

Supplementary Material

Fine-tuning and remodeling of pectins play a key role in the maintenance of cell adhesion

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Supplementary Figure 1. Monomers content, HGs quantity and HGs fragments released (A) Relative content of each monomers hydrolysed by TFA of dried cell wall of 4-days-old dark grown of seedlings Col-0, *qua2-1, qua2-1/esmd1-1 and esmd1-1*. (B) Truncated violin plot of homogalacturonan quantity corresponding to galacturonic acid content minus rhamnose content,

represented in µg/mg of dried cell wall of Col-0, qua2-1, qua2-1/esmd1-1 and esmd1-1. Red line represents the median, black line the quartiles (n=4 biological replicates per genotype), *, P < 0.05, Mann & Whitney test. (C)Truncated violin plot of the quantity of HGs fragments & monomers released by PG Aspergillus aculeatus digestion of dried cell wall seedlings 4 days dark grown of Col-0, qua2-1, qua2-1/esmd1-1 and esmd1-1. Red line represents the median, black line the quartiles, (n ≤ 4 biological replicates per genotype). *, P < 0.058, Mann-Whitney test.







Supplementary Figure 2. HGs fragments released significantly different.

(A) Heatmap of all of the HGs fragments released by *PG Aspergillus aculeatus* significantly different between Col0 and *qua2-1* (48 rows) for 3 samples (columns). Annotation labels refer to HGs fragments structure. Rows are centered; unit variance scaling is applied to rows. Both rows and columns are clustered using McQuitty distance and maximum linkage. (B) Heatmap of all of the HGs fragments released by *PG Aspergillus aculeatus* significantly different between Col0 and *esmd1-1* (39 rows) for 3 samples (columns). (C) Heatmap of all of the HGs fragments released by *PG Aspergillus aculeatus* significantly different setues the HGs fragments released by *PG Aspergillus aculeatus* significantly different between *qua2-1* and *qua2-1/esmd1-1* (46 rows) for 3 samples (columns). -H₂O indicates endogenous pectin lyase action. Methyl-esterified (*), acetyl-esterified (°) and methyl/acetyl-esterified (•) HGs fragments specific *to esmd1-1* are highlighted. For (A), (B), and (C). *n* = 3 *biological replicates per genotype*. Mann-Whitney test.



Supplementary Figure 3. Fine structure of xyloglucans and cellulose content

(A) Truncated violin plot of the quantity of xyloglucans fragment digested by *endo-cellulase* applied on dried cell wall seedlings 4 days dark grown of Col-0, *qua2-1*, *qua2-1/esmd1-1* and *esmd1-1*. Red *line represents the median, black line the quartiles, black dot represents biological replicate* (n = 4*biological replicates per genotype*) *, P < 0.05 Mann-Whitney test. (B) Relative content (%) of xyloglucans fragment digested by *endo-cellulase* applied on dried cell wall seedlings 4 days dark grown of Col-0, *qua2-1*, *qua2-1/esmd1-1* and *esmd1-1*. (C) Truncated violin plot of cellulose content (μ g/mg of dried cell wall) of 4-days-old dark grown seedlings of Col-0, *qua2-1*, *qua2-1/esmd1-1* and *esmd1-1* (n = 4 biological replicates per genotype) Mann-Whitney test.



Supplementary Figure 4. Effect of enzymes on cell adhesion.

Ruthenium red staining of enzyme application on 4 days dark grown seedlings (from day 2) on Col0 : (A) empty vector, (B) PAE7, (C) PAE12, (D) PME41, (E) PME35, (F) PME53 and (G) PMEI4 and *qua2-1/esmd1-1* : (H to N) same order of protein

1.1 Supplementary Table

Gene name	Forward primer sequence	Reverse primer sequence
CLATHRIN	5'-GTTTGGGAGAAGAGCGGTTA-3'	5'-CTGATGTCACTGAACCTGAACTG -3'
(At5g46630)		
APT1	5'-GAGACATTTTGCGTGGGATT-3'	5'-ATTTTAAGTGGAACA -3'
(At1g27450)		
PME53	5'-TTTGTATCTTGGGAGGGCATGGG-3'	5'-ACTGCCCATAGAACACCGTCATC -3'
(At5g19730)		
PME41	5'-TACATCGCCGAACTTCGTTGCC -3'	5'-GCTTCTCTGGTCCAGCGGTATTTC -3'
(At4g02330)		
PME35	5'-CCGCCATGGGAGATGGATTCATAG -3'	5'-AGTTTGGTCCGGCACTGTTCAC -3'
(At3g59010)		
PMEi4	5'-AAACGGCATGCAACTCAACAAC -3'	5'-CGGACTTGATGGTGGAGGAATAGG -3'
(At4g25250)		
PAE7	5'-ATGCGAGCATTGTCACCGGTTC -3'	5'- CCGACTGCTTTCGCAATTCTCG -3'
(At4g19410)		
PAE12	5'-CTTGGTTTGCCGATGATTCT -3'	5'-ACCAAGTTGTGGCAGCTCTT -3'
(At3g05910)		

Supplementary Table 1. List of primers used for qPCR