding, K. L., Britton, T., Ryden, M., Frisen, J., et al. (2010). Adipocyte turnover: Relevance to human adipose tissue morphology. *Diabetes* 59 (1), 105–109. doi:10.2337/db09-0942

Arner, P., Andersson, D. P., Backdahl, J., Dahlman, I., and Ryden, M. (2018). Weight gain and impaired glucose metabolism in women are predicted by inefficient subcutaneous fat cell lipolysis. *Cell Metab.* 28 (1), 45–54. Epub 2018/06/05. doi:10. 1016/j.cmet.2018.05.004

Arner, P., Bernard, S., Salehpour, M., Possnert, G., Liebl, J., Steier, P., et al. (2011). Dynamics of human adipose lipid turnover in health and metabolic disease. *Nature* 478 (7367), 110–113. Epub 2011/09/29. doi:10.1038/nature10426

Bilson, J., Sethi, J. K., and Byrne, C. D. (2023). Heterogeneity of white adipocytes in metabolic disease. *Curr. Opin. Clin. Nutr. Metab. Care* 26 (2), 72–77. Epub 2023/02/03. doi:10.1097/MCO.0000000000885

Busch, R., Neese, R. A., Awada, M., Hayes, G. M., and Hellerstein, M. K. (2007). Measurement of cell proliferation by heavy water labeling. *Nat. Protoc.* 2 (12), 3045–3057. Epub 2007/12/15. doi:10.1038/nprot.2007.420

Cypess, A. M. (2022). Reassessing human adipose tissue. N. Engl. J. Med. 386 (8), 768–779. Epub 2022/02/24. doi:10.1056/NEJMra2032804

Danforth, E., Jr. (2000). Failure of adipocyte differentiation causes type II diabetes mellitus? *Nat. Genet.* 26 (1), 13. doi:10.1038/79111

DeLany, J. P., Dube, J. J., Standley, R. A., Distefano, G., Goodpaster, B. H., Stefanovic-Racie, M., et al. (2014). Racial differences in peripheral insulin sensitivity and mitochondrial capacity in the absence of obesity. J. Clin. Endocrinol. Metab. 99 (11), 4307–4314. Epub 2014/08/12. doi:10.1210/jc.2014-2512

Despres, J. P., Bouchard, C., Savard, R., Tremblay, A., Marcotte, M., and Theriault, G. (1984). The effect of a 20-week endurance training program on adipose-tissue morphology and lipolysis in men and women. *Metabolism* 33 (3), 235–239. Epub 1984/03/01. doi:10.1016/0026-0495(84)90043-x

Hoffstedt, J., Arner, E., Wahrenberg, H., Andersson, D. P., Qvisth, V., Lofgren, P., et al. (2010). Regional impact of adipose tissue morphology on the metabolic profile in morbid obesity. *Diabetologia* 53 (12), 2496–2503. Epub 2010/09/11. doi:10.1007/ s00125-010-1889-3

Johannsen, D. L., Tchoukalova, Y., Tam, C. S., Covington, J. D., Xie, W., Schwarz, J. M., et al. (2014). Effect of 8 weeks of overfeeding on ectopic fat deposition and insulin sensitivity: Testing the "adipose tissue expandability" hypothesis. *Diabetes care* 37 (10), 2789–2797. Epub 2014/07/12. doi:10.2337/dc14-0761

Lessard, J., Laforest, S., Pelletier, M., Leboeuf, M., Blackburn, L., and Tchernof, A. (2014). Low abdominal subcutaneous preadipocyte adipogenesis is associated with visceral obesity, visceral adipocyte hypertrophy, and a dysmetabolic state. *Adipocyte* 3 (3), 197–205. Epub 2014/07/30. doi:10.4161/adip.29385

Mauriege, P., Prud'Homme, D., Marcotte, M., Yoshioka, M., Tremblay, A., and Despres, J. P. (1997). Regional differences in adipose tissue metabolism between sedentary and endurance-trained women. *Am. J. Physiol.* 273, E497–E506. Epub 1997/10/08. doi:10.1152/ajpendo.1997.273.3.E497

McLaughlin, T., Craig, C., Liu, L. F., Perelman, D., Allister, C., Spielman, D., et al. (2016). Adipose cell size and regional fat deposition as predictors of metabolic response to overfeeding in insulin-resistant and insulin-sensitive humans. *Diabetes* 65 (5), 1245–1254. Epub 2016/02/18. doi:10.2337/db15-1213

McLaughlin, T., Lamendola, C., Coghlan, N., Liu, T. C., Lerner, K., Sherman, A., et al. (2014). Subcutaneous adipose cell size and distribution: Relationship to insulin resistance and body fat. *Obes. (Silver Spring)* 22 (3), 673–680. Epub 2013/05/15. doi:10.1002/oby.20209

McLaughlin, T., Sherman, A., Tsao, P., Gonzalez, O., Yee, G., Lamendola, C., et al. (2007). Enhanced proportion of small adipose cells in insulin-resistant vs insulinsensitive obese individuals implicates impaired adipogenesis. *Diabetologia* 50 (8), 1707–1715. Epub 2007/06/06. doi:10.1007/s00125-007-0708-y

Murphy, C. S., Liaw, L., and Reagan, M. R. (2019). *In vitro* tissue-engineered adipose constructs for modeling disease. *BMC Biomed. Eng.* 1, 27. Epub 2019/01/01. doi:10. 1186/s42490-019-0027-7

Neese, R. A., Misell, L. M., Turner, S., Chu, A., Kim, J., Cesar, D., et al. (2002). Measurement *in vivo* of proliferation rates of slow turnover cells by 2H2O labeling of the deoxyribose moiety of DNA. *Proc. Natl. Acad. Sci. U. S. A.* 99 (24), 15345–15350. Epub 2002/11/09. doi:10.1073/pnas.232551499

Pasarica, M., Xie, H., Hymel, D., Bray, G., Greenway, F., Ravussin, E., et al. (2009). Lower total adipocyte number but no evidence for small adipocyte depletion in patients with type 2 diabetes. *Diabetes care* 32 (5), 900–902. Epub 2009/02/21. doi:10.2337/dc08-2240

Sethi, J. K., and Vidal-Puig, A. J. (2007). Thematic review series: Adipocyte biology. Adipose tissue function and plasticity orchestrate nutritional adaptation. *J. lipid Res.* 48 (6), 1253–1262. Epub 2007/03/22. doi:10.1194/jlr.R700005-JLR200

Spalding, K. L., Arner, E., Westermark, P. O., Bernard, S., Buchholz, B. A., Bergmann, O., et al. (2008). Dynamics of fat cell turnover in humans. *Nature* 453 (7196), 783–787. Epub 2008/05/06. doi:10.1038/nature06902

Stanford, K. I., Middelbeek, R. J., Townsend, K. L., Lee, M. Y., Takahashi, H., So, K., et al. (2015). A novel role for subcutaneous adipose tissue in exercise-induced improvements in glucose homeostasis. *Diabetes* 64 (6), 2002–2014. Epub 2015/01/ 22. doi:10.2337/db14-0704

Strawford, A., Antelo, F., Christiansen, M., and Hellerstein, M. K. (2004). Adipose tissue triglyceride turnover, de novo lipogenesis, and cell proliferation in humans measured with 2H2O. *Am. J. Physiol. Endocrinol. Metab.* 286 (4), E577–E588. Epub 2003/11/06. doi:10.1152/ajpendo.00093.2003

Strissel, K. J., Stancheva, Z., Miyoshi, H., Perfield, J. W., 2nd, DeFuria, J., Jick, Z., et al. (2007). Adipocyte death, adipose tissue remodeling, and obesity complications. *Diabetes* 56 (12), 2910–2918. Epub 2007/09/13. doi:10.2337/db07-0767

Tchoukalova, Y. D., Votruba, S. B., Tchkonia, T., Giorgadze, N., Kirkland, J. L., and Jensen, M. D. (2010). Regional differences in cellular mechanisms of adipose tissue gain with overfeeding. *Proc. Natl. Acad. Sci. U. S. A.* 107 (42), 18226–18231. Epub 2010/10/06. doi:10.1073/pnas.1005259107

Virtue, S., and Vidal-Puig, A. (2010). Adipose tissue expandability, lipotoxicity and the Metabolic Syndrome-an allostatic perspective. *Biochim. Biophys. Acta* 1801 (3), 338–349. Epub 2010/01/09. doi:10.1016/j.bbalip.2009.12.006

Weyer, C., Foley, J. E., Bogardus, C., Tataranni, P. A., and Pratley, R. E. (2000). Enlarged subcutaneous abdominal adipocyte size, but not obesity itself, predicts type II diabetes independent of insulin resistance. *Diabetologia* 43 (12), 1498–1506. Epub 2001/ 01/11. doi:10.1007/s001250051560

White, U., Beyl, R. A., and Ravussin, E. (2022). A higher proportion of small adipocytes is associated with increased visceral and ectopic lipid accumulation during weight gain in response to overfeeding in men. *Int. J. Obes. (Lond).* 46 (8), 1560–1563. Epub 2022/05/23. doi:10.1038/s41366-022-01150-y

White, U., and Ravussin, E. (2019). Dynamics of adipose tissue turnover in human metabolic health and disease. *Diabetologia* 62 (1), 17–23. Epub 2018/09/30. doi:10.1007/s00125-018-4732-x

White, U. A., Fitch, M. D., Beyl, R. A., Hellerstein, M. K., and Ravussin, E. (2017). Association of *in vivo* adipose tissue cellular kinetics with markers of metabolic health in humans. *J. Clin. Endocrinol. Metab.* 102, 2171–2178. doi:10.1210/jc.2016-4000

White, U. A., Fitch, M. D., Beyl, R. A., Hellerstein, M. K., and Ravussin, E. (2016). Differences in *in vivo* cellular kinetics in abdominal and femoral subcutaneous adipose tissue in women. *Diabetes* 65, 1642–1647. Epub 2016/03/20. doi:10.2337/db15-1617

White, U. A., Fitch, M. D., Beyl, R. A., Hellerstein, M. K., and Ravussin, E. (2018). Racial differences in *in vivo* adipose lipid kinetics in humans. *J. lipid Res.* 59 (9), 1738–1744. Epub 2018/06/19. doi:10.1194/jlr.P082628