

Supplementary Materials

Table S1: List of primers for virulence and regulatory genes, *Vibrio* seventh pathogenicity island and PCR conditions used in this study.

Primer	Target	Sequence	Size in bp	PCR Cyclic conditions	References
toxR-F	<i>toxR</i>	CCTTCGATCCCCTAAGCAATAC	779	ID: 94°C for 2 min	Singh et al. 2002
toxR-R		AGGGTTAGCAACGATGCGTAAG		D: 94°C for 1 min	
ace-F	<i>ace</i>	TAAGGATGTGCTTATGATGGACACCC	316	A: 62°C for 1 min	
ace-R		CGTGATGAATAAAGATACTCATAGG		E: 72 °C for 1 min	
tcpA-F	<i>tcpA^{class}</i>	CACGATAAGAAAACCGGTCAAGAG		D: 94°C for 1 min	
tcpA-class		TTACCAAATGCAACGCCGAATG	620	A: 54°C for 1 min	
tcpA-El Tor		CGAAAGCACCTTCTTTCACACGTTG	453	E: 72 °C for 1 min	
ctxA-F	<i>ctxA</i>	CGGGCAGATTCTAGACCTCCTG	564	FE:72°C for 10 min	
ctxA-R		CGATGATCTTGGAGCATTCCCAC			
ompU-F	<i>ompU</i>	ACGCTGACGGAATCAACCAAAG	869		
ompU-R		GCGGAAGTTTGGCTTGAAGTAG			
zot-F	<i>zot</i>	TCGCTTAACGATGGCGCGTTTT	947		
zot-R		AACCCCGTTTCACTTCTACCCA			
tcpI-F	<i>tcpI</i>	TAGCCTTAGTTCTCAGCAGGCA	862	D: 94°C for 1 min	Faruque et al. 1998
tcpI-R		GGCAATAGTGTCGAGCTCGTTA		A: 62°C for 1 min E: 72 °C for 1 min	
hlyA ^{ETF}	<i>hlyA^{class/El Tor}</i>	GGCAAACAGCGAAACAAATACC	481	ID: 94°C for 2 min	Rivera et al. 2001
hlyA ^{CF}		GAGCCGGCATTCTCATCTGAAT	727	D: 94°C for 1 min	
hlyA-R		CTCAGCGGGCTAATACGGTTTA		A: 62°C for 1 min E: 72 °C for 1 min EF:72°C for 10 min	

ctxB-F	ctxB	GATACACATAATAGAATTAAGGATG	449	ID: 94°C for 2 min	35 cycles	Mohapatra et al. 2011
ctxB-R		GGTTGCTTCTCATCATCGAACCCAC		D: 94°C for 1 min A: 57°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
rstR ^{class} -F	rstR ^{class}	TCTCATCAGCAAAGCCTCCATC	243	ID: 94°C for 2 min		
rstR ^{class} -R		GTAGCAAATGGTATCGGCGTTGG		D: 94°C for 30 sec		
rstR ^{ET} -F	rstR ^{El Tor}	GCACCATGATTTAAGATGCTC	320	A: 62°C for 30 sec	35 cycles	
rstR ^{ET} -R		GGCAATTAATAAGACTCAGGCAC		E: 72 °C for 30 sec FE:72°C for 10 min		
CII-F	chromosomal	CTCACGCTGAACAGCAAGTC	800	ID: 94°C for 2 min	35 cycles	Maiti et al. 2006.
CII-R	location of CTXphage	TTGCTTGAATCGAAAGGACA		D: 94°C for 1 min A: 52°C for 1 min E: 72 °C for 1 min FE:72°C for 10min		
ctxB-3	ctxB ^{Haiti}	GTTTTACTATCTTCAGCATATGCGA	191	ID: 94°C for 2 min	30 cycles	Naha et al. 2012 (DMAMA-PCR)
Rv-cla		CCTGGTACTTCTACTTGAAACG		D: 94°C for 30 sec A: 56°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
ctxB-4	ctxB ^{class}	GTTTTACTATCTTCAGCATATGCGC	191	ID: 94°C for 2 min		
Rv-cla		CCTGGTACTTCTACTTGAAACG		D: 94°C for 30 sec A: 60°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		

Fw-com	<i>ctxB</i> ^{El Tor}	ACTATCTTCAGCATATGCACATGG	254	ID: 94°C for 2 min	30 cycles	Morita et al.2008 (MAMA-PCR)
Re-elt		CCTGGTACTTCTACTTGAAACA		D: 94°C for 30 sec A: 54°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
smp-F-VC2346	<i>smp</i>	GCAACTGTGCTAGCAGTTGCCGTG	405	ID: 94°C for 2 min D: 94°C for 1 min	30 cycles	Grim et al.2010
smp-R-VC2346		GCCTGTTTCAAACGTGATGCGTA		A: 65°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
dcd-820F	<i>VC0175</i>	GCTTATTCAGC GCCCTCAGG TC	584	ID: 94°C for 2 min D: 94°C for 1 min	30 cycles	
dcd-1403R		TAG CGT CGA GAT GAC ACA CCT TCG		A: 65°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
VSPI-F	VC0180-	GCCGAGAACTCTAAAGCG CTTCTC	331	ID: 94°C for 2 min D: 94°C for 1 min	30 cycles	
VSPI-R	VC0181	CCAAGGTACAGATGAGTACCAGCA		A: 61°C for 30 sec E: 72 °C for 30 sec FE:72°C for 10 min		
VC0174-F	Chr-I –VSPI insertion	AAACTGGCGACCTTTGAGCAAGC	1321	ID: 94°C for 2 min D: 94°C for 1 min	30 cycles	
VC0186-R		GATGGTAGCCTGACGCTGCATCTG		A: 65°C for 1 min E: 72 °C for 1 min FE:72°C for 10 min		

VCA0695-F	Chr-II-VSP II	ATAGCGGGAGTTGGCTCTGCA	957	ID: 94°C for 2 min	
VCA0697-R	insertion	GGTGACTTGGTGCCCATCGTA		D: 94°C for 1 min A: 65°C for 1 min E: 72 °C for 1 min FE:72°C for 10 min	30 cycles

ID: Initial denaturation; D: Denaturation; A: Annealing; E: Extension; FE: Final extension.

References

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Table S2. Multilocus sequence typing (MLST) primers for five housekeeping genes and PCR conditions used in this study.

Gene	Gene Product	Direction	Primer Sequence(s)	PCR cyclic conditions	References
<i>dnaE</i>	DNA polymerase III alpha subunit	Forward	CgRATMACCgCTTTCgCCg	ID: 98°C for 30 sec D: 98°C for 10 sec A: 50-52°C for 30 sec E: 70°C for 30 sec FE: 70°C for 10 min } 35 cycles	Garg et al. 2003
		Reverse	gAKATgTgTgAgCTgTTTgC		
<i>lap</i>	Leucine amino-peptidase	Forward	gAAgAggTCggTTTgCgAgg		
		Reverse	gTTTgAATggTgAgCggTTTgCT		
<i>pgm</i>	Phosphoglucomutase	Forward	CCKTCSCAYAACCCgCC		
		Reverse	TCRACRAACCATTTgAADCC		
<i>recA</i>	Recombination Repair Protein	Forward	gAAACCATTTTCgACCggTTC		Kotetishvili et al. 2003
		Reverse	CCgTTATAgCTgTACCAAgCgCCC		
<i>asd</i>	Aspartate semialdehyde dehydrogenase	Forward	CgACTACgACATTCTC		Karaolis et al. 1995
		Reverse	gTTATCCgCCCACTACCC		

ID: initial denaturation; D: denaturation; A: Annealing; E: extension; FE: final extension.

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Table S3. Temporal effect of DNase I, Proteinase K, and NaIO₄ on biofilm grown in *Leuria* Bertani Broth (Mann-Whitney test).

Sr. No	Treatment of Bio-film with different reagents	Mann-Whitney test (<i>p-value</i> < 0.05)
1	Control Vs DNase I	< 0.0001
2	Control Vs Proteinase K	0.0043
3	Control Vs NaIO ₄	<0.0001

Fig. S1a. Polymorphic sites of *asd* loci among *V. cholerae* isolates. The nucleotide designation was shown for allele one and for rest of alleles sites only polymorphism is shown along with synonymous and non-synonymous polymorphism.

[illegible]

Fig. S1b. Polymorphic sites of *dnaE* loci among *V. cholerae* isolates. The nucleotide designation was shown for allele one and for rest of alleles sites only polymorphism is shown along with synonymous and non-synonymous polymorphism.

[illegible]

Fig. S1c. Polymorphic sites of *lap* loci among *V. cholerae* isolates. The nucleotide designation was shown for allele one and for rest of alleles sites only polymorphism is shown along with synonymous and non-synonymous polymorphism.

[illegible]

pgm

[illegible]

Fig. S1e. Polymorphic sites of *recA* loci among *V. cholerae* isolates. The nucleotide designation was shown for allele one and for rest of alleles sites only polymorphism is shown along with synonymous and non-synonymous polymorphism.

recA

	1 1 1 1 1 1 1 2 2 2 2 2 2 2 3 3 3 3 3 3 4 4 4 5 5 5 5 6 6 6 6 6 6 6 6 6 6 6 6 6																																															
	2 4 5 9 9 0 3 3 5 7 8 9 3 5 5 6 6 6 7 8 0 2 3 6 6 8 5 6 7 5 5 5 9 0 0 1 3 4 4 5 5 5 8																																															
	1 3 5 6 9 4 4 0 8 2 5 6 6 9 3 8 0 2 5 1 4 5 9 7 6 5 9 3 9 3 3 5 9 4 5 8 7 0 8 5 3 0 8 1 5 9 6																																															
Allele_1	T	G	T	G	T	A	A	G	A	A	C	A	G	A	A	T	T	G	C	C	G	A	G	G	T	G	A	A	T	A	T	T	T	G	A	T	G	T	G	T	C	G	G	A				
Allele_2	G	A	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_4	G	A	A	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_5	G	A	A	A	C	T	G	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_6	G	A	A	A	C	T	G	A	T	G	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_7	G	A	A	A	C	T	G	A	T	G	T	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_8	G	A	A	A	C	T	G	A	T	G	T	G	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_9	G	A	A	A	C	T	G	A	T	G	T	G	A	G	.	.	A	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_10	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_11	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_12	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_13	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_14	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_15	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_16	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.		
Allele_17	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.		
Allele_18	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.		
Allele_19	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	.	T	.	.	A	.	.	.	T	.	A	.		
Allele_21	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	.	.	A	.	.	.	T	.	A	.		
Allele_22	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	.	A	.	.	.	T	.	A	.		
Allele_23	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	.	.	.	T	.	A	.		
Allele_24	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	C	.	.	.	T	.	A	.	
Allele_25	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	C	C	.	.	.	T	.	A	.
Allele_26	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	C	C	.	C	T	.	A	.	
Allele_27	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	C	C	.	C	T	T	A	.	
Allele_28	G	A	A	A	C	T	G	A	T	G	T	G	A	G	T	A	G	A	G	T	.	.	C	T	C	A	T	G	T	A	T	C	A	G	T	T	G	A	A	C	C	.	C	T	T	A	G	
Allele_35	G	A	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_36	G	C	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_38	G	A	.	A	G	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_39	G	A	.	A	G	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_41	G	A	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	.	A	.			
Allele_42	G	A	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	A	.	T	.	A	.		
Allele_43	G	A	.	A	C	T	.	A	T	.	.	.	A	.	.	A	.	.	.	.	.	.	T	T	A	.	.	T	A	T	.	A	G	.	T	.	.	A	.	.	.	T	A	A	.			
	S	N	N	S	S	S	S	N	S	S	S	S	N	S	S	N	S	S	N	S	S	N	S	S	N	S	S	S	N	S	S	S	N	S	S	S	S	S	S	S	N	S	S	S	S	S	N	S

Fig. S2. Dendrogram presenting the genomic fingerprint pattern of *V. cholerae* O1 and O139 strains isolated from clinical and environmental sources in India. The cluster analysis was carried out using 1.5 % optimization, 1.5 % tolerance and >95 % similarity matrix of Dice similarity coefficient of pulsed-field gel electrophoresis. The dendrogram was generated by pulsed-field gel electrophoresis of total chromosomal DNA digested with *NotI* restriction enzyme and correlation between their pulsotype and ST types with respect to their sources and year of isolation.

