



# Transcriptome Response and Spatial Pattern of Gene Expression in the Primate Subventricular Zone Neurogenic Niche After Cerebral Ischemia

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The main stem cell niche for neurogenesis in the adult mammalian brain is the subventricular zone (SVZ) that extends along the cerebral lateral ventricles. We aimed at characterizing the initial molecular responses of the macaque monkey SVZ to transient, global cerebral ischemia. We microdissected tissue lining the anterior horn of the lateral ventricle (SVZa) from 7 day post-ischemic and sham-operated monkeys. Transcriptomics shows that in ischemic SVZa, 541 genes were upregulated and 488 genes were down-regulated. The transcription data encompassing the upregulated genes revealed a profile typical for quiescent stem cells and astrocytes. In the primate brain the SVZ is morphologically subdivided in distinct and separate ependymal and subependymal regions. The subependymal contains predominantly neural stem cells (NSC) and differentiated progenitors. To determine in which SVZa region ischemia had evoked transcriptional upregulation, sections through control and ischemic SVZa were analyzed by high-throughput *in situ* hybridization for a total of 150 upregulated genes shown in the [www.monkey-niche.org](http://www.monkey-niche.org) image database. The majority of the differentially expressed genes mapped to the subependymal layers on the striatal or callosal aspect of the SVZa. Moreover, a substantial number of upregulated genes was expressed in the ependymal layer, implicating a contribution of the ependyma to stem cell biology. The transcriptome analysis yielded several novel gene markers for primate SVZa including the apelin receptor that is strongly expressed in the primate SVZa niche upon ischemic insult.

**Keywords:** neural stem cell, global ischemia, non-human primate, subventricular zone, cell marker

**Abbreviations:** Bv, blood vessel; CC, corpus callosum; CG, cingulate gyrus; CSVZ, the callosal aspect of the subventricular zone; EL, ependymal layer; FrCx, frontal cortex; GO, Gene ontology; GSEA, gene set enrichment analysis; ISH, *in situ* hybridization; LV, lateral ventricle; NSC, neural stem cell, PVSEL, perivascular subependymal layer; SEL, subependymal layer on the striatal side; SPT, septum; STR, striatum; SVZa, subventricular zone along the anterior horn of LV; TAP, transit amplifying progenitor; Th, thalamus; aNSC, activated NSC; qNSC, quiescent NSC.

## INTRODUCTION

Neurogenesis in the adult mammalian brain takes place in two niches, the subventricular zone (SVZa) lining the anterior horn of the forebrain lateral ventricle and the subgranular zone (SGZ) of the dentate gyrus. In these microenvironments, astrocyte-like neural stem cells (NSCs) proliferate and produce neural precursors generating neuronal and glial progeny (Obernier and Alvarez-Buylla, 2019). Neurogenic niches host a heterogeneous cellular population that includes NSCs at different stages of development: quiescent NSCs (qNSCs), active NSCs (aNSCs), transit-amplifying progenitors (TAPs), and neuroblasts (NBs) (Bond et al., 2015; Obernier and Alvarez-Buylla, 2019). Single-cell transcriptome analyses of the mouse SVZ have revealed gene expression patterns characteristic for each of these stages but have also uncovered additional intermediate states (Beckervordersandforth et al., 2010; Codega et al., 2014; Llorens-Bobadilla et al., 2015; Dulken et al., 2017; Zywitzka et al., 2018).

The knowledge on transcriptome-wide gene expression in the primate SVZa is limited. Most such studies have mapped gene expression in diverse brain regions during development and in the adult (Bernard et al., 2012; Hawrylycz et al., 2012; Miller et al., 2014; Bakken et al., 2015, 2016), but few studies have focused on gene expression in the neurogenic niches (Miller et al., 2013; Foret et al., 2014; Sandstrom et al., 2014). A comparison of the adult SGZ transcriptomes of mice and rhesus monkeys reported a conservation of the molecular signatures of precursors involved in hippocampal neurogenesis (Miller et al., 2014). Transcriptome analyses of isolated SVZa precursors from adult baboon monkeys revealed genes regulated by active (H3K4me3) or repressive (H3K9me3) epigenetic marks (Foret et al., 2014; Sandstrom et al., 2014).

Brain ischemia is a major cause of death and disability. Several experimental models have been used to study the response of the brain to ischemia, but only a few of those use non-human primates (Tajiri et al., 2013). In ischemic rodent models, the NSCs attempt to replenish lost or damaged cells by enhancing NSC proliferation followed by neuron production (Marques et al., 2019). The capacity of the primate brain to undergo reparative processes is more limited than in rodents (Alunni and Bally-Cuif, 2016). Understanding the molecular cause which underlies this difference between primates and rodents is an important step in the development of strategies for restoring human brain functions after injury (Jessberger and Gage, 2014). We have previously established a model of transient global ischemia in adult macaque monkey. Such ischemia induces regional tissue damage affecting neuronal populations in the hippocampus, striatum, neocortex and cerebellum (Yamashima et al., 1996; Yoshida et al., 2002). In this model, we have observed an enhanced proliferation of bromodeoxyuridine (BrdU) labeled progenitor cells in SVZa, in the second postischemic week (Tonchev et al., 2003, 2005).

To study gene expression changes preceding the onset of proliferation in the SVZa, we investigated the transcriptome-wide response of the SVZa niche following global brain ischemia. Using a combination of transcriptomics and high-throughput *in situ* hybridization (ISH), we examined the changes in gene

expression in the SVZa and its subregions at postischemic day 7. We found that ischemic SVZa shows a transcriptional profile reminiscent of a profile typical for quiescent stem cells, astrocytes and oligodendrocytes. In addition, we determined by ISH the expression of 150 genes in control and ischemic SVZa, digital images are freely available online<sup>1</sup>. A comparative analysis of the expression pattern these 150 genes yielded several novel gene markers for SVZa cells, including the apelin receptor (*APLNR*).

## MATERIALS AND METHODS

### Subjects

Experiments with the 7 monkeys used were approved by the Animal Care and Ethics Committee of Kanazawa University, Japan (Approval protocols AP-031498 and AP-080920). The monkeys (5 female and 2 male *Macaca fuscata*) were kept in air-conditioned cages and had free daily access to food and water. The monkeys were 4–6 years of age at the time of the experiments. Six monkeys were subjected to surgical procedures to induce global brain ischemia or to a sham operation, and sacrificed 7 days later. One additional monkey which was not operated received daily injections (100 mg/kg, *i.v.*) of 5-bromo-2'-deoxyuridine (BrdU, Sigma-Aldrich Corp., St. Louis, MO, United States) for 5 consecutive days. This monkey was sacrificed 2 h after the last BrdU injection.

Optimization of the ISH conditions were performed on sections from *Macaca mulatta* brain tissues obtained from the German Primate Center (Göttingen, Germany). All animals were offsprings of monkeys that have been bred in captivity. The animals were kept under the regulations for non-human primates by the guidelines for the accommodation and care of animals used for experimental and other scientific purposes (2007/526/EC; Appendix A ETS 123).

Mice were sacrificed according to the German Law on Animal Welfare. Sacrifice is licensed by the Veterinary authorities of Göttingen, Germany (392000\_2a/Si/rö, 09/12/2013).

### Surgical Procedures and SVZa Microdissection

Three monkeys underwent transient whole brain ischemia, and 3 monkeys were subjected to a sham surgery (Yamashima et al., 1996; Yoshida et al., 2002; Tonchev et al., 2003, 2005). The brachiocephalic trunk and left subclavian arteries were clipped for 18 min (**Supplementary Figure 1A**). At day 7 after the sham or ischemic surgery, the monkeys were anesthetized with a lethal dose of sodium pentobarbital and intracardially perfused with 0.5L cooled sterile saline. Within less than 20 min from the start of the perfusion, craniotomy was performed and the whole brain was extracted, sectioned into two hemispheres through the corpus callosum, and each hemisphere was snap-frozen in liquid nitrogen and stored at  $-80^{\circ}\text{C}$ . The right hemisphere was used for the subsequent analyses. In the monkey brain, the lateral ventricle spans more than 30 mm, between levels with atlas coordinates +35 mm rostrally to +2 mm

<sup>1</sup>www.monkey-niche.org

caudally (Saleem and Logothetis, 2007). The right hemisphere was sectioned into 5 mm-thick coronal slices. The slice which included the anterior commissure and the head of the caudate nucleus (atlas coordinates +20 mm to +25 mm (Saleem and Logothetis, 2007) was placed under a stereomicroscope and three adjacent tissue fragments (~1 mm × 0.2 mm, 3 mm thick) were dissected with a microscalpel along the striatal side of the right lateral ventricle (**Supplementary Figures 1B1–B3**). The callosal side of the lateral ventricle was not included in the excised tissue sample. All steps were performed on dry ice to avoid complete defreezing and RNA degradation within the samples. The dissected specimens were transferred to an Eppendorf tube, placed in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  until the RNA extraction step.

## RNA Sequencing (RNA Seq) and Data Analysis

### RNA Seq

RNA Seq was performed at the Transcriptome and Genome Analysis Laboratory (TAL, University of Göttingen). The total RNA from each SVZa sample was extracted using the phenol-chloroform extraction protocol and 1  $\mu\text{g}$  of total RNA was converted into Illumina sequencing libraries using the TruSeq RNA Sample Preparation Kit (Illumina, RS-122-2002). The size range of the final cDNA libraries were determined using Bioanalyzer 2100 (Agilent Technologies; Santa Clara, California, United States). The generated cDNA libraries were then amplified and sequenced using the cBot and HiSeq2000 (Illumina Inc., San Diego, California, United States) (SR; 1 × 50 bp; ~ 30 million reads per sample). Finally, the sequence images were transformed to *per* read base call files (fastq files) using the Illumina software BaseCaller and CASAVA v1.8.2. RNA-Sequencing data have been deposited to GEO under the accession ID GSE136036.

We checked the RNA Seq data *via* FastQC quality control tool (v.0.10.0, Babraham Bioinformatics). Raw sequence reads were aligned to the *Macaca mulatta* genome (ensembl 73) by TopHat bioinformatics tool (V2.0.8 custom parameters:  $-\text{no-coverage-search}$ ) (Trapnell et al., 2009). The gene annotation was downloaded from RhesusBase v2.0<sup>2</sup>. This database uses all published Macaque RNA-seq data to refine ensembl Macaque gene annotation. The reads number mapped to each gene were calculated using bedtools (v2.16.2 default parameters) (Quinlan and Hall, 2010).

In order to reduce the false discovery rate due to variations between monkeys, we applied two methods to identify genes differentially expressed after ischemia. First, read numbers were normalized to RPKM (reads per kilobase per million). Since the reads mapped to known transcripts can differ significantly, we used the total number of reads mapped to a known transcript to do normalization. We compared the 9 sham-operated and the 9 ischemic samples with the paired *T*-test to calculate the differentially expressed genes. Second, the R package DESeq was applied to normalize the reads counts and calculate the significance of the differential expression, taking the batch effect into account. The paired *T*-test *p*-value < 0.05, DESeq *p*-value < 0.05, and log<sub>2</sub>-transformed fold change > 0.5 were

<sup>2</sup><https://web.archive.org/web/20190414050927/http://www.rhesusbase.org/>

used as criteria to select the differentially expressed genes with an FDR = 2.1%. It was calculated as the ratio between the number of differentially expressed genes by chance, which is estimated by the median number of significant genes for 1000 random permutation of samples, and the number of significant genes of real data under the same criterion.

### Functional Enrichment Analysis of Monkey SVZa-DE Genes

The lists of monkey SVZa-DE-UP or SVZa-DE-DOWN genes were uploaded to the DAVID functional annotation tool<sup>3</sup>. Then, representative enriched biological functional terms were selected. To perform the Gene Set Enrichment Analysis (GSEA) analysis, the combined list of SVZa-DE-UP and SVZa-DE-DOWN genes was uploaded to GSEA. Using FDR *q*-value < 0.05 as a cut-off generated 166 enriched biological and cellular component terms. The Fisher's exact test was applied to identify the terms showing a statistically significant difference for the SVZa-DE-UP or the SVZa-DE-DOWN genes.

### Protein-Protein Interaction Network

The combined list of monkey SVZa-DE-UP and SVZa-DE-DOWN genes was uploaded to the STRING database<sup>4</sup>, then protein-protein interaction from STRING were visualized by Cytoscape<sup>5</sup>.

### Non-radioactive Automated ISH

Automated ISH was performed on frozen sections (Lein et al., 2007). Coronal tissue blocks approx. 2 × 2 cm in size, which included the caudate nucleus (**Supplementary Figures 1B1,B2**), were dissected from the right hemisphere. The blocks were embedded in Tissue-Tek O.C.T (Sakura), quick frozen, and sectioned at 20- $\mu\text{m}$  thickness. Hybridization was performed on a Tecan (Männedorf, Switzerland) ISH robot as previously described (Lein et al., 2007). Macaque monkey specific templates, 700–1000 nucleotides long, were synthesized. Primer and template and sequences are available online at <http://www.monkey-niche.org>. Digoxigenin-tagged riboprobes, were generated by *in vitro* transcription. Hybridized probes were detected by a two-step chromogenic catalyzed reporter deposition. For FISH staining, the hybridized probe for *APLNR* was visualized using fluorochrome-conjugated reagents. The mouse *Aplnr* mRNA (Entrez Gene reference: NM\_011784.3) was visualized using SP6/T7 primers with a total length of the template 649 of nucleotides. The template was generated using the following specific primers: Forward: TCTAGGCACCACAGGCAATG; Reverse: GGCTACTACAAGCACCACGA.

### Visualization of Colorimetric ISH and Atlas Generation

After colorimetric ISH, slides were cover-slipped and digitally scanned at 0.501  $\mu\text{m}$ /pixel using an Aperio ScanScope AT2 whole slide scanner (Leica Biosystems, Wetzlar, Germany). Images were

<sup>3</sup><https://david.ncifcrf.gov/>

<sup>4</sup><http://string-db.org/>

<sup>5</sup><https://cytoscape.org/>

cropped and stored in Aperio SVS format. Next the images were converted into a zoomable image pyramid format (Zoomify <http://www.zoomify.com/>) with Libvips (Martinez and Cupitt, 2005); <https://libvips.github.io/libvips/>, and the OpenSlide library (Satyanarayanan et al., 2013)<sup>6</sup>. The metadata was extracted from the glass slide label via Optical Character Recognition (Tesseract <https://github.com/tesseract-ocr/tesseract>), checked and stored in a MySQL Database together with the standard image data. The database is accessed using a web interface and the images are rendered via a custom implementation of the Openseadragon viewer<sup>7</sup> which additionally features an image comparison mode. The atlas coordinates of the images shown in the database are according to Saleem and Logothetis, 2007. The atlas coordinates of the sections shown in **Figures 3D–F, 4D–K, 5, 6C–F** are provided in **Supplementary Table S1**.

## Quantitative Measurements of Colorimetric ISH

To quantify the gene expression in the SVZa, we used the custom software Celldetekt (version 2.7; Carson et al., 2005; <https://github.com/tumrod/cellDetekt>). Celldetekt is a python script<sup>8</sup> that uses the Python Imaging Library<sup>9</sup>. The input images for analysis were derived from the *monkey-niche.org* database and imported into Celldetekt. The expression signal strength for ischemic sections was compared to the control and the fold increase is shown in the bar diagrams of **Figures 3D–F, 4D–K, 5, 6C–F**. Celldetekt locates the cells on the images and estimates the level of expression in each cell. The cells are classified according to the ISH precipitates in each cell as: (i) level 3 cells are filled with dye precipitate, (ii) level 2 cells are partially filled with precipitate, (iii) level 1 cells are characterized by scattered minute particles of deposit, and (iv) level 0 cells lack detectable precipitate (Carson et al., 2005). The cells in the RoIs (CSVZ and SEL) were evaluated within an area of 100  $\mu\text{m}$  in width  $\times$  1500  $\mu\text{m}$  in length, starting from the dorsal tip of the lateral ventricle (**Supplementary Figure 2**). The cells in the EL were evaluated within the thickness of the ependymal layer along a length of 1500  $\mu\text{m}$ , starting from the dorsal tip of the lateral ventricle. The number of level 2 and 3 cells out of the number of all cells was calculated as a ratio for one sham-operated and one postischemic SVZa.

## Immunohistochemistry

Immunofluorescence stainings of monkey specimens were performed following the FISH labeling of *APLNR*. The slides were washed 3  $\times$  5 min in PBS and subjected to antigen retrieval using the Vector Antigen Unmasking Solution (Cat. No. H-3300, Vectorlabs, Burlingame, CA, United States). The slides were incubated in a microwave oven at 800 W for 3  $\times$  5 min, each cycle being followed by a cooling step on ice for 30 min. The slides were then washed in PBS 3  $\times$  5 min

and blocked for 1 h in 10% normal goat serum (Cat. No S-1000, Vectorlabs) in PBS with 0.1% Triton-X100. The primary antibodies were applied in blocking solution overnight at 4°C. On the next day, the sections were washed in PBS 3  $\times$  5 min and incubated for 2 h at room temperature in the respective secondary antibody conjugated to AlexaFluor-488 or AlexaFluor-647 fluorochromes (Thermo Fisher Scientific, Germany). The following primary antibodies were used: rat anti-BrdU (1:100, Cat. No Ab6326, Abcam, Cambridge, United Kingdom); mouse anti-GFAP (1:400; Cat. No M0761, Dako-Agilent Technologies GmbH, Hamburg, Germany); chick anti-GFAP (1:1000; Cat. No AB5541, Merck Millipore); rabbit anti-GLUT1/SLC2A1 (1:100; Cat. No HPA031345, Sigma-Aldrich); mouse anti-VIM (1:1000; Cat. No MAB3400, Merck Millipore).

## Microscopy on Fluorescently Labeled Sections and Image Analysis

The slides labeled by FISH and immunofluorescence were imaged using a fully motorized wide-field epifluorescence microscope Zeiss AxioImager Z.2 (Carl Zeiss GmbH) with an AxioCam Mrm rev.3 monochrome CCD camera (Carl Zeiss GmbH), and AxioVision v.4.9 software. A set of 3–5 non-overlapping randomly selected Z-stacks within the SEL of each section were captured through an EC Plan-Neofluar objective 40 $\times$ /0.75 at resolution of 0.325  $\mu\text{m}$ /pixel and Z-axial resolution of 0.55  $\mu\text{m}$ . The number of sections visualized and the number of monkeys from which data are derived are listed in **Supplementary Table S14**.

The camera was set to a binning factor 2  $\times$  2 to reduce both scanning time and camera noise. Shading from the irregular illumination field was corrected during the acquisition via the camera built-in shading correction using prerecorded illumination field images from fluorescent test slides. All camera settings for the different channels were kept constant. The exposure time was allowed to vary only when this was needed to avoid saturation. The images were saved.zvi format (Zeiss) of the AxioVision software, stitched together, and exported to a 16 bit TIFF format for further processing and analysis as described in the following section.

## Quantitation of Cells on Sections Labeled by Multiple Fluorescence Dyes

Because of the large size of the primate SEL, analysis of cell identity required screening thousands of cells. We therefore developed a semi-automated digital image cytometry workflow (Wahlby, 2003; Chieco et al., 2013). The process includes the following steps: (1) Preprocessing, including channel extraction and image denoising, (2) Top-Hat morphological filtering for improving signal/noise ratio, (3) Nuclear segmentation, (4) Channel thresholding, (5) Classification by a pretrained decision tree and counting of the RoIs. This algorithm for the analysis of co-expression might be useful to those who aim at scanning entire brain regions in large-sized brains, such as the primate brain.

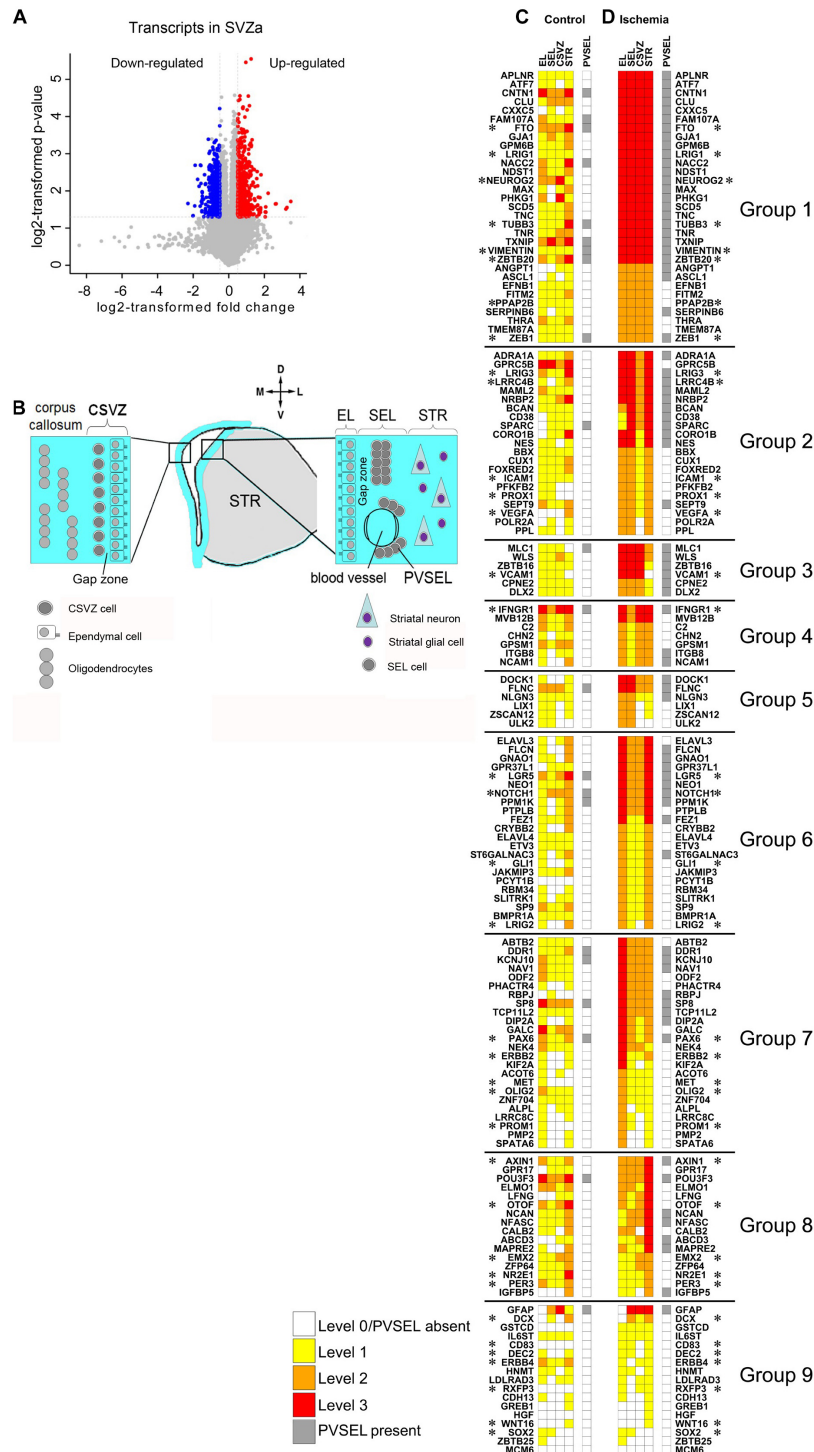
The goal of the workflow was to reduce the operator introduced bias by creating an automated algorithm for

<sup>6</sup><https://openslide.org/>

<sup>7</sup><https://openseadragon.github.io/>

<sup>8</sup><http://www.python.org/>

<sup>9</sup><http://www.pythonware.com/>



**FIGURE 1 |** Molecular changes in the neurogenic niche induced by ischemia. **(A)** Volcano plot displays changes in gene expression in monkey SVZa. Transcripts up- or down-regulated and those unchanged are in red or blue and gray. **(B)** Cellular architecture of SVZa on the striatal and the callosal side of the lateral ventricle. For details see Text. **(C,D)** Heat maps illustrating gene expression changes of 150 genes in EL, SEL, CSVZ, STR and PVSEL upon ischemia. The 36 genes implicated in stem cell biology are marked (\*). Original data are on “monkey-niche.org.” Expression of 150 genes, deposited in the “monkey-niche” database (**Supplementary Table S7**) is shown for five regions of interest (**Figure 1B**). The genes in **(C,D)** are grouped according to their postischemic upregulation: Group 1 (markedly enhanced expression levels in all Rols), Group 2 (predominantly upregulated in the Rols on the striatal side of the ventricle), Group 3 (strongly induced in EL, SEL and CSVZ), Group 4 (strongly induced in EL, CSVZ and STR), Group 5 (strongly induced in EL and SEL), Group 6 (strongly induced in EL and STR), Group 7 (strongly induced in EL), Group 8 (strongly induced in STR), Group 9 (genes with low level of expression or not belonging to the other groups).

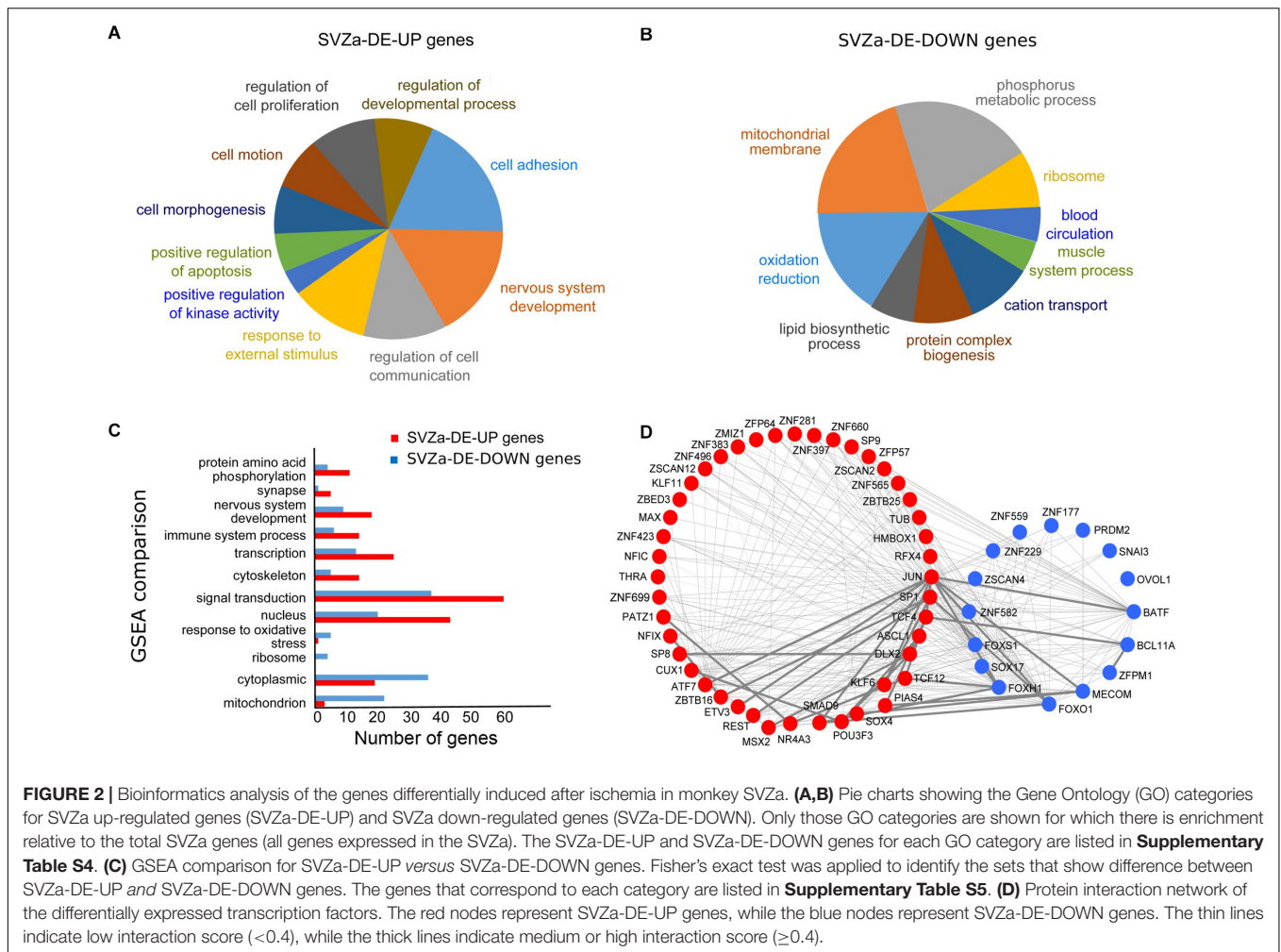


image processing and analysis. We applied a unified, reproducible approach to a set of images with variable brightness distributions of the signal or the background. The workflow includes the following major steps following the image acquisition:

### Preprocessing

Generate a projection of the z-stack on one all-in-focus plane using the Smooth manifold extraction (SME) algorithm (Shihavuddin et al., 2017) as implemented in the SME-projection plugin for Fiji/ImageJ<sup>10</sup>. Perform image denoising using the Non-Local Means algorithm as implemented in the Fiji/ImageJ software for image processing and analysis (Buades et al., 2011; Schindelin et al., 2012); channel extraction and sorting into one of the following channel classes: nuclear antigen, cytoplasmic antigen, mRNA FISH cytoplasmic channel, or DAPI channel.

### Top-Hat Morphological Filtering

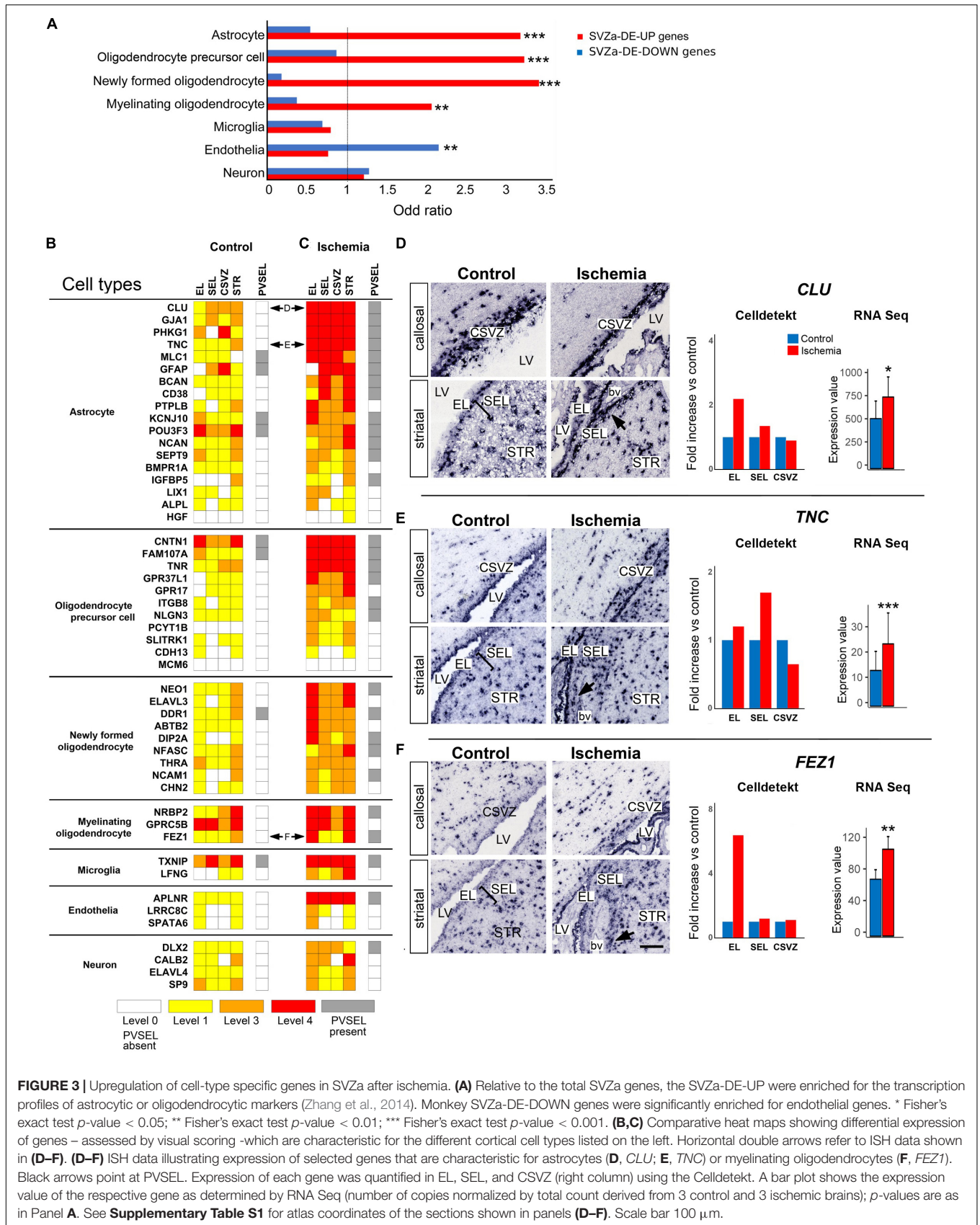
Top-Hat morphological filtering by reconstruction of each channel to suppress background and improve the signal-to-noise

<sup>10</sup><https://github.com/biocompibens/SME/>

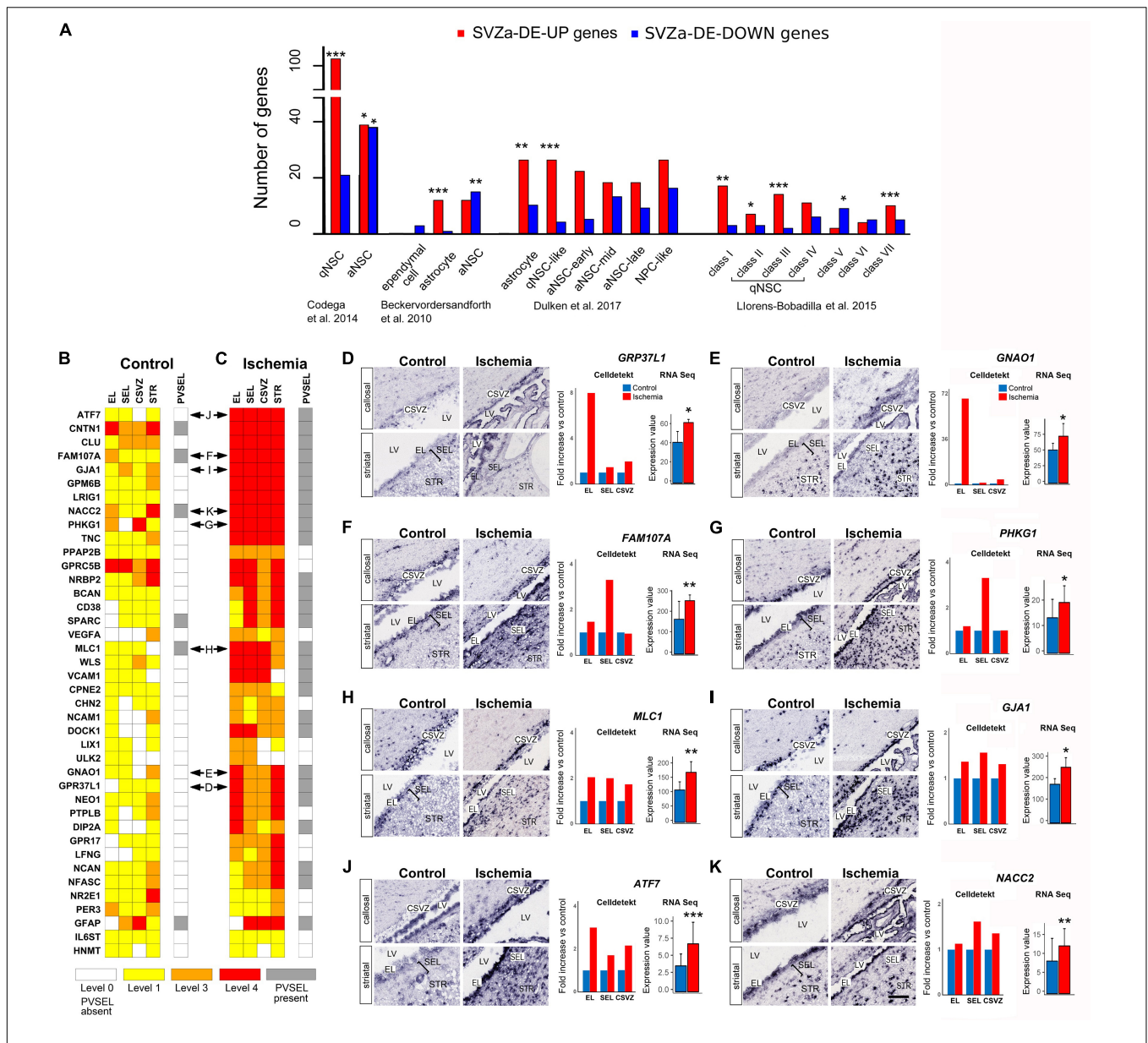
(S/N) ratio. The Top-Hat (“top minus hat”) procedure subtracts an estimate of the background developed by a morphological opening from the original image. To eliminate the influence of dark background variability we added a step of morphological closing with a structuring element of the same size before the opening. This eliminates dark variations and the final image includes only the bright parts in the image that stand above the background. Instead of the classical gray-scale morphological filtering we used reconstructions of the opening and the closing thus reducing further image artefacts arising from the anisotropy of the structuring elements (SEs) (Soille, 1999; Legland et al., 2016). Each channel class (nuclear antigen, cytoplasmic antigen, mRNA FISH cytoplasmic channel, or DAPI channel) was filtered with a disk-shaped SE, of size appropriate for the distributions of signals. The sizes were selected with the help of digital granulometry of the preprocessed channel images (Legland et al., 2016).

### Definition of Cellular Masks

After the top-hat filtering the DAPI channels were processed with the CellProfiler 3 (Bray et al., 2015) to detect, declump



**FIGURE 3 |** Upregulation of cell-type specific genes in SVZa after ischemia. **(A)** Relative to the total SVZa genes, the SVZa-DE-UP were enriched for the transcription profiles of astrocytic or oligodendrocytic markers (Zhang et al., 2014). Monkey SVZa-DE-DOWN genes were significantly enriched for endothelial genes. \* Fisher's exact test  $p$ -value < 0.05; \*\* Fisher's exact test  $p$ -value < 0.01; \*\*\* Fisher's exact test  $p$ -value < 0.001. **(B,C)** Comparative heat maps showing differential expression of genes – assessed by visual scoring – which are characteristic for the different cortical cell types listed on the left. Horizontal double arrows refer to ISH data shown in **(D–F)**. **(D–F)** ISH data illustrating expression of selected genes that are characteristic for astrocytes **(D, *CLU*;** **E, *TNC*)** or myelinating oligodendrocytes **(F, *FEZ1*)**. Black arrows point at PVSEL. Expression of each gene was quantified in EL, SEL, and CSVZ (right column) using the Celldetekt. A bar plot shows the expression value of the respective gene as determined by RNA Seq (number of copies normalized by total count derived from 3 control and 3 ischemic brains);  $p$ -values are as in Panel A. See **Supplementary Table S1** for atlas coordinates of the sections shown in panels **(D–F)**. Scale bar 100  $\mu$ m.

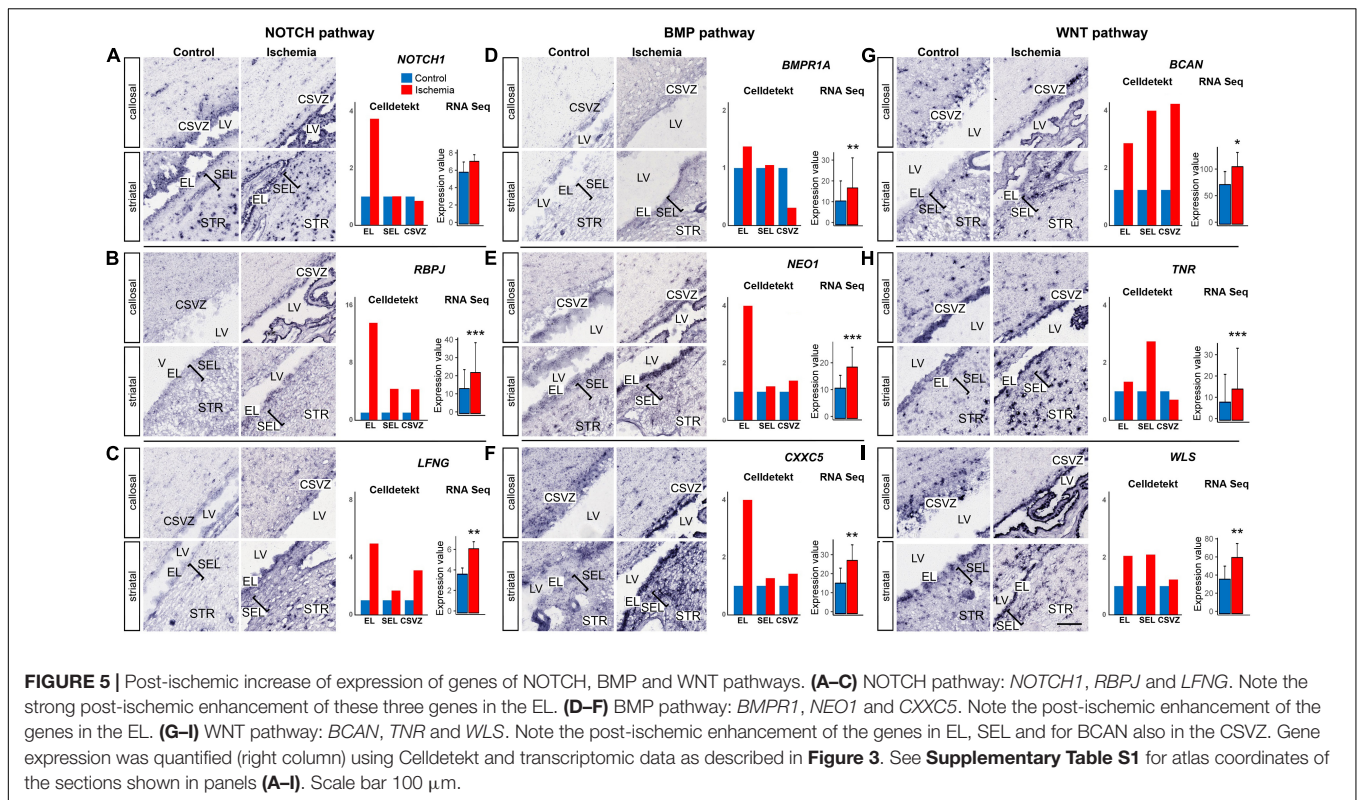


and export binary masks of nuclear connected regions. The unprocessed images were used to manually select areas 150  $\mu$ m below the ependymal layer. All nuclear masks within these bounds were extracted, diluted under restriction to avoid overlap by  $\sim 3$   $\mu$ m and saved as collections of labeled cellular masks for the analysis of the cells in each image.

## Channel Thresholding

The images for analysis were thresholded in two steps. First the top-hat images were Z-transformed – the gray value of each pixel was replaced by its Z-score and all pixels with values  $\leq 1$ , i.e., less than the mean plus one standard deviation, were set to 0. Next these images were thresholded with Li's Minimum Crossentropy algorithm (Li and Tam, 1998) as implemented in Fiji/ImageJ.





## Automatic Classification and Counting of the Cellular Labels

We used a simple decision tree which classifies the cellular labels into positive and negative considering the fraction of the area covered by the corresponding thresholded signal. The limits for acceptance of positivity were selected after a receiver operating characteristic (ROC) analysis of  $\sim 300$  cells classified by an experienced researcher. All used imageJ macro scripts and the steps of CellProfiler pipeline are available in an online github repository<sup>11</sup>.

## RESULTS

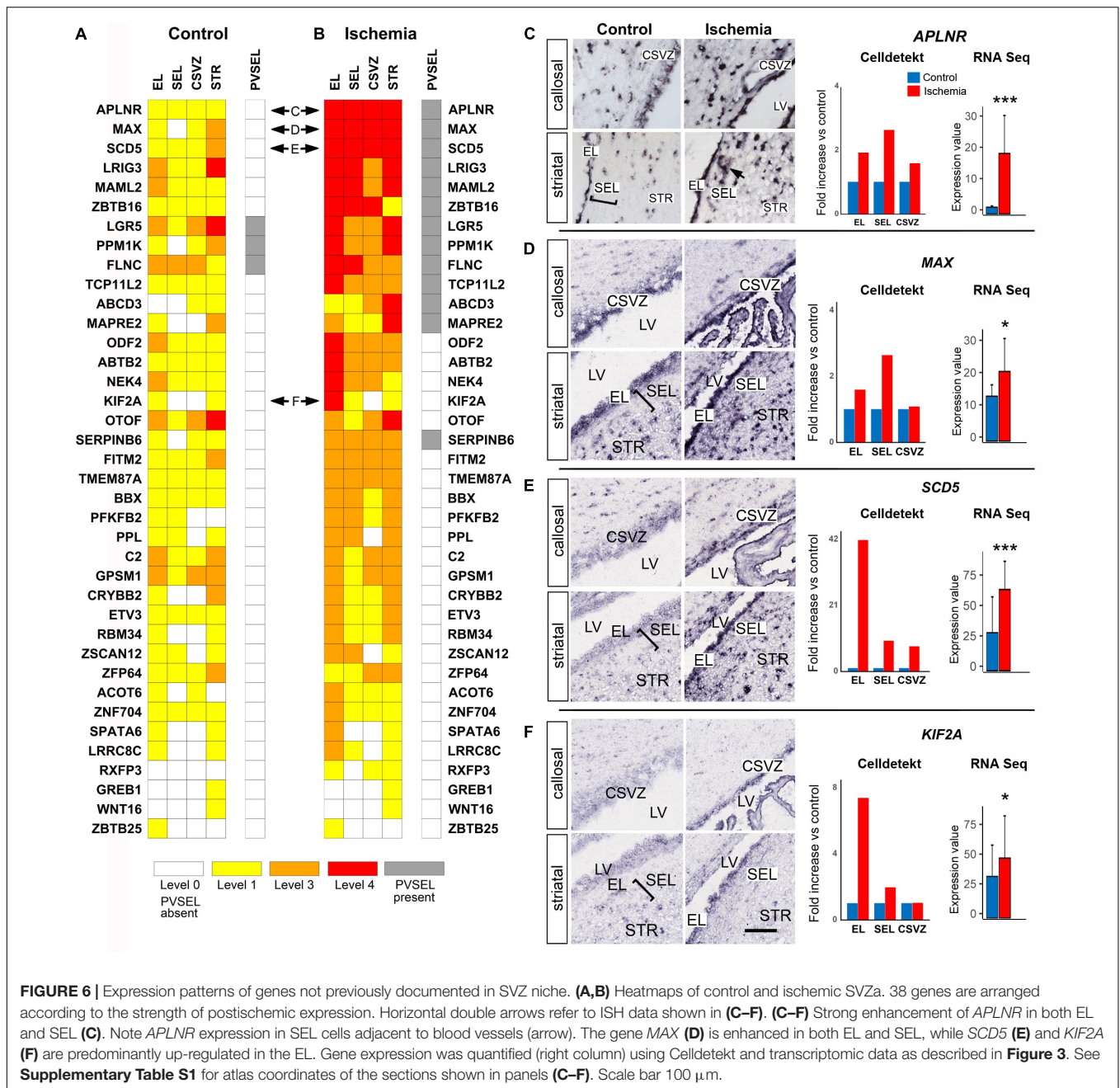
### The Transcriptome Changes in the SVZa Niche Following Ischemia

To uncover the changes in gene expression upon ischemia in the macaque SVZa niche, we performed transient global cerebral ischemia in adult macaque monkeys. In both sham-operated and ischemic tissue  $\sim 25,000$  transcripts corresponding to  $\sim 16,000$  genes were detected (**Supplementary Table S2**). Upon ischemia a total of 1554 transcripts were differentially expressed (DE), 816 (541 genes) of which were upregulated (SVZa-DE-UP genes) and 738 transcripts (488 genes) were down regulated (SVZa-DE-DOWN genes) (**Figure 1A** and **Supplementary Table S3**). Gene ontology (GO) and gene set enrichment analysis (GSEA) revealed that SVZa-DE-UP and SVZa-DE-DOWN genes

encompass distinct functional groups (**Figures 2A–C**). One third (185) of the SVZa-DE-UP genes were associated with GO biological process categories that were over-represented when compared to the total SVZa genes. These SVZa-DE-UP genes were enriched in “cell adhesion”, “nervous system development”, “regulation of cell communication”, “response to external stimulus”, and “cell proliferation” (**Figure 2A** and **Supplementary Table S4**). A GSEA analysis showed enrichment in SVZa-DE-UP genes, in the categories “signal transduction”, “nervous system development”, “cytoskeleton”, and “protein amino acid phosphorylation” (**Figure 2C** and **Supplementary Table S5**).

We looked for transcriptional regulators, which were differentially expressed in SVZa after ischemia. We identified 45 upregulated transcription factors corresponding to 18 protein families and 17 down-regulated transcription factors corresponding to 8 protein families (**Supplementary Table S6**). C2H2 type zinc finger protein family was the most enriched protein family, with 18 upregulated and 11 down-regulated genes, followed by bHLH and homeobox protein families with 5 upregulated genes each. Next we determined which of the transcription factors interact physically or functionally using the STRING database (see foot note 4). This revealed a highly interconnected network formed by both upregulated and down-regulated factors (**Figure 2D**). Several transcription factors formed a network hub, including *JUN*, *DLX2*, *ASCL1*, *SOX4*, *BATF*, *FOXS1*, *SOX17*, and *FOXH1* with *JUN* in the center of the network. This raises the possibility that the components of this transcription factor network are involved in regulating aspects of

<sup>11</sup>[https://github.com/StoyanPv/MKY-FISH\\_scripts](https://github.com/StoyanPv/MKY-FISH_scripts)



the response of the SVZa to ischemia. Such *in silico* interaction data need to be tested for physical or functional interaction.

In the group of the 488 SVZa-DE-DOWN genes, GO analysis revealed that 143 genes were over-represented in the categories “oxidation reduction,” “mitochondrial membrane,” “cation transport,” and “ribosome” (**Figure 2B** and **Supplementary Table S4**). Consistent with the GO results, the GSEA analysis showed enrichment of the gene groups “response to oxidative stress,” and “mitochondrion” (**Figure 2C** and **Supplementary Table S5**). The genes annotated by these GO terms typically included genes involved in mitochondrial function and their down-regulation following ischemia most probably reflects

the dysfunction of mitochondria under hypoxic conditions (Solaini et al., 2010).

### **In situ Analysis of Gene Expression Changes After Ischemia**

The transcriptomic analysis provides a global view of which gene expression levels change after an ischemic insult. To anchor this change to cells and tissues in their natural environment we performed colorimetric ISH on coronal sections through the SVZa (**Supplementary Figures 1B1,B2**). We generated 150 macaque-specific riboprobes that were hybridized to sections using a flow-through robotic system (Lein et al., 2007) to

minimize experimental variability. ISH data from control and ischemic brain sections are freely available on a public database ([www.monkey-niche.org](http://www.monkey-niche.org); **Supplementary Figure 1C**). Of note, the images on [www.monkey-niche.org](http://www.monkey-niche.org) show gene expression the SVZa but also in additional brain regions, including the corpus callosum, the cingulate gyrus and the thalamus. For identification of such structures see Saleem and Logothetis, 2007.

In the monkey, the SVZa is composed of the ependymal layer (EL) and the subependymal layer (SEL). The SEL is separated from the EL by an approximately 100- $\mu$ m-wide, cell-sparse gap zone (**Figure 1B**; Gil-Perotin et al., 2009). Underneath the striatal SEL lies the striatal parenchyma (STR). NSCs reside predominantly in the SEL on the striatal side of the lateral ventricle (Fuentelba et al., 2015). The callosal side of the monkey lateral ventricles contains a few proliferating cells under the ependyma which do not form clusters (Tonchev et al., 2005). We refer to this region as callosal SVZ (CSVZ; **Figure 1B**). In addition, there are perivascular cell clusters in the striatal SEL (PVSEL). The level of expression in the regions of interest (RoIs) EL, SEL, CSVZ, and STR was visually graded as level 0 (not detected), level 1 (low), level 2 (moderate) or level 3 (high) expression. Expression in PVSEL was assessed as “present” or absent (for illustrations of grading see **Supplementary Figure 3**).

In this report we focus on genes that belong to the SVZa-DE-UP group because this group was enriched for genes that relate to progenitor cell biology (**Figure 2A**). From the 541 SVZa-DE-UP genes, we performed ISH for 114 genes from the top 300 highly expressed genes in the transcriptome, and which relate to stem cell biology (signaling pathways, transcription factors, regulators of cell adhesion, etc.). Additionally, we included 36 genes for the ISH analysis because they are implicated in stem cell biology (**Supplementary Table S7**). Of the 114/36 SVZa-DE-UP genes the visual scoring revealed enhanced postischemic expression in at least one RoI for 102/27 genes. A heat map compiled from the data for the annotated 150 genes visualizes the marked effect of ischemia on the gene expression patterns (**Figures 1C,D**). Genes whose expression is strongly enhanced in all RoIs are found on top of the heat map (**Figure 1D**), genes induced strongly in two RoIs are found in the middle of the map. Weakly induced genes are found toward the bottom of the map. The genes in the top half of the “ischemia” (**Figure 1D**) map show strongly enhanced expression in all RoIs, including the STR, suggesting that ischemia considerably affects striatal cells as well. Most of the genes are induced in the SEL, the main stem cell-containing region. Whenever a gene is strongly expressed in the SEL, it is also strongly expressed in 2 or 3 additional RoIs (EL, CSVZ or STR).

## Enrichment of Astrocytic and Oligodendrocytic Transcription Profiles in SVZa-DE-UP Genes

The adult SVZ niche consists of several cell types (Martínez-Cerdeño and Noctor, 2018). In order to gain insight into the cell identity in our SVZa samples, we compared the transcriptome profiles of monkey total SVZa genes or SVZa-DE genes with the transcription profiles of defined cell types isolated from the adult mouse cortex (Zhang et al., 2014). We found

that SVZa-DE-UP genes were significantly enriched for genes typical for oligodendrocytic cells or astrocytes (**Figure 3A** and **Supplementary Table S8**). In contrast, SVZa-DE-DOWN genes were enriched for genes typical for endothelial cells (**Figure 3A**).

For 50 of the cell type-enriched genes, ISH data are available in the “monkey-niche” database, and their expression strength was visually scored. A comparison of the heat maps of control (**Figure 3B**) and ischemic (**Figure 3C**) SVZa shows that 34 out of these 50 genes were upregulated after ischemia in three of four RoIs. In addition, 29/50 genes showed an increased expression in perivascular cell clusters (**Figure 3C**, gray boxes; arrows in **Figures 3D–F**). Among the astrocytic genes upregulated by ischemia were *CLU* (**Figure 3D**), *TNC* (**Figure 3E**), *PHKG1* (**Figure 4G**) and *GJA1* (**Figure 4I**). Among the enhanced oligodendroglial genes were *FEZ1* (**Figure 3F**), *GPR37L1* (**Figure 4D**) and *FAM107A* (**Figure 4F**). Quantification of expression using CellDetekt software was consistent with the visual scoring shown in **Figures 3B,C**. A bar plot shows the expression value of the respective genes that are extracted from the RNA Seq data. In most cases, there is a good agreement between RPKM data, CellDetekt-analysis and visual assessment. Altogether, the transcriptomic and ISH analyses demonstrate an upregulation of astrocytic or oligodendrocytic transcripts in postischemic monkey SVZa.

## Predominance of qNSC-Related Gene Signatures in the Ischemic Primate SVZa

The progenitor pool in adult mouse SVZ represents a heterogeneous population with subtypes, including the quiescent NSCs (qNSCs) which are slowly proliferating, active NSCs (aNSCs) that further proliferate and produce multipotent transit-amplifying progenitors (TAPs). The transcription profiles of the progenitors at each of these stages have been characterized, in the mouse SVZ, using single-cell RNA Seq (Beckervordersandforth et al., 2010; Codega et al., 2014; Llorens-Bobadilla et al., 2015; Dulken et al., 2017).

To search for the presence of gene signatures typical for progenitor subtypes in the monkey SVZa-DE data sets, we compared our data (total SVZa genes or SVZa-DE genes) with the transcription profiles of mouse SVZ subtypes. A comparison with the profiles of murine qNSCs and aNSCs (Codega et al., 2014) revealed a significant over-representation of transcripts (102 genes) typical for qNSCs (**Figure 4A** and **Supplementary Table S9**). Only 39 SVZa-DE-UP genes belonged to murine aNSCs signature. Unlike the SVZa-DE-UP genes, the SVZa-DE-DOWN genes were enriched for aNSC signatures (**Figure 4A** and **Supplementary Table S9**). An enrichment of qNSC-like transcriptional profiles in monkey SVZa-DE-UP genes and of aNSC-like profiles in monkey SVZa-DE-DOWN genes was also observed when we compared the SVZa-DE transcripts with the datasets of two other studies (Beckervordersandforth et al., 2010; Dulken et al., 2017; **Figure 4A** and **Supplementary Tables S10, S11**).

Llorens-Bobadilla et al. (2015) have reported transcriptomes of murine SVZ progenitors isolated from control mice and mice subjected to global brain ischemia after 2 days. The authors

identified gene signatures for seven subclasses of SVZ cells. Class I genes are characteristic for oligodendrocytes, classes II–VI characterize sequential NSC transitory states (dormant qNSC [class II]), primed qNSC [class III]), non-mitotic aNSC [class IV], mitotic aNSC [class V], TAPs [class VI], and class VII characterize neuroblasts. The induction of global cerebral ischemia to the mouse brain upregulated classes I, II and III (Llorens-Bobadilla et al., 2015). A comparison of our data sets with their study revealed that 38 of the monkey SVZa-DE-UP genes were enriched in genes of classes of I, II and III (Figure 4A and Supplementary Table S12) corresponding to gene signatures of oligodendroglial cells and qNSCs. We also observed that 10 of the up-regulated monkey genes showed enrichment in class VII (neuroblast) genes of Llorens-Bobadilla et al., 2015. These results raise the possibility that besides the predominance of the quiescent state of the SVZa cells, a small number of SVZa precursors leave quiescence.

40 of the 116 genes present in the qNSC datasets of mouse SVZ (Codega et al., 2014; Llorens-Bobadilla et al., 2015; Dulken et al., 2017) are available in the “monkey-niche” database. Notably, 23 of these qNSC-typical genes showed ischemia-induced upregulation in three or four of the RoIs (Figures 4B,C). In addition, 28 genes showed an increased expression in perivascular cell clusters along SVZa vessels (Figure 4C, gray boxes). In Figures 4D–K we show sample ISH data for genes with a specific pattern of postischemic upregulation. *GPR37L1* and *GNAO1* are predominantly upregulated in the EL after ischemia (Figures 4D,E). The genes *FAM107A* and *PHKG1* are strongly upregulated in the SEL and to a lesser extent in the EL (Figures 4F,G). The genes *MLC1*, *GJA1*, *ATF7*, and *NACC2* (Figures 4H–K) are upregulated in all three RoIs (EL, SEL, and CSVZ). The transcriptomics and ISH results support the notion of an enhancement of qNSC-related transcripts in monkey EL and SEL.

## Enhanced Postischemic Expression of Genes Belonging to Signaling Pathways

GSEA enrichment analysis revealed that components of several signaling pathways (NOTCH, BMP, and WNT) were significantly enriched in the SVZa-DE-UP genes (Supplementary Table S13). *NOTCH1* (Figure 5A), its transcriptional effector *RBPJ* (Figure 5B) and *LFNG* (Figure 5C), a regulator of Notch signaling in NSCs (Semerci et al., 2017) are strongly induced in the EL. BMP pathway-related transcripts, *BMPRIA* and *NEO1* receptors (Figures 5D,E) as well as the *CXXC5* zinc finger protein (Figure 5F) are also upregulated in the EL. Upregulation of WNT pathway members was evident in EL, SEL and CSVZ (Figures 5G–I). In contrast to the SVZa-DE-UP genes, the SVZa-DE-DOWN genes were not significantly enriched for genes belonging to these pathways (Supplementary Table S13).

## Novel Gene Markers for the Primate SVZa

Single-cell RNA Seq experiments have provided transcriptional profiles of many SVZ cell types (Beckervordersandforth et al., 2010; Codega et al., 2014; Llorens-Bobadilla et al., 2015; Dulken

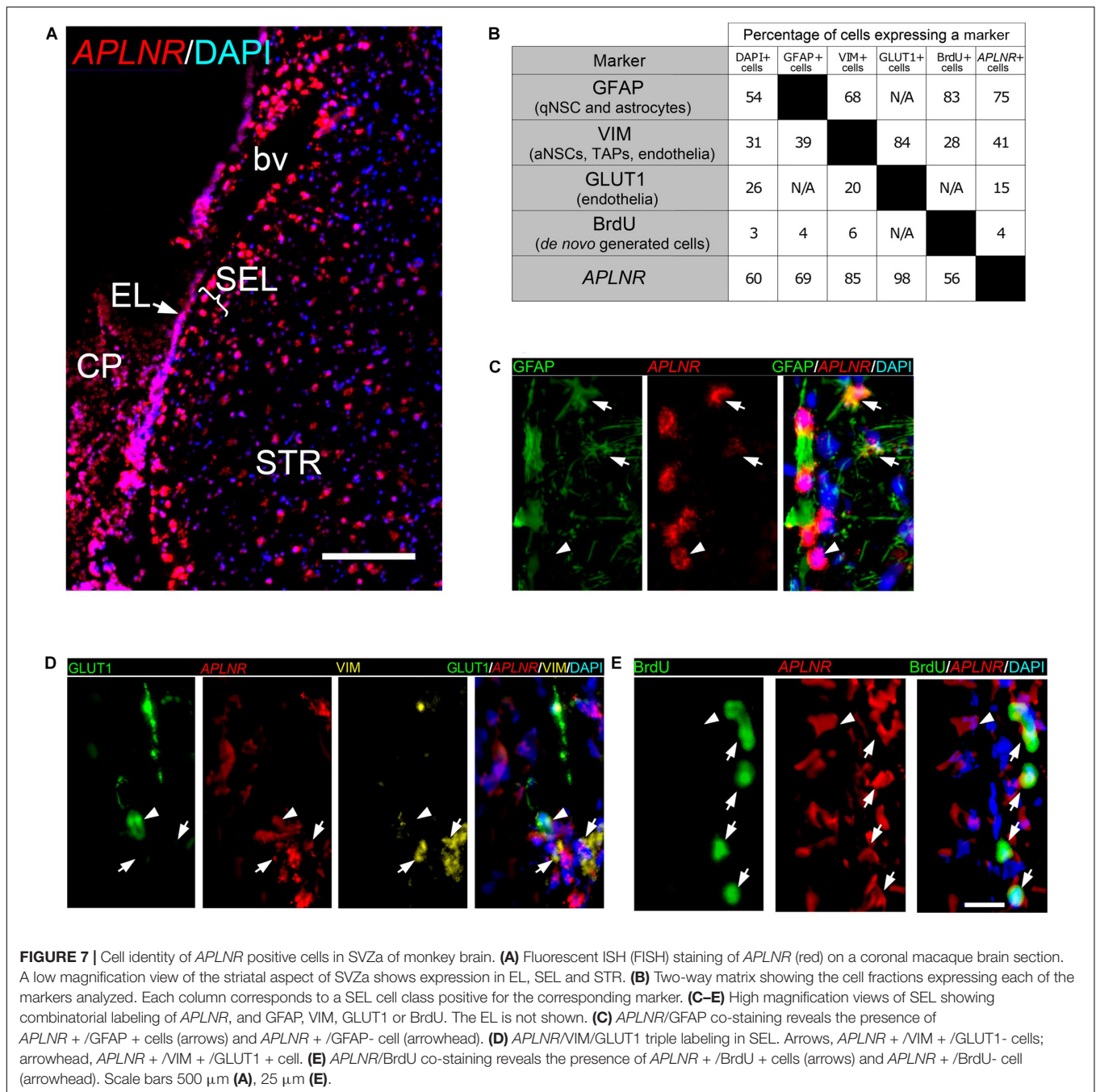
et al., 2017; Zywitza et al., 2018). We found that 38 out of the 150 genes of the “monkey-niche” database were not present in any of these profiles (Figures 6A,B). Examples include *APLNR*, *MAX*, *SCD5*, and *KIF2A* (Figures 6C–F). The apelin receptor (*APLNR*; Figure 6C) is an angiogenesis-promoting cell surface molecule (Wu et al., 2017). Its expression was markedly increased upon ischemia in the EL as well as in non-vascular cells of the SEL (Figure 6C, arrow). Similarly, *MAX*, a transcription factor which is an obligate partner of the proliferation driver *MYC*, was also markedly enhanced after ischemia, particularly in the SEL (Figure 6D). *SCD5* (Figure 6E), encoding a primate-specific isoform of Stearoyl-CoA Desaturase (Wang et al., 2005), and the microtubule-associated protein *KIF2A* (Figure 6F) were both strongly upregulated in the EL after ischemia. Other interesting examples of novel SVZa genes are accessible on monkey-niche database and include the intestinal stem cell marker *LGR5* (Barker et al., 2007) and the hematopoietic progenitor marker *ZBTB16* (Pardrige et al., 1990).

## Apelin Receptor Marks Specific Cell Populations in the Primate SVZa

The transcriptome data showed ~10-fold increase in the expression of *APLNR* (Figures 1C,D, 6C), one of the genes not previously documented in the rodent or primate SVZ. The ISH confirmed a strong induction of *APLNR* in postischemic monkey EL and SEL (Figure 6C). In the brain, *Aplnr* is expressed in vascular cells and neurons (Wu et al., 2017). Recent data indicate that *Aplnr* promotes postischemic angiogenesis and neuroprotection, at least in rodents (Wu et al., 2017). To identify which SEL cell type(s) express *APLNR* in monkey SVZa, we combined fluorescent ISH (FISH) for *APLNR* (Figure 7A) with immunofluorescence staining for markers of cells present in the niche (Figure 7B and Supplementary Table S14). One of the monkeys had been injected with the thymidine analog bromodeoxyuridine (BrdU) that selectively labels *de novo* generated cells (Tonchev et al., 2003, 2005).

To analyze the cell identity of thousands of cells in the primate SEL we developed a semi-automated approach. It involved segmentation of all DAPI stained nuclei, of cytoplasmic fluorescence signals derived from FISH or fluorescence-tagged antibodies against GFAP, VIM or GLUT1, and of nuclear BrdU. We found that 60% of all SEL cells express *APLNR* and 54% of all SEL cells express GFAP. A quarter of the SEL cells expressed VIM or GLUT1. After BrdU infusion, 3% of the SEL cells are BrdU-positive (BrdU+), confirming that only a few primate SEL cells were *de novo* generated (Figure 7B).

Examples of co-expression of *APLNR* with cell-type markers in the SEL (Figures 7C–E) and the percentages of co-expression (Figure 7B) are shown. 75% of the *APLNR*+ cells expressed GFAP (Figures 7B,C, arrows) and 69% of GFAP+ cells expressed *APLNR* (Figure 7B). These data show that *APLNR* is expressed in a significant proportion of SEL cells representing either niche astrocytes or qNSCs. *APLNR*/VIM co-labeling experiments demonstrated that 41% of the *APLNR*+ cells expressed VIM



(Figures 7B,D, arrows), and 85% of the VIM+ cells were positive for *APLNR* (Figure 7B). To distinguish between *APLNR*+ /VIM+ endothelial cells and *APLNR*+ /VIM+ neural progenitors (aNSCs or TAPs), we performed triple-labeling experiments combining *APLNR*/VIM co-staining with labeling for the endothelial marker GLUT1 (Pardrige et al., 1990; Figure 7D). We found that 98% of the GLUT1+ cells also expressed *APLNR* (Figures 7B,D, arrowhead). However, only 15% of the *APLNR*+ cells in the SEL were positive for GLUT1 (Figure 7B). Thus, we clearly detected *APLNR*+ /VIM+ cells, which were negative for GLUT1 in SEL (Figure 7D, arrows),

probably representing activated progenitors. To further support the notion that *APLNR* is expressed by proliferating cells, newly generated cells were labeled by BrdU infusion at five subsequent days. Double-staining for *APLNR* and BrdU revealed that 4% of the *APLNR*+ cells had incorporated BrdU (Figures 7B,E, arrows), and 56% of the BrdU+ cells expressed *APLNR* (Figure 7B). Notably, *APLNR* expression was evident in the SVZ niche not only upon ischemia, but also in the brain of sham-operated macaques. In contrast, in the mouse SVZ *Apnr* mRNA was not expressed in the EL and only weakly expressed in the SEL (Supplementary Figure 4B, dotted lines). Strong

*Aplnr* signal was detected in the murine neurons (**Supplementary Figure 4B**, arrowheads).

## DISCUSSION

### Status of Neurogenic Niche One Week After Ischemia

To identify the response of the SVZ neurogenic niche upon ischemic insult in the adult primate brain is a challenging task. Here, we present the first comprehensive characterization of the molecular responses of the monkey SVZ at a transcriptome level seven days after an ischemic injury. The time point of our analysis precedes the proliferative response of the SVZ niche (Tonchev et al., 2005) by a week. We provide evidence that the predominant progenitor expression profile of SVZ was typical for genes characteristic for the qNSCs state. Furthermore, we found many upregulated genes typical for astrocytes and oligodendrocytes, possibly related to the up-regulation of WNT and BMP signaling pathways (Han et al., 2014; Guo et al., 2015). Because qNSCs and astrocytes share many common gene expression profiles (Götz et al., 2015) the enrichment of astrocytic fate in the ischemic primate SVZ further support a prevalence of progenitors in a quiescent state.

Under steady-state conditions, the SVZa cells in the adult baboon monkey are enriched for the repressive chromatin marks H3K9me3 that point at a heterochromatin state (Foret et al., 2014). Ischemia in macaque brain induced expression of ~1000 genes (15% of the total genes expressed in SVZa) which is likely to be accompanied by changes in the epigenetic state of the induced genes. It will be interesting to explore how ischemia affects the presence of repressive chromatin marks in monkey SVZa and which genes have retained or lost this mark. For example, the macaque transcriptome data did not reveal statistically significant expression changes of genes critical of NSC activation, including *EGFR* (Codega et al., 2014; Dulken et al., 2017) and *INFGRI* (Llorens-Bobadilla et al., 2015). It thus appears that more than a week is necessary to transform the transcriptional landscape of the macaque SVZa from a quiescent toward an active state, while in the rodent this transformation begins to take place on the second day following global ischemia (Llorens-Bobadilla et al., 2015). Another possible reason for the prevalence of qNSC-like gene signatures is an increase of the NOTCH and BMP signaling pathways detected at postischemic day 7, particularly in the EL. These two pathways are responsible for preserving quiescence of both NSCs (Chapouton et al., 2010; Llorens-Bobadilla et al., 2015) and ependymal cells (Carlén et al., 2009; Luo et al., 2015).

The postischemic response of brain stem cells has been studied in rodents (Napoli and Borlongan, 2016; Dillen et al., 2019; Nakagomi et al., 2019). While in the adult rodent brain, the ependymal cells are quiescent and express Notch signaling components, upon stroke they become activated to produce neuroblasts and glial cells. This requires Notch down regulation in the ependyma (Carlén et al., 2009). We find, in the macaque, seven-days after ischemia, an *enhanced* ependymal expression of members of the Notch signaling pathway (*Notch1*, *RBPI*, *LFNG*). This upregulation of Notch signaling may be the reason for the

inability of primates to initiate neurogenesis. A caveat is that down-regulation of Notch signaling could occur but after the 7 day time point.

### Genes With a Novel Expression Pattern in Primate SVZa

We report the expression pattern of 38 genes previously not documented in the mammalian SVZ. An intriguing example is the enzyme Stearoyl-CoA Desaturase 5. *SCD5* is not present in rodents (Wang et al., 2005). We found a strong postischemic activation of *SCD5* in the EL. Interestingly, *SCD5* can inhibit cell proliferation by suppressing the EGFR signaling pathway (Sinner et al., 2012). This could contribute to the prevalence of qNSCs in the postischemic monkey SVZa. A strong postischemic enhancement in the EL was observed for the *Kinesin Family Member 2A (KIF2A)*, a mitotic spindle regulator (Wordeman and Mitchison, 1995) involved in human cortical development (Cavallin et al., 2017). Another interesting example was the transcription factor *Myc-Associated Factor X (MAX)*. The MAX protein forms heterodimers with several other transcription factors, most notably with c-MYC, which is critical in maintaining stemness of embryonic stem cells (Yoshida, 2018).

Two of the novel genes in the primate SVZa were shown to play a role in non-neuronal stem cell niches. These include *LGR5*, expressed in intestinal stem cells (Barker et al., 2007) and *ZBTB16*, which marks immature hematopoietic progenitors (Liu, 2007). Interestingly, we found a strong postischemic expression of both genes in EL and SEL. *LGR5* is not expressed in the mouse SVZ (Yu et al., 2017) and its presence in the primate SVZa may indicate a primate-specific expression. During embryonic chick development, *ZBTB16* maintains the self-renewal of neural progenitors (Gaber et al., 2013), and its potential involvement in adult primate neurogenesis needs to be further studied.

### APLNR Exhibits a Strong Expression in Primate SVZa

Our data show, for the first time, expression of *APLNR* in the EL and SEL of the primate SVZa. This *APLNR* transcription is markedly increased upon ischemia. Studies reporting the transcriptomes of murine SVZ cell types (Beckervordersandforth et al., 2010; Codega et al., 2014; Llorens-Bobadilla et al., 2015; Dulken et al., 2017; Zywitza et al., 2018) did not detect *Aplnr* mRNA. We show here that such expression was very weak in the mouse SEL and undetectable in the EL, while in the control primate brain, both SEL and EL showed moderate presence of *APLNR* transcripts, implicating possible contribution to brain neurogenesis. Noteworthy, genetic *APLNR* variants have been associated with the increased stroke risk in humans (Hata et al., 2007). In rodents, activation of the apelinergic system improves the recovery after experimental stroke by inhibiting neuronal apoptosis and facilitating angiogenesis (Chen et al., 2015; Wu et al., 2017). Our data showing *APLNR* transcripts in *de novo* generated SVZa progenitors suggest that *APLNR* might contribute not only to postischemic neuroprotection, but also to regeneration after brain injury. Because *APLNR* is a druggable

target, this receptor can serve as an entry point of clinically relevant molecules into a wide spectrum of primate SVZa cells.

## DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

## ETHICS STATEMENT

The animal study was reviewed and approved by Animal Care and Ethics Committee of Kanazawa University Graduate School of Medical Sciences, Japan.

## AUTHOR CONTRIBUTIONS

AS, GE, and AT designed the experiments. MC, CT, N-HH, IV, SP, and TY performed the experiments. HW, LW, and SP programed the data. MC, HW, SP, AS, GE, and AT analyzed the data. JY, GE, and AT supervised the project. AS, GE, and AT wrote the manuscript. All contributing authors commented on the manuscript and contributed to the article and approved the submitted version.

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## REFERENCES

- Alunni, A., and Bally-Cuif, L. (2016). A comparative view of regenerative neurogenesis in vertebrates. *Development* 143, 741–753. doi: 10.1242/dev.122796
- Bakken, T. E., Miller, J. A., Ding, S.-L., Sunkin, S. M., Smith, K. A., Ng, L., et al. (2016). A comprehensive transcriptional map of primate brain development. *Nature* 535, 367–375. doi: 10.1038/nature18637
- Bakken, T. E., Miller, J. A., Luo, R., Bernard, A., Bennett, J. L., Lee, C. K., et al. (2015). Spatiotemporal dynamics of the postnatal developing primate brain transcriptome. *Hum. Mol. Genet.* 24, 4327–4339. doi: 10.1093/hmg/ddv166
- Barker, N., Van Es, J. H., Kuipers, J., Kujala, P., Van Den Born, M., Cozijnsen, M., et al. (2007). Identification of stem cells in small intestine and colon by marker gene *Lgr5*. *Nature* 449, 1003–1007. doi: 10.1038/nature06196
- Beckervordersandforth, R., Tripathi, P., Ninkovic, J., Bayam, E., Lepier, A., Stempfhuber, B., et al. (2010). In vivo fate mapping and expression analysis reveals molecular hallmarks of prospectively isolated adult neural stem cells. *Cell Stem Cell* 7, 744–758. doi: 10.1016/j.stem.2010.11.017
- Bernard, A., Lubbers, L. S., Tanis, K. Q., Luo, R., Podtelezhnikov, A. A., Finney, E. M., et al. (2012). Transcriptional architecture of the primate neocortex. *Neuron* 73, 1083–1099. doi: 10.1016/j.neuron.2012.03.002
- Bond, A. M., Ming, G. L., and Song, H. (2015). Adult mammalian neural stem cells and neurogenesis: five decades later. *Cell Stem Cell* 17, 385–395. doi: 10.1016/j.stem.2015.09.003

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## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fcell.2020.584314/full#supplementary-material>

- Bray, M. A., Vokes, M. S., and Carpenter, A. E. (2015). Using Cellprofiler for automatic identification and measurement of biological objects in images. *Curr. Protoc. Mol. Biol.* 2015, 14.17.1–14.17.13. doi: 10.1002/0471142727.mb1417s109
- Buades, A., Coll, B., and Morel, J.-M. (2011). Non-local means denoising. *Image Process. Line* 1, 208–212. doi: 10.5201/ipol.2011.bcm\_nlm
- Carlén, M., Meletis, K., Göritz, C., Darsalia, V., Evergren, E., Tanigaki, K., et al. (2009). Forebrain ependymal cells are Notch-dependent and generate neuroblasts and astrocytes after stroke. *Nat. Neurosci.* 12, 259–267. doi: 10.1038/nn.2268
- Carson, J. P., Eichele, G., and Chiu, W. (2005). A method for automated detection of gene expression required for the establishment of a digital transcriptome-wide gene expression atlas. *J. Microsci.* 217, 275–281. doi: 10.1111/j.1365-2818.2005.01450.x
- Cavallin, M., Bijlsma, E. K., El Morjani, A., Moutton, S., Peeters, E. A. J., Maillard, C., et al. (2017). Recurrent *KIF2A* mutations are responsible for classic lissencephaly. *Neurogenetics* 18, 73–79. doi: 10.1007/s10048-016-0499-8
- Chapouton, P., Skupien, P., Hesel, B., Coolen, M., Moore, J. C., Madeline, R., et al. (2010). Notch activity levels control the balance between quiescence and recruitment of adult neural stem cells. *J. Neurosci.* 30, 7961–7974. doi: 10.1523/JNEUROSCI.6170-09.2010
- Chen, D., Lee, J., Gu, X., Wei, L., and Yu, S. P. (2015). Intranasal delivery of apelin-13 is neuroprotective and promotes angiogenesis after ischemic stroke in mice. *ASN Neuro.* 7:1759091415605114. doi: 10.1177/1759091415605114
- Chieco, P., Jonker, A., De Boer, B. A., Ruijter, J. M., and Van Noorden, C. J. F. (2013). Image cytometry: protocols for 2D and 3D quantification in

- microscopic images. *Prog. Histochem. Cytochem.* 47, 211–333. doi: 10.1016/j.proghi.2012.09.001
- Codega, P., Silva-Vargas, V., Paul, A., Maldonado-Soto, A. R., DeLeo, A. M., Pastrana, E., et al. (2014). Prospective identification and purification of quiescent adult neural stem cells from their in vivo niche. *Neuron* 82, 545–559. doi: 10.1016/j.neuron.2014.02.039
- Dillen, Y., Kempfs, H., Gervois, P., Wolfs, E., and Bronckaers, A. (2019). Adult neurogenesis in the subventricular zone and its regulation after ischemic stroke: implications for therapeutic approaches. *Transl. Stroke Res.* 11, 60–79. doi: 10.1007/s12975-019-00717-8
- Dulken, B. W., Leeman, D. S., Boutet, S. C., Hebestreit, K., and Brunet, A. (2017). Single-cell transcriptomic analysis defines heterogeneity and transcriptional dynamics in the adult neural stem cell lineage. *Cell Rep.* 18, 777–790. doi: 10.1016/j.celrep.2016.12.060
- Foret, M. R., Sandstrom, R. S., Rhodes, C. T., Wang, Y., Berger, M. S., and Lin, C. H. A. (2014). Molecular targets of chromatin repressive mark H3K9me3 in primate progenitor cells within adult neurogenic niches. *Front. Genet.* 5:252. doi: 10.3389/fgene.2014.00252
- Fuentealba, L. C., Rompani, S. B., Parraguez, J. I., Obernier, K., Romero, R., Cepko, C. L., et al. (2015). Embryonic origin of postnatal neural stem cells. *Cell* 161, 1644–1655. doi: 10.1016/j.cell.2015.05.041
- Gaber, Z. B., Butler, S. J., and Novitsch, B. G. (2013). PLZF regulates fibroblast growth factor responsiveness and maintenance of neural progenitors. *PLoS Biol.* 11:e1001676. doi: 10.1371/journal.pbio.1001676
- Gil-Perotin, S., Duran-Moreno, M., Belzunegui, S., Luquin, M. R., and Garcia-Verdugo, J. M. (2009). Ultrastructure of the subventricular zone in *Macaca fascicularis* and evidence of a mouse-like migratory stream. *J. Comp. Neurol.* 514, 533–554. doi: 10.1002/cne.22026
- Götz, M., Sirko, S., Beckers, J., and Irmeler, M. (2015). Reactive astrocytes as neural stem or progenitor cells: in vivo lineage, in vitro potential, and Genome-wide expression analysis. *Glia* 63, 1452–1468. doi: 10.1002/glia.22850
- Guo, F., Lang, J., Sohn, J., Hammond, E., Chang, M., and Pleasure, D. (2015). Canonical Wnt signaling in the oligodendroglial lineage-puzzles remain. *Glia* 63, 1671–1693. doi: 10.1002/glia.22813
- Han, D., Choi, M. R., Jung, K. H., Kim, N., Kim, S. K., Chai, J. C., et al. (2014). Global transcriptome profiling of genes that are differentially regulated during differentiation of mouse embryonic neural stem cells into astrocytes. *J. Mol. Neurosci.* 55, 109–125. doi: 10.1007/s12031-014-0382-8
- Hata, J., Matsuda, K., Ninomiya, T., Yonemoto, K., Matsushita, T., Ohnishi, Y., et al. (2007). Functional SNP in an Sp1-binding site of AGTRL1 gene is associated with susceptibility to brain infarction. *Hum. Mol. Genet.* 16, 630–639. doi: 10.1093/hmg/ddm005
- Hawrylycz, M. J., Levin, E. S., Guillozet-Bongaarts, A. L., Shen, E. H., Ng, L., Miller, J. A., et al. (2012). An anatomically comprehensive atlas of the adult human brain transcriptome. *Nature* 489, 391–399. doi: 10.1038/nature11405
- Jessberger, S., and Gage, F. H. (2014). Adult neurogenesis: bridging the gap between mice and humans. *Trends Cell Biol.* 24, 558–563. doi: 10.1016/j.tcb.2014.07.003
- Legland, D., Arganda-Carreras, I., and Andrey, P. (2016). MorphoLibJ: integrated library and plugins for mathematical morphology with ImageJ. *Bioinformatics* 32, 3532–3534. doi: 10.1093/bioinformatics/btw413
- Lein, E. S., Hawrylycz, M. J., Ao, N., Ayres, M., Bensinger, A., Bernard, A., et al. (2007). Genome-wide atlas of gene expression in the adult mouse brain. *Nature* 445, 168–176. doi: 10.1038/nature05453
- Li, C. H., and Tam, P. K. S. (1998). An iterative algorithm for minimum cross entropy thresholding. *Pattern Recognit. Lett.* 19, 771–776. doi: 10.1016/S0167-8655(98)00057-9
- Liu, T. M. (2007). Concise review: balancing stem cell self-renewal and differentiation with PLZF. *Access* 34, 1339–1347. doi: 10.1634/stemcells.2006-0644
- Llorens-Bobadilla, E., Zhao, S., Baser, A., Saiz-Castro, G., Zwadlo, K., and Martin-Villalba, A. (2015). Single-cell transcriptomics reveals a population of dormant neural stem cells that become activated upon brain injury. *Cell Stem Cell* 17, 329–340. doi: 10.1016/j.stem.2015.07.002
- Luo, Y., Coskun, V., Liang, A., Yu, J., Cheng, L., Ge, W., et al. (2015). Single-cell transcriptome analyses reveal signals to activate dormant neural stem cells. *Cell* 161, 1175–1186. doi: 10.1016/j.cell.2015.04.001
- Marques, B. L., Carvalho, G. A., Freitas, E. M. M., Chiareli, R. A., Barbosa, T. G., Di Araújo, A. G. P., et al. (2019). The role of neurogenesis in neurorepair after ischemic stroke. *Semin. Cell Dev. Biol.* 95, 98–110. doi: 10.1016/j.semcdb.2018.12.003
- Martinez, K., and Cupitt, J. (2005). VIPS - A highly tuned image processing software architecture. *Proc. Int. Conf. Image Process. ICIP* 2, 574–577. doi: 10.1109/ICIP.2005.1530120
- Martínez-Cerdeño, V., and Noctor, S. C. (2018). Neural progenitor cell terminology. *Front. Neuroanat.* 12:104. doi: 10.3389/fnana.2018.00104
- Miller, J. A., Ding, S. L., Sunkin, S. M., Smith, K. A., Ng, L., Szafer, A., et al. (2014). Transcriptional landscape of the prenatal human brain. *Nature* 508, 199–206. doi: 10.1038/nature13185
- Miller, J. A., Nathanson, J., Franjic, D., Shim, S., Dalley, R. A., Shapouri, S., et al. (2013). Conserved molecular signatures of neurogenesis in the hippocampal subgranular zone of rodents and primates. *Dev.* 140, 4633–4644. doi: 10.1242/dev.097212
- Nakagomi, T., Takagi, T., Beppu, M., Yoshimura, S., and Matsuyama, T. (2019). Neural regeneration by regionally induced stem cells within post-stroke brains: novel therapy perspectives for stroke patients. *World J. Stem Cells* 11, 452–463. doi: 10.4252/wjsc.v11.i8.452
- Napoli, E., and Borlongan, C. V. (2016). Recent advances in stem cell-based therapeutics for stroke. *Transl. Stroke Res.* 7, 452–457. doi: 10.1007/s12975-016-0490-6
- Obernier, K., and Alvarez-Buylla, A. (2019). Neural stem cells: origin, heterogeneity and regulation in the adult mammalian brain. *Development* 146, dev.156059. doi: 10.1242/dev.156059
- Pardrige, W. M., Boado, R. J., and Farrell, C. R. (1990). Brain-type glucose transporter the blood-brain barrier (GLUT-1) is selectively localized to the blood-brain barrier. *J. Biol. Chem.* 265, 18035–18040.
- Quinlan, A. R., and Hall, I. M. (2010). BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* 26, 841–842. doi: 10.1093/bioinformatics/btq033
- Saleem, K. S. K., and Logothetis, N. K. N. (2007). *A Combined MRI and Histology Atlas of the Rhesus Monkey Brain in Stereotaxic Coordinates*. Cambridge, MA: Academic Press.
- Sandstrom, R. S., Foret, M. R., Grow, D. A., Haugen, E., Rhodes, C. T., Cardona, A. E., et al. (2014). Epigenetic regulation by chromatin activation mark H3K4me3 in primate progenitor cells within adult neurogenic niche. *Sci. Rep.* 4, 1–9. doi: 10.1038/srep05371
- Satyanarayanan, M., Goode, A., Gilbert, B., Harkes, J., and Jukic, D. (2013). OpenSlide: a vendor-neutral software foundation for digital pathology. *J. Pathol. Inform.* 4:27. doi: 10.4103/2153-3539.119005
- Semerci, F., Tin-Shing Choi, W., Bajic, A., Thakkar, A., Encinas, J. M., Depreux, F., et al. (2017). Lunatic fringe-mediated notch signaling regulates adult hippocampal neural stem cell maintenance. *eLife* 6, 1–28. doi: 10.7554/eLife.24660
- Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., et al. (2012). Fiji: an open-source platform for biological-image analysis. *Nat. Methods* 9, 676–682. doi: 10.1038/nmeth.2019
- Shihavuddin, A., Basu, S., Rexhepaj, E., Delestro, F., Menezes, N., Sigoillot, S. M., et al. (2017). Smooth 2D manifold extraction from 3D image stack. *Nat. Commun.* 8, 1–8. doi: 10.1038/ncomms15554
- Sinner, D. I., Kim, G. J., Henderson, G. C., and Igal, R. A. (2012). StearoylCoA desaturase-5: a novel regulator of neuronal cell proliferation and differentiation. *PLoS One* 7:e039787. doi: 10.1371/journal.pone.0039787
- Soille, P. (1999). *Morphological Image Analysis*. Berlin: Springer.
- Solaini, G., Baracca, A., Lenaz, G., and Sgarbi, G. (2010). Hypoxia and mitochondrial oxidative metabolism. *Biochim. Biophys. Acta Bioenerg.* 1797, 1171–1177. doi: 10.1016/j.bbabi.2010.02.011
- Tajiri, N., Dailey, T., Metcalf, C., Mosley, Y. I., Lau, T., Staples, M., et al. (2013). In vivo animal stroke models. *Transl. Stroke Res.* 4, 308–321. doi: 10.1007/s12975-012-0241-2
- Tonchev, A. B., Yamashima, T., Sawamoto, K., and Okano, H. (2005). Enhanced proliferation of progenitor cells in the subventricular zone and limited neuronal production in the striatum and neocortex of adult macaque monkeys after global cerebral ischemia. *J. Neurosci. Res.* 81, 776–788. doi: 10.1002/jnr.20604
- Tonchev, A. B., Yamashima, T., Zhao, L., Okano, H. J., and Okano, H. (2003). Proliferation of neural and neuronal progenitors after global brain ischemia in



- young adult macaque monkeys. *Mol. Cell. Neurosci.* 23, 292–301. doi: 10.1016/S1044-7431(03)00058-7
- Trapnell, C., Pachter, L., and Salzberg, S. L. (2009). TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* 25, 1105–1111. doi: 10.1093/bioinformatics/btp120
- Wählby, C. (2003). *Algorithms for Applied Digital Image Cytometry*. Available online at: <http://www.diva-portal.org/smash/get/diva2:163446/FULLTEXT01.pdf> (accessed April 29, 2020).
- Wang, J., Yu, L., Schmidt, R. E., Su, C., Huang, X., Gould, K., et al. (2005). Characterization of HSCD5, a novel human stearyl-CoA desaturase unique to primates. *Biochem. Biophys. Res. Commun.* 332, 735–742. doi: 10.1016/j.bbrc.2005.05.013
- Wordeman, L., and Mitchison, T. J. (1995). Identification and partial characterization of mitotic centromere-associated kinesin, a kinesin-related protein that associates with centromeres during mitosis. *J. Cell Biol.* 128, 95–105.
- Wu, Y., Wang, X., Zhou, X., Cheng, B., Li, G., and Bai, B. (2017). Temporal expression of apelin/apelin receptor in ischemic stroke and its therapeutic potential. *Front. Mol. Neurosci.* 10:1. doi: 10.3389/fnmol.2017.00001
- Yamashima, T., Saido, T. C., Takita, M., Miyazawa, A., Yamano, J., Miyakawa, A., et al. (1996). Transient brain ischaemia provokes Ca<sup>2+</sup>, PIP<sub>2</sub> and calpain responses prior to delayed neuronal death in monkeys. *Eur. J. Neurosci.* 8, 1932–1944. doi: 10.1111/j.1460-9568.1996.tb01337.x
- Yoshida, G. J. (2018). Emerging roles of Myc in stem cell biology and novel tumor therapies. *J. Exp. Clin. Cancer Res.* 37, 1–20. doi: 10.1186/s13046-018-0835-y
- Yoshida, M., Yamashima, T., Zhao, L., Tsuchiya, K., Kohda, Y., Tonchev, A. B., et al. (2002). Primate neurons show different vulnerability to transient ischemia and response to cathepsin inhibition. *Acta Neuropathol.* 104, 267–272. doi: 10.1007/s00401-002-0554-4
- Yu, Y., Moberly, A. H., Bhattarai, J. P., Duan, C., Zheng, Q., Li, F., et al. (2017). The stem cell marker *Lgr5* defines a subset of postmitotic neurons in the olfactory bulb. *J. Neurosci.* 37, 9403–9414. doi: 10.1523/JNEUROSCI.0500-17.2017
- Zhang, Y., Chen, K., Sloan, S. A., Bennett, M. L., Scholze, A. R., O’Keeffe, S., et al. (2014). An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J. Neurosci.* 34, 11929–11947. doi: 10.1523/JNEUROSCI.1860-14.2014
- Zywitzka, V., Misios, A., Bunatyan, L., Willnow, T. E., and Rajewsky, N. (2018). Single-cell transcriptomics characterizes cell types in the subventricular zone and uncovers molecular defects impairing adult neurogenesis. *Cell Rep.* 25, 2457–2469.e8. doi: 10.1016/j.celrep.2018.11.003

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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