#### Check for updates

#### **OPEN ACCESS**

EDITED BY Maria Vamvakaki, University of Crete, Greece

#### REVIEWED BY Qixu Zhang, University of Texas MD Anderson Cancer Center, United States Maowen Chen, Anhui University of Chinese Medicine, China

\*CORRESPONDENCE Weiwei Liu, ⊠ liuweiw@jlu.edu.cn

<sup>†</sup>These authors have contributed equally to this work and share first authorship

RECEIVED 17 June 2023 ACCEPTED 21 July 2023 PUBLISHED 03 August 2023

#### CITATION

Hao M, Wang D, Duan M, Kan S, Li S, Wu H, Xiang J and Liu W (2023), Functional drug-delivery hydrogels for oral and maxillofacial wound healing. *Front. Bioeng. Biotechnol.* 11:1241660. doi: 10.3389/fbioe.2023.1241660

#### COPYRIGHT

© 2023 Hao, Wang, Duan, Kan, Li, Wu, Xiang and Liu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Functional drug-delivery hydrogels for oral and maxillofacial wound healing

Ming Hao<sup>1,2†</sup>, Dongxu Wang<sup>3†</sup>, Mengna Duan<sup>4†</sup>, Shaoning Kan<sup>1,2</sup>, Shuangji Li<sup>1,2</sup>, Han Wu<sup>1,2</sup>, Jingcheng Xiang<sup>1,2</sup> and Weiwei Liu<sup>1,2</sup>\*

<sup>1</sup>Department of Oral and Maxillofacial Surgery, Hospital of Stomatology, Jilin University, Changchun, China, <sup>2</sup>Jilin Provincial Key Laboratory of Tooth Development and Bone Remodeling, Hospital of Stomatology, Jilin University, Changchun, China, <sup>3</sup>Laboratory Animal Center, College of Animal Science, Jilin University, Changchun, China, <sup>4</sup>Department of Prosthodontics, Hospital of Stomatology, Jilin University, Changchun, China

The repair process for oral and maxillofacial injuries involves hemostasis, inflammation, proliferation, and remodeling. Injury repair involves a variety of cells, including platelets, immune cells, fibroblasts, and various cytokines. Rapid and adequate healing of oral and maxillofacial trauma is a major concern to patients. Functional drug-delivery hydrogels play an active role in promoting wound healing and have shown unique advantages in wound dressings. Functional hydrogels promote wound healing through their adhesive, antiinflammatory, antioxidant, antibacterial, hemostatic, angiogenic, and reepithelialization-promoting properties, effectively sealing wounds and reducing inflammation. In addition, functional hydrogels can respond to changes in temperature, light, magnetic fields, pH, and reactive oxygen species to release drugs, enabling precise treatment. Furthermore, hydrogels can deliver various cargos that promote healing, including nucleic acids, cytokines, small-molecule drugs, stem cells, exosomes, and nanomaterials. Therefore, functional drugdelivery hydrogels have a positive impact on the healing of oral and maxillofacial injuries. This review describes the oral mucosal structure and healing process and summarizes the currently available responsive hydrogels used to promote wound healing.

#### KEYWORDS

functional hydrogel, skin repair, wound healing, oral and maxillofacial, drug delivery

# **1** Introduction

The oral and maxillofacial region is involved in important physiological functions, such as respiration, mastication, articulation, esthetic maintenance, and facial expression production, and involves complex anatomical structures, including skin, mucosa, periodontal tissue, teeth, and salivary glands (Madani et al., 2014; van Veen et al., 2019; Zhai et al., 2019; Matichescu et al., 2020; Ulaganathan et al., 2020; Chung et al., 2021; Hatcher, 2022; Hong, 2023). Thus, the healing of oral and maxillofacial wounds is of great importance to maintaining the integrity of its anatomical structure and function (Ha et al., 2023). Wounds are defined as the disruption of anatomical structures accompanied by loss of continuity of function, which can be repaired by a unique, timely, and spatially coordinated wound healing process (Chhabra et al., 2017; Li et al., 2022a). Wounds that fail to heal can, even indirectly, lead to death (Zeng et al., 2018). The characteristics of non-healing wounds include long-term or excessive inflammation, persistent infection, microbial biofilm

formation, and the incapacity of dermal or epidermal cells to react to repairing stimuli (Frykberg and Banks, 2015). Non-healing wounds may lead to permanent damage to the normal function and appearance of the oral and maxillofacial region.

Hydrogels have been found to effectively improve wound healing and functional recovery (Zhang et al., 2020). Hydrogels have unique advantages in wound dressings due to their excellent biological, physicochemical, and mechanical properties (Alven and Aderibigbe, 2020), including excellent biocompatibility (Graca et al., 2020), a biological barrier effect blocking harmful factors, such as bacteria, from entering the wound (Norahan et al., 2023), and the ability to deliver cargos that promote wound healing in response to environmental changes (Dimatteo et al., 2018). It is a great concern that antibiotic overuse has led to the development of drug-resistant bacterial strains. Therefore, a variety of antimicrobial hydrogels have recently been synthesized to effectively avoid development of drugresistant bacteria (Farazin et al., 2023). Thus, hydrogels have widespread applications in the field of wound healing, and various functional hydrogels have been developed with adhesive, antioxidant, antibacterial, hemostatic, and tissue regenerative properties (Liang et al., 2021). Oral and maxillofacial wounds require hydrogels with good adhesion to provide wound protection; however, their degradation time and on-demand removal need to be considered to prevent rapid degradation exposing the wound or leading to secondary injury when removing the hydrogel (Ding et al., 2022; Lv et al., 2023). Moreover, hydrogels can respond to stimuli, such as temperature, light, magnetic fields, pH, and reactive oxygen species (ROS), to release active cargos (for example, nucleic acids, growth factors, small-molecule drugs, stem cells, exosomes, and nanomaterials) (Zhang et al., 2023).

This review summarizes the oral and maxillofacial wound healing process as well as the factors that influence healing. Functional and responsive hydrogels which are effective treatments to promote wound healing are also introduced in categories.

# 2 The process of oral and maxillofacial wound healing

### 2.1 Epithelium of skin and oral mucosa

The skin and mucosa of the oral and maxillofacial region, as barriers and protective organs of the body, are important for the maintenance of water–electrolyte balance and protection of internal tissues and organs from microbial infection and external friction, which would affect their function and esthetics (Wang et al., 2017; Park et al., 2018).

The oral mucosa and skin epithelium consist of superficial epithelium and an underlying basement membrane (Toma et al., 2021). The skin consists of a keratinized epidermal layer, dermis, and subcutaneous layer (Glim et al., 2013). The epithelium of the skin contains hair follicles and pores, which serve as a transmembrane absorption pathway providing a therapeutic modality (Glim et al., 2013; Barbero and Frasch, 2017). However, the oral mucosa consists of non-keratinized epithelium, a loose extracellular matrix (ECM), and salivary glands (Groeger and Meyle,

2019; Liu et al., 2019), and may vary according to the different environments in which the skin and mucous membranes are located. Furthermore, the oral mucosa is in a complex, warm, and moist environment with a diverse microbial presence and is subject to friction from mastication (Mark Welch et al., 2020). In contrast, the skin is exposed to changes in air, temperature, and humidity (Waasdorp et al., 2021). Moreover, studies have shown that the oral mucosa heals faster than the skin and that fewer scars are formed, likely due to the lower response to inflammation and the presence of saliva in the oral environment (Brand et al., 2014). Recent research has indicated that saliva contains various substances, such as antimicrobial peptides, histatins, and mucins, that can promote wound healing by interacting with fibroblasts, keratinocytes, and growth factors (Torres et al., 2018). In addition, oral mucosa has a rich blood supply and contains numerous capillaries, which is beneficial for wound healing (Han et al., 2018). There are significant differences in tissue structure and physiological characteristics between oral mucosa and skin that lead to variations in the healing process (Figure 1A; Table 1).

### 2.2 Oral and maxillofacial wound healing

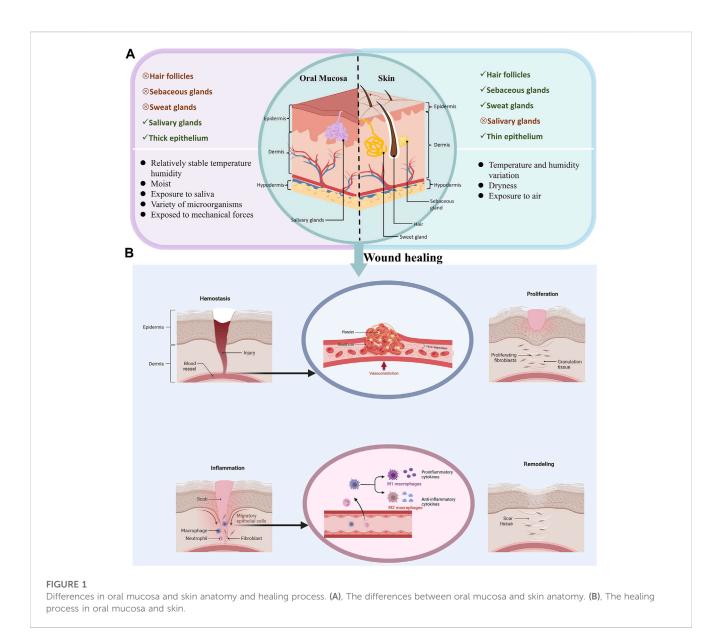
Following traumatic injuries, such as car accidents, knife wounds, cuts, and surgery, wounds heal through hemostasis, inflammation, proliferation, and remodeling, with the involvement of multiple cells and cytokines in the oral and maxillofacial region. The process of healing is described below (Figure 1B).

#### 2.2.1 Hemostasis

Upon skin injury and damage to blood vessels, bleeding occurs immediately. Hemostasis subsequently occurs with the activation of platelets, thus initiating the hemostasis cascade (Burzynski et al., 2019). Subsequently, platelets produce a number of biologically active substances, including proteases, growth factors and cytokines such as epidermal growth factor (EGF), insulin-like growth factor (IGF), platelet-derived growth factor (PDGF), and transforming growth factor- $\beta$  (TGF $\beta$ ) (Politis et al., 2016; Scopelliti et al., 2022). Moreover, blood vessels constrict and platelets aggregate to form a platelet plug (Scopelliti et al., 2022).

#### 2.2.2 Inflammation

Inflammation is caused by fibrin clot formation and aggregated platelet degranulation (Pazyar et al., 2014). In the early stages, neutrophils migrate to the site of injury and secrete enzymes, such as matrix metalloproteinases (MMPs), to remove the damaged ECM. Neutrophils then initiate cytokine and growth factor production to recruit monocytes (Velnar et al., 2009). Monocytes migrate to the wound and differentiate into macrophages (Velnar et al., 2009). Macrophages secrete cytokines, including interleukin-1 (IL-1), interleukin-6 (IL-6), fibroblast growth factor (FGF), PDGF, EGF, tumor necrosis factor (TNF), and TGF $\beta$ , which coordinate the migration of keratinocytes and fibroblasts to the wound (Sindrilaru and Scharffetter-Kochanek, 2013). Macrophages are characterized by their ability to respond to environmental changes and can differentiate into two distinct subpopulations: pro-inflammatory



M1 macrophages and anti-inflammatory M2 macrophages (Gordon and Taylor, 2005; Smigiel and Parks, 2018). M2 macrophages contribute to the upregulation of endogenous anti-inflammatory cytokines and the downregulation of previously secreted proinflammatory cytokines in the vicinity of the wound (Krzyszczyk et al., 2018). Indeed, abnormal macrophage polarization has ability to delay wound healing or lead to chronic wound formation (Louiselle et al., 2021). Nevertheless, excessive infiltration of inflammatory cells, including neutrophils and macrophages, inhibits wound healing (Eming et al., 2007).

#### 2.2.3 Proliferation

At this stage, the matrix formed by platelets is replaced by vascularized granulation tissue and re-epithelialization occurs, leading to wound closure and reestablishment of the epithelial barrier (An et al., 2021). Re-epithelialization proceeds through proliferation and migration of keratinocytes, which is influenced by nitric oxide, EGF, IGF1, keratinocyte growth factor (KGF),

and nerve growth factor (Landen et al., 2016). Reconstruction of the existing vascular network and formation of new vessels are the hallmarks of successful wound healing (Toma et al., 2021). Newly formed blood vessels provide nutrients and oxygen to the tissues (Li et al., 2007). Furthermore, studies have shown that several growth factors participate in angiogenesis, including vascular endothelial growth factor (VEGF), angiopoietin, FGF, and TGF $\beta$  (Li et al., 2003), and that the production of a temporary matrix in the granulation tissue is facilitated by the involvement of fibroblasts (Darby et al., 2014). Furthermore, fibroblasts differentiate muscle actin-expressing into a-smooth myofibroblasts, which initiate the wound closure process (Zhang et al., 2020b). Nevertheless, hyperactivation of fibroblasts and myofibroblasts is associated with abnormal scar formation (Zhang et al., 2020b). Moreover, macrophages are also involved in granulation tissue formation, secreting a variety of growth factors (FGF, TGFβ, EGF and PDGF) (Barrientos et al., 2008).

	Oral mucosa	Skin	References
Structures	Epithelial and lamina propria, part of the oral mucosa has submucosa	Epidermis, dermis and subcutaneous tissue	Glim et al. (2013)
	Keratinization/Incomplete keratinization	Keratinization	Qin et al. (2017)
	Salivary glands	Hair follicles, sebaceous glands, sweat glands	Glim et al. (2013)
	Thicker epithelium	Relatively thin epithelium	Glim et al. (2013)
	High proliferation rate of basal cells	Low proliferation rate of basal cells	Gibbs and Ponec (2000)
	Relatively high degree of vascularization	Relatively low degree of vascularization	Brand et al. (2014)
Surroundings	Epidermal moistening	Epidermal dryness	Brand et al. (2014)
	Exposure to saliva	Exposure to air	Brand et al. (2014)
	Subjected to chewing force and tension, continuously exposed to bacteria		Dutzan et al. (2017)
Healing process	The oral mucosa has less neovascularization than the skin		Szpaderska et al. (2005)
		Fibroblasts are more responsive to stimulus	Mak et al. (2009)
	Lower inflammation levels		Szpaderska et al. (2003), Chen et al. (2010)
	Saliva contains high levels of healing-promoting histone and growth factors		Brand et al. (2014)
	Oral keratinocytes have a proliferative capacity greater than that of skin keratinocytes		Turabelidze et al. (2014)
	Oral wounds exhibit rapid re-epithelialization		Schrementi et al. (2008)

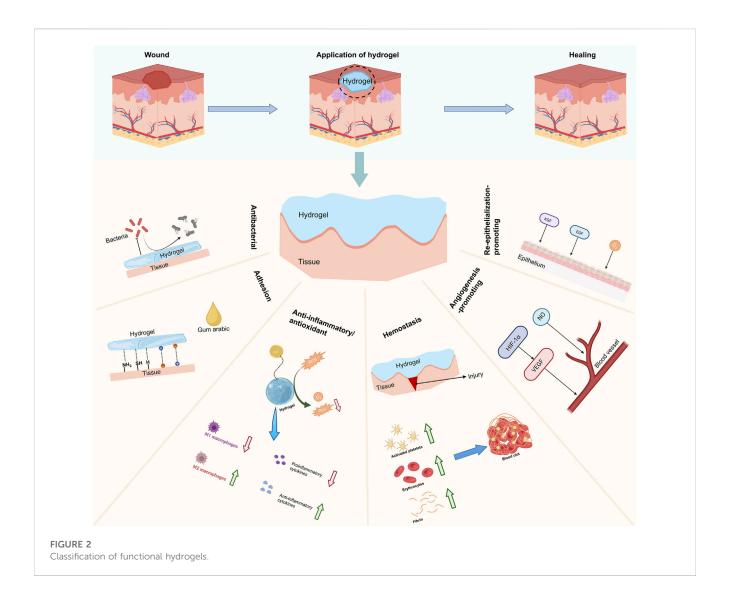
TABLE 1 The difference in structure, surroundings and healing processes between oral mucosa and skin.

#### 2.2.4 Remodeling

The final stage of healing is remodeling, which is intended to remove unnecessary vessels, fibroblasts, and inflammatory cells (Nourian Dehkordi et al., 2019). Wound closure mainly occurs through granulation and re-epithelialization. However, granulation tissue is a temporary matrix. Indeed, an important feature of wound remodeling is the change in composition of the ECM. Thus, the structure of the ECM fibrillar network undergoes organization and alignment (Li et al., 2007). Fibroblasts remodel the ECM by secreting ECMdegrading enzymes. Active proteases, especially MMPs, are involved in wound healing by controlling the synthesis of new collagen and degradation of old collagen (Visse and Nagase, 2003). The previous, temporary ECM then transitions from a loose network of fibronectin tissue to a larger, denser collagen bundle. Type III collagen is gradually transformed into type I collagen (Finnerty et al., 2016). Collagen alignment also tends to be consistent, thus increasing the strength of new tissue. Skin appendages, such as hair, sweat glands, and sebaceous glands, may regenerate (Feng et al., 2019).

# 2.3 The formation of hard-to-heal and poorly healed wounds

When a stage of wound healing is compromised, it can result in chronic wounds. The main features of chronic wounds include a prolonged inflammatory phase, excessive infiltration of neutrophils, persistent infection and high levels of MMPs. Fibroblasts in chronic wounds and ulcers are phenotypically different and have a reduced migratory capacity compared to those in acute wounds (Brumberg et al., 2021). The dysregulation of key pro-inflammatory cytokines, such as IL-1β and TNF, prolongs the inflammatory phase and delays healing. IL-1β and TNF are increased in chronic wounds, which have been shown to lead to elevated MMP levels and excessive degradation of the local ECM, impairing cellular migration (Hesketh et al., 2017; Krzyszczyk et al., 2018). It has been shown that MMPs, such as collagenase and gelatinase A and B, are elevated in the fluid of chronic wounds compared to that in acute wounds (Eming et al., 2010). Additionally, inadequate angiogenesis is considered a potential cause of chronic wounds. This is related to the deprivation of pro-angiogenic factors, such as members of the VEGF family, by proteolytic degradation, followed by subsequent interference with their bioactivity in the microenvironment of chronic wounds (Lauer et al., 2000). The formation of chronic wounds involves multiple cellular and signaling pathways. Several molecular markers downstream of the Wnt signaling pathway, such as activation of the  $\beta$ -catenin/c-myc pathway, promotes healing of damaged tissue by inhibiting the migration of keratinocytes and altering their differentiation to suppress keratinization (Stojadinovic et al., 2005; Stojadinovic et al., 2014). The EGF receptor is inhibited in keratinocytes at the non-healing edge (Brem et al., 2007). In addition, TGF<sup>β</sup> signaling is suppressed in chronic wounds through the downregulation of TGFB receptors and attenuation of Smad signaling (Pastar et al., 2010). Therefore, such cell signaling can serve as a target for rapid wound healing of chronic wounds.



# 3 Functional hydrogels for wound healing

Traditional wound dressings, including gauze and bandages, are commonly used for dry and clean wounds (Brumberg et al., 2021). Compared to conventional wound dressings, hydrogels have great advantages and application prospects due to their porous structure, biodegradability, and drug-carrying and sustained release capacity (Kamoun et al., 2017; Bai et al., 2020b). Hydrogels can be designed by modulating their properties to match the type of wound and trauma process, with the development of hydrogels with adhesive, anti-inflammatory/antioxidant, antibacterial, hemostatic, angiogenesis-promoting, and re-epithelialization-promoting properties (Figure 2).

### 3.1 Adhesive hydrogels

The oral mucosa is affected by functional movements and constant flushing when chewing food and processing saliva, making it difficult for drugs and dressings to stay in a moist environment (Wu et al., 2022a). Therefore, hydrogels that can remain adherent in wet environments have been designed inspired by the superb adhesion of mussels in humid environments. Hydrogels with better adhesion properties than commercial dressings were synthesized by photocrosslinking polyethylene glycol (PEG) monomethyl ether-modified glycidyl methacrylate-functionalized chitosan (CS), double bond-modified dopamine, and zinc ions (Yang et al., 2022). These hydrogels remained adherent to the wound for more than a few hours in a humid environment in the mouth. A light-responsive hydrogel of cyclo-nitrobenzene-modified hyaluronic acid was designed to bind firmly to intraoral wound tissue within seconds of light exposure and was adherent for more than 24 h (Zhang et al., 2021b). Given that the skin and muscles of the oral and maxillofacial region are involved in a variety of important physiological functions, the adherent hydrogel must be tough and self-healing to resist external friction (Sun et al., 2021). A super-tough hydrogel with excellent tissue affinity, adhesion, and self-healing properties was prepared by physical and chemical double cross-linking providing firm protection for wounds (Chen et al., 2018b). Moreover, a common hydrogel prepared from gum Arabic has good adhesion

to the tissue interface and does not affect the organization's movement (Singh et al., 2017). Furthermore, the excellent bonding ability of hydrogels can enhance wound contraction and closure during the early stages of wound healing (Sun et al., 2016). Thus, adhesive hydrogels can adhere to the wound and protect it, thereby promoting healing.

# 3.2 Anti-inflammatory/antioxidant hydrogels

The inflammatory response is a phase of wound healing that is susceptible to interference from internal and external oral surroundings, resulting in excessive inflammation. Moreover, prolonged inflammation leads to the accumulation of ROS in the wound and limits the antioxidant capacity of cells to clear excess ROS (Wagener et al., 2013). Therefore, anti-inflammatory hydrogels that can eliminate ROS, and thus reduce the production of proinflammatory chemokines, are useful for the regulation of wound healing. Natural polysaccharides, polyphenols, and polypeptides have excellent anti-inflammatory and antioxidant abilities (Fernandes et al., 2023). Previous studies have made use of paramylon, a natural algal polysaccharide, to prepare a hydrogel that can effectively reduce wound inflammation, clear ROS, and promote rapid healing of chronic burn wounds (Lei et al., 2022). The hydrogel was synthesized utilizing the antioxidant properties of tannins, natural polyphenols, forming Fe<sup>3+</sup>-tannin complexmodified P(AM-AA) hydrogels (Fe-TA@P(AM-AA)). The hydrogel releases tannin, which reacts with ROS, reversing excessive infiltration of neutrophils and the imbalance between M1 and M2 macrophages, thus promoting rapid wound healing (Sun et al., 2023a). Hydrogels made from pearl peptides enhance cellular viability and resistance to oxidative stress in wound healing (Liu et al., 2022). In addition to natural ingredients, artificially synthesized metallic materials also possess antioxidant abilities. By combining thiol hyaluronic acid and bioactive silver lignin nanoparticles, a hybrid nanocomposite hydrogel was prepared with excellent antioxidant activity and chronic wound healing promotion (Perez-Rafael et al., 2021). MnO2 also possesses bioenzyme-like activities. Hydrobranched poly-L-lysine-modified MnO<sub>2</sub> was cross-linked with hydrophilic poly(PEGMA-co-GMAco-AAm) polymers to produce a hydrogel in which MnO<sub>2</sub> showed peroxidase-like activity to decrease ROS levels and catalyze O2 production. This hydrogel significantly enhanced the polarization of M2 macrophages (Tu et al., 2022). Thus, hydrogels can achieve anti-inflammatory and antioxidant effects through multiple pathways.

# 3.3 Antibacterial hydrogels

The oral cavity contains a diverse range of microorganisms, including many bacteria and fungi, making oral cavity wounds easily susceptible to bacterial infections. The development of suitable biological dressings to maintain a non-sterile wound environment is of great importance for wound healing. Antibacterial hydrogels can be obtained by careful design of the components. PEG diglycidyl ether and  $\epsilon$ -polylysine were used to

synthesize antimicrobial hydrogels, in which  $\varepsilon$ -polylysine is a cationic peptide that kills bacteria and PEG diglycidyl ether prevents bacteria from adhering to the hydrogel, thus achieving an antibacterial and sterile wound environment (Yuan et al., 2021). Compared to traditional dressings, hydrogels also have better therapeutic effects on wounds that are prone to bacterial infections (Shi et al., 2020). Near-infrared (NIR)-responsive wound dressings have been developed, which can generate heat under NIR (808 nm) irradiation to kill bacteria and accelerate deep burn wound healing (Wang et al., 2022c).

The clinical application of antibiotics often leads to the development of drug resistance in bacteria. Hydrogels can easily address this; for example, hydrogels synthesized from bovine serum albumin can effectively kill methicillin-resistant *Staphylococcus aureus* (Ouyang et al., 2022). A hydrogel with excellent antimicrobial properties against extensively drug-resistant bacteria was synthesized from CS and  $\beta$ -glycerophosphate, promoting full wound healing of extensively drug-resistant infections (Aliakbar Ahovan et al., 2020). Therefore, through the design of hydrogel components, it is possible to gain antibacterial effects against a variety of bacteria and antibiotic-resistant strains.

# 3.4 Hemostatic hydrogels

The oral and maxillofacial areas are rich in blood vessels and highly susceptible to bleeding after trauma. Therefore, timely hemostasis is essential to prevent shock and death by bleeding. Hydrogels can promote hemostasis through physical closure and enrichment of coagulation factors (Ghobril and Grinstaff, 2015). Hydrogels for rapid hemostasis have been prepared using catecholfunctionalized quaternized CS (CQCS) and dibenzaldehyde-capped PEG (DB-PEG), which has good adhesion to the wet tissue-blood interface and can cope well with bleeding by physical closure in a variety of situations (Fang et al., 2023). In addition, the hemostatic hydrogel can stop bleeding by activating platelets. Platelet activation and blood cell aggregation can be induced through the electrostatic interaction, thereby promoting hemostasis (Fan et al., 2016). Cationic CS hydrogels can interact with negatively charged blood cells to promote coagulation (Xu et al., 2013) and hemostasis can be achieved through Ca2+ activation of thrombin, significantly improving the hemostatic properties (Weng et al., 2022). Thus, hydrogels provide excellent hemostasis through physical sealing or through the activation of platelets and thrombin.

# 3.5 Angiogenesis-promoting hydrogels

Oral and maxillofacial healing requires blood vessels to deliver nutrients and oxygen, and impaired angiogenesis can lead to difficult-to-heal wounds. Therefore, hydrogels have been developed to have intrinsic properties promoting or stimulating angiogenesis through the loading of pro-angiogenic substances. Bioactive glass nanoparticles containing copper (BGNC) and sodium alginate (ALG) hydrogels synthesized by PEG diacrylate, bioactive glass nanoparticles containing copper, and sodium alginate were able to significantly promote angiogenesis by enhancing HIF1 $\alpha$ and VEGF expression (Li et al., 2020). Amino acids are the basic

units of proteins and have a regulatory effect on the proliferation of vascular endothelial cells during wound healing. Amino acids have been added into CS-collagen hydrogels with excellent proangiogenic effects (Aleem et al., 2019). These hydrogels can also have excellent angiogenesis-promoting ability by loading smallmolecule drugs or bioactive components that promote angiogenesis. Desferrioxamine (DFO), an FDA-approved proangiogenic small molecule, and hydroxy saffron yellow A were loaded into a gelatin-CS hydrogel, which upregulated HIF1a expression and promoted angiogenesis, further promoting rapid healing of diabetic wounds (Gao et al., 2018). Aptamer-Fibrinogen hydrogels with a high affinity and specificity for target molecules have been synthesized to control and regulate VEGF expression and promote angiogenesis (Zhao et al., 2019). Other bioactive components, such as platelet-rich plasma and exosomes, have also been used for the synthesis of pro-angiogenic hydrogels (Shi et al., 2017; Qian et al., 2020). In addition to delivering proangiogenic molecules, vascular networks can be physically engineered to induce blood perfusion from the host tissue to the implant and to promote monocyte function with pro-angiogenic properties (Lee et al., 2020). Thus, hydrogels can be designed to significantly promote angiogenesis in oral and maxillofacial wounds.

# 3.6 Re-epithelialization-promoting hydrogels

Re-epithelialization is an important evaluation index of oral and maxillofacial wound healing and is of great significance for wound closure. Keratinocytes initiate the process of epithelialization, with KGF being an important factor (Ni et al., 2007). However, KGF cannot act directly at the wound site due to its instability. Thus, hydrogel transport of KGF can significantly improve its stability and promote wound re-epithelialization (Wathoni et al., 2019). Epiregulin, a member of the EGF family, is more effective than KGF in promoting wound re-epithelialization. Epiregulincontaining hydrogels can positively promote wound healing by inducing HaCaT cell proliferation and stratification (Reno et al., 2012). In addition to delivering growth factors, the hydrogel delivers oxygen to relieve the hypoxic environment and promote reepithelialization. Hydrogels containing oxygen-releasing microspheres can promote re-epithelialization of diabetic wounds through continuous oxygen delivery to the wound site (Guan et al., 2021). Therefore, the promotion of re-epithelialization in oral and maxillofacial wounds can be achieved through the use of hydrogels carrying cargos such as KGF, EGF and oxygen.

## 4 Functional drug-loaded hydrogel biodressings promote drug release in response to stimulation

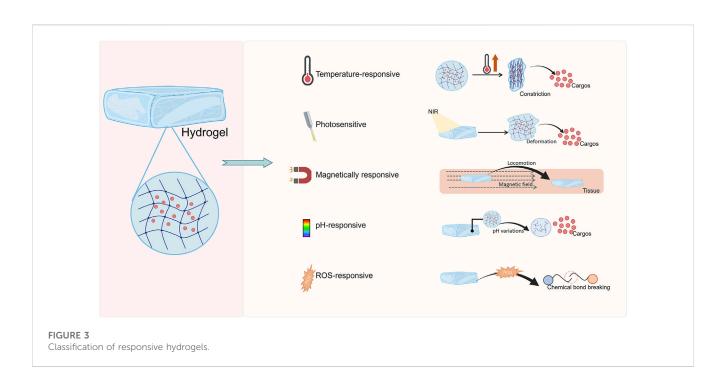
The wound healing process involves a variety of cells and growth factors, and conventional dressings cannot regulate the release of active pharmaceutical ingredients in response to the healing process. Smart wound dressings can selectively release active substances according to changes in temperature, light, magnetic fields, pH, enzyme levels, and ROS at the wound site. Furthermore, the synthetic components of hydrogels and the physical characteristics of their 3D pore-like structure allow good drugloading capacity. Compared to traditional wound dressings, drugloaded hydrogels can release active substances by responding to the local environment to regulate the different stages of wound healing (Figure 3).

#### 4.1 Temperature-responsive hydrogels

Compared with skin, the internal temperature of the oral cavity is relatively high and stable; thus, temperature-sensitive hydrogels can be utilized to achieve responsive release of cargos with temperature variation. Inspired by hot spring shower, hydrogels were prepared from N,O-carboxymethyl CS and favalite, which can stimulate angiogenesis and promote wound healing by releasing bioactive ions under heat stimulation (Sheng et al., 2021). In addition, thermally responsive hydrogels have been designed to release VEGF in response to body temperature stimulation to promote the healing of diabetic wounds (Chen et al., 2022a). The inflammatory response leads to an increased temperature of wound tissue, and the temperature-responsive properties of the hydrogel allow for the intelligent release of FGF to reduce the inflammatory response (Chen et al., 2018a). Hydrogels have been designed that can change from the gel state to liquid state based on temperature sensitivity, which reduces secondary damage that may be caused during dressing changes (Shi et al., 2022). Therefore, temperaturesensitive hydrogels can achieve drug release according to temperature changes inside the mouth and promote angiogenesis and anti-inflammation, avoiding secondary damage during dressing change.

### 4.2 Photosensitive hydrogels

Some hydrogels can be laser irradiated to disrupt the original structure of the hydrogel and facilitate drug release. Smart hydrogels can respond to light stimulation for drug delivery. Metal ions have photothermal conversion ability, and the release of drugs can be achieved by synthesizing metal ion-loaded hydrogels. Au nanorods functionalized with active peptides have been made into bilayer structured hydrogels to achieve NIR-responsive drug release under NIR irradiation (Huang et al., 2023b). Additionally, given the excellent antibacterial properties of Cu, a multifunctional composite hydrogel was prepared using polydopamine and Cudoped calcium silicate ceramics. Cu ions are heated by the photothermal effect and produce a unique 'hot ion effect', providing antibacterial properties, angiogenesis, and collagen deposition during the healing of infected wounds (Xu et al., 2020). In addition to metal nanoparticles, polydopamine has been used as a photothermal agent for the synthesis of photosensitive hydrogels due to its excellent light absorption in the NIR region; in this way, hydrogels have been developed to release Se nanoparticles (Xu et al., 2023). Therefore, photo-responsive hydrogels can be prepared by designing the components of hydrogels, achieving the responsive release of drugs with antibacterial function to promote wound healing in oral and maxillofacial areas.



### 4.3 Magnetically responsive hydrogels

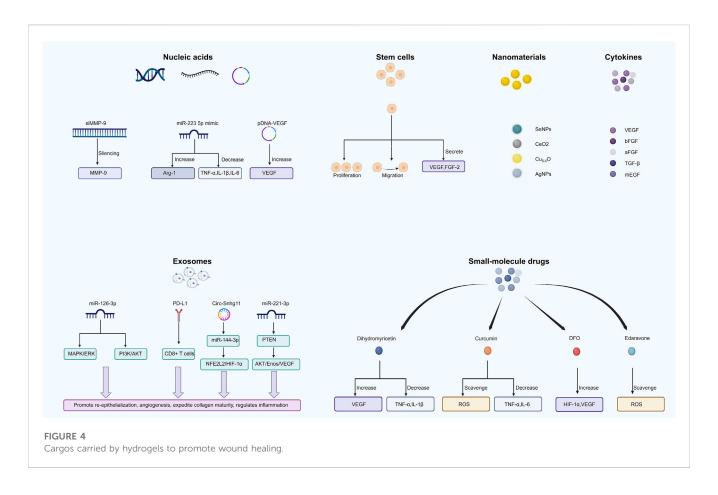
Homogeneous magnetically responsive nanocomposite hydrogels with enhanced mechanical properties have been obtained by tannin-assisted bridge building between magnetically deformable cobalt ferrite nanoparticles and a polyvinyl alcohol (PVA) matrix. In the presence of an external static magnetic field, the composite hydrogels produced mechanical stimulation-induced early blood vessel formation and accelerated wound healing (Wang et al., 2022b). Additionally, by doping the hydrogel sheet with magnetic nanoparticles, endothelial progenitor cells can be promoted to colonize the wound, enhancing the healing effect of hydrogels (Noh et al., 2019). Curcumin has also been used in the synthesis of magnetic hydrogels, and released curcumin is able to promote tissue repair and regeneration by promoting angiogenesis (Daya et al., 2022). Therefore, angiogenesis can be promoted in oral and maxillofacial wound healing by preparing hydrogels responsive to magnetic fields.

# 4.4 pH-responsive hydrogels

Studies have shown that, when the skin barrier is disrupted, it leads to changes in wound pH, which has implications for the design of pH-responsive wound dressings. A hydrogel synthesized by crosslinking a Schiff base bonds and PF127 micelles is protonated in an acidic environment, leading to breakage of the Schiff base and resulting in the release of curcumin (Qu et al., 2018). Magnesium acetate was also loaded for the synthesis of pH-responsive hydrogels with sustained antibacterial and antioxidant activity (Wang et al., 2022a). In addition to traditional Chinese medicines, studies have shown that exosomes have a positive effect on promoting wound healing. There is a need to develop hydrogel materials that can release exosomes on demand and have exosome compatibility. Hydrogels carrying adipose mesenchymal stem cell exosomes were successfully synthesized by the Schiff base reaction, and the Schiff base bond was broken in the acidic environment of diabetic wounds to release exosomes (Wang et al., 2019a). Therefore, according to the characteristics of pH change in the wound healing process, pH-responsive hydrogels can be designed to achieve cargo release and promote antibacterial, antioxidant, and angiogenesis functions.

# 4.5 ROS-responsive hydrogels

ROS levels in chronic wound sites are high, and ROSresponsive hydrogels can be designed according to these characteristics to achieve intelligent release of drugs and thus improve the therapeutic effect. High glucose in diabetic wounds leads to increased levels of ROS, which increases the duration of the inflammatory response (Brizeno et al., 2016). Synthetic polymers, through the modification of molecular components, can confer specific efficacy to hydrogels. By mixing tannic acid, phenylboronic acid-modified polyphosphonitrile, and PVA, the ROS-responsive anti-inflammatory TA-conjugated nanoparticle hydrogels effectively scavenged ROS, reduced the expression of pro-inflammatory factors, and promoted diabetic wound healing (Ni et al., 2022). PVA-based hydrogels cross-linked with a ROSresponsive linker have also been obtained by modifying the composition of the hydrogels, which gradually degrade due to ROS-responsive cleavage to release loaded mupirocin and the growth factor GM-CSF, which can inhibit bacterial growth (Zhao et al., 2020). ROS-responsive hydrogels can be prepared by designing the components of hydrogels to achieve antiinflammatory/antioxidant effects on oral and maxillofacial wounds.



## 4.6 Multiple responsiveness hydrogels

During injury to the oral and maxillofacial region, factors such as pH, enzymes levels, and ROS are altered. Reversible covalent bonds in hydrogels are an important feature in designing responsive hydrogels, and many covalent bonds can respond to pH, ROS levels, and temperature. Based on the characteristics of low pH and increased ROS in diabetic wounds, a pH/ROS dual-responsive hydrogel has been designed that releases diclofenac sodium and caffeic acid to inhibit inflammation and promote angiogenesis, achieving rapid healing of diabetic wounds in their acidic oxidative environment (Wu et al., 2022b). In addition, hyaluronic acid has the ability to respond to a variety of external stimuli and has been used to design a pH-responsive (weakly alkaline environment) and hyaluronidase-responsive Zr4+-releasing hydrogel that can inhibit bacterial growth and biofilm formation, accelerating wound healing (Zhou et al., 2023a). Keratin is an excellent hydrogel candidate given its multi-stimulus responsiveness to pH and glutathione and enzyme levels (Peralta Ramos et al., 2017; Qiu et al., 2020; Wang et al., 2020). Dual network hydrogels exhibit responsiveness to temperature, pH, and ROS levels as well as excellent antibacterial and antioxidant properties. In particular, these hydrogels respond to both acidic and alkaline environments, and can be used in the treatment of both acute and chronic wounds (Han et al., 2021). The synthetic design of peptide structures imparts excellent thermo-responsive, ultrasoundresponsive, and enzymatic-responsive properties to release the anti-inflammatory drug naproxen, which regulates the inflammatory response (Fan et al., 2017). To reduce postoperative complications, including recurrence of residual tumor cells and bacterial infection, a pH, temperature and NIR multiresponsive hydrogel loaded with doxorubicin and indocyanine green has been prepared to kill residual tumor cells on the wound surface and deep in the skin, eliminate harmful bacteria and bacterial biofilms, and promote rapid healing of post-operative tumor wounds (Zhao et al., 2021). In addition, hydrogels can be designed for the treatment of postoperative wounds of oral and maxillofacial tumors (Chen et al., 2022b). Therefore, hydrogels can be designed to be multi-responsive and effective in promoting antibacterial, antioxidant, and angiogenesis properties for a variety of wound types.

# 5 Healing-promoting cargos carried by hydrogels

Hydrogels play an important role in promoting wound healing, and their porous structure and biocompatibility allow them to carry a variety of biologically active cargos such as cells and exosomes (Figure 4; Table 2).

## 5.1 Nucleic acids

The process of wound healing is regulated by a variety of nucleic acids, and therefore the regulation of wound healing can be achieved

#### TABLE 2 Cargos carried by hydrogels to promote wound healing.

Cargos		Function	Wound healing properties of hydrogels	Wound	References
Nucleic acids	MMP-9-targeting siRNA (siMMP-9)	MMP-9 sliencing, reduce the expression of MMP-9.	Increase the collagen content in the granulation tissue and promote the maturation of the granulation tissue.	Diabetic wound	Lan et al. (2021)
	miR-223-5p mimic	Increase expression of Arg-1 and decrease TNF, IL-1 $\beta$ , and IL-6.	Promote the polarization of M2 type macrophages, reduce inflammatory response, and promotes the formation of vascularized skin tissue.	Wound	Saleh et al. (2019)
	pDNA-VEGF	Promote VEGF expression.	Inhibit inflammation and promote angiogenesis.	Burn wound	Wang et al. (2019a)
	PDRN	Improve angiogenesis, promote cellular activity, increase collagen synthesis, and exert anti-inflammatory effects.	Promote angiogenesis, cell proliferation, and collagen synthesis.	Wound	Jing et al. (2021)
	PDRN	Promote tissue regeneration and anti- inflammation.	Promote proliferation and migration of epidermal fibroblasts and increase collagen synthesis.	Wound	Sun et al. (2023a)
	miRNA-146a	Reduce expression of pro-inflammatory genes and promote Col1a2 gene expression.	Reduce inflammation and improve wound healing.	Diabetic wound	Sener et al. (2020)
Cytokines	VEGF	Promote angiogenesis.	Enhanced anti-inflammatory response, angiogenesis and cell proliferation at the wound site to promote rapid wound healing.	Wound	Siebert et al. (2021)
	aFGF, bFGF	Promote cell proliferation, granulation formation, re-epithelization, and angiogenesis.	Promote granulation formation, re- epithelization, and angiogenesis.	Wound	Wu et al. (2016)
	TGFβ-inhibitor	Regulate cell proliferation, migration, ECM production and immune response.	Enhance wound closure and enhance scarless wound repair.	Wound	Zhang et al. (2021a)
	rhEGF	Promote cell proliferation.	Promotes fibroblast proliferation and accelerates diabetic wound closure.	Diabetic wound	Hajimiri et al. (2016)
Small-molecule drugs	Curcumin	Scavenge excess ROS and reduce the expression of pro-inflammatory cytokines (IL-6 and TNF-α).	Reduce the inflammatory response and promote angiogenesis.	Burn wound	Yang et al. (2021)
	Dihydromyricetin	Antioxidant, antibacterial, anti- inflammatory.	Promote the expression of skin-related growth factors, angiogenesis and collagen production and inhibit inflammation.	Wound	Zhao et al. (2022)
	Edaravone	Scavenge excess ROS.	Reduce oxidative stress at the wound site and promote diabetic wound healing.	Diabetic wound	Fan et al. (2019)
	DFO	Regulate HIF-1a.	Promote angiogenesis and reduce inflammation in diabetic wounds, and antibacterial.	Diabetic ulcers	Li et al. (2022a)
	Resveratrol	Anti-inflammatory, antioxidant, antibacterial and anti-aging.	Anti-inflammatory, antioxidant, antibacterial.	Wound	Jia et al. (2022)
	Baicalin	Antioxidant and scavenge excess ROS.	Regulate inflammatory response and promote wound healing.	Wound	Manconi et al. (2018)
Stem cells	Adipose stem cells	Secrete multiple pro-angiogenic factors.	Promote angiogenesis.	Wound	Eke et al. (2017)
	Human adipose stem cells	Promote cell proliferation and regulate inflammatory response.	Promote re-epithelialization, nerve fiber growth and modulate the inflammatory response.	Diabetic ulcers	da Silva et al. (2017)
	Bone marrow mesenchymal stem cells	Promote growth factor secretion and regulate macrophage polarization.	Inhibit inflammation and promote angiogenesis.	Diabetic ulcers	Bai et al. (2020)
	Mesenchymal Stem Cell Spheroids	Angiogenesis, anti-inflammatory, paracrine effects.	Accelerate epithelial remodeling and angiogenesis.	Diabetic wound	Yang et al. (2020b)
Exosomes	Mesenchymal stem cell- derived exosomes	Vascular endothelial growth factor (VEGF) and factor transforming growth factor beta- 1 (TGF $\beta$ -1) expression are upregulated.	Promote angiogenesis.	Diabetic wound	Yang et al. (2020a)

(Continued on following page)

Cargos		Function	Wound healing properties of hydrogels	Wound	References
	hypoxia-pretreated ADSC-Exo	Circ-Snhg11 activates miR-144-3p/ NFE2L2/HIF1α signaling.	Promote angiogenesis.	Diabetic wound	Hu et al. (2023)
	ATV pretreated BMSC derived exosomes	MiR-221-3p activates the AKT/eNOS pathway and promotes endothelial cell proliferation, migration, tube formation and VEGF levels.	Promote angiogenesis.	Diabetic wound	Yu et al. (2020)
	Exosomes with high concentration of PD-L1	Inhibit T-cell activation and promote migration of epidermal cells and dermal fibroblasts.	Regulate immune response, accelerate wound contraction and re-epithelialization.	Wound	Su et al. (2019)
Nanomaterials	Selenium nanoparticles	Antioxidant and anti-inflammatory.	Reduce inflammation and promote wound closure, granulation tissue formation, collagen deposition, angiogenesis, fibroblast activation and differentiation.	Wound	Mao et al. (2021)
	Cerium oxide nanoparticles	Antibacterial, antioxidant, anti- inflammatory and anti-cancer.	Induce collagen deposition and accelerate wound closure.	Wound	Ahmed et al. (2021)
	nanoenzyme-Cu <sub>5.4</sub> O	Scavenge ROS.	Reduce inflammation, increase epidermal regeneration, angiogenesis.	Wound	Peng et al. (2021)
	Silver nanoparticles	Antibacterial and antioxidant.	Promotes diabetic wound healing through antibacterial, accelerated collagen deposition and re-epithelialization.	Diabetic wound	Masood et al. (2019)

TABLE 2 (Continued) Cargos carried by hydrogels to promote wound healing.

through the application of nucleic acid-based drugs. MMPs are involved in tissue remodeling; however, increased levels of MMP9 expression in chronic non-healing wounds lead to local ECM breakdown and delay the wound healing process. Small interfering RNAs can effectively silence gene expression and are highly promising for wound healing applications. Sustained local delivery of MMP9-targeting small interfering RNA (siMMP9) has been achieved through hydrogel loading, which significantly improved diabetic wound closure by silencing MMP9 (Lan et al., 2021). MicroRNAs are capable of targeting multiple genes in specific biological processes to produce lasting effects on disease treatment at the molecular level. Hydrogels have been designed and synthesized loaded with a miR-223-5p mimic, which promotes the polarization of macrophages to M2-type macrophages in wound tissue and upregulates the expression of the anti-inflammatory gene Arg-1 and downregulates the expression of TNF, IL-1β, and IL-6 (Saleh et al., 2019). DNA plasmids have low transfection efficiency, while their transport with hydrogels can improve their delivery efficiency. VEGF gene transfection was promoted by delivering plasmid DNA encoded with VEGF through hydrogels, inhibiting the inflammatory response and promoting microvessel formation (Wang et al., 2019b). In addition, polydeoxyribonucleotides (PDRN) also have a healing-promoting effect, and wound healing has been shown to be significantly accelerated by the synthesis of polysaccharide-based hydrogels containing PDRNcarriers that release the gene carrier PCNP for effective uptake by skin fibroblasts (Jing et al., 2021). Therefore, the hydrogel can be used to carry nucleic acids and improve their stability and utilization, with application in the treatment of oral and maxillofacial wound healing through regulation of the inflammatory response and promotion of angiogenesis.

## 5.2 Cytokines

Cytokines at the wound site play an important regulatory role in the healing process; therefore, the topical delivery of cytokines, such as VEGF, basic FGF (bFGF), TGFβ, and rhEGF, through hydrogels has a significant effect on wound healing (Wu et al., 2016). A 3Dprinted hydrogel patch coated with VEGF and modified by tetrapodal zinc oxide particles has been developed to achieve controlled release of VEGF and regulate angiogenesis during wound healing (Siebert et al., 2021). bFGF is involved in the physiological process of wound healing and has strong biological activity. A hydrogel system with continuous delivery of bFGF was prepared with a significant effect on wound healing in aged rats (Xiao et al., 2022). A microencapsulated hydrogel system that releases TGF $\beta$  inhibitor has been prepared to achieve temporal specific management of skin wounds and to promote scar-free healing (Zhang et al., 2021a). A study has also modified the growth factor and prepared carboxymethyl CS recombinant human epidermal growth factor coupling (NaCMCh-rhEGF), which can protect rhEGF from degradation by protease and improve the application of the drug (Hajimiri et al., 2016). The cytokine-loaded hydrogel can be used for oral and maxillofacial wound healing by reducing the inflammatory response and promoting angiogenesis.

### 5.3 Small-molecule drugs

A variety of small-molecule drugs have been developed for antiinflammatory, antioxidant, and angiogenesis purposes. Drugdelivery hydrogels can improve the utilization rate, stability, and sustained release of drugs to improve their therapeutic effect, whilst reducing their toxic side effects. By loading curcumin into a hydrogel, an improved capacity to reduce ROS and regulate the inflammatory response can be achieved (Yang et al., 2021). polyhydroxyflavonoid Dihydromyricetin, a natural with antioxidant, antibacterial, and anti-inflammatory properties, has been shown to promote skin repair when loaded in a temperature-sensitive hydrogel by promoting the expression of skin-related growth factors and inhibiting the expression of cellular inflammatory factors (Zhao et al., 2022). Diabetic wounds are difficult to heal due to impaired angiogenesis, and the increased inflammatory response caused by excessive ROS in wounds is a major factor affecting healing. Edaravone can effectively remove ROS but its restricted stability limits its application. Loading edaravone onto hydrogels can improve its stability and drug utilization and promote the rapid healing of diabetic wounds through slow release (Fan et al., 2019). DFO is an FDA-approved small-molecule drug that accelerates the healing of diabetic ulcers by regulating VEGF and HIF1a in a long-lasting and stable manner through hydrogel-carrying DFO (Li et al., 2022b). Resveratrol and baicalin have also been used in drug-loaded hydrogels to promote wound healing (Manconi et al., 2018; Jia et al., 2022). Thus, hydrogels can carry various small-molecule drugs to achieve oral and maxillofacial wound healing by promoting antibacterial, antiinflammatory/antioxidant, and angiogenesis properties.

### 5.4 Stem cells

Stem cells can produce and secrete growth factors that stimulate angiogenesis and re-epithelialization (Kim et al., 2009). Moreover, hydrogels can provide an excellent storage environment for stem cells. Studies have shown that the angiogenic effect of hydrogels is increased threefold after the addition of adipose stem cells, which can significantly promote wound healing (Eke et al., 2017; Shiekh et al., 2020). Human umbilical cord mesenchymal stem cells encapsulated in hydrogels can promote angiogenesis and collagen deposition by inhibiting the secretion of inflammatory factors TNF and IL-1β, with accelerated healing effects on diabetic wounds (Xu et al., 2022). Embryonic stem cells, induced pluripotent stem cells, and mesenchymal stem cells have shown good efficacy in promoting wound healing in hydrogel-stem cell therapy by inhibiting inflammation and promoting angiogenesis (Li et al., 2022a; Huang et al., 2022). Thus, hydrogels loaded with stem cells may promote oral and maxillofacial wound healing by inhibiting inflammation and promoting angiogenesis.

### 5.5 Exosomes

Exosomes are small vesicles secreted by cells that carry a variety of nucleic acids and proteins. The addition of exosomes to hydrogels has shown an excellent effect on the healing of diabetic wounds, a reduction in the formation of scar tissue, and promotion of the production of skin appendages, with a positive effect on skin regeneration (Wang et al., 2019a). Exosomes, as bioactive substances, have modifiable properties. By preparing exosomes derived from miR-126-3p overexpressed synovial mesenchymal stem cells, the exosomes obtain re-epithelialization, angiogenesis, and collagen maturation effects, promoting diabetic wound healing (Li et al., 2016). Moreover, as a product of cells, exosomes retain the ability to regulate wound healing and immune tolerance and are more readily taken up by recipient cells; therefore, exosome-loaded hydrogels have the potential to promote wound healing (Safari et al., 2022). Exosomes can be engineered and loaded to promote oral and maxillofacial wound healing by regulating the wound inflammatory microenvironment, promoting angiogenesis, and enhancing reepithelialization (Su et al., 2019; Yu et al., 2020; Hu et al., 2023).

# 5.6 Nanomaterials

Metal nanomaterials with antioxidant, antibacterial, and woundhealing properties have also been used in hydrogels (Hu et al., 2017). Metal nanoenzymes have a large number of catalytically active surface atoms and are therefore highly active and more stable than natural enzymes (Yadi et al., 2018). For example, selenium nanoparticles have significant antibacterial, anti-inflammatory, and antioxidant effects (Mao et al., 2021). The addition of cerium oxide nanoparticles to a CS hydrogel provides antioxidant and antibacterial properties, significantly improving the wound healing process (Ahmed et al., 2021). In addition, a composite hydrogel loaded with ultra-small nanoenzyme-Cu<sub>5.4</sub>O can chelate inflammatory chemokines and inhibit inflammatory cell activity, with the continuous release of Cu<sub>5.4</sub>O promoting angiogenesis (Peng et al., 2021). Most metal nanoparticles have antibacterial effects, especially silver nanoparticles; therefore, the addition of silver nanoparticles into hydrogels and their slow release lead to a continuous antibacterial effect (Masood et al., 2019). The healing of oral and maxillofacial wounds can thus be promoted by hydrogels loaded with nanomaterials through antibacterial, anti-inflammatory, and antioxidant effects.

# 6 Discussion and conclusion

The skin and mucous membranes of the oral and maxillofacial region are located in exposed areas of the body and are susceptible to interference from internal and external environmental factors, resulting in destruction of the epidermal barrier and the formation of wounds. In addition, there are structural biological differences between the oral mucosa and skin (Nikoloudaki et al., 2020). Both oral mucosa and skin are composed of an epithelial layer and connective tissue but part of the oral mucosal epithelium does not contain a keratinized layer (Qin et al., 2017). Moreover, the oral mucosa is in a more humid, friction-prone, and bacteria-rich environment than the skin and has no skin appendages (Glim et al., 2013). Compared to skin, oral mucosa requires dressings and medications with good adhesion in wet environments as well as antimicrobial properties, friction resistance, and anti-inflammatory properties (Table 3). Thus, considering the differences between skin and oral mucosa, functional and suitable hydrogels should provide varying effects of hemostasis, inflammation, proliferation, and remodeling for wound healing.

Traditional dressings are not able to fully meet the needs of oral and maxillofacial wound treatment. Thus, addressing wound healing in the oral and maxillofacial region is urgently required. Hydrogels have shown promising application in wound healing of the oral and maxillofacial region (Zhou et al., 2023b). Hydrogels can accelerate the healing of many types of wounds, including chronic

#### TABLE 3 Characteristics and effects of hydrogels in oral wounds.

Oral wound	The characteristics of hydrogels	Effects of application of hydrogels	References
Humid and highly dynamic oral wound environments.	Photocrosslinking, adhesion and degradability.	The hydrogel adheres to the oral mucosa, protects mucosal wounds from liquid flushing, oral movements and friction, and promotes healing of oral mucosal injuries in rats and pigs.	Zhang et al. (2021b)
Tooth-Extraction Wound Healing.	High strength, quick gel formation, injectable, anti- inflammatory.	The hydrogel is able to rapidly form a gel within seconds, has excellent antimicrobial properties, and is able to reduce the inflammatory infiltration of rat tooth extraction wounds.	Wang et al. (2022)
Moist and dynamic oral wound environment.	Protects the wound and isolates it from the external environment, with good resistance to dissolution, adhesion and adherence.	The hydrogel cross-links <i>in situ</i> to protect the wound, reduce acute inflammation in canine extraction wounds, and promote rapid healing of intraoral wounds.	Wu et al. (2022b)
Moist and dynamic oral wound environment.	Favorable adhesion properties, anti-oxidation.	The hydrogel was able to adhere to moist oral mucosal wounds and possessed excellent antioxidant ability, which could effectively shorten the inflammation period and promote the healing of oral wounds in diabetic rats.	Sun et al. (2023)
Dark, persistent bacterial irritation of oral mucosal wounds	Antibacterial.	The hydrogel possesses favorable antimicrobial ability and can promote the rapid healing of oral mucosal wounds in rats.	Qi et al. (2023)
Moist and dynamic oral wound environment.	Hemostatic and antimicrobial.	The hydrogel reduces inflammatory reactions, bleeding, and antimicrobial activity, thereby promoting rapid wound healing in patients with oral mucosal injuries.	de Jesus et al. (2023)
Salivary gland excision wound.	Promotes salivary gland regeneration.	The hydrogel sheet was able to promote the regeneration of salivary glands at the wound of submandibular gland excision in rats.	Miyake et al. (2020)
Periodontal damage.	Antibacterial, osteogenic.	The hydrogel resists <i>Porphyromonas gingivalis</i> , promotes osteogenesis and accelerates periodontal wound healing.	Zang et al. (2019)
Bacterial irritation.	Antibacterial, protects wounds.	The hydrogel is antimicrobial but does not affect the oral commensal microbiota and accelerates healing of infected extraction wounds.	Chen et al. (2023)
Direct exposure to the external environment.	Regulating macrophages.	The hydrogel modulates macrophage differentiation, inhibits transitional inflammatory responses, and promotes rapid repair of oral mucosal damage.	Wen et al. (2023)

difficult-to-heal wounds such as diabetic wounds, inflammation, and burns, through good adhesion, anti-inflammatory/antioxidant effects, antibacterial effects, hemostasis, promotion of angiogenesis, and re-epithelialization (Li et al., 2022a). The oral mucosa is moist and the environment with saliva and food flushing requires the hydrogel to have excellent tissue adhesion ability and be able to maintain a long adhesion time on tissues. In addition, the oral mucosa is in direct contact with the external environment and is prone to inflammation and oxidation, and prolonged inflammation that is difficult to suppress may lead to tissue necrosis and reduced function of the oral and maxillofacial area (Senel, 2021; Ngeow et al., 2022). The anti-inflammatory and antioxidant properties of hydrogels help to alleviate oxidative stress and reduce the inflammatory response (Luneva et al., 2022). Bacteria in the oral cavity can become pathogenic when the body is compromised or immunocompromised, causing slow wound healing. Hydrogels can prevent bacterial colonization and inhibit bacterial growth. Therefore, hydrogels should have adhesive, anti-inflammatory/ antioxidant, and antibacterial properties. In the complex oral environment, hydrogels adhere to moist wounds and protect them from the external environment without destroying oral microflora (Zhang et al., 2021b; Chen et al., 2023). For periodontal, extraction and salivary gland tissues, hydrogels

promote periodontal repair, bone regeneration, and salivary gland regeneration as well as resisting bacterial infection, which has a positive effect on the restoration of oral tissue structure and function (Zang et al., 2019; Miyake et al., 2020).

Further, functional hydrogels have the ability to respond to magnetic fields, light, temperature, and pH changes in wound healing. For example, given that oral and maxillofacial wounds are located on the surface of the body, an applied magnetic field and light can directly modulate hydrogel release without additional damage to the tissue. The temperature inside the oral cavity is relatively constant at ~37°C, such that the temperature-sensitive hydrogel can be designed to apply to the oral cavity (Sund-Levander et al., 2002). In addition, the temperature of the wound increases during inflammation, and the drug can be released specifically in response to this change (Zhou et al., 2019). Thus, responsive hydrogels can be designed to respond to the particular internal environmental changes of wounds, such as pH, enzymes, peroxide environment, high ROS levels, and high glucose levels, thus achieving on-demand release of the drug-loaded hydrogel. The healing process of oral and maxillofacial trauma is complex. Most smart hydrogels only regulate one of these stages, and hydrogels should be designed that can track and regulate all of the different healing stages (Huang et al., 2023).

Changes in temperature, light, magnetic field, pH, and ROS in functional hydrogels can also be exploited to achieve effective cargo release where required. Indeed, the effects of hemostasis, inflammation, proliferation, and remodeling of oral and maxillofacial wound healing depend on the delivery of cargos such as RNAs, growth factors, drugs, stem cells, and exosomes (McCarthy et al., 2021; Teixeira et al., 2021; Qi et al., 2023). Previous reports indicated that miR-31, miR-21, miR-23b, miR-200 and VEGF, EGF, HGF growth factors promote cell proliferation and influence inflammation in both oral mucosa and skin (Nagy et al., 2001; Chen et al., 2019; Simoes et al., 2019). Furthermore, evidence has suggested that functional stem cell-loaded and exosome-loaded hydrogels significantly promote wound healing through MAPK, Wnt, HIF-1, IGF1R/AKT/mTOR, and TGFβ signaling pathways in oral and skin wounds (Prime et al., 2004; Eslami et al., 2009; Zhang et al., 2009; Steenhuis et al., 2011; Calenic et al., 2015; Houschyar et al., 2020; Zhou et al., 2022).

Oral and maxillofacial wound healing undergoes hemostasis, inflammation, proliferation, and remodeling and can be improved by functional and responsive hydrogels that can be applied to oral wounds and promote wound healing.

## Author contributions

MH and DW wrote the manuscript. MH, DW, MD, SK, SL, HW, JX, and WL searched PubMed and Web of Science for citations and prepared figures. All authors contributed to the article and approved the submitted version.

# References

Ahmed, H. E., Iqbal, Y., Aziz, M. H., Atif, M., Batool, Z., Hanif, A., et al. (2021). Green synthesis of CeO(2) nanoparticles from the abelmoschus esculentus extract: Evaluation of antioxidant, anticancer, antibacterial, and wound-healing activities. *Molecules* 26 (15), 4659. doi:10.3390/molecules26154659

Aleem, A. R., Shahzadi, L., Tehseen, S., Alvi, F., Chaudhry, A. A., Rehman, I. U., et al. (2019). Amino acids loaded chitosan/collagen based new membranes stimulate angiogenesis in chorioallantoic membrane assay. *Int. J. Biol. Macromol.* 140, 401–406. doi:10.1016/j.ijbiomac.2019.08.095

Aliakbar Ahovan, Z., Khosravimelal, S., Eftekhari, B. S., Mehrabi, S., Hashemi, A., Eftekhari, S., et al. (2020). Thermo-responsive chitosan hydrogel for healing of full-thickness wounds infected with XDR bacteria isolated from burn patients: *In vitro* and *in vivo* animal model. *Int. J. Biol. Macromol.* 164, 4475–4486. doi:10.1016/j.ijbiomac. 2020.08.239

Alven, S., and Aderibigbe, B. A. (2020). Chitosan and cellulose-based hydrogels for wound management. Int. J. Mol. Sci. 21 (24), 9656. doi:10.3390/ijms21249656

An, Y., Lin, S., Tan, X., Zhu, S., Nie, F., Zhen, Y., et al. (2021). Exosomes from adiposederived stem cells and application to skin wound healing. *Cell. Prolif.* 54 (3), e12993. doi:10.1111/cpr.12993

Bai, H., Kyu-Cheol, N., Wang, Z., Cui, Y., Liu, H., Liu, H., et al. (2020a). Regulation of inflammatory microenvironment using a self-healing hydrogel loaded with BM-MSCs for advanced wound healing in rat diabetic foot ulcers. *J. Tissue Eng.* 11, 204173142094724. doi:10.1177/2041731420947242

Bai, Q., Han, K., Dong, K., Zheng, C., Zhang, Y., Long, Q., et al. (2020b). <p&gt; Potential applications of nanomaterials and Technology for diabetic wound healing</ p>. Int. J. Nanomedicine 15, 9717–9743. doi:10.2147/IJN.S276001

Barbero, A. M., and Frasch, H. F. (2017). Effect of stratum corneum heterogeneity, anisotropy, asymmetry and follicular pathway on transdermal penetration. *J. Control Release* 260, 234–246. doi:10.1016/j.jconrel.2017.05.034

Barrientos, S., Stojadinovic, O., Golinko, M. S., Brem, H., and Tomic-Canic, M. (2008). Perspective article: Growth factors and cytokines in wound healing: Growth factors and cytokines in wound healing. *Wound Repair Regen.* 16 (5), 585–601. doi:10. 1111/j.1524-475X.2008.00410.x

## Funding

This work was supported by the Jilin Province Department of Finance (Grant No. jcsz2021893-13), the Jilin Scientific and Technological Development Program (Grant Nos. YDZJ202301ZYTS166 and 20200801077GH), Jilin University Baikouen Program Project (Grant No. 2023B26), Jilin Provincial Education Department Science and Technology Research Project (Grant No. JJKH20221093KJ), Jilin University Cross disciplinary Cultivation Program for Young Faculty and Students (Grant No. 2022-JCXK-10), Jilin Province Development and Reform Commission (Grant No. 2023C028-6), and the Changchun Scientific and Technological Development Program (Grant No. 21ZY26).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Brand, H. S., Ligtenberg, A. J., and Veerman, E. C. (2014). Saliva and wound healing. *Monogr. Oral Sci.* 24, 52–60. doi:10.1159/000358784

Brem, H., Stojadinovic, O., Diegelmann, R. F., Entero, H., Lee, B., Pastar, I., et al. (2007). Molecular markers in patients with chronic wounds to guide surgical debridement. *Mol. Med.* 13 (1-2), 30–39. doi:10.2119/2006-00054. Brem

Brizeno, L. A., Assreuy, A. M., Alves, A. P., Sousa, F. B., de, B. S. P. G., de Sousa, S. C., et al. (2016). Delayed healing of oral mucosa in a diabetic rat model: Implication of TNF-α, IL-1β and FGF-2. *Life Sci.* 155, 36–47. doi:10.1016/j.lfs.2016.04.033

Brumberg, V., Astrelina, T., Malivanova, T., and Samoilov, A. (2021). Modern wound dressings: Hydrogel dressings. *Biomedicines* 9 (9), 1235. doi:10.3390/biomedicines9091235

Burzynski, L. C., Humphry, M., Pyrillou, K., Wiggins, K. A., Chan, J. N. E., Figg, N., et al. (2019). The coagulation and immune systems are directly linked through the activation of interleukin-1a by thrombin. *Immunity* 50 (4), 1033–1042.e6. doi:10.1016/j. immuni.2019.03.003

Calenic, B., Greabu, M., Caruntu, C., Tanase, C., and Battino, M. (2015). Oral keratinocyte stem/progenitor cells: Specific markers, molecular signaling pathways and potential uses. *Periodontol* 69(1), 68–82. doi:10.1111/prd.12097

Chen, C., Liu, Y., Wang, H., Chen, G., Wu, X., Ren, J., et al. (2018a). Multifunctional chitosan inverse opal particles for wound healing. *ACS Nano* 12 (10), 10493–10500. doi:10.1021/acsnano.8b06237

Chen, C., Wang, Y., Zhang, H., Zhang, H., Dong, W., Sun, W., et al. (2022a). Responsive and self-healing structural color supramolecular hydrogel patch for diabetic wound treatment. *Bioact. Mater* 15, 194–202. doi:10.1016/j.bioactmat. 2021.11.037

Chen, L., Arbieva, Z. H., Guo, S., Marucha, P. T., Mustoe, T. A., and DiPietro, L. A. (2010). Positional differences in the wound transcriptome of skin and oral mucosa. *BMC Genomics* 11, 471. doi:10.1186/1471-2164-11-471

Chen, L., Peng, M., Zhou, J., Hu, X., Piao, Y., Li, H., et al. (2023). Supramolecular photothermal cascade nano-reactor enables photothermal effect, cascade reaction, and

in situ hydrogelation for biofilm-associated tooth-extraction wound healing. Adv. Mater, e2301664. doi:10.1002/adma.202301664

Chen, L., Simoes, A., Chen, Z., Zhao, Y., Wu, X., Dai, Y., et al. (2019). Overexpression of the oral mucosa-specific microRNA-31 promotes skin wound closure. *Int. J. Mol. Sci.* 20 (15), 3679. doi:10.3390/ijms20153679

Chen, T., Chen, Y., Rehman, H. U., Chen, Z., Yang, Z., Wang, M., et al. (2018b). Ultratough, self-healing, and tissue-adhesive hydrogel for wound dressing. ACS Appl. Mater Interfaces 10 (39), 33523–33531. doi:10.1021/acsami.8b10064

Chen, X., Chen, D. R., Liu, H., Yang, L., Zhang, Y., Bu, L. L., et al. (2022b). Local delivery of gambogic acid to improve anti-tumor immunity against oral squamous cell carcinoma. *J. Control Release* 351, 381–393. doi:10.1016/j.jconrel.2022.09.010

Chhabra, S., Chhabra, N., Kaur, A., and Gupta, N. (2017). Wound healing concepts in clinical practice of OMFS. *J. Maxillofac. Oral Surg.* 16 (4), 403–423. doi:10.1007/s12663-016-0880-z

Chung, M. K., Wang, S., Oh, S. L., and Kim, Y. S. (2021). Acute and chronic pain from facial skin and oral mucosa: Unique neurobiology and challenging treatment. *Int. J. Mol. Sci.* 22 (11), 5810. doi:10.3390/ijms22115810

da Silva, L. P., Santos, T. C., Rodrigues, D. B., Pirraco, R. P., Cerqueira, M. T., Reis, R. L., et al. (2017). Stem cell-containing hyaluronic acid-based spongy hydrogels for integrated diabetic wound healing. *J. Investig. Dermatol* 137 (7), 1541–1551. doi:10.1016/j.jid.2017.02.976

Darby, I. A., Laverdet, B., Bonte, F., and Desmouliere, A. (2014). Fibroblasts and myofibroblasts in wound healing. *Clin. Cosmet. Investig. Dermatol* 7, 301–311. doi:10. 2147/CCID.S50046

Daya, R., Xu, C., Nguyen, N. T., and Liu, H. H. (2022). Angiogenic hyaluronic acid hydrogels with curcumin-coated magnetic nanoparticles for tissue repair. ACS Appl. Mater Interfaces 14 (9), 11051–11067. doi:10.1021/acsami.1c19889

de Jesus, G., Marques, L., Vale, N., and Mendes, R. A. (2023). The effects of chitosan on the healing process of oral mucosa: An observational cohort feasibility split-mouth study. *Nanomater. (Basel)* 13 (4), 706. doi:10.3390/nano13040706

Dimatteo, R., Darling, N. J., and Segura, T. (2018). *In situ* forming injectable hydrogels for drug delivery and wound repair. *Adv. Drug Deliv. Rev.* 127, 167–184. doi:10.1016/j. addr.2018.03.007

Ding, T., Qi, J., Zou, J., Dan, H., Zhao, H., and Chen, Q. (2022). A multifunctional supramolecular hydrogel for infected wound healing. *Biomater. Sci.* 10 (2), 381–395. doi:10.1039/d1bm01575c

Dutzan, N., Abusleme, L., Bridgeman, H., Greenwell-Wild, T., Zangerle-Murray, T., Fife, M. E., et al. (2017). On-going mechanical damage from mastication drives homeostatic Th17 cell responses at the oral barrier. *Immunity* 46 (1), 133–147. doi:10.1016/j.immuni.2016.12.010

Eke, G., Mangir, N., Hasirci, N., MacNeil, S., and Hasirci, V. (2017). Development of a UV crosslinked biodegradable hydrogel containing adipose derived stem cells to promote vascularization for skin wounds and tissue engineering. *Biomaterials* 129, 188–198. doi:10.1016/j.biomaterials.2017.03.021

Eming, S. A., Koch, M., Krieger, A., Brachvogel, B., Kreft, S., Bruckner-Tuderman, L., et al. (2010). Differential proteomic analysis distinguishes tissue repair biomarker signatures in wound exudates obtained from normal healing and chronic wounds. J. Proteome Res. 9 (9), 4758–4766. doi:10.1021/pr100456d

Eming, S. A., Krieg, T., and Davidson, J. M. (2007). Inflammation in wound repair: Molecular and cellular mechanisms. *J. Investig. Dermatol* 127 (3), 514–525. doi:10.1038/ sj.jid.5700701

Eslami, A., Gallant-Behm, C. L., Hart, D. A., Wiebe, C., Honardoust, D., Gardner, H., et al. (2009). Expression of integrin  $\alpha\nu\beta6$  and TGF- $\beta$  in scarless vs scar-forming wound healing. *J. Histochem Cytochem* 57 (6), 543–557. doi:10.1369/jbc.2009.952572

Fan, J., Li, R., Wang, H., He, X., Nguyen, T. P., Letteri, R. A., et al. (2017). Multiresponsive polypeptide hydrogels derived from N-carboxyanhydride terpolymerizations for delivery of nonsteroidal anti-inflammatory drugs. *Org. Biomol. Chem.* 15 (24), 5145–5154. doi:10.1039/c7ob00931c

Fan, L., Yang, H., Yang, J., Peng, M., and Hu, J. (2016). Preparation and characterization of chitosan/gelatin/PVA hydrogel for wound dressings. *Carbohydr. Polym.* 146, 427–434. doi:10.1016/j.carbpol.2016.03.002

Fan, Y., Wu, W., Lei, Y., Gaucher, C., Pei, S., Zhang, J., et al. (2019). Edaravone-loaded alginate-based nanocomposite hydrogel accelerated chronic wound healing in diabetic mice. *Mar. Drugs* 17 (5), 285. doi:10.3390/md17050285

Fang, W., Yang, L., Chen, Y., and Hu, Q. (2023). Bioinspired multifunctional injectable hydrogel for hemostasis and infected wound management. *Acta Biomater*. 161, 50–66. doi:10.1016/j.actbio.2023.01.021

Farazin, A., Shirazi, F. A., and Shafiei, M. (2023). Natural biomarocmolecule-based antimicrobial hydrogel for rapid wound healing: A review. *Int. J. Biol. Macromol.* 244, 125454. doi:10.1016/j.ijbiomac.2023.125454

Feng, C. J., Lin, C. H., Tsai, C. H., Yang, I. C., and Ma, H. (2019). Adipose-derived stem cells-induced burn wound healing and regeneration of skin appendages in a novel skin island rat model. *J. Chin. Med. Assoc.* 82 (8), 635–642. doi:10.1097/JCMA. 00000000000134

Fernandes, A., Rodrigues, P. M., Pintado, M., and Tavaria, F. K. (2023). A systematic review of natural products for skin applications: Targeting inflammation, wound healing, and photo-aging. *Phytomedicine* 115, 154824. doi:10.1016/j.phymed.2023. 154824

Finnerty, C. C., Jeschke, M. G., Branski, L. K., Barret, J. P., Dziewulski, P., and Herndon, D. N. (2016). Hypertrophic scarring: The greatest unmet challenge after burn injury. *Lancet* 388 (10052), 1427–1436. doi:10.1016/S0140-6736(16)31406-4

Frykberg, R. G., and Banks, J. (2015). Challenges in the treatment of chronic wounds. *Adv. Wound Care (New Rochelle)* 4 (9), 560–582. doi:10.1089/wound.2015.0635

Gao, S. Q., Chang, C., Li, J. J., Li, Y., Niu, X. Q., Zhang, D. P., et al. (2018). Co-delivery of deferoxamine and hydroxysafflor yellow A to accelerate diabetic wound healing via enhanced angiogenesis. *Drug Deliv.* 25 (1), 1779–1789. doi:10.1080/10717544.2018. 1513608

Ghobril, C., and Grinstaff, M. W. (2015). The chemistry and engineering of polymeric hydrogel adhesives for wound closure: A tutorial. *Chem. Soc. Rev.* 44 (7), 1820–1835. doi:10.1039/c4cs00332b

Gibbs, S., and Ponec, M. (2000). Intrinsic regulation of differentiation markers in human epidermis, hard palate and buccal mucosa. *Arch. Oral Biol.* 45 (2), 149–158. doi:10.1016/s0003-9969(99)00116-8

Glim, J. E., van Egmond, M., Niessen, F. B., Everts, V., and Beelen, R. H. (2013). Detrimental dermal wound healing: What can we learn from the oral mucosa? *Wound Repair Regen.* 21 (5), 648–660. doi:10.1111/wrr.12072

Gordon, S., and Taylor, P. R. (2005). Monocyte and macrophage heterogeneity. Nat. Rev. Immunol. 5 (12), 953–964. doi:10.1038/nri1733

Graca, M. F. P., Miguel, S. P., Cabral, C. S. D., and Correia, I. J. (2020). Hyaluronic acid-based wound dressings: A review. *Carbohydr. Polym.* 241, 116364. doi:10.1016/j. carbpol.2020.116364

Groeger, S., and Meyle, J. (2019). Oral mucosal epithelial cells. Front. Immunol. 10, 208. doi:10.3389/fimmu.2019.00208

Guan, Y., Niu, H., Liu, Z., Dang, Y., Shen, J., Zayed, M., et al. (2021). Sustained oxygenation accelerates diabetic wound healing by promoting epithelialization and angiogenesis and decreasing inflammation. *Sci. Adv.* 7 (35), eabj0153. doi:10.1126/ sciadv.abj0153

Ha, P., Liu, T. P., Li, C., and Zheng, Z. (2023). Novel strategies for orofacial soft tissue regeneration. *Adv. Wound Care (New Rochelle)* 12 (6), 339–360. doi:10.1089/wound. 2022.0037

Hajimiri, M., Shahverdi, S., Esfandiari, M. A., Larijani, B., Atyabi, F., Rajabiani, A., et al. (2016). Preparation of hydrogel embedded polymer-growth factor conjugated nanoparticles as a diabetic wound dressing. *Drug Dev. Ind. Pharm.* 42 (5), 707–719. doi:10.3109/03639045.2015.1075030

Han, X., Yang, R., Wan, X., Dou, J., Yuan, J., Chi, B., et al. (2021). Antioxidant and multi-sensitive PNIPAAm/keratin double network gels for self-stripping wound dressing application. *J. Mater Chem. B* 9 (31), 6212–6225. doi:10.1039/d1tb00702e

Han, Y., Xu, S., Jin, J., Wang, X., Liu, X., Hua, H., et al. (2018). Primary clinical evaluation of photodynamic therapy with oral leukoplakia in Chinese patients. *Front. Physiol.* 9, 1911. doi:10.3389/fphys.2018.01911

Hatcher, D. C. (2022). Anatomy of the mandible, temporomandibular joint, and dentition. *Neuroimaging Clin. N. Am.* 32 (4), 749–761. doi:10.1016/j.nic.2022.07.009

Hesketh, M., Sahin, K. B., West, Z. E., and Murray, R. Z. (2017). Macrophage phenotypes regulate scar formation and chronic wound healing. *Int. J. Mol. Sci.* 18 (7), 1545. doi:10.3390/ijms18071545

Hong, S. O. (2023). Cosmetic treatment using botulinum toxin in the oral and maxillofacial area: A narrative review of esthetic techniques. *Toxins (Basel)* 15 (2), 82. doi:10.3390/toxins15020082

Houschyar, K. S., Tapking, C., Puladi, B., Popp, D., Duscher, D., Rein, S., et al. (2020). Wnt signaling in cutaneous wound healing. *Handchir Mikrochir Plast. Chir.* 52 (2), 151–158. doi:10.1055/a-1017-3600

Hu, M., Korschelt, K., Daniel, P., Landfester, K., Tremel, W., and Bannwarth, M. B. (2017). Fibrous nanozyme dressings with catalase-like activity for H(2)O(2) reduction to promote wound healing. *ACS Appl. Mater Interfaces* 9 (43), 38024–38031. doi:10. 1021/acsami.7b12212

Hu, N., Cai, Z., Jiang, X., Wang, C., Tang, T., Xu, T., et al. (2023). Hypoxia-pretreated ADSC-derived exosome-embedded hydrogels promote angiogenesis and accelerate diabetic wound healing. *Acta Biomater.* 157, 175–186. doi:10.1016/j.actbio.2022.11.057

Huang, C., Yuan, W., Chen, J., Wu, L. P., and You, T. (2023a). Construction of smart biomaterials for promoting diabetic wound healing. *Molecules* 28 (3), 1110. doi:10.3390/molecules28031110

Huang, J. N., Cao, H., Liang, K. Y., Cui, L. P., and Li, Y. (2022). Combination therapy of hydrogel and stem cells for diabetic wound healing. *World J. Diabetes* 13 (11), 949–961. doi:10.4239/wjd.v13.i11.949

Huang, X., Xu, L., Yu, X., Li, Y., Huang, Z., Xu, R., et al. (2023b). Near-infrared lightresponsive multifunctional hydrogel releasing peptide-functionalized gold nanorods sequentially for diabetic wound healing. *J. Colloid Interface Sci.* 639, 369–384. doi:10. 1016/j.jcis.2023.02.078 Jia, Y., Shao, J. H., Zhang, K. W., Zou, M. L., Teng, Y. Y., Tian, F., et al. (2022). Emerging effects of resveratrol on wound healing: A comprehensive review. *Molecules* 27 (19), 6736. doi:10.3390/molecules27196736

Jing, X., Sun, Y., Liu, Y., Ma, X., and Hu, H. (2021). Alginate/chitosan-based hydrogel loaded with gene vectors to deliver polydeoxyribonucleotide for effective wound healing. *Biomater. Sci.* 9 (16), 5533–5541. doi:10.1039/d1bm00911g

Kamoun, E. A., Kenawy, E. S., and Chen, X. (2017). A review on polymeric hydrogel membranes for wound dressing applications: PVA-based hydrogel dressings. J. Adv. Res. 8 (3), 217–233. doi:10.1016/j.jare.2017.01.005

Kim, W. S., Park, B. S., and Sung, J. H. (2009). The wound-healing and antioxidant effects of adipose-derived stem cells. *Expert Opin. Biol. Ther.* 9 (7), 879–887. doi:10. 1517/14712590903039684

Krzyszczyk, P., Schloss, R., Palmer, A., and Berthiaume, F. (2018). The role of macrophages in acute and chronic wound healing and interventions to promote prowound healing phenotypes. *Front. Physiol.* 9, 419. doi:10.3389/fphys.2018.00419

Lan, B., Zhang, L., Yang, L., Wu, J., Li, N., Pan, C., et al. (2021). Sustained delivery of MMP-9 siRNA via thermosensitive hydrogel accelerates diabetic wound healing. *J. Nanobiotechnology* 19 (1), 130. doi:10.1186/s12951-021-00869-6

Landen, N. X., Li, D., and Stahle, M. (2016). Transition from inflammation to proliferation: A critical step during wound healing. *Cell. Mol. Life Sci.* 73 (20), 3861–3885. doi:10.1007/s00018-016-2268-0

Lauer, G., Sollberg, S., Cole, M., Flamme, I., Sturzebecher, J., Mann, K., et al. (2000). Expression and proteolysis of vascular endothelial growth factor is increased in chronic wounds. *J. Investig. Dermatol* 115 (1), 12–18. doi:10.1046/j.1523-1747.2000.00036.x

Lee, J. B., Kim, D. H., Yoon, J. K., Park, D. B., Kim, H. S., Shin, Y. M., et al. (2020). Microchannel network hydrogel induced ischemic blood perfusion connection. *Nat. Commun.* 11 (1), 615. doi:10.1038/s41467-020-14480-0

Lei, H., Zhao, J., Li, H., and Fan, D. (2022). Paramylon hydrogel: A bioactive polysaccharides hydrogel that scavenges ROS and promotes angiogenesis for wound repair. *Carbohydr. Polym.* 289, 119467. doi:10.1016/j.carbpol.2022.119467

Li, J., Chen, J., and Kirsner, R. (2007). Pathophysiology of acute wound healing. Clin. Dermatol 25 (1), 9-18. doi:10.1016/j.clindermatol.2006.09.007

Li, J., Zhang, Y. P., and Kirsner, R. S. (2003). Angiogenesis in wound repair: Angiogenic growth factors and the extracellular matrix. *Microsc. Res. Tech.* 60 (1), 107–114. doi:10.1002/jemt.10249

Li, M., Ke, Q. F., Tao, S. C., Guo, S. C., Rui, B. Y., and Guo, Y. P. (2016). Fabrication of hydroxyapatite/chitosan composite hydrogels loaded with exosomes derived from miR-126-3p overexpressed synovial mesenchymal stem cells for diabetic chronic wound healing. *J. Mater Chem. B* 4 (42), 6830–6841. doi:10.1039/c6tb01560c

Li, Q., Wang, D., Jiang, Z., Li, R., Xue, T., Lin, C., et al. (2022a). Advances of hydrogel combined with stem cells in promoting chronic wound healing. *Front. Chem.* 10, 1038839. doi:10.3389/fchem.2022.1038839

Li, S., Wang, X., Chen, J., Guo, J., Yuan, M., Wan, G., et al. (2022b). Calcium ion crosslinked sodium alginate hydrogels containing deferoxamine and copper nanoparticles for diabetic wound healing. *Int. J. Biol. Macromol.* 202, 657–670. doi:10.1016/j.ijbiomac. 2022.01.080

Li, Y., Xu, T., Tu, Z., Dai, W., Xue, Y., Tang, C., et al. (2020). Bioactive antibacterial silica-based nanocomposites hydrogel scaffolds with high angiogenesis for promoting diabetic wound healing and skin repair. *Theranostics* 10 (11), 4929–4943. doi:10.7150/thno.41839

Liang, Y., He, J., and Guo, B. (2021). Functional hydrogels as wound dressing to enhance wound healing. ACS Nano 15 (8), 12687-12722. doi:10.1021/acsnano.1c04206

Liu, G., Zhang, F., Wang, R., London, S. D., and London, L. (2019). Salivary gland immunization via Wharton's duct activates differential T-cell responses within the salivary gland immune system. *FASEB J.* 33 (5), 6011–6022. doi:10.1096/fj.201801993R

Liu, M., Chen, Y., Zhu, Q., Tao, J., Tang, C., Ruan, H., et al. (2022). Antioxidant thermogelling formulation for burn wound healing. *Chem. Asian J.* 17 (16), e202200396. doi:10.1002/asia.202200396

Louiselle, A. E., Niemiec, S. M., Zgheib, C., and Liechty, K. W. (2021). Macrophage polarization and diabetic wound healing. *Transl. Res.* 236, 109–116. doi:10.1016/j.trsl. 2021.05.006

Luneva, O., Olekhnovich, R., and Uspenskaya, M. (2022). Bilayer hydrogels for wound dressing and tissue engineering. *Polym. (Basel)* 14 (15), 3135. doi:10.3390/polym14153135

Lv, Y., Cai, F., He, Y., Li, L., Huang, Y., Yang, J., et al. (2023). Multi-crosslinked hydrogels with strong wet adhesion, self-healing, antibacterial property, reactive oxygen species scavenging activity, and on-demand removability for seawater-immersed wound healing. *Acta Biomater.* 159, 95–110. doi:10.1016/j.actbio.2023.01.045

Madani, M., Berardi, T., and Stoopler, E. T. (2014). Anatomic and examination considerations of the oral cavity. *Med. Clin. North Am.* 98 (6), 1225–1238. doi:10.1016/j. mcna.2014.08.001

Mak, K., Manji, A., Gallant-Behm, C., Wiebe, C., Hart, D. A., Larjava, H., et al. (2009). Scarless healing of oral mucosa is characterized by faster resolution of inflammation and control of myofibroblast action compared to skin wounds in the red Duroc pig model. J. Dermatol Sci. 56 (3), 168–180. doi:10.1016/j.jdermsci.2009.09.005 Manconi, M., Manca, M. L., Caddeo, C., Cencetti, C., di Meo, C., Zoratto, N., et al. (2018). Preparation of gellan-cholesterol nanohydrogels embedding baicalin and evaluation of their wound healing activity. *Eur. J. Pharm. Biopharm.* 127, 244–249. doi:10.1016/j.ejpb.2018.02.015

Mao, L., Wang, L., Zhang, M., Ullah, M. W., Liu, L., Zhao, W., et al. (2021). *In situ* synthesized selenium nanoparticles-decorated bacterial cellulose/gelatin hydrogel with enhanced antibacterial, antioxidant, and anti-inflammatory capabilities for facilitating skin wound healing. *Adv. Healthc. Mater* 10 (14), e2100402. doi:10.1002/adhm. 202100402

Mark Welch, J. L., Ramirez-Puebla, S. T., and Borisy, G. G. (2020). Oral microbiome geography: Micron-scale habitat and niche. *Cell. Host Microbe* 28 (2), 160–168. doi:10. 1016/j.chom.2020.07.009

Masood, N., Ahmed, R., Tariq, M., Ahmed, Z., Masoud, M. S., Ali, I., et al. (2019). Silver nanoparticle impregnated chitosan-PEG hydrogel enhances wound healing in diabetes induced rabbits. *Int. J. Pharm.* 559, 23–36. doi:10.1016/j.ijpharm.2019. 01.019

Matichescu, A., Ardelean, L. C., Rusu, L. C., Craciun, D., Bratu, E. A., Babucea, M., et al. (2020). Advanced biomaterials and techniques for oral tissue engineering and regeneration-A review. *Mater. (Basel)* 13 (22), 5303. doi:10.3390/ma13225303

McCarthy, P. C., Zhang, Y., and Abebe, F. (2021). Recent applications of dual-stimuli responsive chitosan hydrogel nanocomposites as drug delivery tools. *Molecules* 26 (16), 4735. doi:10.3390/molecules26164735

Miyake, Y., Shimizu, O., Shiratsuchi, H., Tamagawa, T., and Tonogi, M. (2020). Wound healing after applying a gelatin-based hydrogel sheet to resected rat submandibular gland. *J. Oral Sci.* 62 (2), 222–225. doi:10.2334/josnusd.19-0244

Nagy, A., Nagashima, H., Cha, S., Oxford, G. E., Zelles, T., Peck, A. B., et al. (2001). Reduced oral wound healing in the NOD mouse model for type 1 autoimmune diabetes and its reversal by epidermal growth factor supplementation. *Diabetes* 50 (9), 2100–2104. doi:10.2337/diabetes.50.9.2100

Ngeow, W. C., Tan, C. C., Goh, Y. C., Deliberador, T. M., and Cheah, C. W. (2022). A narrative review on means to promote oxygenation and angiogenesis in oral wound healing. *Bioeng. (Basel)* 9 (11), 636. doi:10.3390/bioengineering9110636

Ni, Y., Turner, D., Yates, K., and Tizard, I. (2007). Stabilization of growth factors relevant to wound healing by a plant cell wall biomaterial. *Planta Med.* 73 (12), 1260–1266. doi:10.1055/s-2007-990225

Ni, Z., Yu, H., Wang, L., Huang, Y., Lu, H., Zhou, H., et al. (2022). Multistage ROSresponsive and natural polyphenol-driven prodrug hydrogels for diabetic wound healing. ACS Appl. Mater Interfaces 14 (47), 52643–52658. doi:10.1021/acsami.2c15686

Nikoloudaki, G., Creber, K., and Hamilton, D. W. (2020). Wound healing and fibrosis: A contrasting role for periostin in skin and the oral mucosa. *Am. J. Physiol. Cell. Physiol.* 318 (6), C1065–C1077. doi:10.1152/ajpcell.00035.2020

Noh, M., Choi, Y. H., An, Y. H., Tahk, D., Cho, S., Yoon, J. W., et al. (2019). Magnetic nanoparticle-embedded hydrogel sheet with a groove pattern for wound healing application. *ACS Biomater. Sci. Eng.* 5 (8), 3909–3921. doi:10.1021/acsbiomaterials. 8b01307

Norahan, M. H., Pedroza-Gonzalez, S. C., Sanchez-Salazar, M. G., Alvarez, M. M., and Trujillo de Santiago, G. (2023). Structural and biological engineering of 3D hydrogels for wound healing. *Bioact. Mater* 24, 197–235. doi:10.1016/j.bioactmat.2022.11.019

Nourian Dehkordi, A., Mirahmadi Babaheydari, F., Chehelgerdi, M., and Raeisi Dehkordi, S. (2019). Skin tissue engineering: Wound healing based on stem-cell-based therapeutic strategies. *Stem Cell. Res. Ther.* 10 (1), 111. doi:10.1186/s13287-019-1212-2

Ouyang, J., Bu, Q., Tao, N., Chen, M., Liu, H., Zhou, J., et al. (2022). A facile and general method for synthesis of antibiotic-free protein-based hydrogel: Wound dressing for the eradication of drug-resistant bacteria and biofilms. *Bioact. Mater* 18, 446–458. doi:10.1016/j.bioactmat.2022.03.033

Park, J. Y., Chung, H., DiPalma, D. T., Tai, X., and Park, J. H. (2018). Immune quiescence in the oral mucosa is maintained by a uniquely large population of highly activated Foxp3(+) regulatory T cells. *Mucosal Immunol.* 11 (4), 1092–1102. doi:10. 1038/s41385-018-0027-2

Pastar, I., Stojadinovic, O., Krzyzanowska, A., Barrientos, S., Stuelten, C., Zimmerman, K., et al. (2010). Attenuation of the transforming growth factor beta-signaling pathway in chronic venous ulcers. *Mol. Med.* 16 (3-4), 92–101. doi:10.2119/molmed.2009.00149

Pazyar, N., Yaghoobi, R., Rafiee, E., Mehrabian, A., and Feily, A. (2014). Skin wound healing and phytomedicine: A review. *Skin. Pharmacol. Physiol.* 27 (6), 303–310. doi:10. 1159/000357477

Peng, Y., He, D., Ge, X., Lu, Y., Chai, Y., Zhang, Y., et al. (2021). Construction of heparin-based hydrogel incorporated with Cu5.4O ultrasmall nanozymes for wound healing and inflammation inhibition. *Bioact. Mater* 6 (10), 3109–3124. doi:10.1016/j. bioactmat.2021.02.006

Peralta Ramos, M. L., Gonzalez, J. A., Fabian, L., Perez, C. J., Villanueva, M. E., and Copello, G. J. (2017). Sustainable and smart keratin hydrogel with pH-sensitive swelling and enhanced mechanical properties. *Mater Sci. Eng. C Mater Biol. Appl.* 78, 619–626. doi:10.1016/j.msec.2017.04.120

Perez-Rafael, S., Ivanova, K., Stefanov, I., Puiggali, J., Del Valle, L. J., Todorova, K., et al. (2021). Nanoparticle-driven self-assembling injectable hydrogels provide a multi-

factorial approach for chronic wound treatment. Acta Biomater. 134, 131–143. doi:10. 1016/j.actbio.2021.07.020

Politis, C., Schoenaers, J., Jacobs, R., and Agbaje, J. O. (2016). Wound healing problems in the mouth. *Front. Physiol.* 7, 507. doi:10.3389/fphys.2016.00507

Prime, S. S., Pring, M., Davies, M., and Paterson, I. C. (2004). TGF-beta signal transduction in oro-facial health and non-malignant disease (part I). *Crit. Rev. Oral Biol. Med.* 15 (6), 324–336. doi:10.1177/154411130401500602

Qi, W., Dong, N., Wu, L., Zhang, X., Li, H., Wu, H., et al. (2023). Promoting oral mucosal wound healing using a DCS-RuB(2)A(2) hydrogel based on a photoreactive antibacterial and sustained release of BMSCs. *Bioact. Mater* 23, 53–68. doi:10.1016/j. bioactmat.2022.10.027

Qian, Z., Wang, H., Bai, Y., Wang, Y., Tao, L., Wei, Y., et al. (2020). Improving chronic diabetic wound healing through an injectable and self-healing hydrogel with plateletrich plasma release. ACS Appl. Mater Interfaces 12 (50), 55659–55674. doi:10.1021/acsami.0c17142

Qin, R., Steel, A., and Fazel, N. (2017). Oral mucosa biology and salivary biomarkers. *Clin. Dermatol* 35 (5), 477–483. doi:10.1016/j.clindermatol.2017.06.005

Qiu, J., Wilkens, C., Barrett, K., and Meyer, A. S. (2020). Microbial enzymes catalyzing keratin degradation: Classification, structure, function. *Biotechnol. Adv.* 44, 107607. doi:10.1016/j.biotechadv.2020.107607

Qu, J., Zhao, X., Liang, Y., Zhang, T., Ma, P. X., and Guo, B. (2018). Antibacterial adhesive injectable hydrogels with rapid self-healing, extensibility and compressibility as wound dressing for joints skin wound healing. *Biomaterials* 183, 185–199. doi:10. 1016/j.biomaterials.2018.08.044

Reno, F., Rizzi, M., and Cannas, M. (2012). Effect of a gelatin hydrogel incorporating epiregulin on human keratinocyte growth. *J. Biomater. Sci. Polym. Ed.* 23 (16), 2025–2038. doi:10.1163/092050611X603872

Safari, B., Aghazadeh, M., Davaran, S., and Roshangar, L. (2022). Exosome-loaded hydrogels: A new cell-free therapeutic approach for skin regeneration. *Eur. J. Pharm. Biopharm.* 171, 50–59. doi:10.1016/j.ejpb.2021.11.002

Saleh, B., Dhaliwal, H. K., Portillo-Lara, R., Shirzaei Sani, E., Abdi, R., Amiji, M. M., et al. (2019). Local immunomodulation using an adhesive hydrogel loaded with miRNA-laden nanoparticles promotes wound healing. *Small* 15 (36), e1902232. doi:10.1002/smll.201902232

Schrementi, M. E., Ferreira, A. M., Zender, C., and DiPietro, L. A. (2008). Site-specific production of TGF-beta in oral mucosal and cutaneous wounds. *Wound Repair Regen.* 16 (1), 80–86. doi:10.1111/j.1524-475X.2007.00320.x

Scopelliti, F., Cattani, C., Dimartino, V., Mirisola, C., and Cavani, A. (2022). Platelet derivatives and the immunomodulation of wound healing. *Int. J. Mol. Sci.* 23 (15), 8370. doi:10.3390/ijms23158370

Senel, S. (2021). An overview of physical, microbiological and immune barriers of oral mucosa. *Int. J. Mol. Sci.* 22 (15), 7821. doi:10.3390/ijms22157821

Sener, G., Hilton, S. A., Osmond, M. J., Zgheib, C., Newsom, J. P., Dewberry, L., et al. (2020). Injectable, self-healable zwitterionic cryogels with sustained microRNA - cerium oxide nanoparticle release promote accelerated wound healing. *Acta Biomater.* 101, 262–272. doi:10.1016/j.actbio.2019.11.014

Sheng, L., Zhang, Z., Zhang, Y., Wang, E., Ma, B., Xu, Q., et al. (2021). A novel "hot spring"-mimetic hydrogel with excellent angiogenic properties for chronic wound healing. *Biomaterials* 264, 120414. doi:10.1016/j.biomaterials.2020.120414

Shi, C., Wang, C., Liu, H., Li, Q., Li, R., Zhang, Y., et al. (2020). Selection of appropriate wound dressing for various wounds. *Front. Bioeng. Biotechnol.* 8, 182. doi:10.3389/fbioe.2020.00182

Shi, Q., Qian, Z., Liu, D., Sun, J., Wang, X., Liu, H., et al. (2017). GMSC-derived exosomes combined with a chitosan/silk hydrogel sponge accelerates wound healing in a diabetic rat skin defect model. *Front. Physiol.* 8, 904. doi:10.3389/fphys.2017.00904

Shi, X., Chen, Z., He, Y., Lu, Q., Chen, R., Zhao, C., et al. (2022). Dual light-responsive cellulose nanofibril-based *in situ* hydrogel for drug-resistant bacteria infected wound healing. *Carbohydr. Polym.* 297, 120042. doi:10.1016/j.carbpol.2022.120042

Shiekh, P. A., Singh, A., and Kumar, A. (2020). Exosome laden oxygen releasing antioxidant and antibacterial cryogel wound dressing OxOBand alleviate diabetic and infectious wound healing. *Biomaterials* 249, 120020. doi:10.1016/j.biomaterials.2020. 120020

Siebert, L., Luna-Ceron, E., Garcia-Rivera, L. E., Oh, J., Jang, J., Rosas-Gomez, D. A., et al. (2021). Light-controlled growth factors release on tetrapodal ZnO-incorporated 3D-printed hydrogels for developing smart wound scaffold. *Adv. Funct. Mater* 31 (22), 2007555. doi:10.1002/adfm.202007555

Simoes, A., Chen, L., Chen, Z., Zhao, Y., Gao, S., Marucha, P. T., et al. (2019). Differential microRNA profile underlies the divergent healing responses in skin and oral mucosal wounds. *Sci. Rep.* 9 (1), 7160. doi:10.1038/s41598-019-43682-w

Sindrilaru, A., and Scharffetter-Kochanek, K. (2013). Disclosure of the culprits: Macrophages-versatile regulators of wound healing. *Adv. Wound Care (New Rochelle)* 2 (7), 357–368. doi:10.1089/wound.2012.0407

Singh, B., Sharma, S., and Dhiman, A. (2017). Acacia gum polysaccharide based hydrogel wound dressings: Synthesis, characterization, drug delivery and biomedical properties. *Carbohydr. Polym.* 165, 294–303. doi:10.1016/j.carbpol.2017.02.039

Smigiel, K. S., and Parks, W. C. (2018). Macrophages, wound healing, and fibrosis: Recent insights. Curr. Rheumatol. Rep. 20 (4), 17. doi:10.1007/s11926-018-0725-5

Steenhuis, P., Huntley, R. E., Gurenko, Z., Yin, L., Dale, B. A., Fazel, N., et al. (2011). Adrenergic signaling in human oral keratinocytes and wound repair. *J. Dent. Res.* 90 (2), 186–192. doi:10.1177/0022034510388034

Stojadinovic, O., Brem, H., Vouthounis, C., Lee, B., Fallon, J., Stallcup, M., et al. (2005). Molecular pathogenesis of chronic wounds: The role of beta-catenin and c-myc in the inhibition of epithelialization and wound healing. *Am. J. Pathol.* 167 (1), 59–69. doi:10.1016/s0002-9440(10)62953-7

Stojadinovic, O., Pastar, I., Nusbaum, A. G., Vukelic, S., Krzyzanowska, A., and Tomic-Canic, M. (2014). Deregulation of epidermal stem cell niche contributes to pathogenesis of nonhealing venous ulcers. *Wound Repair Regen.* 22 (2), 220–227. doi:10.1111/wrr.12142

Su, D., Tsai, H. I., Xu, Z., Yan, F., Wu, Y., Xiao, Y., et al. (2019). Exosomal PD-L1 functions as an immunosuppressant to promote wound healing. *J. Extracell. Vesicles* 9 (1), 1709262. doi:10.1080/20013078.2019.1709262

Sun, F., Li, R., Jin, F., Zhang, H., Zhang, J., Wang, T., et al. (2021). Dopamine/zinc oxide doped poly(N-hydroxyethyl acrylamide)/agar dual network hydrogel with super self-healing, antibacterial and tissue adhesion functions designed for transdermal patch. *J. Mater Chem. B* 9 (27), 5492–5502. doi:10.1039/d1tb00822f

Sun, J., Chen, T., Zhao, B., Fan, W., Shen, Y., Wei, H., et al. (2023a). Acceleration of oral wound healing under diabetes mellitus conditions using bioadhesive hydrogel. ACS Appl. Mater Interfaces 15 (1), 416–431. doi:10.1021/acsami.2c17424

Sun, L., Huang, Y., Bian, Z., Petrosino, J., Fan, Z., Wang, Y., et al. (2016). Sundewinspired adhesive hydrogels combined with adipose-derived stem cells for wound healing. ACS Appl. Mater Interfaces 8 (3), 2423–2434. doi:10.1021/acsami.5b11811

Sun, Y., Jing, X., Liu, Y., Yu, B., Hu, H., Cong, H., et al. (2023). A chitosan derivativecrosslinked hydrogel with controllable release of polydeoxyribonucleotides for wound treatment. *Carbohydr. Polym.* 300, 120298. doi:10.1016/j.carbpol.2022.120298

Sund-Levander, M., Forsberg, C., and Wahren, L. K. (2002). Normal oral, rectal, tympanic and axillary body temperature in adult men and women: A systematic literature review. *Scand. J. Caring Sci.* 16 (2), 122–128. doi:10.1046/j.1471-6712.2002. 00069.x

Szpaderska, A. M., Walsh, C. G., Steinberg, M. J., and DiPietro, L. A. (2005). Distinct patterns of angiogenesis in oral and skin wounds. *J. Dent. Res.* 84 (4), 309–314. doi:10. 1177/154405910508400403

Szpaderska, A. M., Zuckerman, J. D., and DiPietro, L. A. (2003). Differential injury responses in oral mucosal and cutaneous wounds. *J. Dent. Res.* 82 (8), 621–626. doi:10. 1177/154405910308200810

Teixeira, M. O., Antunes, J. C., and Felgueiras, H. P. (2021). Recent advances in fiberhydrogel composites for wound healing and drug delivery systems. *Antibiot. (Basel)* 10 (3), 248. doi:10.3390/antibiotics10030248

Toma, A. I., Fuller, J. M., Willett, N. J., and Goudy, S. L. (2021). Oral wound healing models and emerging regenerative therapies. *Transl. Res.* 236, 17–34. doi:10.1016/j.trsl. 2021.06.003

Torres, P., Castro, M., Reyes, M., and Torres, V. A. (2018). Histatins, wound healing, and cell migration. *Oral Dis.* 24 (7), 1150–1160. doi:10.1111/odi.12816

Tu, C., Lu, H., Zhou, T., Zhang, W., Deng, L., Cao, W., et al. (2022). Promoting the healing of infected diabetic wound by an anti-bacterial and nano-enzyme-containing hydrogel with inflammation-suppressing, ROS-scavenging, oxygen and nitric oxide-generating properties. *Biomaterials* 286, 121597. doi:10.1016/j.biomaterials.2022. 121597

Turabelidze, A., Guo, S., Chung, A. Y., Chen, L., Dai, Y., Marucha, P. T., et al. (2014). Intrinsic differences between oral and skin keratinocytes. *PLoS One* 9 (9), e101480. doi:10.1371/journal.pone.0101480

Ulaganathan, G., Babu, S. S., Senthilmoorthy, M., Prasad, V., Kalaiselvan, S., and Kumar, R. S. A. (2020). Retrospective analysis of oral and maxillofacial biopsies: An institutional study. *J. Pharm. Bioallied Sci.* 12 (1), S468–S471. doi:10.4103/jpbs.JPBS\_ 141\_20

van Veen, M. M., Tavares-Brito, J., van Veen, B. M., Dusseldorp, J. R., Werker, P. M. N., Dijkstra, P. U., et al. (2019). Association of regional facial dysfunction with facial palsy-related quality of life. *JAMA Facial Plast. Surg.* 21 (1), 32–37. doi:10.1001/jamafacial.2018.0804

Velnar, T., Bailey, T., and Smrkolj, V. (2009). The wound healing process: An overview of the cellular and molecular mechanisms. *J. Int. Med. Res.* 37 (5), 1528–1542. doi:10.1177/147323000903700531

Visse, R., and Nagase, H. (2003). Matrix metalloproteinases and tissue inhibitors of metalloproteinases: Structure, function, and biochemistry. *Circ. Res.* 92 (8), 827–839. doi:10.1161/01.RES.0000070112.80711.3D

Waasdorp, M., Krom, B. P., Bikker, F. J., van Zuijlen, P. P. M., Niessen, F. B., and Gibbs, S. (2021). The bigger picture: Why oral mucosa heals better than skin. *Biomolecules* 11 (8), 1165. doi:10.3390/biom11081165

Wagener, F. A., Carels, C. E., and Lundvig, D. M. (2013). Targeting the redox balance in inflammatory skin conditions. *Int. J. Mol. Sci.* 14 (5), 9126–9167. doi:10.3390/ ijms14059126 Wang, C., Wang, M., Xu, T., Zhang, X., Lin, C., Gao, W., et al. (2019a). Engineering bioactive self-healing antibacterial exosomes hydrogel for promoting chronic diabetic wound healing and complete skin regeneration. *Theranostics* 9 (1), 65–76. doi:10.7150/ thno.29766

Wang, L., Du, J., Han, X., Dou, J., Shen, J., and Yuan, J. (2020). Self-crosslinked keratin nanoparticles for pH and GSH dual responsive drug carriers. *J. Biomater. Sci. Polym. Ed.* 31 (15), 1994–2006. doi:10.1080/09205063.2020.1788371

Wang, L., Hu, C., and Shao, L. (2017). The antimicrobial activity of nanoparticles: Present situation and prospects for the future. *Int. J. Nanomedicine* 12, 1227–1249. doi:10.2147/IJN.S121956

Wang, M., Huang, H., Huang, C., Liu, S., and Peng, X. (2022a). pH-responsive magnolol nanocapsule-embedded magnolol-grafted-chitosan hydrochloride hydrogels for promoting wound healing. *Carbohydr. Polym.* 292, 119643. doi:10.1016/j.carbpol. 2022.119643

Wang, P., Huang, S., Hu, Z., Yang, W., Lan, Y., Zhu, J., et al. (2019b). *In situ* formed anti-inflammatory hydrogel loading plasmid DNA encoding VEGF for burn wound healing. *Acta Biomater*. 100, 191–201. doi:10.1016/j.actbio.2019.10.004

Wang, P., Lv, C., Zhou, X., Wu, Z., Wang, Z., Wang, Y., et al. (2022b). Tannin-bridged magnetic responsive multifunctional hydrogel for enhanced wound healing by mechanical stimulation-induced early vascularization. *J. Mater Chem. B* 10 (38), 7808–7826. doi:10.1039/d2tb01378a

Wang, X., Tang, X., Li, N., Sun, Y., Yu, R., Zhang, J., et al. (2022c). A multifunctional black phosphorus-based adhesive patch intrinsically induces partial EMT for effective burn wound healing. *Biomater. Sci.* 11 (1), 235–247. doi:10.1039/d2bm01625g

Wang, Z., Zhang, Y., Yin, Y., Liu, J., Li, P., Zhao, Y., et al. (2022). High-strength and injectable supramolecular hydrogel self-assembled by monomeric nucleoside for tooth-extraction wound healing. *Adv. Mater* 34 (13), e2108300. doi:10.1002/adma. 202108300

Wathoni, N., Rusdiana, T., Hasanah, A. N., Muhtadi, A., Pratiwi, E. D., Mahmudah, R., et al. (2019). Sacran hydrogel film containing keratinocyte growth factor accelerates wound healing by stimulating fibroblast migration and Re-epithelization. *Chem. Pharm. Bull. (Tokyo)* 67 (8), 849–854. doi:10.1248/cpb.c19-00291

Wen, X., Xi, K., Tang, Y., Bian, J., Qin, Y., Xiao, W., et al. (2023). Immunized microspheres engineered hydrogel membrane for reprogramming macrophage and mucosal repair. *Small* 19 (15), e2207030. doi:10.1002/smll.202207030

Weng, H., Jia, W., Li, M., and Chen, Z. (2022). New injectable chitosan-hyaluronic acid based hydrogels for hemostasis and wound healing. *Carbohydr. Polym.* 294, 119767. doi:10.1016/j.carbpol.2022.119767

Wu, J., Pan, Z., Zhao, Z. Y., Wang, M. H., Dong, L., Gao, H. L., et al. (2022a). Anti-Swelling, robust, and adhesive extracellular matrix-mimicking hydrogel used as intraoral dressing. *Adv. Mater* 34 (20), e2200115. doi:10.1002/adma.202200115

Wu, J., Zhu, J., He, C., Xiao, Z., Ye, J., Li, Y., et al. (2016). Comparative study of heparinpoloxamer hydrogel modified bFGF and aFGF for *in vivo* wound healing efficiency. ACS Appl. Mater Interfaces 8 (29), 18710–18721. doi:10.1021/acsami.6b06047

Wu, Y., Wang, Y., Long, L., Hu, C., Kong, Q., and Wang, Y. (2022b). A spatiotemporal release platform based on pH/ROS stimuli-responsive hydrogel in wound repairing. *J. Control Release* 341, 147–165. doi:10.1016/j.jconrel.2021.11.027

Xiao, Y., Zhao, H., Ma, X., Gu, Z., Wu, X., Zhao, L., et al. (2022). Hydrogel dressing containing basic fibroblast growth factor accelerating chronic wound healing in aged mouse model. *Molecules* 27 (19), 6361. doi:10.3390/molecules27196361

Xu, H., Wang, J., Wu, D., and Qin, D. (2022). A hybrid hydrogel encapsulating human umbilical cord mesenchymal stem cells enhances diabetic wound healing. *J. Mater Sci. Mater Med.* 33 (8), 60. doi:10.1007/s10856-022-06681-4

Xu, J., Younis, M. R., Zhang, Z., Feng, Y., Su, L., Que, Y., et al. (2023). Mild heatassisted polydopamine/alginate hydrogel containing low-dose nanoselenium for facilitating infected wound healing. *ACS Appl. Mater Interfaces* 15 (6), 7841–7854. doi:10.1021/acsami.2c21516

Xu, L., Zhang, X., Zhu, C., Zhang, Y., Fu, C., Yang, B., et al. (2013). Nonionic polymer cross-linked chitosan hydrogel: Preparation and bioevaluation. *J. Biomater. Sci. Polym. Ed.* 24 (13), 1564–1574. doi:10.1080/09205063.2013.781934

Xu, Q., Chang, M., Zhang, Y., Wang, E., Xing, M., Gao, L., et al. (2020). PDA/Cu bioactive hydrogel with "hot ions effect" for inhibition of drug-resistant bacteria and enhancement of infectious skin wound healing. ACS Appl. Mater Interfaces 12 (28), 31255–31269. doi:10.1021/acsami.0c08890

Yadi, M., Mostafavi, E., Saleh, B., Davaran, S., Aliyeva, I., Khalilov, R., et al. (2018). Current developments in green synthesis of metallic nanoparticles using plant extracts: A review. *Artif. Cells Nanomed Biotechnol.* 46 (3), S336–S343. doi:10.1080/21691401. 2018.1492931

Yang, C., Chen, Y., Huang, H., Fan, S., Yang, C., Wang, L., et al. (2021). ROSeliminating carboxymethyl chitosan hydrogel to enhance burn wound-healing efficacy. *Front. Pharmacol.* 12, 679580. doi:10.3389/fphar.2021.679580

Yang, J., Chen, Z., Pan, D., Li, H., and Shen, J. (2020a). <p&gt;Umbilical cord-derived mesenchymal stem cell-derived exosomes combined pluronic F127 hydrogel promote

chronic diabetic wound healing and complete skin regeneration</p&gt;. Int. J. Nanomedicine 15, 5911–5926. doi:10.2147/IJN.S249129

Yang, M., He, S., Su, Z., Yang, Z., Liang, X., and Wu, Y. (2020b). Thermosensitive injectable chitosan/collagen/ $\beta$ -glycerophosphate composite hydrogels for enhancing wound healing by encapsulating mesenchymal stem cell spheroids. *ACS Omega* 5 (33), 21015–21023. doi:10.1021/acsomega.0c02580

Yang, Y., Liang, Y., Chen, J., Duan, X., and Guo, B. (2022). Mussel-inspired adhesive antioxidant antibacterial hemostatic composite hydrogel wound dressing via photopolymerization for infected skin wound healing. *Bioact. Mater* 8, 341–354. doi:10.1016/j.bioactmat.2021.06.014

Yu, M., Liu, W., Li, J., Lu, J., Lu, H., Jia, W., et al. (2020). Exosomes derived from atorvastatin-pretreated MSC accelerate diabetic wound repair by enhancing angiogenesis via AKT/eNOS pathway. *Stem Cell. Res. Ther.* 11 (1), 350. doi:10.1186/s13287-020-01824-2

Yuan, J., Zhang, D., He, X., Ni, Y., Che, L., Wu, J., et al. (2021). Cationic peptide-based salt-responsive antibacterial hydrogel dressings for wound healing. *Int. J. Biol. Macromol.* 190, 754–762. doi:10.1016/j.ijbiomac.2021.09.019

Zang, S., Mu, R., Chen, F., Wei, X., Zhu, L., Han, B., et al. (2019). Injectable chitosan/ $\beta$ -glycerophosphate hydrogels with sustained release of BMP-7 and ornidazole in periodontal wound healing of class III furcation defects. *Mater Sci. Eng. C Mater Biol. Appl.* 99, 919–928. doi:10.1016/j.msec.2019.02.024

Zeng, R., Lin, C., Lin, Z., Chen, H., Lu, W., Lin, C., et al. (2018). Approaches to cutaneous wound healing: Basics and future directions. *Cell. Tissue Res.* 374 (2), 217–232. doi:10.1007/s00441-018-2830-1

Zhai, Q., Dong, Z., Wang, W., Li, B., and Jin, Y. (2019). Dental stem cell and dental tissue regeneration. *Front. Med.* 13 (2), 152–159. doi:10.1007/s11684-018-0628-x

Zhang, A., Liu, Y., Qin, D., Sun, M., Wang, T., and Chen, X. (2020a). Research status of self-healing hydrogel for wound management: A review. *Int. J. Biol. Macromol.* 164, 2108–2123. doi:10.1016/j.ijbiomac.2020.08.109

Zhang, D. L., Gu, L. J., Liu, L., Wang, C. Y., Sun, B. S., Li, Z., et al. (2009). Effect of Wnt signaling pathway on wound healing. *Biochem. Biophys. Res. Commun.* 378 (2), 149–151. doi:10.1016/j.bbrc.2008.11.011

Zhang, J., Zheng, Y., Lee, J., Hua, J., Li, S., Panchamukhi, A., et al. (2021a). A pulsatile release platform based on photo-induced imine-crosslinking hydrogel promotes scarless wound healing. *Nat. Commun.* 12 (1), 1670. doi:10.1038/s41467-021-21964-0

Zhang, S., Ge, G., Qin, Y., Li, W., Dong, J., Mei, J., et al. (2023). Recent advances in responsive hydrogels for diabetic wound healing. *Mater Today Bio* 18, 100508. doi:10. 1016/j.mtbio.2022.100508

Zhang, T., Wang, X. F., Wang, Z. C., Lou, D., Fang, Q. Q., Hu, Y. Y., et al. (2020b). Current potential therapeutic strategies targeting the TGF- $\beta$ /Smad signaling pathway to attenuate keloid and hypertrophic scar formation. *Biomed. Pharmacother.* 129, 110287. doi:10.1016/j.biopha.2020.110287

Zhang, W., Bao, B., Jiang, F., Zhang, Y., Zhou, R., Lu, Y., et al. (2021b). Promoting oral mucosal wound healing with a hydrogel adhesive based on a phototriggered S-nitrosylation coupling reaction. *Adv. Mater* 33 (48), e2105667. doi:10.1002/adma. 202105667

Zhao, C., Chen, R., Chen, Z., Lu, Q., Zhu, H., Bu, Q., et al. (2021). Bioinspired multifunctional cellulose nanofibril-based *in situ* liquid wound dressing for multiple synergistic therapy of the postoperative infected wound. *ACS Appl. Mater Interfaces* 13 (43), 51578–51591. doi:10.1021/acsami.1c18221

Zhao, H., Huang, J., Li, Y., Lv, X., Zhou, H., Wang, H., et al. (2020). ROS-scavenging hydrogel to promote healing of bacteria infected diabetic wounds. *Biomaterials* 258, 120286. doi:10.1016/j.biomaterials.2020.120286

Zhao, N., Coyne, J., Xu, M., Zhang, X., Suzuki, A., Shi, P., et al. (2019). Assembly of bifunctional aptamer-fibrinogen macromer for VEGF delivery and skin wound healing. *Chem. Mater* 31 (3), 1006–1015. doi:10.1021/acs.chemmater.8b04486

Zhao, Y., Liu, X., Peng, X., Zheng, Y., Cheng, Z., Sun, S., et al. (2022). A poloxamer/ hyaluronic acid/chitosan-based thermosensitive hydrogel that releases dihydromyricetin to promote wound healing. *Int. J. Biol. Macromol.* 216, 475–486. doi:10.1016/j.ijbiomac.2022.06.210

Zhou, S., Yang, D., Yang, D., Guo, Y., Hu, R., Li, Y., et al. (2023a). Injectable, selfhealing and multiple responsive histamine modified hyaluronic acid hydrogels with potentialities in drug delivery, antibacterial and tissue engineering. *Macromol. Rapid Commun.* 44 (3), e2200674. doi:10.1002/marc.202200674

Zhou, W. C., Tan, P. F., Chen, X. H., Cen, Y., You, C., Tan, L., et al. (2019). Berberine-Incorporated shape memory fiber applied as a novel surgical suture. *Front. Pharmacol.* 10, 1506. doi:10.3389/fphar.2019.01506

Zhou, X., Guo, Y., Yang, K., Liu, P., and Wang, J. (2022). The signaling pathways of traditional Chinese medicine in promoting diabetic wound healing. *J. Ethnopharmacol.* 282, 114662. doi:10.1016/j.jep.2021.114662

Zhou, Y., Yang, Y., Liu, R., Zhou, Q., Lu, H., and Zhang, W. (2023b). Research progress of polydopamine hydrogel in the prevention and treatment of oral diseases. *Int. J. Nanomedicine* 18, 2623–2645. doi:10.2147/IJN.S407044