Supplementary Material

An allosteric-probe for detection of alkaline phosphatase activity and its application in immunoassay

Jingjing Guo1, Mingxuan Gao1, Yanling Song2\*, Li Lin1, Kaifeng Zhao1, Tian Tian1, Dan Liu1, Zhi Zhu1\*, Chaoyong James Yang1,2

1 MOE Key Laboratory of Spectrochemical Analysis & Instrumentation, Collaborative Innovation Center of Chemistry for Energy Materials, Key Laboratory for Chemical Biology of Fujian Province, State Key Laboratory of Physical Chemistry of Solid Surfaces, College of Chemistry and Chemical Engineering, Xiamen University, Xiamen 361005 (China)

2 Institute of Molecular Medicine, Renji Hospital, School of Medicine, Shanghai Jiao Tong University, Shanghai 200240 (China)

**\* Correspondence:** Zhi Zhu, [zhuzhi@xmu.edu.cn](mailto:zhuzhi@xmu.edu.cn);

Yanling Song, [songyanling2012@hotmail.com](mailto:songyanling2012@hotmail.com).

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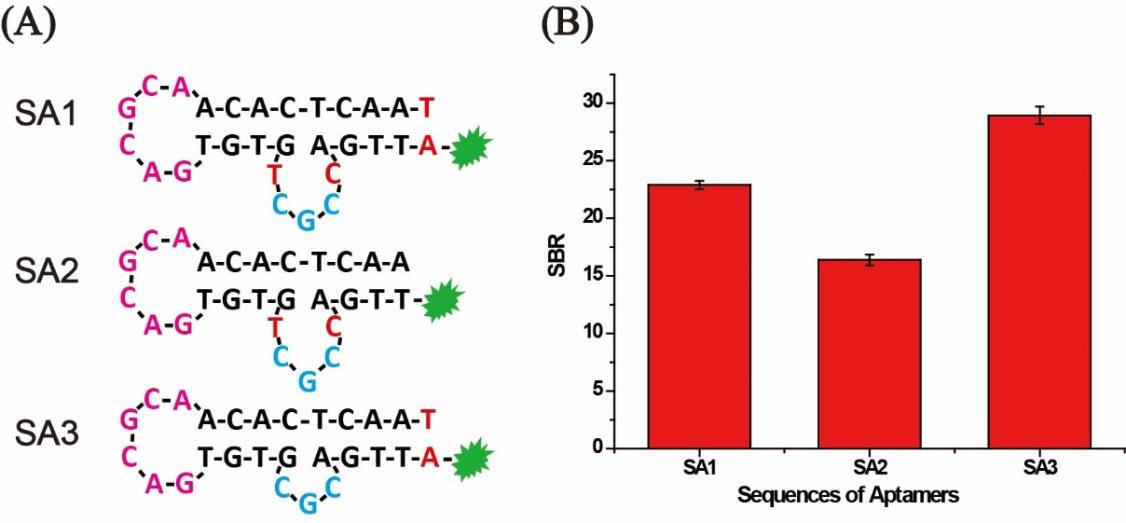
**References**  S-9

|  |  |
| --- | --- |
|  | Sequences |
| SA1 | FAM-5’-ATTGACCGCTGTGTGACGCAACACTCAAT-3’ |
| cDNA1 | PO4-5’- ATTGAGTGTTGCGTCACACAGCGGTCAAT-3’ |
| SA2 | FAM-5’-TTGACCGCTGTGTGACGCAACACTCAA-3’ |
| cDNA2 | PO4-5’- TTGAGTGTTGCGTCACACAGCGGTCAA-3’ |
| SA3 | FAM-5’-ATTGACGCGTGTGACGCAACACTCAAT-3’ |
| cDNA3 | PO4-5’-ATTGAGTGTTGCGTCACACGCGTCAAT-3’ |

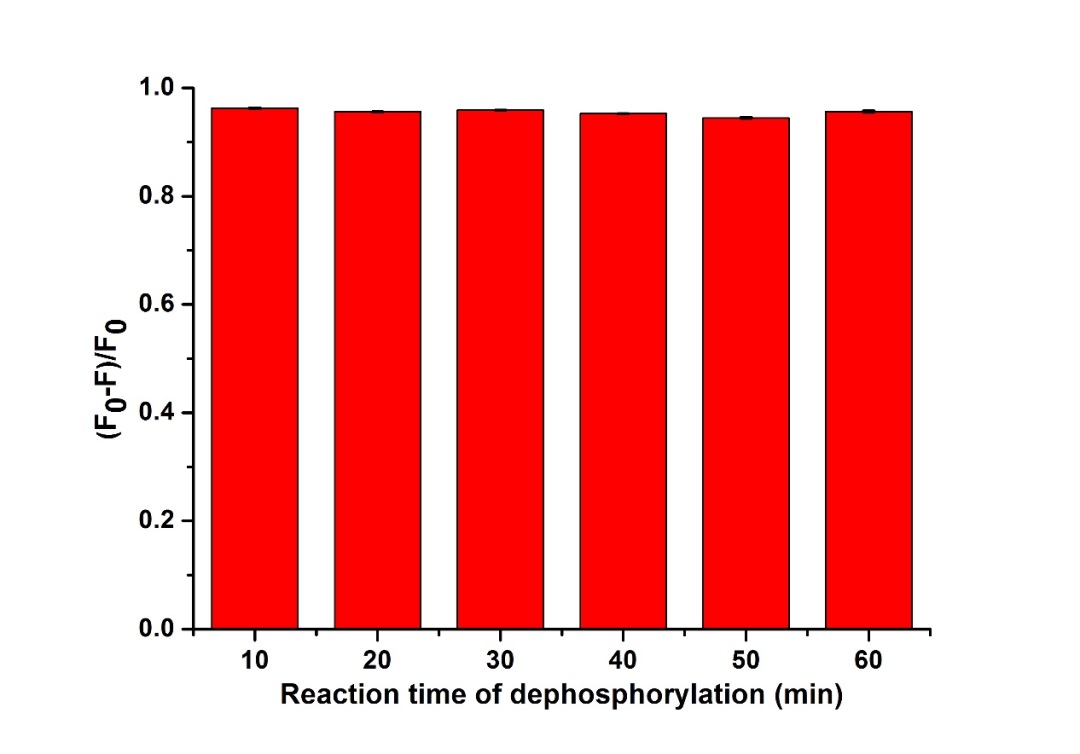
**Table S1.** DNA sequences list.

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| Detection  methods | Limit of Detection (LOD) | The minimum incubation time required for ALP detection | Biological Sample | Reference |
| Fluorescence | 0.19 U/L | ~1 h | 1% human serum | 1 |
| Fluorescence | 2.17 U/L | ~1.5 h | 2% human serum | 2 |
| Fluorescence | 60 U/L | ~2 h | Living cells | 3 |
| Fluorescence | 10 U/L | 20 min | Not given | 4 |
| Absorbance | 32 U/L | ~1 h | 1% A549 cell lysate | 5 |
| Fluorescence | 5 U/L | ~2 h | 1% bovine serum | 6 |
| Absorbance | 0.1 U/L | ~4 h | 1% human serum | 7 |
| Fluorescence | 12 U/L | 30 min | 20% human serum and cell lysate | This work |

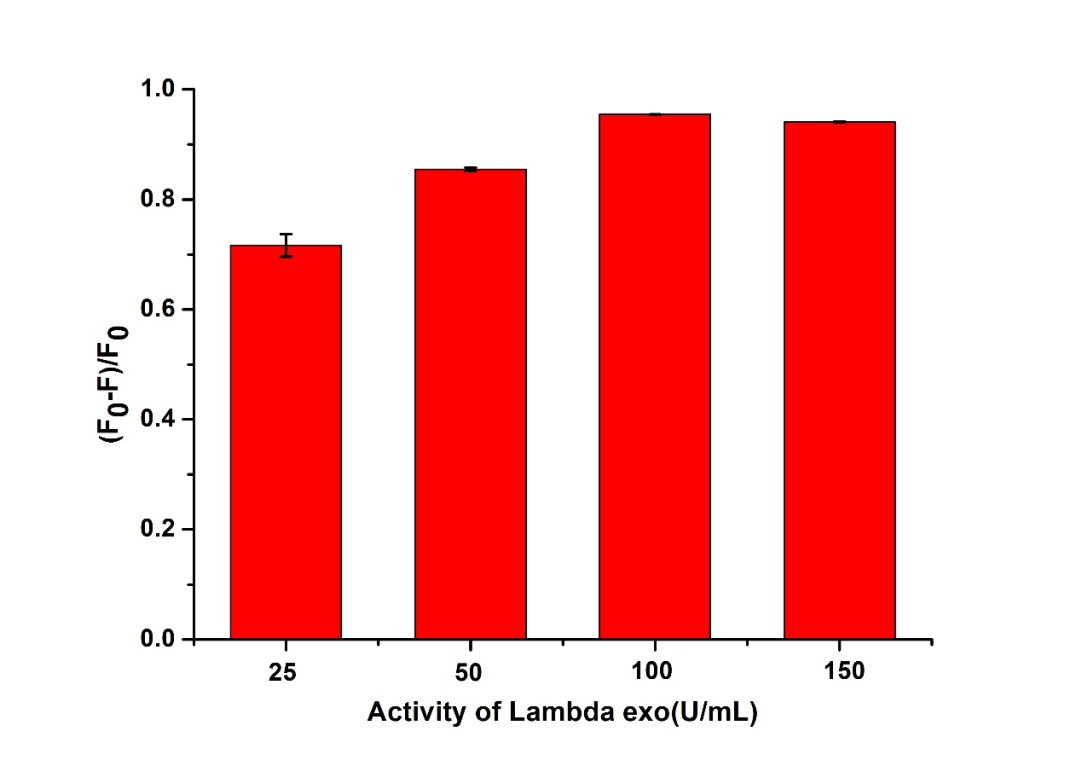
**Table S2**. Comparison of several recent detection methods for alkaline phosphatase.



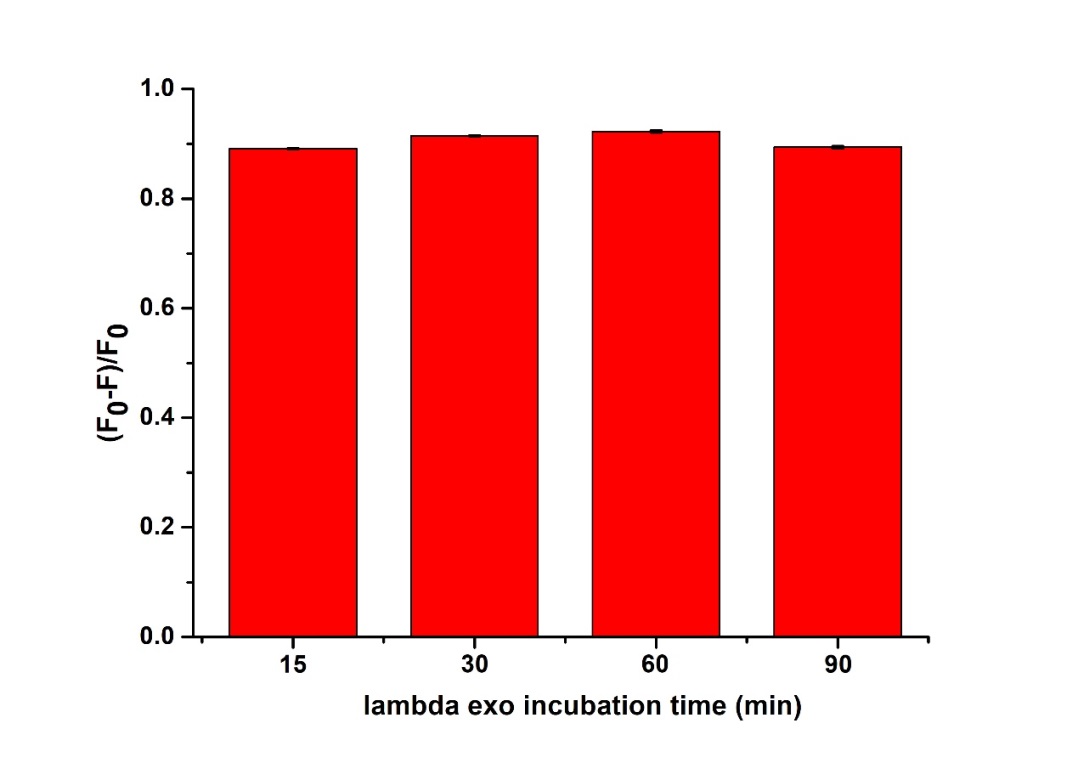
**Figure S1**. The optimization of SA aptamer sequences. (A) Sequences and secondary structures of three designed SA aptamers. Bases in pink, blue and red, respectively, represent the loop, the bulge, and different bases among aptamers. (B) Signal-Background ratio (SBR) of various AP (SA aptamers and their corresponding cDNA) incubated with or without 10 U/mL ALP. SBR was calculated by F0/F, where F0 and F are fluorescence intensities without and with ALP, respectively. Error bars indicate the standard deviations of three samples.



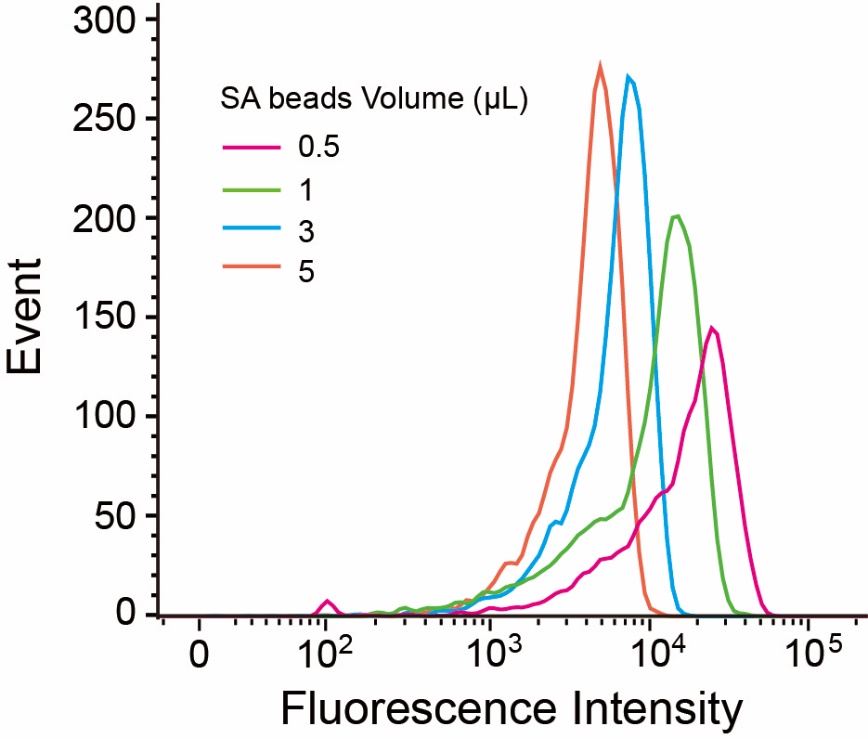
**Figure S2.** Response of AP to ALP with different ALP incubation times in a buffer solution. F0 and F are fluorescence intensities without and with ALP, respectively. The concentrations of ALP and λ exo are 10 U/mL and 100 U/mL, respectively. Error bars indicate the standard deviations of three samples.



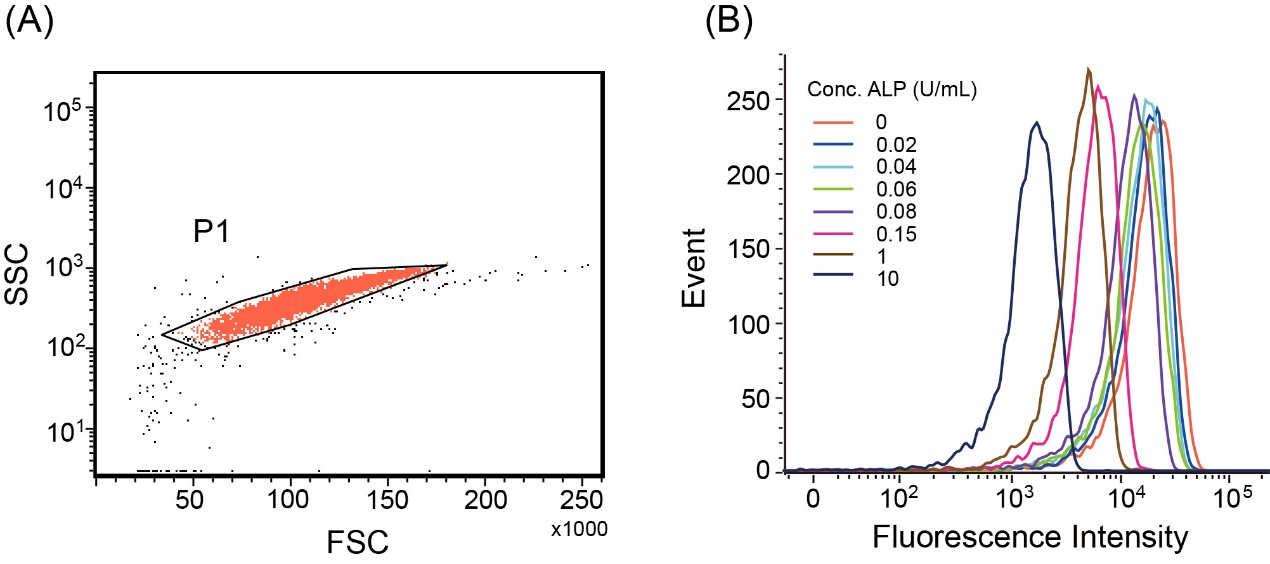
**Figure S3.** Response of AP to ALP with different concentrations of λ exo in a buffer solution. F0 and F are fluorescence intensities without and with ALP, respectively. The concentration of ALP is 10 U/mL. Error bars indicate the standard deviations of three samples.



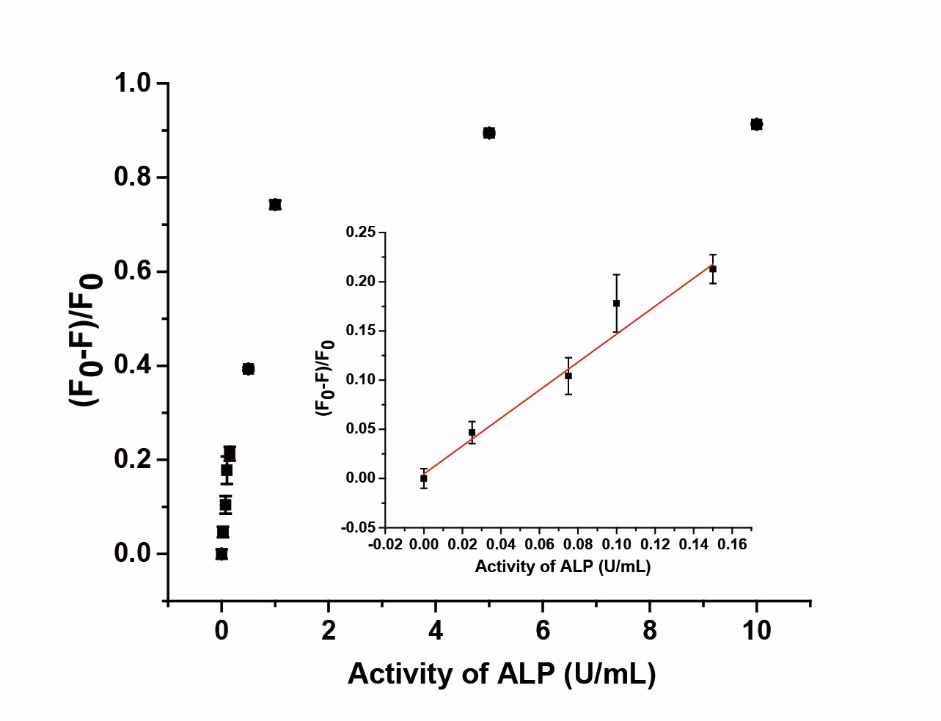
**Figure S4.** Response of AP to ALP with different incubation time of λ exo in a buffer solution. F0 and F are fluorescence intensities without and with ALP, respectively. The concentration of ALP is 1 U/mL. Error bars indicate the standard deviations of three samples.



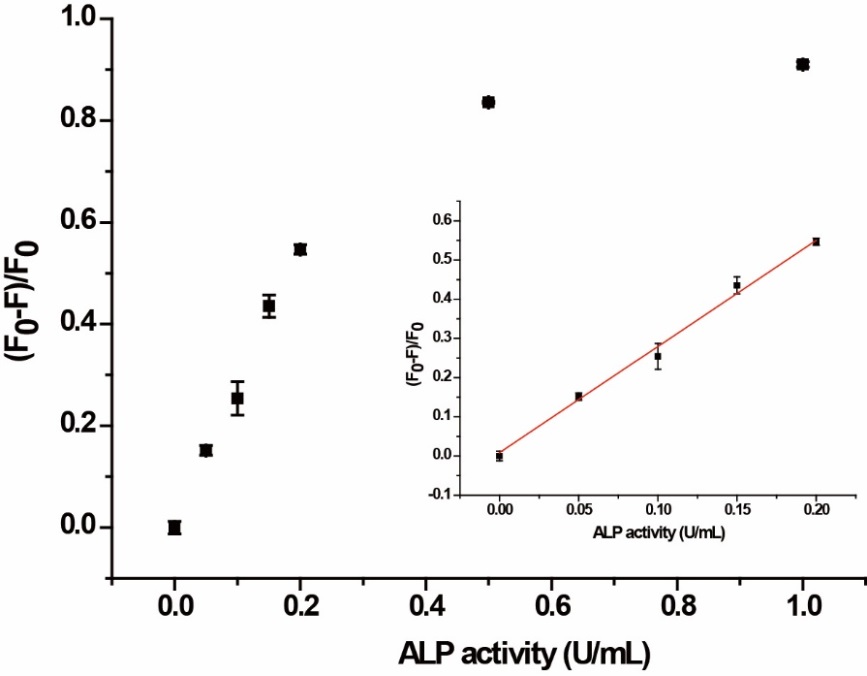
**Figure S5.** Fluorescence Intensity of different concentrations of SA beads for binding to 200 nM SA3 in a buffer solution.



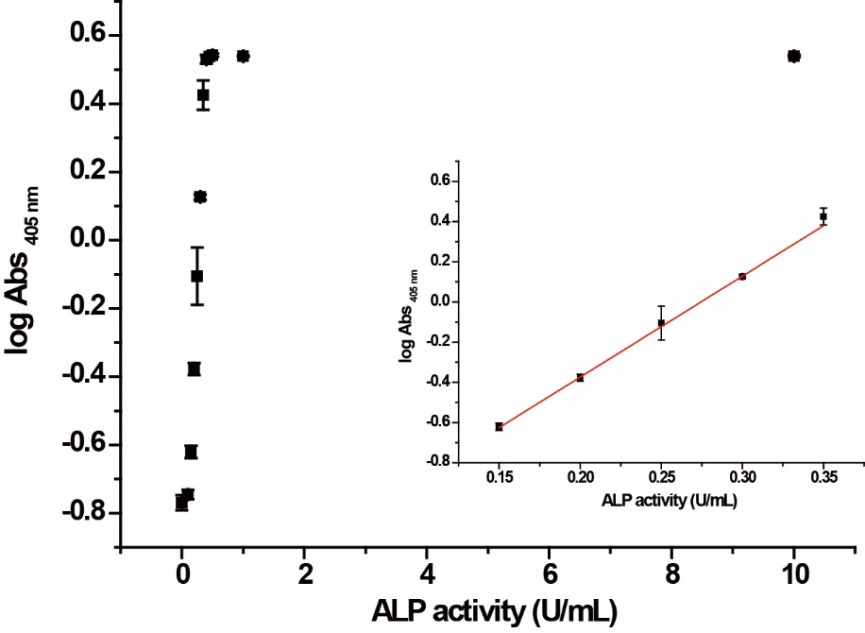
**Figure S6.** (A) A typical scatter diagram of beads. (B) Original flow cytometry data of detection of ALP for plotting Figure 3A.



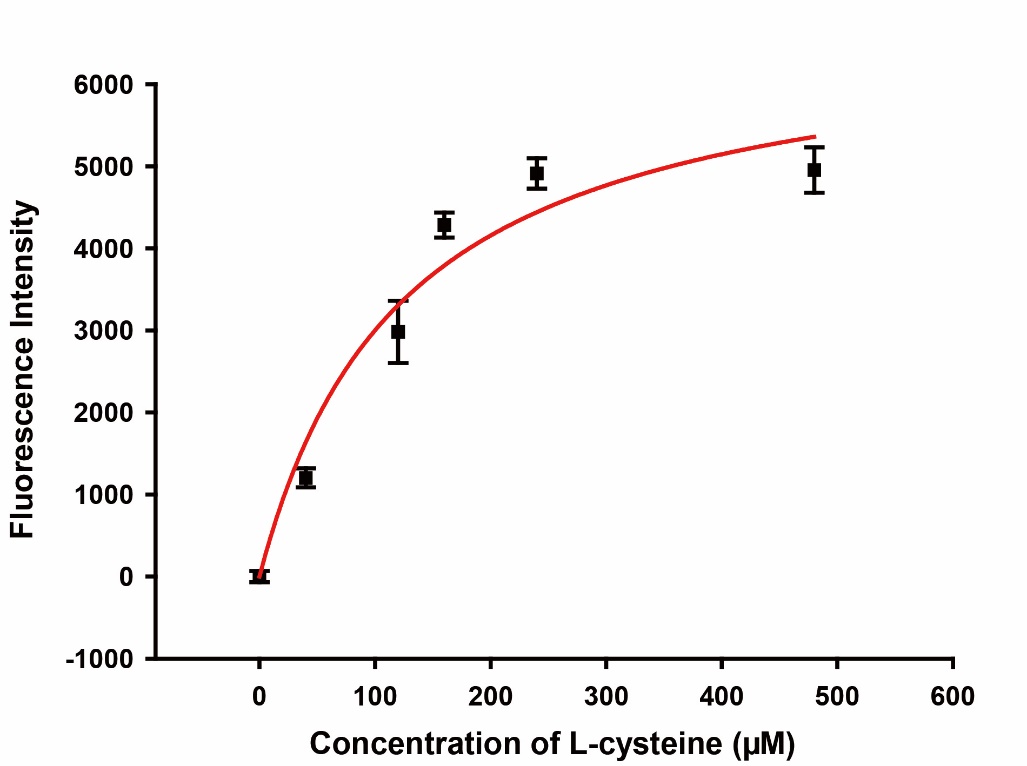
**Figure S7.** Response of quantitative detection of ALP range from 0 to 10 U/mL in 20% human serum monitored by flow cytometry. Inset is the calibration curve range from 0.025 to 0.15 U/mL. F0 and F are fluorescence intensities without and with ALP, respectively. Error bars indicate the standard deviations of three samples.



**Figure S8.** Quantitative detection of ALP ranging from 0 to 1 U/mL in spiked Hela cell lysate of 105 cell/mL with AP monitored by flow cytometry. The calibration curve ranges from 0.05 to 0.20 U/mL and the limit of detection is 0.014 U/mL (3σ/S). Error bars indicate the standard deviations of three samples.



**Figure S9.** Quantitative detection of ALP ranging from 0 to 10 U/mL with PNPP substrate by monitoring the absorption at 405 nm. The calibration curve ranges from 0.15 to 0.35 U/mL and the limit of detection is 0.13 U/mL (3σ/S). Error bars indicate the standard deviations of three samples.

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**Figure S10.** Response of the AP to 2 U/mL ALP with inhibitor L-cysteine. Error bars indicate the standard deviations of three samples.

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